

Dickkopf-3 and β -catenin play opposite roles in the Wnt/ β -catenin pathway during the abnormal subchondral bone formation of human knee osteoarthritis

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Abstract. Osteoarthritis (OA) is condition which poses a main concern to the aging population and its severity is expected to increase with the increasing life expectancy. In the future, several possible targets for OA treatment need to be defined. Dickkopf-related protein 3 (DKK3) is an atypical member of the Wnt-antagonistic dickkopf-related protein (DKK) family. The availability of research into the role of DKK3 in the abnormal remodeling of subchondral bone in human knee joints is currently limited. Thus, the aim of the present study was the evaluation of DKK3 expression in the abnormal bone remodeling of subchondral bone in human knee OA in order to clarify the role of DKK3 in subchondral bone remodeling and to acknowledge its potential relevance to β -catenin. In total, 38 specimens were collected from osteotomies of the medial tibial plateau of the human knee. The patient samples were then divided into the normal, mild, moderate and severe symptom groups, according to the Osteoarthritis Research Society International (OARSI) score. Following hematoxylin and eosin (H&E) and Safranin O-fast green staining for alkaline phosphatase (AZO method), changes in the distribution and number of osteocytes in the subchondral bone and the degree of sclerosis of the subchondral bone were observed. Immunohistochemical staining, immunofluorescence, western blot analysis and reverse-transcription quantitative PCR (RT-qPCR) were used for the detection of DKK3 and β -catenin expression level changes in osteoblasts in the subchondral bone of the medial tibial plateau. H&E and alkaline phosphatase staining revealed that the total number of osteocytes in the

subchondral bone increased with the severity of the disease. The samples were also evaluated using Safranin O-Fast Green staining and were attributed a score according to the OARSI scoring system: The scoring number and cartilage damage increased along with OA severity. Immunohistochemistry and immunofluorescence assays demonstrated that β -catenin expression in osteocytes increased from mild to moderate, whereas DKK3 expression decreased with the development of arthritis from normal, mild to moderate. According to the results of western blot analysis, β -catenin expression was higher in moderate OA and then decreased in severe OA. On the other hand, the DKK3 levels decreased along with the progression from normal, mild to moderate OA. The results of RT-qPCR demonstrated that β -catenin and DKK3 gene expression differed with the degree of OA. On the whole, the present study demonstrates that DKK3 and β -catenin may play opposite roles in OA subchondral bone remodeling.

Introduction

The most common orthopedic diseases include osteoarthritis (OA) and osteoporosis. OA is defined as a joint disease, characterized by cartilage degeneration, abnormal remodeling of subchondral bone and joint pain (1), and occurs in aging populations. However, previous research has demonstrated that subchondral bone hardens and cartilage degenerates during disease progression. Anderson-MacKenzie *et al* (2) reported that the role of subchondral bone was mainly to reduce impact forces and mitigate cartilage damage. In addition, subchondral bone density and cartilage thickness are positively associated in knees without OA, and negatively associated in knees with OA, in support of the subchondral bone/chondral functional unit theory. These conclusions also contribute to the improvement of the understanding of the pathophysiology of OA (3). Furthermore, Neve *et al* (4) studied osteoblasts, which are mononuclear mesenchymal stem cell-derived cells responsible for osteogenesis and mineralization in early bone formation and subsequent bone remodeling, while OA osteoblasts exhibit an abnormal metabolic behavior. Maruotti *et al* (5) revealed that osteoblast disorders play a crucial role in the pathogenesis

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of OA, stimulating the osteoblast-induced production of large transcription factor quantities, growth factors and other proteins involved in OA pathogenesis. In addition, Di Pompo *et al* (6) previously reported that osteoblasts promoted the secretion of osteoclastic cytokines and inflammatory mediators in an acidic environment (pH 6.8), further exacerbating OA.

The signaling pathways form a complex network of molecular interactions in osteoblasts. It has been reported that the Wnt signaling pathway is involved in OA, in which signaling molecules and regulatory factors are aberrantly activated or inhibited (7). The activation of the classical Wnt/ β -catenin signaling pathway increases subchondral bone remodeling and osteophyte formation, which are two pathological processes in bone also associated with OA progression (8). Similarly, decreased levels of Wnt activity are important for maintaining the viability of osteocytes (both chondrocytes and osteoblasts); however, excessive activation has been found to be associated with joint damage (9).

Osteoblasts and their precursors can control various Wnt ligands and are involved in the regulation of bone remodeling; the Wnt signaling may regulate chondrocyte and osteoblast differentiation and function via classical (β -catenin-dependent) or non-classical Wnt signaling pathways, as well as cross-linking with other signaling pathways, affecting cartilage and bone metabolism. In addition, the excessive activation of the Wnt signaling pathway may exacerbate cartilage OA-like degeneration (10,11). β -catenin has been reported to be essential for the differentiation of osteoblasts, and the overexpression of constitutively active β -catenin forms may also lead to the development of OA, characterized by increased chondrocyte hypertrophy (12,13). Thus, the dysregulation of the Wnt/ β -catenin signaling pathway may lead to OA.

The Dickkopf (DKK) family has been reported to regulate the Wnt signaling pathway by binding to the Wnt receptor complex (14). Chen *et al* (15) demonstrated that the local injection of DKK1 knockout lentivirus into the rat femur alleviated the progression of AONFH; Wnt/ β -catenin signalling was activated, the number of empty fat cavities was reduced, the blood supply was restored and bone formation was significantly increased. Among DKK family members, Dickkopf-related protein 3 (DKK3) is an atypical member of the DKK family of Wnt antagonists; human DKK-1, -2 and -4 are located in the same homology group, whereas DKK3 does not belong to this group (16). Furthermore, the mechanisms of action of DKK3 differ from those of other proteins in the DDK gene family. Lee *et al* (17) demonstrated that DKK3 attenuated β -catenin protein expression and its transcriptional activity via the interaction with β -transducin repeat-containing protein (β -TrCP) and prevented the translocation of β -catenin to the nucleus. DKK3 expression levels in the subchondral bone, as well as its role in osteocyte formation have not yet been fully elucidated. Therefore, the aim of the present study was to investigate the DKK3 expression levels in osteoblasts in subchondral bone from human knees with OA, in order to clarify the role of DKK3 in abnormal bone remodeling in subchondral bone and to determine its potential association with β -catenin expression level changes.

Materials and methods

Materials. A total of 38 osteotomy specimens (Table I) were collected from the medial tibial plateau of the knee joint of

patients from January, 2019 to October, 2020 at the Third Department of Orthopedics, General Hospital of Ningxia Medical University (Yinchuan, China). Tissue samples from three cases with a normal and five cases with a mild OA symptomatology were obtained from young patients who underwent lower limb amputations. Tissue samples from 10 cases of moderate OA symptomatology were collected from patients diagnosed with OA and who underwent knee arthroplasty, including single condyle replacement. Tissue samples from 20 cases with severe OA symptomatology were also collected, including a case of tibial plateau osteotomy that was diagnosed with OA in a patient undergoing total knee replacement. Samples were acquired from 16 male, and 22 female patients, aged 20-80 years, with an average age of 62 years. The patients included in the present study had no family history of malignant tumors, hemiplegia, or death from stroke, and no patients had undergone surgery on the ipsilateral hip. In total, 23 patients were diagnosed with hypertension (20 patients had grade I and 3 had grade II hypertension), and none were diagnosed with type 2 diabetes. In addition, 1 patient had a giant cell tumor of the ipsilateral pelvis (Jaffe grade II). During tissue collection, all samples were immediately stored in liquid nitrogen, at -147°C . The sample cohort was divided into two equal parts, following sample collection. The samples in the first group were fixed with neutral formalin for 24 h, decalcified with fast decalcification liquid for 1 week, and then embedded in paraffin, as described below. Subchondral bone was retained after cartilage removal in the other sample group, and stored in a refrigerator at -80°C for later use. The present study was approved by the Ethics Committee of the General Hospital of Ningxia Medical University (approval no. KYLL-2020-20; approval date, January 14, 2021), and written informed consent was obtained from all the enrolled patients.

Experimental methods. The samples were divided into the normal, mild, moderate and severe symptom groups, according to the Osteoarthritis Research Society International (OARSI) (18), and the distribution and sclerosis of medial osteoblasts were observed following hematoxylin and eosin (H&E), Safranin O/fast green staining and alkaline phosphatase (ALP; AZO method) as described below. Along with the increase in the osteoarthritis pathology degree, alterations in the number of subchondral osteoblasts were also observed. Changes in DKK3 and β -catenin expression in the subchondral osteoblasts of the medial tibial plateau were observed and detected using immunohistochemical staining, immunofluorescence, western blot analysis and reverse transcription-quantitative PCR (RT-qPCR) as described below.

Main reagents. Anti-DKK3 antibody (EPR15611; ab186409; rabbit monoclonal) was purchased from Abcam, and β -catenin antibody (ab32572; rabbit monoclonal) and GAPDH antibody (ab9485) were obtained from Abcam. RIPA lysate, 50X cocktail, PMSF (100 mM), phosphorylated protease inhibitor, BCA protein quantitative detection kit, 5X protein loading buffer and the SDS-PAGE gel preparation kit were acquired from Wuhan Google Biotechnology Co., Ltd.; the protein marker was acquired from Thermo Fisher Scientific, Inc. The PVDF membrane was acquired from MilliporeSigma; and TRIzol

Table I. Clinicopathological data of the patients in the present study.

Patient no.	Sex	Age (years)	Severity of symptoms	Left or right knee
1	M	65	Moderate	Right
2	F	62	Severe	Right
3	M	59	Severe	Right
4	F	40	Normal	Left
5	M	68	Severe	Left
6	F	41	Mild	Left
7	M	80	Severe	Right
8	F	72	Moderate	Right
9	M	32	Normal	Right
10	F	66	Severe	Right
11	F	80	Moderate	Left
12	M	80	Severe	Left
13	F	78	Severe	Right
14	F	59	Moderate	Left
15	M	55	Moderate	Right
16	M	63	Severe	Left
17	F	61	Moderate	Right
18	M	69	Severe	Right
19	F	44	Mild	Left
20	M	64	Moderate	Left
21	M	72	Moderate	Right
22	M	78	Severe	Right
23	F	67	Severe	Left
24	M	20	Normal	Left
25	M	62	Severe	Left
26	F	67	Severe	Right
27	F	62	Moderate	Right
28	M	78	Severe	Left
29	F	58	Severe	Left
30	M	70	Severe	Left
31	F	43	Mild	Right
32	M	64	Moderate	Left
33	F	68	Severe	Left
34	M	45	Mild	Right
35	M	70	Severe	Right
36	M	79	Severe	Left
37	M	76	Severe	Right
38	M	40	Mild	Right

Universal and Trizol[®] Reagent was purchased from Invitrogen; Thermo Fisher Scientific, Inc.

Histological analysis. All obtained tissues were rinsed with normal saline three times for 15 min each and fixed in 4% neutral formaldehyde solution for 24 h. Subsequently, the tissues were incubated in a rapid decalcification solution (Beijing Zhuoding Biological Company) for decalcification for 7 days and immersed in PBS solution for 6 h, with PBS replacement every 1 h. Tissues were trimmed to a size of ~5x5x3 mm and placed in a plastic embedding box for subsequent processing. The tissues were sequentially placed in 50,

70, 80, 95 and 100% ethanol for 10 min each at first, and subsequently in a 100% ethanol/xylene (2:1) solution, 100% ethanol/xylene (1:1) solution, 100% ethanol/xylene (1:2) solution, xylene I, II, and III for 10 min per step. The paraffin wax was melted at 60°C in an electric heating constant temperature incubator (constant temperature chamber; Xiaogan Yaguang Medical-electronic Technology Co., Ltd.), placed afterwards on tissue paraffin glass for embedding, and xylene was then used to melt the paraffin wax in order to create a mixed solution. The tissues were then placed consecutively in xylene/paraffin (2:1) solution, xylene/paraffin (1:1) solution, and xylene/paraffin (1:2) solution for 30 min per step. Finally,

the tissues were placed in paraffin I and II in a thermostat at 60°C for 1.5 h, respectively.

H&E staining. Tissue samples were dewaxed by soaking the sections in xylene I and II for 10 min, rinsing with distilled water for 30 sec, then soaking consecutively in 100 (I, II), 90, 80 and 70% alcohol for 5 min, and finally rinsing with tap water for 5 min three times. The samples were stained with hematoxylin (Beijing Solarbio Science & Technology Co., Ltd.) for 5 min at room temperature and rinsed with running water. Samples were differentiated with 5% acetic acid for 1 min, washed with running water for 10 min, stained with eosin (Beijing Solarbio Science & Technology Co., Ltd.) for 1 min at room temperature, and rinsed three times with running water, for 5 min. The slices were dehydrated by placing them in 70, 80, 90 and 100% alcohol and xylene for 1 min each, and were then sealed with neutral glue, and covered with a cover slip. The tissues were examined using a Leica microscope (Leica DM2700 M; Leica Microsystems GmbH).

Safranin O/fast green staining. The dewaxing procedure was performed according to the H&E staining protocol, as described above. The sections were stained with hematoxylin for 5 min and rinsed with tap water for 10 min. The sections were dyed with 0.3% solid green for 3 min and then washed with tap water for 5 min three times. The slices were placed in 1% acetic acid for 5 sec, then immersed in 1% safranin staining solution (Beijing Solarbio Science & Technology Co., Ltd.) for 2 min at room temperature, and then rinsed three times with distilled water for 5 min. Finally, the samples were dehydrated and sealed as process as described above for H&E staining. The samples were examined using a Leica microscope (Leica Microsystems GmbH).

The final score was based on the Safranin O/fast green staining results and the OARSI scoring items. The OARIS score was obtained by multiplying the histological grading of cartilage degeneration (in 6 grades) by the histological staging of cartilage degeneration (in five stages) following Safranin O/fast green staining.

ALP staining (AZO coupling method). The dewaxing procedure was performed as described above for the H&E staining protocol. Firstly, the working fluid was prepared. ALP staining solution B1 (AS-BI staining solution) and alkaline phosphatase staining solution B2 (FBB staining solution) (both stains from Beijing Solarbio Science & Technology Co., Ltd.) were mixed in a volume ratio of 1:1, and the prepared ALP incubation solution was then dropped onto the cut sections. The sections were then incubated in a dark incubator for 20 min at 37°C and washed with water for 5 min. The sections were immersed in nuclear hard red staining solution for counterstaining for 3 min at room temperature. Finally, samples were dehydrated and sealed as described above previously described. The samples were examined using a Leica microscope (Leica Microsystems GmbH).

Immunohistochemical staining. The dewaxing procedure was performed as described above. A total of 20 μ l 0.1% trypsin solution was added to the tissue, incubated in an incubator at 37°C for 15 min, and soaked in PBS for 15 min. Endogenous

peroxidase blocker (Beijing Zhongshan Jinqiao Biological Co., Ltd.) was then added (20 μ l) to the tissue, incubated at 37°C for 10 min, and soaked in PBS solution for 15 min, followed by the addition of goat serum (Beijing Solarbio Science & Technology Co., Ltd.) and incubation at 37°C for 30 min. Afterwards, the primary antibodies [DKK3 antibody (goat anti-rabbit, 1:100, ab186409, Abcam) and β -catenin antibody (goat anti-rabbit, 1:150, ab32572, Abcam)] were added in a dropwise manner to the tissue and incubated overnight in a refrigerator at 4°C. The secondary antibody used was enhanced goat anti-rabbit IgG polymer (1:500, PV-9000, 37°C, 40 min; Beijing Zhongshan Jinqiao Biological Co., Ltd.). The slides were removed warmed to 37°C for 40 min the following day. Immunohistochemical detection reagent 2 (Beijing Zhongshan Jinqiao Biological Co., Ltd.) was then added in a dropwise manner to the tissues. The tissues were incubated in an incubator at 37°C for 30 min and soaked in PBS solution for 15 min. Immunohistochemical detection reagent 3 (Beijing Zhongshan Jinqiao Biological Co., Ltd.) was dropped onto the tissues. The samples were placed in an incubator at 37°C and incubated for 40 min. Subsequently, the samples were removed from the incubator and soaked in PBS solution for 15 min at room temperature. DAB chromogenic solution (Beijing Zhongshan Jinqiao Biological Co., Ltd.) was then added in a dropwise manner to the tissues, and they were observed under an optical microscope (Leica Microsystems GmbH) for 1 min, and soaked in distilled water for 5 min. The cells were then stained with hematoxylin for 5 min at room temperature, differentiated with 5% acetic acid for 1 min, and rinsed with tap water for 10 min. Samples were dehydrated and sealed as described in the H&E staining section above. Finally, analysis was performed based on the experimental results. Tissues were counted in areas of high expression, and with the use of ImageJ (1.8.0) software (National Institutes of Health) was used to determine the intensity of protein expression. Afterwards, the histological score was calculated as the average optical density of positively stained osteoblasts, which equals to the ratio of the overall optical density of the positive cells to the positive area. Positive cells are those with clear brownish-yellow granules in the cytoplasm or nucleus.

Immunofluorescence. The procedure for the first day of the immunofluorescence protocol was the same as that used for immunohistochemistry (described above). The following day, the slices were warmed to room temperature for 40 min. Under dark conditions, goat anti-rabbit IgG H&L (Cy3; S0011, Affinity Biosciences, Ltd.) fluorescent secondary antibody (1:500) was added to the tissues and incubated for 1 h at 37°C. The samples were then soaked and washed in PBS for 15 min. DAPI (Beijing Solarbio Science & Technology Co., Ltd.) was then added in a dropwise manner in the dark and incubated at room temperature for 5 min at room temperature. The samples were then soaked in PBS solution for 4 min, followed by anti-fluorescence attenuation sealer (containing DAPI) drops on the tissue for mounting. The samples were examined using a Leica microscope (DM4B; Leica Microsystems GmbH). For immunofluorescence analysis, in a place with high light intensity, under a 40X lens, the optical density value was analyzed under the same area and was equal to the ratio of the overall optical density to the area.

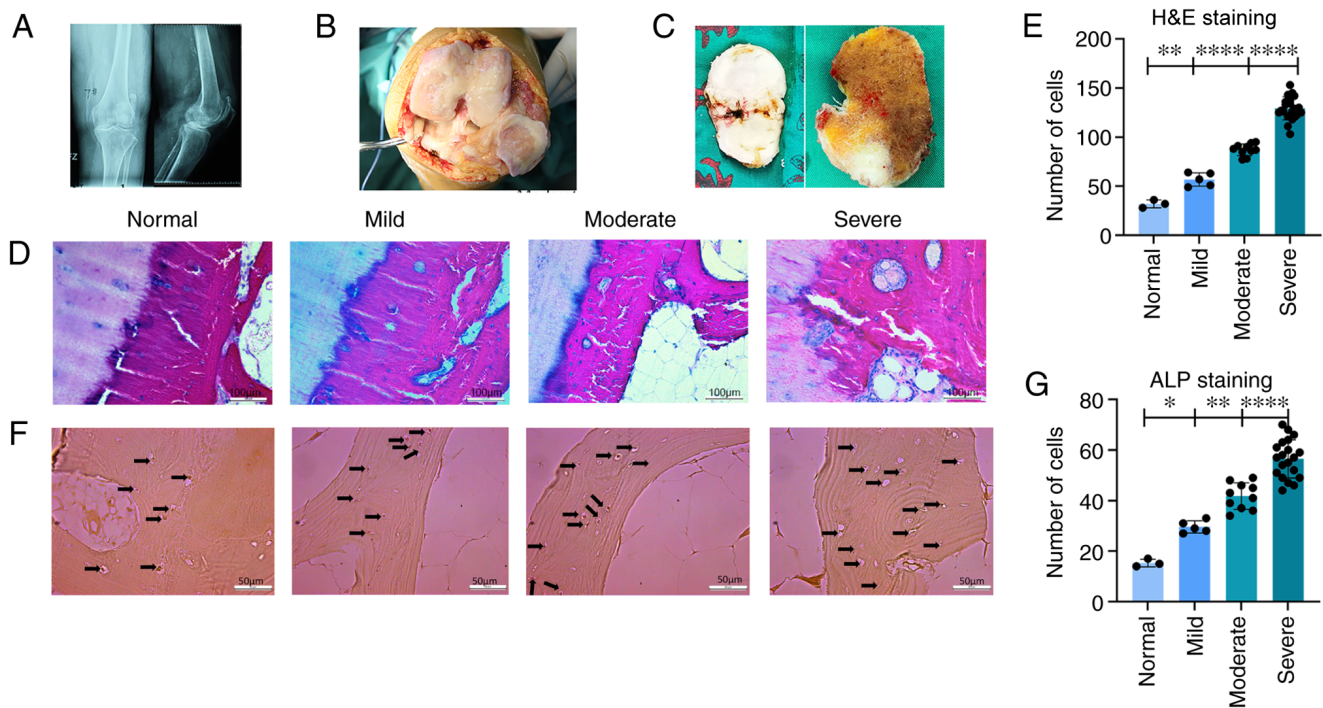


Figure 1. (A) Preoperative radiographs of the patient. (B) Intraoperative gross pathological images. (C) Intraoperative intercepted bone plates of the tibial plateau including cartilage. (D) Subchondral bone H&E staining. (F) Subchondral bone ALP staining of the subchondral bone tissue. The arrows indicate cells positive for ALP staining. (E and G) Results of cell number statistical analysis. In H&E staining and ALP staining, there was a statistically significant difference between the groups ($^*P<0.05$, $^{****}P<0.0001$, and $^{**}P<0.01$). (E and G) The data were analyzed using one-way ANOVA; mean of the 95% confidence interval: Normal (n=3), mild (n=5), moderate (n=10) and severe (n=20). ALP, alkaline phosphatase; H&E, hematoxylin and eosin.

Western blot analysis. All tissues used were removed from -80°C . Subsequently, the samples were washed with cold PBS 2-3 times for blood stain removal, cut into small sections (3x3x5 mm) using a tissue grinder, and placed in a homogenization tube (stored in a liquid nitrogen tank). Subsequently, a total protein extraction kit [containing lysate, phosphatase inhibitor (100X), protease inhibitor (100X), PMSF (100X; Beijing Zhongshan Jinqiao Biological Co.)] was applied for protein extraction. The homogenization tube was placed in the homogenizer, the program was selected to homogenize thoroughly so that the bone texture was 120 sec and was intermittently cooled during the period (liquid nitrogen was automatically added intermittently). The homogenized sample tube was removed, iced for 30 min, and shaken every 5 min to ensure that the tissue was completely lysed. The supernatant, which is the total protein solution, was collected and centrifuged at 4°C at $16,000 \times g$ for 10 min. The undenatured protein solution was determined using the BCA protein concentration determination kit method and the microplate reader [ESCO (Shanghai) Enterprise Development Co., Ltd.] was used to measure the protein concentration (referring to the kit instructions for the specific method), and the protein concentration was then calculated. The protein solution was added to 5X protein loading buffer at a ratio of 4:1 and denatured in a boiling water bath for 15 min. Subsequently, a 10% SDS-PAGE gel was prepared and 15 μl protein per sample was loaded in equal volumes of sample (including markers) into the gel wells. Electrophoresis was performed (80 V for concentrated gel, 120 V for separation gel, 300 mA for 40 min) and gel was transferred to a PVDF membrane (MilliporeSigma). The membrane was washed with TBST

three times for 5 min, blocked with 5% skim milk for 1.5 h at room temperature, and incubated with the primary antibodies [anti-DKK3 antibody (ab186409; rabbit monoclonal, 1:300), anti-GAPDH antibody (ab9485, Abcam, rabbit monoclonal, 1:300) and anti- β -catenin antibody (ab32572, Abcam, rabbit monoclonal, 1:200)] at 4°C overnight. On the second day, the samples were moved to room temperature conditions for 40 min and the membrane was then washed with TBST three times for 5 min, incubated with the secondary antibody [anti-horseradish peroxidase antibody (1:1,000, ab181658, Abcam)] at room temperature for 1.5 h. Afterwards, the membranes were washed with TBST three times for 5 min. Finally, ECL luminescent agent (Beijing Solarbio Science & Technology Co., Ltd.) was used for exposure and observation in a chemical exposure machine. Densitometry was performed using ImageJ (1.8.0) software (National Institutes of Health). Western blot analysis utilized statistical analysis based on the gray value obtained, which is equal to the ratio of the gray value of the target protein to the gray value of the internal reference.

Reverse transcription-quantitative polymerase chain reaction (RT-qPCR). Total RNA was isolated from tissues using a high-speed homogenizer for cancellous bone tissue comminution, using TRIzol[®] reagent (Invitrogen; Thermo Fisher Scientific, Inc.). RNA (1 μg) was then reverse transcribed into cDNA using the PrimeScript RT kit (Affinity Biosciences, Ltd.) according to the manufacturer's protocol. The cDNA samples were then analyzed using LightCycler 480 SYBR-Green I Master (Roche Diagnostics) for RT-qPCR on a LightCycler 480 system (Roche Diagnostics). The reaction

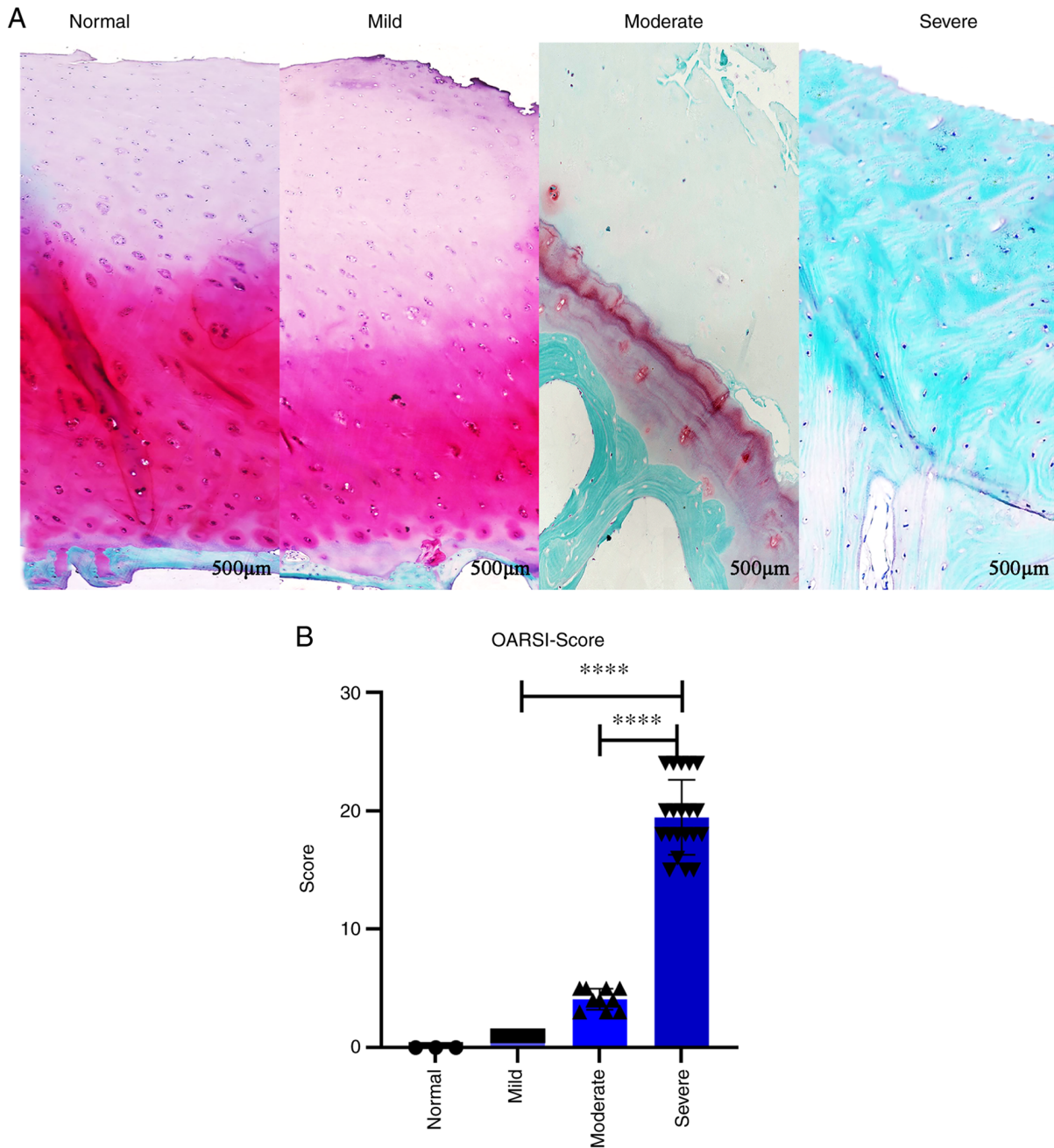


Figure 2. (A) Safranin O staining: Each image demonstrates the OARSI histological score of the medial tibial plateau cartilage. (B) Statistical analysis results. The difference between the mild and moderate groups was statistically significant (**** $P < 0.0001$). (B) Data were analyzed using the Kruskal-Wallis test. OARSI score equals cartilage degenerative tissue grading times staging. Normal (n=3), mild (n=5), moderate (n=10) and severe (n=20). OARSI, Osteoarthritis Research Society International.

conditions were as follows: 95°C for 30 sec, followed by 5 sec at 95°C, 20 sec at 60°C and 20 sec at 75°C 40 cycles, and GAPDH expression was used as an internal control. The data was analyzed for expression changes using the $2^{-\Delta\Delta Cq}$ formula (19). The primer sequences for β -catenin, DKK3 and GAPDH were as follows: β -catenin forward, 5'-CAT CTACACAGTTTGTGCTGCT-3' and reverse, 5'-GCA GTTTTGTCAGTTCAGGGA-3'; DKK3 forward, 5'-CAA TGGGACCATCTGTGACAAC-3' and reverse, 5'-ATCGGT CCAAGGCTCCATCA-3'; GAPDH forward, 5'-TGCCAA ATGATGACATCAAGAA-3' and reverse, 5'-GGAGTGGT GTCGCTGTTG-3'.

Statistical analysis. All data were analyzed using SPSS 22.0 (IBM Corp.) statistical software. Graphs were drawn using GraphPad Prism software (version 8.0). ImageJ (1.8.0) (National Institutes of Health) software was used for immunohistochemistry, immunofluorescence, and western blot analysis. All data were tested for normality of distribution using the Shapiro-Wilk test. One-way analysis of variance with Tukey's post hoc test was used to compare the parametric data between groups. For non-parametric data, the Kruskal-Wallis test with Dunn's post hoc test were used. A value of $P < 0.05$ was considered to indicate a statistically significant difference. Data from all studies were subjected to before statistical analysis.

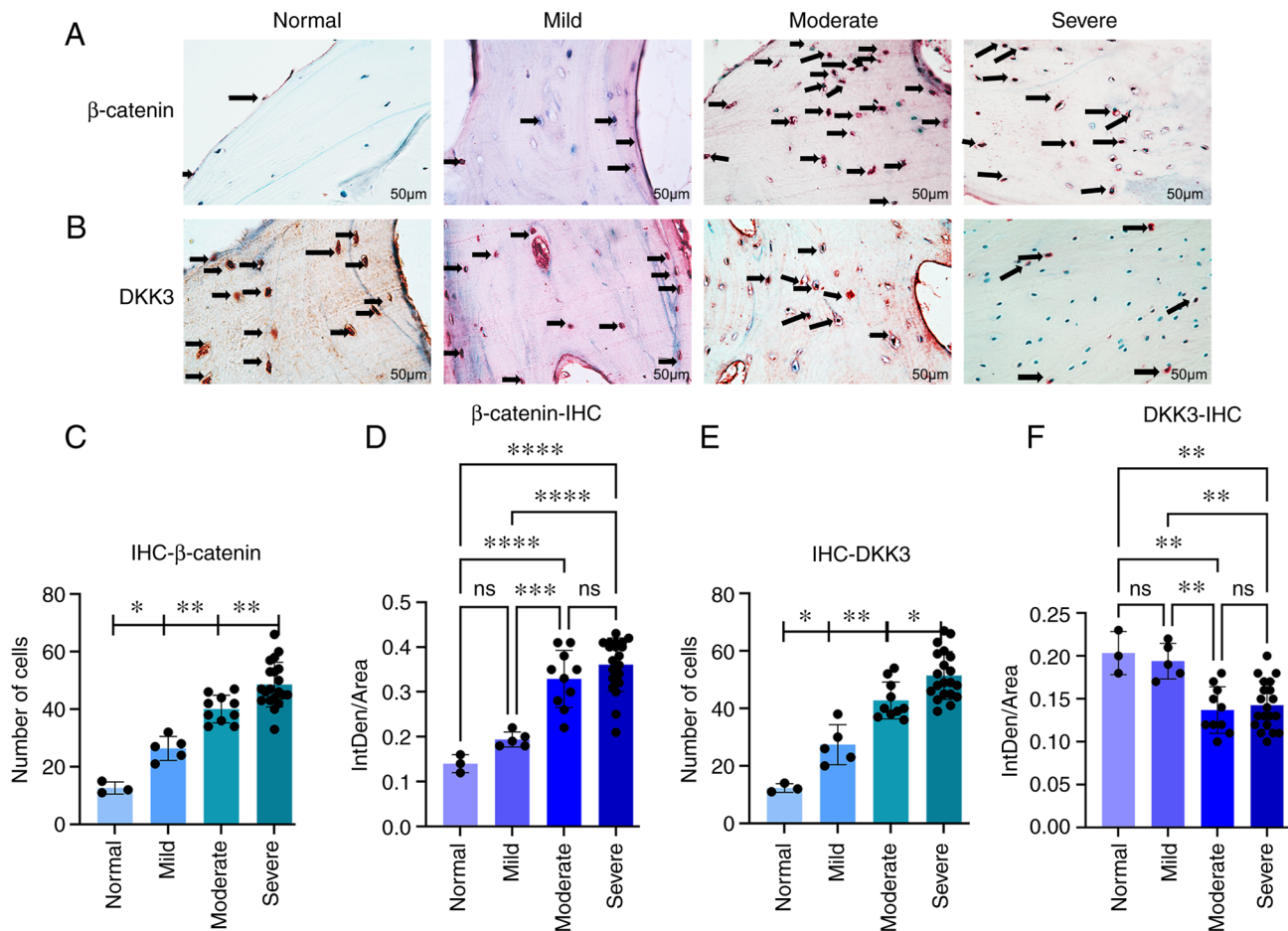


Figure 3. (A and B) Immunohistochemical staining. Arrows indicate the number of positively stained cells. (C) In the β -catenin group, the difference in cell number was statistically significant between the normal and mild, mild and moderate, and moderate and severe symptom groups ($P < 0.01$, $**P < 0.01$ and $**P < 0.01$ respectively). (D) Statistically significant differences between the mild and moderate symptom groups according to the average optical density values ($***P < 0.001$ and $****P < 0.0001$). (E) In the DKK3 group, cell numbers were statistically significant between the normal and mild, mild and moderate, and moderate and severe symptom groups ($P < 0.05$, $**P < 0.01$ and $*P < 0.05$, respectively). (F) Statistically significant differences between the mild and moderate symptom groups according to the average optical density values ($**P < 0.01$). (C-F) Data were analyzed using one-way ANOVA; mean of the 95% confidence interval: Normal (n=3), mild (n=5), moderate (n=10) and severe (n=20). DKK3, Dickkopf-related protein 3; ns, not significant.

Results

Cell count and changes in osteoblast activation. Histological samples of the medial tibial plateau were collected and divided into the normal (n=3), mild (n=5), moderate (n=10) and severe symptom groups (n=20). From the analysis of the pre-operative radiographs from the patients (Fig. 1A), gross pathological images (Fig. 1B) and intraoperative cuttings of the bone plate of the tibial plateau, including cartilage and subchondral bone (Fig. 1C), the medial subchondral bone was found to be markedly sclerotic and cartilage damage was evident. To detect changes in the number of bone cells in the subchondral bone, H&E staining was performed. The results revealed changes in the H&E staining of the subchondral bone tissue (Fig. 1D), and combined with statistical analysis (Fig. 1E) there was a statistically significant difference in cell numbers between the normal, mild and moderate/severe symptom groups. The total number of bone cells (including osteoblasts and osteoclasts) increased with the progression of OA. To observe the changes in the number of osteoblasts in the subchondral bone tissue, ALP staining of the subchondral bone tissue was performed (Fig. 1F). By observing the results of ALP staining

and combining these with statistical analysis (Fig. 1G), an increase in the number of osteoblasts with calcaneal nodules in the subchondral bone was observed, in parallel with the aggravation of the disease. There was a statistically significant difference between the mild and moderate symptom groups and between the moderate and severe symptom groups. The arrows in Fig. 1F indicate the calcareous nodules within the osteoblasts, following ALP staining.

OARSI score. Cartilage damage was investigated by performing Safranin O/fast green staining (Fig. 2A). The results revealed no notable damage in the mild symptom group, mild damage in the moderate symptom group and severe damage in the severe symptom group. In the latter group, some of the subchondral bone was no longer covered by cartilage and the subchondral bone was completely exposed. The cartilage was then scored according to the severity of cartilage damage and in combination with the OARSI scoring system (OARSI score was equal to the cartilage degenerative tissue grading multiplied by the stage). Statistical analysis was then performed, with higher scores indicating more severe cartilage injury. The differences between the mild and moderate symptom groups, and

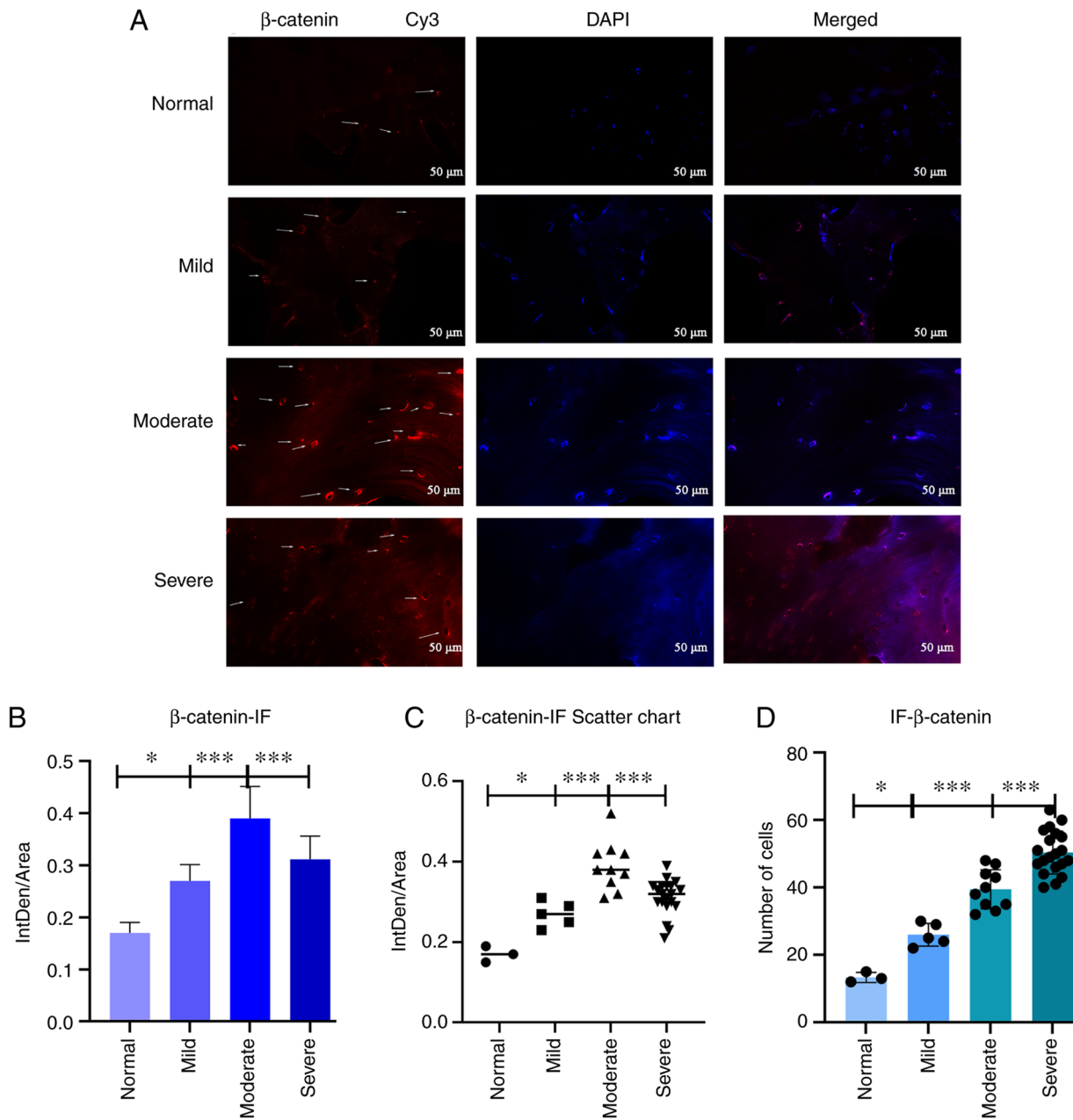


Figure 4. (A) Immunofluorescence staining (β -catenin). Arrows indicate the number of positively stained cells. (B and C) Statistical analysis results. Statistical significant differences in β -catenin expression were revealed between normal and mild, mild and moderate, and moderate and severe OA in the β -catenin group (* P <0.05, *** P <0.001 and *** P <0.001, respectively). (D) Statistical analysis results of cell numbers. The number of cells was statistically significant between the normal and mild, mild and moderate, and moderate and severe symptom groups (* P <0.05, *** P <0.001 and *** P <0.001, respectively). (B-D) Data were analyzed using one-way ANOVA. Mean 95% confidence interval for average score: Normal (n=3), mild (n=5), moderate (n=10) and severe (n=20). OA, osteoarthritis.

the moderate and severe symptom groups were statistically significant (both P <0.0001; Fig. 2B).

β -catenin and DKK3 expression evaluation using immunohistochemistry. To observe the expression patterns of β -catenin and DKK3 in the osteoblasts in the subchondral bone, immunohistochemistry was performed, and the results were statistically analyzed (Fig. 3). For β -catenin (Fig. 3A), the differences in cell counts between the normal and mild groups, mild and moderate groups, and moderate and severe groups were statistically significant (P <0.05, P <0.01 and P <0.01, respectively) (Fig. 3C). For DKK3 (Fig. 3B), there were statistically significant differences in cell numbers between

the normal and mild, mild and moderate, and moderate and severe symptom groups (P <0.05, P <0.01 and P <0.05, respectively) (Fig. 3E). In addition, the differences between the mild and moderate symptom groups were statistically significant according to the analysis of optical density values (P <0.001 and P <0.01, respectively; Fig. 3D and F).

Evaluation of β -catenin expression and DKK3 by immunofluorescence. To further observe β -catenin and DKK3 expression levels in osteoblasts from the subchondral bone, immunofluorescence staining was used. The results revealed the optical density changes in β -catenin and DKK3 expression in the subchondral osteoblasts. Immunofluorescence staining

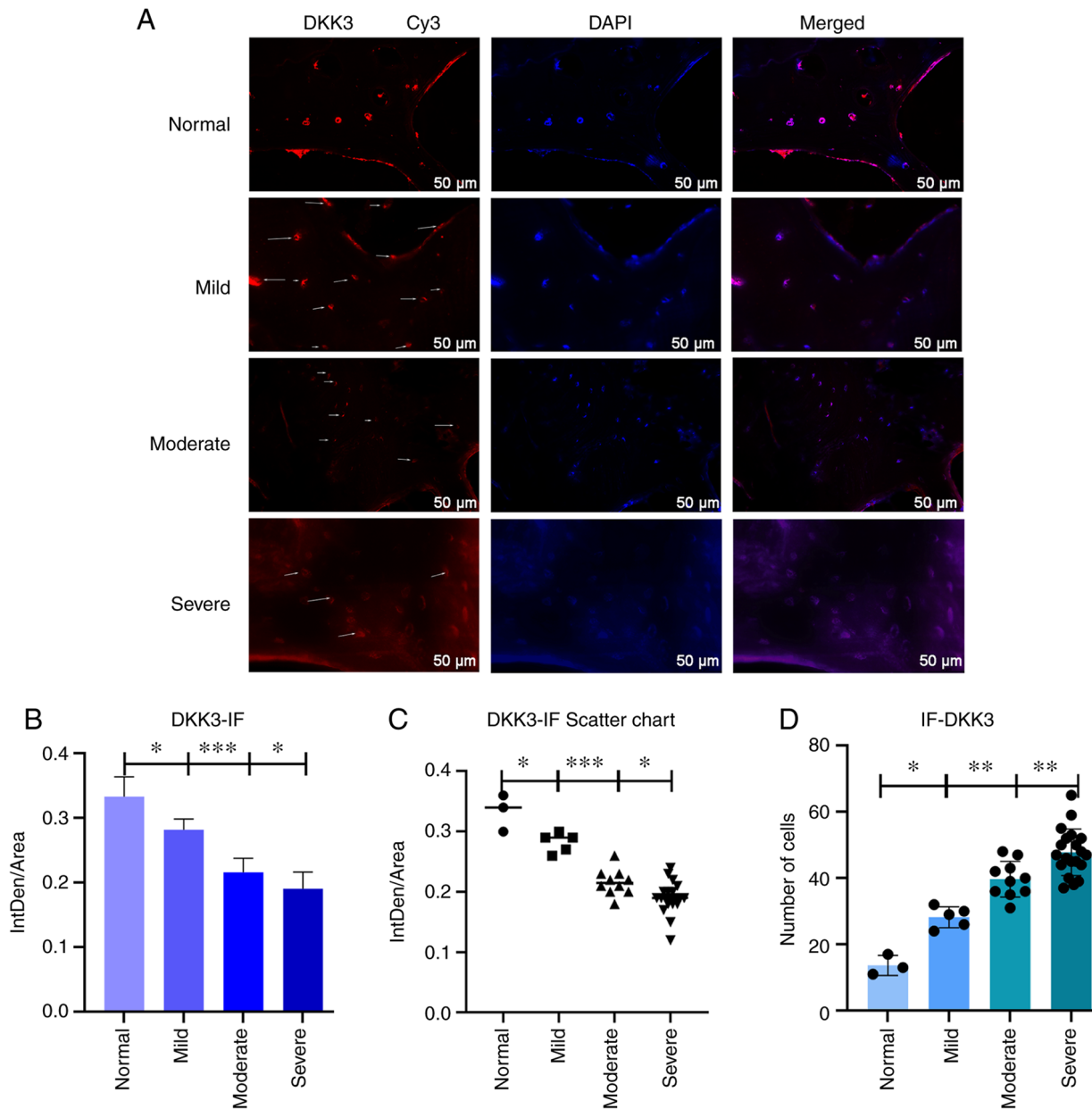


Figure 5. (A) Immunofluorescence staining for DKK3. Arrows indicate the number of positively stained cells. (B and C) Statistical analysis results. The difference in DKK3 expression between the normal and mild, mild and moderate, and moderate and severe groups was statistically significant ($P < 0.05$, $***P < 0.001$ and $*P < 0.05$, respectively). (D) Cell count statistical analysis results. The differences in cell numbers was statistically significant between the normal and mild, mild and moderate, and moderate and severe symptom groups ($P < 0.05$, $***P < 0.01$, and $**P < 0.01$, respectively). (B-D) Data were analyzed using one-way ANOVA; mean 95% confidence interval for average score: Normal (n=3), mild (n=5), moderate (n=10) and severe (n=20). DKK3, Dickkopf-related protein 3.

was performed using Cy3, which is excited at 532 nm and emits orange light, then images of the samples were captured using a 40X fluorescence microscope. The results of β -catenin (Fig. 4A) and DKK3 expression (Fig. 5A) were observed and statistically analyzed (Figs. 4B-D, and 5B and C). The cell counts were statistically significantly different between the normal and mild, mild and moderate, and moderate and severe symptom groups. The differences in β -catenin and DKK3 expression were statistically significant between the mild and moderate symptom groups by analyzing the mean optical density ($P < 0.001$ and $P < 0.001$, respectively).

Changes in β -catenin and DKK3 protein and mRNA expression. To detect and quantify β -catenin and DKK3 protein

expression in the subchondral bone tissue, western blot analysis was performed. By analyzing the results (Fig. 6A-C) using statistical analysis, significant differences in the protein expression level of β -catenin were observed between the mild and moderate symptom groups, and between the moderate and severe symptom groups ($P < 0.001$ and $P < 0.01$, respectively). The β -catenin expression levels were more increased in the moderate symptom group. There were also statistically significant differences in the protein expression level of DKK3 between the mild and moderate symptom groups, and between the moderate and severe symptom groups ($P < 0.01$ and $P < 0.0001$, respectively). Increased DKK3 expression levels were observed in the normal and mild symptom groups, whereas decreased expression levels were observed in the

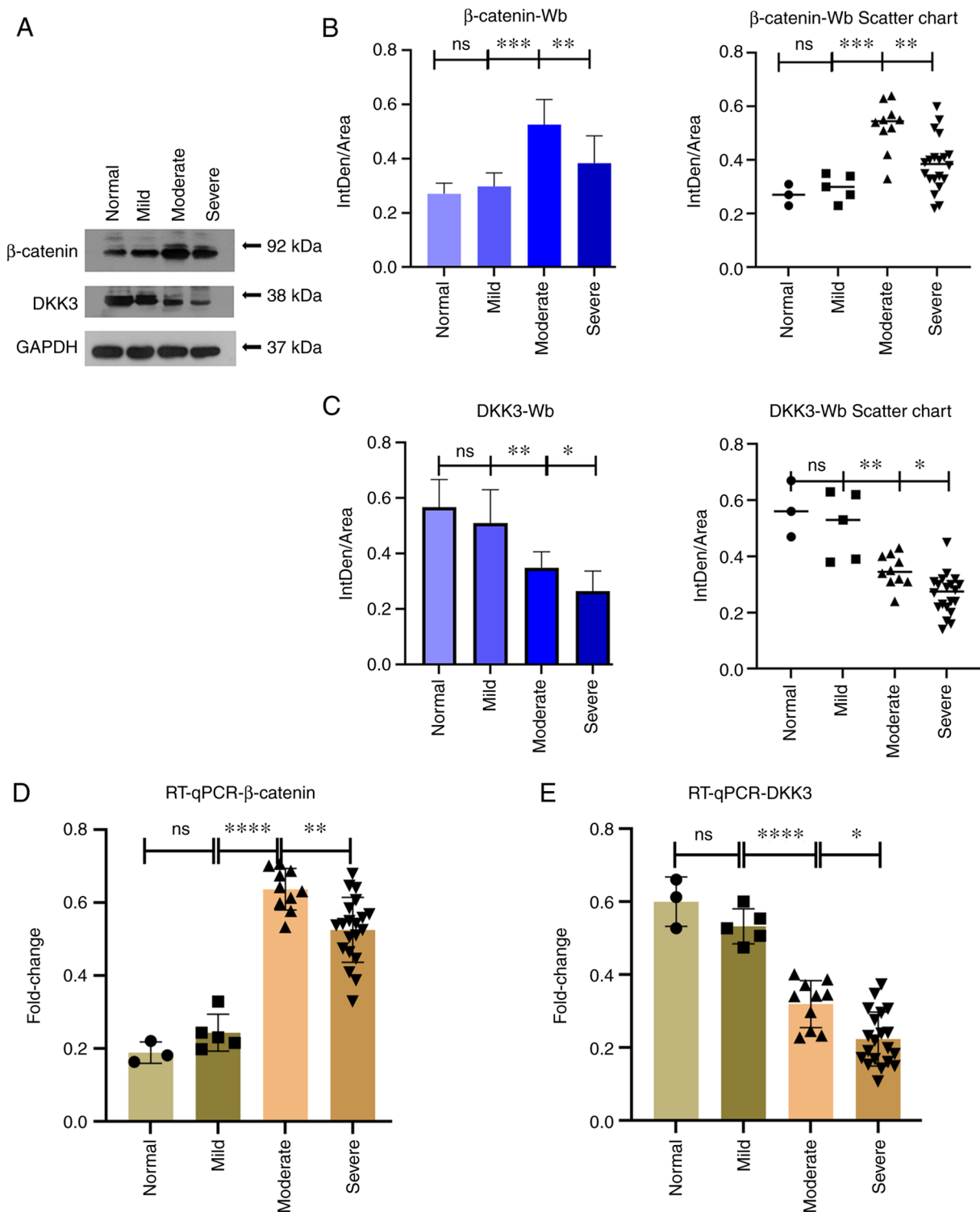


Figure 6. (A-C) The results of western blot analysis revealed that in the β -catenin group, there was a statistically significant difference between the mild and moderate symptom groups and between the moderate and severe symptom groups ($***P < 0.001$ and $**P < 0.01$, respectively). In the DKK3 group, there were statistically significant differences between the mild and moderate symptom groups, as well as between the moderate and severe symptom groups ($**P < 0.01$ and $*P < 0.05$, respectively). (D and E) The results of RT-qPCR revealed significant differences between the mild and moderate groups as well as between the moderate and severe symptom groups in the β -catenin group ($****P < 0.0001$, and $**P < 0.01$, respectively). In the DKK3 group, there was a statistically significant difference between the mild and moderate symptom groups and between the moderate and severe symptom groups ($****P < 0.0001$ and $*P < 0.01$, respectively). (B-E) Data were analyzed using one-way ANOVA. 95% confidence interval for average score: Normal (n=3), mild (n=5), moderate (n=10) and severe (n=20). DKK3, Dickkopf-related protein 3; RT-qPCR, reverse transcription quantitative PCR; ns, not significant.

moderate and severe symptom groups, exhibiting a trend towards a decrease in expression levels with the severity of OA. Finally, in order to detect the gene expression level of

β -catenin and DKK3 in the bone tissue from subchondral bone, RT-qPCR was performed. The results (Fig. 6D and E) demonstrated significant differences between the mild and

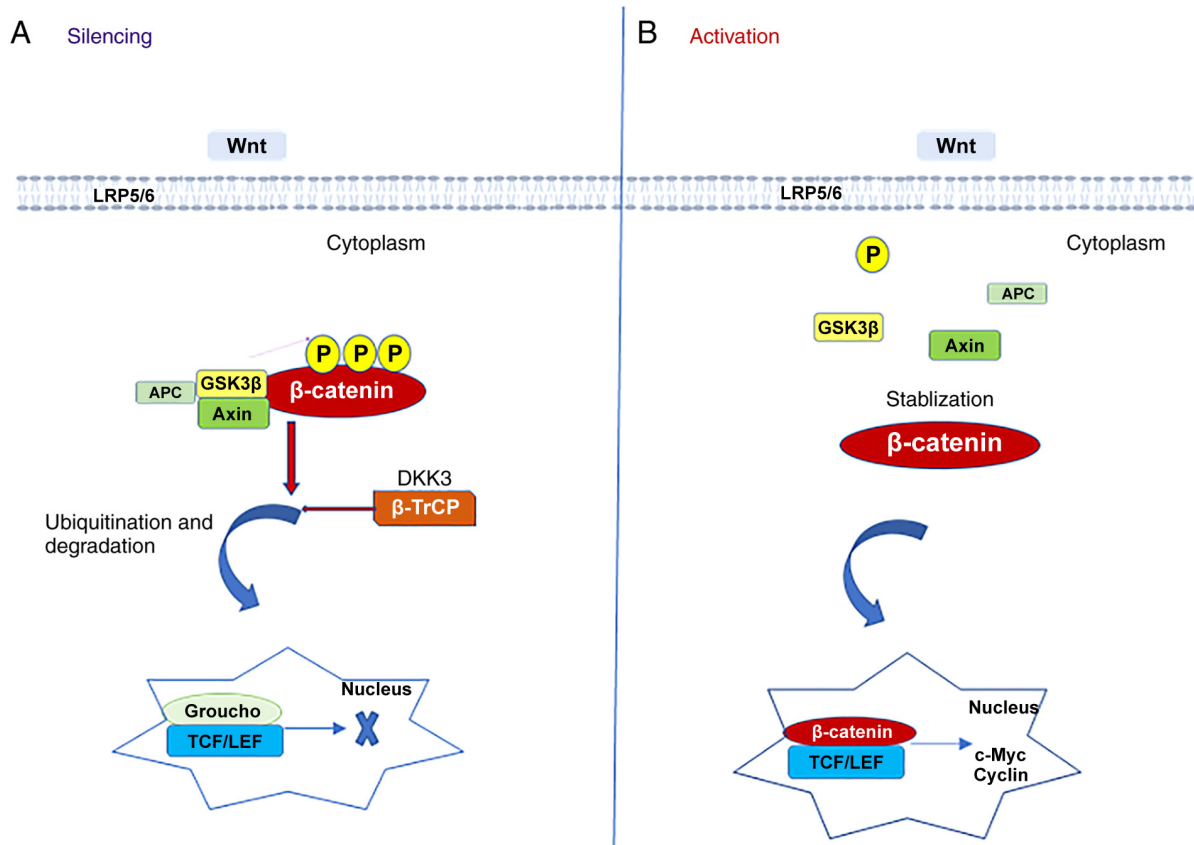


Figure 7. Overview of the Wnt signaling pathway. (A) Through the silencing the Wnt pathway, DKK3 competes with β -catenin in the cytoplasm and promotes its degradation, inhibiting the Wnt pathway and blocking nuclear transcription. (B) Activation of the Wnt pathway β -catenin aggregates in the cytoplasm. Stable β -catenin is then transferred to the nucleus and stimulates the transcription of Wnt target genes. DKK3, Dickkopf-related protein 3.

moderate symptom groups, and between the moderate and severe symptom groups concerning the β -catenin expression evaluation group ($P < 0.0001$ and $P < 0.01$, respectively). Concerning DKK3 mRNA expression, there was a statistically significant difference between the mild and moderate symptom groups and between the moderate and severe symptom groups ($P < 0.0001$ and $P < 0.01$ respectively), demonstrating an increased expression in the normal and mild symptom groups, and a decreased expression in the moderate and severe symptom groups, exhibiting a trending towards a decreased expression with the increasing severity of OA.

Discussion

Previous research has characterized early-stage OA with increased subchondral bone remodeling and late OA with decreased bone resorption and increased bone formation (5). Abnormal subchondral bone remodeling and osteophyte formation are hallmarks of OA progression (20).

Human β -catenin protein is an 88-kDa cytoskeletal protein, 781-amino acid long, encoded by the catenin beta-1 gene (CTNNB1) and located on the 3p21 human chromosome (21). Numerous studies have shown that β -catenin is a major signaling molecule in the Wnt signaling pathway (22–24). Wnt signaling pathway activation depends on β -catenin expression levels in the cell (Fig. 7). When β -catenin levels are decreased, the Wnt pathway is not activated; however, when the levels are increased, the Wnt pathway is activated (25).

DKK3 is a member of the DKK family, among which DKK-1, -2 and -4 compete for binding to the Wnt protein low-density lipoprotein 5/6 receptor (26). DKK3 negatively regulates β -catenin and its downstream signaling pathways by reducing β -catenin aggregation in the cytoplasm (27). The graphs presented in Fig. 1 depict the changing patterns of β -catenin and DKK3 expression when the Wnt/ β -catenin pathway is inhibited and activated. In addition, DKK3 indirectly inhibited its role in the Wnt/ β -catenin pathway by reducing the nuclear transcription of β -catenin. Suwa *et al* (28) reported that various genes in the Wnt signaling pathway were expressed in the adrenal cortex. Based on the regional distribution pattern of DKK3 expression, it may be associated with the Wnt/ β -catenin signaling pathway, affecting regional differentiation or growth. However, it is unclear whether DKKs are associated with bone formation. Subsequently, Niehrs (29) reported that, in adults, DKKs were associated with bone formation in bone disease. Furthermore, Aslan *et al* (30) further demonstrated that DKK3 expression affected the osteogenesis process *in vivo* and *in vitro*, suggesting that it may exert an inhibitory effect on osteogenesis.

As the number of studies on the involvement of the Wnt signaling pathway in the developmental mechanisms of OA have increased, it has been reported that the activation of the classical Wnt/ β -catenin pathway accelerates the progression of OA (31). According to a previously published study by Funck-Brentano *et al* (32), it was revealed by using a mouse model of OA that the classic Wnt pathway was activated

to promote subchondral bone formation osteophytes and inhibition of the Wnt/ β -catenin pathway signaling improved subchondral bone structure and biomechanical function, thereby alleviating the degeneration of articular cartilage. This may be attributed to active osteoblasts in OA altering bone microarchitecture through aberrant bone remodeling and reduced mineralization. However, this alteration notably reduced the biomechanical function of the subchondral bone, depriving it of its cushioning effect against mechanical loads and its protective function against cartilage, further aggravating cartilage damage and accelerating the development of OA.

In the present study, a total of 38 subchondral bone tissue specimens were collected from the tibial plateau in patients with severe OA. In addition, intraoperative anatomical tissues and postoperative tibial plateau osteotomy specimens were also collected, demonstrating severe cartilage damage in the femoral condyle and tibial plateau, with some areas of cartilage lost and replaced by sclerotic subchondral bone. H&E and ALP staining of the subchondral bone tissue revealed that the total number of osteocytes and osteoblasts in the subchondral bone increased along with the degree of OA, including various osteoblasts (pre-osteoblasts, osteoclasts and bone lining cells). Neve *et al* (4) reported that the abnormal differentiation and increased activity of osteoblasts were observed in OA. In addition, dysfunctional osteoblasts produce in turn numerous transcription factors, growth factors and other proteins involved in the pathogenesis of OA (33), further promoting the development of OA.

It has been found that osteoblast differentiation requires the classical Wnt/ β -catenin signaling pathway (14). It has been reported that some osteoblasts contain ALP, which can be used as a biochemical marker to assess bone formation (34); however, mechanical loading increases ALP activity (35). Thus, in the present study, it was demonstrated using ALP staining (AZO method) that the number of ALP-positive cells in the subchondral bone specimens, to be increased during the progression of OA from mild to moderate; in addition, in severe OA, there was no significant increase in active osteoblasts compared with that in moderate OA.

Cartilage changes may be visualized using Safranin O/fast green staining, revealing that the hyaline cartilage cracks, thins and eventually disappears, as the degree of OA increases. It has been reported that articular cartilage is very sensitive to mechanical loading, subchondral bone provides cartilage support and cartilage degeneration is a sign of OA aggravation (36). The role of the subchondral bone in the development of OA remains under investigation; however, there is evidence to indicate that changes in the subchondral bone are accompanied by cartilage degeneration (37).

Another study revealed that metabolic modulators produced by subchondral osteoblasts directly cause chondrocyte degeneration by affecting the microstructure between the bone and cartilage, while abnormal expression of osteoblasts can indirectly expose cartilage to higher stress (38). Thus, this direct and indirect mode of action of subchondral bone and cartilage accelerates cartilage degeneration and further aggravates the progression of OA. Fell *et al* (39) reported that the association between the morphology of the subchondral bone tissue and the viscoelasticity, and thickness of cartilage may contribute to the development and progression of

OA by altering the ability of cartilage to dissipate energy. Furthermore, Smieszek *et al* (40) demonstrated that osteoblasts are inhibited by high expression of microRNA-21 in OA, through the reduction of subchondral bone mineralization. Therefore, altering and alleviating the abnormal remodeling of subchondral bone may mitigate further cartilage damage and delay the progression of OA.

Due to this supportive role of subchondral bone on cartilage, the question on whether DKK3 could reduce subchondral bone sclerosis via β -catenin inhibition of the Wnt pathway, preventing further damage to cartilage due to subchondral bone sclerosis and delaying the progression of OA was raised. Among the numerous regulators of the classical Wnt pathway, β -catenin is a key factor, while DKK3 is a downstream inhibitor of the Wnt pathway. In malignant tumor cells, DKK3 was associated with the induction of apoptosis and inhibition of invasion by regulating β -catenin signaling, and c-Jun N-terminal kinase-dependent cellular pathways, as reported by Lee *et al* (17). It has been reported that β -catenin may be a central molecule mediating osteoblast viability and differentiation (41). In addition, DKK3 inhibited the Wnt signaling pathway by hydrolyzing β -catenin/APC/GSK3 β , thereby inhibiting the entry of β -catenin into the nucleus for transcription (17). Therefore, in OA, DKK3 degrades β -catenin via ubiquitination, thereby degrading the downstream signaling pathway factors of β -catenin and inhibiting nuclear transcription of the Wnt pathway (15), delaying abnormal remodeling of subchondral bone.

According to Huang *et al* (42) the regulation of the Wnt signaling pathway reduced the risk of OA. Charlier *et al* (43) reported that the DKK3 regulation of the Wnt/ β -catenin pathway increased the dedifferentiation of OA chondrocytes in the human hip and promoted the remission of OA. Thus, maintaining normal Wnt/ β -catenin signaling is critical for orthogenic bone tissue differentiation and in OA, once this pathway is activated, it can accelerate OA progression.

In conclusion, the results of the present study demonstrated that with the increase in the pathological changes of OA in the knee, the number of osteoblasts may also gradually increase; however, the number of active osteoblasts was highest in moderate OA and slightly more increased in severe OA. The aforementioned association analysis revealed that DKK3 expression was high in the normal and mild groups, decreased in the moderate and severe groups, while β -catenin expression increased significantly in the moderate OA group after activation of the Wnt/ β -catenin pathway. In subchondral bone tissue, DKK3 may play an opposite role to β -catenin by inhibiting the Wnt/ β -catenin signaling pathway in osteoblasts, reducing subchondral bone sclerosis, while reducing cartilage damage and delaying OA progression. In future studies, the authors aim to verify whether DKK3 can inhibit bone formation *in vitro*. Furthermore, it was unexpectedly found that, in the late stages of OA, some osteoblasts can still be activated and play a role in abnormal bone remodeling, leading to increased subchondral bone sclerosis and worsening of pain.

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Availability of data and materials

The datasets used and/or analyzed during the current study are available from the corresponding author on reasonable request.

Authors' contributions

XL and QJ contributed to the conception and design of the present study. XL, WJ and XY performed the experiments. XL performed the statistical analysis and contributed to the manuscript preparation. QJ and XL confirm the authenticity of all the raw data. QJ contributed to funding acquisition. All authors have read and approved the final version of the manuscript.

Ethics approval and consent to participate

The present study was approved by the Ethics Committee of the General Hospital of Ningxia Medical University (approval no. KYLL-2020-20; approval date, January 14, 2021), and informed consent was obtained from all the enrolled patients.

Patient consent for publication

The patient whose images are shown in Fig. 1 consents to the publication of these images.

Competing interests

The authors declare that they have no competing interests.

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