

Original paper

Estimation of the risk of fibrosis in children with nonalcoholic fatty liver disease

Paweł Małecki, Magdalena Figlerowicz, Paweł Kemnitz, Katarzyna Mazur-Melewska, Wojciech Stuzewski, Anna Mania

Department of Infectious Diseases and Child Neurology, Poznan University of Medical Sciences, Poznan, Poland

Abstract

Aim of the study: Assessment of liver fibrosis as a predictive factor of liver-related mortality in children with nonalcoholic fatty liver disease (NAFLD) is crucial. This study aims to estimate the risk of fibrosis using noninvasive markers.

Material and methods: The study group of 49 children with NAFLD (age range 3-16, mean 10.51 ± 3.18 years) was created. The diagnosis was based on clinical picture, abdominal ultrasound, and laboratory tests; four children underwent liver biopsy. Then homeostatic model assessment (HOMA-IR) and noninvasive hepatic fibrosis scores were calculated, and patients were divided into four groups depending on body mass index (BMI, obese vs. lean) and aminotransferases.

Results: 71.43% of patients were obese, and 14.29% were overweight. We found that overweight children had lower mean corpuscular volume (MCV) than lean patients. In a group of patients with a high risk of fibrosis or significant fibrosis due to pediatric NAFLD fibrosis score (PNFS), higher alanine aminotransferase (ALT) to platelet ratio (APRI) values were observed. The highest values of APRI were found in a group of lean patients with elevated aminotransferases and the highest value of PNFS – among obese patients with elevated aminotransferases. A strong significant correlation between APRI and PNFS was found ($r = 0.88$).

Conclusions: Apart from aminotransferase activity, complete blood count should be assessed looking for lower MCV caused by iron deficiency. In contrast to FIB-4 (fibrosis score), PNFS and APRI proved to be more accurate in our group. PNFS seems to be appropriate to evaluate fibrosis in a noninvasive diagnostic algorithm.

Key words: children, fatty liver, serum markers of fibrosis, laboratory indexes.

Address for correspondence:

Dr. Paweł Małecki, Department of Infectious Diseases and Child Neurology, Poznan University of Medical Sciences, 27/33 Szpitalna St., 60-572 Poznan, Poland, e-mail: pmalecki@ump.edu.pl

Introduction

Nonalcoholic fatty liver disease (NAFLD) is the most common chronic liver disease in children in developed countries [1]. The estimated prevalence of NAFLD in children varies from 7.6% in the general population to 34.2% in obese patients [2].

The spectrum of disorders from the fatty liver disease group extends from simple steatosis, nonalcoholic fatty liver (NAFL) to nonalcoholic steatohepatitis (NASH) with the presence of injury, inflammation, and fibrosis of the liver tissue. The diagnosis of NAFLD is based on histopathological or imaging testing in the absence of other causes leading to liver damage (e.g., alcohol intake, drug

poisoning, viral hepatitis, metabolic diseases) [3]. Liver biopsy remains the gold standard of definitive diagnosis (steatosis involving > 5% of hepatocytes); however, because of its invasive character and possible complications it is unacceptable as an initial diagnostic tool. It is not suitable for repeated long-term monitoring in children.

In children, life-threatening complications of NAFLD are rare, but it is essential to emphasize that other conditions associated with chronic liver disease or metabolic syndrome can increase overall mortality in adulthood. The studies of the natural history of patients with NAFLD revealed that fibrosis is an independent predictive factor of liver-related mortality [4]. Over the past years, scientists have attempted to create a universal tool

allowing for repetitive long-term non-invasive diagnostics, including fibrosis assessment, using primary laboratory and imaging tests or new biochemical markers.

According to published reports, markers used in adults may not be suitable for children and adolescents [5].

A model based on age, waist circumference and triglyceride (pediatric NAFLD fibrosis index – PNFI) was found to be inadequately useful [6]. Therefore the pediatric NAFLD fibrosis score (PNFS) was developed in 2014 [7]. The authors created a new model based on alanine aminotransferase (ALT), alkaline phosphatase, platelets (PLT) and γ -glutamyltransferase (GGT); AUROC was 0.74 (95% CI: 0.66-0.82) – higher than AUROC for APRI (aspartate transaminase [AST] to platelet ratio index), NAFLD fibrosis score, and FIB-4 (fibrosis-4) index.

This study aimed to assess the clinical usefulness of noninvasive markers of fibrosis in children with NAFLD in daily care.

Material and methods

Patients

Forty-nine children with NAFLD aged 3-16 years (mean 10.51 \pm 3.18 years) who visited the Hepatology Outpatient Clinic at the Department of Infectious Diseases and Child Neurology were consecutively recruited

to this study. The diagnosis of NAFLD was based on the clinical picture, abdominal ultrasound, necessary laboratory tests (elevated liver function tests) and, in individual cases, liver biopsy. In differential diagnosis viral hepatitis (type B, C, Epstein-Barr virus and cytomegalovirus infection), autoimmune hepatitis, α 1-antitrypsin deficiency, hemochromatosis, celiac disease, Wilson disease and alcohol intake were considered. In children younger than 5 years, inborn errors of metabolism were excluded [8]. Diagnosis of liver diseases in the differential diagnosis mentioned above was the study exclusion criterion. Table 1 summarizes the clinical characteristics of the study group.

Nonalcoholic fatty liver (NAFL) was defined as hyperechogenicity of the liver ultrasound and absence of alcohol consumption and concomitant diseases included in the differential diagnosis list.

For further analysis, patients were divided into four groups: HAO (group with elevated serum ALT and obesity), HAL (lean patients with elevated ALT), NAO (patients with ALT below upper limit of normal [ULN] and obesity) and NAL (lean patients without elevation of serum ALT level).

Obesity was defined as a body mass index (BMI) higher than the 97th percentile for the child's age and sex, overweight as BMI between the 90th and 97th percentiles. Weight was measured on an electronic digital

Table 1. Characteristics of the group

Parameter	Number	Percentage	Mean \pm SD	Median	Range
Age (years)	–	–	10.51 \pm 3.18	11	3-16
Gender M/F	35/14	71.4/28.6	–	–	–
Weight (kg)	–	–	68.23 \pm 25.1	62.5	24.5-122.5
Height (cm)	–	–	153.42 \pm 18.27	153	114.5-190
BMI	–	–	28.08 \pm 5.84	27.51	15.43-42.13
BMI z-score	–	–	2.11 \pm 0.98	2.28	–0.89-3.29
BMI > 97 percentile	35	71.43	–	–	–
BMI 90-97 percentile	7	14.29	–	–	–
WHR	–	–	0.94 \pm 0.07	0.93	0.82-1.06
ALT (IU/l)	–	–	61.35 \pm 57.54	42.5	12-304
AST (IU/l)	–	–	46.21 \pm 28.65	35.5	19-149
GGTP (IU/l)	–	–	26.83 \pm 17.96	21	9-75
Total cholesterol (mg/dl)	–	–	166.39 \pm 40.29	162	56-249
LDL (mg/dl)	–	–	99.07 \pm 32.72	101.5	18-151
HDL (mg/dl)	–	–	46.56 \pm 21.99	41	25-148
TG (mg/dl)	–	–	137.85 \pm 62.05	126.5	23-265
HOMA-IR \geq 2 (n = 20)	–	–	4.82 \pm 4.64	3.65	1.4-23.1

BMI – body mass index, WHR – waist to hip ratio, ALT – alanine aminotransferase, AST – aspartate aminotransferase, GGTP – γ -glutamyltranspeptidase, LDL – low-density lipoprotein, HDL – high-density lipoprotein, TG – triglycerides, HOMA-IR – homeostatic model assessment–international ratio

scale, with 150 kg capacity and 10 g accuracy. Height was measured with a 2 m vertical wall stadiometer graduated in millimeters. The measurements were performed following recommended procedures.

Data collection

Anthropometric data of the participants, including weight, height, waist circumference, and hip circumference, were collected. A blood sample following 8-10 hours fasting was obtained to assess complete blood count, fasting blood sugar, insulin level, ALP and ASP (ALT normal value < 12 years old: boys < 40 IU/l, girls < 35 IU/l, > 12 years old: boys < 26 IU/l, girls < 22 IU/l), GGT (normal value 4-24 IU/l) and serum lipid profile.

Complete blood count (CBC), glucose and insulin levels, ALT, AST, GGT and serum lipid levels were determined on a standard laboratory analyzer.

HOMA-IR (homeostatic model assessment) index was calculated using the formula [9]: $HOMA-IR = [insulin (U/ml) * glucose (mg/dl)]/405$. The cutoff point for HOMA-IR was 2 [10, 11].

Noninvasive hepatic fibrosis scores

APRI was calculated as $AST/ULN \text{ platelets (G/L)} * 100$ [12]. The cutoff point indicating fibrosis was 0.5, while a cut-off of 1.5 may indicate severe risk of liver cirrhosis [13]. Formula: $FIB-4 = (age * AST)/[platelet (G/L) * \sqrt{ALT}]$ was used for FIB-4 index calculation [14]. Pediatric NAFLD fibrosis score (PNFS) was calculated using the following formula:

$PNFS (\text{probability distribution}) = [e^z/1 + e^z] * 100$;

$z = 1.1 + (0.34 * \sqrt{ALT}) + (0.002 * \text{alkaline phosphatase}) - (1.1 * \log(\text{platelets G/l})) - (0.02 * GGT)$ [8].

The risk of the presence of fibrosis was estimated by this parameter, and patients were divided into groups with and without the risk of fibrosis (F) – PNFS between 13 and 24 (equivalent to estimated $F \geq 2$) and severe fibrosis (SF) – PNFS higher than 24 (equivalent to F3-F4) [8].

Invasive diagnostics

Biopsy specimens were taken in 4 children with suspicion of the most progressed liver injury in local anesthesia using Menghini needle (Braun). The specimens were evaluated by pathologists blinded for history and clinical data.

Ultrasonography

All examinations were performed by an experienced radiologist on a Toshiba Aplio i700 and Esaote MyLab Twice.

Results were based on four hallmarks of liver steatosis: echogenicity of the liver, visualization of the diaphragm, intrahepatic vessels and posterior part of the right liver lobe.

Statistical analysis

Data were presented as the mean \pm standard deviation (SD). Student's *t* test or the Mann-Whitney test was used to compare continuous variables, where appropriate. Moreover, Pearson's χ^2 test was applied to categorical variables. Fisher's exact test was used in the analysis of contingency tables. The Kruskal-Wallis test was used for testing whether samples originate from the same distribution. Correlations were performed using the Spearman rank correlation test. For all tests, a *p*-value of < 0.05 was considered statistically significant. STATISTICA (version 13, TIBCO Software Inc., Palo Alto, USA) was used for all analyses.

Compliance with ethics guidelines

The study has been approved by Ethics Committee at the University of Medical Sciences in Poznan (no. 1074/17 from November 9.2017). We have adhered to all recommendations of the committee.

Results

In the study group, 35 (71.43%) of patients were obese, and 7 (14.29%) were overweight. There were no statistically significant differences between overweight and lean patients regarding laboratory parameters and fibrosis markers except lower mean corpuscular volume (MCV, *p* = 0.034) found in overweight patients. ALT activity exceeded ULN in both groups of patients. Higher AST was observed in lean patients only. No statistically significant difference was observed regarding these parameters. There were no statistically significant difference in PNFS, APRI and FIB-4 between lean and overweight patients (Table 2).

Furthermore, the group of patients with a risk of fibrosis assessed according to the PNFS index was compared to the group without such a risk (F vs. no F). Significantly higher aminotransferase and GGT activity were observed in the group with fibrosis (*n* = 11), while triglyceride (TG) levels were lower in this group compared to children without risk of fibrosis. Moreover, APRI and FIB-4 values were also significantly higher in the group with a risk of fibrosis (Table 3). Mean APRI value exceeds the cut-off of 0.5. Similar observations in groups with and without risk of fibrosis (PNFS ≥ 24 , *n* = 4) were made regarding aminotransferase and GGT activity (Table 4). However, only

Table 2. Comparison of laboratory parameters and noninvasive hepatic fibrosis scores in overweight and lean patient groups

Parameter	Overweight <i>n</i> = 42	Lean <i>n</i> = 7	<i>p</i>
Age (years)	10.4 ±3.07	11.14 ±3.53	0.59
Gender M/F	30/12	5/2	
Weight (kg)	71.77 ±24.17	48.04 ±21.77	0.024
Height (cm)	153.41 ±18.13	153.43 ±20.57	0.98
BMI (kg/m ²)	29.61 ±4.62	19.36 ±4.35	0.0002
BMI z-score	2.41 ±0.65	0.41 ±0.86	0.000031
HGB (g/dl)	13.75 ±1.22	13.75 ±1.63	1
HCT (%)	39.7 ±3.12	39.9 ±4.17	0.91
MCV (fl)	80.62 ±4.53	85.6 ±8.47	0.034
WBC (G/l)	7.73 ±1.9	6.82 ±1.47	0.26
PLT (G/l)	294.59 ±66.63	268.71 ±107.15	0.28
ALT (U/l)	61.05 ±48.35	74.71 ±101.71	0.88
AST (U/l)	44.18 ±24.32	63.71 ±45.77	0.28
Total cholesterol (mg/dl)	170.47 ±35.57	148.2 ±101.71	0.97
LDL (mg/dl)	102.56 ±27.92	81.6 ±51.27	0.54
HDL (mg/dl)	46.63 ±23.77	46.2 ±11.65	0.53
TG (mg/dl)	147 ±54.32	101 ±96.3	0.92
APRI	0.39 ±0.21	0.67 ±0.49	0.26
FIB-4	0.21 ±0.08	0.45 ±0.42	0.31
PNFS	9.22 ±9.29	11.73 ±16.52	0.72

BMI – body mass index, HGB – hemoglobin, HCT – hematocrit, MCV – mean corpuscular volume, WBC – white blood cells, PLT – platelets, ALT – alanine aminotransferase, AST – aspartate aminotransferase, LDL – low-density lipoprotein, HDL – high-density lipoprotein, TG – triglycerides, APRI – AST to platelet ratio, FIB-4 – fibrosis-4 index, PNFS – pediatric NAFLD fibrosis score

APRI differed significantly between these groups, with a mean value exceeding 1.

In the next step, the four previously described groups were compared (HAL, HAO, NAL, NAO). Higher values of APRI were found in the group of lean patients (with and without aminotransferase elevations respectively) compared to the corresponding groups of obese patients. The highest values of APRI were found in a group of lean patients with elevated aminotransferase activity. In obese patients with aminotransferase activity within reference values, the APRI index was below the cutoff for significant fibrosis (Fig. 1).

Patients with elevated aminotransferases (HAO, HAL) developed higher BMI compared to the appropriate groups with the liver function tests within reference limits. In the lean group with increased aminotransferase activity, a higher BMI was found. Between HAO and NAO, the mean BMI values were similar. The group of the leanest patients comprised those with normal aminotransferase activity (Fig. 2).

Significantly higher mean PNFS index values were found in the HAO group compared to other groups. Among the other groups, the mean values of the PNFS

index were similar. In these groups, the threshold for fibrosis (F) was not exceeded (Fig. 3).

A strong significant correlation between APRI and PNFS indexes was found ($r = 0.88$, $p < 0.05$) (Fig. 4).

Liver biopsy was performed in 4 patients with the highest risk of fibrosis estimated by chosen indexes. In all cases, more than 5% macrovesicular steatosis was found (30-50%). In 1 case with suspected NASH, periportal and pericellular fibrosis, lobular inflammation, and hepatocyte ballooning were observed by a pathologist. Fibrosis was found in all assessed cases.

Discussion

Nonalcoholic fatty liver disease, the most common liver chronic disease, constitutes a growing health problem in developed countries. Despite this, there are few reports about the prevalence of NAFLD in children. Boys were the majority in the described population (71.4%), which is consistent with the previously published observations [2]. These data are also in compliance with the higher prevalence of obesity among boys observed in Poland [15].

Table 3. Comparison of clinical and laboratory parameters and noninvasive hepatic fibrosis scores in groups of patients with and without a risk of fibrosis (PNFS ≥ 13)

Parameter	Group with fibrosis <i>n</i> = 11	No fibrosis <i>n</i> = 38	<i>p</i>
Age (years)	9.56 \pm 2.65	10.68 \pm 3.03	0.25
Gender M/F	8/3	27/11	0.62
Weight (kg)	66.39 \pm 25.38	68.22 \pm 26.24	0.87
Height (cm)	146.83 \pm 14.98	154 \pm 16.69	0.21
BMI (kg/m ²)	29.82 \pm 6.23	27.76 \pm 6.2	0.2
BMI z-score	2.29 \pm 0.82	2.05 \pm 1.22	0.12
HGB (g/dl)	13.43 \pm 1.03	13.76 \pm 1.27	0.49
HCT (%)	39.44 \pm 3.11	39.63 \pm 3.2	0.75
MCV (fl)	82.41 \pm 5.52	80.61 \pm 4.47	0.36
WBC (G/l)	7.54 \pm 1.75	7.67 \pm 1.83	0.9
PLT (G/l)	297.78 \pm 44.21	289.65 \pm 80.34	0.52
ALT (U/l)	146.22 \pm 71.87	40.84 \pm 30.82	0.00002
AST (U/l)	81.89 \pm 33.35	36.61 \pm 19.35	0.0001
ALP (U/l)	340.33 \pm 106.14	249.58 \pm 87.3	0.013
GGTP (U/l)	39.44 \pm 20.79	23.16 \pm 15.58	0.004
Total cholesterol (mg/dl)	184.5 \pm 20.22	165.76 \pm 43.99	0.17
LDL (mg/dl)	118 \pm 13.14	95.7 \pm 34.78	0.23
HDL (mg/dl)	49.25 \pm 6.55	46.27 \pm 25.05	0.15
TG (mg/dl)	104 \pm 20.22	153.82 \pm 66.5	0.049
HOMA-IR	2.27 \pm 0.76	5.59 \pm 5.33	0.068
APRI	0.7 \pm 0.3	0.34 \pm 0.2	0.0003
FIB-4	31.13 \pm 16.85	9.84 \pm 8.85	0.0002
PNFS	24.37 \pm 13.77	5.32 \pm 2.57	0.000007

BMI – body mass index, HGB – hemoglobin, HCT – hematocrit, MCV – mean corpuscular volume, WBC – white blood cells, PLT – platelets, ALT – alanine aminotransferase, AST – aspartate aminotransferase, LDL – low-density lipoprotein, HDL – high-density lipoprotein, TG – triglycerides, APRI – AST to platelet ratio, FIB-4 – fibrosis-4 index, PNFS – pediatric NAFLD fibrosis score

On the authority of expert opinion published in 2007 (Barlow SE), overweight children over ten years of age with additional risk factors should be screened for liver disease (serum ALT and AST) twice a year [16]. These guidelines match the trends observed in our group – patients with elevated serum ALT and AST had higher BMI than patients with liver function tests within reference values.

Significantly lower MCV values in overweight patients in the study group were documented. These observations are compatible with the Vuong report [16]. They compared complete blood count (CBC) parameters in patients with and without an increased waist circumference. According to those data, waist circumference (WC) was positively correlated with red blood cells (RBC), hemoglobin and hematocrit and negatively correlated with MCV, mean corpuscular hemoglobin (MCH), and mean corpuscular hemoglobin concentration (MCHC). Lower iron levels in a group with

higher WC were detected. Iron deficiency in obese patients may be caused by increased production of hepcidin. The concentration of this protein grows during inflammation, leading to impairment of iron transport from enterocytes to the portal circulation [17]. Similar observations in a group of patients undergoing bariatric surgery were made – a high percentage of anemia was noted [18].

Initially, three hepatic fibrosis scores were taken into consideration in our study – APRI, FIB-4, and PNFS. We found that FIB-4 was adequate only in a group with a risk of fibrosis, not significant fibrosis in the study group. Therefore this marker was not found reliable in children with NAFLD. In the study by American authors from 2015 [5], of the four analyzed indicators (AST/ALT ratio, APRI, NAFLD fibrosis score and FIB-4) only APRI had good predictive value for diagnosing any fibrosis in children but not to distinguish a population with significant or advanced fi-

Table 4. Comparison of clinical and laboratory parameters and noninvasive hepatic fibrosis scores in groups of patients with and without a significant risk of fibrosis (PNFS > 23)

Parameter	Group with SF n = 4	No SF n = 45	p
Age (years)	8.33 ± 1.54	10.59 ± 3	0.13
Gender M/F	2/2	33/12	0.32
Weight (kg)	51.67 ± 22.19	69.12 ± 25.82	0.30
Height (cm)	139.33 ± 6.66	153.51 ± 18.25	0.14
BMI (kg/m ²)	25.99 ± 9.2	28.41 ± 6.12	0.92
BMI z-score	1.77 ± 1.29	2.13 ± 1.05	0.89
HGB (g/dl)	13.43 ± 0.92	13.70 ± 1.24	0.82
HCT (%)	38.83 ± 3.08	39.65 ± 3.18	0.58
MCV (fl)	83.63 ± 3.33	80.82 ± 4.8	0.34
WBC (G/l)	7.46 ± 1.28	7.65 ± 1.84	0.93
PLT (G/l)	289.67 ± 56.32	291.62 ± 75.16	0.96
ALT (U/l)	227.33 ± 68.79	51.35 ± 37.96	0.0047
AST (U/l)	118.67 ± 26.5	40.97 ± 21.26	0.0065
ALP (U/l)	403 ± 148.2	259.22 ± 87.39	0.057
GGTP (U/l)	54.3 ± 26.27	24.6 ± 15.6	0.031
Total cholesterol (mg/dl)	175.33 ± 26.63	168.37 ± 42.65	0.68
LDL (mg/dl)	115 ± 21.21	97.72 ± 34	0.55
HDL (mg/dl)	51 ± 5.66	46.37 ± 24.01	0.21
TG (mg/dl)	92 ± 16.1	150.16 ± 64.27	0.78
HOMA-IR	1.40 ± 0	5.23 ± 5.06	1
APRI	1.04 ± 0.2	0.37 ± 0.2	0.0055
FIB-4	0.23 ± 0.06	0.23 ± 0.13	0.72
PNFS	42.3 ± 4.6	9.95 ± 4.52	0.0047

BMI – body mass index, HGB – hemoglobin, HCT – hematocrit, MCV – mean corpuscular volume, WBC – white blood cells, PLT – platelets, ALT – alanine aminotransferase, AST – aspartate aminotransferase, SF – significant fibrosis, LDL – low-density lipoprotein, HDL – high-density lipoprotein, TG – triglycerides, APRI – AST to platelet ratio, FIB-4 – fibrosis-4 index, PNFS – pediatric NAFLD fibrosis score

bro sis. The authors found the uselessness of indicators intended for adults in the pediatric population and the necessity to create a new one.

In 2014 the first study regarding PNFS was published. The authors gave a first and probable explanation of the high utility of PNFS to distinguish a group of children with significant fibrosis: low PLT associated with more advanced liver damage, ALT as a marker of necroinflammatory activity and the unexplained background of changes in alkaline phosphatase and GGT activity [7]. Due to the probable effectiveness of PNFS in the diagnosis of fibrosis and significant fibrosis, we chose this indicator for further analysis. Indirect confirmation of this observation may be significantly higher PNFS values in the group of obese patients with elevated aminotransferase activity.

The utility of APRI for distinguishing groups with and without risk of fibrosis was found, and the values

were significantly higher in children with severe fibrosis, which coincides with previous reports. In the latter group of patients the mean APRI value exceeded 1.0. Although a value of 1.5 (cut-off for cirrhosis) has not been crossed the risk of significant fibrosis was confirmed by histopathological evaluation. An interesting fact is that lean patients with elevated aminotransferase activity had higher values of APRI than overweight ones. Similar alterations in groups without elevated activity of liver function tests were noted.

There is a need for further studies comparing the effectiveness of non-invasive markers of fibrosis in children NAFLD. In our analysis, a high correlation between APRI and PNFS was found ($r = 0.88$). It seems reasonable to create a commonly available, low-cost tool to identify a group of children requiring a more accurate and, consequently, more expensive diagnosis of liver disease. An example of such a tool is an

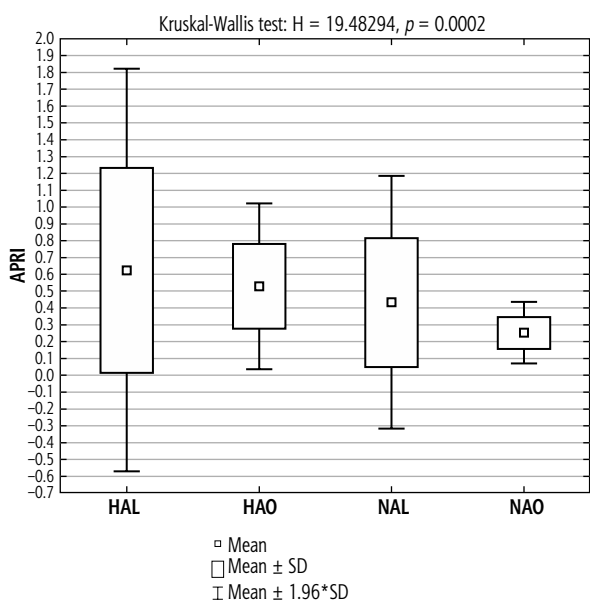


Fig. 1. Mean APRI index in the groups of lean and obese patients with and without elevated aminotransferase activity

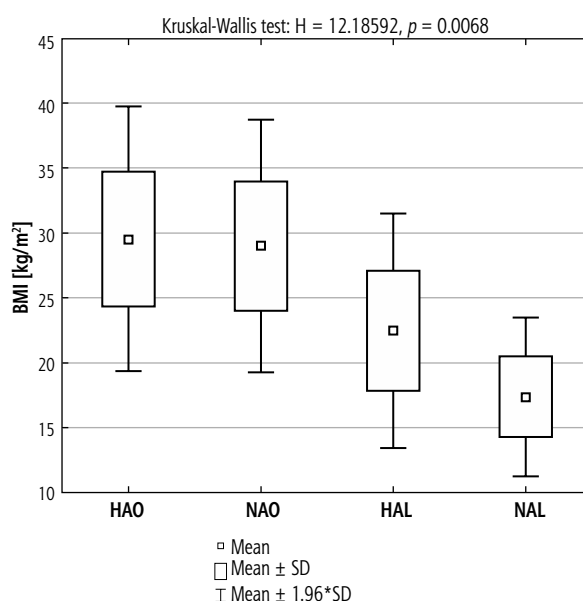


Fig. 2. Mean BMI in the groups of lean and obese patients with and without elevated aminotransferase activity

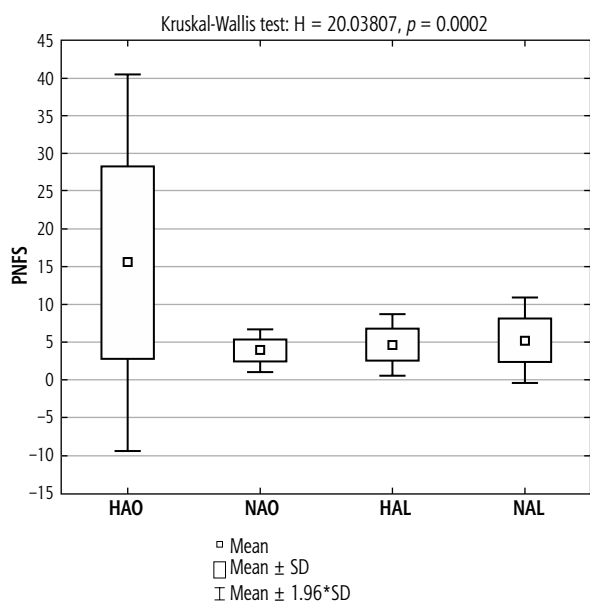


Fig. 3. Mean PNFS index in the groups of lean and obese patients with and without elevated aminotransferase activity

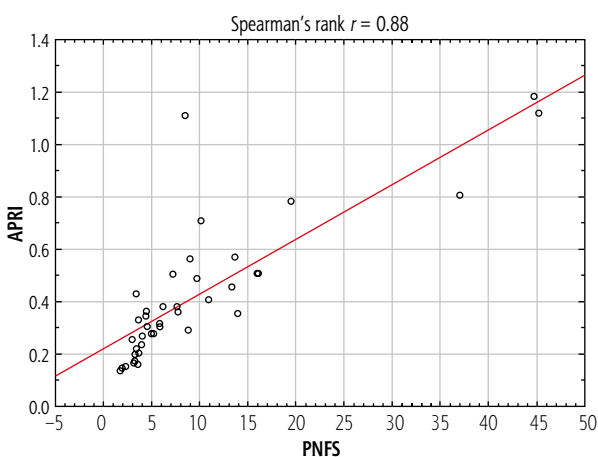


Fig. 4. Spearman rank correlation between APRI and PNFS in the study group

algorithm created for adult patients with NAFLD and suspected risk of advanced fibrosis published this year [19]. In patients with abnormal liver function tests, evidence of fat accumulation in diagnostic imaging, risk factors (e.g., obesity) and excluded secondary causes of steatosis and significant alcohol consumption, the next step is to rule out advanced fibrosis by the NAFLD fibrosis score or FIB-4 index. According to the risk calculated by indicators, these patients will carry out an appropriate further diagnostic path – e.g., pa-

tients with low risk will repeat PNFS/FIB-4 every two years and patients with intermediate risk will undergo more sophisticated examinations such as the ELF test. When attempting to create a similar tool for children, APRI and PNFS scores can be considered as one of the steps. In our study children with a risk of significant fibrosis developed higher ALT, AST and GGT activity. APRI was also higher in this group of patients. Therefore, these patients required a further, more detailed diagnostic approach including a liver biopsy.

We are aware that our research has limitations, mainly concerning the small population from one outpatient clinic in a tertiary hospital. All children were from one ethnic group, and only 4 underwent liver

biopsy, all of them having noninvasive markers of liver fibrosis.

However, our observations may be continued on a larger group of children with NAFLD.

Conclusions

Apart from aminotransferase activity, complete blood count should be assessed, looking for lower MCV caused by iron deficiency. In contrast to FIB-4 (fibrosis score), PNFS and APRI proved to be more accurate in our group. PNFS seems to be appropriate to evaluate fibrosis in a noninvasive diagnostic algorithm.

Disclosure

The authors declare no conflict of interest.

References

- Schwimmer JB, Deutsch R, Kahen T, et al. Prevalence of fatty liver in children and adolescents. *Pediatrics* 2006; 118: 1388-1393.
- Anderson EL, Howe LD, Jones HE, et al. The prevalence of non-alcoholic fatty liver disease in children and adolescents: a systematic review and meta-analysis. *PLoS One* 2015; 10: e0140908.
- Chalasanani N, Younossi Z, Lavine JE, et al. The diagnosis and management of nonalcoholic fatty liver disease: practice guidance from the American Association for the Study of Liver Diseases. *Hepatology* 2018; 67: 328-357.
- Angulo P, Kleiner DE, Dam-Larsen S, et al. Liver fibrosis, but no other histologic features, is associated with long-term outcomes of patients with nonalcoholic fatty liver disease. *Gastroenterology* 2015; 149: 389-397.
- Mansoor S, Yerian L, Kohli R, et al. The evaluation of hepatic fibrosis scores in children with nonalcoholic fatty liver disease. *Dig Dis Sci* 2015; 60: 1440-1447.
- Valerio N, Anna A, Andrea V, et al. The pediatric NAFLD fibrosis index: a predictor of liver fibrosis in children with non-alcoholic fatty liver disease. *BMC Med* 2009; 7: 21.
- Alkhouiri N, Mansoor S, Giammaria P, et al. The development of pediatric NAFLD fibrosis score (PNFS) to predict the presence of advanced fibrosis in children with nonalcoholic fatty liver disease. *PLoS One* 2014; 9: e104558.
- Vajro P, Lenta S, Socha P, et al. Diagnosis of nonalcoholic fatty liver disease in children and adolescents: position paper of the ESPGHAN Hepatology Committee. *J Pediatr Gastroenterol Nutr* 2012; 54: 700-713.
- Matthews DR, Hosker JP, Rudenski AS, et al. Homeostasis model assessment: insulin resistance and beta-cell function from fasting plasma glucose and insulin concentrations in man. *Diabetologia* 1985; 28: 412-419.
- Salgado AL, Carvalho L, Oliveira AC, et al. Insulin resistance index (HOMA-IR) in the differentiation of patients with non-alcoholic fatty liver disease and healthy individuals. *Arq Gastroenterol* 2010; 47: 165-169.
- Singh Y, Garg MK, Trandon N, et al. A study of insulin resistance by HOMA-IR and its cut-off value to identify metabolic syndrome in urban Indian adolescents. *J Clin Res Pediatr Endocrinol* 2013; 5: 245-251.
- Wai CT, Greenon JK, Fontana RJ, et al. A simple noninvasive index can predict both significant fibrosis and cirrhosis in patients with chronic hepatitis C. *Hepatology* 2003; 38: 518-526.
- Chou R, Wasson N. Blood tests to diagnose fibrosis or cirrhosis in patients with chronic hepatitis C virus infection: a systematic review. *Ann Intern Med* 2013; 158: 807-820.
- Sterling RK, Lissen E, Clumeck N, et al. Development of a simple noninvasive index to predict significant fibrosis in patients with HIV/HCV coinfection. *Hepatology* 2006; 43: 1317-1325.
- Kulaga Z, Grajda A, Gurzowska B, et al. The prevalence of overweight and obesity among polish school-aged children and adolescents. *Przegl Epidemiol* 2016; 70: 641-651.
- Vuong J, Qiu Y, La M, et al. Reference intervals of complete blood count constituents are highly correlated to waist circumference: Should obese patients have their own "normal values?". *Am J Hematol* 2014; 89: 671-677.
- Tussing-Humphreys L, Pusatcioglu C, Nemeth E, et al. Rethinking iron regulation and assessment in iron deficiency, anemia of chronic disease, and obesity: introducing hepcidin. *J Acad Nutr Diet* 2012; 112: 391-400.
- Khanbhai M, Dubb S, Patel K, et al. The prevalence of iron deficiency anaemia in patients undergoing bariatric surgery. *Obes Res Clin Pract* 2013; 9: 45-49.
- Vilar-Gomez E, Chalasanani N. Non-invasive assessment of non-alcoholic fatty liver disease: Clinical prediction rules and blood-based biomarkers. *J Hepatol* 2018; 68: 305-315.