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Dispersal and niche evolution jointly shape the geographic turnover of phylogenetic clades across continents

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The turnover of phylogenetic clades across space is a fundamental biodiversity pattern that may depend on long-term evolutionary processes, and that has downstream effects on other aspects of diversity including species richness and community structure. Limited niche evolution and limited dispersal are two major processes causing spatial restriction, and thus turnover, of clades. We studied the determinants of clade turnover within the World's richest floristic kingdom, the Neotropics, using the palm family (Arecaceae) as a model. We show that continental-scale clade turnover is driven by a combination of limited niche evolution — with respect to temperature and soil tolerances — and limited dispersal. These findings are consistent with strong dispersal barriers within the Neotropics, and the observation that some palm lineages are most diverse in certain biomes or climates. The importance of such deep-time effects suggest that palms might be slow to adapt or disperse in response to anthropogenic climate change.

Most groups of organisms are geographically restricted in their occurrence, sometimes becoming emblematic for a region, like the lemurs for Madagascar or the palms for the tropics. This turnover of phylogenetic clades in space is a fundamental biodiversity pattern that has long fascinated biogeographers¹. Two major factors seem to restrict the geographic distribution of clades: limited dispersal and limited β -niche evolution (β -niche refers to the niche dimensions that determine a species' distribution in space²). By limited dispersal we mean the failure of individuals to reach certain environmentally suitable areas, whereas by limited β -niche evolution we mean the failure to evolve adaptations to certain environments. The relative importance of these two factors is largely unstudied and likely to vary among taxa, regions and biomes³. Of note, geographic turnover of clades can lead to other diversity patterns, such as variation in species richness or the functional composition and diversity of assemblages, if there is variation in species richness or morphology among clades^{4–6}. The factors leading to among-clade differences in species richness have received considerable attention⁷, while the factors constraining those clades in space — although a crucial piece of the puzzle — have mainly been discussed for a few prominent examples^{8,9}.

Tropical America harbours the most diverse flora of the world¹⁰, and the origin of this wealth of plant species has received intense scientific scrutiny^{3,11–13}. Scenarios of Neotropical plant evolution involve effects of both dispersal barriers and ecological factors, such as biome conservatism^{3,14}. Influential historical events include the long-time separation of South America from North and Central America and the subsequent Great American Biotic Interchange, marine incursions into the Amazon basin, and the uplift of the Andes^{11,15}. These geomorphological reorganizations caused strong changes in dispersal opportunities and habitat distribution, with abundant dispersal barriers and steep ecological gradients. Here, we provide new insights into the complex history of Neotropical plant diversification by analysing the geographic turnover of clades within a major tropical plant family, the palms (Arecaceae).

Palms are a pantropical group largely restricted to megathermal climates¹⁶ and a textbook example of tropical niche conservatism¹⁹. Approaching one third of the family's 2,400 species is found in the New World¹⁷, with all but one of native species being endemic. American palms are distributed between 34°N and 34°S (Fig. 1a), from sea level to 3600 m.a.s.l., and occur under a wide variety of environmental conditions¹⁸. It is known that the broad-scale distributions of palm species depend on both environment — mainly climate — and dispersal¹⁹. In the

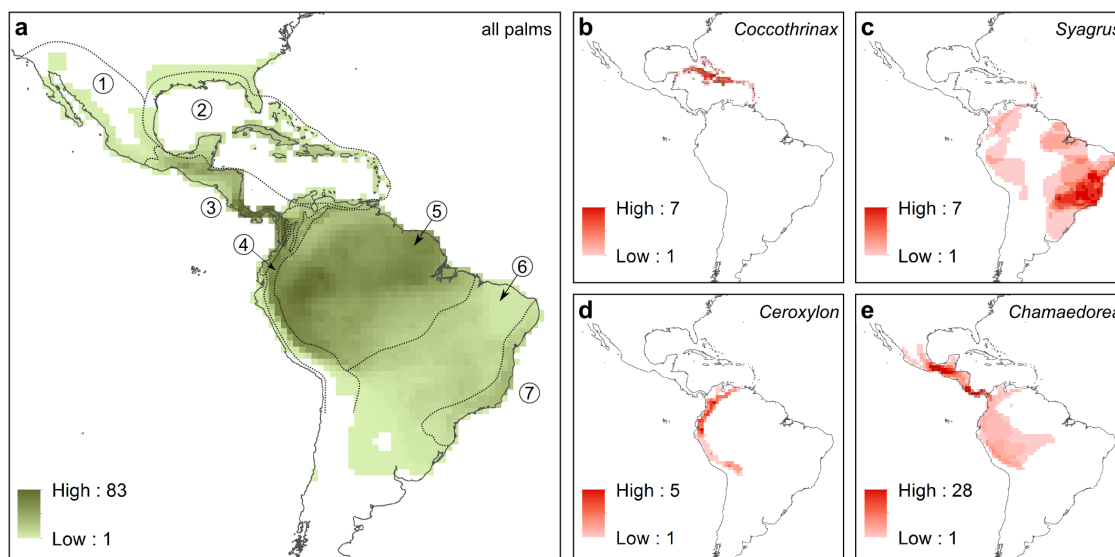


Figure 1 | a: Distribution, species richness, and biogeographic regions of palms in the Americas. Green shading indicates the species richness of palms in $1^\circ \times 1^\circ$ grid cells. Stippled lines indicate the biogeographic “palm regions” suggested by A. Henderson¹⁸: (1) Mexican region, (2) Caribbean region, (3) Central American region, (4) Andean region, (5) Amazon region, (6) Central Brazilian region, (7) Atlantic coastal forest. b–e: Examples of palm lineages with a spatially restricted distribution. b: *Coccothrinax* (14 spp., stem age: 10 mya), c: *Syagrus* (30 spp., 8.5 mya), d: *Ceroxylon* (11 spp., 11 mya), e: *Chamaedorea* (77 spp., 19 mya)^{18,58}.

Americas, many palm lineages have a spatially restricted distribution (e.g. Fig. 1 b–e). Based on the distribution of palm genera, seven biogeographic “palm regions” were proposed as putative centres of diversification (Fig. 1a)¹⁸. Geographic turnover of palm clades is also reflected in strong phylogenetic clustering of palm assemblages in “botanical countries” (TDWG level 3 units)²⁰. A previous study indicated that the turnover of larger palm clades (subfamilies) is shaped by both β -niche evolution and dispersal limitation¹⁴. The subfamilies Arecoideae and Calamoideae were most species-rich in areas with high humidity and fertile soils, indicating limited capacity for adaptation to other environments¹⁴. Species diversity within the palm subfamilies peaked in different regions after controlling for the current environment, suggesting dispersal-limited diversification around different areas of origin¹⁴. However, the numerically dominant Arecoideae appeared to be close to equilibrium with the current environment, showing little evidence for dispersal limitation at the clade level¹⁴. Since this previous study only analysed a few predefined clades (the subfamilies), it remains unclear to what degree American palm lineages in general are spatially restricted by limited β -niche evolution or by dispersal.

Limited dispersal and limited β -niche evolution should lead to predictable relationships between phylogenetic turnover (often called “phylogenetic beta diversity”²¹, but see ref. 22) and environmental and spatial distances among assemblages (Table 1). Analysing these relationships has previously provided important insights into community assembly, albeit at much smaller scales than

in the present study^{23–25}. Applied at large spatial scales, this approach may complement insights from biogeographic analyses²⁶ and phylogenetic assemblage structure²⁰. At regional scales, phylogenetic turnover is often influenced by speciation (mechanisms M_1 and M_2 in Table 1), causing closely related species to have strongly disjunct distributions (allopatric speciation) or strongly divergent β -niches (adaptive speciation along spatio-environmental gradients)²⁴. At larger (continental) scales this effect is likely less pronounced, and phylogenetic turnover is dominated by limited β -niche evolution and/or limited dispersal (mechanisms M_3 and M_4 in Table 1).

The extent to which limited dispersal leads to predictable relationships between phylogenetic turnover and spatial distance should depend strongly on the environmental heterogeneity of the study region. Even spatially close locations may be separated by strong dispersal barriers (e.g. mountain ranges, seaways), whereas even distant locations may be connected by dispersal corridors (i.e. habitat similar to the locations themselves). If a correlation between phylogenetic turnover and spatial distance is indeed due to limited dispersal (M_3 in Table 1), it should increase when spatial distance is weighted with the occurrence of barriers between locations. Measuring barriers is not trivial, since any given habitat (e.g. mountain peaks) may constitute a barrier for some species (e.g. lowland rainforest species), but a dispersal corridor for others (e.g. alpine species). We developed a new measure of “barrier distance” that accounts for such differences. For each pair of locations, we mapped habitat dissimilarity — relative to the habitat at the locations themselves — throughout the study region.

Table 1 | Determinants of phylogenetic turnover among assemblages (see also^{21,47})

Mechanism	Prediction for environmental dissimilarity (d_e)	Prediction for spatial distance (d_s)
M_1 : Allopatric speciation	No relationship between phylogenetic turnover and d_e , if controlled for d_s	Phylogenetic turnover <i>decreases</i> with d_s
M_2 : Adaptive speciation along spatio-environmental gradients	Phylogenetic turnover <i>decreases</i> with d_e	No relationship between phylogenetic turnover and d_s , if controlled for d_e
M_3 : Dispersal limitation	No relationship between phylogenetic turnover and d_e , if controlled for d_s	Phylogenetic turnover <i>increases</i> with d_s
M_4 : Limited β -niche evolution	Phylogenetic turnover <i>increases</i> with d_e	No relationship between phylogenetic turnover and d_s , if controlled for d_e



We then quantified the minimum amount of dissimilar habitat that needs to be crossed in order to get from one location to the other (i.e. the sum of dissimilarity scores of all grid cells crossed). Deviations from a straight line were allowed if this led to a lower barrier distance value. This captures the occurrence of dispersal corridors that indirectly connect locations. Of note, this barrier distance is inherently correlated with environmental distance between locations, and it can only be compared to pure spatial distance if environmental distance is controlled.

Based on previous knowledge of the geographical ecology of palms, we derived the following specific hypotheses for our study: *i.* Clade turnover among palm assemblages in the Americas is determined by limited dispersal (M_3) and limited β -niche evolution (M_4). Speciation patterns (M_1 and M_2) are, if at all, of minor importance at this scale, resulting in an overall positive relationship between both environmental dissimilarity and spatial distance and phylogenetic turnover. *ii.* The β -niche evolution effect is strongest (based on the numerical dominance of Arecoideae, see above). *iii.* Climate, particularly temperature extremes and seasonality, is most important for this β -niche evolution effect. *iv.* If an observed relationship between phylogenetic turnover and spatial distance is caused by dispersal (M_3 in Table 1), this relationship should be even stronger when spatial distance is replaced with a distance measure that takes dispersal barriers into account.

Results

The turnover of clades among palm assemblages in $1 \times 1^\circ$ grid cells (in the following: phylogenetic turnover) was significantly positively related to (a) spatial distance, (b) a measure combining spatial distance and the presence of barriers in between the cells (“barrier distance”), and (c) the overall environmental dissimilarity among the cells integrating climate, soils, topography, and remotely sensed vegetation structure (Fig. 2a–c). All assessments of significance were based on a phylogenetic null model (see Methods). Phylogenetic turnover was also significantly positively related to environmental

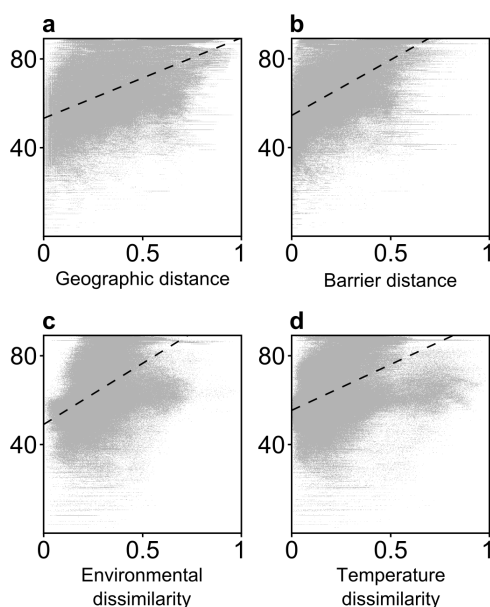


Figure 2 | Relationships between phylogenetic turnover (y-axis, in mya) and different distance measures. a: geographic (great circle) distance, b: barrier distance, an integrated measure of geographic distance and dispersal barriers, c: environmental dissimilarity based on 31 environmental variables describing climate, soil, topography, and vegetation structure, d: temperature dissimilarity based on 11 temperature variables only. All distances are rescaled to a range of [0, 1]. Dashed lines are regression lines.

dissimilarities that included only one aspect of the environment (i.e. temperature dissimilarity [Fig. 2d], precipitation dissimilarity, soil dissimilarity, topographic dissimilarity, and vegetation dissimilarity). When using individual environmental variables (e.g. temperature seasonality, isothermality, etc. – see Supplementary Table S1) to compute dissimilarity, 20 out of the 31 single-variable dissimilarities significantly predicted phylogenetic turnover. All results of the aforementioned models can be found in Supplementary Table S1.

The largest amount of variation in phylogenetic turnover (49%) was explained by a model that included barrier distance and dissimilarity in temperature seasonality (Supplementary Table S2). Models that included barrier distance consistently outperformed (i.e. explained 1.8–7.4% more variation, median 6.6%) the corresponding models with simple geographic distance. The fraction of variation independently explained by spatial or barrier distance (R_{pd}) was always significantly larger than R_{pd} of the null model ($P \leq 0.001$). The fractions of variation that were independently explained by environmental dissimilarity (R_{pe}) were generally smaller when barrier distance was used than when spatial distance was used, with the exception of two environmental variables that anyway showed negligible R_{pe} values; however, the median difference was small (0.06%). The degree to which the environmental variables co-varied with spatial/barrier distance was also highly variable; e.g. the fraction of variation explained by both environmental and barrier distance ranged from 0.2% to 25%, depending on the way environmental dissimilarity was computed. The different environmental variables also differed widely in their independent ability to explain variation in phylogenetic turnover (R_{pe} of 0–19% with spatial distance, 0–14% with barrier distance). Observed R_{pe} was not always significantly larger than R_{pe} of the null model (Fig. 3, Table 2). The single-variable

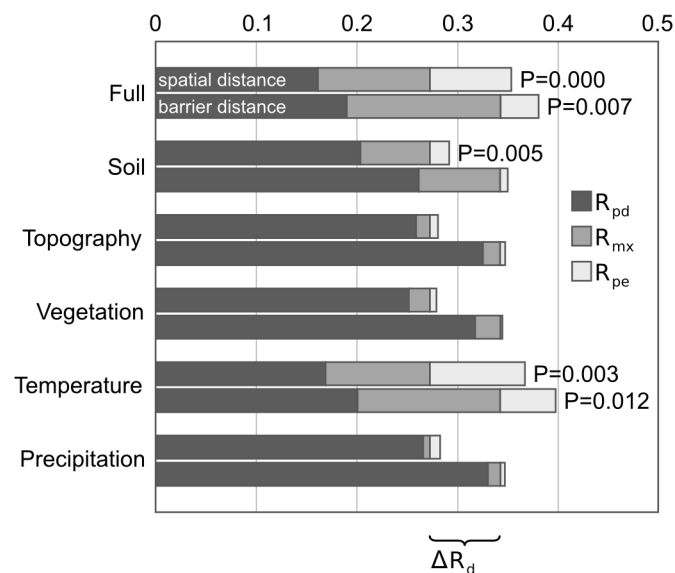


Figure 3 | Variation in phylogenetic turnover among palm assemblages explained by environmental dissimilarity and spatial/barrier distance. Pairs of bars correspond to different measures of environmental dissimilarity (Full = all variables PCA, Soil, Topography, Vegetation structure, Temperature, and Precipitation). The upper bar in each pair is based on spatial distance and environmental dissimilarity; the lower bar on barrier distance and environmental dissimilarity. R_{pd} : fraction of variation explained by spatial/barrier distance only, R_{mx} : fraction of variation explained by either spatial/barrier distance or environmental dissimilarity, R_{pe} : fraction of variation explained by environmental dissimilarity only. P values indicate the probability that the R_{pe} fraction is not larger than the R_{pe} fraction of the null model (H_0). Only P-values < 0.05 are shown. x-axis: fraction of variation in phylogenetic turnover.



dissimilarities of temperature seasonality, temperature annual range, and isothermality consistently had the highest independent ability to explain variation in phylogenetic turnover (Table 2). Mean temperature of the coldest quarter and minimum temperature of the coldest month also performed better than any other single-variable dissimilarity. However, the individual effects of these variables cannot be separated, because they are highly intercorrelated (Spearman's $|\rho| \geq 0.7$; see Supplementary Table S3). Temperature dissimilarity also explained more variation independently (R_{pe}) than any other category of variables (i.e. precipitation, soil, topographic, or vegetation dissimilarity), and performed even better than the measure including all environmental variables (temperature and non-temperature). Some non-temperature single-variable dissimilarities also had significant R_{pe} fractions, albeit with low amounts of explained variation. These included soil pH and base saturation, total exchangeable bases (only with spatial distance), precipitation of the wettest quarter and month, annual precipitation (only with spatial distance), and annual mean QSCAT, a microwave backscatter measure of vegetation structure. Soil as a group also had a significant effect ($P = 0.005$) when analysed together with spatial distance.

Discussion

Our expectation (hypothesis *i*) that both limited β -niche evolution and dispersal limitation would contribute to the spatial turnover of palm clades was strongly supported by the results (Fig. 3). Of note, the influence of the two factors cannot be fully separated due to

spatial autocorrelation in the environment; this is evidenced by the R_{mx} fractions of the variation partitioning (Fig. 3). However, the independent effects of limited niche evolution (R_{pe}) and dispersal limitation (R_{pd}) are sufficient to conclude that both factors have a significant and quantitatively non-negligible effect on the geographic turnover of clades.

Due to dispersal limitation, lineages often diversify within continents or islands, but fail to spread to other environmentally suitable landmasses. Within continents, dispersal limitation is more difficult to detect, because barriers are not always obvious and differ among species depending on their niche²⁷. Pure spatial distance can serve as a crude proxy for the amount of barriers between two locations²⁸, but this correlation is imperfect. Of note, pure spatial distance will also capture “time-for-dispersal” effects²⁹, i.e. a limitation that is not imposed by barriers, but by the limited speed at which populations can spread across a uniformly suitable landscape. This mechanism appears to be relevant sometimes even on evolutionary timescales³⁰. The significant effect of spatial distance on clade turnover in American palms (R_{pd}) can thus be interpreted as evidence for dispersal limitation on evolutionary timescales caused either by barriers or by time. We were able to gain additional insights by devising a complex measure of “barrier distance” that aimed to incorporate the species-specific suitability of areas as dispersal corridors. The fact that barrier distance explains more variation of phylogenetic turnover than spatial distance shows that the more complex measure better reflects dispersal barriers, and confirms that barriers play an important role for the distribution

Table 2 | Amount of variation in phylogenetic turnover explained by environmental dissimilarity based on separate environmental variables. Letters indicate variable category: T = Temperature, P = Precipitation, S = Soil, R = Remote sensing, To = Topography. R_{pe} is the amount of variation in phylogenetic turnover explained by environmental dissimilarity independent of spatial/barrier distance. The environmental variables were ranked according to the size of R_{pe} (“Rank”). The P-value is calculated as $n(R_{pe}^0 > R_{pe})/999$, where $n(R_{pe}^0 > R_{pe})$ indicates how many randomized phylogenetic turnover matrices produced higher R_{pe} than the non-randomized matrix in the null model. Bold variables have a significant R_{pe} at the $\alpha = 0.05$ level with both spatial and barrier distance

Variable	Category	Spatial distance			Barrier distance		
		R_{pe}	Rank	P	R_{pe}	Rank	P
Temperature Seasonality	T	0.198	1	0.006	0.144	1	0.012
Temperature Annual Range	T	0.166	2	0.002	0.121	2	0.005
Isothermality	T	0.142	3	0.002	0.104	3	0.007
Mean Temperature of Coldest Quarter	T	0.080	4	0.015	0.051	5	0.034
Min Temperature of Coldest Month	T	0.080	5	0.005	0.052	4	0.016
Soil pH	S	0.051	6	0.001	0.036	6	0.003
Soil Base Saturation	S	0.040	7	0	0.034	7	0
Precipitation of Wettest Quarter	P	0.035	8	0	0.022	9	0.004
Mean Temperature of Driest Quarter	T	0.035	9	0.059	0.022	8	0.108
Precipitation of Wettest Month	P	0.030	10	0.002	0.018	10	0.011
Annual Mean QSCAT	V	0.028	11	0.001	0.014	12	0.042
Annual Precipitation	P	0.027	12	0.009	0.016	11	0.052
Annual Mean Temperature	T	0.023	13	0.051	0.010	14	0.144
Mean Diurnal Temperature Range	T	0.019	14	0.029	0.011	13	0.089
Soil Total Exchangeable Bases	S	0.012	15	0.024	0.005	15	0.164
Altitudinal Range	To	0.008	16	0.145	0.004	16	0.273
Slope	To	0.006	17	0.221	0.003	17	0.378
Soil CaCO ₃	S	0.006	18	0.229	0	24	0.807
Mean Temperature of Wettest Quarter	T	0.005	19	0.317	0	25	0.898
Max Temperature of Warmest Month	T	0.004	20	0.277	0	21	0.722
Precipitation of Coldest Quarter	P	0.003	21	0.401	0.003	18	0.441
Annual Mean NDVI	V	0.003	22	0.486	0	31	0.991
Mean Temperature of Warmest Quarter	T	0.003	23	0.394	0	26	0.91
Soil Cation Exchange Capacity	S	0.001	24	0.316	0	30	0.929
Precipitation of Driest Quarter	P	0.001	25	0.61	0	27	0.913
Precipitation of Driest Month	P	0.001	26	0.621	0	29	0.944
Seasonal Variation of NDVI	V	0.001	27	0.592	0	23	0.729
Precipitation of Warmest Quarter	P	0.001	28	0.553	0.002	20	0.381
Precipitation Seasonality	P	0	29	0.704	0.002	19	0.393
Soil Sand Content	S	0	30	0.556	0	22	0.442
Seasonal Variation of QSCAT	V	0	31	0.878	0	28	0.929



of palm clades. This clear result is remarkable since barrier distance (i) is based on mean environmental conditions within $1 \times 1^\circ$ grid cells, which cannot capture the occurrence of smaller-scale dispersal barriers or corridors, (ii) ignores differences in niche breadth among species, and (iii) assumes that the environment of a given grid cell is optimal for all species occurring there. In spite of these limitations, barrier distance as calculated here is clearly better suited for modelling dispersal limitation than great circle distance. Our findings likely reflect the influence of strong large-scale barriers such as seaways (e.g. separation between N and S America, isolated islands of the Caribbean) and the Andes^{31–33}, but potentially also of less visible environmental barriers (e.g. the savannahs separating the Brazilian Atlantic Rainforest from the Amazon).

The geographic turnover of palm clades in the Americas is clearly shaped by β -niche evolution. Our finding that assemblages in different environments tended to be phylogenetically distinct, even when spatially close and not separated by measurable barriers, and that phylogenetic distinctness increases with environmental dissimilarity indicates that niche evolution is overall slow compared to the tempo of lineage diversification (Table 1: M_4). Effects of limited niche evolution (sometimes called “phylogenetic niche conservatism”, but the definition of this term is controversial^{34,35}) on clade distributions have been demonstrated previously^{36,37}. There is also evidence showing that limited niche evolution may, via its effect on clade distributions, have shaped patterns of community structure⁶ or diversity^{4,38} in terrapins, salamanders, and mammals. Our results supported the hypothesis *iii* that evolution of the climate niche, in particular related to temperature, had the strongest effect in American palms (Fig. 3, Table 2). It is well known that temperature is important in limiting species distributions³⁹; this was previously documented in palms⁴⁰. Our results are consistent with the fact that palms are inherently ill-suited for tolerating extreme temperatures^{9,41,42}. Of note, while water availability probably limits the geographic ranges of American palms^{14,43}, the evolution of those range limits does not seem to be a limiting factor for clade distributions (Fig. 3). Although scientific evidence has been lacking, the distributions of American palm species are thought to be influenced by edaphic conditions^{14,19}. This was confirmed by our results; furthermore, edaphic range limits appeared to evolve slowly compared to the speed of clade diversification, although the effect was much weaker than for temperature (Fig. 3, Table 2). Slow evolution (“conservatism”) of edaphic niches has also been shown for other plant groups and regions⁴⁴. In contrast, vegetation cover and structure either do not limit American palm ranges, or these constraints are phylogenetically labile (Fig. 3). Vegetation structure characteristics measured by remote sensing have been successfully used to model species distributions and diversity⁴⁵, but probably influence palm distributions primarily at local scales¹⁹.

Our results tentatively suggest that dispersal limitation has an equal or higher influence on palm clade turnover than niche evolution, rejecting our hypothesis *ii*. Even if the variation in phylogenetic turnover that could be due to either of the factors (R_{mx}) is entirely attributed to niche evolution, the influence of both factors would be about equal (Fig. 3). The dominance of dispersal limitation is also supported by the observation that $R_{pd} > R_{pe}$ except for the two models with highest R_{pe} (Supplementary Table S2). However, comparisons of the magnitude of R_{pe} and R_{pd} should be interpreted with caution since these fractions were affected by different error sources. The magnitude of R_{pe} is sensitive to the inclusion of relevant environmental predictors and their quality; the magnitude of R_{pd} depends on how well we were able to quantify the presence and strength of dispersal barriers between locations. Our results concerning the relative importance of dispersal and niche evolution rest on the assumption that these two error sources were of similar size. Of note, dispersal limitation on evolutionary timescales often requires some degree of limited niche evolution, since barriers are unsuitable areas

as defined by the niche²⁷. If niches were highly evolvable, barriers could easily be crossed by adaptation to the environment *on* the barrier. Any signals of dispersal limitation, such as correlations between phylogenetic turnover and spatial/barrier distance, are thus also evidence of limited niche evolution. However, this effect — mediated by dispersal — is different from the direct niche evolution effect (Table 1 M_4).

In general, we consider all our estimates conservative. Uncertainty was introduced at various points, including the distribution maps, the interpolated environmental predictors, the molecular dating procedure, and the artificial species level resolution. Relevant environmental predictors could also be missing⁴⁶, although the set of included variables was extensive. Neither spatial distance nor our measure of “barrier distance” are perfect proxies of dispersal limitation. Any further improvement in the data sources would likely increase the amount of variation explained by dispersal limitation or niche evolution. When it becomes possible to model biotic interactions, the influence of other species on the distribution patterns and niche estimates can be added to the equation.

Besides the effects outlined above, the phylogenetic structure of assemblages may be influenced by other factors, notably speciation patterns^{21,24} (Table 1). At regional scales, repeated allopatric speciation across a barrier might lead to a *negative* correlation between phylogenetic turnover and spatial or barrier distance⁴⁷. This pattern was found in a study on phylogenetic turnover among hummingbird communities in Ecuador (ca. 700 km in extent)²⁴. Since allopatric speciation is the outcome of dispersal limitation, the effect of dispersal on the relationship between phylogenetic turnover and spatial/barrier distance appears to reverse in direction between regional and continental scales. However, the effect of allopatric speciation on phylogenetic turnover should also be relatively ephemeral⁴⁸, as soon as sister lineages on each side of the barrier start to diversify into clades, phylogenetic turnover becomes positively related to barrier distance. Cases of relatively recent allopatric speciation are documented in American palms³¹ but clearly do not dominate our results. This might partly be due to the lack of terminal (intra-generic) phylogenetic resolution and/or our choice of phylogenetic turnover metric, which is known to emphasize ‘basal’ phylogenetic relationships⁴⁹. However, given the large extent of the study and the long presence of palms in the Americas, it is likely that any signal of allopatric speciation would be weak compared to the diversification of clades within regions that are delineated by barriers¹⁸. Still, there might be some effect of allopatric speciation that decreases the correlation between phylogenetic turnover and spatial/barrier distance. A similar logic applies to adaptive speciation: if sister species segregate along spatially structured environmental gradients⁵⁰ phylogenetic turnover might actually decrease with environmental dissimilarity (M_2 in Table 1). Again, this is most likely to occur at spatial scales much smaller than the scale of our study. In any case, our interpretations are conservative: if allopatric or adaptive speciation influenced phylogenetic turnover in our study system, it would bias the slopes and fractions of variation presented here towards zero and insignificance.

Our findings allow some thoughts on the future of American palm diversity under climate change. If niches evolve slowly even on evolutionary timescales species will not readily adapt to rapidly changing environments. Climate-related genetic variation within species appears to be widespread⁵¹, and rapid adaptation to climate change occurs⁵². Adaptation at the population level through selection acting on existing genotypes may be rapid, but adaptation to truly new environments is limited by the supply of new genotypes through mutation⁵¹. The former is conceivable even for groups showing strong phylogenetic niche conservatism, but only until the climatic changes extend beyond the range of conditions that is covered by existing intra-specific niche variation. At the end of the day, adaptation to

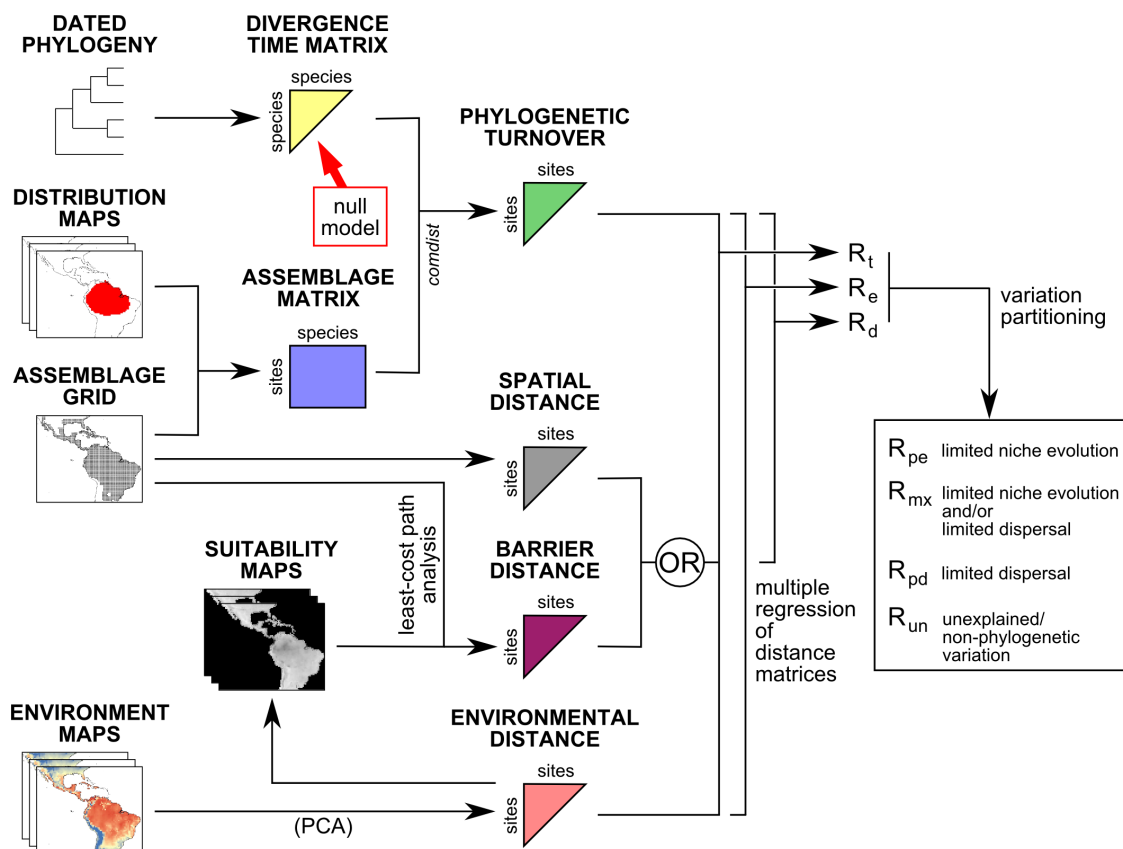


Figure 4 | Workflow of the analyses carried out for this study. Each individual step is described in the text (Methods section). R_c : variation in phylogenetic turnover explained by environmental dissimilarity and spatial/barrier distance, R_d : explained by spatial/barrier distance, R_t : explained by environmental dissimilarity and spatial/barrier distance. R_{pe} : explained by environmental dissimilarity *alone*, R_{pd} : explained by spatial/barrier distance *alone*, R_{mx} : explained by either environmental dissimilarity or spatial/barrier distance, R_{un} : unexplained. PCA = Principal Components Analysis.

environments that are not already part of the species' niche may be the key to surviving dramatic, rapid climate change. Our data suggested that this type of adaptation might be limited in American palms; however, more studies are required to improve our understanding of the adaptive capacities within this charismatic, ecologically and economically important plant group towards a changing world.

In conclusion, we found that the turnover of clades within American palms is controlled by a combination of two factors: 1) tolerances to temperature extremes and seasonality, and soil preferences, evolve slowly compared to the tempo of lineage diversification and 2) dispersal is limited on evolutionary timescales, definitely by barriers and possibly by time. The results corroborate that phylogenetic history is pivotal for understanding large-scale diversity patterns, especially since the turnover of clades can have important downstream effects on patterns of species richness and functional diversity. Future studies on other groups of organisms should also take into account that diversity patterns may be strongly shaped by restricted lineage mobility during diversification. Of note, the balance of dispersal limitation and phylogenetic niche conservatism, as well as the relevant niche factors, might be very different in other taxa due to ecological and life history differences, and in other regions due to differences in geomorphology, environment, and history. We found that 'barrier distance', our measure of dispersal barriers between locations, is a more relevant predictor of phylogenetic turnover than spatial distance. This measure should be similarly useful for studies on species richness and functional diversity. Finally, because their dispersal and niche evolution are limited on evolutionary timescales, palms might fail to migrate or adapt in response to rapidly changing climates. This

would have serious consequences for tropical biodiversity and for cultures that depend on palms as a resource^{53,54}.

Methods

An overview of the analyses performed for this article is shown in Fig. 4.

Distribution data. We used distribution maps of all American palms¹⁸, excluding three that are endemic to remote islands with no other palms (*Brahea edulis*, *Juania australis*, *Sabal bermudana*). This resulted in a total of 547 species. Due to taxonomic developments, close to 800 species of palms are currently recognized for the Americas¹⁷. However, there are no comparable distribution data that match the more recent taxonomy, and we are not aware of any bias in taxonomic changes since 1995 that might influence our results. We registered presence or absence of the included 547 species within $1^\circ \times 1^\circ$ grid cells ($n = 1701$ cells with palms, Fig. 1a). We preferred this grid to an equal area grid because it has previously been used in analyses of palm species richness^{14,55}, turnover⁵⁶, and net diversification⁵⁷. Thus, we accepted small area differences among grid cells for the sake of comparability. The richness per grid cell ranged from 1 to 83 species (Fig. 1a)¹⁴. These cells were used as assemblages in the analysis of phylogenetic turnover.

Phylogenetic tree. We modified a recently published dated version⁵⁸ of a genus level phylogenetic tree of all palms⁵⁹ to fit our assemblage dataset. We deleted genera that did not occur in the Americas and replaced terminal branches that represented genera with > 1 species with polytomies that included all species in our dataset. The crown age of those polytomies had to be chosen arbitrarily; to test the influence of this parameter on our results we produced four versions: crown ages equalling 10%, 50%, and 90% of the genus stem age, and crown ages computed with a simulation approach⁶⁰. The variation caused by those different assumptions was orders of magnitude smaller than the values of interest, and did not influence the qualitative outcome of our study. Thus, we present only the results for genus crown ages set to 50% of the stem age. Other related measures of phylogenetic community structure have been found equally insensitive to this parameter²⁰.

Phylogenetic turnover. For all pairwise combinations of grid cells, the *comdist* metric⁴⁸ was calculated in R (<http://www.r-project.org/>). We modified the *comdist*



function of the package *picante*⁶¹ to improve computational performance (see Supplementary Methods online). *Comdist* is defined as:

$$comdist_{AB} = \frac{1}{AB} \sum_{i=1}^s \sum_{j=1}^s d_{ij} p_{iA} p_{jB}, \quad (1)$$

where p_{iA} is the presence (= 1) or absence (= 0) of species i in assemblage A ; d_{ij} is the phylogenetic distance between species i and j ; A is the species richness of assemblage A . In our case, d_{ij} was proportional to pairwise divergence time; therefore, *comdist* represented the time in the past when the members of assemblage A diverged, on average, from the members of assemblage B .

Environmental variables. We extracted environmental information for each grid cell from Geographic Information System (GIS) layers with ArcInfo 9.3 (ESRI Inc., Redlands, California, USA). We obtained a total of 19 climatic variables, 6 soil variables, 2 topographic variables, and 4 measures of vegetation structure (see Supplementary Table S4 for information on the sources). All variables were averaged for each grid cell, except elevation, where a range was calculated to reflect topographic heterogeneity. All variables were standardised and centred.

Environmental dissimilarity. First, we computed a measure of environmental dissimilarity where all variables were included on an equal basis. We subjected the dataset to a principal components analysis (PCA) to remove multicollinearity between variables; all components with standard deviations < 10% of the standard deviation of the first component were omitted. Environmental dissimilarity was then calculated as the Euclidean distance in the space spanned by the PCA axes. Second, we repeated this procedure for five separate sets of variables, including temperature (bioclimatic variables 1–11), precipitation (bioclimatic variables 12–19), soil, topography, and vegetation structure (QSCAT and NDMI). Third, we computed a separate dissimilarity matrix for each environmental factor, with Euclidean distances taken directly from the standardised variables. This resulted in a total of 37 matrices of environmental dissimilarity for all pairwise combinations of grid cells.

Dispersal limitation. We computed two different measures to represent the degree to which dispersal among grid cells was limited. First, we calculated geodesic distance between grid cell midpoints of all pair-wise combinations ('spatial distance'). Second, we calculated a 'barrier distance' that takes the environmental insuitability of interjacent areas (i.e. dispersal barriers *sensu* ref. 27) into account. For each grid cell, a GIS layer was calculated for the entire study area representing how different the environment is from the environment of the focal grid cell ('cost surface'). For each pair of grid cells, environment-based dispersal distance was calculated as the average length of the least-cost path between them, based on their respective cost surfaces. Details on the calculation can be found in the Supplementary Methods online.

Analyses. We used ordinary least squares linear regression models to determine whether environmental dissimilarity (d_e) and spatial/barrier distance (d_d) could explain the variation in phylogenetic turnover (d_ϕ). As d_d we used either spatial distance or barrier distance. Three models were built, with $d_\phi \sim d_e$, $d_\phi \sim d_d$, and $d_\phi \sim d_e + d_d$. All variables were centred and standardised to obtain comparable effect sizes. We used variation partitioning⁶² to divide the variance of phylogenetic turnover into four fractions, according to whether they could be explained by environmental dissimilarity alone (R_{pe}), spatial/barrier distance alone (R_{pd}), environmental dissimilarity and dispersal distance (R_{mx}), or unexplained (R_{un}). These fractions were computed from the R^2 values of the $d_\phi \sim d_e$ model (R_e), the $d_\phi \sim d_d$ model (R_d), and the multiple predictor model (R_t) as follows: $R_{pe} = R_t - R_d$; $R_{pd} = R_t - R_e$; $R_{mx} = R_e + R_d - R_t$; and $R_{un} = 1 - R_t$.

Significance testing. Since pairwise measures of dissimilarity are not statistically independent, we used a permutation procedure to assess significances. This allowed us to account simultaneously for a second problem, namely that phylogenetic turnover between assemblages is confounded by the number of shared species (i.e. species turnover) as well as differences in species richness. For the purpose of our study, we were only interested in the turnover of higher-level lineages, not species. We therefore employed a null model where we re-shuffled the species labels on the phylogeny 999 times and re-calculated phylogenetic turnover from these randomised phylogenies. For each of these null matrices, we performed the regression analysis and variation partitioning described above. In these null analyses, *comdist* still correlated with environmental dissimilarity and dispersal distance due to shared species and richness differences. The proportion of times that the R_{pe} and R_{pd} fractions, respectively, of the null analyses were higher than the R_{pe}/R_{pd} fractions of the real data was used as the probability of the hypothesis H_0 that niche evolution/dispersal limitation did not affect phylogenetic turnover. This procedure has been previously used to test for relationships between phylogenetic turnover and altitudinal difference independent of species turnover²³.

- Lomolino, M. V., Riddle, B. R., Whittaker, R. J. & Brown, J. H. *Biogeography*. 4 edn, (Sinauer Associates, 2010).
- Ackerly, D. D., Schwilk, D. W. & Webb, C. O. Niche evolution and adaptive radiation: Testing the order of trait divergence. *Ecology* **87**, S50–S61 (2006).

- Pennington, R. T., Lavin, M. & Oliveira, A. Woody plant diversity, evolution, and ecology in the tropics: Perspectives from seasonally dry tropical forests. *Annu. Rev. Ecol. Evol. S.* **40**, 437–457 (2009).
- Buckley, L. B. *et al.* Phylogeny, niche conservatism and the latitudinal diversity gradient in mammals. *Proc. Roy. Soc. B-Biol. Sci.* **277**, 2131–2138 (2010).
- Roncal, J., Blach-Overgaard, A., Borchsenius, F., Balslev, H. & Svenning, J. C. A dated phylogeny complements macroecological analysis to explain the diversity patterns in *Geonoma* (Arecaceae). *Biotropica* **43**, 324–334 (2011).
- Stephens, P. R. & Wiens, J. J. Bridging the gap between community ecology and historical biogeography: niche conservatism and community structure in emydid turtles. *Mol. Ecol.* **18**, 4664–4679 (2009).
- Wiens, J. J. The causes of species richness patterns across space, time, and clades and the role of "ecological limits". *Q. Rev. Biol.* **86**, 75–96 (2011).
- Baselga, A., Lobo, J. M., Svenning, J.-C., Aragón, P. & Araújo, M. B. Dispersal ability modulates the strength of the latitudinal richness gradient in European beetles. *Global Ecol. Biogeogr.* **21**, 1106–1113 (2012).
- Wiens, J. J. & Donoghue, M. J. Historical biogeography, ecology and species richness. *Trends Ecol. Evol.* **19**, 639–644 (2004).
- Antonelli, A. & Sanmartin, I. Why are there so many plant species in the Neotropics? *Taxon* **60**, 403–414 (2011).
- Hoorn, C. *et al.* Amazonia Through Time: Andean Uplift, Climate Change, Landscape Evolution, and Biodiversity. *Science* **330**, 927–931 (2010).
- Pennington, R. T., Richardson, J. E. & Lavin, M. Insights into the historical construction of species-rich biomes from dated plant phylogenies, neutral ecological theory and phylogenetic community structure. *New Phytol.* **172**, 605–616 (2006).
- Rull, V. Neotropical biodiversity: timing and potential drivers. *Trends Ecol. Evol.* **26**, 508–513 (2011).
- Bjorholm, S., Svenning, J. C., Baker, W. J., Skov, F. & Balslev, H. Historical legacies in the geographical diversity patterns of New World palm (Arecaceae) subfamilies. *Bot. J. Linn. Soc.* **151**, 113–125 (2006).
- Webb, S. D. The Great American Biotic Interchange: Patterns and processes. *Ann. Mo. Bot. Gard.* **93**, 245–257 (2006).
- Kissling, W. D. *et al.* Quaternary and pre-Quaternary historical legacies in the global distribution of a major tropical plant lineage. *Global Ecol. Biogeogr.* **21**, 909–921 (2012).
- Balslev, H. *et al.* Species diversity and growth forms in tropical American palm communities. *Bot. Rev.* **77**, 381–425 (2011).
- Henderson, A., Galeano, G. & Bernal, R. *Field guide to the palms of the Americas*. (Princeton University Press, 1995).
- Eiserhardt, W. L., Svenning, J.-C., Kissling, D. & Balslev, H. Geographical ecology of the palms (Arecaceae): determinants of diversity and distributions across spatial scales. *Ann. Bot.* **108**, 1391–1416 (2011).
- Kissling, W. D. *et al.* Cenozoic imprints on the phylogenetic structure of palm species assemblages worldwide. *P. Natl. Acad. Sci. USA* **109**, 7379–7384 (2012).
- Graham, C. H. & Fine, P. V. A. Phylogenetic beta diversity: linking ecological and evolutionary processes across space in time. *Ecol. Lett.* **11**, 1265–1277 (2008).
- Tuomisto, H. A diversity of beta diversities: straightening up a concept gone awry. Part I. Defining beta diversity as a function of alpha and gamma diversity. *Ecography* **33**, 2–22 (2010).
- Bryant, J. A. *et al.* Microbes on mountainsides: Contrasting elevational patterns of bacterial and plant diversity. *P. Natl. Acad. Sci. USA* **105**, 11505–11511 (2008).
- Graham, C. H., Parra, J. L., Rahbek, C. & McGuire, J. A. Phylogenetic structure in tropical hummingbird communities. *P. Natl. Acad. Sci. USA* **106**, 19673–19678 (2009).
- Parmentier, I. & Hardy, O. J. The impact of ecological differentiation and dispersal limitation on species turnover and phylogenetic structure of inselberg's plant communities. *Ecography* **32**, 613–622 (2009).
- Baker, W. J. & Couvreur, T. L. P. Global biogeography and diversification of palms sheds light on the evolution of tropical lineages. II. diversification history and origin of regional assemblages. *J. Biogeogr.* **40**, 286–298 (2013).
- Wiens, J. J. & Graham, C. H. Niche conservatism: Integrating evolution, ecology, and conservation biology. *Annu. Rev. Ecol. Evol. S.* **36**, 519–539 (2005).
- Nekola, J. C. & White, P. S. The distance decay of similarity in biogeography and ecology. *J. Biogeogr.* **26**, 867–878 (1999).
- Svenning, J.-C. & Skov, F. Could the tree diversity pattern in Europe be generated by postglacial dispersal limitation? *Ecol. Lett.* **10**, 453–460 (2007).
- Paul, J. R., Morton, C., Taylor, C. M. & Tonsor, S. J. Evolutionary time for dispersal limits the extent but not the occupancy of species' potential ranges in the tropical plant genus *Psychotria* (Rubiaceae). *Am. Nat.* **173**, 188–199 (2009).
- Barfod, A. S., Trelen, P. & Borchsenius, F. In *Diversity, Phylogeny, and Evolution of the Monocotyledons*. (eds O. Seberg, G. Petersen, A. S. Barfod, & J. I. Davis) 225–243 (Aarhus University Press, 2010).
- Cuenca, A., Asmussen-Lange, C. B. & Borchsenius, F. A dated phylogeny of the palm tribe Chamaedoreae supports Eocene dispersal between Africa, North and South America. *Mol. Phylogenet. Evol.* **46**, 760–775 (2008).
- Roncal, J., Borchsenius, F., Asmussen-Lange, C. B. & Balslev, H. In *Diversity, phylogeny, and evolution of the monocotyledons*. (eds O. Seberg, G. Petersen, A. S. Barfod, & J. I. Davis) 245–265 (Aarhus University Press, 2010).
- Losos, J. B. Seeing the forest for the trees: The limitations of phylogenies in comparative biology. *Am. Nat.* **177**, 709–727 (2011).



35. Wiens, J. J. *et al.* Niche conservatism as an emerging principle in ecology and conservation biology. *Ecol. Lett.* **13**, 1310–1324 (2010).
36. Peterson, A. T., Soberon, J. & Sanchez-Cordero, V. Conservatism of ecological niches in evolutionary time. *Science* **285**, 1265–1267 (1999).
37. Kellermann, V. *et al.* Upper thermal limits of *Drosophila* are linked to species distributions and strongly constrained phylogenetically. *P. Natl. Acad. Sci. USA*, early online, doi: 10.1073/pnas.1207553109.
38. Kozak, K. H. & Wiens, J. J. Niche conservatism drives elevational diversity patterns in Appalachian salamanders. *Am. Nat.* **176**, 40–54 (2010).
39. Woodward, F. I., Fogg, G. E. & Heber, U. The impact of low temperatures in controlling the geographical distribution of plants [and discussion]. *Philos. T. R. Soc. B* **326**, 585–593 (1990).
40. Walther, G. R. *et al.* Palms tracking climate change. *Global Ecol. Biogeogr.* **16**, 801–809 (2007).
41. Salm, R., Salles, N. V. D., Alonso, W. J. & Schuck-Paim, C. Cross-scale determinants of palm species distribution. *Acta Amazonica* **37**, 17–25 (2007).
42. Tomlinson, P. B. The uniqueness of palms. *Bot. J. Linn. Soc.* **151**, 5–14 (2006).
43. Blach-Overgaaard, A., Svenning, J. C., Dransfield, J., Greve, M. & Balslev, H. Determinants of palm species distributions across Africa: the relative roles of climate, non-climatic environmental factors, and spatial constraints. *Ecography* **33**, 380–391 (2010).
44. Prinzing, A., Durka, W., Klotz, S. & Brandl, R. The niche of higher plants: evidence for phylogenetic conservatism. *Proc. Roy. Soc. B-Biol. Sci.* **268**, 2383–2389 (2001).
45. Buermann, W. *et al.* Predicting species distributions across the Amazonian and Andean regions using remote sensing data. *J. Biogeogr.* **35**, 1160–1176 (2008).
46. Tuomisto, H. & Ruokolainen, K. Analyzing or explaining beta diversity? Reply. *Ecology* **89**, 3244–3256 (2008).
47. Hardy, O. J. & Senterre, B. Characterizing the phylogenetic structure of communities by an additive partitioning of phylogenetic diversity. *J. Ecol.* **95**, 493–506 (2007).
48. Webb, C. O., Cannon, C. H. & Davies, R. G. in *Tropical forest community ecology*. (eds W. P. Carson & S. A. Schnitzer) 79–97 (Wiley & Sons, 2008).
49. Swenson, N. G. Phylogenetic beta diversity metrics, trait evolution and inferring the functional beta diversity of communities. *PLoS ONE* **6**, e21264 (2011).
50. Fine, P. V. A., Daly, D. C., Munoz, G. V., Mesones, I. & Cameron, K. M. The contribution of edaphic heterogeneity to the evolution and diversity of Burseraceae trees in the western Amazon. *Evolution* **59**, 1464–1478 (2005).
51. Jump, A. S. & Peñuelas, J. Running to stand still: adaptation and the response of plants to rapid climate change. *Ecol. Lett.* **8**, 1010–1020 (2005).
52. Franks, S. J., Sim, S. & Weis, A. E. Rapid evolution of flowering time by an annual plant in response to a climate fluctuation. *P. Natl. Acad. Sci. USA* **104**, 1278–1282 (2007).
53. Balick, M. J. & Beck, H. T. *Useful palms of the world: a synoptic bibliography* (Columbia University Press, New York, 1990).
54. Balslev, H. Palm harvest impacts in north-western South America. *Bot. Rev.* **77**, 370–380 (2011).
55. Eiserhardt, W. L., BJORHOLM, S., Svenning, J. C., Rangel, T. F. & Balslev, H. Testing the water-energy theory on American palms (Arecaceae) using geographically weighted regression. *PLoS ONE* **6**, e27027 (2011).
56. BJORHOLM, S., Svenning, J.-C., Skov, F. & Balslev, H. To what extent does Tobler's 1st law of geography apply to macroecology? A case study using American palms (Arecaceae). *BMC Ecology* **8**, 11 (2008).
57. Svenning, J. C., Borchsenius, F., BJORHOLM, S. & Balslev, H. High tropical net diversification drives the New World latitudinal gradient in palm (Arecaceae) species richness. *J. Biogeogr.* **35**, 394–406 (2008).
58. Couvreur, T. L. P., Forest, F. & Baker, W. J. Origins and global diversification patterns of tropical rain forests: inferences from a complete genus-level phylogeny of palms. *BMC Biol.* **9**, 44 (2011).
59. Baker, W. J. *et al.* Complete generic-level phylogenetic analyses of palms (Arecaceae) with comparisons of supertree and supermatrix approaches. *Syst. Biol.* **58**, 240–256 (2009).
60. Eiserhardt, W. L., Svenning, J.-C., Borchsenius, F., Kristiansen, T. & Balslev, H. Separating environmental and geographical determinants of phylogenetic community structure in Amazonian palms (Arecaceae). *Bot. J. Linn. Soc.* **171**, 244–259 (2013).
61. Kembel, S. W. *et al.* Picante: R tools for integrating phylogenies and ecology. *Bioinformatics* **26**, 1463–1464 (2010).
62. Legendre, P. & Legendre, P. *Numerical Ecology*. 2nd. Edition edn (Elsevier, 1998).

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Author contributions

W.L.E., J.-C.S. and H.B. developed the idea. W.J.B. and T.L.P.C. constructed the dated phylogeny. W.L.E. performed the analyses and led the writing. All authors contributed substantially to revisions.

Additional information

Supplementary information accompanies this paper at <http://www.nature.com/scientificreports>

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