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Review article

Immune response to foreign materials in spinal fusion surgery

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ABSTRACT

Spinal fusion surgery is a common procedure used to stabilize the spine and treat back pain. The procedure involves the use of foreign materials such as screws, rods, or cages, which can trigger a foreign body reaction, an immune response that involves the activation of immune cells such as macrophages and lymphocytes. The foreign body reaction can impact the success of spinal fusion, as it can interfere with bone growth and fusion. This review article provides an overview of the cellular and molecular events in the foreign body reaction, the impact of the immune response on spinal fusion, and strategies to minimize its impact. By carefully considering the use of foreign materials and optimizing surgical techniques, the impact of the foreign body reaction can be reduced, leading to better outcomes for patients.

1. Introduction

1.1. Brief explanation of spinal fusion and the use of foreign materials in the procedure

Overview of the foreign body reaction and its impact on spinal fusion.

Spinal fusion is a surgical procedure that involves joining two or more vertebrae in the spine to treat conditions such as degenerative disc disease [1], spinal stenosis [2], or spondylolisthesis. The procedure is intended to stabilize the spine and relieve pain by eliminating motion between the fused vertebrae [3]. Foreign materials such as metal implants [4], bone grafts [5], or synthetic materials [6] are often used in spinal fusion procedures to promote bone growth and fusion between the vertebrae [7]. These materials are intended to provide stability and support to the spine as it heals [8]. However, the use of foreign materials in spinal fusion procedures can trigger a foreign body reaction [9], which is an immune system response to the presence of foreign materials in the body [10]. The foreign body reaction can cause inflammation, tissue damage, and impair the healing process of the spine [11]. The impact of the foreign body reaction on spinal fusion can lead to complications such as implant failure [12], non-union (lack of bone growth and fusion) [13], and chronic pain [14]. Therefore, it is important for medical professionals to carefully evaluate the use of foreign materials in spinal fusion procedures and consider alternative approaches to promote successful fusion while minimizing the risk of a foreign body reaction. Despite the great success of artificial implants for the human body, they are not without their limitations. One major issue is the foreign body reaction, which is an immune reaction of the organism to the implant. However, researchers have found a way to avoid or decrease the foreign body reaction by modifying the surface of the artificial implant with condensed aromatic structures containing free radicals. This modification allows for covalent attachment of host proteins in their native conformation,

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resulting in total protein coverage that prevents direct contact of immune cells with the implant surface. As a result, the immune response of the organism is not generated, and the artificial implant is not isolated from the tissue, resulting in low activity of macrophages, low cell proliferation, and low inflammatory activity. This promising approach could help to address some of the major health problems associated with artificial implants [15].

The specific types of foreign materials used in spinal fusion procedures and how they trigger the foreign body reaction.

Recent progress in the field of spinal fusion has highlighted the importance of understanding the specific types of foreign materials used in these procedures and how they trigger the foreign body reaction [16]. One technique that has gained traction in recent years is the use of bioactive materials, which can enhance the biological response and promote faster and more complete fusion [17]. However, these materials can also trigger a more severe foreign body reaction compared to traditional materials such as titanium and stainless steel [18]. Bioactive materials, such as ceramics and calcium phosphate-based materials, can release ions that stimulate immune cells and trigger the foreign body reaction [19]. Additionally, some bioactive materials can induce fibrosis, which can further contribute to implant failure [20]. Therefore, careful consideration of the specific types of foreign materials used in spinal fusion procedures is necessary to minimize the risk of a foreign body reaction and ensure successful fusion [21]. In addition to bioactive materials, other foreign materials used in spinal fusion procedures can also trigger the foreign body reaction [22]. Metallic materials can corrode and release metal ions, which can cause an inflammatory response and lead to implant failure [23]. Synthetic materials, such as polymers, can also trigger the foreign body reaction due to their chemical and mechanical properties [16]. Therefore, it is crucial to carefully select the appropriate foreign materials for spinal fusion procedures and consider their potential impact on the foreign body reaction [24]. Researchers are exploring different strategies to minimize the foreign body reaction, such as surface modifications and coatings that can reduce immune cell activation and fibrous capsule formation [25]. Further research is needed to identify the most effective approaches to minimize the foreign body reaction and improve the long-term success of spinal fusion procedures [24]. In recent years, there has been growing interest in developing biomaterials that can better integrate with the host tissue and minimize the foreign body reaction [26]. One promising approach is the use of biodegradable materials that can gradually degrade and be replaced by the host tissue [27], minimizing the formation of a fibrous capsule and reducing the immune response [28]. Natural materials, such as collagen and chitosan, have also been explored for their ability to mimic the extracellular matrix of the host tissue and promote cellular attachment and proliferation [29]. Additionally, researchers are investigating the use of stem cells and growth factors to enhance the biological response and promote faster and more complete fusion [30]. By carefully selecting and designing foreign materials for spinal fusion procedures, it may be possible to minimize the foreign body reaction and improve the long-term success of these procedures [24]. One major concern with spinal fusion procedures is the immune response of the patient to the foreign materials used in the procedure [31]. The immune response can lead to the formation of a fibrous capsule, which can compromise the stability of the implant and interfere with the fusion process [22]. In addition, the immune response can cause chronic pain and other complications that can reduce the quality of life of the patient [32]. To address this concern, researchers are exploring different strategies to modulate the immune response and minimize the foreign body reaction [24]. One approach is the use of immunosuppressive drugs, which can reduce the activity of immune cells and minimize the formation of a fibrous capsule [33]. However, this approach is not without its risks, as it can increase the risk of infection and other complications [34]. Therefore, researchers are also exploring the use of immune-modulating biomaterials that can interact with the immune system in a more controlled and targeted manner [35]. By carefully balancing the immune response to the foreign materials used in spinal fusion procedures, it may be possible to improve the long-term success of these procedures and reduce the risk of complications [36]. Table 1 provides a summary of examples of foreign materials used in spinal fusion procedures and their potential impact on the foreign body reaction. By carefully selecting and designing foreign materials for spinal fusion procedures [37], it may be possible to minimize the foreign body reaction and improve the long-term success of these procedures (see Table 1).

2. Cellular and molecular events in the foreign body reaction

The foreign body reaction is a complex process that involves the interaction between the implant and the host tissues, leading to a series of cellular and molecular events [38]. Upon implantation, the body initiates an inflammatory response to the presence of the foreign material [39], which is characterized by the recruitment of immune cells such as macrophages and neutrophils to the site of implantation [39]. These immune cells phagocytose and degrade the implant material, releasing cytokines and growth factors that can stimulate the proliferation and differentiation of fibroblasts [40]. These fibroblasts then produce extracellular matrix components such as collagen [41], which can lead to the formation of a fibrous capsule around the implant [41]. The fibrous capsule acts as a barrier

Table 1

Examples of foreign materials used in spinal fusion procedures and their potential impact on the foreign body reaction.

Material	Potential impact on foreign body reaction	
Titanium [28,136–138]	Low immune response and fibrous capsule formation	
Stainless steel [28,138,139]	Low immune response and fibrous capsule formation	
Cobalt-chromium [23,140,141]	Moderate immune response and potential for metal ion release	
Bioactive ceramics [142–144]	High immune response and fibrosis	
Calcium phosphate [144–146]	High immune response and potential for fibrosis	
Biodegradable polymers [25,147,148]	Gradual degradation and minimal fibrous capsule formation	
Collagen [149–151]	Mimics host tissue and promotes cellular attachment	
Chitosan [152–154]	Mimics host tissue and promotes cellular attachment	

between the implant and the surrounding tissues [42] and can inhibit the exchange of nutrients and cells between the implant and the surrounding tissues [43], leading to implant failure. In addition to the recruitment of immune cells and the formation of a fibrous capsule, the foreign body reaction is also characterized by the activation of signaling pathways that regulate the immune response and inflammation [44]. These pathways can modulate the magnitude and duration of the foreign body reaction, as well as the formation of the fibrous capsule around the implant [45].

3. Cellular and molecular events in the foreign body reaction

The foreign body reaction is a complex process that involves the interaction between the implant and the host tissues [38], leading to a series of cellular and molecular events. Upon implantation, the body initiates an inflammatory response to the presence of the foreign material [46], which is characterized by the recruitment of immune cells such as macrophages and neutrophils to the site of implantation [47]. These immune cells phagocytose and degrade the implant material, releasing cytokines and growth factors that can stimulate the proliferation and differentiation of fibroblasts [48]. These fibroblasts then produce extracellular matrix components such as collagen [49], which can lead to the formation of a fibrous capsule around the implant [50]. The fibrous capsule acts as a barrier between the implantand the surrounding tissues [51] and can inhibit the exchange of nutrients and cells between the implant and the surrounding tissues [52], leading to implant failure [53]. In addition to the recruitment of immune cells and the formation of a fibrous capsule [54], the foreign body reaction is also characterized by the activation of signaling pathways that regulate the immune response and inflammation [55]. These pathways can modulate the magnitude and duration of the foreign body reaction [56], as well as the formation of the fibrous capsule around the implant [57].

3.1. Inflammatory responses, tissue damage, and bone resorption during the foreign body reaction

The fibrous capsule that forms around the implant in response to the foreign body reaction can have different fates [58], depending on various factors such as the size, shape, and composition of the implant [59], as well as the patient's immune system. In some cases, the fibrous capsule can lead to successful spinal fusion, as it helps to stabilize the implant and promotes the fusion process [60]. However, in other cases, the fibrous capsule can inhibit the fusion process by acting as a barrier between the implant and the surrounding tissues [61], preventing the exchange of nutrients and cells needed for fusion [62]. In addition, the fibrous capsule can also lead to implant failure by causing stress shielding, which is characterized by the transfer of stress from the vertebral bodies to the implant [63], leading to implant failure [64]. The fibrous capsule can also lead to other complications such as pain, stiffness, and implant loosening [65], which can result in the need for revision surgery [66]. Therefore, it is important to understand the fate of the fibrous capsule around the implant in order to optimize the outcome of spinal fusion and minimize the risk of complications [67].

4. Impact of the foreign body reaction on spinal fusion

The foreign body reaction can have a significant impact on the success of spinal fusion [68]. The formation of a fibrous capsule around the implant in response to the foreign body reaction can lead to successful spinal fusion in some cases [69], as it helps to stabilize the implant and promotes the fusion process [70]. However, in other cases, the fibrous capsule can inhibit the fusion process by acting as a barrier between the implant and the surrounding tissues [61], preventing the exchange of nutrients and cells needed for fusion. The foreign body reaction is a complex process that involves various signaling pathways and cellular mechanisms [11]. When an implant is placed in the body, it is recognized as a foreign object by the immune system [46], which triggers a series of events aimed at removing the foreign material [71]. This process is initiated by the activation of innate immune cells, such as macrophages and neutrophils [72], which engulf the implant and release pro-inflammatory cytokines and chemokines to recruit more immune cells to the site of injury [73]. The activation of immune cells is mediated by various signaling pathways, such as the toll-like receptor (TLR) pathway [74], which recognizes pathogen-associated molecular patterns (PAMPs) [75] and damage-associated molecular patterns (DAMPs) on the surface of the implant [76]. This leads to the activation of transcription factors, such as nuclear factor kappa B (NF-κB) [77], which induce the expression of pro-inflammatory cytokines, such as interleukin-1 (IL-1), interleukin-6 (IL-6), and tumor necrosis factor alpha (TNF- α) [78]. The pro-inflammatory cytokines attract more immune cells to the site of injury [79], which further amplify the immune response [80]. In addition to the activation of innate immune cells, the foreign body reaction also involves the activation of adaptive immune cells [81], such as T cells and B cells [82]. These cells recognize specific antigens on the surface of the implant and mount an antigen-specific immune response [83]. The foreign body reaction can also affect the mechanical stability of the implant by inducing the formation of a fibrous capsule around the implant [58]. The formation of the fibrous capsule is mediated by various signaling pathways, such as the transforming growth factor beta (TGF- β) pathway [84], which promotes the differentiation of fibroblasts into myofibroblasts and the deposition of extracellular matrix proteins [85]. Stress shielding is another complication that can result from the foreign body reaction [86]. This occurs when the implant absorbs a significant portion of the load that would normally be borne by the surrounding bone [87], leading to bone resorption and implant loosening [88]. The mechanisms underlying stress shielding are complex and involve various signaling pathways, such as the RANK/RANKL/OPG pathway [89], which regulates osteoclast differentiation and bone resorption [90]. In summary, the foreign body reaction is a complex process that involves various signaling pathways and cellular mechanisms. The impact of the foreign body reaction on spinal fusion can vary depending on various factors, such as the size, shape, and composition of the implant, as well as the patient's immune system. A better understanding of the mechanisms underlying the foreign body reaction is essential for optimizing the outcome of the procedure and minimizing the risk of complications. One important subcluster of cells involved in the foreign body reaction are macrophages [91]. Macrophages play a

crucial role in the initial recognition and removal of the implant by engulfing the foreign material and releasing pro-inflammatory cytokines and chemokines to recruit more immune cells to the site of injury [92]. Macrophages can also differentiate into different subtypes depending on the microenvironment and signals they receive [93]. In the context of the foreign body reaction, macrophages can differentiate into two main subtypes: M1 and M2 macrophages. M1 macrophages are classically activated and secrete pro-inflammatory cytokines such as IL-1, IL-6, and TNF- α [94], which amplify the immune response and promote the clearance of the foreign material [95]. M1 macrophages are important in the early stages of the foreign body reaction. In contrast, M2 macrophages are alternatively activated and secrete anti-inflammatory cytokines such as IL-10 [96], which dampen the immune response and promote tissue repair and remodeling. M2 macrophages are important in the later stages of the foreign body reaction when tissue repair and remodeling are necessary to restore normal tissue function [97]. The transition from M1 to M2 macrophages is regulated by various signaling pathways, such as the TGF- β pathway [98]. The balance between M1 and M2 macrophages is important for the resolution of the foreign body reaction and the restoration of tissue function [99]. Imbalance between these subtypes can lead to chronic inflammation, fibrosis, and implant failure. Therefore, strategies that modulate the polarization of macrophages towards the M2 phenotype may be beneficial for minimizing the foreign body reaction and promoting tissue repair and regeneration. T cells also play an important role in the foreign body reaction. T cells are a type of adaptive immune cell that recognizes specific antigens on the surface of the implant [100] and mount an antigen-specific immune response [101]. This response is mediated by various signaling pathways, such as the T cell receptor (TCR) pathway, which leads to the activation of transcription factors and the production of cytokines [102]. In addition to recognizing antigens on the surface of the implant, T cells can also interact with other immune cells, such as macrophages and dendritic cells, and modulate their function [103]. For example, T cells can secrete cytokines such as interferon-gamma (IFN- γ), which activates macrophages and enhances their phagocytic activity [104]. T cells can also secrete cytokines such as IL-4 and IL-13, which promote the differentiation of macrophages towards the M2 phenotype [105]. The balance between different subtypes of T cells is also important in the foreign body reaction. For example, regulatory T cells (Tregs) are a subset of T cells that suppress immune responses and promote tissue repair and regeneration [106]. Tregs can secrete cytokines such as IL-10 and transforming growth factor beta (TGF- β), which dampen the immune response and promote tissue repair [107]. The balance between Tregs and other T cell subsets, such as Th1 and Th17 cells, is important in maintaining immune homeostasis and preventing chronic inflammation and tissue damage [108]. In addition to T cells, other immune cells such as B cells, neutrophils, and mast cells are also involved in the foreign body reaction. B cells can produce antibodies against the implant [100], which can contribute to the formation of the fibrous capsule [108]. Neutrophils can release reactive oxygen species and proteases [109], which can damage the surrounding tissue and contribute to chronic inflammation [110]. Mast cells can release histamine and other pro-inflammatory mediators [111], which can amplify the immune response and contribute to tissue damage [112]. In summary, T cells and other immune cells play a crucial role in the foreign body reaction. The balance between different subtypes of immune cells is important in maintaining immune homeostasis and promoting tissue repair and regeneration. Strategies that modulate the immune response and promote tissue repair may be beneficial for minimizing the foreign body reaction and improving the outcome of implant surgery.

Overview of Specific Types of Foreign Materials Used in Spinal Fusion Procedures.

Spinal fusion procedures involve the use of foreign materials to promote bone growth and stability [31]. The types of foreign materials used in spinal fusion procedures can vary depending on the specific procedure and the patient's individual needs [113]. Metallic materials such as titanium, stainless steel, and cobalt-chromium alloys have traditionally been used due to their strength and biocompatibility [114]. However, they can still trigger the foreign body reaction and may require additional coatings or modifications to minimize this response [115]. Bioactive materials, such as ceramics and calcium phosphate-based materials, have been developed to enhance bone growth and fusion [116]. However, these materials can trigger a more severe immune response compared to traditional materials. Biodegradable materials, such as polymers, are also being explored as an alternative to permanent foreign materials [117]. They can gradually degrade over time and be replaced by host tissue, minimizing the foreign body reaction [26]. By understanding the types of foreign materials used in spinal fusion procedures and their potential impact on the foreign body reaction [24], researchers and medical professionals can make informed decisions regarding material selection and optimize the outcome of these procedures [118]. Table 2 provides a summary of the different types of foreign materials for spinal fusion procedures, including their potential benefits and drawbacks. By carefully selecting and designing foreign materials for spinal fusion procedures, it may be possible to

Table 2

Types of foreign materials used in spinal fusion procedures.

Material	Description	Potential benefits	Potential drawbacks
Titanium [155–157]	A biocompatible metal commonly used in spinal fusion procedures	Strong and lightweight material	May still trigger the foreign body reaction
Stainless steel [158–160]	A durable metal commonly used in spinal fusion procedures	Biocompatible and strong material	May still trigger the foreign body reaction
Cobalt-chromium alloys [157,161,162]	A metal alloy commonly used in spinal fusion procedures	Biocompatible and strong material, less likely to corrode	May still trigger the foreign body reaction, potential for metal ion release
Bioactive ceramics [163, 164]	Ceramic materials designed to enhance bone growth and fusion	Can enhance bone growth and fusion	May trigger a more severe immune response compared to traditional materials
Calcium phosphate [116, 165,166]	A synthetic material that can enhance bone growth and fusion	Can enhance bone growth and fusion	May trigger a more severe immune response compared to traditional materials
Biodegradable polymers [27,167,168]	Materials that can gradually degrade and be replaced by host tissue	Can minimize the foreign body reaction and promote tissue integration	May not provide sufficient strength or stability for some spinal pathologies

minimize the foreign body reaction and improve the long-term success of these procedures [24](see Table 2). Medical professionals can use this information to make informed decisions regarding material selection and optimize the outcome of spinal fusion procedures.

4.1. Factors influencing the success of spinal fusion

The success of spinal fusion is influenced by a variety of factors, including the surgical technique, the type of implant used, and the patient's underlying medical conditions [119]. The surgical technique used for spinal fusion can impact the success of the procedure, as different techniques may result in varying levels of spinal stability and fusion rates [120]. For example, minimally invasive techniques, such as tubular retractor-assisted lumbar interbody fusion, have been shown to result in higher fusion rates compared to open techniques, such as posterolateral fusion [121]. The type of implant used for spinal fusion can also impact the success of the procedure, as different implants may have varying levels of biomechanical stability and biocompatibility [122]. For example, titanium implants have been shown to have higher biomechanical stability compared to polyetheretherketone (PEEK) implants, while PEEK implants have been shown to have a lower foreign body reaction compared to titanium implants [123]. Additionally, underlying medical conditions, such as osteoporosis, can also impact the success of spinal fusion, as they can weaken the vertebral bodies and reduce the stability of the implant [124]. Therefore, it is important to consider the various factors that can influence the success of spinal fusion in order to optimize the outcome of the procedure.

5. Strategies to minimize the impact of the foreign body reaction

There are several strategies that have been proposed to minimize the impact of the foreign body reaction on spinal fusion procedures. One approach is to modify the surface properties of the implant to reduce the foreign body reaction. This can be achieved by coating the implant with biocompatible materials or by modifying the surface roughness of the implant. Another approach is to use biologically active agents such as growth factors and cytokines to stimulate the fusion process and reduce the foreign body reaction. In addition, using minimally invasive surgical techniques, such as tubular retractor-assisted lumbar interbody fusion, has been shown to result in a lower foreign body reaction compared to open techniques, such as posterolateral fusion. Moreover, optimizing the patient's underlying medical conditions, such as osteoporosis, can also help to minimize the impact of the foreign body reaction on spinal fusion. Therefore, it is important to consider the various strategies that can be used to minimize the impact of the foreign body reaction in order to optimize the outcome of spinal fusion procedures.

5.1. Use of biocompatible materials in spinal fusion

The use of biocompatible materials in spinal fusion has been proposed as a strategy to reduce the foreign body reaction and improve the success of the procedure [125]. Biocompatible materials are materials that are compatible with the host tissues and elicit a minimal foreign body reaction [126]. The use of biocompatible materials in spinal fusion can reduce the magnitude and duration of the foreign body reaction [127], leading to a smaller fibrous capsule and improved bone growth and fusion [128]. For example, biocompatible coatings, such as hydroxyapatite coatings, have been shown to reduce the foreign body reaction and improve the success of spinal fusion compared to uncoated implants [129]. Additionally, the use of biocompatible materials, such as PEEK, has been shown to result in a lower foreign body reaction compared to titanium implants [130]. Therefore, the use of biocompatible materials in spinal fusion can be an effective strategy to reduce the foreign body reaction and improve the success of the procedure.

5.2. Optimization of surgical techniques to reduce tissue damage

Optimization of surgical techniques to reduce tissue damage is important in order to minimize the foreign body reaction and improve the success of spinal fusion [131]. Minimally invasive surgical techniques, such as tubular retractor-assisted lumbar interbody fusion, have been shown to result in reduced tissue damage compared to open techniques, such as posterolateral fusion [132]. Additionally, the use of smaller incisions and specialized instruments can help to reduce tissue damage and minimize the foreign body reaction [11]. Furthermore, the use of techniques such as nerve monitoring can help to minimize the risk of nerve damage during spinal fusion procedures [133]. The optimization of surgical techniques can help to reduce tissue damage and minimize the foreign body reaction, leading to improved outcomes and reduced risk of complications in spinal fusion procedures [134]. The underlying mechanism of minimizing the foreign body reaction in spinal fusion procedures is to reduce the interaction between the implant and the host tissues [135]. This can be achieved by reducing the magnitude and duration of the immune response, reducing the formation of a fibrous capsule around the implant, and improving the exchange of nutrients and cells between the implant and the surrounding tissues [65]. Minimizing the foreign body reaction can be achieved through various strategies, such as using biocompatible materials, optimizing surgical techniques, using biologically active agents such as growth factors and cytokines, and addressing underlying medical conditions such as osteoporosis [11]. By reducing the foreign body reaction, the success of spinal fusion can be improved by promoting bone growth and fusion, reducing the risk of implant failure, and minimizing the risk of complications such as pain, stiffness, and implant loosening.

Application of anti-inflammatory agents to reduce the immune response.

6. Conclusion and future directions

The application of anti-inflammatory agents to reduce the immune response in spinal fusion procedures is an emerging strategy to minimize the foreign body reaction and improve the success of the procedure. Anti-inflammatory agents, such as nonsteroidal anti-inflammatory drugs (NSAIDs) and corticosteroids, have been shown to reduce the magnitude and duration of the immune response in spinal fusion procedures. Additionally, the use of biologic agents, such as cytokine inhibitors, has also been proposed as a strategy to reduce the immune response in spinal fusion procedures. However, further studies are needed to fully understand the mechanisms underlying the application of anti-inflammatory agents in spinal fusion and to determine the optimal dosing and administration of these agents. In conclusion, the foreign body reaction can have a significant impact on the success of spinal fusion procedures, and minimizing the foreign body reaction of anti-inflammatory agents, and addressing underlying medical conditions are among the strategies that have been proposed to minimize the foreign body reaction in spinal fusion. Further research is needed to fully understand the mechanisms underlying these strategies and to determine the optimal approach to minimize the foreign body reaction and improve the success of spinal fusion.

6.1. Summary of the key points

The application of anti-inflammatory agents to reduce the immune response in spinal fusion procedures is an emerging strategy to minimize the foreign body reaction and improve the success of the procedure. The foreign body reaction can have a significant impact on the success of spinal fusion, and minimizing the foreign body reaction is essential to improve the outcome of the procedure. Strategies to minimize the foreign body reaction include the use of biocompatible material, optimization of surgical techniques, application of anti-inflammatory agents and addressing underlying medical conditions such as osteoporosis. The application of anti-inflammatory agents, such as nonsteroidal anti-inflammatory drugs (NSAIDs) and corticosteroids, as well as biologic agents, such as cytokine inhibitors, have been proposed to reduce the immune response and minimize the foreign body reaction in spinal fusion. Further research is needed to fully understand the mechanisms underlying the application of anti-inflammatory agents and to determine the optimal approach to minimize the foreign body reaction and improve the success of spinal fusion procedures.

6.2. Discussion of future research directions and potential improvements in spinal fusion surgery

Future advancements in spinal fusion surgery aim to address the challenges posed by the foreign body reaction, which can negatively impact the success of the procedure. The development of biocompatible materials and the optimization of surgical techniques, such as minimally invasive surgical techniques, are among the strategies being explored to reduce tissue damage and minimize the foreign body reaction. The application of anti-inflammatory agents, such as nonsteroidal anti-inflammatory drugs (NSAIDs) and corticosteroids, as well as biologic agents, such as cytokine inhibitors, may also play a role in reducing the immune response and improving the success of spinal fusion. The potential use of regenerative medicine and tissue engineering represents another area of investigation for promoting the growth of new, functional tissue and improving the success of spinal fusion. However, further research is needed to fully understand these strategies and determine the optimal approach to improve the success of spinal fusion procedures. In recent years, there has been growing interest in exploring alternative approaches to spinal fusion procedures that can minimize the risk of complications and improve patient outcomes. One promising approach is dynamic stabilization, which uses flexible implants to stabilize the spine and preserve range of motion. This approach can help reduce the risk of adjacent segment disease and preserve patient mobility. Another alternative approach is disc replacement, which replaces the damaged disc with an artificial disc. This approach can preserve motion and reduce the risk of adjacent segment disease, although it is still a relatively new procedure and may require further research to establish long-term safety and efficacy. By providing more detailed information on alternative approaches to spinal fusion procedures and their potential benefits and drawbacks, medical professionals and patients can make more informed decisions about treatment options and optimize the outcomes of spinal fusion procedures.

Author contribution statement

All authors listed have significantly contributed to the development and the writing of this article.

Data availability statement

No data was used for the research described in the article.

Additional information

No additional information is available for this paper.

Declaration of competing interest

The authors declare that they have no known competing financial interests or personal relationships that could have appeared to

influence the work reported in this paper.

References

- B.I. Martin, et al., Trends in lumbar fusion procedure rates and associated hospital costs for degenerative spinal diseases in the United States, 2004 to 2015, Spine 44 (2019).
- [2] J.L. Melancia, A.F. Francisco, J.L. Antunes, Chapter 35 spinal stenosis, in: J. Biller, J.M. Ferro (Eds.), Handbook of Clinical Neurology, vol. 119, Elsevier, 2014, pp. 541–549.
- [3] P.H. Newman, K.H. Stone, The etiology of spondylolisthesis, The Journal of Bone & Joint Surgery British 45-B (1963) 39-59.
- [4] M.T. Mathew, M.A. Wimmer, 13 tribocorrosion in artificial joints: in vitro testing and clinical implications, in: D. Landolt, S. Mischler (Eds.), Tribocorrosion of Passive Metals and Coatings, Woodhead Publishing, 2011, pp. 368–400.
- [5] Y. Fillingham, J. Jacobs, Bone grafts and their substitutes, The Bone & Joint Journal 98-B (2016) 6-9.
- [6] I. Lostalé-Seijo, J. Montenegro, Synthetic materials at the forefront of gene delivery, Nat. Rev. Chem 2 (2018) 258–277.
- [7] S. Virk, S. Qureshi, H. Sandhu, History of spinal fusion: where we came from and where we are going, HSS J. 16 (2020) 137–142.
- [8] B.A. Frost, S. Camarero-Espinosa, E.J. Foster, Materials for the spine: anatomy, problems, and solutions, Materials 12 (2019).
 [9] A.J. Vegas, et al., Combinatorial hydrogel library enables identification of materials that mitigate the foreign body response in primates, Nat. Biotechnol. 34 (2016) 345–352.
- [10] R. Trindade, et al., Osseointegration and foreign body reaction: titanium implants activate the immune system and suppress bone resorption during the first 4 weeks after implantation, Clin. Implant Dent. Relat. Res. 20 (2018) 82–91.
- [11] J.M. Anderson, A. Rodriguez, D.T. Chang, Foreign body reaction to biomaterials, Semin. Immunol. 20 (2008) 86-100.
- [12] P.J. Lennarson, F.T. Guillen, Spinal cord compression from a foreign body reaction to spinal cord stimulation: a previously unreported complication, Spine 35 (2010).
- [13] R.t. Watkins, R. Watkins 3rd, R. Hanna, Non-union rate with stand-alone lateral lumbar interbody fusion, Medicine 93 (2014) e275.
- [14] J.H. Healey, H.K. Brown, Complications of bone metastases, Cancer 88 (2000) 2940–2951.
- [15] I. Kondyurina, A.J.a.p.a. Kondyurin, Foreign Body Reaction (Immune Respond) for Artificial Implants Can Be Avoided, 2019.
- [16] T.R. Kyriakides, et al., Foreign body response to synthetic polymer biomaterials and the role of adaptive immunity 17 (2022), 022007.
- [17] A. Suwardi, et al., Machine Learning-Driven Biomaterials Evolution 34 (2022), 2102703.
- [18] S. Ivanovski, P.M. Bartold, Y.S.J.P. Huang, The role of foreign body response in peri-implantitis: What is the evidence? 90 (2022) 176–185.
- [19] Ö. Demir-Oğuz, A.R. Boccaccini, D.J.B.M. Loca, Injectable Bone Cements: what Benefits the Combination of Calcium Phosphates and Bioactive Glasses Could Bring?, vol. 19, 2023, pp. 217–236.
- [20] J.H. Park, et al., Materials and extracellular matrix rigidity highlighted in tissue damages and diseases: Implication for biomaterials design and therapeutic targets 20 (2023) 381-403.
- [21] M. Laubach, P. Kobbe, D.W.J.B. Hutmacher, Biodegradable Interbody Cages for Lumbar Spine Fusion: Current Concepts and Future Directions, 2022, 121699.
- [22] F. Eslami-Kaliji, N. Hedayat Nia, J.R. Lakey, A.M. Smink, M.J.P. Mohammadi, Mechanisms of Foreign Body Giant Cell Formation in Response to Implantable Biomaterials 15 (2023) 1313.
- [23] H. Matusiewicz, M.J. Richter, Metal ions release from metallic orthopedic implants exposed to tribocorrosion and electrochemical corrosion conditions in simulated body fluids: clinical context and in vitro experimental investigations, W.J.o.A.R. & Reviews 14 (2022) 261–283.
- [24] S. Capuani, G. Malgir, C.Y.X. Chua, A.J.B. Grattoni, T. Medicine, Advanced strategies to thwart foreign body response to implantable devices 7 (2022), e10300.
 [25] F.T. Foroushani, et al., Advances in Surface Modifications of the Silicone Breast Implant and Impact on its Biocompatibility and Biointegration, vol. 26, 2022.
- pp. 1–27.
- [26] C. Chu, et al., Application of biomaterials in periodontal tissue repair and reconstruction in the presence of inflammation under periodontitis through the foreign body response, Recent progress and perspectives 110 (2022) 7–17.
- [27] K.M. Naegeli, et al., Bioengineering human tissues and the future of vascular replacement 131 (2022) 109-126.
- [28] R.M. Visalakshan, et al., Antibacterial Nanostructured Surfaces Modulate Protein Adsorption, Inflammatory Responses, and Fibrous Capsule Formation, 2022.
- [29] R. Hama, et al., Recent Tissue Engineering Approaches to Mimicking the Extracellular Matrix Structure for Skin Regeneration 8 (2023) 130.
- [30] M.L. Wickramasinghe, G.J. Dias, K.M. Premadasa, G.P.J. J, B.M. o, R.P.B.A. B, A novel classification of bone graft materials 110 (2022) 1724–1749.
- [31] J. Litak, et al., Hydroxyapatite Use in Spine Surgery—Molecular and Clinical Aspect 15 (2022) 2906.
- [32] A. Lahousse, et al., Lifestyle and Pain Following Cancer: State-Of-The-Art and Future Directions, vol. 11, 2022, p. 195.
- [33] Z. Samsonchi, et al., Transplantation of Islet-Containing microcapsules modified with constitutional isomers of sulfated alginate in diabetic mice to mitigate fibrosis for Long-term glycemic control 432 (2022), 134298.
- [34] J.E. Rosen, N. Agrawal, D.R. Flum, J.M.J. Liao, Verbal descriptions of the probability of treatment complications lead to high variability in risk perceptions: a survey study, A.o.S (2022).
- [35] N. Su, C. Villicana, F.J.B. Yang, Immunomodulatory strategies for bone regeneration, A review from the perspective of disease types 286 (2022), 121604.
- [36] A.A. Khan, et al., Evaluation and management of hypoparathyroidism summary statement and guidelines from the second international workshop 37 (2022) 2568–2585.
- [37] M.N. Kravtsov, V.A. Manukovsky, G.G. Bulyshchenko, S.D. Mirzametov, V.A. Byvaltsev, J.F.i. S, Case report: full-endoscopic surgery for bullet wounds of the spine, a report of three cases 9 (2022).
- [38] J.J. Li, H. Zreiqat, Tissue response to biomaterials, in: R. Narayan (Ed.), Encyclopedia of Biomedical Engineering, Elsevier, Oxford, 2019, pp. 270–277.
- [39] B. Zhang, Y. Su, J. Zhou, Y. Zheng, D. Zhu, Toward a better regeneration through implant-mediated immunomodulation: harnessing the immune responses, Adv. Sci. 8 (2021), 2100446.
- [40] S. Jimi, A. Jaguparov, A. Nurkesh, B. Sultankulov, A. Saparov, Sequential delivery of cryogel released growth factors and cytokines accelerates wound healing and improves tissue regeneration, Front. Bioeng. Biotechnol. 8 (2020) 345.
- [41] T.N. Wight, S. Potter-Perigo, The extracellular matrix: an active or passive player in fibrosis? Am. J. Physiol. Gastrointest. Liver Physiol. 301 (2011) G950–G955.
- [42] R. Sridharan, A.R. Cameron, D.J. Kelly, C.J. Kearney, F.J. O'Brien, Biomaterial based modulation of macrophage polarization: a review and suggested design principles, Mater. Today 18 (2015) 313–325.
- [43] T. Rademakers, J.M. Horvath, C.A. van Blitterswijk, V.L.S. LaPointe, Oxygen and nutrient delivery in tissue engineering: approaches to graft vascularization, Jtissue eng regen med 13 (2019) 1815–1829.
- [44] O. Veiseh, A.J. Vegas, Domesticating the foreign body response: recent advances and applications, Adv. Drug Deliv. Rev. 144 (2019) 148–161.
- [45] B.N. Kharbikar, G.S. Chendke, T.A. Desai, Modulating the foreign body response of implants for diabetes treatment, Adv. Drug Deliv. Rev. 174 (2021) 87–113.
- [46] A. Carnicer-Lombarte, S.-T. Chen, G.G. Malliaras, D.G. Barone, Foreign Body Reaction to Implanted Biomaterials and Its Impact in Nerve Neuroprosthetics 9 (2021).
- [47] J.O. Abaricia, et al., Control of innate immune response by biomaterial surface topography, energy, and stiffness, Acta Biomater. 133 (2021) 58–73.
- [48] S. Franz, S. Rammelt, D. Scharnweber, J.C. Simon, Immune responses to implants a review of the implications for the design of immunomodulatory biomaterials, Biomaterials 32 (2011) 6692–6709.
- [49] L.E. Tracy, R.A. Minasian, E.J. Caterson, Extracellular matrix and dermal fibroblast function in the healing wound, Adv. Wound Care 5 (2016) 119–136.
- [50] J.K. Laitung, J. McClure, C.A. Shuttleworth, The fibrous capsules around static and dynamic implants: their biochemical, histological, and ultrastructural characteristics, Ann. Plast. Surg. 19 (1987) 208–216.

- [51] C.R. Bartoli, M.M. Nadar, J.J. Godleski, Capsule thickness correlates with vascular density and blood flow within foreign-body capsules surrounding surgically implanted subcutaneous devices, Artif. Organs 34 (2010) 857–861.
- [52] N.J. Hickok, I.M. Shapiro, Immobilized antibiotics to prevent orthopaedic implant infections, Adv. Drug Deliv. Rev. 64 (2012) 1165–1176.
- [53] M. Yuan, et al., Preimplantation exposure to bisphenol A and triclosan may lead to implantation failure in humans, BioMed Res. Int. 2015 (2015), 184845.
- [54] J.L. Hernandez, et al., Effect of tissue microenvironment on fibrous capsule formation to biomaterial-coated implants, Biomaterials 273 (2021), 120806.
- [55] F.-j. Zhu, Y.-I. Tong, Z.-y. Sheng, Y.-m. Yao, Role of dendritic cells in the host response to biomaterials and their signaling pathways, Acta Biomater. 94 (2019) 132–144.
- [56] W.J. Geelhoed, L. Moroni, J.I. Rotmans, Utilizing the foreign body response to grow tissue engineered blood vessels in vivo, J Cardiovascular Translational Res 10 (2017) 167–179.
- [57] P.O. Rujitanaroj, et al., Controlling fibrous capsule formation through long-term down-regulation of collagen type I (COL1A1) expression by nanofibermediated siRNA gene silencing, Acta Biomater. 9 (2013) 4513–4524.
- [58] D. Zhang, et al., Dealing with the foreign-body response to implanted biomaterials: strategies and applications of new materials, Adv. Funct. Mater. 31 (2021), 2007226.
- [59] L. Baggi, I. Cappelloni, M. Di Girolamo, F. Maceri, G. Vairo, The influence of implant diameter and length on stress distribution of osseointegrated implants related to crestal bone geometry: a three-dimensional finite element analysis, J. Prosthet. Dent 100 (2008) 422–431.
- [60] P.C. McAfee, et al., Symposium: a critical discrepancy—a criteria of successful arthrodesis following interbody spinal fusions, Spine 26 (2001).
- [61] S. Kligman, et al., The impact of dental implant surface modifications on osseointegration and biofilm formation, J. Clin. Med. 10 (2021).
- [62] A.W. Bridges, et al., Chronic inflammatory responses to microgel-based implant coatings, J. Biomed. Mater. Res. 94 (2010) 252-258.
- [63] E. Wintermantel, J. Mayer, T.N. Goehring, Composites for biomedical applications, in: K.H.J. Buschow, et al. (Eds.), Encyclopedia of Materials: Science and Technology, Elsevier, Oxford, 2001, pp. 1371–1376.
- [64] S. Gupta, H. Gupta, A. Tandan, Technical complications of implant-causes and management: a comprehensive review, Natl. J. Maxillofac. Surg. 6 (2015) 3–8.
- [65] N. Noskovicova, B. Hinz, P. Pakshir, Implant fibrosis and the underappreciated role of myofibroblasts in the foreign body reaction, Cells 10 (2021). [66] M. Corradi, M. Frattini, B. Panno, S. Tocco, F. Pogliacomi, Linked semi-constrained total elbow prosthesis in chronic arthritis: results of 18 cases,
- MUSCULOSKELETAL SURGERY 94 (2010) 11-23.
- [67] R. Bayston, Capsule formation around breast implants, JPRAS open 31 (2022) 123-128.
- [68] A. Carnicer-Lombarte, S.T. Chen, G.G. Malliaras, D.G. Barone, Foreign body reaction to implanted biomaterials and its impact in nerve neuroprosthetics, Front. Bioeng. Biotechnol. 9 (2021), 622524.
- [69] L. Luo, et al., Polycaprolactone nanofibrous mesh reduces foreign body reaction and induces adipose flap expansion in tissue engineering chamber, Int. J. Nanomed. 11 (2016) 6471–6483.
- [70] E.E. Rutherford, L.J. Tarplett, E.M. Davies, J.M. Harley, L.J. King, Lumbar spine fusion and stabilization: hardware, techniques, and imaging appearances, Radiographics 27 (2007) 1737–1749.
- [71] J.M. Anderson, A. Rodriguez, D.T. Chang, Foreign body reaction to biomaterials, Semin. Immunol. 20 (2008) 86-100.
- [72] N. Germic, Z. Frangez, S. Yousefi, H.-U. Simon, Regulation of the innate immune system by autophagy: neutrophils, eosinophils, mast cells, NK cells, Cell Death Differ. 26 (2019) 703–714.
- [73] B. Zhang, Y. Su, J. Zhou, Y. Zheng, D. Zhu, Toward a better regeneration through implant-mediated immunomodulation: harnessing the immune responses, Adv. Sci. 8 (2021), e2100446.
- [74] T. Duan, Y. Du, C. Xing, H.Y. Wang, R.-F. Wang, Toll-Like Receptor Signaling and its Role in Cell-Mediated Immunity, vol. 13, 2022.
- [75] M.T. Heise, Viral pathogenesis, in: Reference Module in Biomedical Sciences, Elsevier, 2014.
- [76] A. Gupta, et al., Identification of damage associated molecular patterns and extracellular matrix proteins as major constituents of the surface proteome of lung implantable silicone/nitinol devices, Acta Biomater. 141 (2022) 209–218.
- [77] R. van den Berg, G.R.M.M. Haenen, H. van den Berg, A. Bast, Transcription factor NF-kB as a potential biomarker for oxidative stress, Br. J. Nutr. 86 (2001) S121–S127.
- [78] T. Tanaka, M. Narazaki, T. Kishimoto, IL-6 in inflammation, immunity, and disease, Cold Spring Harbor Perspect. Biol. 6 (2014) a016295.
- [79] J.M. Zhang, J. An, Cytokines, inflammation, and pain, Int. Anesthesiol. Clin. 45 (2007) 27-37.
- [80] C. Wenzek, et al., The interplay of thyroid hormones and the immune system where we stand and why we need to know about it, Eur. J. Endocrinol. 186 (2022) R65–R77.
- [81] J.S. Marshall, R. Warrington, W. Watson, H.L. Kim, An introduction to immunology and immunopathology, Allergy Asthma Clin. Immunol. 14 (2018) 49.
- [82] J. Černý, I. Stříž, Adaptive innate immunity or innate adaptive immunity? Clin. Sci. 133 (2019) 1549–1565.
 [83] A. Limmer, et al., Efficient presentation of exogenous antigen by liver endothelial cells to CD8+ T cells results in antigen-specific T-cell tolerance, Nat. Med. 6
- (2000) 1348–1354. [84] K.J. Gordon, G.C. Blobe, Role of transforming growth factor-β superfamily signaling pathways in human disease, Biochim. Biophys. Acta (BBA) - Mol. Basis Dis.
- 1782 (2008) 197–228.
- [85] L. Moretti, J. Stalfort, T.H. Barker, D. Abebayehu, The interplay of fibroblasts, the extracellular matrix, and inflammation in scar formation, J. Biol. Chem. 298 (2022), 101530.
- [86] P.K. Givissis, S.I. Stavridis, P.J. Papagelopoulos, P.D. Antonarakos, A.G. Christodoulou, Delayed foreign-body reaction to absorbable implants in metacarpal fracture treatment, Clin. Orthop. Relat. Res. 468 (2010) 3377–3383.
- [87] S.M. Heckmann, M.G. Wichmann, W. Winter, M. Meyer, H.-P. Weber, Overdenture attachment selection and the loading of implant and denture-bearing area. Part 2: a methodical study using five types of attachment, Clin. Oral Implants Res. 12 (2001) 640–647.
- [88] P. Sahoo, S.K. Das, J. Paulo Davim, 1 tribology of materials for biomedical applications, in: J.P. Davim (Ed.), Mechanical Behaviour of Biomaterials, Woodhead Publishing, 2019, pp. 1–45.
- [89] M. Tobeiha, M.H. Moghadasian, N. Amin, S. Jafarnejad, RANKL/RANK/OPG pathway: a mechanism involved in exercise-induced bone remodeling, BioMed Res. Int. 2020 (2020), 6910312.
- [90] N. Udagawa, et al., Osteoclast differentiation by RANKL and OPG signaling pathways, J. Bone Miner. Metabol. 39 (2021) 19-26.
- [91] Z. Sheikh, P.J. Brooks, O. Barzilay, N. Fine, M. Glogauer, Macrophages, foreign body giant cells and their response to implantable biomaterials, Materials 8 (2015) 5671–5701.
- [92] L. Davenport Huyer, et al., Advanced strategies for modulation of the material-macrophage interface, Adv. Funct. Mater. 30 (2020), 1909331.
- [93] M. Augsten, Cancer-Associated Fibroblasts as Another Polarized Cell Type of the Tumor Microenvironment 4 (2014).
- [94] C. Atri, F.Z. Guerfali, D. Laouini, Role of human macrophage polarization in inflammation during infectious diseases, Int. J. Mol. Sci. 19 (2018).
- [95] E. Mariani, G. Lisignoli, R.M. Borzì, L. Pulsatelli, Biomaterials: foreign bodies or tuners for the immune response? Int. J. Mol. Sci. 20 (2019).
- [96] M.D. da Silva, et al., IL-10 cytokine released from M2 macrophages is crucial for analgesic and anti-inflammatory effects of acupuncture in a model of inflammatory muscle pain, Mol. Neurobiol. 51 (2015) 19–31.
- [97] P. Krzyszczyk, R. Schloss, A. Palmer, F. Berthiaume, The role of macrophages in acute and chronic wound healing and interventions to promote pro-wound healing phenotypes, Front. Physiol. 9 (2018) 419.
- [98] Y. Zhou, et al., β-elemene regulates M1-M2 macrophage balance through the ERK/JNK/P38 MAPK signaling pathway, Commun. Biol. 5 (2022) 519.
- [99] M.M. Alvarez, et al., Delivery strategies to control inflammatory response: modulating M1-M2 polarization in tissue engineering applications, J. Contr. Release : official journal of the Controlled Release Society 240 (2016) 349–363.
- [100] T.R. Kyriakides, et al., Foreign body response to synthetic polymer biomaterials and the role of adaptive immunity, Biomed. Mater. 17 (2022), 022007.
- [101] G.E. Kaiko, J.C. Horvat, K.W. Beagley, P.M. Hansbro, Immunological decision-making: how does the immune system decide to mount a helper T-cell response? Immunology 123 (2008) 326–338.

- [102] W. Huang, A. August, The signaling symphony: T cell receptor tunes cytokine-mediated T cell differentiation, J. Leukoc. Biol. 97 (2015) 477–485.
- [103] B.G. Keselowsky, J.S. Lewis, Dendritic cells in the host response to implanted materials, Semin. Immunol. 29 (2017) 33–40.
- [104] G. Arango Duque, A. Descoteaux, Macrophage cytokines: involvement in immunity and infectious diseases, Front. Immunol. 5 (2014) 491.
- [105] N. Tong, et al., Tumor associated macrophages, as the dominant immune cells, are an indispensable target for immunologically cold tumor-glioma therapy? Front. Cell Dev. Biol. 9 (2021), 706286.
- [106] A. Sharma, D. Rudra, Emerging Functions of Regulatory T Cells in Tissue Homeostasis, 2018, p. 9.
- [107] Y.Y. Wan, R.A. Flavell, TGF-beta and regulatory T cell in immunity and autoimmunity, J. Clin. Immunol. 28 (2008) 647–659.
- [108] S. Zhang, et al., The alterations in and the role of the Th17/treg balance in metabolic diseases, Front. Immunol. 12 (2021), 678355.
- [109] G.T. Nguyen, E.R. Green, J. Mecsas, Neutrophils to the ROScue: mechanisms of NADPH oxidase activation and bacterial resistance, Front. Cell. Infect.
 - Microbiol. 7 (2017) 373.
- [110] L. Chen, et al., Inflammatory responses and inflammation-associated diseases in organs, Oncotarget 9 (2018) 7204–7218.
- [111] T.C. Moon, A.D. Befus, M. Kulka, Mast cell mediators, Their Differential Release and the Secretory Pathways Involved 5 (2014).
- [112] S.J. Galli, M. Grimbaldeston, M. Tsai, Immunomodulatory mast cells: negative, as well as positive, regulators of immunity, Nat. Rev. Immunol. 8 (2008) 478–486.
- [113] A. Benzakour, et al., Artificial Intelligence in Spine Surgery, 2022, pp. 1-9.
- [114] F. Cakir, F.M. Ozkal, E.J.A. Sensoz, A.B. M, Performance assessment of biocompatible metals used in the treatment of femoral neck fractures 5 (2022) 3013–3022.
- [115] Z. Xiao, et al., Microfluidic Production of Zwitterion Coating Microcapsules with Low Foreign Body Reactions for Improved Islet Transplantation 18 (2022), 2202596.
- [116] A. Mussatto, et al., High strength bioinspired calcium phosphate-based material for bone repair applications 33 (2022), 104693.
- [117] H.M. El-Husseiny, et al., Hybrid biodegradable polymeric scaffolds for cardiac tissue engineering, in: Handbook Of Biodegradable Materials 1045-1092, Springer, 2023.
- [118] K.A. Aschbrenner, G. Kruse, J.J. Gallo, V.L.J.P. Plano Clark, F. Studies, Applying mixed methods to pilot feasibility studies to inform intervention trials 8 (2022) 217.
- [119] A.F. Mannion, A. Elfering, Predictors of surgical outcome and their assessment, European spine journal, in: The European Spinal Deformity Society, and the European Section of the Cervical Spine Research Society, vol. 15official publication of the European Spine Society, 2006, pp. S93–S108. Suppl 1.
- [120] C.S. Lee, C.J. Hwang, D.H. Lee, Y.T. Kim, H.S. Lee, Fusion rates of instrumented lumbar spinal arthrodesis according to surgical approach: a systematic review of randomized trials, Clinics in orthopedic surgery 3 (2011) 39–47.
- [121] S. Son, B.R. Yoo, S.G. Lee, W.K. Kim, J.M. Jung, Full-Endoscopic versus minimally invasive lumbar interbody fusion for lumbar degenerative diseases : a systematic review and meta-analysis, Journal of Korean Neurosurgical Society 65 (2022) 539–548.
- [122] A. Warburton, S.J. Girdler, C.M. Mikhail, A. Ahn, S.K. Cho, Biomaterials in spinal implants: a review, Neurospine 17 (2020) 101-110.
- [123] T.A. El Awadly, et al., A histomorphometric study on treated and untreated ceramic filled PEEK implants versus titanium implants: preclinical in vivo study, Clin. Oral Implants Res. 31 (2020) 246-254.
- [124] F. Tomé-Bermejo, A.R. Piñera, L. Alvarez-Galovich, Osteoporosis and the management of spinal degenerative disease (I), The archives of bone and joint surgery 5 (2017) 272–282.
- [125] A.R. Vaccaro, et al., The use of bioabsorbable implants in the spine, Spine J. 3 (2003) 227-237.
- [126] M. Durisin, 4 bioabsorbable behaviour of magnesium alloys an in vivo approach, in: T.S.N.S. Narayanan, I.-S. Park, M.-H. Lee (Eds.), Surface Modification of Magnesium and its Alloys for Biomedical Applications, Woodhead Publishing, Oxford, 2015, pp. 123–178.
- [127] H. Zhang, et al., Biomaterials for interbody fusion in bone tissue engineering, Front. Bioeng. Biotechnol. 10 (2022), 900992.
- [128] S. Rosy, R. Paulus, Bone development and growth, in: Y. Haisheng (Ed.), Osteogenesis and Bone Regeneration, vol. 1, IntechOpen, Rijeka, 2018. Ch.
- [129] S. Bohara, J. Suthakorn, Surface coating of orthopedic implant to enhance the osseointegration and reduction of bacterial colonization: a review, Biomater. Res. 26 (2022) 26.
- [130] R. Eftekhar Ashtiani, M. Alam, S. Tavakolizadeh, K. Abbasi, The role of biomaterials and biocompatible materials in implant-supported dental prosthesis, Evid. base Compl. Alternative Med. : eCAM 2021 (2021), 3349433.
- [131] K.T. Kim, S.H. Lee, K.S. Suk, S.C. Bae, The quantitative analysis of tissue injury markers after mini-open lumbar fusion, Spine (Phila Pa 31 (1976) 712–716, 2006.
- [132] B. Zheng, et al., Endoscopic techniques for lumbar interbody fusion: principles and context, BioMed Res. Int. 2022 (2022), 4979231.
- [133] J.H. Park, S.J. Hyun, Intraoperative neurophysiological monitoring in spinal surgery, World journal of clinical cases 3 (2015) 765–773.
- [134] B. Debono, et al., Consensus statement for perioperative care in lumbar spinal fusion: enhanced Recovery after Surgery (ERAS®) Society recommendations, Spine J. : official journal of the North American Spine Society 21 (2021) 729–752.
- [135] D.G. Barone, et al., Prevention of the foreign body response to implantable medical devices by inflammasome inhibition, Proc. Natl. Acad. Sci. USA 119 (2022), e2115857119.
- [136] M. Li, et al., Tuning the surface potential to reprogram immune microenvironment for bone regeneration 282 (2022), 121408.
- [137] K. Giragosyan, I. Chenchev, V. Ivanova, S.J.F.M. Zlatev, Immunological response to nonresorbable barrier membranes used for guided bone regeneration and formation of pseudo periosteum, a Narrative Review 64 (2022) 13–20.
- [138] O. Suljevic, S.F. Fischerauer, A.M. Weinberg, N.G.J. Sommer, B.M. T, Immunological reaction to magnesium-based implants for orthopedic applications. What do we know so far? A systematic review on in vivo studies (2022), 100315.
- [139] J. Choi, et al., Micro-textured silicone-based implant fabrication using electrospun fibers as a sacrificial template to suppress fibrous capsule formation 135 (2022), 112687.
- [140] T. Khodaei, E. Schmitzer, A.P. Suresh, A.P. Acharya, J.B. M, Immune response differences in degradable and non-degradable alloy implants 24 (2023) 153–170.
- [141] B. Grosgogeat, A. Vaicelyte, R. Gauthier, C. Janssen, M.J.M. Le Borgne, Toxicological risks of the cobalt-chromium alloys in dentistry, A Systematic Review 15 (2022) 5801.
- [142] A. Hansda, S. Mukherjee, K. Dixit, S. Dhara, G. Mukherjee, Immunological perspectives involved in tissue engineering, in: Regenerative Medicine: Emerging Techniques To Translation Approaches 37-55, Springer, 2023.
- [143] Y. Yang, et al., Strategies for advanced particulate bone substitutes regulating the osteo-immune microenvironment 17 (2022), 022006.
- [144] J. Wang, et al., CD301b+ macrophages mediate angiogenesis of calcium phosphate bioceramics by, CaN/NFATc1/VEGF axis 15 (2022) 446–455.
- [145] M. Iafisco, et al., Biocompatible antimicrobial colistin loaded calcium phosphate nanoparticles for the counteraction of biofilm formation in cystic fibrosis related infections 230 (2022), 111751.
- [146] N. Rana, et al., Systemic and local innate immune responses to surgical co-transplantation of mesenchymal stromal cells and biphasic calcium phosphate for bone regeneration 141 (2022) 440–453.
- [147] P. Fan, H. Yu, B. Xi, W.J.E.I. Tan, A Review on the Occurrence and Influence of Biodegradable Microplastics in Soil Ecosystems: Are Biodegradable Plastics Substitute or Threat?, 2022, 107244.
- [148] S. Tajvar, A. Hadjizadeh, S.S.J.I.B. Samandari, Biodegradation, Scaffold degradation in bone tissue engineering, An overview 180 (2023), 105599.
- [149] M. Samiei, et al., Application of collagen and mesenchymal stem cells in regenerative dentistry 17 (2022) 606–620.
- [150] P. Kumari, et al., Biological and Physicochemical Characterization of Flax Seed Mucilage Collagen Bio-Composite for Potential Use as Tissue Regenerative Scaffold, 2023, 105426.
- [151] M. Zheng, et al., A Review of Recent Progress on Collagen-based Biomaterials, 2022, 2202042.

- [152] E. Stocco, et al., Development of Two-Layer Hybrid Scaffolds Based on Oxidized Polyvinyl Alcohol and Bioactivated Chitosan Sponges for Tissue Engineering Purposes 23 (2022), 12059.
- [153] W. Xiang, et al., Applications of Chitosan-Based Biomaterials: from Preparation to Spinal Cord Injury Neuroprosthetic Treatment, 2023, 123447.
- [154] Y. Kim, et al., Chitosan-Based Biomaterials for Tissue Regeneration 15 (2023) 807.
- [155] B. Estupiñan, J. Adler, J.J.D. Swan, Titanium Hypersensitivity Diagnosed by Patch Testing After Anterior Cervical Spine Fusion 33 (2022) e59–e60.
- [156] M.-H. Wu, et al., In vitro and in vivo comparison of bone growth characteristics in additive-manufactured porous titanium, nonporous titanium, and porous tantalum interbody cages 15 (2022) 3670.
- [157] A. Bandyopadhyay, S. Ciliveri, S.J.J. Bose, I.o.S.o.t. I, Metal additive manufacturing for load-bearing implants 102 (2022) 561-584.
- [158] G. Szczęsny, et al., A review on biomaterials for orthopaedic surgery and traumatology, Past Present 15 (2022) 3622.
- [159] S. Dadkhahfar, M. Chehrassan, C.J.M.s. Faldini, Hypersensitivity Reactions to Metals in Spine Surgery, 2022, pp. 1-7.
- [160] S.R. Giri, B.K. Khamari, B.R.J. Moharana, M.T P, Joining of Titanium and Stainless Steel by Using Different Welding Processes: A Review, 2022.
- [161] J. Litak, et al., Metallic implants used in lumbar interbody fusion 15 (2022) 3650.
- [162] N. K Singh, N. Kumar Singh, D. Pandit, Saxena, K.K.J.A.i.M. & technologies, P, Recent trends in bio-materials and advances in design of spinal fusion implants 8 (2022) 2122–2141.
- [163] S. Gowtham, et al., A Survey on Additively Manufactured Nanocomposite Biomaterial for Orthopaedic Applications, 2022, p. 2022.
- [164] Z. Liu, X. He, S. Chen, H.J.C.I. Yu, Advances in the Use of Calcium Silicate-Based Materials in Bone Tissue Engineering, 2023.
- [165] X. Li, et al., Enhanced bone regenerative properties of calcium phosphate ceramic granules in rabbit posterolateral spinal fusion through a reduction of grain size 11 (2022) 90–106.
- [166] T. Fusco, et al., Arthrodesis of the Subtalar Joint Using a Novel Biphasic Calcium Phosphate Bone Graft, vol. 2, 2022, 100150.
- [167] G.C. Miceli, F.S. Palumbo, F.P. Bonomo, M. Zingales, M.J.P. Licciardi, Polybutylene Succinate Processing and Evaluation as a Micro Fibrous Graft for Tissue Engineering Applications, vol. 14, 2022, p. 4486.
- [168] M. Dziadek, et al., Polyphenolic Compounds Affect the Long-Term Degradation Behaviour of Polymer and Composite Materials Based on PCL, PLGA, and Bioactive Glass, 2023, e00568.