

Combined Identification of Novel Markers for Diagnosis and Prognostic of Classic Hodgkin Lymphoma

Zhixing Kuang^{1,*}
Jiannan Tu^{2,*}
Xun Li³

¹Department of Radiation Oncology, Nanping First Hospital Affiliated to Fujian Medical University, Nanping, People's Republic of China; ²Department of Oncology, Nanping First Hospital Affiliated to Fujian Medical University, Nanping, People's Republic of China; ³Department of Oncology, Changzhou Tumor Hospital Affiliated to Soochow University, Changzhou, People's Republic of China

*These authors contributed equally to this work

Background: An effective diagnostic and prognostic marker based on the gene expression profile of classic Hodgkin lymphoma (cHL) has not yet been developed. The aim of the present study was to investigate potential markers for the diagnosis and prediction of cHL prognosis.

Methods: The gene expression profiles with all available clinical features were downloaded from the Gene Expression Omnibus (GEO) database. Then, multiple machine learning algorithms were applied to develop and validate a diagnostic signature by comparing cHL with normal control. In addition, we identified prognostic genes and built a prognostic model with them to predict the prognosis for 130 patients with cHL which were treated with first-line treatment (ABVD chemotherapy or an ABVD-like regimen).

Results: A diagnostic prediction signature was constructed and showed high specificity and sensitivity (training cohort: AUC=0.981,95% CI 0.933–0.998, P<0.001, validation cohort: AUC=0.955,95% CI 0.895–0.986, P<0.001). Additionally, nine prognostic genes (LAMP1, STAT1, MMP9, C1QB, ICAM1, CD274, CCL19, HCK and LILRB2) were screened and a prognostic prediction model was constructed with them, which had been confirmed effectively predicting prognosis (P<0.001). Furthermore, the results of the immune infiltration assessment indicated that the high scale of the fraction of CD8 + T cells, M1 macrophages, resting mast cells associated with an adverse outcome in cHL, and naive B cells related to prolonged survival. In addition, a nomogram that combined the prognostic prediction model and clinical characteristics is also suggested to have a good predictive value for the prognosis of patients.

Conclusion: The new markers found in this study may be helpful for the diagnosis and prediction of the prognosis of cHL.

Keywords: novel markers, diagnosis, prognosis, classic Hodgkin lymphoma

Introduction

Classic Hodgkin lymphoma (cHL) is a kind of B-cell neoplasm which characterized by the paucity of Hodgkin/Reed-Sternberg (HRS) cells in conjunction with a microenvironment which contains abundant infiltrating reactive cells. cHL is one type of HL, which constitutes more than 90% of HL.¹ The incidence of HL is 2.8 per 100,000 people, it is estimated that 8,830 people in the United States will be diagnosed with HL and 960 people will die from this disease in 2021.² Currently, the main treatments of cHL include multi-agent chemotherapy and involved field radiation therapy, which can significantly reduce the rate of death and the long-term remission of classic Hodgkin lymphoma can achieve 85%–95%, however,

Correspondence: Xun Li
Department of Oncology, Changzhou Tumor Hospital Affiliated to Soochow University, Changzhou, 213002, People's Republic of China
Email smxlixun@sina.com



approximately 20%–30% of patients will be seen with disease progression or death within 5 years.³ Until now, many efforts have been made in the area of new drug development to improve treatment efficacy and reduce toxicity; the best known drugs for us are anti-CD30 inhibitors and PD-1 antibodies, which have been reported to show promising results for patients with relapsed and refractory cHL.⁴ Anti-CD30 inhibitors, called rentuximab vedotin, have been approved for the treatment of classic Hodgkin lymphoma patients after failure with autologous stem cell transplantation or multi-agent chemotherapy regimens,⁵ but only approximately 17% to 29% of those patients who experienced failure or relapsed with both brentuximab vedotin (CD30-targeted therapy) and autologous stem cell transplantation (ASCT) can expect complete remission (CR) after accepting the treatment of PD-1 antibodies.⁶ To meet the patient's needs, our ability to develop newly approaches for patients with cHL should keep pace with the development of those suffering from cHL. Expanding knowledge of the changes in cHL gene expression levels may be helpful for the discovery of potential therapeutic targets.

On the other hand, in light of the tremendous developments in the treatment of cHL, an appropriate attention should be paid to novel markers for diagnostic and prognostic factors.

Novel diagnostic markers will be helpful for accurate diagnosis and the development of new therapeutic targets. Excessive medication or radiation therapy may compromise survival due to adverse effects, so a suitable and accurate risk stratification is essential for the treatment of all patients. The most widely used clinical indicator to evaluate the risk of cHL is the international prognostic score (IPS) and is used primarily to evaluate advanced cHL.⁷ In recent years, the value of interim fluorodeoxyglucose positron emission tomography (PET)/computer tomography (CT) was gradually established to differentiate the risk of cHL.⁸ However, given the heterogeneity among individual patients with cHL, all these tools that we mentioned use for risk stratification provide limited information underlying the HL biology.⁹ In light of the pathogenesis of cHL is multifactorial and complicated, therefore, a better understanding of the molecular mechanism in the pathogenesis of cHL is essential to us to find a new prognostic factor.

In recent years, the development of medical technologies that allow gene expression profiling from archival paraffin-

embedded tissues pushed forward the recognition of pathogenetic and contributed to the creation of new markers for predicting diagnosis and prognosis based on gene expression. Here, multiple statistical methods were utilized to identify novel markers for diagnostic and prognostic prediction (Figure 1). The discovery of the present research may be helpful in establishing new diagnostic and prognostic markers and to put forward new therapeutic targets.

Materials and Methods

Data Sources and Data Processing

The raw “CEL” data of GSE12453 (12 samples of cHL and 25 samples of normal B cells), GSE39133 (29 samples of cHL and 5 cases of normal B cells), GSE25986 (5 cHL and 5 normal B cell) and GSE17920 (130 case of cHL), which all based on the platform of GPL570 [HG-U133_Plus_2] and were downloaded from the Gene Expression Omnibus (<http://www.ncbi.nlm.nih.gov/geo/>) database. All raw chip data undergoes quality assessment and quality control to identify unqualified samples, and then background correction and normalization were performed. The process which mentioned above were completed by “simpleaffy”,¹⁰ “affyPLM”, “arrayQualityMetrics”¹¹ packages. After accomplishment of preprocessing and analysis, one disqualified sample in GSE12453 was excluded from our following analysis.

Identification of a mRNA Signature Discriminating Between cHL and Normal Control

To enhance the reliability of the research, four data sets were merged to a big one with 210 case samples and the package of “sva” was used to get rid of batch effects. Subsequently, all 210 cases of cHL patients were randomly divided into a training group and a testing group with a ratio of 1:1. The package of “limma” was utilized to identify differentially expressed genes (DEGs) in the training cohort, while $P < 0.05$ and $|\log_2 FC| > 1$ were set as cutoff values. Then, the random forest (RF) algorithm and least absolute shrinkage and selection operator (LASSO) analysis were used to reduce the dimensionality of DEGs. The optimal values of the penalty parameter λ were determined by the results of 10 times cross-validations. Random forest is a collection of decision trees; these decision trees are generated by recursive binary division of different random sub-samples of training data. In the random forest analysis, the optimal genes were selected according to the mean decrease in accuracy to select optimal genes. Finally, the overlapping markers from the two algorithms were

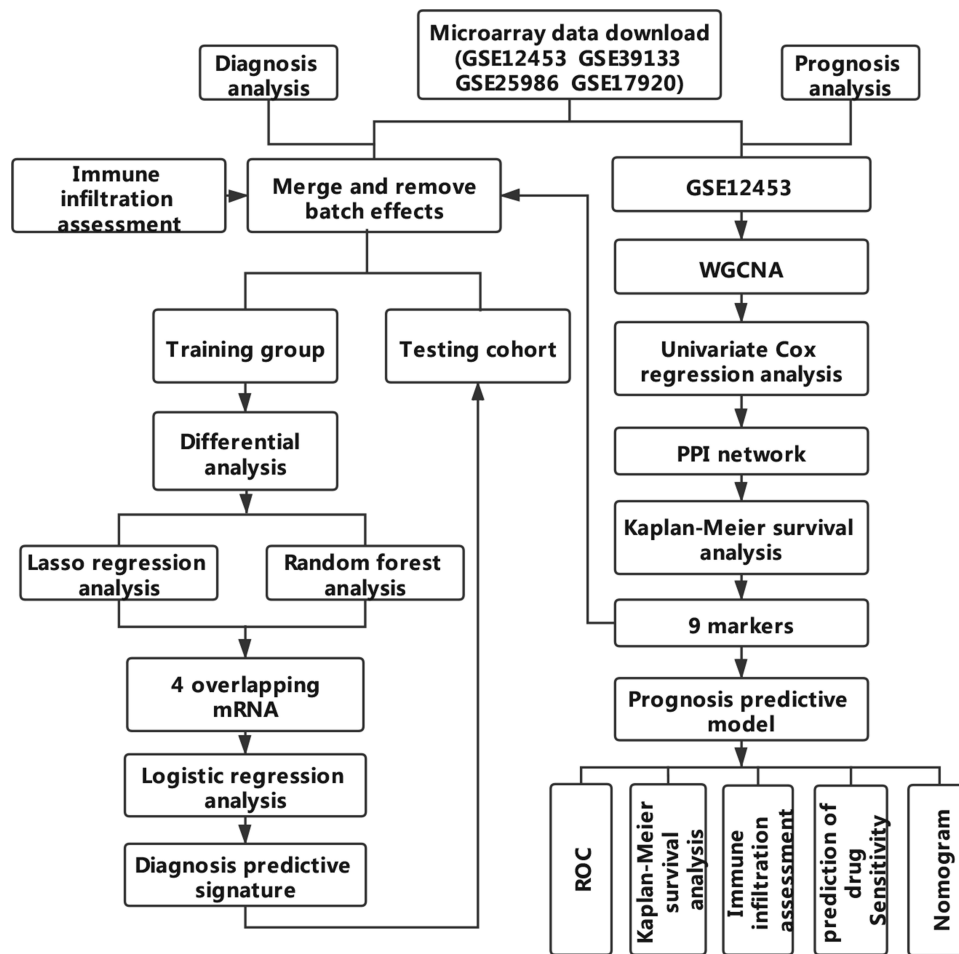


Figure 1 Workflow chart of data generation and analysis.

obtained and were used to build a diagnostic signature and get the diagnostic score by using the mRNA coefficients that obtained in logistic regression. Diagnostic score was constructed as follows diagnostic score = $\sum_{i=1}^n \exp_i * \beta_i + \text{constant value}$. Where β represents the regression coefficient of mRNA which derived from the analysis of logistic regression, and i represents the expression level of mRNA. LASSO analysis was performed by the “glmnet” package, random forest analysis was performed by “randomForest” package.

Co-Expression Network Construction and Identity Candidate Genes

The top 6000 variant of expression profiles of 24 samples of normal B cells and 12 samples of cHL in GSE12453 were selected and utilized to construct a co-expression network by using “WGCNA” package. The best soft thresholding parameter was selected to make our gene distribution conform to

the scale-free network. To counteract the effects of missing or spurious connections between the nodes in the network, the topological overlap matrix (TOM) was transformed by adjacent matrices. According to the measure of TOM-based dissimilarity measure, the genes that highly correlated were clustered into the same module. The Gene Significance (GS) values represented the relationship of each gene with cHL, and the module membership (MM) was represented the correlation of genes with module eigengenes (MEs). Finally, only the module most relevant to cHL was used for further analysis. In our study, to screen the hub genes in the module and ensure the reliability of the results, the criterion of genes of GS > 0.2 and MM > 0.8 were seated.

Construction of a PPI Network and Detection of Prognosis-Related Genes

To reduce the number of genes, the univariate Cox proportional hazards regression analysis was performed by

assessing the relationship between overall survival of patients with cHL (OS) and the expression of genes retained with WGCNA, only genes with $p < 0.05$ in the analysis results were sorted out and used to build a protein-protein interactions (PPI) network. The Search Tool for the Retrieval of Interacting Genes (STRING 10.0) is a database that was designed to predict the protein-protein interactions of genes, and only combination scores > 0.4 were chosen to be significant. Only candidate genes with degree > 10 will be considered hub genes. To investigate real hub mRNAs, Kaplan-Meier survival analysis was performed using the median expression value as the cut-off point in GSE17920. In addition, the Wilcoxon Signed-Rank test was used to calculate the expression difference between cHL and normal B cells for real hub genes in the combinational dataset with 210 case samples.

Multivariate Cox Regression Conduction and Identification of a Risk Model

Multivariate Cox regression analysis was performed to construct a prognosis model using the nine genes related to the prognosis of cHL. The risk score of the prognostic risk model for each patient with cHL was calculated as follows: risk score = $\beta_1 e_1 + \beta_2 e_2 + \beta_3 e_3 + \dots + \beta_n e_n$, e represents the expression level of gene n , where β represents the expression level derived from the multivariate Cox regression analysis. All patients with cHL were divided into high- and low-risk groups according to the median value of the risk score as the cutoff value. Survival curves were calculated and visualized by the Kaplan-Meier analysis, and the package of “survivalROC” was utilized to illustrate the accuracy of the survival prediction based on the risk score. Apart from this, we also studied the correlation between risk score and clinical characteristics. Furthermore, the IC50 of six commonly used cytotoxicity drugs was predicted for all high and low-risk groups using the “pRRophetic” package. Finally, the disagreement of immune landscape between the patients with different risk was analysed by using the tool of CIBERSORT.

Integrated Analysis by Combining the Clinical Characteristic and the Risk Model

The analyses of univariate and multivariate Cox regression were performed to investigate the effect of the risk model on the prognosis of cHL patients. The risk stratification which obtained from the risk model and other

clinicopathological characteristics including age, gender, albumin, hemoglobin, stage, white cell count and lymphocyte rate were used as covariates. In addition to this, the risk model with clinical characteristics was used to build a nomogram. Calibration curves were used to estimate the conformity between the real outcomes and the predicted outcomes for the nomogram, and the discrimination for the nomogram was evaluated by C-index. The package of ‘rms’ was applied to construct the nomogram and calibration curves. Statistical analyzes in our study were performed using R software.

Results

Pre-Processing of the Data Sets and Identification of DEGs in cHL

In total, 34 samples of normal B cells and 176 cases of classical Hodgkin lymphoma(cHL) in four cohorts were chosen for subsequent analysis after excluding a disqualified sample. After completion from the preprocessing, all data of microarray were converted into an expression matrix. To reduce the differences between batches, we regroup the data from four batches after the effects of the removed batch, so that the downstream analysis can only consider biological differences ([Figure S1](#)). After that, all 210 case samples were randomly divided into training data set and validation data set. The training group was utilized to identify DEGs, and a total of 61 upregulated and 114 downregulated genes were filtered out ([Figure S2](#)).

Building and Validation of an mRNA Signature for Diagnosis and Prediction of cHL

We obtained the top 11 markers with the maximum value of the mean decrease in accuracy by using the analysis of RandomForest, and 27 markers were obtained by using analysis of LASSO ([Figure S3](#)). There were eight overlapping markers between these two methods. We obtained four markers by using the method of logistic regression variable selection and constructed a diagnostic prediction signature with the four markers ([Table 1](#)). Applying the signature in the training cohort yielded a sensitivity of 98.86% and a specificity of 82.35% for cHL in the training dataset ([Figure 2A](#)) and a specificity of 88.32% and sensitivity of 89.77% in the testing dataset ([Figure 2B](#)). We also found that this signature also was demonstrated could effectively distinguish cHL from normal controls both in the training cohort (AUC=0.981) and the testing dataset (AUC=0.955) ([Figure 2C and D](#)).

Table 1 Characteristics of Four Gene Markers and Their Coefficients in the Diagnosis of cHL

Variable	Coefficient	Std. Error	Wald	P
ADD3	-3.03945	1.18793	6.5465	0.0105
FOXC1	3.48986	1.29980	7.2088	0.0073
LAPTM5	-2.59629	0.86762	8.9546	0.0028
MAL	1.92006	0.81152	5.5980	0.0180
Constant	31.23070	11.98438	6.7910	0.0092

Unsupervised hierarchical clustering analysis of ADD3, FOXC1, LAPTM5 and MAL showed high specificity and sensitivity to differentiate cHL from normal controls (Figure 2E and F).

Construction of Weighted Co-Expression Network and Identification of Hub Genes

Six thousand genes in GSE12453 were obtained and utilized to construct the co-expression network (Figure 3A). The power of $\beta = 28$ was chosen as the best soft-thresholding parameter (Figure 3B and C). Additionally, the closely

correlative modules were merged into a bigger one by setting the MEDissThreasas parameter as 0.25 (Figure 3D). Ultimately, 10 modules were generated and the yellow modules showed significantly positive correlation with cHL patients (weighted correlation = 0.95, $P=2e^{-19}$, Figure 3E). Five hundred and seventy candidate genes selected by WGCNA were used to screen survival-related mRNA by conducting the analysis of univariable Cox survival. One hundred and seventy survival-related mRNA were screened out and used to construct the PPI, only the 18 genes 'degree in the PPI network > 10 were figure out (Supplement Table 1).

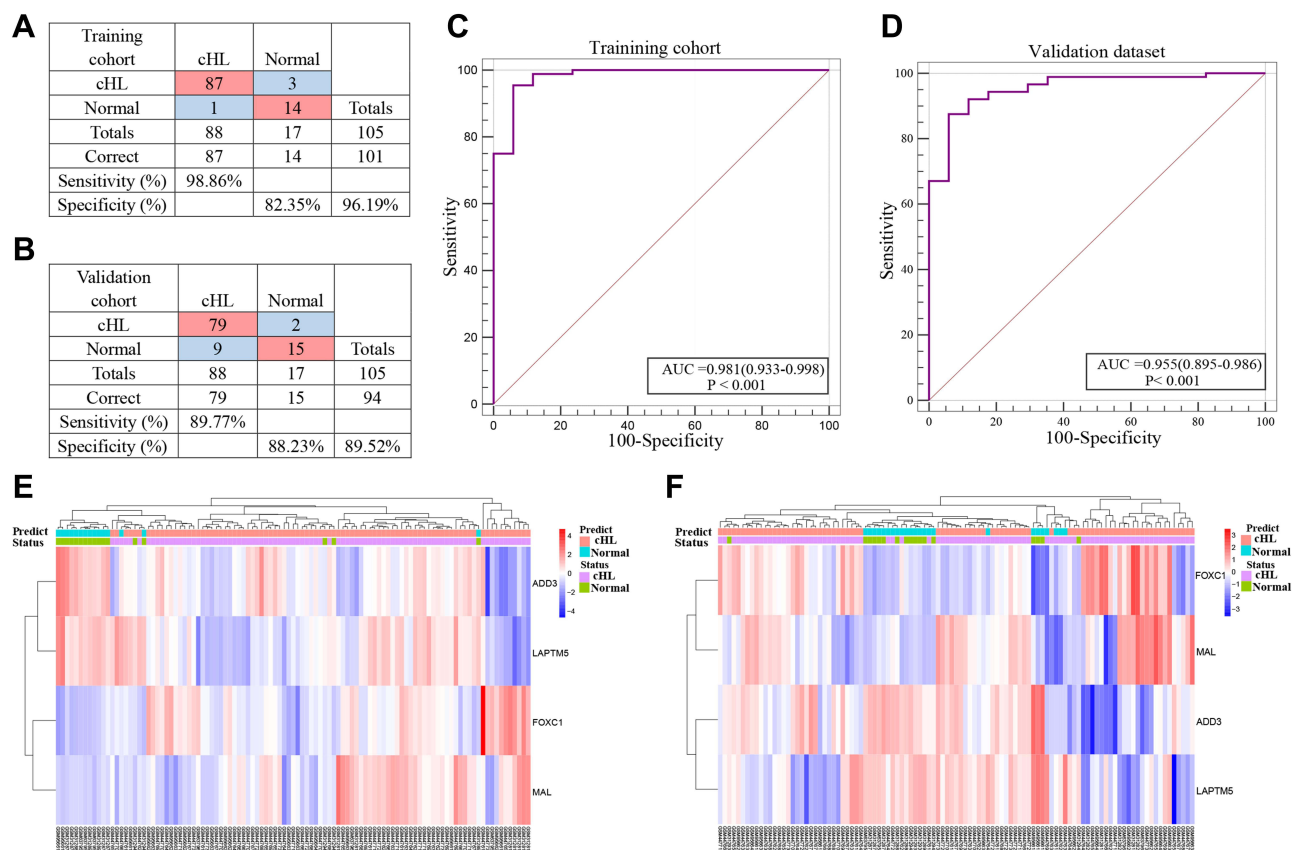


Figure 2 mRNA expression analysis of cHL diagnosis. (A and B) Confusion tables of binary results of the diagnostic prediction signature in the training and validation cohorts. (C and D) ROC of the diagnostic prediction signature with four mRNA markers in the training and validation cohorts. (E and F) Unsupervised hierarchical clustering of four mRNA markers in the diagnostic prediction model in the training and validation cohorts.

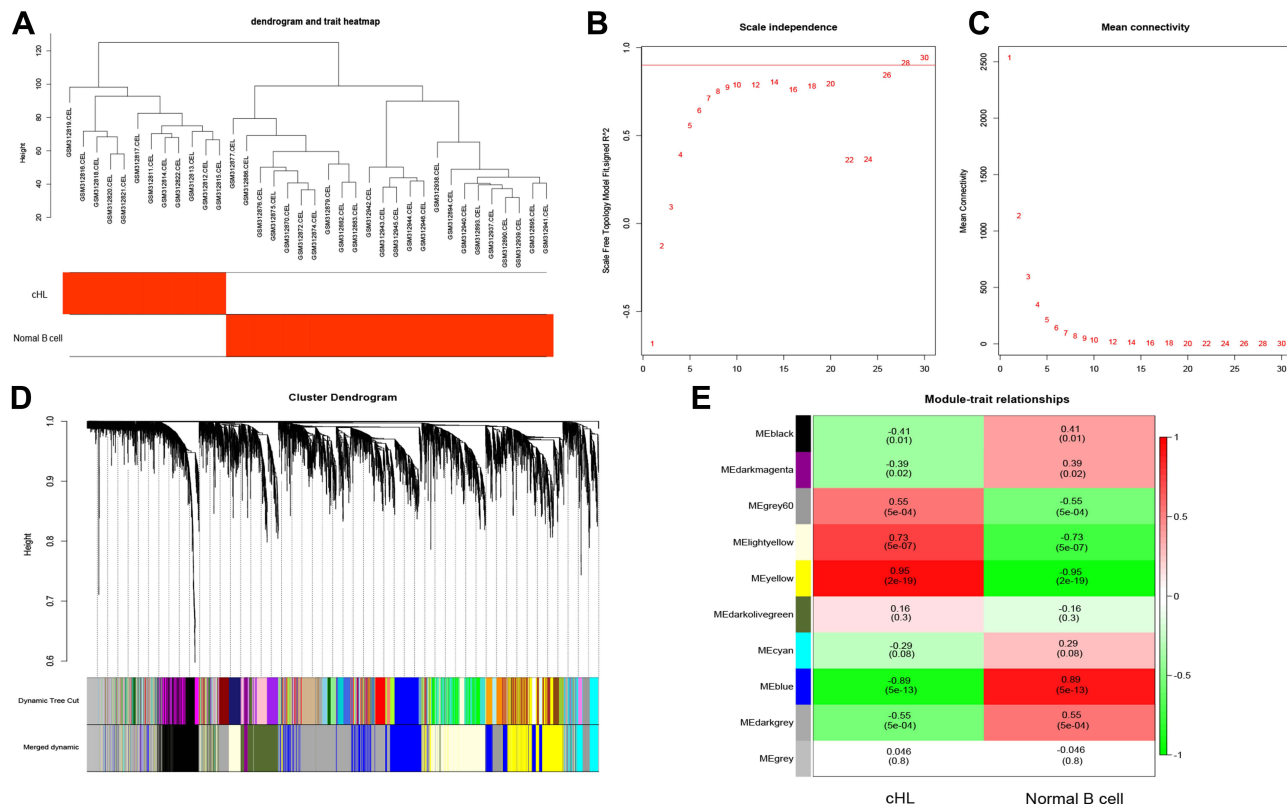


Figure 3 Identification of candidate genes in cHL. **(A)** Clustering dendrogram of cHL and normal B cell. **(B and C)** analysis of scale-free fit for soft thresholding powers and 28 was selected as the best value. **(D)** Dendrogram of all 6000 genes clustered on a dissimilarity measure. **(E)** Heatmap of the relationships between modules and cHL by Pearson correlation.

We evaluated the relationship between the expression of 18 genes and the survival of the patient, and the results showed that 9 genes including LAMP1, STAT1, MMP9, C1QB, ICAM1, CD274, CCL19, HCK, and LILRB2 were significantly negatively correlated with the prognosis of the patient with cHL in GSE17920 (Figure 4). The 9 genes had significantly high expression in cHL compared to normal B cells within the combinational data set with 210 case samples except HCK (Figure S4A).

Identification of the Nine-Gene Risk Prognostic Model for Survival

The nine hub genes were used to perform multivariate regression analysis and build a risk prognostic model in the GSE17920 cohort. The risk score of prognostic model was calculated with the following formula: risk score = (0.6891×STAT1 expression) + (5.1633× LAMP1 expression) + (0.1953× GBP2 expression) + (0.4689 × ICAM1 expression) + (1.8556×C1QB expression) + (0.7998×MMP9 expression) + (0.6767× LILRB2 expression) + (0.9942× HCK expression) + (1.5728× CCL19 expression) + (1.5587× 1.5587 expression). All patients diagnosed with

cHL were categorized into a high- and low-risk groups according to the median value of the risk score, which was illustrated by Kaplan-Meier curves (Figure 5A). The Kaplan-Meier curves also showed a significantly different prognosis in patients separated by staging (p<0.001, Figure 5B). The 1-year, 3-year, and 5-year areas under the curve were 0.735, 0.768 and 0.785, respectively (Figure 5C). Among these, nine genes, all significantly elevated in the group with high-risk (Figure S4B). We also found that the risk score is positively correlated with staging, and the risk score significantly increased in the group of hemoglobin <10.5 g/dl, serum albumin <4 g/dl and age ≥45 Years (Figure 5D, E, H, J). There is no statistical difference in the distribution of the risk score in the groups with different lymphocyte rate, white cell, and sex (Figure 5F, G, I). The distribution of patients with different clinical characteristics between high risk and low risk patients was calculated, and found that patients who younger than 45 years old, with stage I–II and albumin greater than 40mg/DL were obviously concentrated on low-risk group. And there is no statistical difference between different gender, hemoglobin, white cell count and

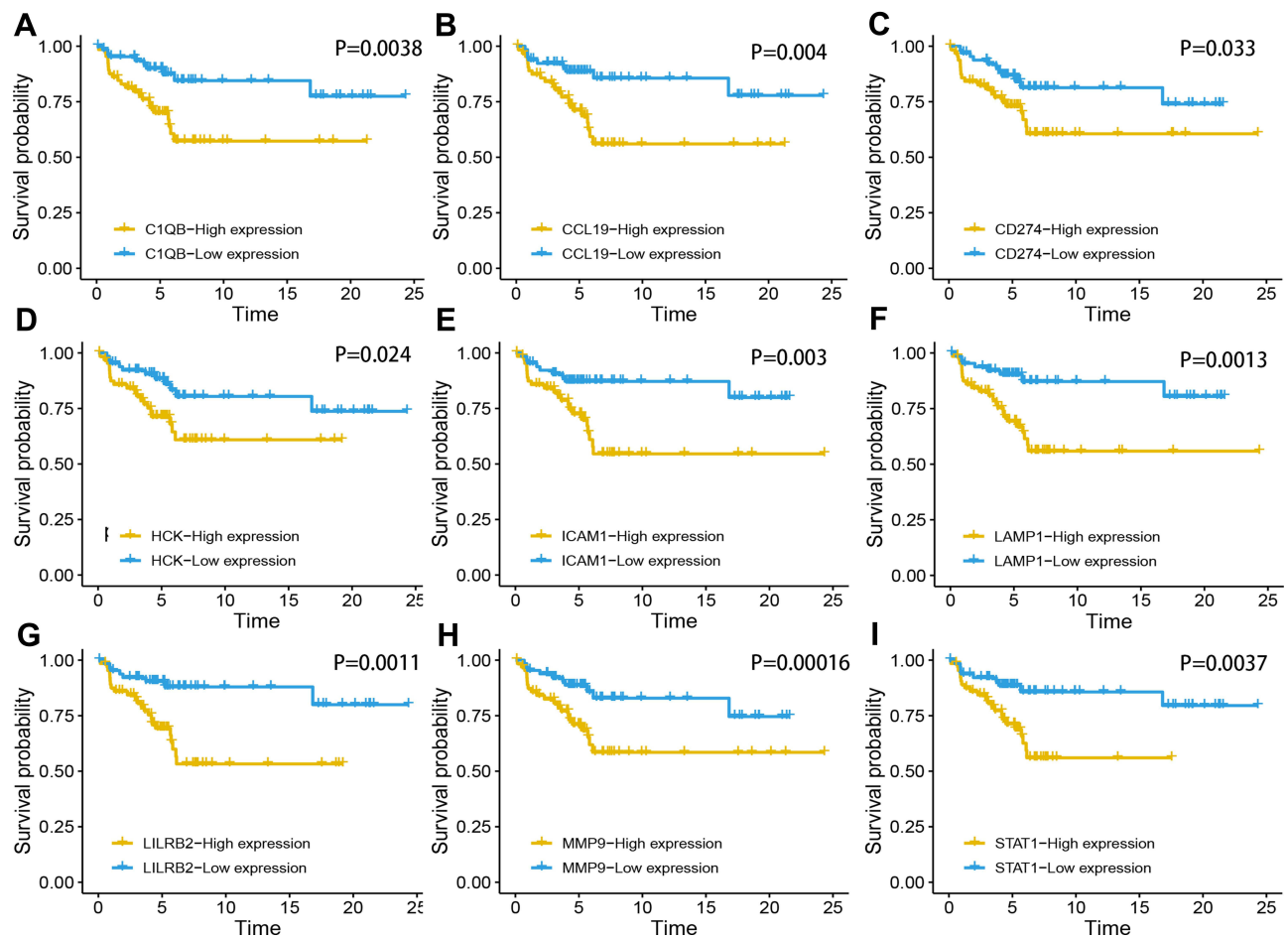


Figure 4 Survival analysis of nine genes in cHL. Kaplan–Meier survival curves were generated for (A) C1QB, (B) CCL19, (C) CCL19, (D) HCK, (E) ICAM1, (F) LAMP1, (G) LILRB2, (H) MMP9 and (I) STAT1.

lymphocyte rate in different risk groups (Figure S5A–G). Furthermore, cytarabine and vinblastine in our study had a lower IC50 in low-risk patients, suggesting that low-risk patients are more sensitive to cytotoxic drugs (Figure 6D and E), and there is no difference in the drug sensitivity of bleomycin doxorubicin, etoposide, and methotrexate between the high-risk category and the low-risk category (Figure 6A, B, C, F).

Immune Infiltration Assessment Based on Sample Type and Risk Model

After the analysis of CIBERSORT immune was completed, we found that the distribution of immune cells in cHL compared to normal tissues is similar to the distribution of immune cells in the high-risk group compared to the low-risk group. In the combinational cohort incorporated by all cHL and normal samples, 176 cases of cHL and 34 normal samples had shown a substantial divergence in the

proportion of nine types of immune cells (M1 macrophages, activated CD4+ memory T cells, CD8+T cell, activated dendritic cells, naive B cells, resting mast cells, memory B cells, resting NK cells and activated NK cells, Figure 7A). The presence of nine immune cells types (activated dendritic cells, naive B cells, M1 macrophages, CD8+T cells, resting mast cells, M0 macrophages, helper follicular T cells, resting memory CD4+ T cells, eosinophils, Figure 7B) in the low-risk category and the high-risk category also showed a considerable discrepancy. Among the 22 types of immune cell, the content of CD8+T cells, resting mast cells and M1 macrophages significantly high in cHL and high-risk category contrast to normal controls and low-risk category, which contrary to the distribution of naive B cells. The results indicating that the high scale of the fraction of the CD8+T cells, M1 macrophages and resting mast cells led to adverse outcome in cHL and naive B cells related to prolonged survival.

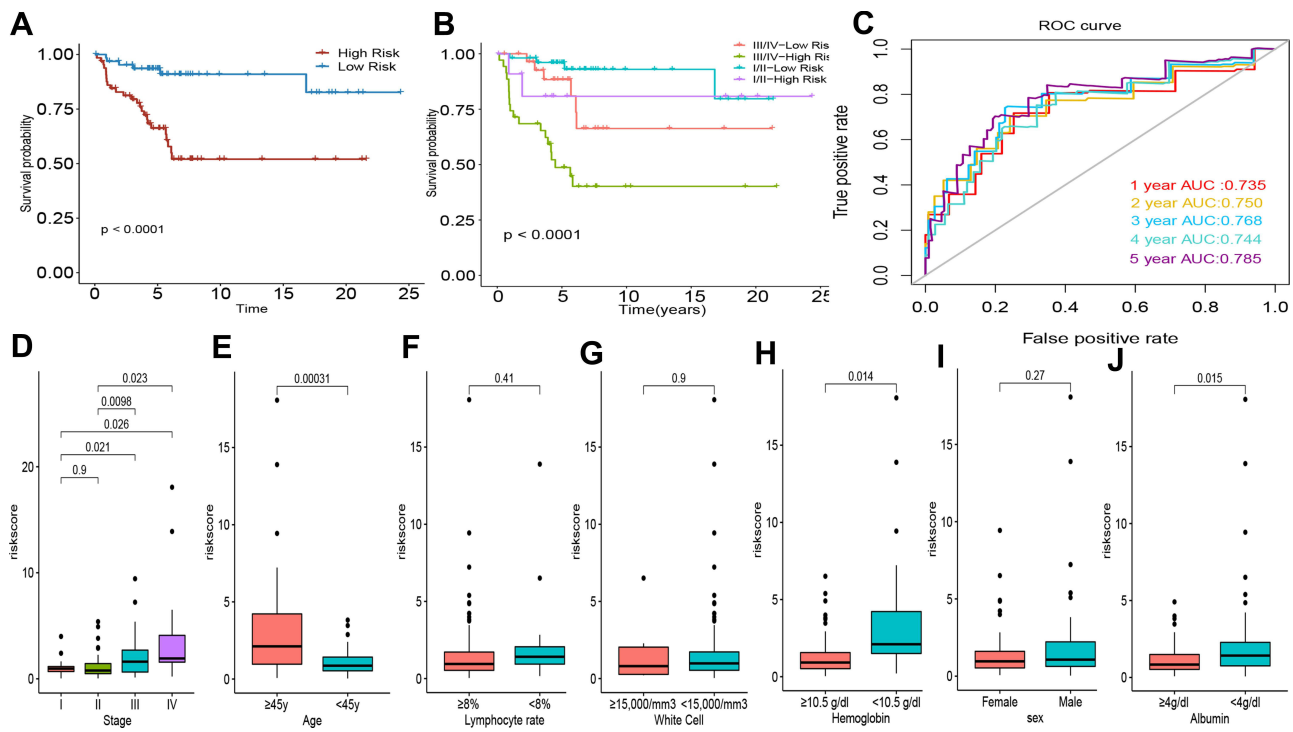


Figure 5 Analysis of the prognostic risk model in cHL. (A) Kaplan-Meier survival curves for all cHL patients; (B) Survival curves of cHL patients with combinations of risk core and stage. (C) time-dependent ROC curves for the nine-mRNA risk model in the cohorts of GSE17920. (D) Risk score in patients with different stage. (E) Risk score in patients with age of ≥ 45 and < 45 y. (F) Risk score in patients with lymphocyte rate $< 8\%$ and $\geq 8\%$. (G) Risk score in patients with white Cell of $\geq 15,000$ and $< 15,000/\text{mm}^3$. (H) Risk score in patients with hemoglobin < 10.5 g/L and ≥ 10.5 g/L. (I) Risk score in female and male patients. (J) Risk score in patients with Albumin < 40 g/L and ≥ 40 g/L.

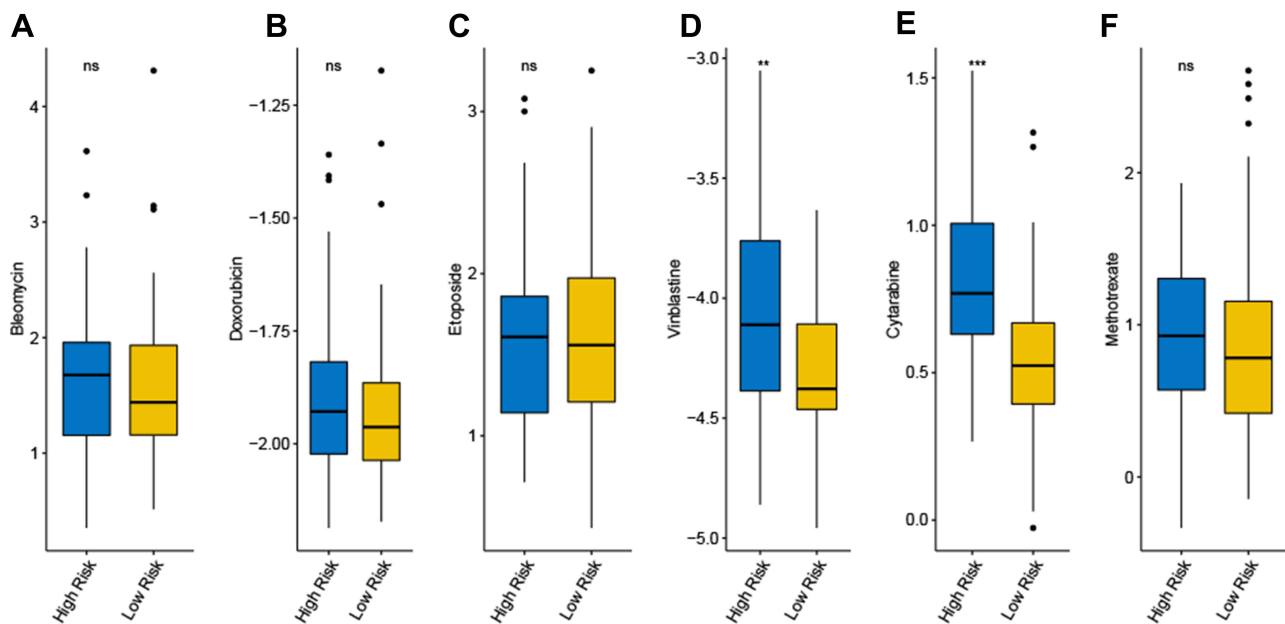


Figure 6 Prediction of chemotherapy of six common therapeutic drugs in patients of high risk and low risk. (A) Bleomycin; (B) Doxorubicin; (C) Etoposide; (D) Vinblastine; (E) Cytarabine (F) Methotrexate.(ns not significant, $*P < 0.05$; $**P < 0.01$ and $***P < 0.001$).

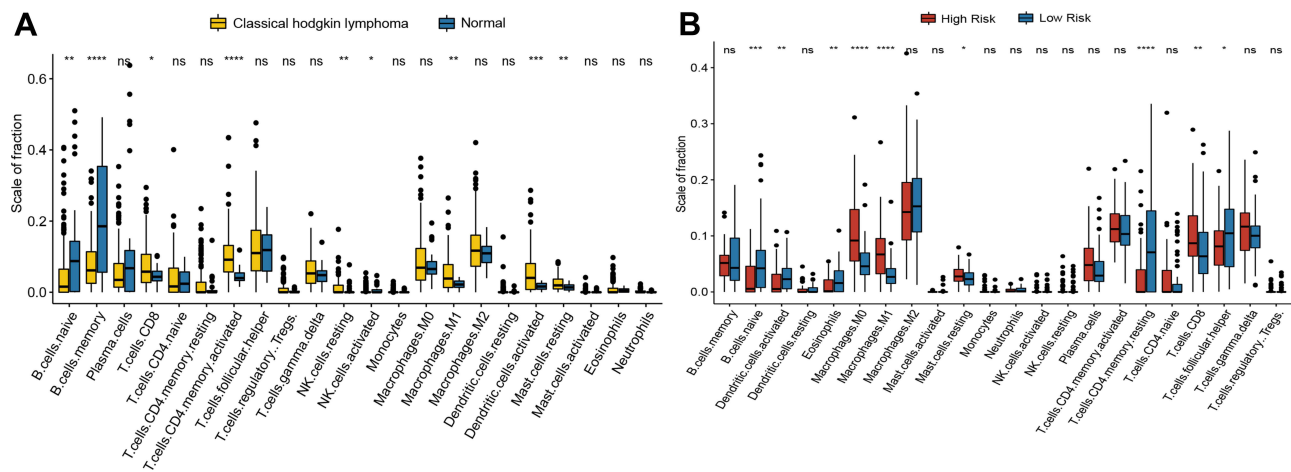


Figure 7 Immune infiltration assessment based on sample type and risk model. **(A)** Differential distribution of immune cells between the cHL and low normal control in the merged cohort. **(B)** Distribution of immune cells between the high risk and low risk group.(ns not significant, *P < 0.05; **P < 0.01; ***P < 0.001; and ****P < 0.0001).

Independent Role of the Risk Model in Patients with cHL

In order to evaluate whether the risk model has independent role in evaluating the prognosis of cHL patients, univariate and multivariate Cox regression analysis were performed in GSE17920 dataset by including the risk stratification and all available clinical characteristics as explanatory variables. According to the international prognostic score (IPS) criterion, all clinical characteristics parameters were grouped by hemoglobin <10.5 g/dl, age ≥45 years, male sex, lymphocyte count <8% of white cell count, white cell count (WCC) ≥15,000/mm³, serum albumin <4 g/dL, stage III/ IV disease by Ann Arbor classification. In univariate Cox regression, age, stage, hemoglobin, WCC and risk stratification were significantly related to OS. After multivariate adjustment was completed, risk stratification, age, stage, and WCC also showed an independent role in affecting the prognosis (risk stratification: HR = 4.4, 95% CI = 1.57–12.35,

P=0.0048, Table 2). A nomogram which including all clinical characteristic parameters and risk stratification was constructed and the calibration curve also exhibited high consistency between predictive outcome and observation survival time in the cHL cohort, and C-index for OS were 0.852 (Figure S6A and B).

Discussion

cHL is a kind of malignancy that originates from B cells and typically involved lymphonodus, sometimes other organs. The pathogenic mechanisms of cHL are still not very clear. In the present research, we investigate the molecular mechanism of cHL by using bioinformatics methods and try to provide a new clue to the development of diagnosis and risk stratification. In the present study, we built and validated a mRNA signature with four genes (MAL, FOXC1, LAPTM5, and ADD3) for the diagnosis and prediction of cHL, with a sensitivity of 98.86%,

Table 2 Univariate and Multivariate Cox Regression Analyses of the Nine-Genes Risk Model in Patients with cHL

Variable	Univariate Analysis			Multivariate Analysis		
	HR	95% CI	P	HR	95% CI	P
Risk Score (High/Low)	5.4	2.10–13.0	0.00033	4.4	1.57–12.35	0.0048
Gender (Male/Female)	5.0	2.40–11.0	0.058			
Age (≥45/<45y)	2.1	0.97–4.60	2.60e-05	2.6	1.03–5.51	0.041
Stage (III–IV/I–II)	3.2	1.50–7.00	0.0033	1.2	0.35–2.46	0.88
Hemoglobin (<105 g/L/≥105 g/L)	5.4	2.50–12.0	1.90e-05	2.6	0.96–5.59	0.061
Albumin (<40 g/L/≥40 g/L)	2.1	0.84–5.10	0.11			
White Cell (≥15,000 /<15,000/mm ³)	5.4	2.00–15.0	0.00074	6.4	2.43–24.5	0.0005
Lymphocyte rate (<8% /≥8%)	0.98	0.34–2.80	0.97			

a specificity of 82.35%, and an AUC of 0.981 (95% CI, 0.933 to 0.998) in the training dataset, and with a sensitivity of 89.77%, a specificity of 88.23% in the validation dataset. In addition, we identified nine genes that included LAMP1, STAT1, MMP9, C1QB, ICAM1, CD274, CCL19, HCK and LILRB2 with a significant prognosis of cHL, and a nine-mRNA-based risk model was developed to better assess the risk of the patient with cHL.

Several candidate genes for gene expression or genome methylation-based markers have been proposed to predict cancer diagnosis, such as the one study conducted by Xu et al in which a diagnosis signature was used to discriminate hepatocellular carcinoma and healthy controls, and achieved AUC values of 0.966 and 0.944, respectively.¹² Another research to assess the accuracy of a signature for detecting colorectal cancer patients in the training dataset and validation dataset, the results showed the AUC of 0.851 and 0.923, respectively.¹³ In the present study, our diagnostic prediction signature showed better performance in differentiating cHL from normal controls and yielded new insights into the diagnosis of cHL. FOXC1 in our signature is an essential component of FOX family members which involved in carcinogenesis and the growth tumor cell and express a higher level in a variety of carcinomas.¹⁴ Overexpression of FOXC1 has also been observed in the HL.¹⁵ Furthermore, FOXC1 has been demonstrated to activate the NOTCH and NF- κ B signaling pathway, and these pathways are the two most important pathways in cHL to promote the survival of HRS cells.¹⁶ cHL and mediastinal large B-cell lymphoma (MLBL) have similar molecular genetics, histopathology and clinical manifestations. Myelin and lymphocytes (MAL) has been reported to overexpressed in the majority of MLBL and a minority of cHL, but the role of MAL in those diseases have not reach definitive agreement. In this context, elevated expressions of FOXC1 and MAL were found in the cHL group compared to the normal group, consistent with the observation mentioned in the above research. Widespread loss of classic B lineage phenotype markers in HRS cells is the most important features of cHL,⁹ which in line with our results that the proportion of memory B cells and naïve B cells in cHL is significantly lower than which in the control group. There is evidence that lysosomal-associated protein transmembrane 5 (LAPTM5) is abundantly expressed in mature B cells.¹⁷ In our study, LAPTM5 was significantly down-regulated in cHL compared to the normal control; this may be attributed to the

decrease in the number of normal mature B cells and leads to decreased expression of LAPTM5. Adducin 3 (ADD3) is a crucial membrane skeleton protein and has been confirmed to be aberrantly low expressed in multitype cancers, and loss of ADD3 can promote tumor cell growth and angiogenesis that occurs in glioblastoma multiforme.¹⁸ Moreover, ADD3 deletions were significantly negatively correlated with prognosis.¹⁹

Epidemiological and serologic studies have implicated that approximately 40% of Hodgkin lymphoma cases are associated with Epstein-Barr virus (EBV). Epstein-Barr virus infection and genetic alterations at 9p24.1 contribute to overexpression of CD274/PD-L1,⁶ which in turn contributes to HRS cells evading T cell immune surveillance.²⁰ Overexpression of PD-L1 expression has been detected and associated with poor prognosis in diffuse large B-cell lymphoma, nasopharyngeal carcinoma and colorectal cancer,^{21–23} similar conclusions have also been reported in cHL.⁹ However, the interesting thing is that was a trend with higher HRS cell CD274/PD-L1 expression, patients exhibit more favorable responses to anti-programmed death-1 monoclonal antibody and had superior outcome.^{3,24} In our study, CD274 has significantly high expression in patients with cHL and high-risk group, which means that patients in the high-risk group will benefit more from immunotherapy. STAT1, ICAM1 and CCL19 which also have been found to be significantly elevated in patients with cHL and correlated with a short period of survival of the cHL patient. High levels of STAT1 contribute to proliferation and bad outcomes of breast cancer,^{25,26} however, it has also been demonstrated that high levels of STAT1 has tumor suppressive functions and are beneficial to prognosis in some selected cancer.^{27,28} The divergent role of STAT1 in tumorigenesis and development can be due to heterogeneity in different type of cancer and Different tumor microenvironment.²⁹ Accumulating evidences suggest that Intercellular cell adhesion molecule-1 (ICAM-1) upregulated in multitype types of cancer and plays an important role in tumor invasion and metastasis,^{30,31} but it also has been proposed that elevated ICAM1 expression in colorectal cancers, gastric cancer and breast cancer is correlated with more favorable prognosis.³² CCL19 is one of the chemokines and has been found to be overexpressed in many tumors, it can not only modulate the immune response and take part in the proliferation of cancer,³³ but also recruit tumor cells to the T cell zone, leading to lymph node metastasis,^{34,35} in addition, it was discovered that CCL19 in signaling

pathway of NF- κ B can promoting central nervous system lymphoma (CNSL) formation.³⁶

Chemokines and cytokines which produced by the HRS cells as and inflammatory cells in the microenvironment of cHL work in some kind manner to promote the survival of HRS cells.³⁷ Based on current research it is evident that chemokines secreted by HRS cells and inflammatory cells contribute to mediate the expression of MMPs that needed for malignant cell invasion and metastasis via degradate the extracellular matrix (ECM).³⁸ MMP9 is a major constituent of the MMPs and has been identified as a prognostic factor in glioblastoma and clear cell renal carcinoma.^{39,40} Lysosome-associated membrane protein-1 (LAMP1) is a member of the lysosomal membrane protein that protects the lysosomal membrane from hydrolysis, growing evidence has shown LAMP1 participate in process of tumor cell and overexpression of it correlates with adverse outcome of diffuse Large B-cell Lymphoma and breast cancer,^{41,42} but the exact role involved in cancer is still unknown. Leukocyte immunoglobulin-like receptor sub-family B member 2 (LILRB2) has also been reported to be associated with promoted tumor cell growth and correlates with poor prognosis,^{43–45} which is consistent with our study. LILRB2 antibodies can not only inhibit AKT and STAT6 activation but also increase the efficacy of anti-PD-L1 and enhance antitumor immunity,⁴⁶ suggesting that LILRB2 may act as a novel potential immune-targeted therapeutic checkpoint.

Hematopoietic cell kinase (HCK) is a kind of protein tyrosine that belongs to SRC kinase family, which originally found in the cells of hematopoietic and now it also been studied in other solid tumors and lymphoma.⁴⁷ Blocking HCK can reduce the activation of MAPK/ERK and PI3K/AKT pathways which has been widely recognized to promote the formation of various cancers.⁴⁸ Highly expressed HCK is negatively correlated with colorectal cancer patients' prognosis,⁴⁹ which in line with our results in cHL. Complement component 1, Q subcomponent B chain (C1QB) is a submember of the complement classical pathway; it has been demonstrated to enhance chemokine secretion that are expected to promote immunosuppression and play a role in tumor growth and progression.⁵⁰ To date, C1QB overexpression in gliomas has been associated with a reduced prognosis,⁵¹ however, recent research revealed that high C1QB in breast cancer is related to a prolonged prognosis.⁵² Implementing a further study on where C1QB can have a protective or deleterious effect on cancer progression is necessary. In this context, the expression of C1QB increased

significantly in cHL patients compared to the normal group and was associated with poor outcomes.

Advances in bioinformatics technology contribute to the construction of multigene risk signature by using public data sets that have been used,⁵³ however, the gene-based risk signatures and the nomogram based on it that as a prognostic tool for cHL has not yet been investigated. In this report, a risk model was constructed and showed powerful performance to stratify all cHL patients into low-risk and high-risk groups, and exhibit an independent impact on the prognostic value of cHL. To further investigate the prognostic value of the risk model, a nomogram was built to predict the personalized risk of every cHL patient. The c index used to evaluate the effectiveness of the nomogram in cHL patients was 0.852, significantly better than previous research used to predict the outcome of other cancer patients.⁵⁴ Having the ability to accurately assess individual risk and prognosis is the basis of personalized treatment for patients, and the risk model may add complementary value to clinicians in making treatment decisions.

However, the limitations of the present study should also not be ignored. First, the sample size to identify the marker for diagnosis and prognosis needs to be further expanded to avoid selection bias. Secondly, the lack of available validation groups to further confirm the reliability of the risk model in the study. Finally, further experimental research is needed in our future study to elucidate the mechanism of these markers in the carcinogenesis, proliferation and progression of cHL.

Collectively, this is the first study that has been used to investigate the ability of the signature based on mRNA as a novel biomarker for the diagnosis and prediction of the prognosis for cHL. In present study, we identified a diagnostic prediction signature, nine prognostic genes, and a prognostic prediction model, which has been confirmed the usefulness in diagnosis and prognosis prediction of cHL. The novel genes may be helpful for researchers to find new therapeutic targets.

Data Sharing Statement

All data generated or analyzed during this study are included in this article.

Ethics Approval and Consent to Participate

The need for ethics approval was waived by the Department of Scientific Research Management, Changzhou Tumor Hospital Affiliated to Soochow University.

Acknowledgments

This work was supported by a grant from Startup Fund for scientific research, Fujian Medical University (2018QH1171). And all authors would like to thank Christian Steidl, MD, for sharing survival information of dataset GSE17920.

Disclosure

The authors declare that they have no competing interests.

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