Open Access



Glutathione S-transferase pi 1 variant and squamous cell carcinoma susceptibility: a meta-analysis of 52 case-control studies

Shuang Wang¹, Jingqi Zhang¹, Fan Jun¹ and Zhijie Bai^{2*}

Abstract

Background: There are several meta-analyses on the genetic relationship between the rs1695 polymorphism within the *GSTP1* (glutathione S-transferase pi 1) gene and the risk of different SCC (squamous cell carcinoma) diseases, such as ESCC (oesophageal SCC), HNSCC (head and neck SCC), LSCC (lung SCC), and SSCC (skin SCC). Nevertheless, no unified conclusions have been drawn.

Methods: Herein, an updated meta-analysis was performed to evaluate the probable impact of *GSTP1* rs1695 on the susceptibility to different SCC diseases under six genetic models (allele, carrier, homozygote, heterozygote, dominant, and recessive). Three online databases, namely, PubMed, WOS (Web of Science), and Embase (Excerpta Medica Database), were searched.

Results: Initially, we obtained a total of 497 articles. Based on our selection criteria, we eventually included 52 casecontrol studies (9763 cases/15,028 controls) from 47 eligible articles. As shown in the pooling analysis, there was no difference in the risk of overall SCC disease between cases and controls [allele, P_a (P value of association test) = 0. 601; carrier, $P_a = 0.587$; homozygote, $P_a = 0.689$; heterozygote, $P_a = 0.167$; dominant, $P_a = 0.289$; dominant, $P_a = 0.548$]. Similar results were obtained after stratification by race (Asian/Caucasian), genotyping, control source, and disease type (ESCC/HNSCC/LSCC/SSCC) (all $P_a > 0.05$).

Conclusion: The rs1695 polymorphism within the *GSTP1* gene is not associated with the risk of overall SCC or a specific SCC type, including ESCC, HNSCC, LSCC, and SSCC.

Keywords: GSTP1, Polymorphism, Squamous cell carcinoma, Susceptibility

Background

SCC (squamous cell carcinoma), also termed "epidermal carcinoma," is a malignant tumour that takes part in epidermis or adnexal cells and exhibits distinct degrees of keratosis [1-3]. SCC exists in the squamous epithelium of several places, e.g., skin, mouth, lung, lips, oesophagus, cervix, and vagina [4-6]. Based on GWAS (genome-wide association study) data, more and more reported genetic polymorphisms are believed to contribute to the aetiologies of different SCC types. For instance, a series of genes, including *CADM*1 (cell adhesion molecule 1), *AHR* (aryl hydrocarbon receptor), and *SEC16A* (SEC16 homolog A,

endoplasmic reticulum export factor), may be related with the risk of SCC [7]. Two variants within the *KLF5* (Kruppel-like factor 5) gene on chromosome 13q22.1, namely, rs1924966 and rs115797771, may be relevant to ESCC (oesophageal SCC) susceptibility [8]. Herein, we determined whether *GSTP1* (glutathione S-transferase pi 1) gene polymorphism is associated with the susceptibility to different SCC patterns.

GSTP1, a member of the GST (glutathione S-transferase) family in humans, is associated with the biological detoxification or biotransformation process through catalysing the conjugation of many hydrophobic and electrophilic compounds with reduced glutathione [9, 10]. The *GSTP1* gene, which is located on human chromosome 11q13, comprises seven exons and six introns [11]. Two common polymorphisms, namely, rs1695 A/G polymorphism in exon five



© The Author(s). 2019 **Open Access** This article is distributed under the terms of the Creative Commons Attribution 4.0 International License (http://creativecommons.org/licenses/by/4.0/), which permits unrestricted use, distribution, and reproduction in any medium, provided you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons license, and indicate if changes were made. The Creative Commons Public Domain Dedication waiver (http://creativecommons.org/publicdomain/zero/1.0/) applies to the data made available in this article, unless otherwise stated.

^{*} Correspondence: bzjky1127@126.com

²Department of Urology Surgery, Tianjin First Center Hospital, Tianjin 300192, China

Full list of author information is available at the end of the article

(p.Ile105Val) and rs1138272 C/T polymorphism in exon six (p.Ala114Val), have been reported [12, 13].

Several SCC/GSTP1 rs1695-associated meta-analyses with conflicting conclusions have been reported. For instance, in 2009, Zendehdel et al. enrolled three case-control studies [14-16], performed a meta-analysis to assess the association between GSTP1 rs1695 and ESCC risk in Caucasian populations, and found a borderline significant association [16]. In 2014, Song et al. enrolled 21 case-control studies to perform a meta-analysis concerning the role of the GSTP1 rs1695 polymorphism in the risk of oesophageal cancers, including EAC (oesophageal adenocarcinoma) and ESCC [17]. The subgroup meta-analysis of ESCC containing thirteen case-control studies showed a positive correlation, particularly in the Caucasian population [17]. However, in 2015, Tan et al. performed another meta-analysis with twenty case-control studies on overall oesophageal cancer and reported negative results in both ESCC and EAC subgroups [18]. Accordingly, we performed an updated meta-analysis with a relatively larger sample size to reevaluate the potential impact of the GSTP1 rs1695 A/G polymorphism on the susceptibility to SCC diseases, mainly including ESCC, SSCC, HNSCC (head and neck SCC), and LSCC (lung SCC).

Methods

Electronic database retrieval

We reviewed three on-line databases, including PubMed, WOS (Web of Science), and Embase (Excerpta Medica Database), through January 2018 using the following main search keywords: Carcinoma, Squamous Cell; Carcinomas, Squamous Cell; Squamous Cell Carcinomas; Squamous Cell Carcinoma; Carcinoma, Squamous; Carcinomas, Squamous; Squamous Carcinoma; Squamous Carcinomas; Carcinoma, Epidermoid; Carcinomas, Epidermoid; Epidermoid Carcinoma; Epidermoid Carcinomas; Carcinoma, Planocellular; Carcinomas, Planocellular; Planocellular Carcinoma; Planocellular Carcinomas; SCC; GSTP1; Glutathione S-Transferase pi; Glutathione S Transferase pi; GST Class-phi; Class-phi, GST; GST Class phi; Glutathione Transferase P1-1; Glutathione Transferase P1 1; Transferase P1-1, Glutathione; GSTP1 Glutathione D-Transferase; D-Transferase, GSTP1 Glutathione; GSTP1 Glutathione D Transferase; Glutathione D-Transferase, GSTP1; Polymorphism; Polymorphism, Genetic; Polymorphisms, Genetic; Genetic Polymorphisms; Genetic Polymorphism; Polymorphism (Genetics); Polymorphisms (Genetics); and Polymorphism; Polymorphisms.

Eligible article screening

We performed a literature search and screened the retrieved articles as per the PRISMA (preferred reporting items for systematic reviews and meta-analyses) guidelines [19]. Selection criteria included duplicated articles; data from animal or cell experiments; meeting abstract or meta-analysis; review, trials or case reports; data of GSTP1 expression; not SCC or GSTP1; lack confirmed histopathological data; combined GA + AA genotype frequency; without the control data; and P value of HWE (Hardy-Weinberg equilibrium) less than 0.05. Eligible case-control studies provided sufficient genotype fre-

Data extraction

each case and control group.

Two investigators independently extracted the data and evaluated the methodological quality of each article by means of the NOS (Newcastle-Ottawa Scale) system. One table contains the following basic information: first author, publication year, region, race, genotyping assay, genotype frequency, disease type, control source, P values of HWE, study number, and sample size of the case/control.

quency data of the GSTP1 gene rs1695 polymorphism in

Data synthesis

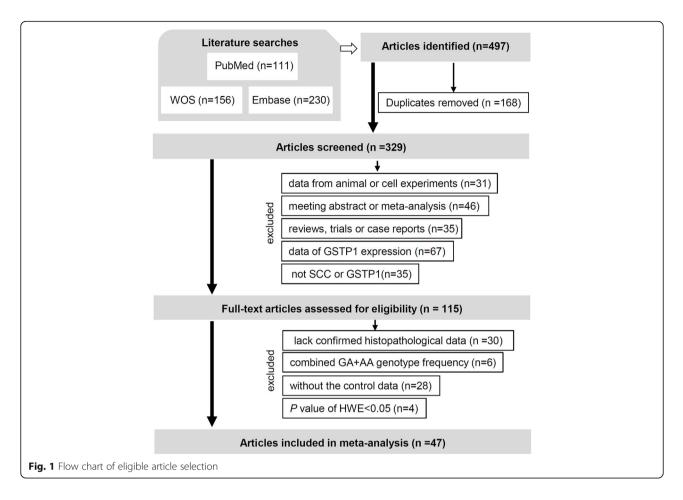
We utilized STATA software (StataCorp LP, College Station, TX, USA) for the following statistical analyses. The allele (allele G vs. A), carrier (carrier G vs. A), homozygote (GG vs. AA), heterozygote (AG vs. AA), dominant (AG + GG vs. AA), and recessive (GG vs. AA+AG) models were utilized to target the *GSTP1* gene rs1695 G/A polymorphism. We calculated the OR (odds ratio), 95% CIs (confidence intervals) and P_a (*P* value of association test) values to estimate the association. When the P_h (*P* value of heterogeneity) was > 0.1 or I² was < 50.0%, a fixed-effects model was adopted. Otherwise, a random-effects model was selected.

Considering the factors of race, genotyping assay, control source, and disease type, we performed the corresponding subgroup meta-analyses. We also carried out Egger's/Begg's tests to determine a potential publication bias. The presence of a publication bias was considered when $P_{\rm E}$ (*P* value of Egger's test) and $P_{\rm B}$ (*P* value of Begg's test) were below 0.05. Sensitivity analysis was applied to assess data stability and robustness.

Results

Article retrieval and screening

The article retrieval and selection processes during our meta-analysis were conducted as described in the flow chart shown in Fig. 1. After our literature search, a total of 497 articles were obtained. Then, 168 articles with duplicated data and 214 articles meeting the exclusion criteria were excluded. Next, we assessed the eligibility of the remaining 115 full-text articles. After the exclusion of 68 ineligible articles, a total of 47 articles containing 52 case-control studies [14–16, 20–63] were ultimately



recruited for our meta-analysis. Table 1 summarizes the extracted basic information.

Overall meta-analysis

First, we performed the overall meta-analysis, which included 52 case-control studies with 9763 cases and 15,028 controls (Table 2). The fixed-effects model was applied in all meta-analyses, because no substantial between-study heterogeneity was detected [Table 2, I² value < 50.0%, $P_{\rm h} > 0.1$]. As shown in Table 2, no altered susceptibility to SCC disease in cases was observed compared with controls [allele, $P_{\rm a} = 0.601$; carrier, $P_{\rm a} = 0.587$; homozygote, $P_{\rm a} = 0.689$; heterozygote, $P_{\rm a} = 0.167$; dominant, $P_{\rm a} = 0.289$; dominant, $P_{\rm a} = 0.548$]. These data suggest that the rs1695 polymorphism within the *GSTP1* gene does not contribute to the risk of overall SCC.

Subgroup analysis

Next, we performed additional subgroup meta-analyses according to the factors of race (Asian/Caucasian), genotyping assay (PCR-RFLP), control source (PB/HB), and disease type (ESCC/HNSCC/LSCC/SSCC). As shown in Tables 3 and 4, there were no significant associations in any subgroup analysis for all genetic models tested (all $P_{\rm a}$ > 0.05). The forest plot of the subgroup analysis by disease type under the allele model is shown in Fig. 2.

Furthermore, we included all case-controls studies regarding the specific SCC type and conducted a series of subgroup analyses by race and control source. However, similar results were obtained (data not shown). As a result, the *GSTP1* gene rs1695 polymorphism is not likely related to the genetic susceptibility of a specific SCC type, including ESCC, HNSCC, LSCC, and SSCC.

Publication bias and sensitivity analysis

The publication bias analysis data obtained from Egger's and Begg's tests are shown in Table 2. There was no remarkable publication bias in most genetic models ($P_{\rm E} > 0.05$, $P_{\rm B} > 0.05$), except for the heterozygote ($P_{\rm E} = 0.022$, $P_{\rm B} = 0.049$) and dominant ($P_{\rm E} = 0.036$) models. The funnel plot (allele model) is displayed in Fig. 3a-b. Moreover, our sensitivity analysis led us to consider the stability of the data. Figure 4 shows a representative example of the sensitivity analysis (allele model).

Discussion

In the current meta-analysis, we first focused on the genetic relationship between the *GSTP1* rs1695 A/G

Table 1 Basic information of the eligible articles in the meta-analysis

First author	Year	Region	Race	Assay	Case			Disease	Cont	rol		Control	P _{HWE}
					AA	AG	GG	type	AA	AG	GG	source	
Abbas	2004	France	Caucasian	PCR-RFLP	21	21	3	ESCC	59	56	9	PB	0.38
Cabelguenne	2001	France	Caucasian	PCR-RFLP	89	57	16	HNSCC	146	139	25	HB	0.31
Cai	2006	China	Asian	PCR-RFLP	143	58	3	ESCC	265	116	12	PB	0.87
Cho	2006	Korea	Asian	Gene sequencing	201	85	7	HNSCC	211	112	10	HB	0.29
Dura	2013	Netherlands	Caucasian	PCR	48	42	15	ESCC	246	261	84	PB	0.27
Dzian	2012	Netherlands	Caucasian	PCR-RFLP	56	45	11	LSCC	153	115	22	PB/HB	0.95
Evans	2004	USA	Caucasian	PCR-RFLP	123	132	27	HNSCC	97	85	24	PB	0.42
Fryer	2005	Australia	Caucasian	PCR-RFLP	59	51	18	SSCC	95	90	25	HB	0.60
Harth	2008	Germany	Caucasian	PCR-melting-curve	145	122	45	HNSCC	130	138	32	HB	0.62
Jain	2006	India	Asian	PCR-RFLP	46	23	7	ESCC	72	56	9	HB	0.67
Jourenkova	1999a	France	Caucasian	PCR-RFLP	49	53	15	HNSCC	86	64	22	HB	0.07
Jourenkova	1999b	France	Caucasian	PCR-RFLP	62	52	15	HNSCC	86	64	22	HB	0.07
Jourenkova	1998	France	Caucasian	PCR-RFLP	46	41	11	LSCC	86	64	22	HB	0.07
Kelders	2002	Netherlands	Caucasian	PCR-RFLP	36	38	13	HNSCC	26	18	7	HB	0.20
Kihara	1999	Japan	Asian	PCR-RFLP	84	32	9	LSCC	184	65	8	HB	0.45
Larsen	2006	Australia	Caucasian	PCR-RFLP	230	213	51	LSCC	161	169	49 ^a	HB	0.66
		Australia	Caucasian	PCR-RFLP	230	213	51	LSCC	112	100	35 ^b	PB	0.11
Leichsenring	2006	Brazil	Mixed	PCR-RFLP	30	34	8	HNSCC	30	25	5	PB	0.95
Leite	2007	Brazil	Mixed	PCR-RFLP	14	13	2	SSCC	60	46	18	PB	0.07
Lewis	2002	UK	Caucasian	PCR-RFLP	14	17	1	LSCC	64	74	13	HB	0.19
Li	2010	South African	Black African	PCR-RFLP	56	59	26	ESCC	76	83	27	PB	0.58
			Mixed	PCR-RFLP	34	52	11	ESCC	30	51	13	PB	0.24
Li	2007	USA	Caucasian	PCR-RFLP	336	356	111	HNSCC	333	385	121	PB	0.57
Liang	2005	China	Asian	diASA-AMP	58	32	4	LSCC	132	86	9	HB	0.27
Liu	2010	China	Asian	PCR-RFLP	66	29	0	ESCC	61	27	3	PB	1.00
Malik	2010	India	Asian	PCR-RFLP	53	36	14	ESCC	111	75	9	PB	0.41
Matejcic	2011	South African	Black African	TaqMan genotyping	79	155	91	ESCC	100	242	132	PB	0.57
		South African	Mixed	TaqMan genotyping	69	112	48	ESCC	145	191	92	PB	0.05
McWilliams	2000	USA	Mixed	PCR-RFLP	60	73	13	HNSCC	58	51	15	HB	0.47
Miller	2006	USA	Caucasian	PCR-RFLP	190	173	49	LSCC	579	623	141	PB	0.16
Moaven	2010	Iran	Asian	PCR-RFLP	84	50	14	ESCC	74	54	8	PB	0.65
Nazar	2003	USA	Mixed	PCR-RFLP	35	29	9	LSCC	199	234	54	PB	0.23
Olshan	2000	USA	Mixed	PCR-RFLP	40	62	7	HNSCC	68	80	20	HB⊂	0.63
		USA	Mixed	PCR-RFLP	18	38	7	HNSCC	7	13	5	HB^d	0.82
Oude	2003	Netherlands	Caucasian	PCR-RFLP	116	90	29	HNSCC	125	121	39	PB	0.27
Peters	2006	USA	Mixed	PCR-RFLP	303	311	76	HNSCC	333	329	86	PB	0.73
Ramsay	2001	UK	Caucasian	SSCP	10	10	0	SSCC	53	71	17	HB	0.36
Risch	2001	Germany	Caucasian	PCR-RFLP	76	77	18	LSCC	167	151	35	HB	0.92
Rossini	2007	Brazil	Mixed	PCR-RFLP	42	65	18	ESCC	116	108	28	PB	0.71
Ruwali	2009	India	Caucasian	PCR-RFLP	224	112	14	HNSCC	199	138	13	PB	0.06
Ruwali	2011	India	Caucasian	PCR-RFLP	316	162	22	HNSCC	285	195	20	PB	0.06
Ryberg	1997	Norway	Caucasian	PCR-RFLP	20	34	13	LSCC	153	117	27	PB	0.50
Schneider	2004	Germany	Caucasian	PCR-melting-curve	81	75	27	LSCC	298	254	70	PB/HB	0.16

 Table 1 Basic information of the eligible articles in the meta-analysis (Continued)

First author	Year	Region	Race	Assay	Case			Disease	Cont	rol		Control	P _{HWE}
					AA	AG	GG	type	AA	AG	GG	source	
Soucek	2010	Czech/Polish	Caucasian	TaqMan drug metabolism genotyping	56	53	7	HNSCC	57	50	10	PB	0.52
Soya	2007	India	Asian	PCR-RFLP	219	162	27	UADTSCC	120	88	12	PB	0.42
Stücker	2002	France	Caucasian	PCR-RFLP	54	46	15	LSCC	124	120	20	HB	0.22
Tan	2000	China	Asian	PCR-RFLP	93	48	9	ESCC	91	53	6	PB	0.62
То	2002	Spain	Caucasian	PCR-RFLP	101	84	19	HNSCC	100	78	23	PB	0.20
То	1999	Spain	Caucasian	PCR-RFLP	29	20	3	LSCC	64	54	14	PB ^b	0.61
		Spain	Caucasian	PCR-RFLP	29	20	3	LSCC	90	90	20	PB ^e	0.72
van	1999	Netherlands	Caucasian	PCR-RFLP	5	6	2	ESCC	146	89	12	PB	0.74
Zendehdel	2009	Sweden	Caucasian	Pyrosequencing	26	42	10	ESCC	208	207	38	PB	0.18

PCR polymerase chain reaction, PCR-RFLP polymerase chain reaction-restriction fragment length polymorphism, diASA-AMP di-allele-specific-amplification with artificially modified primers assay, SSCP Single-stranded conformational polymorphism, ESCC oesophageal squamous cell carcinoma, HNSCC head and neck squamous cell carcinoma, LSCC lung squamous cell carcinoma, SSCC skin squamous cell carcinoma, OSCC oral squamous cell carcinoma, UADTSCC upper aerodigestive tract squamous cell carcinoma, PB population-based, HB hospital-based, P_{HWE} P value of hardy-weinberg equilibrium

^aCOPD patients without LSCC, ^bhealthy smokers; ^ccontrol from Caucasian population; ^dcontrol from Black African population; ^econtrol from general population

polymorphism and the risk of overall SCC and then conducted subgroup analyses by the specific histological status. After rigorous screening, four main types of SCC, namely, ESCC, HNSCC, ESCC, and SSCC, were targeted.

ESCC, a type of squamous epithelium differentiation of a malignant tumour within the oesophagus, accounts for the vast majority of oesophageal cancers [64, 65]. ESCC often presents in physiological or pathological stenosis of the oesophagus, and genetic factors, carcinogens, and/or chronic irritants may contribute to the pathogenesis of ESCC [64, 65]. The *GSTP1* rs1695 A/G polymorphism is significantly related to the risk of ESCC in the Kashmiri population [42]. Similarly, *GSTP1* rs1695 may be an independent risk factor for ESCC in Western populations [53]. Nevertheless, different

Table 2 Meta-analysis of the GSTP1 rs1695 A/G polymorphism

associations were detected in other reports. For instance, no difference between unrelated controls and ESCC cases was observed in a French population [14] or a Chinese population [61]. Therefore, a meta-analysis was required to comprehensively evaluate the role of the GSTP1 rs1695 A/G polymorphism in ESCC risk. Herein, we recruited 15 case-control studies involving 1934 and 3951 controls and performed a new cases meta-analysis to examine the association between the GSTP1 rs1695 A/G polymorphism and ESCC susceptibility. The carrier (carrier G vs. A) model, as well as the allele, homozygote, heterozygote, dominant and recessive genetic models, was used. Our results in the stratified analysis of specific ESCCs are consistent with the data of Tan et al. [18].

Statistical analysis	Index	Allele	Carrier	Homozygote	Heterozygote	Dominant	Recessive
Association	OR	0.99	0.99	1.02	0.96	0.97	1.03
	95% Cls	0.95~1.03	0.94~1.03	0.93~1.12	0.91~1.02	0.92~1.03	0.94~1.12
	Pa	0.601	0.587	0.689	0.167	0.289	0.548
Sample size	case	9763	9763	9763	9763	9763	9763
	control	15,028	15,028	15,028	15,028	15,028	15,028
	study	52	52	52	52	52	52
Heterogeneity	²	15.5%	0.0%	9.7%	7.7%	11.8%	1.2%
	P_h	0.174	0.999	0.278	0.318	0.239	0.450
	Model	Fixed	Fixed	Fixed	Fixed	Fixed	Fixed
Egger's test	t	1.14	1.38	0.13	2.36	2.16	-0.31
	P_E	0.259	0.175	0.899	0.022	0.036	0.760
Begg's test	Z	0.53	0.84	0.77	1.96	1.82	1.29
	P_B	0.597	0.398	0.444	0.049	0.068	0.198

OR odds ratio, Cls confidence intervals, Pa, P value of association test, Ph, P value of heterogeneity test, PE, P value of Egger's test, PB, P value of Begg's test

Table 3 Subgroup analysis of the GSTP1 rs1695 A/G polymorphism by race, genotyping assay and control source

Factor	Subgroup	Index	Allele	Carrier	Homozygote	Heterozygote	Dominant	Recessive
Race	Asian	OR (95% Cls)	1.00 (0.89~1.12)	0.98 (0.86~1.11)	1.29 (0.94~1.76)	0.90 (0.78~1.04)	0.94 (0.82~1.08)	1.35 (0.99~1.83)
		Pa	0.948	0.716	0.114	0.139	0.361	0.058
		Case/control	1696/2139	1696/2139	1696/2139	1696/2139	1696/2139	1696/2139
		Study number	10	10	10	10	10	10
Race	Caucasian	OR (95% CIs)	0.98 (0.93~1.03)	0.98 (0.82~1.04)	1.00 (0.89~1.12)	0.94 (0.87~1.01)	0.95 (0.89~1.02)	1.02 (0.91~1.14)
		Pa	0.358	0.447	0.984	0.099	0.153	0.716
		Case/control	5968/9719	5968/9719	5968/9719	5968/9719	5968/9719	5968/9719
		Study number	30	30	30	30	30	30
genotyping assay	PCR-RFLP	OR (95% CIs)	0.99 (0.94~1.03)	0.99 (0.93~1.04)	1.01 (0.91~1.12)	0.96 (0.90~1.03)	0.97 (0.91~1.03)	1.01 (0.91~1.12)
		Pa	0.542	0.579	0.874	0.260	0.351	0.824
		Case/control	8008/11,342	8008/11,342	8008/11,342	8008/11,342	8008/11,342	8008/11,342
		Study number	42	42	42	42	42	42
control source	PB	OR (95% Cls)	0.98 (0.94~1.03)	0.98 (0.93~1.04)	1.00 (0.90~1.12)	0.96 (0.89~1.03)	0.96 (0.90~1.03)	1.02 (0.92~1.13)
		Pa	0.519	0.572	0.943	0.214	0.287	0.751
		Case/control	6697/10,170	6697/10,170	6697/10,170	6697/10,170	6697/10,170	6697/10,170
		Study number	31	31	31	31	31	31
control source	HB	OR (95% CIs)	0.98 (0.91~1.06)	0.98 (0.90~1.07)	1.00 (0.84~1.20)	0.95 (0.86~1.06)	0.96 (0.87~1.07)	1.01 (0.85~1.19)
		Pa	0.586	0.638	0.977	0.377	0.461	0.944
		Case/control	2771/3946	2771/3946	2771/3946	2771/3946	2771/3946	2771/3946
		Study number	19	19	19	19	19	19

 P_{a} , P value of association test

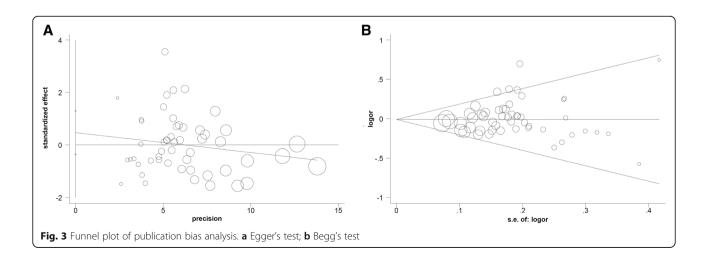
PCR-RFLP polymerase chain reaction-restriction fragment length polymorphism, PB population-based, HB hospital-based, OR odds ratio, CIs confidence intervals

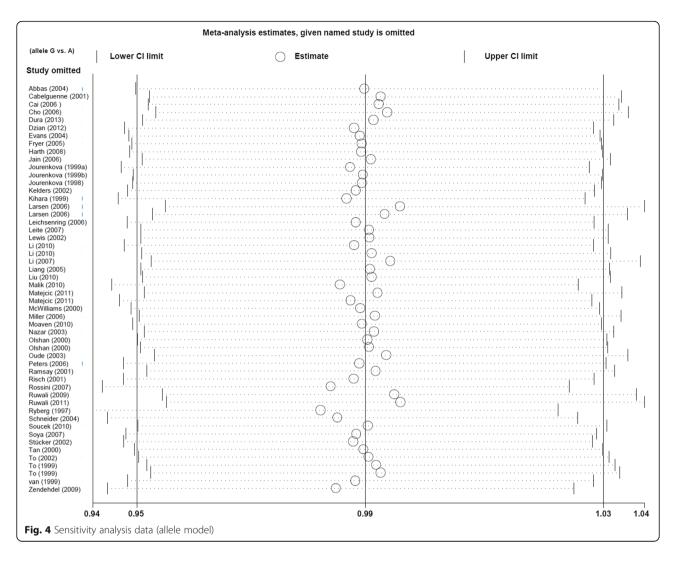
Table 4 Subgroup analysis of the GSTP1 rs1695 A/G polymorphism by SCC type

Subgroup	Index	Allele	Carrier	Homozygote	Heterozygote	Dominant	Recessive
ESCC	OR (95% CIs)	1.05 (0.96~1.15)	1.03 (0.93~1.14)	1.15 (0.95~1.39)	1.00 (0.88~1.14)	1.03 (0.92~1.17)	1.13 (0.95~1.34)
	Pa	0.263	0.568	0.155	0.970	0.575	0.160
	Case/control	1934/3951	1934/3951	1934/3951	1934/3951	1934/3951	1934/3951
	Study number	15	15	15	15	15	15
HNSCC	OR (95% CIs)	0.95 (0.89~1.01)	0.96 (0.89~1.03)	0.94 (0.82~1.09)	0.94 (0.87~1.02)	0.93 (0.86~1.01)	0.95 (0.83~1.09)
	Pa	0.112	0.247	0.408	0.131	0.102	0.459
	Case/control	4671/4961	4671/4961	4671/4961	4671/4961	4671/4961	4671/4961
	Study number	18	18	18	18	18	18
LSCC	OR (95% CIs)	1.00 (0.93~1.08)	1.00 (0.92~1.09)	1.04 (0.88~1.24)	0.97 (0.87~1.07)	0.98 (0.89~1.09)	1.06 (0.90~1.25)
	Pa	0.940	0.973	0.616	0.526	0.741	0.485
	Case/control	2574/5421	2574/5421	2574/5421	2574/5421	2574/5421	2574/5421
	Study number	15	15	15	15	15	15
SSCC	OR (95% CIs)	0.91 (0.70~1.19)	0.94 (0.69~1.28)	0.83 (0.46~1.49)	0.94 (0.64~1.36)	0.91 (0.64~1.30)	0.86 (0.49~1.51)
	Pa	0.493	0.688	0.532	0.728	0.605	0.597
	Case/control	177/475	177/475	177/475	177/475	177/475	177/475
	Study number	3	3	3	3	3	3

ESCC oesophageal squamous cell carcinoma, HNSCC head and neck squamous cell carcinoma, LSCC lung squamous cell carcinoma, SSCC skin squamous cell carcinoma, OR odds ratio, CIs confidence intervals, P_a, P value of association test

Bludy D allele G vs. A OR (95%, C) % ESCC 0 <t< th=""><th></th><th></th><th></th><th></th></t<>				
Abba (2004) Cal (2006) Dura (2013) Li (2010) Li (20	Study ID	allele G vs. A	OR (95% CI)	% Weight
Cabeiguenne (2001) Cho (2006) Harth (2007) Jourenkova (1999a) Jourenkova (1999b) Kelders (2002) Leichsenrig (2006) Leichsenrig (2006) Leichsenrig (2006) Leichsenrig (2006) Leichsenrig (2000) Olshan (2000) Olshan (2000) Peters (2000) Peters (2000) Peters (2000) Distan (2000)	Abbas (2004) Gai (2006) Dura (2013) Jain (2006) Li (2010) Li (2010) Malik (2010) Malik (2011) Matejcic (2011) Matejcic (2011) Matejcic (2011) Tan (2000) van (1999) Zendehdel (2009) Subtotal (I-squared = 21.5%, p = 0.214)		0.86 (0.62, 1.19) 0.82 (0.67, 1.25) 0.87 (0.55, 1.37) 1.11 (0.81, 1.53) 0.89 (0.55, 1.34) 0.81 (0.47, 1.40) 1.44 (0.99, 2.10) 0.94 (0.77, 1.15) 1.07 (0.85, 1.34) 1.03 (0.71, 1.50) 1.41 (1.03, 1.92) 1.02 (0.66, 1.50) 1.41 (0.93, 4.77) 1.45 (1.02, 2.06)	1.77 1.86 0.87 1.57 1.06 0.97 4.34 3.12 1.18 1.42 1.11 0.15 1.10
Dzian (2012) Jourenkova (1998) Kihara (1999) Larsen (2006) Larsen (2006) Lewis (2002) Miler (2006) Miler (2007) Schneider (2002) To (1998) Subtotal (I-squared = 45.1%, p = 0.030) SSCC Fryer (2005) Leite (2007) Subtotal (I-squared = 7.9%, p = 0.338) Leite (2007) Subtotal (I-squared = 7.9%, p = 0.338) Leite (2007) Subtotal (I-squared = .%, p = .) Leite (squared = .%, p = .)	Cabelguenne (2001) Cho (2006) Evans (2004) Harth (2008) Jourenkova (1999a) Jourenkova (1999b) Kelders (2002) Leichsenring (2006) Li (2007) McVVilliams (2000) Olshan (2000) Olshan (2000) Olshan (2000) Ruwai (2009) Ruwai (2011) Soucek (2010) To (2002) Subtotal (I-squared = 0.0%, p = 0.879)		$\begin{array}{c} 1.03 \ (0.79, 1.36) \\ 1.01 \ (0.80, 1.28) \\ 1.20 \ (0.85, 1.71) \\ 1.22 \ (0.72, 1.44) \\ 1.27 \ (0.76, 2.14) \\ 1.29 \ (0.77, 2.18) \\ 0.94 \ (0.82, 1.09) \\ 1.06 \ (0.74, 1.51) \\ 0.96 \ (0.67, 1.38) \\ 0.82 \ (0.43, 1.60) \\ 0.82 \ (0.43, 1.60) \\ 0.82 \ (0.68, 1.11) \\ 1.00 \ (0.68, 1.17) \\ 0.82 \ (0.68, 1.10) \\ 0.95 \ (0.64, 1.42) \\ 0.95 \ (0.64, 1.42) \\ 0.95 \ (0.64, 1.42) \\ 0.95 \ (0.64, 1.42) \\ 0.95 \ (0.64, 1.42) \\ 0.95 \ (0.64, 1.42) \\ 0.95 \ (0.64, 1.42) \\ 0.95 \ (0.74, 1.29) \\ 0.95 \ (0.74, 1$	2.25 2.25 2.98 1.23 1.38 0.56 0.55 8.59 1.27 1.35 0.42 2.70 6.99 2.87 4.08 1.08 1.92
Fryer (2005) 1.03 (0.74, 1.43) 1.53 Leite (2007) 0.84 (0.45, 1.57) 0.48 Ramsay (2001) 0.56 (0.26, 1.20) 0.43 Subtotal (I-squared = 7.9%, p = 0.338) 0.91 (0.70, 1.19) 2.44 UADTSCC 1.05 (0.81, 1.37) 2.34 Subtotal (I-squared = .%, p = .) 1.05 (0.81, 1.37) 2.34	Dzian (2012) Jourenkova (1998) Kihara (1999) Larsen (2006) Lewis (2002) Liang (2005) Miller (2003) Risch (2001) Ryberg (1997) Schneider (2004) Stücker (2002) To (1999) To (1999)		$\begin{array}{c} 1.04 \ (0.71, 1.51) \\ 1.34 \ (0.90, 1.97) \\ 0.86 \ (0.70, 1.05) \\ 0.89 \ (0.71, 1.12) \\ 0.85 \ (0.47, 1.53) \\ 0.91 \ (0.60, 1.37) \\ 0.96 \ (0.32, 1.14) \\ 0.88 \ (0.61, 1.27) \\ 1.08 \ (0.22, 1.14) \\ 0.88 \ (0.61, 1.27) \\ 1.08 \ (0.22, 1.41) \\ 1.08 \ (0.22, 1$	1.16 0.93 4.50 3.38 0.54 1.05 6.24 1.32 2.11 0.76 2.54 1.42 0.76 0.88
Soya (2007) 1.05 (0.81, 1.37) 2.34 Subtotal (I-squared = .%, p = .) 1.05 (0.81, 1.37) 2.34	Fryer (2005) Leite (2007) Ramsay (2001)		0.84 (0.45, 1.57) 0.56 (0.26, 1.20)	0.48 0.43
	Soya (2007)	*		2.34 2.34
Overalii (I-squared = 15.5%, p = 0.174) 0.99 (0.95, 1.03) 100.00	Overall (I-squared = 15.5%, p = 0.174)	4	0.99 (0.95, 1.03)	100.00
	Data of subgroup analysis by SCC type (allele mo	odel)		





Similarly, inconsistent results regarding an association between the *GSTP1* rs1695 A/G polymorphism and LSCC risk have been reported in different races and geographical locations [24, 31, 33, 34, 37, 40, 45, 47, 52, 56, 57, 60, 63]. Here, we failed to detect a positive correlation between *GSTP1* rs1695 and LSCC susceptibility, consistent with the prior meta-analysis of Feng in 2013 [66] and Xu in 2014 [67].

Head and neck cancer comprises cancers of the mouth, nose, sinuses, salivary glands, throat, and lymph nodes in the neck, and HNSCC is the major pathologic type [68]. In 2012, Lang et al. enrolled 28 case-control studies to perform a meta-analysis regarding the genetic effect of the *GSTP1* rs1695 A/G polymorphism on overall head and neck cancer [69]. The authors were unable to identify a positive association between the *GSTP1* rs1695 A/G polymorphism and the risk of overall head and neck cancer. Nevertheless, the potential role of *GSTP1* rs1695 in the susceptibility to HNSCC was not assessed. Therefore, we performed a subgroup meta-analysis of HNSCC involving

18 case-control studies, but did not identify an association between *GSTP1* rs1695 and HNSCC risk.

SSCC, SBCC (skin basal cell carcinoma) and (MM malignant melanoma) are the three main types of cutaneous cancer [4]. Herein, we did not identify an association between the *GSTP1* rs1695 A/G polymorphism and SSCC risk, consistent with the prior meta-analyses regarding the correlation between *GSTP1* rs1695 and the susceptibility to cutaneous cancer in 2015 [70, 71].

Human GST family genes, mainly including *GSTA* (glutathione S-transferase alpha), *GSTM1* (glutathione S-transferase mu 1), *GSTT1* (glutathione S-transferase theta 1) and *GSTP1*, encode phase II enzymes and are thus important for the body defence, metabolic detoxification of mutagens or chemical drugs, or cellular elimination of carcinogens [9, 10]. The rs1695 A/G polymorphism within the *GSTP1* gene can result in the substitution of Ile (isoleucine) for Val (valine) at amino acid position 105, which may lower the cytosolic enzyme activity of *GSTP1* protein [72, 73]. Although significant associations were not

obtained in our overall meta-analysis or subgroup analyses by pathological type, we cannot rule out the potential genetic effect of the *GSTP1* rs1695 A/G polymorphism.

There are still some limitations to our meta-analysis that should be clarified. Even though our findings were considered reliable by our sensitivity analysis and publication bias assessment, more eligible investigations are still warranted to further enhance the statistical power. We note that population-based controls were not utilized in each case-control study. The currently available data of genotypic and allelic frequency from the on-line databases led us to only target the rs1695 polymorphism of the *GSTP1* gene. Other possible functional polymorphisms of the *GSTP1* gene, such as rs1138272, or relative haplotypes will be important to examine in the future. We should also pay attention to the genetic relationship between GSTP1/GSTM1/GSTT1 polymorphisms and the risk of SCC.

Conclusion

In general, based on the currently published data, the *GSTP1* gene rs1695 polymorphism is not associated with the susceptibility to overall SCC diseases, including ESCC, HNSCC, LSCC, and skin SCC. The confirmation or refutation of this conclusion merits further evidence.

Abbreviations

AHR: Aryl hydrocarbon receptor; CADM1: Cell adhesion molecule 1; diASA-AMP: Di-allele-specific- amplification with artificially modified primers assay; Embase: Excerpta Medica Database; ESCC: Oesophageal squamous cell carcinoma; GST: Glutathione S-transferase; GSTA: Glutathione S-transferase alpha; GSTM1: Glutathione S-transferase mu 1; GSTP1: Glutathione Stransferase pi 1; GSTT1: Glutathione S-transferase theta 1; GWAS: Genomewide association study; HB: Hospital-based; HNSCC: Head and neck squamous cell carcinoma; HWE: Hardy-Weinberg equilibrium; KLF5: Kruppel like factor 5; LSCC: Lung squamous cell carcinoma; MM: Malignant melanoma; OSCC: Oral squamous cell carcinoma; PB: Population-based; PCR: Polymerase chain reaction; PCR-RFLP: Polymerase chain reactionrestriction fragment length polymorphism; SBCC: Skin basal cell carcinoma; SCC: Squamous cell carcinoma; SEC16A: SEC16 homolog A, endoplasmic reticulum export factor; SSCC: Skin squamous cell carcinoma; SSCP: Singlestranded conformational polymorphism; UADTSCC: Upper aerodigestive tract squamous cell carcinoma; WOS: Web of Science

Acknowledgements

Not applicable.

Funding

This study was supported in part by a grant of Science Foundation from Tianjin Municipal Commission of Health and Family Planning (2015KY11).

Availability of data and materials

The datasets used and/or analyzed during the current study are available from the corresponding author on reasonable request.

Authors' contributions

SW and ZB designed the study. SW, JZ and FJ extracted, analyzed, and interpreted the data. SW and ZB drafted the manuscript. All authors read and approved the final version of the manuscript.

Ethics approval and consent to participate

Not applicable.

Consent for publication

Not applicable.

Competing interests

The authors declare that they have no competing interests.

Publisher's Note

Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.

Author details

¹Department of Plastic and Burn Surgery, Tianjin First Center Hospital, Tianjin 300192, China. ²Department of Urology Surgery, Tianjin First Center Hospital, Tianjin 300192, China.

Received: 18 July 2018 Accepted: 11 January 2019 Published online: 21 January 2019

References

- Que SKT, Zwald FO, Schmults CD. Cutaneous squamous cell carcinoma: management of advanced and high-stage tumors. J Am Acad Dermatol. 2018;78(2):249–61.
- Wang C, Wang J, Chen Z, Gao Y, He J. Immunohistochemical prognostic markers of esophageal squamous cell carcinoma: a systematic review. Chin J Cancer. 2017;36(1):65.
- Bann DV, Deschler DG, Goyal N. Novel Immunotherapeutic Approaches for Head and Neck Squamous Cell Carcinoma. Cancers (Basel). 2016;8(10).
- Liu N, Liu GJ, Liu J. Genetic association between TNF-alpha promoter polymorphism and susceptibility to squamous cell carcinoma, basal cell carcinoma, and melanoma: a meta-analysis. Oncotarget. 2017;8(32):53873–85.
- Zhang X, He R, Ren F, Tang R, Chen G. Association of miR-146a rs2910164 polymorphism with squamous cell carcinoma risk: a meta-analysis. J buon. 2015;20(3):829–41.
- Yu H, Li H, Zhang J, Liu G. Influence of MDM2 polymorphisms on squamous cell carcinoma susceptibility: a meta-analysis. Onco Targets Ther. 2016;9: 6211–24.
- Chahal HS, Lin Y, Ransohoff KJ, Hinds DA, Wu W, Dai HJ, Qureshi AA, Li WQ, Kraft P, Tang JY, et al. Genome-wide association study identifies novel susceptibility loci for cutaneous squamous cell carcinoma. Nat Commun. 2016;7:12048.
- Chang J, Wei L, Miao X, Yu D, Tan W, Zhang X, Wu C, Lin D. Two novel variants on 13q22.1 are associated with risk of esophageal squamous cell carcinoma. Cancer Epidemiol Biomark Prev. 2015;24(11):1774–80.
- Schnekenburger M, Karius T, Diederich M. Regulation of epigenetic traits of the glutathione S-transferase P1 gene: from detoxification toward cancer prevention and diagnosis. Front Pharmacol. 2014;5:170.
- Marchewka Z, Piwowar A, Ruzik S, Dlugosz A. Glutathione S transferases class pi and mi and their significance in oncology. Postepy Hig Med Dosw (Online). 2017;71(0):541–50.
- Yuan Y, Qian ZR, Sano T, Asa SL, Yamada S, Kagawa N, Kudo E. Reduction of GSTP1 expression by DNA methylation correlates with clinicopathological features in pituitary adenomas. Mod Pathol. 2008;21(7):856–65.
- Hollman AL, Tchounwou PB, Huang HC. The association between Geneenvironment interactions and diseases involving the human GST superfamily with SNP variants. Int J Environ Res Public Health. 2016;13(4): 379.
- Karaca S, Karaca M, Cesuroglu T, Erge S, Polimanti R. GSTM1, GSTP1, and GSTT1 genetic variability in Turkish and worldwide populations. Am J Hum Biol. 2015;27(3):310–6.
- Abbas A, Delvinquiere K, Lechevrel M, Lebailly P, Gauduchon P, Launoy G, Sichel F. GSTM1, GSTT1, GSTP1 and CYP1A1 genetic polymorphisms and susceptibility to esophageal cancer in a French population: different pattern of squamous cell carcinoma and adenocarcinoma. World J Gastroenterol. 2004;10(23):3389–93.
- van Lieshout EM, Roelofs HM, Dekker S, Mulder CJ, Wobbes T, Jansen JB, Peters WH. Polymorphic expression of the glutathione S-transferase P1 gene and its susceptibility to Barrett's esophagus and esophageal carcinoma. Cancer Res. 1999;59(3):586–9.
- 16. Zendehdel K, Bahmanyar S, McCarthy S, Nyren O, Andersson B, Ye W. Genetic polymorphisms of glutathione S-transferase genes GSTP1, GSTM1,

and GSTT1 and risk of esophageal and gastric cardia cancers. Cancer Causes Control. 2009;20(10):2031–8.

- Song Y, Du Y, Zhou Q, Ma J, Yu J, Tao X, Zhang F. Association of GSTP1 lle105Val polymorphism with risk of esophageal cancer: a meta-analysis of 21 case-control studies. Int J Clin Exp Med. 2014;7(10):3215–24.
- Tan X, Chen M. Association between glutathione S-transferases P1 lle105Val polymorphism and susceptibility to esophageal cancer: evidence from 20 case-control studies. Mol Biol Rep. 2015;42(2):399–408.
- Moher D, Liberati A, Tetzlaff J, Altman DG. Preferred reporting items for systematic reviews and meta-analyses: the PRISMA statement. PLoS Med. 2009;6(7):e1000097.
- Cabelguenne A, Loriot MA, Stucker I, Blons H, Koum-Besson E, Brasnu D, Beaune P, Laccourreye O, Laurent-Puig P, De Waziers I. Glutathioneassociated enzymes in head and neck squamous cell carcinoma and response to cisplatin-based neoadjuvant chemotherapy. Int J Cancer. 2001; 93(5):725–30.
- Cai L, Mu LN, Lu H, Lu QY, You NC, Yu SZ, Le AD, Zhao J, Zhou XF, Marshall J, et al. Dietary selenium intake and genetic polymorphisms of the GSTP1 and p53 genes on the risk of esophageal squamous cell carcinoma. Cancer Epidemiol Biomark Prev. 2006;15(2):294–300.
- 22. Cho CG, Lee SK, Nam SY, Lee MS, Lee SW, Choi EK, Park HJ, Kim SY. Association of the GSTP1 and NQO1 polymorphisms and head and neck squamous cell carcinoma risk. J Korean Med Sci. 2006;21(6):1075–9.
- Dura P, Salomon J, Te Morsche RH, Roelofs HM, Kristinsson JO, Wobbes T, Witteman BJ, Tan AC, Drenth JP, Peters WH. No role for glutathione Stransferase genotypes in Caucasian esophageal squamous cell or adenocarcinoma etiology: an European case-control study. BMC Gastroenterol. 2013;13:97.
- Dzian A, Halasova E, Matakova T, Kavcova E, Smolar M, Dobrota D, Hamzik J, Mistuna D. Lung adenocarcinoma and squamous cell carcinoma in association with genetic polymorphisms of GSTs in Slovak population. Neoplasma. 2012;59(2):160–7.
- Evans AJ, Henner WD, Eilers KM, Montalto MA, Wersinger EM, Andersen PE, Cohen JI, Everts EC, McWilliams JE, Beer TM. Polymorphisms of GSTT1 and related genes in head and neck cancer risk. Head Neck. 2004;26(1):63–70.
- Fryer AA, Ramsay HM, Lovatt TJ, Jones PW, Hawley CM, Nicol DL, Strange RC, Harden PN. Polymorphisms in glutathione S-transferases and nonmelanoma skin cancer risk in Australian renal transplant recipients. Carcinogenesis. 2005;26(1):185–91.
- Harth V, Schafer M, Abel J, Maintz L, Neuhaus T, Besuden M, Primke R, Wilkesmann A, Thier R, Vetter H, et al. Head and neck squamous-cell cancer and its association with polymorphic enzymes of xenobiotic metabolism and repair. J Toxicol Environ Health A. 2008;71(13–14):887–97.
- Jain M, Kumar S, Rastogi N, Lal P, Ghoshal UC, Tiwari A, Pant MC, Baiq MQ, Mittal B. GSTT1, GSTM1 and GSTP1 genetic polymorphisms and interaction with tobacco, alcohol and occupational exposure in esophageal cancer patients from North India. Cancer Lett. 2006;242(1):60–7.
- Jourenkova-Mironova N, Voho A, Bouchardy C, Wikman H, Dayer P, Benhamou S, Hirvonen A. Glutathione S-transferase GSTM1, GSTM3, GSTP1 and GSTT1 genotypes and the risk of smoking-related oral and pharyngeal cancers. Int J Cancer. 1999a;81(1):44–8.
- Jourenkova-Mironova N, Voho A, Bouchardy C, Wikman H, Dayer P, Benhamou S, Hirvonen A. Glutathione S-transferase GSTM3 and GSTP1 genotypes and larynx cancer risk. Cancer Epidemiol Biomark Prev. 1999b; 8(2):185–8.
- Jourenkova-Mironova N, Wikman H, Bouchardy C, Voho A, Dayer P, Benhamou S, Hirvonen A. Role of glutathione S-transferase GSTM1, GSTM3, GSTP1 and GSTT1 genotypes in modulating susceptibility to smokingrelated lung cancer. Pharmacogenetics. 1998;8(6):495–502.
- Kelders WP, Oude Ophuis MB, Roelofs HM, Peters WH, Manni JJ. The association between glutathione S-transferase P1 genotype and plasma level in head and neck cancer. Laryngoscope. 2002;112(3):462–6.
- Kihara M, Kihara M, Noda K. Lung cancer risk of the GSTM1 null genotype is enhanced in the presence of the GSTP1 mutated genotype in male Japanese smokers. Cancer Lett. 1999;137(1):53–60.
- Larsen JE, Colosimo ML, Yang IA, Bowman R, Zimmerman PV, Fong KM. CYP1A1 Ile462Val and MPO G-463A interact to increase risk of adenocarcinoma but not squamous cell carcinoma of the lung. Carcinogenesis. 2006;27(3):525–32.
- Leichsenring A, Losi-Guembarovski R, Maciel ME, Losi-Guembarovski A, Oliveira BW, Ramos G, Cavalcanti TC, Bicalho MG, Cavalli IJ, Colus IM, et al.

CYP1A1 and GSTP1 polymorphisms in an oral cancer case-control study. Braz J Med Biol Res. 2006;39(12):1569–74.

- Leite JL, Morari EC, Granja F, Campos GM, Guilhen AC, Ward LS. Influence of the glutathione s-transferase gene polymorphisms on the susceptibility to basal cell skin carcinoma. Rev Med Chil. 2007;135(3):301–6.
- Lewis SJ, Cherry NM, Niven RM, Barber PV, Povey AC. GSTM1, GSTT1 and GSTP1 polymorphisms and lung cancer risk. Cancer Lett. 2002;180(2):165–71.
- Li D, Dandara C, Parker MI. The 341C/T polymorphism in the GSTP1 gene is associated with increased risk of oesophageal cancer. BMC Genet. 2010;11:47.
- Li DH, Wang LE, Chang P, El-Naggar AK, Sturgis EM, Wei QY. In vitro benzo a pyrene diol epoxide-induced DNA adducts and risk of squamous cell carcinoma of head and neck. Cancer Res. 2007;67(12):5628–34.
- Liang G, Pu Y, Yin L. Rapid detection of single nucleotide polymorphisms related with lung cancer susceptibility of Chinese population. Cancer Lett. 2005;223(2):265–74.
- Liu R, Yin L, Pu Y, Li Y, Liang G, Zhang J, Li X. Functional alterations in the glutathione S-transferase family associated with enhanced occurrence of esophageal carcinoma in China. J Toxicol Environ Health A. 2010;73(7):471–82.
- Malik MA, Upadhyay R, Mittal RD, Zargar SA, Mittal B. Association of xenobiotic metabolizing enzymes genetic polymorphisms with esophageal cancer in Kashmir Valley and influence of environmental factors. Nutr Cancer. 2010;62(6):734–42.
- Matejcic M, Li D, Prescott NJ, Lewis CM, Mathew CG, Parker MI. Association of a deletion of GSTT2B with an altered risk of oesophageal squamous cell carcinoma in a south African population: a case-control study. PLoS One. 2011;6(12):e29366.
- McWilliams JE, Evans AJ, Beer TM, Andersen PE, Cohen JI, Everts EC, Henner WD. Genetic polymorphisms in head and neck cancer risk. Head Neck. 2000; 22(6):609–17.
- Miller DP, Asomaning K, Liu G, Wain JC, Lynch TJ, Neuberg D, Su L, Christiani DC. An association between glutathione S-transferase P1 gene polymorphism and younger age at onset of lung carcinoma. Cancer. 2006; 107(7):1570–7.
- Moaven O, Raziee HR, Sima HR, Ganji A, Malekzadeh R, A'Rabi A, Abdollahi A, Memar B, Sotoudeh M, Naseh H, et al. Interactions between glutathione-S-transferase M1, T1 and P1 polymorphisms and smoking, and increased susceptibility to esophageal squamous cell carcinoma. Cancer Epidemiol. 2010;34(3):285–90.
- Nazar-Stewart V, Vaughan TL, Stapleton P, Van Loo J, Nicol-Blades B, Eaton DL. A population-based study of glutathione S-transferase M1, T1 and P1 genotypes and risk for lung cancer. Lung Cancer. 2003;40(3):247–58.
- Olshan AF, Weissler MC, Watson MA, Bell DA. GSTM1, GSTT1, GSTP1, CYP1A1, and NAT1 polymorphisms, tobacco use, and the risk of head and neck cancer. Cancer Epidemiol Biomark Prev. 2000;9(2):185–91.
- Oude Ophuis MB, Roelofs HM, van den Brandt PA, Peters WH, Manni JJ. Polymorphisms of the glutathione S-transferase P1 gene and head and neck cancer susceptibility. Head Neck. 2003;25(1):37–43.
- Peters ES, McClean MD, Marsit CJ, Luckett B, Kelsey KT. Glutathione S-transferase polymorphisms and the synergy of alcohol and tobacco in oral, pharyngeal, and laryngeal carcinoma. Cancer Epidemiol Biomark Prev. 2006;15(11):2196–202.
- Ramsay HM, Harden PN, Reece S, Smith AG, Jones PW, Strange RC, Fryer AA. Polymorphisms in glutathione S-transferases are associated with altered risk of nonmelanoma skin cancer in renal transplant recipients: a preliminary analysis. J Invest Dermatol. 2001;117(2):251–5.
- Risch A, Wikman H, Thiel S, Schmezer P, Edler L, Drings P, Dienemann H, Kayser K, Schulz V, Spiegelhalder B, et al. Glutathione-S-transferase M1, M3, T1 and P1 polymorphisms and susceptibility to non-small-cell lung cancer subtypes and hamartomas. Pharmacogenetics. 2001;11(9):757–64.
- Rossini A, Rapozo DCM, Soares Lima SC, Guimarães DP, Ferreira MA, Teixeira R, Kruel CDP, Barros SGS, Andreollo NA, Acatauassú R, et al. Polymorphisms of GSTP1 and GSTT1, but not of CYP2A6, CYP2E1 or GSTM1, modify the risk for esophageal cancer in a western population. Carcinogenesis. 2007;28(12): 2537–42.
- Ruwali M, Pant MC, Shah PP, Mishra BN, Parmar D. Polymorphism in cytochrome P450 2A6 and glutathione S-transferase P1 modifies head and neck cancer risk and treatment outcome. Mutat Res. 2009;669(1–2):36–41.
- Ruwali M, Singh M, Pant MC, Parmar D. Polymorphism in glutathione Stransferases: susceptibility and treatment outcome for head and neck cancer. Xenobiotica. 2011;41(12):1122–30.
- Ryberg D, Skaug V, Hewer A, Phillips DH, Harries LW, Wolf CR, Ogreid D, Ulvik A, Vu P, Haugen A. Genotypes of glutathione transferase M1 and P1

and their significance for lung DNA adduct levels and cancer risk. Carcinogenesis. 1997;18(7):1285–9.

- Schneider J, Bernges U, Philipp M, Woitowitz HJ. GSTM1, GSTT1, and GSTP1 polymorphism and lung cancer risk in relation to tobacco smoking. Cancer Lett. 2004;208(1):65–74.
- Soucek P, Susova S, Mohelnikova-Duchonova B, Gromadzinska J, Moraviec-Sztandera A, Vodicka P, Vodickova L. Polymorphisms in metabolizing enzymes and the risk of head and neck squamous cell carcinoma in the Slavic population of the Central Europe. Neoplasma. 2010;57(5):415–21.
- Soya SS, Vinod T, Reddy KS, Gopalakrishnan S, Adithan C. Genetic polymorphisms of glutathione-S-transferase genes (GSTM1, GSTT1 and GSTP1) and upper aerodigestive tract cancer risk among smokers, tobacco chewers and alcoholics in an Indian population. Eur J Cancer. 2007;43(18): 2698–706.
- Stücker I, Hirvonen A, De Waziers I, Cabelguenne A, Mitrunen K, Cénée S, Koum-Besson E, Hémon D, Beaune P, Loriot MA. Genetic polymorphisms of glutathione S-transferases as modulators of lung cancer susceptibility. Carcinogenesis. 2002;23(9):1475–81.
- Tan W, Song N, Wang GQ, Liu Q, Tang HJ, Kadlubar FF, Lin DX. Impact of genetic polymorphisms in cytochrome P450 2E1 and glutathione Stransferases M1, T1, and P1 on susceptibility to esophageal cancer among high-risk individuals in China. Cancer Epidemiol Biomark Prev. 2000;9(6):551–6.
- To-Figueras J, Gene M, Gomez-Catalan J, Pique E, Borrego N, Caballero M, Cruellas F, Raya A, Dicenta M, Corbella J. Microsomal epoxide hydrolase and glutathione S-transferase polymorphisms in relation to laryngeal carcinoma risk. Cancer Lett. 2002;187(1–2):95–101.
- To-Figueras J, Gene M, Gomez-Catalan J, Pique E, Borrego N, Carrasco JL, Ramon J, Corbella J. Genetic polymorphism of glutathione S-transferase P1 gene and lung cancer risk. Cancer Causes Control. 1999;10(1):65–70.
- Song Q, Jiang D, Wang H, Huang J, Liu Y, Xu C, Hou Y. Chromosomal and genomic variations in esophageal squamous cell carcinoma: a review of technologies, applications, and prospections. J Cancer. 2017;8(13):2492–500.
- Luo LN, He LJ, Gao XY, Huang XX, Shan HB, Luo GY, Li Y, Lin SY, Wang GB, Zhang R, et al. Evaluation of preoperative staging for esophageal squamous cell carcinoma. World J Gastroenterol. 2016;22(29):6683–9.
- Feng X, Zhou HF, Zheng BS, Shi JJ, Luo C, Qin JJ. Association of glutathione S-transferase P1 gene polymorphism with the histological types of lung cancer: a meta-analysis. Mol Biol Rep. 2013;40(3):2439–47.
- Xu CH, Wang Q, Zhan P, Qian Q, Yu LK. GSTP1 Ile105Val polymorphism is associated with lung cancer risk among Asian population and smokers: an updated meta-analysis. Mol Biol Rep. 2014;41(7):4199–212.
- Szyszko TA, Cook GJR. PET/CT and PET/MRI in head and neck malignancy. Clin Radiol. 2018;73(1):60–9.
- Lang J, Song X, Cheng J, Zhao S, Fan J. Association of GSTP1 lle105Val Polymorphism and Risk of Head and Neck Cancers: A Meta-Analysis of 28 Case-Control Studies. PLoS One. 2012;7(11):e48132.
- Lei Z, Liu T, Li X, Xu X, Fan D. Contribution of glutathione S-transferase gene polymorphisms to development of skin cancer. Int J Clin Exp Med. 2015; 8(1):377–86.
- Zhou CF, Ma T, Zhou DC, Shen T, Zhu QX. Association of glutathione Stransferase pi (GSTP1) Ile105Val polymorphism with the risk of skin cancer: a meta-analysis. Arch Dermatol Res. 2015;307(6):505–13.
- Watson MA, Stewart RK, Smith GB, Massey TE, Bell DA. Human glutathione S-transferase P1 polymorphisms: relationship to lung tissue enzyme activity and population frequency distribution. Carcinogenesis. 1998;19(2):275–80.
- Zhong SL, Zhou SF, Chen X, Chan SY, Chan E, Ng KY, Duan W, Huang M. Relationship between genotype and enzyme activity of glutathione Stransferases M1 and P1 in Chinese. Eur J Pharm Sci. 2006;28(1–2):77–85.

Ready to submit your research? Choose BMC and benefit from:

- fast, convenient online submission
- thorough peer review by experienced researchers in your field
- rapid publication on acceptance
- support for research data, including large and complex data types
- gold Open Access which fosters wider collaboration and increased citations
- maximum visibility for your research: over 100M website views per year

At BMC, research is always in progress.

Learn more biomedcentral.com/submissions

