



# **Effects of Thyroid Hormones on Lipid Metabolism Pathologies in Non-Alcoholic Fatty Liver Disease**

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Abstract: The typical modern lifestyle contributes to the development of many metabolic-related disorders, as exemplified by metabolic syndrome. How to prevent, resolve, or avoid subsequent deterioration of metabolic disturbances and the development of more serious diseases has become an important and much-discussed health issue. Thus, the question of the physiological and pathological roles of thyroid hormones (THs) in metabolism has never gone out of fashion. Although THs influence almost all organs, the liver is one of the most important targets as well as the hub of metabolic homeostasis. When this homeostasis is out of balance, diseases may result. In the current review, we summarize the common features and actions of THs, first focusing on their effects on lipid metabolism in the liver. In the second half of the review, we turn to a consideration of non-alcoholic fatty liver disease (NAFLD), a disease characterized by excessive accumulation of fat in the liver that is independent of heavy alcohol consumption. NAFLD is a growing health problem that currently affects ~25% of the world's population. Unfortunately, there are currently no approved therapies specific for NAFLD, which, if left uncontrolled, may progress to more serious diseases, such as cirrhosis or liver cancer. This absence of effective treatment can also result in the development of non-alcoholic steatohepatitis (NASH), an aggressive form of NAFLD that is the leading cause of liver transplantation in the United States. Because THs play a clear role in hepatic fat metabolism, their potential application in the prevention and treatment of NAFLD has attracted considerable research attention. Studies that have investigated the use of TH-related compounds in the management of NAFLD are also summarized in the latter part of this review. An important take-home point of this review is that a comprehensive understanding of the physiological and pathological roles of THs in liver fat metabolism is possible, despite the complexities of this regulatory axis—an understanding that has clinical value for the specific management of NAFLD.

Keywords: thyroid hormone; lipid metabolism; non-alcoholic fatty liver disease (NAFLD)

# 1. Thyroid Hormones and Action

## 1.1. Thyroid Hormones

Thyroid hormones (THs) are amino acid-derived hormones formed by the addition of iodine to thyroglobulin, the tyrosine component of a glycoprotein, that influence the growth, development, and metabolism of vertebrates. The production of THs is regulated by the hypothalamic–pituitary–thyroid (HPT) axis [1]. Specifically, thyroid-releasing hormone



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**Copyright:** © 2022 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (https:// creativecommons.org/licenses/by/ 4.0/). (TRH), released by the hypothalamus, stimulates the anterior pituitary gland to release thyroid-stimulating hormone (TSH), which, in turn, stimulates the thyroid to synthesize and release THs. A negative feedback loop is present in the HPT axis such that THs act on the pituitary and hypothalamus to inhibit TSH production. There are two main forms of THs secreted by the thyroid gland: T4 (3,5,3',5'-tetraiodothyroxine), or thyroxine, the major product and precursor form of THs, and 3,5,3'-triiodothyronine (T3). T3, most of which can be generated in peripheral tissues by deiodination of T4 by iodothyronine deiodinases (DIO), is considered the potent active form. There are three main types of DIO (DIO1-3) in humans: DIO1 and DIO2 are responsible for the activating conversion of T4 to T3, whereas DIO3 converts T4 and T3 to the inactive metabolites reverse T3 (rT3) and rT2, respectively [2]. Therefore, modulation of the activity of DIOs and tissue-specific expression of DIOs can both positively and negatively regulate the functions of THs.

# 1.2. TH Actions

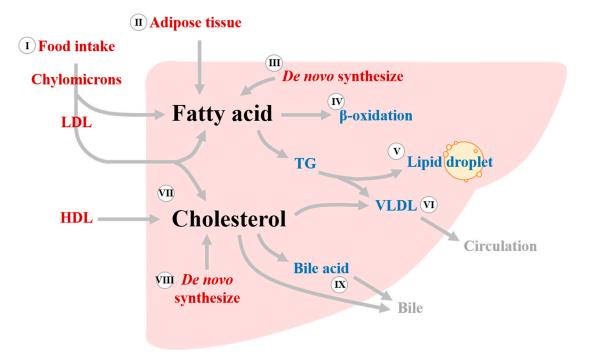
Canonically, THs act as ligands for TH receptor  $\alpha$  (TR $\alpha$ ) and TR $\beta$  encoded by THRA and *THRB* genes, respectively. TRs belong to the nuclear receptor superfamily, whose members can either induce or suppress gene expression by forming heterodimers with another nuclear receptor, retinoid X receptor (RXR), and binding to the TR-response element (TRE) on the promoter of target genes. Besides, TRs bind to promoters of target genes as monomer or homodimer, however, the binding is not as stable as the heterodimers of TR and RXR [3,4]. Notably, TRs regulate downstream target both in the presence or absence of THs. In the absence of THs, nuclear receptor corepressor 1 (NCoR1) and NCoR2 (also known as silencing mediator of retinoid and thyroid hormone receptors [SMRT]) serve as corepressors which are recruited to TR-RXR heterodimers bound to the TRE to prevent gene transcription. Further, in the presence of THs leads to conformational change in TRs and results in releasing of corepressors and recruiting of coactivators, such as SRC (steroid receptor coactivator) and CREB-binding protein/p300, and the target genes are turned on. T3 is the main active ligand for TRs because of the affinity of T3 is about 10-fold higher than that of T4. Binding of THs to TRs to transcriptionally regulate gene expression is considered the canonical pathway of TH action. However, some effects stimulated by THs occur within minutes, indicating that THs can act without directly binding the regulatory domain of target promoters, that is, though noncanonical pathways [5]. Flamant et al. precisely defined four types of TH action [6]. The canonical pathway is defined as type 1 action. Type 2 action refers to indirect binding of TH-TRs to DNA via other DNAbinding proteins, such as activating protein-1 [7], cyclic AMP response element-binding protein [8], and nuclear factor-kapa B [9], to interfere with the activity of THs in regulating the expression of genes. The third type of TH action corresponds to regulation of kinase pathways by TH-TRs through phosphorylation, rather than direct or indirect regulation of transcription. For example, phosphatidylinositol 3-kinase (PI3K) and mitogen-activated protein kinase (MAPK) pathways have been demonstrated to be phosphorylated and activated by TH-TRs [10,11]. Finally, type 4 action describes the signaling of THs through TR-independent pathways. Integrin  $\alpha V\beta 3$  has been identified as one such TH-responsive receptor [12,13]. For more details on TH and TR molecular mechanisms, refer to our previous publication [14].

#### 2. TH and Lipid Metabolism in the Liver

#### 2.1. Lipid Metabolism in the Liver

The liver plays crucial roles in lipid metabolism, including acquisition of lipid, lipid storage, and lipid consumption. Lipids metabolized in the liver can be roughly divided into two types: fatty acid and cholesterol. There are two main sources of fatty acid that are sent to the liver for metabolism. The first is chylomicrons, which are derived from the intestine following digestion and absorption of lipid from food intake. The second is fatty acids released from adipose tissue by lipolysis. In addition to the uptake of fatty acids from extrahepatic sources, de novo lipogenesis (DNL) also occurs in the liver when

excess glucose accumulates. After uptake or synthesis by the liver, fatty acids can be used for energy production via  $\beta$ -oxidation in mitochondria or peroxisomes, triglyceride (TG) synthesis and storage in lipid droplets, or TG secretion into the circulation in the form of very low-density lipoproteins (VLDL) as source of cholesterol for peripheral tissues. Cholesterol is another type of lipid metabolized mainly in the liver. Approximately 30% of cholesterol comes from food intake, whereas 70% comes from de novo synthesis, mainly in the liver. The fates of cholesterol in the liver include conversion to bile acid, packing in lipoproteins and secretion into the bloodstream, and use as a plasma membrane component. Hepatic lipid metabolism is summarized schematically in Figure 1.



**Figure 1.** Lipid metabolism in the liver. The liver is the main site of fatty acid and cholesterol metabolism. Fatty acids come from three main sources: food intake (I), adipose tissue lipolysis (II), and de novo lipogenesis (III). Fatty acids are then used for the production of energy via  $\beta$ -oxidation (IV), synthesis of TGs and storage in lipid droplets (V), or packing into VLDLs and secretion into the circulation (VI). Hepatic cholesterol comes from circulating lipoproteins (LDL and HDL, VII) or de novo synthesis in the liver (VIII). Cholesterol is subsequently used as a component of the plasma membrane of hepatocytes, packaged into VLDLs and released as a cholesterol source for extrahepatic organs (VI), or directly released or converted to bile acid and released into the bile duct (IX). TG, triglyceride; VLDL, very low-density lipoprotein; LDL, low-density lipoprotein; and HDL, high-density lipoprotein.

#### 2.2. THs and Lipid Metabolism in the Liver

The liver is one of the major target organs of THs, reflecting the high expression of TRs, especially TR $\beta$ , in this organ. Reports of TH effects on lipid metabolism have been documented as early as the end of the 19th and early 20th centuries [15]. Initially, it was noted that reversible hyperlipemia, with elevated levels of plasma total cholesterol (TC) and low-density lipoprotein cholesterol (LDL), were frequently observed in patients with hypothyroidism [16,17]. It was subsequently found that blood lipids were regulated by THs in patients with hypothyroidism who underwent thyroid replacement therapy [18–20], and shown that treatment with THs significantly reduce plasma cholesterol and lipoproteins. Furthermore, THs action through TR $\beta$  to regulate hepatic lipid metabolism was demonstrated in mice carrying a dominant-negative mutation in Thrb, the Thrb<sup>PV/PV</sup> mice [21]. Decreased fatty acid  $\beta$ -oxidation and activated peroxisome proliferator-activated receptor- $\gamma$  (PPAR- $\gamma$ ) due to dominant-negative effects of Thrb<sup>PV/PV</sup> on TR $\beta$  contribute

to hepatic lipid accumulation and steatosis. Collectively, these studies have established the critical roles of THs in lipid metabolism. The mechanisms by which THs affect lipid metabolism processes in the liver are summarized and described below.

## 2.2.1. Regulation of Fatty Acid Uptake by THs

In general, the liver takes up about 25% of total free fatty acids from the blood flowing through it [22], a process mediated and regulated by translocases and transporters on the membrane of hepatocytes. Adipose differentiation-related protein (ADRP, also known as adipophilin, perilipin-2), fatty acid transporter proteins (FATPs), fatty acid translocase (FAT, also known as CD36), and fatty acid-binding proteins (FABPs) are the transporters and translocases responsible for fatty acid uptake by the liver [23]. A study in hypothyroid rats showed that fatty acid uptake in the liver is decreased relative to that in euthyroid rats [24]. Decreased hepatic FABPs and FAT expression have been demonstrated in rats with postnatal hypothyroidism [25]. In addition, another study in rats demonstrated that T3 inhibits fatty acid transport and utilization by the liver by decreasing the expression of liver FABP [26]. Collectively, these studies indicate that THs can regulate lipid metabolism by affecting fatty acid uptake through regulation of the expression of transporters or translocases. In addition to regulation of liver FABP by THs, it has been demonstrated that adipocyte FABP increases thermogenesis in brown adipose tissues by promoting the conversion of inactive T4 to active T3 [27]. It would be interesting to determine whether a positive feedback mechanism of FABP on THs operates in the liver.

#### 2.2.2. De Novel Lipogenesis (DNL)

In addition to promoting uptake of exogenous fatty acid, excess glucose can stimulate DNL in the liver. Glucose entering the glycolytic pathway is converted to pyruvate, which is then transported to mitochondria through the mitochondria pyruvate carrier (MPC). After the action of dehydrogenase and subsequent condensation with oxaloacetate (OAA), pyruvate forms citrate, which can enter the tricarboxylic (TCA) cycle to produce energy. Alternatively, if the cell has excess energy, citrate can be sent from mitochondria via the citrate carrier (CiC) to the cytoplasm, where it is reconverted to OAA and acetyl-CoA by ATP citrate lyase (ACLY). Acetyl-CoA, a substrate for DNL, is then catalyzed in turn by two critical enzymes, acetyl-CoA carboxylase (ACC) and fatty acid synthase (FAS). Many of the above-mentioned key factors that participate in DNL have been shown to be regulated by THs. Decreased expression of MPC and CiC is observed in hypothyroid rats, an effect opposite that observed in rats with hyperthyroidism [28–31]. ACYL expression is decreased in cultured primary hepatocytes isolated from thyroidectomized rats and is restored by treating cells with T3 [32]. In rats with propylthiouracil-induced hypothyroidism, ACYL is significant decreased relative to that in euthyroid rats [33]. ACYL is transactivated by sterol regulatory element-binding protein-1 (SREBP-1) [34], a member of the SREBP family of important lipogenic transcription factors. Although SREBP-1 is a bone fide TH-regulated gene [35,36], some contradictory results have been reported. For example, in one study using a mouse model, the authors demonstrated that T3 acts through a negative TRE on the promoter to directly inhibit SREBP-1 expression [35], whereas another study in human HepG2 cells reported that T3 activates SREBP-1 via non-genomic actions [36]. In addition, T3 induces SREBP-1 expression and also increases the activated form of SREBP-1 in chick embryo hepatocytes; this increased SREBP-1 acts in conjunction with TR to regulate ACC transcription [37]. FAS, another critical lipogenic enzyme, shares the same SREBP-1- and TH-independent or coordinated regulatory mechanisms as ACC [33,38,39].

## 2.2.3. $\beta$ -Oxidation

 $\beta$ -oxidation is the process that take place in mitochondria by which fatty acids are broken down to produce energy. In the liver, the release of fatty acids from TGs is mainly executed by hepatic lipase (HL) and adipose triglyceride lipase (ATGL). Levothyroxine, a TH replacement, significantly induces HL in individuals with subclinical hypothyroidism [40].

T3 induces recruitment of ATGL to lipid droplet (LD) surfaces to promote the release of lipids from LDs and entry into the  $\beta$ -oxidation pathway [41]. Transporting of fatty acids, especially long-chain fatty acids, from the cytosol to mitochondria is mediated by carnitine palmitoyl acyl-CoA transferase 1 (CPT1), the rate-limiting enzyme in  $\beta$ -oxidation. CPT1 converts fatty acyl-CoA to acylcarnitine, which can be transported across the inner mitochondria membrane via carnitine translocase (CAT). Once in mitochondria, acylcarnitine can be converted back to fatty acyl-CoA by CPT2 and then enters  $\beta$ -oxidation followed by the TCA cycle to produce energy. There is evidence that hepatic CPT1 activity is decreased in rats with hypothyroidism and is also a direct target of TH-TR [42–44]. Fibroblast growth factor 21 (FGF21), expressed predominantly in the liver, adipose tissue and pancreas, is involved in many metabolic processes, including lipid metabolism. It has been found that transgenic mice overexpressing FGF21 exhibit markedly increased hepatic  $\beta$ -oxidation [45] and reduced hepatic steatosis [46]. Notably, FGF21 is a demonstrated T3-target gene and its regulation is PPAR $\alpha$  dependent [46].

#### 2.2.4. Cholesterol Homeostasis

Chylomicron remnants (CRs) and lipoproteins (LDL and HDL), another source of liver lipids, are also critical for the maintenance of cholesterol homeostasis by the liver. CRs and lipoproteins enter the liver via LDL receptor (LDLR)- and LDLR-related protein (LRP)-mediated endocytosis [47]. It has long been known that there is a negative correlation between TH and cholesterol in plasma [15], an observation that led to the discovery that levels of LDLRs and LRPs are regulated by THs [48,49]. In addition to being directly regulated at the transcriptional level by THs through TR $\beta$ , LDLRs can be indirectly regulated by THs through SREBP2, a key factor in cholesterol biosynthesis in the liver that is transcriptionally regulated by THs [50]. Proprotein convertase subtilisin/kexin type 9 (PCSK9), a secreted protein mainly expressed in the liver, impairs cholesterol uptake by the liver by inducing LDLR degradation [51]. It has been shown that PCSK9 is significantly increased in patients with subclinical hypothyroidism relative to euthyroid controls [52], and that THs and PCSK9 are negatively correlated in humans [52,53]. Therefore, in addition to direct and indirect transcriptional regulation, preventing degradation of LDLR might also be a mechanism by which THs augment cholesterol uptake by the liver. It has further been proposed that THs may regulate plasma cholesterol in an LDLR-independent manner [54]. In mice lacking LDLRs, T3 and TRβ agonists markedly reduce plasma cholesterol by inducing expression of cholesterol 7- $\alpha$ -hydroxylase (CYP7A1) and subsequent promotion of bile acid synthesis. The conversion of cholesterol to bile acid is the major mechanism for clearance of cholesterol by the liver. In an in vitro model, it was shown that the gene encoding CYP7A1, a key enzyme involved in bile acid synthesis, is a direct target of TR $\beta$  and is regulated by THs [55]. In addition to increasing the conversion of cholesterol to bile acid, THs can promote biliary elimination of cholesterol. In hypophysectomized rats, administration of T3 ameliorates hypercholesterolemia in association with increased hepatic secretion of cholesterol and upregulation of ABCG5 (ATP-binding cassette, subfamily G, member 5) and ABCG8 [56]. ABCG5 and ABCG8 form a heterodimeric transporter located at the apical plasma membrane of hepatocytes that is responsible for transport of hepatic cholesterol to bile [57]. Studies performed in ABCG5-knockout mice subsequently demonstrated the importance of ABCG5/ABCG8 in biliary cholesterol secretion [58]. In addition to processing cholesterol coming from outside the liver, cholesterol can be synthesized in the liver. HMG-CoA reductase (HMGR) is the rate-limiting enzyme in cholesterol synthesis in the liver, and its expression has been reported to be induced by SREBP2 [59]. As noted above, the gene encoding SREBP2 is a target of THs; thus, THs also affect cholesterol biosynthesis in the liver.

## 3. Pathological Roles of TH Associated with Lipid Metabolism in the Liver

## 3.1. Lipid Metabolism and Liver Diseases: Non-Alcoholic Fatty Liver Disease (NAFLD)

Dysregulation of physiological lipid metabolism is associated with various diseases, such as obesity, diabetes, cardiovascular diseases, and dyslipidemia, among others, that affect different organs [60,61]. Abnormal lipid metabolism in the liver results in the accumulation of excess fat—the so-called fatty liver syndrome; in particular, non-alcoholic fatty liver disease (NAFLD), which is not caused by excess alcohol consumption or viral infection. NAFLD, which affects approximately a quarter of the world's population, is the most prevalent chronic liver disease worldwide [62]. The pathogenesis of NAFLD is complicated and not fully elucidated. Lifestyle, dietary habits, genetic and epigenetic factors, and microbiota can all result in metabolic dysfunction and contribute to chronic liver disease.

Liver biopsy and pathological assessment is currently the gold standard for diagnosis of NAFLD, nevertheless, this examination is invasive. As a prescreen, or to avoid invasive testing, NAFLD is usually diagnosed based on imaging and is defined as the presence of at least 5% hepatic steatosis in the context of little or no alcohol consumption or other chronic liver disease [63]. NAFLD can be classified into two types: non-alcoholic fatty liver (NAFL) and non-alcoholic steatohepatitis (NASH). In most patients, NAFLD persists as NAFL, which is considered non-progressive, but about 20% of NAFLD cases progress to NASH [64]. Among the typical features of NASH observed in histological assessments are steatosis, lobular inflammation, and ballooning (hepatocellular injury) [65]. Paik et. al. indicated that, although viral hepatitis remains the leading cause of liver-related death, deaths due to NAFLD are the fastest increasing among non-viral liver-related causes [66]. During the course of NAFLD, a small proportion of individuals will progress to cirrhosis and even hepatocellular carcinoma (HCC) [64]. Nevertheless, there is currently no approved therapy for NAFLD. The increasing prevalence of NAFLD and absence of effective treatment has resulted in NASH becoming the leading cause of liver transplantation in the United State [67]. Both establishment of a prognostic system for NAFLD for identifying the population at high risk of progressing to NASH and the development of therapeutic drugs for NAFLD are currently urgent needs.

#### 3.2. Correlations between THs and NAFLD

NAFLD is bidirectionally associated with metabolic diseases such as type 2 diabetes mellitus, dyslipidemia, and hypertension [68], which are also associated with TH dysfunction. Evidence suggests that imbalanced energy metabolism, especially excess carbohydrates and fat delivery into the liver, is a crucial driver of the development of NAFLD [69]. As noted above, THs play crucial roles in energy and metabolic homeostasis; thus, it is reasonable to speculate that THs also play pathogenic roles during the development of NAFLD. Consistent with this, associations between NAFLD and thyroid diseases have been demonstrated. For example, a cross-sectional study of 4648 subjects categorized as controls or patients with subclinical or overt hypothyroidism demonstrated a significant association of NAFLD with hypothyroidism [70]. A higher prevalence of hypothyroidism in patients with NAFLD compared with the control group has also been demonstrated, and patients with hypothyroidism were shown to be more susceptible to NASH [71]. In addition, higher free T4 levels in plasma are associated with a decreased risk of NAFLD [72]. Moreover, in patients diagnosed with NAFLD, low free T3 is independently associated with a high probability of fibrosis and liver stiffness [73]. Collectively, these observations suggest a close association between thyroid function and the development and progression of NAFLD. Nevertheless, discrepant findings have also emerged [74–76], possibly owing to variations in the ethnicity of study populations and criteria used for diagnosing hypothyroidism and NAFLD. A large-scale meta-analysis that enrolled 44,140 individuals across 12 cross-sectional and three longitudinal studies revealed that variably defined primary hypothyroidism is significantly and independently associated with the presence and severity of NAFLD, although a causal relationship could not be definitely established. The authors suggested that screening for primary hypothyroidism could be beneficial in distinguishing NAFLD patients who are at high risk of progressing [77]. In sum, multiple lines of evidence support pathological roles of THs in the course of NAFLD, although some conflicting findings remain to be reconciled. Notably, hypothyroidism-induced NAFLD (HIN) is considered a distinct disease entity [78]. Additional large, well-designed studies are warranted to elucidate the relationship between THs and NAFLD and the underlying mechanism linking them.

# 4. Drugs Targeting THs and Their Clinical Applications in NAFLD

## 4.1. Management of NAFLD

In the absence of approved NAFLD-specific treatments, lifestyle modifications with the goal of weight loss, such as improved diet quality and increased physical activity, are the mainstays of treatment for NAFLD [79]. Because visceral obesity promotes insulin resistance and proinflammatory signaling, which are important risk factors for NAFLD, weight loss is directly correlated with substantial improvement in histological outcomes, even fibrosis. For the severely obese population, bariatric surgery may be a therapeutic option [80]. Several drugs developed for other indications have also been tested for potential clinical effects on NAFLD. In one example, pioglitazone, approved for the treatment of type 2 diabetes, combined with vitamin E demonstrated some benefit in randomized trials [81]. However, potent and safe therapeutic strategies specific to NAFLD, the focus of numerous ongoing studies, await discovery.

## 4.2. Therapeutic Potential of THs, TH Mimetics, and TH Metabolites in NAFLD

Given the functions of THs in weight loss and metabolic and energy homeostasis, it is reasonable to anticipate that THs might have potential for the control and treatment of NAFLD. As expected, exogenously administered T3 has been demonstrated to lower hepatic fat content in various experimental animal models of NAFLD [26,82,83]. However, the physiological actions of THs affect almost every organ; thus, administration of THs is accompanied by adverse extrahepatic effects (especially in the heart, muscle, and bone), hampering clinical applications of THs in NAFLD [84]. Expression of TR isoforms is tissue dependent, with TR $\alpha$  being the predominant form expressed in bone and heart and TR $\beta$ predominating in the liver [85]. Therefore, compounds that specifically target TR $\beta$  were thought to be a solution to the problem of undesired adverse effects. In recent decades, several TR $\beta$ -selective agonists have been developed and investigated, including GC-1 and its derivatives, KB2115, MB07811, and MGI-3196 (Table 1), and the potential of several TH metabolites in the treatment of NAFLD have been demonstrated, as detailed below.

Table 1. Summary	y of thyromimetics.
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Compound	Туре	Favorable Effects	Unfavorable Effects	Clinical Trials
GC-1 (Sobetirome)	TRβ- and liver- selective thyromimetic	10-fold lower TRα affinity [86] ↓ triglyceride [87,88] ↓ cholesterol [87] Prevent hepatic steatosis in rodent model [26,88,89] ↑β-oxidation [26]	Fasting hyperglycemia and hyper- insulinemia [89]	
GC-24	TRβ-selective thyromimetic	40-fold higher TRβ affinity than TRα [90] ↑ energy expenditure [91] Eliminate the increase in adiposity [91] ↑ insulin sensitivity [91] ↓ triglyceride [91]	No improve in hepatic cholesterol [91] Less potent on liver target [92]	

Compound	Туре	Favorable Effects	Unfavorable Effects	Clinical Trials
KB141	TRβ-selective thyromimetics	14-fold higher affinity to TRβ than TRα [93] ↑ metabolic rate [93] ↓ cholesterol [93] ↓ lipoprotein [93] ↓ body weight [93]	↑ heart rate [93] No liver- selective [93,94]	
KB-2115 (Eprotirome)	TRβ- and liver- selective thyromimetic	↓ triglyceride [88] Ameliorate hepatic steatosis [88,89] ↓ LDL cholesterol [95] ↑ bile acid synthesis [95]	Fasting hyperinsulinemia [89] Liver injury and cartilage damage [96]	NCT00593047 (phase 2) [97] NCT00776321 (phase 2) NCT00677248 (phase 2) NCT01410383 (phase 3) [96]
MB07811 (VK2809)	TRβ- and liver-selective prodrug	↓ cholesterol [98] ↓ triglyceride [82,98] ↓ hepatic steatosis [82] ↓ free fatty acid [82] ↑ β-oxidation [82]		NCT02927184 (phase 2) NCT04173065 (phase 2)
MGL-3196 (Resmetirom)	TRβ- and liver- selective thyromimetic	28-fold higher affinity to TRβ than TRα [99] ↓ cholesterol [99,100] ↓ triglyceride [99,100] ↓ lipoprotein [100] ↓ fibrosis [100]	Mild diarrhea and nausea [100]	NCT03900429 (phase 3) [101] NCT04643795 (phase 1) NCT04951219 (phase 3) NCT04197479 (phase 3)

## Table 1. Cont.

## 4.2.1. GC-1

GC-1, (3,5-dimethyl 1-4(4'-hydroxy-3'-isopropylbenzul- phenoxy) acetic acid), also known as sobetirome, was the first synthetic TR-selective TH analog developed [86]. GC-1 is a shape-mimic of T3 with several structural modifications that weaken the affinity of GC-1 for TR $\alpha$  by ~10-fold but without compromising its affinity for TR $\beta$ . In a mouse model of hypothyroidism, GC-1 showed better TG-lowering and similar cholesterol-lowering effects compared with T3 [87]; it also exhibited enhanced hepatic targeting. Both TR $\beta$ - and liverselectivity contribute to the ability of GC-1 to affect lipid metabolism without altering heart rate or weight [87]. The potential of GC-1 to ameliorate NAFLD was reported in several experimental rodent models [26,88]. In a high-fat-diet (HFD) rat model, GC-1 treatment was shown to reduce hepatic TGs by ~75%, but it also caused fasting hyperglycemia and hyperinsulinemia, reflecting a decrease in insulin sensitivity [89]. Similar results were also observed in a study using an ob/ob mice model [88]. These results suggest that, although GC-1 may have potential to treat NAFLD, the effects of GC-1 on insulin sensitivity must be considered.

#### 4.2.2. GC-24

The GC-24, a derivative of GC-1, obtained by modifying the structure of GC-1 exhibits ~40-fold higher binding affinity for TR $\beta$  than TR $\alpha$  [90]. In a HFD rat model, administration of GC-24 eliminated the HFD-induced increase in adiposity, increased insulin sensitivity, reduced plasma TG levels, and partially reduced plasma cholesterol levels without causing cardiac hypertrophy. However, hepatic cholesterol content remained high in animals treated with GC-24 [91], which was, subsequently, reported to be less potent against liver targets than GC-1 [92].

# 4.2.3. KB141

KB141 (3,5-dichloro-4-(4-isopropy acetic acid)), another derivative of GC-1, achieved a ~14-fold higher affinity for TR $\beta$  than TR $\alpha$  by virtue of structural modifications [93]. In

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preclinical animal models, KB141 was shown to increase metabolic rate in mice and rats, and significantly reduce cholesterol, lipoprotein, and body weight in primates [93]. It also has only minimal effects on the heart, but, unlike GC-1, does not preferentially accumulate in the liver [93,94].

## 4.2.4. KB2115

KB2115 (3-(3,5-dibromo-4-(4-hydroxy-3-(1-methylethyl)-phenoxy)-phenyl)-amino-3oxopropanoic acid), also known as eprotirome, is a brominated TRβ-selective TH replacement drug that is more liver-specific than GC-1 and has no deleterious effects on the heart [95,97]. Similar to GC-1, administration of KB2115 effectively prevented hepatic steatosis in rodents [88,89]. However, unlike the case for GC-1, administration of KB2115 did not lead to fasting hyperglycemia, although fasting hyperinsulinemia was still noted. A randomized phase 3 study showed that KB2115 lowered cholesterol levels in patients with familial hypercholesterolemia, but also reported evidence of liver injury and cartilage damage [96].

## 4.2.5. MB07811

MB07811, ((2R,4S)-4-(3-chlorophenyl)-2-((3,5-dimethyl-4-(4'-hydroxy-3'-isopropylbenzyl) phenoxy)methyl)-2-oxido-(1,3,2)-dioxaphosphonane), also known as VK2809, is a prodrug can be processed by hepatic cytochrome P450 enzymes to the active TR agonist, MB07344 ((3,5-dimethyl-4-(4'-hydroxy-3'-isopropylbenzyl)phenoxy) methylphosphonic acid). Amelio-rated liver targeting without unfavorable effects on cardiac of MB07811 was demonstrated. Also, administration of MB07811 can reduce hepatic TG levels and also plasma cholesterol and TG [98]. In several experimental models of NAFLD, MB07811 was demonstrated to significantly reduce hepatic steatosis, plasma fatty acids, and TGs without causing liver fibrosis or other liver damage [82]. These experimental findings support the potential of MB0711 for use in treating NAFLD. Two phase 2 clinical trials—NCT02927184 and NCT04173065—have been registered and their results are eagerly awaited.

## 4.2.6. MGL-3196

MGL-3196 (2-(3,5-dichloro-4-(5-isopropyl-6-oxo-1,6-dihydropyridazin-3-yloxy) phenyl)-3,5-dioxo-2,3,4,5-tetrahydro (1,2,4) triazine-6-carbonitrile), also known as resmetirom, shows a ~28-fold selectivity for TR $\beta$  relative to TR $\alpha$ . MGL-3196 lacks adverse cardiac effect and is safe for the central thyroid axis. A study in 53 healthy volunteers showed that MGL-3196 was well-tolerated and decreased LDL cholesterol and TGs without causing adverse reactions [99]. In phase II clinical trials, MGL-3196 reduced LDL cholesterol, TG, and lipoprotein, and, most adverse events noted, including a higher incidence of transient diarrhea and nausea, were mild [100]. Another recent phase II clinical trials also supports the safety and efficacy of MGL-3196 [101], revealing a reduction in LDL cholesterol, apolipoprotein B, TGs, and markers of fibrosis. As of April 2022, there are four ongoing phase 3 trials (NCT03900429, NCT04643795, NCT04951219, and NCT04197479) of MGL-3196 that should provide important data.

## 4.2.7. 3,5,-L-diiodothyronine (T2)

T2, produced by deiodination of T3 by DIO2, shows ~50–100 times lower affinity for the THR than T3 [102]. T2 was previously considered a mere catabolite of T3, but it has since been demonstrated that T2 can mimic some T3 functions on lipid metabolism. In a HFD rat model, T2 was shown to reduce hepatic fat accumulation, increase the rate of  $\beta$ -oxidation, improve the oxidation-stress status of mitochondria, and reverse liver steatosis without causing notable adverse effects on the heart [103,104]. Subsequent studies revealed that T2 administration prevented HFD-induced insulin resistance [105]. In the same study, the authors suggested that, rather than acting on THR $\beta$ , T2 directly activates hepatic nuclear sirtuin 1 (SIRT1), which was previously shown to be capable of modulating lipid metabolism and enhancing mitochondrial activity [106]. In this context, it has been established that direct interactions of SIRT1 with THR $\beta$  modulates the expression of T3regulated hepatic genes [107,108]. T2 acts to regulate lipid metabolism in a different manner compared with T3. Specifically, T2 decreases lipogenesis and increases  $\beta$ -oxidation, whereas the actions of T3 primarily depend on the latter [109]. Although high doses of T2 were found to be associated with serious adverse heart effects [110], overall, most studies have shown beneficial effects of T2 on lipid metabolism, suggesting that T2 is worthy of further study for the prevention and treatment of diseases like obesity and NAFLD.

#### 4.2.8. 3-iodothyronamine (T1AM)

T1AM is another TH metabolite found in blood that is concentrated in some organs, such as the liver, brain, and muscle. Although its biosynthetic pathway is currently unclear, T1AM could theoretically be produced from T4 by enzymatic decarboxylation and deiodination [111]. T1AM mediates its functions through the trace amine-associated receptor (TAAR1) rather than the TR [112]. T1AM was first reported to be an antagonist of T3 [113]; however, recent studies have demonstrated that it induces antilipogenic effects in liver and muscle, and reduces hepatic TG and cholesterol levels in a polycystic ovary rat model [114]. In a spontaneously overweight mouse model, administration of T1AM was shown to increase lipolysis in association with significant weight loss [115]. Additional studies are needed to resolve controversies surrounding the functions of T1AM.

#### 4.3. Clinical Application for Thyroid Hormones Status and NAFLD

Thyroid dysfunctions strongly associated with NAFLD and a high prevalence of NAFLD in hypothyroidism patients compared to euthyroid individuals were demonstrated [70–73,77]. Similarly, the thyroid functions (usually assessed by detecting the levels of serum T3, T4, or TSH) were evaluated in NAFLD patients. However, the results were conflicting. Some studies are showing that circulating T3 or T4 was significantly decreased in NAFLD than that in non-NAFLD [116,117]. The prevalence of hypothyroidism was higher in patients with NAFLD compared to controls [71,118]. In contrast, some studies show inconsistent results [74,119]. Regardless of levels of THs, studies suggest NAFLD patients may acquire resistance to THs [120,121]. All this evidence suggest regulations and actions of THs are extremely complicated and it may be too hasty to conclude using TH replacements or mimetic as an intervention in NAFLD even though close associations between thyroid derangement and NAFLD were established. Fortunately, there are two trials (NCT01848171, and NCT03281083) that started to evaluate the benefits of levothyroxine, the standard TH replacement, on NAFLD in hypothyroidism patients and acquired encouraging results. In phase 4 randomized control trial (NCT01848171) for subclinical hypothyroidism patients revealed the prevalence of NAFLD was significantly reduced after treatment with levothyroxine [122]. Another phase 2b single-arm study (NCT03281083) for euthyroid patients with type 2 diabetes and steatosis revealed that administration of levothyroxine makes a decrease in 12% of baseline intrahepatic lipid content and a small decrease in body mass index, and visceral and subcutaneous adiposity [123]. These results suggest thyroid function should be considered when engaged with NAFLD patients, which may help to find the subpopulations suitable for adjuvant THs analogs therapy to effectively control or even reverse the progression of NAFLD.

Collectively, TH replacement, metabolites, and mimetics show promise for the treatment of NAFLD, however, the safely and efficacy of these compounds need further evaluation in well-designed studies.

#### 5. Conclusions and Perspectives

THs are among the most important hormones that regulate multiple physiological activities, including growth, development, and metabolism, throughout life. They remain the focus of research efforts, with numerous studies continuously attempting to more thoroughly understand their functions. Studies on the role of THs in lipid metabolism have revealed their systemic effects in addition to their involvement in intrahepatic pathways. Fully understanding the underlying mechanisms of THs' actions is challenging. Fortu-

nately, the "omics" field has reached a more mature stage and has become more accessible, providing a powerful tool for elucidating such complex issues. In previous decades, data from small-scale analyses of genes/proteins or comprehensive omics studies, including metabolome studies of blood biochemistry, have most frequently been used to investigate the effects of interventions. More recently, the microbiota/microbiome, especially the gut microbiota/microbiome, has been an emergent topic, with studies demonstrating that gut microbiota influence almost all organs of the body through bidirectional interactions. The microbiota is so thoroughly integrated into human physiology that it is considered an essential organ in humans [124]. Food intake, lifestyle, the microbiota, and human metabolism are all closely related and cannot be considered separately. Future trends for comprehensive studies of metabolic issues are likely to include using "big data" approaches that integrate information from genomic, transcriptomic, proteomic, and metabolomic studies, deep phenotyping, and investigations of the microbiota/microbiome and lifestyle. Such an approach may help unveil the link between thyroid dysfunction and diseases, such as NAFLD, and help to discover new strategies for the prevention, early diagnosis and treatment of NAFLD, and those that improve the prognosis of NAFLD patients. Although such big data applications are still in their infancy, some NAFLD-related studies using this approach have appeared in recent publications [125–127].

In addition to the biomarkers and genetic markers, medical imaging technologies have also made great strides in the diagnosis or monitoring of NAFLD. For example, magnetic resonance imaging is considered the gold standard for hepatic fat quantification and shows prominent accuracy to discriminate between different grades of fatty liver [128]. However, innate limitations of medical imaging should be considered when applied to the diagnosis or monitoring of NAFLD. The advantages and limitations of different medical imaging technologies have been summarized elsewhere [129,130]. In sum, the achievements of medical imaging technologies and molecular biologic research could complement each other and help to obtain the best benefit in basic research or clinical application for fatty liver diseases.

The development of thyromimetics and exploration of their potential in the prevention or treatment of NAFLD have continued for decades. Despite these efforts, compounds that combine efficacy and safety have not yet been found. Currently, the main strategy for improving efficacy and reducing adverse effects is to introduce modifications of the base in the backbone of endogenous T3 that increase TR $\beta$ - and liver-selectivity. However, the main limit to obtaining a TR $\beta$ -specific modification based on the native structure of T3 is the similarity of the ligand-binding domains of TR $\alpha$  and TR $\beta$ , which share ~82% homology [86]. In addition, as noted above, metabolism is a complicated systematic issue rather than an issue that involves a single organ. Even if a liver- and  $TR\beta$ -specific compound is found, extrahepatic factors might interfere with its functions in complicated and unforeseen ways, further increasing the difficulty of research design and limiting the validity and credibility of findings obtained using the current research model. Although many difficulties remain to be overcome, THs clearly have potential in the management of NAFLD and other metabolism-related diseases. Incorporating new research strategies, such as big data, into ongoing and future studies is expected to provide greater insight into the causative relationship between TH function and NAFLD and other related diseases.

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# References

- 1. Ortiga-Carvalho, T.M.; Sidhaye, A.R.; Wondisford, F.E. Thyroid hormone receptors and resistance to thyroid hormone disorders. *Nat. Rev. Endocrinol.* **2014**, *10*, 582–591. [CrossRef] [PubMed]
- Hernandez, A.; Martinez, M.E.; Ng, L.; Forrest, D. Thyroid Hormone Deiodinases: Dynamic Switches in Developmental Transitions. *Endocrinology* 2021, 162, bqab091. [CrossRef] [PubMed]
- 3. Lazar, M.A.; Berrodin, T.J.; Harding, H.P. Differential DNA binding by monomeric, homodimeric, and potentially heteromeric forms of the thyroid hormone receptor. *Mol. Cell. Biol.* **1991**, *11*, 5005–5015. [CrossRef] [PubMed]
- Berrodin, T.J.; Marks, M.S.; Ozato, K.; Linney, E.; Lazar, M.A. Heterodimerization among thyroid hormone receptor, retinoic acid receptor, retinoid X receptor, chicken ovalbumin upstream promoter transcription factor, and an endogenous liver protein. *Mol. Endocrinol.* 1992, 6, 1468–1478. [CrossRef] [PubMed]
- 5. Segal, J.; Ingbar, S.H. In vivo stimulation of sugar uptake in rat thymocytes. An extranuclear action of 3,5,3'-triiodothyronine. *J. Clin. Investig.* **1985**, *76*, 1575–1580. [CrossRef]
- Flamant, F.; Cheng, S.Y.; Hollenberg, A.N.; Moeller, L.C.; Samarut, J.; Wondisford, F.E.; Yen, P.M.; Refetoff, S. Thyroid Hormone Signaling Pathways: Time for a More Precise Nomenclature. *Endocrinology* 2017, 158, 2052–2057. [CrossRef]
- Perez, P.; Schönthal, A.; Aranda, A. Repression of c-fos gene expression by thyroid hormone and retinoic acid receptors. *J. Biol. Chem.* 1993, 268, 23538–23543. [CrossRef]
- 8. Méndez-Pertuz, M.; Sánchez-Pacheco, A.; Aranda, A. The thyroid hormone receptor antagonizes CREB-mediated transcription. *EMBO J.* **2003**, *22*, 3102–3112. [CrossRef]
- 9. Lasa, M.; Gil-Araujo, B.; Palafox, M.; Aranda, A. Thyroid hormone antagonizes tumor necrosis factor-alpha signaling in pituitary cells through the induction of dual specificity phosphatase 1. *Mol. Endocrinol.* **2010**, *24*, 412–422. [CrossRef]
- 10. Davis, P.J.; Davis, F.B.; Cody, V. Membrane receptors mediating thyroid hormone action. *Trends Endocrinol. Metab.* 2005, 16, 429–435. [CrossRef]
- 11. Hiroi, Y.; Kim, H.H.; Ying, H.; Furuya, F.; Huang, Z.; Simoncini, T.; Noma, K.; Ueki, K.; Nguyen, N.H.; Scanlan, T.S.; et al. Rapid nongenomic actions of thyroid hormone. *Proc. Natl. Acad. Sci. USA* **2006**, *103*, 14104–14109. [CrossRef] [PubMed]
- 12. Bergh, J.J.; Lin, H.Y.; Lansing, L.; Mohamed, S.N.; Davis, F.B.; Mousa, S.; Davis, P.J. Integrin alphaVbeta3 contains a cell surface receptor site for thyroid hormone that is linked to activation of mitogen-activated protein kinase and induction of angiogenesis. *Endocrinology* **2005**, *146*, 2864–2871. [CrossRef]
- 13. Davis, P.J.; Davis, F.B.; Mousa, S.A.; Luidens, M.K.; Lin, H.Y. Membrane receptor for thyroid hormone: Physiologic and pharmacologic implications. *Annu. Rev. Pharm. Toxicol.* **2011**, *51*, 99–115. [CrossRef] [PubMed]
- 14. Chi, H.C.; Chen, C.Y.; Tsai, M.M.; Tsai, C.Y.; Lin, K.H. Molecular functions of thyroid hormones and their clinical significance in liver-related diseases. *Biomed. Res. Int.* 2013, 2013, 601361. [CrossRef] [PubMed]
- 15. Mason, R.L.; Hunt, H.M.; Hurxthal, L. Blood Cholesterol Values in Hyperthyroidism and Hypothyroidism—Their Significance. *N. Engl. J. Med.* **1930**, 203, 1273–1278. [CrossRef]
- 16. Muls, E.; Rosseneu, M.; Blaton, V.; Lesaffre, E.; Lamberigts, G.; de Moor, P. Serum lipids and apolipoproteins A-I, A-II and B in primary hypothyroidism before and during treatment. *Eur. J. Clin. Investig.* **1984**, *14*, 12–15. [CrossRef]
- 17. Friis, T.; Pedersen, L.R. Serum lipids in hyper- and hypothyroidism before and after treatment. *Clin. Chim. Acta* **1987**, *162*, 155–163. [CrossRef]
- 18. Packard, C.J.; Shepherd, J.; Lindsay, G.M.; Gaw, A.; Taskinen, M.R. Thyroid replacement therapy and its influence on postheparin plasma lipases and apolipoprotein-B metabolism in hypothyroidism. *J. Clin. Endocrinol. Metab.* **1993**, *76*, 1209–1216. [CrossRef]
- 19. Wiseman, S.A.; Powell, J.T.; Humphries, S.E.; Press, M. The magnitude of the hypercholesterolemia of hypothyroidism is associated with variation in the low density lipoprotein receptor gene. *J. Clin. Endocrinol. Metab.* **1993**, *77*, 108–112. [CrossRef]
- 20. Pazos, F.; Alvarez, J.J.; Rubiés-Prat, J.; Varela, C.; Lasunción, M.A. Long-term thyroid replacement therapy and levels of lipoprotein(a) and other lipoproteins. *J. Clin. Endocrinol. Metab.* **1995**, *80*, 562–566. [CrossRef]
- 21. Araki, O.; Ying, H.; Zhu, X.G.; Willingham, M.C.; Cheng, S.Y. Distinct dysregulation of lipid metabolism by unliganded thyroid hormone receptor isoforms. *Mol. Endocrinol.* **2009**, *23*, 308–315. [CrossRef] [PubMed]
- 22. Grefhorst, A.; van de Peppel, I.P.; Larsen, L.E.; Jonker, J.W.; Holleboom, A.G. The Role of Lipophagy in the Development and Treatment of Non-Alcoholic Fatty Liver Disease. *Front. Endocrinol.* **2020**, *11*, 601627. [CrossRef] [PubMed]
- 23. Hajri, T.; Abumrad, N.A. Fatty acid transport across membranes: Relevance to nutrition and metabolic pathology. *Annu. Rev. Nutr.* **2002**, *22*, 383–415. [CrossRef] [PubMed]
- Klieverik, L.P.; Coomans, C.P.; Endert, E.; Sauerwein, H.P.; Havekes, L.M.; Voshol, P.J.; Rensen, P.C.; Romijn, J.A.; Kalsbeek, A.; Fliers, E. Thyroid hormone effects on whole-body energy homeostasis and tissue-specific fatty acid uptake in vivo. *Endocrinology* 2009, 150, 5639–5648. [CrossRef]
- Santana-Farré, R.; Mirecki-Garrido, M.; Bocos, C.; Henríquez-Hernández, L.A.; Kahlon, N.; Herrera, E.; Norstedt, G.; Parini, P.; Flores-Morales, A.; Fernández-Pérez, L. Influence of neonatal hypothyroidism on hepatic gene expression and lipid metabolism in adulthood. *PLoS ONE* 2012, 7, e37386. [CrossRef]

- Perra, A.; Simbula, G.; Simbula, M.; Pibiri, M.; Kowalik, M.A.; Sulas, P.; Cocco, M.T.; Ledda-Columbano, G.M.; Columbano, A. Thyroid hormone (T3) and TRbeta agonist GC-1 inhibit/reverse nonalcoholic fatty liver in rats. *FASEB J.* 2008, 22, 2981–2989. [CrossRef]
- Shu, L.; Hoo, R.L.C.; Wu, X.; Pan, Y.; Lee, I.P.C.; Cheong, L.Y.; Bornstein, S.R.; Rong, X.; Guo, J.; Xu, A. A-FABP mediates adaptive thermogenesis by promoting intracellular activation of thyroid hormones in brown adipocytes. *Nat. Commun.* 2017, *8*, 14147. [CrossRef]
- 28. Giudetti, A.M.; Leo, M.; Siculella, L.; Gnoni, G.V. Hypothyroidism down-regulates mitochondrial citrate carrier activity and expression in rat liver. *Biochim. Biophys. Acta* 2006, 1761, 484–491. [CrossRef]
- Siculella, L.; Sabetta, S.; Giudetti, A.M.; Gnoni, G.V. Hypothyroidism reduces tricarboxylate carrier activity and expression in rat liver mitochondria by reducing nuclear transcription rate and splicing efficiency. J. Biol. Chem. 2006, 281, 19072–19080. [CrossRef]
- 30. Paradies, G.; Ruggiero, F.M. Effect of hyperthyroidism on the transport of pyruvate in rat-heart mitochondria. *Biochim. Biophys. Acta* **1988**, *935*, 79–86. [CrossRef]
- 31. Paradies, G.; Ruggiero, F.M. Enhanced activity of the tricarboxylate carrier and modification of lipids in hepatic mitochondria from hyperthyroid rats. *Arch. Biochem. Biophys.* **1990**, *278*, 425–430. [CrossRef]
- 32. Spence, J.T.; Pitot, H.C.; Zalitis, G. Regulation of ATP-citrate lyase in primary cultures of adult rat hepatocytes. *J. Biol. Chem.* **1979**, 254, 12169–12173. [CrossRef]
- Gnoni, A.; Siculella, L.; Paglialonga, G.; Damiano, F.; Giudetti, A.M. 3,5-diiodo-L-thyronine increases de novo lipogenesis in liver from hypothyroid rats by SREBP-1 and ChREBP-mediated transcriptional mechanisms. *IUBMB Life* 2019, 71, 863–872. [CrossRef] [PubMed]
- 34. Moon, Y.-A.; Lee, J.-J.; Park, S.-W.; Ahn, Y.-H.; Kim, K.-S. The Roles of Sterol Regulatory Element-binding Proteins in the Transactivation of the Rat ATP Citrate-Lyase Promoter. *J. Biol. Chem.* **2000**, *275*, 30280–30286. [CrossRef]
- Hashimoto, K.; Yamada, M.; Matsumoto, S.; Monden, T.; Satoh, T.; Mori, M. Mouse Sterol Response Element Binding Protein-1c Gene Expression Is Negatively Regulated by Thyroid Hormone. *Endocrinology* 2006, 147, 4292–4302. [CrossRef]
- 36. Gnoni, G.V.; Rochira, A.; Leone, A.; Damiano, F.; Marsigliante, S.; Siculella, L. 3,5,3'triiodo-L-thyronine induces SREBP-1 expression by non-genomic actions in human HEP G2 cells. *J. Cell. Physiol.* **2012**, 227, 2388–2397. [CrossRef]
- 37. Zhang, Y.; Yin, L.; Hillgartner, F.B. SREBP-1 integrates the actions of thyroid hormone, insulin, cAMP, and medium-chain fatty acids on ACCalpha transcription in hepatocytes. *J. Lipid Res.* **2003**, *44*, 356–368. [CrossRef]
- Radenne, A.; Akpa, M.; Martel, C.; Sawadogo, S.; Mauvoisin, D.; Mounier, C. Hepatic regulation of fatty acid synthase by insulin and T3: Evidence for T3 genomic and nongenomic actions. *Am. J. Physiol.-Endocrinol. Metab.* 2008, 295, E884–E894. [CrossRef]
- Jump, D.B.; Thelen, A.P.; Mater, M.K. Functional Interaction between Sterol Regulatory Element-binding Protein-1c, Nuclear Factor Y, and 3,5,3'-Triiodothyronine Nuclear Receptors. J. Biol. Chem. 2001, 276, 34419–34427. [CrossRef]
- Brenta, G.; Berg, G.; Miksztowicz, V.; Lopez, G.; Lucero, D.; Faingold, C.; Murakami, M.; Machima, T.; Nakajima, K.; Schreier, L. Atherogenic Lipoproteins in Subclinical Hypothyroidism and Their Relationship with Hepatic Lipase Activity: Response to Replacement Treatment with Levothyroxine. *Thyroid* 2016, 26, 365–372. [CrossRef]
- Grasselli, E.; Voci, A.; Demori, I.; Vecchione, G.; Compalati, A.D.; Gallo, G.; Goglia, F.; De Matteis, R.; Silvestri, E.; Vergani, L. Triglyceride Mobilization from Lipid Droplets Sustains the Anti-Steatotic Action of Iodothyronines in Cultured Rat Hepatocytes. *Front. Physiol.* 2016, *6*, 418. [CrossRef] [PubMed]
- 42. Saggerson, E.D.; Carpenter, C.A. Carnitine palmitoyltransferase in liver and five extrahepatic tissues in the rat. Inhibition by dl-2-bromopalmitoyl-CoA and effect of hypothyroidism. *Biochem. J.* **1986**, *236*, 137–141. [CrossRef] [PubMed]
- Mynatt, R.L.; Park, E.A.; Thorngate, F.E.; Das, H.K.; Cook, G.A. Changes in carnitine palmitoyltransferase-I mRNA abundance produced by hyperthyroidism and hypothyroidism parallel changes in activity. *Biochem. Biophys. Res. Commun.* 1994, 201, 932–937. [CrossRef] [PubMed]
- Jackson-Hayes, L.; Song, S.; Lavrentyev, E.N.; Jansen, M.S.; Hillgartner, F.B.; Tian, L.; Wood, P.A.; Cook, G.A.; Park, E.A. A thyroid hormone response unit formed between the promoter and first intron of the carnitine palmitoyltransferase-Ialpha gene mediates the liver-specific induction by thyroid hormone. *J. Biol. Chem.* 2003, 278, 7964–7972. [CrossRef]
- 45. Potthoff Matthew, J.; Inagaki, T.; Satapati, S.; Ding, X.; He, T.; Goetz, R.; Mohammadi, M.; Finck Brian, N.; Mangelsdorf David, J.; Kliewer Steven, A.; et al. FGF21 induces PGC-1α and regulates carbohydrate and fatty acid metabolism during the adaptive starvation response. *Proc. Natl. Acad. Sci. USA* **2009**, *106*, 10853–10858. [CrossRef]
- 46. Kharitonenkov, A.; Shiyanova, T.L.; Koester, A.; Ford, A.M.; Micanovic, R.; Galbreath, E.J.; Sandusky, G.E.; Hammond, L.J.; Moyers, J.S.; Owens, R.A.; et al. FGF-21 as a novel metabolic regulator. *J. Clin. Investig.* **2005**, *115*, 1627–1635. [CrossRef]
- 47. Yu, K.C.; Chen, W.; Cooper, A.D. LDL receptor-related protein mediates cell-surface clustering and hepatic sequestration of chylomicron remnants in LDLR-deficient mice. *J. Clin. Investig.* **2001**, 107, 1387–1394. [CrossRef]
- 48. Lopez, D.; Abisambra Socarrás, J.F.; Bedi, M.; Ness, G.C. Activation of the hepatic LDL receptor promoter by thyroid hormone. *Biochim. Biophys. Acta* 2007, 1771, 1216–1225. [CrossRef]
- Moon, J.H.; Kim, H.J.; Kim, H.M.; Choi, S.H.; Lim, S.; Park, Y.J.; Jang, H.C.; Cha, B.S. Decreased expression of hepatic low-density lipoprotein receptor-related protein 1 in hypothyroidism: A novel mechanism of atherogenic dyslipidemia in hypothyroidism. *Thyroid* 2013, 23, 1057–1065. [CrossRef]
- Shin, D.J.; Osborne, T.F. Thyroid hormone regulation and cholesterol metabolism are connected through Sterol Regulatory Element-Binding Protein-2 (SREBP-2). J. Biol. Chem. 2003, 278, 34114–34118. [CrossRef]

- Alborn, W.E.; Cao, G.; Careskey, H.E.; Qian, Y.-W.; Subramaniam, D.R.; Davies, J.; Conner, E.M.; Konrad, R.J. Serum Proprotein Convertase Subtilisin Kexin Type 9 Is Correlated Directly with Serum LDL Cholesterol. *Clin. Chem.* 2007, 53, 1814–1819. [CrossRef] [PubMed]
- 52. Fazaeli, M.; Khoshdel, A.; Shafiepour, M.; Rohban, M. The influence of subclinical hypothyroidism on serum lipid profile, PCSK9 levels and CD36 expression on monocytes. *Diabetes Metab. Syndr. Clin. Res. Rev.* **2019**, *13*, 312–316. [CrossRef] [PubMed]
- Yildirim, A.M.; Koca, A.O.; Beyan, E.; Dogan, O.; Karakaya, S.; Aksoz, Z.; Ertuğrul, D.T. Association of serum proprotein convertase Subtilisin/Kexin Type 9 (PCSK9) level with thyroid function disorders. *Eur. Rev. Med. Pharm. Sci.* 2021, 25, 5511–5517. [CrossRef]
- Lin, J.Z.; Martagón, A.J.; Hsueh, W.A.; Baxter, J.D.; Gustafsson, J.-Å.; Webb, P.; Phillips, K.J. Thyroid Hormone Receptor Agonists Reduce Serum Cholesterol Independent of the LDL Receptor. *Endocrinology* 2012, 153, 6136–6144. [CrossRef] [PubMed]
- 55. Lammel Lindemann, J.A.; Angajala, A.; Engler, D.A.; Webb, P.; Ayers, S.D. Thyroid hormone induction of human cholesterol 7 alpha-hydroxylase (Cyp7a1) in vitro. *Mol. Cell. Endocrinol.* **2014**, *388*, 32–40. [CrossRef]
- 56. Gälman, C.; Bonde, Y.; Matasconi, M.; Angelin, B.; Rudling, M. Dramatically increased intestinal absorption of cholesterol following hypophysectomy is normalized by thyroid hormone. *Gastroenterology* **2008**, *134*, 1127–1136. [CrossRef]
- Graf, G.A.; Yu, L.; Li, W.P.; Gerard, R.; Tuma, P.L.; Cohen, J.C.; Hobbs, H.H. ABCG5 and ABCG8 are obligate heterodimers for protein trafficking and biliary cholesterol excretion. *J. Biol. Chem.* 2003, 278, 48275–48282. [CrossRef]
- 58. Bonde, Y.; Plösch, T.; Kuipers, F.; Angelin, B.; Rudling, M. Stimulation of murine biliary cholesterol secretion by thyroid hormone is dependent on a functional ABCG5/G8 complex. *Hepatology* **2012**, *56*, 1828–1837. [CrossRef]
- 59. Tao, R.; Xiong, X.; DePinho, R.A.; Deng, C.-X.; Dong, X.C. Hepatic SREBP-2 and cholesterol biosynthesis are regulated by FoxO3 and Sirt6. *J. Lipid Res.* 2013, *54*, 2745–2753. [CrossRef]
- 60. Markovic, M.; Ben-Shabat, S.; Aponick, A.; Zimmermann, E.M.; Dahan, A. Lipids and Lipid-Processing Pathways in Drug Delivery and Therapeutics. *Int. J. Mol. Sci.* 2020, *21*, 3248. [CrossRef]
- 61. Tang, Q.-Q. Lipid metabolism and diseases. Sci. Bull. 2016, 61, 1471–1472. [CrossRef]
- 62. Powell, E.E.; Wong, V.W.; Rinella, M. Non-alcoholic fatty liver disease. Lancet 2021, 397, 2212–2224. [CrossRef]
- Chalasani, N.; Younossi, Z.; Lavine, J.E.; Charlton, M.; Cusi, K.; Rinella, M.; Harrison, S.A.; Brunt, E.M.; Sanyal, A.J. The diagnosis and management of nonalcoholic fatty liver disease: Practice guidance from the American Association for the Study of Liver Diseases. *Hepatology* 2018, 67, 328–357. [CrossRef] [PubMed]
- 64. Estes, C.; Razavi, H.; Loomba, R.; Younossi, Z.; Sanyal, A.J. Modeling the epidemic of nonalcoholic fatty liver disease demonstrates an exponential increase in burden of disease. *Hepatology* **2018**, *67*, 123–133. [CrossRef] [PubMed]
- Kleiner, D.E.; Brunt, E.M.; Van Natta, M.; Behling, C.; Contos, M.J.; Cummings, O.W.; Ferrell, L.D.; Liu, Y.C.; Torbenson, M.S.; Unalp-Arida, A.; et al. Design and validation of a histological scoring system for nonalcoholic fatty liver disease. *Hepatology* 2005, 41, 1313–1321. [CrossRef]
- Paik, J.M.; Golabi, P.; Younossi, Y.; Mishra, A.; Younossi, Z.M. Changes in the Global Burden of Chronic Liver Diseases from 2012 to 2017: The Growing Impact of NAFLD. *Hepatology* 2020, 72, 1605–1616. [CrossRef]
- Goldberg, D.; Ditah, I.C.; Saeian, K.; Lalehzari, M.; Aronsohn, A.; Gorospe, E.C.; Charlton, M. Changes in the Prevalence of Hepatitis C Virus Infection, Nonalcoholic Steatohepatitis, and Alcoholic Liver Disease Among Patients with Cirrhosis or Liver Failure on the Waitlist for Liver Transplantation. *Gastroenterology* 2017, 152, 1090–1099.e1. [CrossRef]
- Lonardo, A.; Nascimbeni, F.; Mantovani, A.; Targher, G. Hypertension, diabetes, atherosclerosis and NASH: Cause or consequence? J. Hepatol. 2018, 68, 335–352. [CrossRef]
- 69. Al-Muzafar, H.M.; Alshehri, F.S.; Amin, K.A. The role of pioglitazone in antioxidant, anti-inflammatory, and insulin sensitivity in a high fat-carbohydrate diet-induced rat model of insulin resistance. *Braz. J. Med. Biol. Res.* **2021**, *54*, e10782. [CrossRef]
- Chung, G.E.; Kim, D.; Kim, W.; Yim, J.Y.; Park, M.J.; Kim, Y.J.; Yoon, J.H.; Lee, H.S. Non-alcoholic fatty liver disease across the spectrum of hypothyroidism. *J. Hepatol.* 2012, *57*, 150–156. [CrossRef]
- Pagadala, M.R.; Zein, C.O.; Dasarathy, S.; Yerian, L.M.; Lopez, R.; McCullough, A.J. Prevalence of hypothyroidism in nonalcoholic fatty liver disease. *Dig. Dis. Sci.* 2012, 57, 528–534. [CrossRef]
- 72. Bano, A.; Chaker, L.; Plompen, E.P.; Hofman, A.; Dehghan, A.; Franco, O.H.; Janssen, H.L.; Darwish Murad, S.; Peeters, R.P. Thyroid Function and the Risk of Nonalcoholic Fatty Liver Disease: The Rotterdam Study. J. Clin. Endocrinol. Metab. 2016, 101, 3204–3211. [CrossRef] [PubMed]
- 73. Manka, P.; Bechmann, L.; Best, J.; Sydor, S.; Claridge, L.C.; Coombes, J.D.; Canbay, A.; Moeller, L.; Gerken, G.; Wedemeyer, H.; et al. Low Free Triiodothyronine Is Associated with Advanced Fibrosis in Patients at High Risk for Nonalcoholic Steatohepatitis. *Dig. Dis. Sci.* **2019**, *64*, 2351–2358. [CrossRef] [PubMed]
- 74. Eshraghian, A.; Dabbaghmanesh, M.H.; Eshraghian, H.; Fattahi, M.R.; Omrani, G.R. Nonalcoholic fatty liver disease in a cluster of Iranian population: Thyroid status and metabolic risk factors. *Arch. Iran. Med.* **2013**, *16*, 584–589. [PubMed]
- 75. Posadas-Romero, C.; Jorge-Galarza, E.; Posadas-Sánchez, R.; Acuña-Valerio, J.; Juárez-Rojas, J.G.; Kimura-Hayama, E.; Medina-Urrutia, A.; Cardoso-Saldaña, G.C. Fatty liver largely explains associations of subclinical hypothyroidism with insulin resistance, metabolic syndrome, and subclinical coronary atherosclerosis. *Eur. J. Endocrinol.* 2014, 171, 319–325. [CrossRef]
- Jaruvongvanich, V.; Sanguankeo, A.; Upala, S. Nonalcoholic Fatty Liver Disease Is Not Associated with Thyroid Hormone Levels and Hypothyroidism: A Systematic Review and Meta-Analysis. *Eur. Thyroid. J.* 2017, *6*, 208–215. [CrossRef] [PubMed]

- 77. Mantovani, A.; Nascimbeni, F.; Lonardo, A.; Zoppini, G.; Bonora, E.; Mantzoros, C.S.; Targher, G. Association Between Primary Hypothyroidism and Nonalcoholic Fatty Liver Disease: A Systematic Review and Meta-Analysis. *Thyroid* 2018, 28, 1270–1284. [CrossRef]
- 78. Lonardo, A.; Ballestri, S.; Mantovani, A.; Nascimbeni, F.; Lugari, S.; Targher, G. Pathogenesis of hypothyroidism-induced NAFLD: Evidence for a distinct disease entity? *Dig. Liver. Dis.* **2019**, *51*, 462–470. [CrossRef]
- 79. Romero-Gómez, M.; Zelber-Sagi, S.; Trenell, M. Treatment of NAFLD with diet, physical activity and exercise. J. Hepatol. 2017, 67, 829–846. [CrossRef]
- 80. Głuszyńska, P.; Lemancewicz, D.; Dzięcioł, J.B.; Razak Hady, H. Non-Alcoholic Fatty Liver Disease (NAFLD) and Bariatric/Metabolic Surgery as Its Treatment Option: A Review. J. Clin. Med. 2021, 10, 5721. [CrossRef]
- 81. Bril, F.; Biernacki, D.M.; Kalavalapalli, S.; Lomonaco, R.; Subbarayan, S.K.; Lai, J.; Tio, F.; Suman, A.; Orsak, B.K.; Hecht, J.; et al. Role of Vitamin E for Nonalcoholic Steatohepatitis in Patients with Type 2 Diabetes: A Randomized Controlled Trial. *Diabetes Care* 2019, 42, 1481–1488. [CrossRef] [PubMed]
- Cable, E.E.; Finn, P.D.; Stebbins, J.W.; Hou, J.; Ito, B.R.; van Poelje, P.D.; Linemeyer, D.L.; Erion, M.D. Reduction of hepatic steatosis in rats and mice after treatment with a liver-targeted thyroid hormone receptor agonist. *Hepatology* 2009, 49, 407–417. [CrossRef] [PubMed]
- 83. Iannucci, L.F.; Cioffi, F.; Senese, R.; Goglia, F.; Lanni, A.; Yen, P.M.; Sinha, R.A. Metabolomic analysis shows differential hepatic effects of T2 and T3 in rats after short-term feeding with high fat diet. *Sci. Rep.* **2017**, *7*, 2023. [CrossRef] [PubMed]
- 84. Kowalik, M.A.; Columbano, A.; Perra, A. Thyroid Hormones, Thyromimetics and Their Metabolites in the Treatment of Liver Disease. *Front. Endocrinol.* 2018, *9*, 382. [CrossRef] [PubMed]
- 85. Ritter, M.J.; Amano, I.; Hollenberg, A.N. Thyroid Hormone Signaling and the Liver. Hepatology 2020, 72, 742–752. [CrossRef]
- 86. Chiellini, G.; Apriletti, J.W.; Yoshihara, H.A.; Baxter, J.D.; Ribeiro, R.C.J.; Scanlan, T.S. A high-affinity subtype-selective agonist ligand for the thyroid hormone receptor. *Chem. Biol.* **1998**, *5*, 299–306. [CrossRef]
- Trost, S.U.; Swanson, E.; Gloss, B.; Wang-Iverson, D.B.; Zhang, H.; Volodarsky, T.; Grover, G.J.; Baxter, J.D.; Chiellini, G.; Scanlan, T.S.; et al. The Thyroid Hormone Receptor-β-Selective Agonist GC-1 Differentially Affects Plasma Lipids and Cardiac Activity. *Endocrinology* 2000, 141, 3057–3064. [CrossRef]
- Martagón, A.J.; Lin, J.Z.; Cimini, S.L.; Webb, P.; Phillips, K.J. The amelioration of hepatic steatosis by thyroid hormone receptor agonists is insufficient to restore insulin sensitivity in ob/ob mice. *PLoS ONE* 2015, 10, e0122987. [CrossRef]
- Vatner, D.F.; Weismann, D.; Beddow, S.A.; Kumashiro, N.; Erion, D.M.; Liao, X.H.; Grover, G.J.; Webb, P.; Phillips, K.J.; Weiss, R.E.; et al. Thyroid hormone receptor-β agonists prevent hepatic steatosis in fat-fed rats but impair insulin sensitivity via discrete pathways. *Am. J. Physiol. Endocrinol. Metab.* 2013, 305, E89–E100. [CrossRef]
- Borngraeber, S.; Budny, M.J.; Chiellini, G.; Cunha-Lima, S.T.; Togashi, M.; Webb, P.; Baxter, J.D.; Scanlan, T.S.; Fletterick, R.J. Ligand selectivity by seeking hydrophobicity in thyroid hormone receptor. *Proc. Natl. Acad. Sci. USA* 2003, 100, 15358–15363. [CrossRef]
- Amorim, B.S.; Ueta, C.B.; Freitas, B.C.; Nassif, R.J.; Gouveia, C.H.; Christoffolete, M.A.; Moriscot, A.S.; Lancelloti, C.L.; Llimona, F.; Barbeiro, H.V.; et al. A TRbeta-selective agonist confers resistance to diet-induced obesity. *J. Endocrinol.* 2009, 203, 291–299. [CrossRef] [PubMed]
- 92. Grijota-Martínez, C.; Samarut, E.; Scanlan, T.S.; Morte, B.; Bernal, J. In Vivo Activity of the Thyroid Hormone Receptor β- and α-Selective Agonists GC-24 and CO23 on Rat Liver, Heart, and Brain. *Endocrinology* 2011, 152, 1136–1142. [CrossRef] [PubMed]
- 93. Grover, G.J.; Mellström, K.; Ye, L.; Malm, J.; Li, Y.L.; Bladh, L.G.; Sleph, P.G.; Smith, M.A.; George, R.; Vennström, B.; et al. Selective thyroid hormone receptor-beta activation: A strategy for reduction of weight, cholesterol, and lipoprotein (a) with reduced cardiovascular liability. *Proc. Natl. Acad. Sci. USA* 2003, *100*, 10067–10072. [CrossRef] [PubMed]
- Grover, G.J.; Mellstrom, K.; Malm, J. Development of the thyroid hormone receptor beta-subtype agonist KB-141: A strategy for body weight reduction and lipid lowering with minimal cardiac side effects. *Cardiovasc. Drug Rev.* 2005, 23, 133–148. [CrossRef] [PubMed]
- 95. Berkenstam, A.; Kristensen, J.; Mellström, K.; Carlsson, B.; Malm, J.; Rehnmark, S.; Garg, N.; Andersson Carl, M.; Rudling, M.; Sjöberg, F.; et al. The thyroid hormone mimetic compound KB2115 lowers plasma LDL cholesterol and stimulates bile acid synthesis without cardiac effects in humans. *Proc. Natl. Acad. Sci. USA* 2008, 105, 663–667. [CrossRef]
- Sjouke, B.; Langslet, G.; Ceska, R.; Nicholls, S.J.; Nissen, S.E.; Öhlander, M.; Ladenson, P.W.; Olsson, A.G.; Hovingh, G.K.; Kastelein, J.J. Eprotirome in patients with familial hypercholesterolaemia (the AKKA trial): A randomised, double-blind, placebo-controlled phase 3 study. *Lancet Diabetes Endocrinol.* 2014, 2, 455–463. [CrossRef]
- 97. Ladenson, P.W.; Kristensen, J.D.; Ridgway, E.C.; Olsson, A.G.; Carlsson, B.; Klein, I.; Baxter, J.D.; Angelin, B. Use of the Thyroid Hormone Analogue Eprotirome in Statin-Treated Dyslipidemia. *N. Engl. J. Med.* **2010**, *362*, 906–916. [CrossRef]
- Erion, M.D.; Cable, E.E.; Ito, B.R.; Jiang, H.; Fujitaki, J.M.; Finn, P.D.; Zhang, B.H.; Hou, J.; Boyer, S.H.; van Poelje, P.D.; et al. Targeting thyroid hormone receptor-beta agonists to the liver reduces cholesterol and triglycerides and improves the therapeutic index. *Proc. Natl. Acad. Sci. USA* 2007, 104, 15490–15495. [CrossRef]
- Kelly, M.J.; Pietranico-Cole, S.; Larigan, J.D.; Haynes, N.-E.; Reynolds, C.H.; Scott, N.; Vermeulen, J.; Dvorozniak, M.; Conde-Knape, K.; Huang, K.-S.; et al. Discovery of 2-[3,5-Dichloro-4-(5-isopropyl-6-oxo-1,6-dihydropyridazin-3-yloxy)phenyl]-3,5-dioxo-2,3,4,5-tetrahydro[1,2,4]triazine-6-carbonitrile (MGL-3196), a Highly Selective Thyroid Hormone Receptor β Agonist in Clinical Trials for the Treatment of Dyslipidemia. *J. Med. Chem.* 2014, *57*, 3912–3923. [CrossRef]

- Harrison, S.A.; Bashir, M.R.; Guy, C.D.; Zhou, R.; Moylan, C.A.; Frias, J.P.; Alkhouri, N.; Bansal, M.B.; Baum, S.; Neuschwander-Tetri, B.A.; et al. Resmetirom (MGL-3196) for the treatment of non-alcoholic steatohepatitis: A multicentre, randomised, double-blind, placebo-controlled, phase 2 trial. *Lancet* 2019, 394, 2012–2024. [CrossRef]
- 101. Harrison, S.A.; Bashir, M.; Moussa, S.E.; McCarty, K.; Pablo Frias, J.; Taub, R.; Alkhouri, N. Effects of Resmetirom on Noninvasive Endpoints in a 36-Week Phase 2 Active Treatment Extension Study in Patients with NASH. *Hepatol. Commun.* 2021, *5*, 573–588. [CrossRef] [PubMed]
- 102. Ball, S.G.; Sokolov, J.; Chin, W.W. 3,5-Diiodo-L-thyronine (T2) has selective thyromimetic effects in vivo and in vitro. *J. Mol. Endocrinol.* **1997**, *19*, 137–147. [CrossRef] [PubMed]
- 103. Mollica, M.P.; Lionetti, L.; Moreno, M.; Lombardi, A.; De Lange, P.; Antonelli, A.; Lanni, A.; Cavaliere, G.; Barletta, A.; Goglia, F. 3,5-diiodo-l-thyronine, by modulating mitochondrial functions, reverses hepatic fat accumulation in rats fed a high-fat diet. *J. Hepatol.* 2009, *51*, 363–370. [CrossRef] [PubMed]
- 104. Lanni, A.; Moreno, M.; Lombardi, A.; de Lange, P.; Silvestri, E.; Ragni, M.; Farina, P.; Baccari, G.C.; Fallahi, P.; Antonelli, A.; et al. 3,5-diiodo-L-thyronine powerfully reduces adiposity in rats by increasing the burning of fats. *FASEB J.* 2005, 19, 1552–1554. [CrossRef] [PubMed]
- 105. de Lange, P.; Cioffi, F.; Senese, R.; Moreno, M.; Lombardi, A.; Silvestri, E.; De Matteis, R.; Lionetti, L.; Mollica, M.P.; Goglia, F.; et al. Nonthyrotoxic prevention of diet-induced insulin resistance by 3,5-diiodo-L-thyronine in rats. *Diabetes* 2011, 60, 2730–2739. [CrossRef]
- 106. Michan, S.; Sinclair, D. Sirtuins in mammals: Insights into their biological function. Biochem. J. 2007, 404, 1–13. [CrossRef]
- 107. Thakran, S.; Sharma, P.; Attia, R.R.; Hori, R.T.; Deng, X.; Elam, M.B.; Park, E.A. Role of sirtuin 1 in the regulation of hepatic gene expression by thyroid hormone. *J. Biol. Chem.* 2013, 288, 807–818. [CrossRef]
- 108. Suh, J.H.; Sieglaff, D.H.; Zhang, A.; Xia, X.; Cvoro, A.; Winnier, G.E.; Webb, P. SIRT1 is a direct coactivator of thyroid hormone receptor β1 with gene-specific actions. *PLoS ONE* **2013**, *8*, e70097. [CrossRef]
- Senese, R.; Cioffi, F.; de Lange, P.; Leanza, C.; Iannucci, L.F.; Silvestri, E.; Moreno, M.; Lombardi, A.; Goglia, F.; Lanni, A. Both 3,5-Diiodo-L-Thyronine and 3,5,3'-Triiodo-L-Thyronine Prevent Short-term Hepatic Lipid Accumulation via Distinct Mechanisms in Rats Being Fed a High-Fat Diet. *Front. Physiol.* 2017, *8*, 706. [CrossRef]
- Jonas, W.; Lietzow, J.; Wohlgemuth, F.; Hoefig, C.S.; Wiedmer, P.; Schweizer, U.; Köhrle, J.; Schürmann, A. 3,5-Diiodo-L-Thyronine (3,5-T2) Exerts Thyromimetic Effects on Hypothalamus-Pituitary-Thyroid Axis, Body Composition, and Energy Metabolism in Male Diet-Induced Obese Mice. *Endocrinology* 2015, 156, 389–399. [CrossRef]
- 111. Scanlan, T.S. Minireview: 3-Iodothyronamine (T1AM): A new player on the thyroid endocrine team? *Endocrinology* **2009**, *150*, 1108–1111. [CrossRef]
- 112. Zucchi, R.; Chiellini, G.; Scanlan, T.S.; Grandy, D.K. Trace amine-associated receptors and their ligands. *Br. J. Pharmacol.* 2006, 149, 967–978. [CrossRef] [PubMed]
- 113. Scanlan, T.S.; Suchland, K.L.; Hart, M.E.; Chiellini, G.; Huang, Y.; Kruzich, P.J.; Frascarelli, S.; Crossley, D.A.; Bunzow, J.R.; Ronca-Testoni, S.; et al. 3-Iodothyronamine is an endogenous and rapid-acting derivative of thyroid hormone. *Nat. Med.* 2004, 10, 638–642. [CrossRef] [PubMed]
- 114. Selen Alpergin, E.S.; Bolandnazar, Z.; Sabatini, M.; Rogowski, M.; Chiellini, G.; Zucchi, R.; Assadi-Porter, F.M. Metabolic profiling reveals reprogramming of lipid metabolic pathways in treatment of polycystic ovary syndrome with 3-iodothyronamine. *Physiol. Rep.* **2017**, *5*, e13097. [CrossRef]
- 115. Haviland, J.A.; Reiland, H.; Butz, D.E.; Tonelli, M.; Porter, W.P.; Zucchi, R.; Scanlan, T.S.; Chiellini, G.; Assadi-Porter, F.M. NMR-based metabolomics and breath studies show lipid and protein catabolism during low dose chronic T(1)AM treatment. *Obesity* 2013, 21, 2538–2544. [CrossRef] [PubMed]
- 116. Xu, C.; Xu, L.; Yu, C.; Miao, M.; Li, Y. Association between thyroid function and nonalcoholic fatty liver disease in euthyroid elderly Chinese. *Clin. Endocrinol.* **2011**, *75*, 240–246. [CrossRef]
- 117. Wang, B.; Wang, B.; Yang, Y.; Xu, J.; Hong, M.; Xia, M.; Li, X.; Gao, X. Thyroid function and non-alcoholic fatty liver disease in hyperthyroidism patients. *BMC Endocr. Disord.* 2021, 21, 27. [CrossRef] [PubMed]
- 118. Liangpunsakul, S.; Chalasani, N. Is hypothyroidism a risk factor for non-alcoholic steatohepatitis? *J. Clin. Gastroenterol.* **2003**, 37, 340–343. [CrossRef]
- 119. Mazo, D.F.; Lima, V.M.; Stefano, J.T.; Rabelo, F.; Faintuch, J.; Oliveira, C.P. Gluco-lipidic indices in treated hypothyroidism associated with nonalcoholic fatty liver disease. *Arq. Gastroenterol.* **2011**, *48*, 186–189. [CrossRef]
- 120. Krause, C.; Grohs, M.; El Gammal, A.T.; Wolter, S.; Lehnert, H.; Mann, O.; Mittag, J.; Kirchner, H. Reduced expression of thyroid hormone receptor β in human nonalcoholic steatohepatitis. *Endocr. Connect.* **2018**, *7*, 1448–1456. [CrossRef]
- Lai, S.; Li, J.; Wang, Z.; Wang, W.; Guan, H. Sensitivity to Thyroid Hormone Indices Are Closely Associated with NAFLD. Front. Endocrinol. 2021, 12, 766419. [CrossRef] [PubMed]
- 122. Liu, L.; Yu, Y.; Zhao, M.; Zheng, D.; Zhang, X.; Guan, Q.; Xu, C.; Gao, L.; Zhao, J.; Zhang, H. Benefits of Levothyroxine Replacement Therapy on Nonalcoholic Fatty Liver Disease in Subclinical Hypothyroidism Patients. *Int. J. Endocrinol.* 2017, 2017, 5753039. [CrossRef] [PubMed]
- 123. Bruinstroop, E.; Dalan, R.; Cao, Y.; Bee, Y.M.; Chandran, K.; Cho, L.W.; Soh, S.B.; Teo, E.K.; Toh, S.-A.; Leow, M.K.S.; et al. Low-Dose Levothyroxine Reduces Intrahepatic Lipid Content in Patients with Type 2 Diabetes Mellitus and NAFLD. *J. Clin. Endocrinol. Metab.* 2018, 103, 2698–2706. [CrossRef] [PubMed]

- 124. Eckburg, P.B.; Bik, E.M.; Bernstein, C.N.; Purdom, E.; Dethlefsen, L.; Sargent, M.; Gill, S.R.; Nelson, K.E.; Relman, D.A. Diversity of the human intestinal microbial flora. *Science* 2005, *308*, 1635–1638. [CrossRef]
- 125. Goossens, N.; Jornayvaz, F.R. Translational Aspects of Diet and Non-Alcoholic Fatty Liver Disease. *Nutrients* **2017**, *9*, 1077. [CrossRef]
- 126. Schattenberg, J.M.; Ekstedt, M. Assessing the disease burden of non-alcoholic fatty liver disease in the real world—Big data and big numbers. *BMC Med.* **2019**, *17*, 123. [CrossRef]
- 127. Al Taii, H.; Yaqoob, Z.; Al-Kindi, S.G. Underutilization of Statin Therapy Among Patients with NAFLD in the USA: Validation with Big Data. *Dig. Dis. Sci.* 2016, *61*, 2760. [CrossRef]
- Roldan-Valadez, E.; Favila, R.; Martínez-López, M.; Uribe, M.; Ríos, C.; Méndez-Sánchez, N. In vivo 3T spectroscopic quantification of liver fat content in nonalcoholic fatty liver disease: Correlation with biochemical method and morphometry. *J Hepatol.* 2010, 53, 732–737. [CrossRef]
- Roldan-Valadez, E.; Favila, R.; Martínez-López, M.; Uribe, M.; Méndez-Sánchez, N. Imaging techniques for assessing hepatic fat content in nonalcoholic fatty liver disease. *Ann. Hepatol.* 2008, 7, 212–220. [CrossRef]
- Wong, V.W.; Adams, L.A.; de Lédinghen, V.; Wong, G.L.; Sookoian, S. Noninvasive biomarkers in NAFLD and NASH—Current progress and future promise. *Nat. Rev. Gastroenterol. Hepatol.* 2018, 15, 461–478. [CrossRef]