

SHORT REPORT

Open Access

# *In-vitro* renal epithelial cell infection reveals a viral kidney tropism as a potential mechanism for acute renal failure during Middle East Respiratory Syndrome (MERS) Coronavirus infection

Isabella Eckerle<sup>1</sup>, Marcel A Müller<sup>1</sup>, Stephan Kallies<sup>1</sup>, Daniel N Gotthardt<sup>2</sup> and Christian Drosten<sup>1\*</sup>

## Abstract

**Background:** The Middle East Respiratory Syndrome Coronavirus (MERS-CoV) causes symptoms similar to Severe Acute Respiratory Syndrome Coronavirus (SARS-CoV), yet involving an additional component of acute renal failure (ARF) according to several published case reports. Impairment of the kidney is not typically seen in Coronavirus infections. The role of kidney infection in MERS is not understood.

**Findings:** A systematic review of communicated and peer-reviewed case reports revealed differences in descriptions of kidney involvement in MERS versus SARS patients. In particular, ARF in MERS patients occurred considerably earlier after a median time to onset of 11 days (SD  $\pm 2,0$  days) as opposed to 20 days for SARS, according to the literature. *In-situ* histological staining of the respective cellular receptors for MERS- and SARS-Coronavirus showed highly similar staining patterns with a focus of a receptor-specific signal in kidney epithelial cells. Comparative infection experiments with SARS- and MERS-CoV in primary human kidney cells versus primary human bronchial epithelial cells showed cytopathogenic infection only in kidney cells, and only if infected with MERS-CoV. Kidney epithelial cells produced almost 1000-fold more infectious MERS-CoV progeny than bronchial epithelial cells, while only a small difference was seen between cell types when infected with SARS-CoV.

**Conclusion:** Epidemiological studies should analyze kidney impairment and its characteristics in MERS-CoV. Virus replication in the kidney with potential shedding in urine might constitute a way of transmission, and could explain untraceable transmission chains leading to new cases. Individual patients might benefit from early induction of renoprotective treatment.

**Keywords:** Middle East Respiratory Syndrome, Acute renal failure, Human Coronavirus, Renal epithelial cells, Dipeptidyl-peptidase-4, Angiotensin-converting-enzyme-2

Coronaviruses (CoVs) cause human disease with symptoms ranging from mild respiratory symptoms to severe pneumonia [1]. In September 2012, a novel CoV termed the Middle East Respiratory Syndrome (MERS)-CoV emerged on the Arabian Peninsula with 163 laboratory-confirmed cases including 71 deaths so far (World Health Organization, December 2nd, [http://www.who.int/csr/don/2013\\_12\\_02/en/index.html](http://www.who.int/csr/don/2013_12_02/en/index.html)). Due to its distribution in several countries of the Arabian Peninsula there is a risk of global spread through travel and pilgrimage, posing a potential threat to global public health.

The clinical picture of MERS-CoV infection is characterized by acute atypical pneumonia and respiratory failure, resembling symptoms caused by SARS-CoV [2-5]. Acute renal failure (ARF) was described in a number of MERS cases, with potential influence on disease severity [2,3,5-7]. ARF has neither been a typical feature of SARS, nor has it been commonly observed in infections caused by any other CoV, including human HCoV-NL63, -229E, -OC43,

The clinical picture of MERS-CoV infection is characterized by acute atypical pneumonia and respiratory failure, resembling symptoms caused by SARS-CoV [2-5]. Acute renal failure (ARF) was described in a number of MERS cases, with potential influence on disease severity [2,3,5-7]. ARF has neither been a typical feature of SARS, nor has it been commonly observed in infections caused by any other CoV, including human HCoV-NL63, -229E, -OC43,

\* Correspondence: [drosten@virology-bonn.de](mailto:drosten@virology-bonn.de)

<sup>1</sup>Institute of Virology, University of Bonn Medical Centre, Sigmund-Freud-Strasse 25, Bonn 53127, Germany

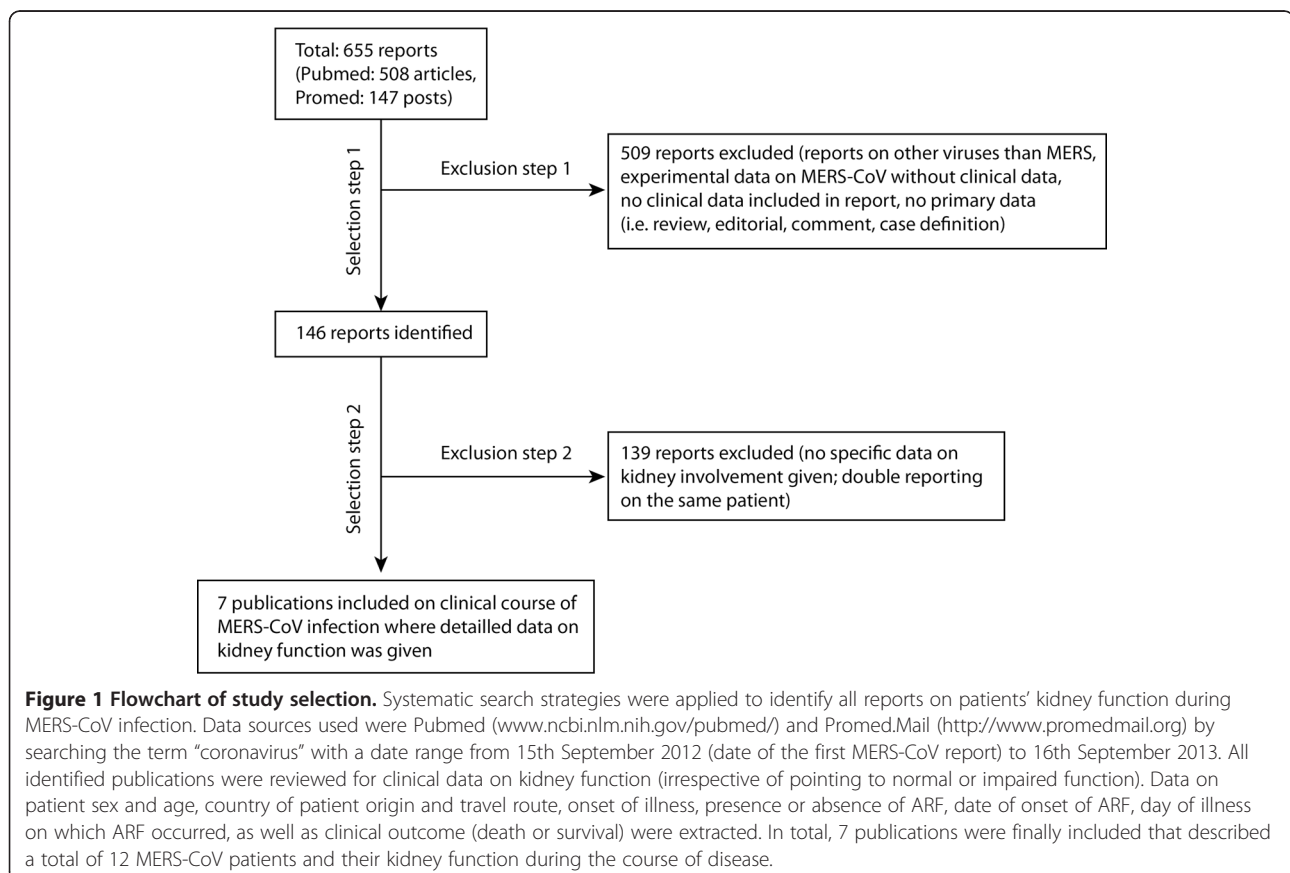
Full list of author information is available at the end of the article

and -HKU1. Of note, the majority (76%) of MERS-CoV patients were reported to have underlying medical conditions such as diabetes, chronic cardiac disease and chronic renal disease [4,8].

It has been shown recently that MERS-CoV enters target cells not via the SARS-CoV receptor Angiotensin-converting-enzyme-2 (ACE-2), but via binding to Dipeptidyl-peptidase 4 (DPP-4) [9,10]. Both, ACE-2 and DPP-4 are expressed in several human tissues, including the kidney [11,12]. However, kidney infection has not been compared between MERS-CoV and SARS-CoV, and no post-mortem investigations on patients who succumbed to MERS-CoV have been done. To assess the potential role of kidney affection in MERS-CoV infection, we reviewed all peer-reviewed or communicated reports on MERS cases for information on kidney involvement. The search was performed in PubMed and Promed.Mail (<http://www.promedmail.org>) using the search term “coronavirus” on items published between 15th September 2012 and 16th September 2013. The broad search term was chosen due to inconsistencies of nomenclature before the consensus name MERS-CoV was announced [13].

In total, the search revealed 655 publications, including 508 articles in PubMed, and 147 reports in ProMed (Figure 1). All identified publications were reviewed in

order to identify clinical descriptions of MERS-CoV infection that included information on kidney function, irrespective of whether they were pointing to normal or impaired function. In cases where two reports on the same patient were available, only the report with the more detailed clinical description was selected. From all identified reports, we extracted information on patient sex and age, country of patient origin and travel route, onset of illness, presence or absence of ARF, date of onset of ARF, day of illness on which ARF occurred, as well as clinical outcome (death or survival). Among 21 publications describing a total of 111 MERS-CoV patients (double-reporting of the same patients could not be excluded), a total of 7 publications were identified that specifically addressed kidney function [2,3,5-7,14,15]. In these 7 publications, a total of 12 MERS patients were described with disease onset dates ranging from April 2012 to March 2013 (Table 1). All patients were male with a mean age of 47 years (range 25–73 years), originating from Jordan, Saudi Arabia, and Qatar, including imported cases to the United Kingdom (UK), Germany and France. Nine of those 12 patients (75%) developed ARF during the acute course of MERS-CoV infection. For 6 of those 9 patients, dates of onset of ARF were available, suggesting a median time of 11 days (SD  $\pm 2,0$  days) from first symptoms to onset of



**Table 1 MERS-CoV patients with explicit reports on presence or absence of acute renal failure (ARF)**

Nr.	Patient	Country	Onset of illness	ARF	Onset ARF	ARF on day of illness	Outcome	Reference
1	25 y m	Jordan	Apr 12, day nd	Yes	nd	nd	Fatal	[7]
2	60 y m	Saudi Arabia	7 Jun 12	Yes	16 Jun 12	10	Fatal	[2]
3	49 y m	UK ex Qatar	3 Sep 12	Yes	14 Sep 12	12	Fatal	[15]
4	45 y m	Saudi Arabia	Nov 12, day nd	Yes	nd	nd	Recovered	[7]
5	45 y m	Germany ex Qatar	5 Oct 12	Yes	nd	nd	Recovered	[14]
6	70 y m	Saudi Arabia	5 Oct 12	Yes	16 Oct 12	12	Fatal	[3]
7	39 y m	Saudi Arabia	24 Oct 12	No	-	-	Fatal	[3]
8	16 y m	Saudi Arabia	3 Nov 12	No	-	-	Recovered	[3]
9	31 y m	Saudi Arabia	4 Nov 12	No	-	-	Recovered	[3]
10	64 y m	France ex Saudi Arabia	22 Apr 13	Yes	30 Apr 13	9	Fatal	[6]
11	51 y m	France	8 May 13	Yes	14 May 13	7	Recovered	[6]
12	73 y m	Germany ex Saudi Arabia	8 Mar 13	Yes	22 Mar 13	12	Fatal	[5]

y – years of age; m – male; nd – no data available.

ARF. Six of 9 (67%) patients with ARF had a fatal outcome, while one of three patients without ARF died. However, the differences found for the fatality rates in cases with and without ARF during MERS-CoV infection were not statistically significant.

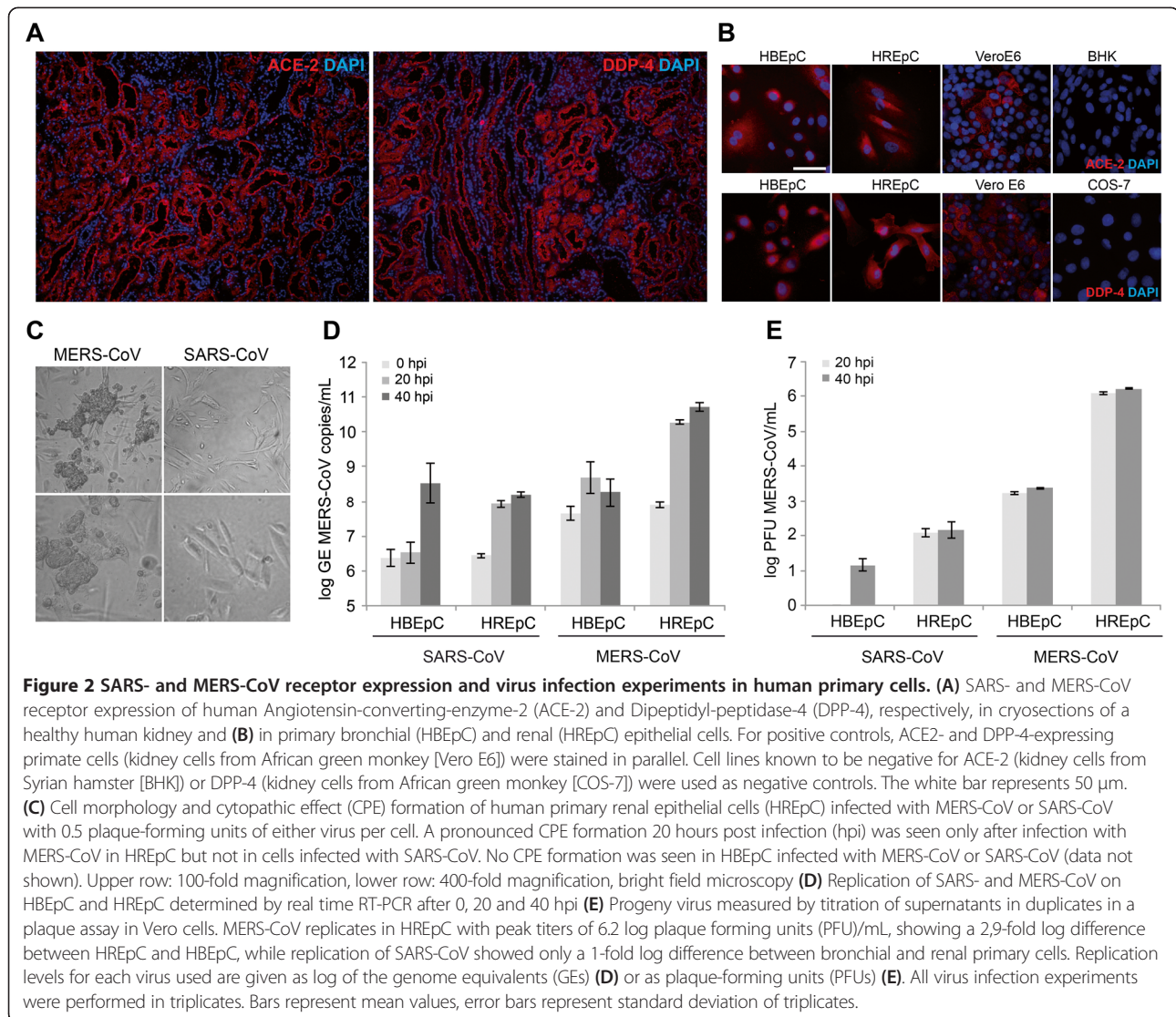
To investigate a laboratory surrogate for susceptibility of the kidney to MERS-CoV in comparison to SARS-CoV, we analyzed cryoslides of a healthy human kidney for ACE-2 and DPP-4 expression by immunofluorescence. Staining using polyclonal goat-anti-human ACE-2 immunoglobulin (R&D systems) and polyclonal goat-anti-human DPP-4 immunoglobulin (R&D systems) showed that both receptors are abundantly expressed in the healthy human kidney, and both receptors are predominantly found in the epithelial layer of the renal ducts (Figure 2A), as well as in human primary epithelial cells derived from these ducts and in epithelial cells from the human airway tract (Figure 2B).

To test whether kidney epithelial cells can actually be infected *in-vitro*, we performed comparative infection experiments with MERS- and SARS-CoV, using defined cultures of primary kidney epithelial cells (HREpC, Promocell, Heidelberg). As the bronchoalveolar epithelium of the lung constitutes the primary target compartment for both viruses, infection of human primary bronchial epithelial cells (HBEPc, Promocell, Heidelberg) was studied in parallel. As summarized in Figure 2, primary cells up to passage 4 were seeded at densities of  $4 \times 10^5$ /mL and inoculated with either MERS-CoV strain EMC/2012, or SARS-CoV strain Frankfurt-1 at multiplicities of infection (MOI) of 0.5 for 1 hour. Cells were washed twice and supernatants were harvested immediately after washing, as well as 20 and 40 hours later. Cells were checked regularly for appearance of cytopathic effects (CPE) by light microscopy. Replication of both viruses was quantified by titration of supernatants in duplicate plaque assays on Vero cells as described previously, as well as by real-time RT-PCR [9,16].

Pronounced CPE consisting of visible and strong cell lysis occurred only in HREpC, and only upon infection with MERS-CoV (Figure 2C). Neither in HBEPc upon MERS-CoV infection, nor in HBEPc or HREpC upon SARS-CoV infection any CPE was observed. Virus replication was nevertheless detectable by real time RT-PCR in all experiments, confirming successful infection (Figure 1D). The CPE in HREpC consisted of rounding and detachment of cells, leading to cell death in the majority of cells already after 20 hours. Comparative quantification of infectious virus production between SARS- and MERS-CoV showed that MERS-CoV replicates to high titers in HREpC with almost 1000-fold higher concentrations of infectious MERS-CoV progeny than in HBEPc. In SARS-CoV infection, only a small difference between primary kidney and bronchial epithelial cells was seen (Figure 1E).

Our results from *in-vitro* infection experiments suggest differences in kidney cell involvement might exist between SARS- and MERS-CoV infection. In a retrospective analysis of 536 SARS cases, only in 36 (6.7%) patients acute renal impairment was diagnosed. ARF occurred rather late in the course of disease after a median of 20 days from the appearance of first symptoms [17]. In our literature review of MERS cases, ARF occurred much earlier, namely in 6 of 9 sufficiently-described patients after a median of 11 days. To date, it is difficult to compare these clinical data with experiences from animal models. The only animal that exhibits clinical signs similar to human MERS-CoV infection is the rhesus macaque, but none of the available studies reported kidney failure in macaques [18-20]. Only very small amounts of viral RNA were detected in kidney tissue, and none in urine. Macaques might be of limited value for the study of MERS-CoV-specific kidney pathology as seen in humans.

While no autopsy data have become available for patients infected with MERS-CoV, post-mortem studies



have been done systematically in SARS victims. Histopathological findings revealed mainly acute tubular necrosis without evidence of glomerular pathology, which is considered to be a consequence of systemic inflammatory response in the context of multi-organ failure, rather than a specific effect of viral infection of the kidney [17]. SARS-CoV has never been successfully isolated from *post mortem* kidney tissue of infected patients [21]. *In-vitro* studies of SARS-CoV with immortalized human proximal tubular epithelial cells showed replication without cell impairment as seen in our study, while no infection of podocyte cell lines and only low-level replication in glomerular mesangial cells (MC) was seen, providing further evidence against specific involvement of the renal tract in SARS-CoV infection [22]. In contrast, MERS-CoV was shown to efficiently replicate in a broad range of bat, primate and also human kidney epithelial cell lines that

are commonly used as laboratory models [9]. The abundant expression of both viruses' entry receptors in kidney epithelium argues against receptor-dependent limitations to viral kidney tropism. Specific interference of MERS-CoV with the induction of the interferon response provides one of many possible explanations for an increased capacity of MERS-CoV to replicate in kidney cells [23]. The clear differences in viral permissiveness of primary kidney epithelial cells suggest these cells to be appropriate models for the identification of host-specific restriction factors in the future. We are aware that cell culture resembles a limited model of viral infection which will not reflect the complexity of infection *in-vivo*. Nevertheless, our results have some congruence with clinical observations in a well-investigated MERS case [5]. Virus in the urine has been detected as late as day 12 of illness, albeit at low concentrations. Virus was detected in the urine but not in

the blood, which could indicate autonomous virus replication in the kidneys. Of note, by using a model to assess the ability of CoVs to persist in the environment, it was shown that MERS-CoV has similarities to viruses that are transmitted via the fecal-oral route [24]. The authors suggest that this could hint for oral-urine transmission of MERS-CoV. Further data on virus shedding in the urine over the course of disease are urgently needed. If the kidney should indeed constitute a site of primary virus replication, shedding of virus in the urine might provide a possible source for human-to-human transmission, especially in health-care settings or among close family contacts [3,6,7]. Clinical guidelines should consider the possibility that MERS patients may benefit from early induction of renoprotective treatment.

#### Competing interests

The authors declare that they have no competing interests.

#### Authors' contributions

IE, MAM designed the experiments, IE, MAM, SK performed experiments and analysis, IE, CD wrote the manuscript, IE, DNG, CD contributed to the final version of the manuscript. All authors read and approved the final manuscript.

#### Acknowledgements

This work was funded by FP7 European research project ANTIGONE (contract number 278976), as well as an infrastructural grant from the German Centre for Infection Research (DZIF). We thank the National Center for Tumor Diseases (NCT), Heidelberg, for tissue sections. We thank Bart Haagmans, Department of Virology, Erasmus Medical Center, Rotterdam, for COS-7 cells.

#### Author details

<sup>1</sup>Institute of Virology, University of Bonn Medical Centre, Sigmund-Freud-Strasse 25, Bonn 53127, Germany. <sup>2</sup>Department of Internal Medicine IV, University Hospital Heidelberg, Im Neuenheimer Feld 410, Heidelberg 69120, Germany.

Received: 17 October 2013 Accepted: 13 December 2013

Published: 23 December 2013

#### References

1. Peiris JS, Guan Y, Yuen KY: Severe acute respiratory syndrome. *Nat Med* 2004, **10**:S88–S97.
2. Zaki AM, van Boheemen S, Bestebroer TM, Osterhaus AD, Fouchier RA: Isolation of a novel coronavirus from a man with pneumonia in Saudi Arabia. *N Engl J Med* 2012, **367**:1814–1820.
3. Memish ZA, Zumla AI, Al-Hakeem RF, Al-Rabeeh AA, Stephens GM: Family cluster of Middle East respiratory syndrome coronavirus infections. *N Engl J Med* 2013, **368**:2487–2494.
4. Assiri A, McGeer A, Perl TM, Price CS, Al Rabeeh AA, Cummings DA, Alabdullatif ZN, Assad M, Almulhim A, Makhdoom H, et al: Hospital outbreak of Middle East respiratory syndrome coronavirus. *N Engl J Med* 2013, **369**:407–416.
5. Drosten C, Seilmaier M, Corman VM, Hartmann W, Scheible G, Sack S, Guggemos W, Kallies R, Muth D, Junglen S, et al: Clinical features and virological analysis of a case of Middle East respiratory syndrome coronavirus infection. *Lancet Infect Dis* 2013, **13**:745–751.
6. Guery B, Poissy J, el Mansouf L, Sejourne C, Ettahar N, Lemaire X, Vuotto F, Goffard A, Behillil S, Enouf V, et al: Clinical features and viral diagnosis of two cases of infection with Middle East respiratory syndrome coronavirus: a report of nosocomial transmission. *Lancet* 2013, **381**:2265–2272.
7. Pollack MP, Pringle C, Madoff LC, Memish ZA: Latest outbreak news from ProMED-mail: novel coronavirus – Middle East. *Int J Infect Dis* 2013, **17**:e143–e144.
8. The WHO MERS-CoV Research Group: State of knowledge and data gaps of Middle East Respiratory Syndrome Coronavirus (MERS-CoV) in Humans. *PLOS Currents Outbreaks* 2013.
9. Muller MA, Raj VS, Muth D, Meyer B, Kallies S, Smits SL, Wolny R, Bestebroer TM, Specht S, Suliman T, et al: Human coronavirus EMC does not require the SARS-coronavirus receptor and maintains broad replicative capability in mammalian cell lines. *M Bio* 2012, **3**. doi:10.1128/mBio.00515-12.
10. Raj VS, Mou H, Smits SL, Dekkers DH, Muller MA, Dijkman R, Muth D, Demmers JA, Zaki A, Fouchier RA, et al: Dipeptidyl peptidase 4 is a functional receptor for the emerging human coronavirus-EMC. *Nature* 2013, **495**:251–254.
11. Hamming I, Cooper ME, Haagmans BL, Hooper NM, Korstanje R, Osterhaus AD, Timens W, Turner AJ, Navis G, van Goor H: The emerging role of ACE2 in physiology and disease. *J Pathol* 2007, **212**:1–11.
12. Lambeir AM, Durinx C, Scharpe S, De Meester I: Dipeptidyl-peptidase IV from bench to bedside: an update on structural properties, functions, and clinical aspects of the enzyme DPP IV. *Crit Rev Clin Lab Sci* 2003, **40**:209–294.
13. de Groot RJ, Baker SC, Baric RS, Brown CS, Drosten C, Enjuanes L, Fouchier RA, Galiano M, Gorbalenya AE, Memish ZA, et al: Middle East respiratory syndrome coronavirus (MERS-CoV): announcement of the Coronavirus Study Group. *J Virol* 2013, **87**:7790–7792.
14. Buchholz U, Muller MA, Nitsche A, Sanewski A, Wevering N, Bauer-Balci T, Bonin F, Drosten C, Schweiger B, Wolff T, et al: Contact investigation of a case of human novel coronavirus infection treated in a German hospital, October–November 2012. *Euro Surveill* 2013, **18**.
15. Bermingham A, Chand MA, Brown CS, Aarons E, Tong C, Langrish C, Hoschler K, Brown K, Galiano M, Myers R, et al: Severe respiratory illness caused by a novel coronavirus, in a patient transferred to the United Kingdom from the Middle East, September 2012. *Euro Surveill* 2012, **17**:20290.
16. Corman VM, Eckerle I, Bleicker T, Zaki A, Landt O, Eschbach-Bludau M, van Boheemen S, Gopal R, Ballhause M, Bestebroer TM, et al: Detection of a novel human coronavirus by real-time reverse-transcription polymerase chain reaction. *Euro Surveill* 2012, **17**.
17. Chu KH, Tsang WK, Tang CS, Lam MF, Lai FM, To KF, Fung KS, Tang HL, Yan WW, Chan HW, et al: Acute renal impairment in coronavirus-associated severe acute respiratory syndrome. *Kidney Int* 2005, **67**:698–705.
18. Falzarano D, de Wit E, Rasmussen AL, Feldmann F, Okumura A, Scott DP, Brining D, Bushmaker T, Martellaro C, Baseler L, et al: Treatment with interferon-alpha2b and ribavirin improves outcome in MERS-CoV-infected rhesus macaques. *Nat Med* 2013, **19**:1313–1317.
19. Munster VJ, de Wit E, Feldmann H: Pneumonia from human coronavirus in a macaque model. *N Engl J Med* 2013, **368**:1560–1562.
20. de Wit E, Rasmussen AL, Falzarano D, Bushmaker T, Feldmann F, Brining DL, Fischer ER, Martellaro C, Okumura A, Chang J, et al: Middle East respiratory syndrome coronavirus (MERS-CoV) causes transient lower respiratory tract infection in rhesus macaques. *Proc Natl Acad Sci* 2013, **110**:16598–16603.
21. Tang JW, To KF, Lo AW, Sung JJ, Ng HK, Chan PK: Quantitative temporal-spatial distribution of severe acute respiratory syndrome-associated coronavirus (SARS-CoV) in post-mortem tissues. *J Med Virol* 2007, **79**:1245–1253.
22. Pacciarini F, Ghezzi S, Canducci F, Sims A, Sampaolo M, Ferioli E, Clementi M, Poli G, Conaldi PG, Baric R, Vicenzi E: Persistent replication of severe acute respiratory syndrome coronavirus in human tubular kidney cells selects for adaptive mutations in the membrane protein. *J Virol* 2008, **82**:5137–5144.
23. Niemeyer D, Zillinger T, Muth D, Zieleski F, Horvath G, Suliman T, Barchet W, Weber F, Drosten C, Muller MA: Middle East respiratory syndrome coronavirus accessory protein 4a is a type I interferon antagonist. *J Virol* 2013. doi:10.1128/JVI.01845-13. Epub 2013 Sep 11.
24. Goh GK, Dunker AK, Uversky V: Prediction of intrinsic disorder in MERS-CoV/HCoV-EMC supports a high oral-fecal transmission. *PLOS Currents Outbreaks* 2013.

doi:10.1186/1743-422X-10-359

Cite this article as: Eckerle et al.: *In-vitro* renal epithelial cell infection reveals a viral kidney tropism as a potential mechanism for acute renal failure during Middle East Respiratory Syndrome (MERS) Coronavirus infection. *Virology Journal* 2013 **10**:359.