

Review



NADPH Oxidase and Epidermal Growth Factor Receptor Are Promising Targets of Phytochemicals for Ultraviolet-Induced Skin Carcinogenesis

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Copyright: © 2021 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (https:// creativecommons.org/licenses/by/ 4.0/). **Abstract:** The skin acts as the primary defense organ that protects the body from the external environment. Skin cancer is one of the most common cancers in the world. Skin carcinogenesis is usually caused by cell degeneration due to exposure to ultraviolet (UV) radiation, which causes changes in various signaling networks, disrupting the homeostasis of single skin cells. In this review, we summarize the roles of nicotinamide adenine dinucleotide phosphate oxidase (NOX) and epidermal growth factor receptor (EGFR) in UV-induced skin carcinogenesis. Furthermore, we describe the crosstalk that exists between NOX, EGFR, and protein tyrosine phosphatase κ and its oncogenic downstream signaling pathways. Chemoprevention is the use of chemical compounds to recover the healthy status of the skin or delay cancer development. Current evidence from in vitro and in vivo studies on chemopreventive phytochemicals that target NOX, EGFR, or both, as major regulators of skin carcinogenesis will also be discussed.

Keywords: nicotinamide adenine dinucleotide phosphate oxidase (NADPH oxidase); epidermal growth factor receptor (EGFR); protein tyrosine phosphatase κ (PTPκ); reactive oxygen species (ROS)

1. Introduction

Based on recent cancer statistics, 10 million people died of skin cancer (9.9 million excluding nonmelanoma skin cancers [NMSCs]) in 2020 [1]. The most frequently diagnosed types of skin cancers, collectively known as NMSCs, are basal cell carcinoma (BCC) and squamous cell carcinoma (SCC) [2]. While NMSC is the most frequently detected skin malignancy, which accounts for 40% of all cancers diagnosed in the United States [3], malignant melanomas are detected in only 4% of all skin cancers [4]. BCCs and SCCs develop from within the basal layer of the epidermis. While BCCs have slow growth and rarely metastasize, SCCs are more invasive and metastasize 10% of the time [5,6].

Ultraviolet (UV) light, inevitably met in daily environments and substantially affecting skin homeostasis, is one of the factors that can harm skin health. Epidemiological studies have proven that solar UV irradiation is the most important environmental carcinogen contributing to skin cancer development [7]. UVC is absorbed in the stratospheric ozone. By contrast, UVB (1–10%) and UVA (90–99%) reach the surface of the earth and ultimately cause DNA damage, erythema, sunburn, immunosuppression, and skin cancer in humans [8]. The UVA irradiation to which we are exposed appears to be weakly carcinogenic and causes photoaging and wrinkling of the skin. On the other hand, UVB is recognized as a complete carcinogen relevant to human skin cancer [9].

Prolonged exposure to UV impairs skin homeostasis and stimulates responses to environmental changes, including the production of reactive oxygen species (ROS). These responses lead to the activation of multiple signaling cascades, which switch on the expression of oncogenic and tumor suppressor genes. A number of studies have proven that antioxidants contribute to the prevention of skin carcinogenesis. However, a part of phytochemicals with low antioxidant capacity demonstrates relatively high chemopreventive effects. Furthermore, several compounds, including vitamin D and lycopene, can act as pro-oxidants. We reported in various studies that phytochemicals can prevent cancer by acting as inhibitors of the oncogenic molecules, Fyn, Src, aryl hydrocarbon receptor (AhR), phosphoinositide 3-kinase (PI3K), Raf, and extracellular signal-regulated kinase (ERK)2 both in vitro and in vivo. In this review, we introduce nicotinamide adenine dinucleotide phosphate (NADPH) oxidase and epidermal growth factor receptor (EGFR) as molecular targets that can complement antioxidant activity and effectively inhibit skin cancer via the regulation of various signaling networks. In addition, based on recent research [10], we highlight the function of protein tyrosine phosphatase κ (PTP κ) as a mediator of EGFR signal activation by NADPH oxidase (NOX).

The sentence, "prevention is better than a cure" highlights the importance of prevention. Many scientists and laboratories are working to develop chemopreventive materials. However, materials that have proven to be effective in counteracting diseases, such as cancer, are still needed. By summarizing the results of our research and that of other scientists, we aim to introduce materials that could prevent or control skin cancer and skin diseases through the regulation of NOX and EGFR.

2. Skin Carcinogenesis and Chemoprevention

Chemoprevention is defined as the use of a drug or appropriate treatment to delay the onset of cancer or to return cancer to normal [11]. It takes over a decade for cancer to develop [12,13]. Skin cancer develops in multiple stages, which is similarly observed in other cancers. Specifically, in skin carcinogenesis, there are three stages: initiation, promotion, and progression. Acute UV exposure can initiate skin carcinogenesis, which is characterized by DNA mutations and abnormal gene activation or suppression. Promotion is the clonal expansion of damaged cells via aberrant signaling pathways, which takes place over many years. It involves uncontrolled tumor growth that can invade and metastasize to secondary organs [14–16]. Because initiation is an irreversible step lasting only for a short duration (seconds or minutes), the prevention of initiation is difficult to achieve using chemopreventive agents. However, targeting the promotion stage has attracted significant interest because of its characteristic reversibility and long duration (a decade) [15,16]. During the promotion stage, mutations of tumor suppressor genes, including p53, and hyperactivation of inflammatory transcription factors, such as activator protein 1 (AP-1) and nuclear factor kappa-light-chain-enhancer of activated B cells (NF-κB), can occur. Subsequently, abnormal production of the inflammatory enzyme, cyclooxygenase-2 (COX-2), and proinflammatory cytokines, interleukin (IL)-1 β , IL-6, and transforming growth factor β (TGF β), are some of the other features of promotion. Therefore, a vast of research aimed at the development of chemopreventive phytochemicals has focused on targeting oncogenic signaling molecules.

3. Role of Dinucleotide Phosphate Oxidases (NOXs) and Their Downstream Signaling Pathways in UV Radiation-Induced Skin Carcinogenesis

Excessive cellular ROS play a critical role in skin carcinogenesis by inducing genotoxicity via DNA damage, which causes inactivation of tumor suppressor genes and activation of proto-oncogenes. ROS also promote skin carcinogenesis by non-genotoxicity through hyperactivation of oncogenic signaling pathways [17]. NOXs, which are membrane-bound enzymes, respond to UVB and are essential for cellular ROS production [18]. NOXs consist of a catalytic subunit (NOX1 through NOX5, dual oxidase (Duox)1, and Duox2), p22phox, p47phox, p67phox, and the small guanosine triphosphatase, Rac1 [19]. NOX1–NOX3 must bind to their corresponding subunits in order to become activated; however, NOX4 is constitutively active, while NOX5 and the Duox enzymes are calcium-dependent via EF-hands [20,21]. NOX1, 2, 4, and 5 mRNA and NOX1, 2, and four proteins have been identified and are the principal producers of ROS in immortalized human keratinocyte HaCaT cells [22,23].

Generally, UVB irradiation triggers a DNA damage response network that includes cell apoptosis through excessive cell cycle progression and damage to DNA repair machinery [24]. A specific type of UVB-mediated DNA damage increases the occurrence of cyclobutane pyrimidine dimers [25–27]. Therefore, NOX-mediated malfunction of nucleotide excision repair, an essential proofreading system to repair UVB-induced DNA damage, plays a crucial role in skin carcinogenesis [25,28]. A clinical study proposed that N-acetyl cysteine (NAC), a NOX inhibitor, could prevent pro-oncogenic oxidative stress in nevi and alleviate the risk of long-term melanoma when taken by patients [29]. However, a phase II randomized placebo-controlled trial reported that a single oral dose of NAC did not show a significant protective effect on UV-induced nevi due to oxidative stress [30]. The development of skin cancer is observable in SKH-1 hairless mice that are subjected to chronic UVB exposure. However, topical treatment with apocynin, which inhibits NOX activity, prevents two stages of UVB-induced skin carcinogenesis in these mice [31]. Furthermore, direct inhibition of NOX1 with a specific peptide inhibitor, lnhNOX1, has been shown to suppress UVB-induced bursts of ROS production, increase nucleotide excision repair efficiency, decrease apoptosis in HaCaT cells, and decrease skin carcinogenesis in Xpc wild-type mice [24]. These results indicate that NOX plays a critical role in UV-induced skin carcinogenesis and could be a promising target for the generation of chemopreventive materials (Figure 1).

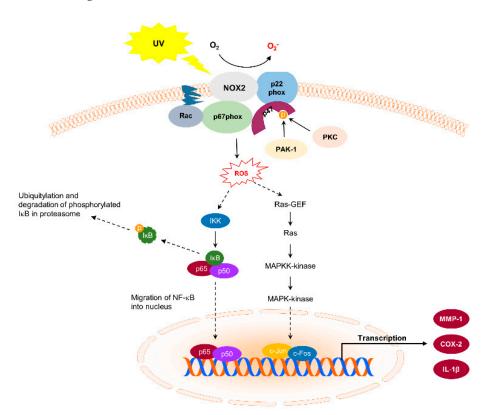


Figure 1. Nicotinamide adenine dinucleotide phosphate oxidase (NOX) and its downstream signaling pathways in ultraviolet (UV)-induced skin carcinogenesis.

UV irradiation and excessive ROS production cause mutations in the protein patched homolog 1 (PTCH) gene, encoding a membrane receptor, which results in the activation of the Hedgehog signaling pathway involved in the pathogenesis of BCC [32]. Once DNA is damaged by acute UV irradiation, p53 makes the DNA repair protein for DNA replication, repairs and regulates the cell cycle, or activates apoptosis-signaling pathways if the DNA damage is irreversible [33,34]. In addition, prolonged UV exposure induces the accumulation of mutations in phosphatase and tensin homolog (PTEN), a tumor suppressor gene that regulates the PI3K/Akt signaling pathway through its phosphatase activity. The results in low expression levels of PTEN in SCC [35,36]. Therefore, damage to p53 and PTEN, due to ROS makes it hard to repair DNA damage caused by UV radiation, ultimately leading to skin cancer. Based on the use of pharmaceutical inhibitors for NOX, Ha et al. observed that NAC suppressed UVB-induced phosphorylation of mitogenactivated protein kinases (MAPKs), Src, and EGFR in HaCaT cells [10]. Therefore, in addition to directly scavenging ROS using antioxidants, modulating EGFR, PI3K, Src, and MAPK activity, such as by phosphorylation, would be a remarkable strategy to prevent UV-induced skin damage and skin cancer.

4. Role of Epidermal Growth Factor Receptor (EGFR) and Its Downstream Signaling Pathways in UV-Induced Skin Carcinogenesis

The epidermal growth factor (EGF) receptors include: ErbB-1 (HER1), ErbB-2 (HER2), ErbB3 (HER3), and ErbB4 (HER4) [37]. These receptors are composed of extracellular ligand-binding, transmembrane, and intracellular tyrosine kinase domains [38,39]. Because hyperactivity and various mutations of EGFR contribute to the development of cancer and drug resistance, many anticancer drugs targeting EGFR are being studied [40–43]. EGFR homodimers and heterodimers are formed when a soluble ligand binds to the receptor's ectodomain. This step is essential for the activation of tyrosine kinase receptors, such as EGFR, via phosphorylation of the C-terminal tail. We have confirmed that UV irradiation upregulates EGFR phosphorylation at Tyr1068, Tyr1045, and Tyr845 in HaCaT cells [10,44].

As a principle of EGFR activation after UV exposure, abnormal ROS production activates a disintegrin and metalloproteinase (ADAM), which performs cleavage to form the EGFR ligand, pro-amphiregulin, at the cell membrane [45]. Free amphiregulin then binds to EGFR and triggers a conformational change of the receptor. EGFR subsequently activates itself through dimerization. We confirmed that amphiregulin expression increases in response to UV irradiation [10] (Figure 2A). Activated EGFR regulates multiple signaling pathways, including p38, c-Jun NH2-terminal kinase (JNK), ERK, PI3K/Akt, and protein kinase C. PI3K/Akt are phosphorylated following UV irradiation in HaCaT cells. Interestingly, UV irradiation brings about the dissociation of AhR and Src complexes, enabling Src to phosphorylate EGFR at Tyr845. Consequently, this phosphorylation increases cyp1a1 mRNA and COX-2 protein expression in HaCaT cells [44]. In short, UV-induced EGFR activation accelerates multiple signaling pathways involved in skin carcinogenesis. Agents with the ability to modulate these signaling cascades could serve as promising chemopreventive materials.

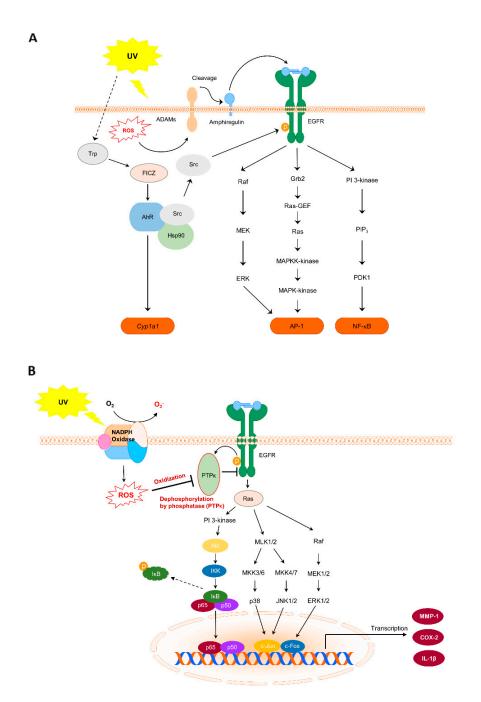


Figure 2. Epidermal growth factor receptor (EGFR) and its downstream signaling pathways, and the possible mechanisms of UV radiation-induced skin carcinogenesis. (**A**) EGFR and its downstream signaling pathways in UV-induced skin carcinogenesis. (**B**) NOX and PTPκ signaling pathways in UV-induced skin carcinogenesis.

5. Role of Protein Tyrosine Phosphatase κ (PTP κ) in NOX and EGFR Signal Transmission in Skin Cancer

An early study suggested that hydrogen peroxide induces EGFR activity via the inhibition of protein tyrosine phosphatase (PTP). This was demonstrated by the addition of catalase, an enzyme that uses hydrogen peroxide as a substrate, which inhibited EGFinduced tyrosine phosphorylation of phospholipase C γ 1 (PLC γ 1), a physiological substrate of the EGFR [46]. Furthermore, several studies have proven that ROS produced by NOX regulates EGFR. NOX4 mediates the oxidation and inactivation of PTP1B in the endoplasmic reticulum (ER) and switches on the EGFR signaling pathways [47]. In addition, activation of NOX by active mutation of TRA4 induces oxidative damage of focal contact phosphatase PTP-PEST [48]. David et al. described the relationship of PTP and EGFR with NOX, the signaling network of EGFR, and the interrelationships within the redox system [49,50]. A recent study revealed the direct relationship between UVB-induced ROS accumulation and the inactivation of PTP κ , which is a phosphatase specific for EGFR [51]. UVB irradiation increases ROS production and NOX activity. It also downregulates PTP κ activity, subsequently upregulating MAPKK and MAPK signaling cascades, and MMP-1 and COX-2 expression in HaCaT cells [10]. Exposure to chronic UVB irradiation in SKH-1 hairless mice led to the development of skin cancer in a two-stage skin carcinogenesis model [10]. Therefore, the interaction of NOX, EGFR, and PTP κ is a crucial factor in skin carcinogenesis. Controlling this interaction is a core approach to prevent UV-induced skin cancer (Figure 2B).

6. Botanical Extracts and Phytochemicals Preventive in UV-Induced Skin Damage via the Regulation of NOX

UVB induces NOX expression in skin tissue, thereby increasing oxidant production. NOX can be assessed both directly, by its activity, and indirectly, by measuring catalase, superoxide dismutase activities, and glutathione depletion (Table 1). To assess whether or not botanical extracts or phytochemicals altered NOX activity, previous studies measured NOX activity, NOX gp91phox mRNA expression, and p47phox translocation.

Name	Stimulus	Measurement of NOX Activity	Enzyme Activity or Transcription Factor	Target Disease	Reference
Eupafolin	LPS	NOX activity, p47 ^{phox} translocation, COX-2 expression using knockdown of NOX2 (gp91 ^{phox}) and p47 ^{phox}	Phosphorylation of c-Fos and c-Jun, AP-1 DNA binding activity	Skin inflammation	[52]
<i>trans-</i> chalcone	UV source	NOX2 gp91 ^{phox} mRNA expression	Myeloperoxidase (MPO) activity, Metalloproteinase-9 (MMP-9) activity, GSH depletion, Catalase (CAT) activity	Skin inflammation Oxidative stress	[53]
Apocynin	UVB	NOX activity	AP-1 promoter activity, NF-кВ promoter activity	Skin carcinogenesis	[31]
Syringic acid	UVB	NOX activity	Metalloproteinase-1 (MMP-1) expression, AP-1 promoter activity	Skin carcinogenesis	[10]
Tannic acid	UVB	NOX activity	Catalase (CAT) activity, Superoxide dismutase (SOD) activity, GSH depletion, Metalloproteinase-1, 9 (MMP-1, 9) expression	Photoaging	[54]
Protocatechuic acid	LIVB NOX a		Catalase (CAT) activity, Superoxide dismutase (SOD) activity, GSH depletion, Lipid peroxidation, Metalloproteinase-1, 9 (MMP-1, 9) expression	Oxidative injuries Photoaging	[55]
Rosmarinic acid in Lycopus europaeus		NOX2 and NOX4 activity		Photoaging	[56]

Table 1. Phytochemicals and their effects on NOX inactivation, enzyme activity, and transcription.

Name	Stimulus	Measurement of NOX Activity	Enzyme Activity or Transcription Factor	Target Disease	Reference
Pimenta pseudocaryophyllus	UVB	NOX2 gp91 ^{phox} mRNA expression	Glutathione reductase mRNA expression, Myeloperoxidase (MPO) activity, Metalloproteinase-9 (MMP-9) activity, GSH depletion, Superoxide anion production, Lipid peroxidation	Skin inflammation Oxidative stress	[57]

Table 1. Cont.

Eupafolin, 6-methoxy 5,7,3',4'-tetrahydroxyflavone, is a vital component of the methanolic extract of *Phyla nodiflora* [52]. Human dermal fibroblasts (Hs68) treated with eupafolin in vitro showed reduced LPS-induced ROS generation via deterioration of $p47^{phox}$ activity and decrease translocation of $p47^{phox}$ from the cytosol to the plasma membrane. The inhibition of LPS-induced COX-2 expression was found to be mediated by transfection with scrambled siRNAs targeting the cytosolic regulatory subunit, $p47^{phox}$, and membrane catalytic subunit, Nox2 (gp91^{phox}). The COX-2 gene promoter contains consensus-binding sites for both AP-1 and NF- κ B; however, eupafolin only reduces the LPS-induced increase in AP-1 activity through the phosphorylation of MAPK. In an in vivo study, male C57BL6 mice were injected subcutaneously with LPS for 24 h following a 30 min pre-treatment with eupafolin (administered intraperitoneally). The results demonstrated that eupafolin induced lower COX-2 expression in the mouse dermis than LPS alone [52]. Overall, the LPS-induced inflammatory response in the skin can be attenuated by inhibiting the Nox2/p47^{phox}/ROS/JNK/AP-1-dependent pathway.

Trans-chalcone [53], 1,3-diphenyl-2-propen-1-one, is the biphenolic precursor of flavonoids in plants. An in vivo experiment was performed using male, hairless mice (HRS/J) treated intraperitoneally with *trans*-chalcone for 1 h before UV irradiation. *Trans*-chalcone reduced UVB-induced excessive generation of superoxide anion and gp91^{phox} mRNA overexpression in these mice. The relationship between acute inflammation and oxidative stress, including the critical roles of NOX2 and ROS as secondary messengers in pro-inflammatory signaling cascades, has been previously described. *Trans*-chalcone reduces proinflammatory Th2 and Th17 cytokines. In addition, *Trans*-chalcone diminishes epidermal thickness and the number of sunburnt cells in the skin of hairless mice exposed to UV radiation. *Pimenta pseudocaryophyllus* belongs to the Myrtaceae family and contains flavonoids and other polyphenolic compounds [57]. *P. pseudocaryophyllus* ethanolic extract (PPE) inhibited UVB-mediated increase in skin edema, gp91^{phox} mRNA overexpression, and IL-1β production. It also increased IL-10 levels in the same model experiment relative to *trans*-chalcone. This compound and extract were suggested to prevent skin diseases, such as oxidative stress and skin inflammation.

Apocynin, a known NOX inhibitor, is an active compound derived from the roots of the medicinal plant *Picrorhiza kurroa* [31]. In vitro, UVB-irradiated epidermal JB6 P⁺ cells from mouse skin were used to study skin carcinogenesis. Pretreatment with apocynin suppressed NOX activity, phosphorylation of MAPK and Akt signaling, and COX-2 activity compared with UVB- irradiated JB6 P⁺ cells. Development of skin tumors was observed within 21 weeks by exposure to UVB, whereas apocynin attenuated UVB-induced tumors in female SKH-1 hairless mice.

Syringic acid is a component of the açaí palm (*Euterpe oleracea*), which contains hydroxybenzoic acid derivatives [10]. UVB-induced NOX activity and phosphorylation of MAPKs, MAPKKs, B-Raf, Akt, and Src were inhibited by syringic acid treatment, in addition to a reduction in the excessive expression of MMP-1 and COX-2 in HaCaT cells. Syringic acid inhibited COX-2 and MMP-13 expression, mouse epidermal thickness, and the num-

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ber of tumors per mouse in SKH-1 hairless mice exposed to UVB (0.2 J/cm^2) irradiation 3 days/week for 22 weeks.

Apocynin and syringic acid have been shown to inhibit NOX and AP-1 or NF- κ B promoter activity, preventing skin carcinogenesis. Thus, phytochemicals can be used to target UV light-induced activation of transcription factors, such as NF- κ B and AP-1 in skin carcinogenesis.

7. Botanical Extracts and Phytochemicals Preventive in UV-Induced Skin Damage via the Regulation of EGFR

Because EGFR has a central role in the growth and survival of cancer cells, many materials have been developed to target EGFR directly. However, the long-term use of EGFR-targeting drugs, such as gefitinib and erlotinib, was accompanied by adverse drug reactions, particularly in the skin. Furthermore, these drugs ultimately failed in female Asian patients with non-small cell lung cancer [58]. Therefore, we investigated the effects of extracts or phytochemicals in the control of upstream or downstream signaling of EGFR, rather than focusing on its direct regulation.

Several studies have explored the effects of nutraceuticals on UV-stimulated skin, in vitro and in vivo, concerning EGFR phosphorylation. However, ERBB activation, dys-regulation of amphiregulin and AhR, and excessive phosphorylation of Src can all be targeted to prevent skin diseases. Therefore, various materials can inhibit skin inflammation or cancer by reducing factors linked to EGFR activation (Table 2).

Name		Stimulus	Measurement of EGFR Activity	Dephosphorylation Targets	Target Disease	Reference
Taxifolin		UVB and solar UV	Phosphorylation (T1068) in vitro EGFR kinase assay	P38, JNK, ERK Akt, p70 ^{s6k} , p90RSK, MSK	Skin carcino- genesis	[59]
Syringic acid		UVB	Phosphorylation (T1068, T1045) Activity of the EGFR phosphatase PTP-к	P38, JNK1/2, ERK1/2 MEK1/2, MKK4/7, MKK3/6, B-Raf, Akt, Src	Skin carcino- genesis	[10]
TGa Cellulose nanocrystal (CNC)		UVB	Phosphorylation (T1068, T1045)	P38, JNK1/2, ERK1/2 MEK1/2, MKK4/7, B-Raf	Skin inflammation	[60]
Carnosi	ic acid	UVB	Phosphorylation	ERK, MEK	Photoaging	[61]
Plant polyphenols	Verbascoside Resveratrol Polydatin Rutin Quercetin	TGFα TNFα + IFNγ UVA + UVB LPS	Phosphorylation nuclear translocation	ERK, p65, Akt	Skin inflammation Skin cancer	[62]
Cyanidin-3-O-glucoside		UVB	Phosphorylation	P38, JNK, ERK, Akt	Epidermal cell apoptosis Skin cancer	[63]
1,8-Cineole		UVB	Phosphorylation (Y845)	P38, JNK1/2, ERK1/2 MEK1/2, B-Raf, C-Raf	Skin carcino- genesis	[44]
Rhus javanica extracts		UVB	Phosphorylation (T1068, T1045) EGFR activity	P38, JNK1/2, ERK1/2 MKK3/6, MKK4/7, MEK1/2, B-Raf, Akt, Src, PKD/PKCµ	Skin inflammation Photoaging	[64]
Curcuma zedoaria extracts		UVB	Phosphorylation (T1068, T1045)	P38, JNK1/2, ERK1/2 MKK3/6, MEK1/2, B-Raf, Akt, Src	Skin inflammation Photoaging	[65]

Table 2. Phytochemicals and their effects on EGFR activation and the phosphorylation of various signaling molecules.

Name	Stimulus	Measurement of EGFR Activity	Dephosphorylation Targets	Target Disease	Reference
Curcumin	EGF	Phosphorylation (T1068) Surface plasmon resonance competition analysis			[66]
Silymarin	EGF	EGFR activity EGFR kinase activity			[67]
Epigallocatechin 3-Gallate	EGF	Phosphorylation of erbB1	ERK1/2		[68]

Table 2. Cont.

The AhR signaling pathway is one of the major UVB-induced signaling pathways, and AhR mediates COX-2 overexpression. We demonstrated that 1,8-cineole, a monoterpene cyclic ether, inhibits c-Src-dependent EGFR activation induced by UVB, thereby suppressing the RAF-MEK1/2-ERK1/2 signaling pathway and cyp1a1 mRNA expression, a target gene of activated AhR [44]. This was demonstrated by the knockdown of AhR, which suppresses COX-2 expression in HaCaT cells. 1,8-cineole also reduced UVB-induced epidermal thickness and skin tumorigenesis in hairless SKH-1 mice. Overall, UVB-induced skin carcinogenesis could be suppressed by directly targeting AhR.

Oi et al. demonstrated via in silico modeling that taxifolin, an ATP-competitive inhibitor, can bind to the ATP-binding pocket of EGFR (Lys 721, Met 769, and Asp 831) and suppress UVB and solar UV-induced carcinogenesis [59]. They also showed that taxifolin regulates COX-2 through EGFR inhibition. In addition, TGa cellulose nanocrystal [60], carnosic acid [61], and cyanidin-3-O-glucoside [63] decreased downstream kinases and Akt or MAPK signaling pathways by reducing UVB-induced phosphorylation of EGFR.

In a recent study, Ha et al. identified that the phosphorylation of EGFR and PTP κ , a regulator of EGFR activation, could be modulated by syringic acid [10]. Notably, oxidized PTP κ is a factor involved in UVB-induced EGFR activation. Among numerous observations, they noted that syringic acid inhibited NOX activity, mainly by its Nox/PTP κ /EGFR axis-driven skin carcinogenic activity [10]. Thus, they suggested that nutraceuticals could regulate a link between the impairment of PTP κ and its rescue. Moreover, it was confirmed that the inhibition of PTP κ oxidation suppressed downstream signaling factors, including Ras, Akt, and MAPK.

The appearance of wrinkles coinciding with MMP-13 and COX-2 expression has also been observed using models of UVB-mediated skin inflammation and photoaging in SKH-1 hairless mice. Both *Rhus javanica* [64] and *Curcuma zedoaria* [65] extracts, administered orally, improved the outcomes of these models. *Curcuma zedoaria* extract reduces MMP-1 expression, phosphorylation of JNK1/2, MKK3/6/p38, B-Raf/MEK1/2/ERK1/2, Src, EGFR (Y1068 and Y1045), and Akt after UVB irradiation in HaCaT cells. Furthermore, *R. javanica* extract demonstrates the same reduction effect as *C. zedoaria* extract and inhibits the phosphorylation of PKD/PKC_µ and Tyr. It also significantly inhibits EGFR kinase activity.

Curcumin, a dietary polyphenol, is the main component of turmeric and is extracted from the East Indian plant, *Curcuma longa*. According to Hosur et al., curcumin treatment inhibited an increase in ear swelling, ERK phosphorylation, and inflammatory cytokine production in a dermatitis model established by dosing female BALB/c mice with 2, 4, 6-trinitrochlorobenzene (TNCB) [66]. Furthermore, amphiregulin gene overexpression was also attenuated by curcumin treatment in the TNCB-challenged ear. Moreover, Capalbo et al. reported preclinical evidence that while conventional EGFR-targeted treatments, such as cetuximab, show resistance, oral curcumin phospholipid supplements can overcome resistance to anti-EGFR treatment. However, the mechanism by which curcumin interacts with EGFR has not been fully elucidated.

8. Conclusions

In this paper, we have described NOX, EGFR, and the crosstalk between these two molecules, as crucial regulators of skin carcinogenesis. In addition, we have discussed the phytochemicals and botanical extracts that exert a preventive effect against UV-induced skin carcinogenesis through their regulation of these molecules (Figures 3 and 4).

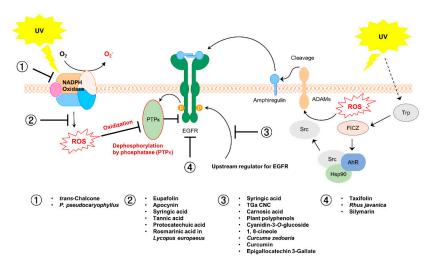


Figure 3. Summary of the mechanism of the effect of botanical extracts and phytochemicals in response to UV irradiation.

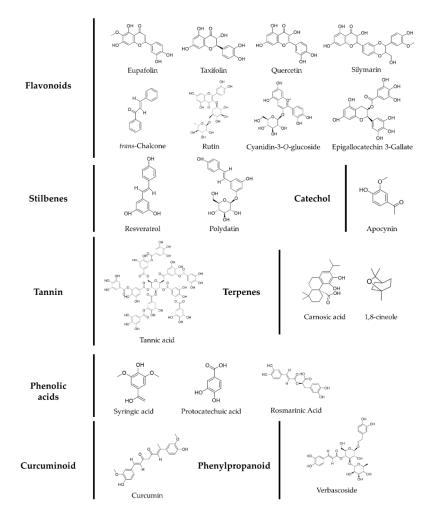


Figure 4. Classification and structures of phytochemicals with regulatory effects on EGFR and NOX activity.

Various materials have been discussed regarding the prevention of skin cancer via the regulation of NOX and EGFR following activation with UV and subordinate factors; however, research on the direct interplay between the materials and these molecules is incomplete. Therefore, to aid the development of more efficient chemopreventive materials in the future, various aspects should be studied, such as the interactions between materials and their target molecules and their mechanisms of action at the molecular and cellular levels.

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Conflicts of Interest: The authors declare no conflict of interest.

References

- 1. Sung, H.; Ferlay, J.; Siegel, R.L.; Laversanne, M.; Soerjomataram, I.; Jemal, A.; Bray, F. Global cancer statistics 2020: GLOBOCAN estimates of incidence and mortality worldwide for 36 cancers in 185 countries. *CA Cancer J. Clin.* **2021**, *71*, 209–249. [CrossRef]
- Fontanillas, P.; Alipanahi, B.; Furlotte, N.A.; Johnson, M.; Wilson, C.H.; Pitts, S.J.; Gentleman, R.; Auton, A. Disease risk scores for skin cancers. *Nat. Commun.* 2021, 12, 160. [CrossRef] [PubMed]
- 3. Lomas, A.; Leonardi-Bee, J.; Bath-Hextall, F. A systematic review of worldwide incidence of nonmelanoma skin cancer. *Br. J. Dermatol.* **2012**, *166*, 1069–1080. [CrossRef]
- 4. Didona, D.; Paolino, G.; Bottoni, U.; Cantisani, C. Non melanoma skin cancer pathogenesis overview. *Biomedicines* **2018**, *6*, 6. [CrossRef]
- 5. Matsumura, Y.; Ananthaswamy, H.N. Short-term and long-term cellular and molecular events following UV irradiation of skin: Implications for molecular medicine. *Expert Rev. Mol. Med.* **2002**, *4*, 1–22. [CrossRef] [PubMed]
- 6. Bowden, G.T. Prevention of non-melanoma skin cancer by targeting ultraviolet-B-light signalling. *Nat. Rev. Cancer* 2004, *4*, 23–35. [CrossRef]
- Armstrong, B.K.; Kricker, A. The epidemiology of UV induced skin cancer. J. Photochem. Photobiol. B Biol. 2001, 63, 8–18. [CrossRef]
- 8. Fisher, G.J.; Talwar, H.S.; Lin, J.; Lin, P.; McPhillips, F.; Wang, Z.; Li, X.; Wan, Y.; Kang, S.; Voorhees, J.J. Retinoic acid inhibits induction of c-Jun protein by ultraviolet radiation that occurs subsequent to activation of mitogen-activated protein kinase pathways in human skin in vivo. *J. Clin. Investig.* **1998**, *101*, 1432–1440. [CrossRef]
- 9. Wlaschek, M.; Tantcheva-Poór, I.; Naderi, L.; Ma, W.; Schneider, L.A.; Razi-Wolf, Z.; Schüller, J.; Scharffetter-Kochanek, K. Solar UV irradiation and dermal photoaging. *J. Photochem. Photobiol. B Biol.* **2001**, *63*, 41–51. [CrossRef]
- 10. Ha, S.J.; Lee, J.; Park, J.; Kim, Y.H.; Lee, N.H.; Kim, Y.E.; Song, K.M.; Chang, P.S.; Jeong, C.H.; Jung, S.K. Syringic acid prevents skin carcinogenesis via regulation of NoX and EGFR signaling. *Biochem. Pharmacol.* 2018, 154, 435–445. [CrossRef] [PubMed]
- 11. Savoia, P.; Zavattaro, E.; Cremona, O. Chemoprevention of skin carcinomas in high-risk transplant recipients. *Curr. Med. Chem.* **2018**, 25, 687–697. [CrossRef]
- 12. Stratton, S.P.; Dorr, R.T.; Alberts, D.S. The state-of-the-art in chemoprevention of skin cancer. *Eur. J. Cancer* 2000, *36*, 1292–1297. [CrossRef]
- 13. Blagodatski, A.; Klimenko, A.; Jia, L.; Katanaev, V.L. Small molecule Wnt pathway modulators from natural sources: History, State of the Art and Perspectives. *Cells* **2020**, *9*, 589. [CrossRef]
- 14. Singh, S.; Singh, P.P.; Roberts, L.R.; Sanchez, W. Chemopreventive strategies in hepatocellular carcinoma. *Nat. Rev. Gastroenterol. Hepatol.* **2014**, *11*, 45–54. [CrossRef]
- 15. Lee, K.W.; Bode, A.M.; Dong, Z. Molecular targets of phytochemicals for cancer prevention. *Nat. Rev. Cancer* 2011, *11*, 211–218. [CrossRef]
- 16. Surh, Y.J. Cancer chemoprevention with dietary phytochemicals. Nat. Rev. Cancer 2003, 3, 768–780. [CrossRef]
- Xian, D.; Lai, R.; Song, J.; Xiong, X.; Zhong, J. Emerging perspective: Role of increased ROS and redox imbalance in skin carcinogenesis. Oxidative Med. Cell. Longev. 2019, 2019, 8127362. [CrossRef] [PubMed]
- 18. Sedeek, M.; Nasrallah, R.; Touyz, R.M.; Hebert, R.L. NADPH oxidases, reactive oxygen species, and the kidney: Friend and foe. *J. Am. Soc. Nephrol.* **2013**, *24*, 1512–1518. [CrossRef] [PubMed]
- 19. Hordijk, P.L. Regulation of NADPH oxidases: The role of Rac proteins. Circ. Res. 2006, 98, 453–462. [CrossRef] [PubMed]

- Bánfi, B.; Tirone, F.; Durussel, I.; Knisz, J.; Moskwa, P.; Molnár, G.Z.; Krause, K.H.; Cox, J.A. Mechanism of Ca²⁺ activation of the NADPH oxidase 5 (NOX5). *J. Biol. Chem.* 2004, 279, 18583–18591. [CrossRef]
- Ameziane-El-Hassani, R.; Morand, S.; Boucher, J.L.; Frapart, Y.M.; Apostolou, D.; Agnandji, D.; Gnidehou, S.; Ohayon, R.; Noël-Hudson, M.S.; Francon, J.; et al. Dual oxidase-2 has an intrinsic Ca²⁺-dependent H₂O₂-generating activity. *J. Biol. Chem.* 2005, 280, 30046–30054. [CrossRef]
- 22. Lee, T.M.; Chen, C.C.; Hsu, Y.J. Differential effects of NADPH oxidase and xanthine oxidase inhibition on sympathetic reinnervation in postinfarct rat hearts. *Free. Radic. Biol. Med.* **2011**, *50*, 1461–1470. [CrossRef]
- Chamulitrat, W.; Stremmel, W.; Kawahara, T.; Rokutan, K.; Fujii, H.; Wingler, K.; Schmidt, H.H.; Schmidt, R. A constitutive NADPH oxidase-like system containing gp91phox homologs in human keratinocytes. J. Investig. Dermatol. 2004, 122, 1000–1009. [CrossRef]
- Raad, H.; Serrano-Sanchez, M.; Harfouche, G.; Mahfouf, W.; Bortolotto, D.; Bergeron, V.; Kasraian, Z.; Dousset, L.; Hosseini, M.; Taieb, A.; et al. NADPH oxidase-1 plays a key role in keratinocyte responses to UV radiation and UVB-induced skin carcinogenesis. J. Investig. Dermatol. 2017, 137, 1311–1321. [CrossRef]
- Pollet, M.; Shaik, S.; Mescher, M.; Frauenstein, K.; Tigges, J.; Braun, S.A.; Sondenheimer, K.; Kaveh, M.; Bruhs, A.; Meller, S.; et al. The AHR represses nucleotide excision repair and apoptosis and contributes to UV-induced skin carcinogenesis. *Cell Death Differ.* 2018, 25, 1823–1836. [CrossRef] [PubMed]
- Huang, K.M.; Liang, S.; Yeung, S.; Oiyemhonlan, E.; Cleveland, K.H.; Parsa, C.; Orlando, R.; Meyskens, F.L., Jr.; Andresen, B.T.; Huang, Y. Topically applied carvedilol attenuates solar ultraviolet radiation induced skin carcinogenesis. *Cancer Prev. Res.* 2017, 10, 598–606. [CrossRef] [PubMed]
- 27. Berton, T.R.; Mitchell, D.L.; Fischer, S.M.; Locniskar, M.F. Epidermal proliferation but not quantity of DNA photodamage is correlated with UV-induced mouse skin carcinogenesis. *J. Investig. Dermatol.* **1997**, *109*, 340–347. [CrossRef]
- Nijhof, J.G.; van Pelt, C.; Mulder, A.A.; Mitchell, D.L.; Mullenders, L.H.; de Gruijl, F.R. Epidermal stem and progenitor cells in murine epidermis accumulate UV damage despite NER proficiency. *Carcinogenesis* 2007, 28, 792–800. [CrossRef]
- 29. Goodson, A.G.; Cotter, M.A.; Cassidy, P.; Wade, M.; Florell, S.R.; Liu, T.; Boucher, K.M.; Grossman, D. Use of oral *N*-acetylcysteine for protection of melanocytic nevi against UV-induced oxidative stress: Towards a novel paradigm for melanoma chemoprevention. *Clin. Cancer Res.* **2009**, *15*, 7434–7440. [CrossRef] [PubMed]
- Cassidy, P.B.; Liu, T.; Florell, S.R.; Honeggar, M.; Leachman, S.A.; Boucher, K.M.; Grossman, D. A Phase II randomized placebocontrolled trial of oral *N*-acetylcysteine for protection of melanocytic nevi against UV-induced oxidative stress in vivo. *Cancer Prev. Res.* 2017, *10*, 36–44. [CrossRef]
- 31. Byun, S.; Lee, E.; Jang, Y.J.; Kim, Y.; Lee, K.W. The NADPH oxidase inhibitor apocynin inhibits UVB-induced skin carcinogenesis. *Exp. Dermatol.* **2016**, *25*, 489–491. [CrossRef] [PubMed]
- Pellegrini, C.; Maturo, M.G.; Di Nardo, L.; Ciciarelli, V.; Garcia-Rodrigo, C.G.; Fargnoli, M.C. Understanding the molecular genetics of basal cell carcinoma. *Int. J. Mol. Sci.* 2017, 18, 2485. [CrossRef]
- 33. Benjamin, C.L.; Ananthaswamy, H.N. p53 and the pathogenesis of skin cancer. *Toxicol. Appl. Pharmacol.* 2007, 224, 241–248. [CrossRef] [PubMed]
- 34. Herbert, K.J.; Cook, A.L.; Snow, E.T. SIRT1 inhibition restores apoptotic sensitivity in p53-mutated human keratinocytes. *Toxicol. Appl. Pharmacol.* **2014**, 277, 288–297. [CrossRef]
- 35. Ming, M.; Feng, L.; Shea, C.R.; Soltani, K.; Zhao, B.; Han, W.; Smart, R.C.; Trempus, C.S.; He, Y.Y. PTEN positively regulates UVB-induced DNA damage repair. *Cancer Res.* **2011**, *71*, 5287–5295. [CrossRef]
- 36. Kitagishi, Y.; Matsuda, S. Redox regulation of tumor suppressor PTEN in cancer and aging (Review). *Int. J. Mol. Med.* **2013**, *31*, 511–515. [CrossRef] [PubMed]
- 37. Wieduwilt, M.J.; Moasser, M.M. The epidermal growth factor receptor family: Biology driving targeted therapeutics. *Cell. Mol. Life Sci.* **2008**, *65*, 1566–1584. [CrossRef]
- 38. Ono, M.; Kuwano, M. Molecular mechanisms of epidermal growth factor receptor (EGFR) activation and response to gefitinib and other EGFR-targeting drugs. *Clin. Cancer Res.* **2006**, *12*, 7242–7251. [CrossRef]
- Ogiso, H.; Ishitani, R.; Nureki, O.; Fukai, S.; Yamanaka, M.; Kim, J.H.; Saito, K.; Sakamoto, A.; Inoue, M.; Shirouzu, M.; et al. Crystal structure of the complex of human epidermal growth factor and receptor extracellular domains. *Cell* 2002, 110, 775–787. [CrossRef]
- Sajadimajd, S.; Bahramsoltani, R.; Iranpanah, A.; Kumar Patra, J.; Das, G.; Gouda, S.; Rahimi, R.; Rezaeiamiri, E.; Cao, H.; Giampieri, F.; et al. Advances on natural polyphenols as anticancer agents for skin cancer. *Pharmacol. Res.* 2020, 151, 104584. [CrossRef]
- 41. Li, L.; Fan, P.; Chou, H.; Li, J.; Wang, K.; Li, H. Herbacetin suppressed MMP9 mediated angiogenesis of malignant melanoma through blocking EGFR-ERK/AKT signaling pathway. *Biochimie* **2019**, *162*, 198–207. [CrossRef]
- 42. Herbst, R.S.; Hong, W.K. IMC-C225, an anti-epidermal growth factor receptor monoclonal antibody for treatment of head and neck cancer. *Semin. Oncol.* 2002, 29, 18–30. [CrossRef] [PubMed]
- 43. Albanell, J.; Rojo, F.; Averbuch, S.; Feyereislova, A.; Mascaro, J.M.; Herbst, R.; LoRusso, P.; Rischin, D.; Sauleda, S.; Gee, J.; et al. Pharmacodynamic studies of the epidermal growth factor receptor inhibitor ZD1839 in skin from cancer patients: Histopathologic and molecular consequences of receptor inhibition. *J. Clin. Oncol.* **2002**, *20*, 110–124. [CrossRef] [PubMed]

- Lee, J.; Ha, S.J.; Park, J.; Kim, Y.H.; Lee, N.H.; Kim, Y.E.; Kim, Y.; Song, K.M.; Jung, S.K. 1,8-cineole prevents UVB-induced skin carcinogenesis by targeting the aryl hydrocarbon receptor. *Oncotarget* 2017, *8*, 105995–106008. [CrossRef] [PubMed]
- 45. Nakayama, H.; Fukuda, S.; Inoue, H.; Nishida-Fukuda, H.; Shirakata, Y.; Hashimoto, K.; Higashiyama, S. Cell surface annexins regulate ADAM-mediated ectodomain shedding of proamphiregulin. *Mol. Biol. Cell* **2012**, *23*, 1964–1975. [CrossRef] [PubMed]
- Bae, Y.S.; Kang, S.W.; Seo, M.S.; Baines, I.C.; Tekle, E.; Chock, P.B.; Rhee, S.G. Epidermal growth factor (EGF)-induced generation of hydrogen peroxide. Role in EGF receptor-mediated tyrosine phosphorylation. *J. Biol. Chem.* 1997, 272, 217–221. [CrossRef] [PubMed]
- 47. Chen, K.; Kirber, M.T.; Xiao, H.; Yang, Y.; Keaney, J.F., Jr. Regulation of ROS signal transduction by NADPH oxidase 4 localization. J. Cell Biol. 2008, 181, 1129–1139. [CrossRef]
- Wu, R.F.; Xu, Y.C.; Ma, Z.; Nwariaku, F.E.; Sarosi, G.A., Jr.; Terada, L.S. Subcellular targeting of oxidants during endothelial cell migration. J. Cell Biol. 2005, 171, 893–904. [CrossRef]
- 49. Heppner, D.E.; Hristova, M.; Dustin, C.M.; Danyal, K.; Habibovic, A.; van der Vliet, A. The NADPH oxidases DUOX1 and NOX2 play distinct roles in redox regulation of epidermal growth factor receptor signaling. *J. Biol. Chem.* **2016**, *291*, 23282–23293. [CrossRef]
- 50. Heppner, D.E.; van der Vliet, A. Redox-dependent regulation of epidermal growth factor receptor signaling. *Redox Biol.* **2016**, *8*, 24–27. [CrossRef]
- Xu, Y.; Shao, Y.; Voorhees, J.J.; Fisher, G.J. Oxidative inhibition of receptor-type protein-tyrosine phosphatase kappa by ultraviolet irradiation activates epidermal growth factor receptor in human keratinocytes. J. Biol. Chem. 2006, 281, 27389–27397. [CrossRef] [PubMed]
- Tsai, M.-H.; Lin, Z.-C.; Liang, C.-J.; Yen, F.-L.; Chiang, Y.-C.; Lee, C.-W. Eupafolin inhibits PGE2 production and COX2 expression in LPS-stimulated human dermal fibroblasts by blocking JNK/AP-1 and Nox2/p47phox pathway. *Toxicol. Appl. Pharmacol.* 2014, 279, 240–251. [CrossRef] [PubMed]
- Martinez, R.M.; Pinho-Ribeiro, F.A.; Steffen, V.S.; Caviglione, C.V.; Fattori, V.; Bussmann, A.J.; Bottura, C.; Fonseca, M.J.; Vignoli, J.A.; Baracat, M.M. *trans*-Chalcone, a flavonoid precursor, inhibits UV-induced skin inflammation and oxidative stress in mice by targeting NADPH oxidase and cytokine production. *Photochem. Photobiol. Sci.* 2017, *16*, 1162–1173. [CrossRef]
- 54. Daré, R.G.; Nakamura, C.V.; Ximenes, V.F.; Lautenschlager, S.O.S. Tannic acid, a promising anti-photoaging agent: Evidences of its antioxidant and anti-wrinkle potentials, and its ability to prevent photodamage and MMP-1 expression in L929 fibroblasts exposed to UVB. *Free Radic. Biol. Med.* **2020**, *160*, 342–355. [CrossRef]
- 55. Daré, R.G.; Oliveira, M.M.; Truiti, M.C.T.; Nakamura, C.V.; Ximenes, V.F.; Lautenschlager, S.O.S. Abilities of protocatechuic acid and its alkyl esters, ethyl and heptyl protocatechuates, to counteract UVB-induced oxidative injuries and photoaging in fibroblasts L929 cell line. *J. Photochem. Photobiol. B Biol.* **2020**, *203*, 111771. [CrossRef]
- Revoltella, S.; Baraldo, G.; Waltenberger, B.; Schwaiger, S.; Kofler, P.; Moesslacher, J.; Huber-Seidel, A.; Pagitz, K.; Kohl, R.; Jansen-Duerr, P.; et al. Identification of the NADPH oxidase 4 inhibiting principle of *Lycopus europaeus*. *Molecules* 2018, 23, 653. [CrossRef] [PubMed]
- 57. Campanini, M.Z.; Pinho-Ribeiro, F.A.; Ivan, A.L.; Ferreira, V.S.; Vilela, F.M.; Vicentini, F.T.; Martinez, R.M.; Zarpelon, A.C.; Fonseca, M.J.; Faria, T.J. Efficacy of topical formulations containing *Pimenta pseudocaryophyllus* extract against UVB-induced oxidative stress and inflammation in hairless mice. *J. Photochem. Photobiol. B Biol.* **2013**, *127*, 153–160. [CrossRef]
- 58. Sasaki, H.; Endo, K.; Konishi, A.; Takada, M.; Kawahara, M.; Iuchi, K.; Matsumura, A.; Okumura, M.; Tanaka, H.; Kawaguchi, T.; et al. EGFR mutation status in japanese lung cancer patients: Genotyping analysis using lightcycler. *Clin. Cancer Res. Off. J. Am. Assoc. Cancer Res.* 2005, 11, 2924–2929. [CrossRef]
- 59. Oi, N.; Chen, H.; Ok Kim, M.; Lubet, R.A.; Bode, A.M.; Dong, Z. Taxifolin suppresses UV-induced skin carcinogenesis by targeting EGFR and PI3K. *Cancer Prev. Res.* 2012, *5*, 1103–1114. [CrossRef] [PubMed]
- 60. So, B.R.; Yeo, H.J.; Lee, J.J.; Jung, Y.H.; Jung, S.K. Cellulose nanocrystal preparation from *Gelidium amansii* and analysis of its anti-inflammatory effect on the skin in vitro and in vivo. *Carbohydr. Polym.* **2021**, 254, 117315. [CrossRef]
- 61. Park, M.; Han, J.; Lee, C.S.; Heung Soo, B.; Lim, K.M.; Ha, H. Carnosic acid, a phenolic diterpene from rosemary, prevents UV-induced expression of matrix metalloproteinases in human skin fibroblasts and keratinocytes. *Exp. Dermatol.* **2013**, *22*, 336–341. [CrossRef]
- 62. Potapovich, A.I.; Lulli, D.; Fidanza, P.; Kostyuk, V.A.; De Luca, C.; Pastore, S.; Korkina, L.G. Plant polyphenols differentially modulate inflammatory responses of human keratinocytes by interfering with activation of transcription factors NFκB and AhR and EGFR–ERK pathway. *Toxicol. Appl. Pharmacol.* **2011**, 255, 138–149. [CrossRef]
- 63. He, Y.; Hu, Y.; Jiang, X.; Chen, T.; Ma, Y.; Wu, S.; Sun, J.; Jiao, R.; Li, X.; Deng, L. Cyanidin-3-O-glucoside inhibits the UVB-induced ROS/COX-2 pathway in HaCaT cells. *J. Photochem. Photobiol. B Biol.* **2017**, *177*, 24–31. [CrossRef] [PubMed]
- 64. Ha, S.J.; Lee, J.; Kim, H.; Song, K.-M.; Lee, N.H.; Kim, Y.E.; Lee, H.; Kim, Y.H.; Jung, S.K. Preventive effect of *Rhus javanica* extract on UVB-induced skin inflammation and photoaging. *J. Funct. Foods* **2016**, *27*, 589–599. [CrossRef]
- 65. Jeong Ha, S.; Song, K.-M.; Lee, J.; Ho Kim, Y.; Hyouck Lee, N.; Eon Kim, Y.; Lee, S.; Keun Jung, S. Preventive effect of *Curcuma zedoaria* extract on UVB-induced skin inflammation and photoaging. J. Food Biochem. **2018**, 42, e12598. [CrossRef]
- 66. Starok, M.; Preira, P.; Vayssade, M.; Haupt, K.; Salomé, L.; Rossi, C. EGFR inhibition by curcumin in cancer cells: A dual mode of action. *Biomacromolecules* 2015, *16*, 1634–1642. [CrossRef]

- Ahmad, N.; Gali, H.; Javed, S.; Agarwal, R. Skin cancer chemopreventive effects of a flavonoid antioxidant silymarin are mediated via impairment of receptor tyrosine kinase signaling and perturbation in cell cycle progression. *Biochem. Biophys. Res. Commun.* 1998, 247, 294–301. [CrossRef] [PubMed]
- 68. Bhatia, N.; Agarwal, C.; Agarwal, R. Differential responses of skin cancer-chemopreventive agents silibinin, quercetin, and epigallocatechin 3-gallate on mitogenic signaling and cell cycle regulators in human epidermoid carcinoma A431 cells. *Nutr. Cancer* **2001**, *39*, 292–299. [CrossRef]