EDITORIAL

Taylor & Francis Taylor & Francis Group

OPEN ACCESS OPEN ACCESS

Antibody-based intervention against the pore-forming toxins of *Staphylococcus aureus*

Isaac P. Thomsen 🕩

Department of Pediatrics, Division of Pediatric Infectious Diseases and the Vanderbilt Vaccine Research Program, Vanderbilt University Medical Center, Nashville, Tennessee, USA

ARTICLE HISTORY Received 30 November 2017; Accepted 6 December 2017 **KEYWORDS** antibody; leukocidin; *Staphylococcus aureus*; toxin

Staphylococcus aureus is nearly unrivaled among bacterial pathogens, both in the diversity of clinical syndromes it causes (from skin abscesses and food-borne illness to endocarditis and sepsis) and in its complex array of virulence mechanisms. In addition, antimicrobial resistance rates continue to increase, both in methicillin-susceptible (MSSA) and methicillin-resistant (MRSA) isolates [1]. *S. aureus* harbors numerous gene regulatory and quorum sensing systems, and features substantial genomic plasticity and frequent redundancy among specific virulence factors. For these reasons, this pathogen represents a daunting target for the development of novel therapeutic and preventive measures, dating back to the first published attempt at a *S. aureus* vaccine in 1902 [2].

Since the onset of the community-associated MRSA epidemic, a family of toxins produced by S. aureus, the leukocidins, have gained increasing attention as important virulence determinants and potential targets of intervention against this pathogen. The leukocidins known to be produced by clinical isolates of S. aureus include Panton-Valentine leukocidin (PVL), LukAB (also known as LukGH), LukED, and the γ -hemolysins HlgAB and HlgCB [3]. Each of these two-component toxins is secreted as a pair of monomers that oligomerize to form a pore on the surface of phagocytes, lymphocytes, and natural killer cells, and they are important mediators of staphylococcal evasion of innate host defenses. The neutrophil represents the primary innate defense against S. aureus infection in humans, as evidenced in part by the clear predilection toward invasive S. aureus disease in patients with neutrophil defects [4]. The leukocidins exert their effect at the level of the neutrophil and other phagocytes, binding receptors in the chemokine and complement receptor families [5-8], forming a pore, and potently lysing these cells, thereby

facilitating infection in a variety of *in vivo* models [9–13]. *S. aureus* elaborates a number of additional poreforming toxins outside the leukocidin family, prominently including alpha-hemolysin (Hla), which primarily targets erythrocytes, epithelial and endothelial cells, and lymphocytes. While the role of Hla has been carefully elucidated in numerous animal models [14,15], most of the leukocidins exhibit a markedly increased tropism for human leukocytes in comparison to murine cells [6,16,17], likely resulting in a previous underappreciation of the importance of the leukocidins when extrapolating from murine data.

Since its discovery by two independent groups in 2010 [12,18], LukAB/LukGH has garnered attention as an important *S. aureus* virulence factor based on its clear role in both *ex vivo* and *in vivo* models of disease [6,12,13,19]. Infection of human neutrophils with diverse *S. aureus* strains indicates that LukAB/LukGH is the dominant toxin responsible for neutrophil targeting and killing [12]. This toxin is also highly conserved, being present in the genome of all known clinical isolates tested to date [20,21]. Finally, LukAB/LukGH is clearly produced during human infection, as evidenced by its recognition by the humoral response following invasive human disease [21,22].

In this issue of *Virulence*, Rouha *et al* have thoroughly evaluated the capacity of a pair of human monoclonal antibodies to inhibit the cytotoxicity of the leukocidins and Hla [23]. These antibodies, termed ASN-1 and ASN-2, were isolated by screening a human IgG1 antibody library using a yeast selection system; ASN-1 exhibits cross-reactive neutralizing activity against Hla and four of the leukocidins (PVL, LukED, and the γ -hemolysins), while ASN-2 neutralizes LukAB/LukGH. The authors have previously reported the cross-reactive

© 2018 The Author. Published by Informa UK Limited, trading as Taylor & Francis Group

This is an Open Access article distributed under the terms of the Creative Commons Attribution License (http://creativecommons.org/licenses/by/4.0/), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

CONTACT Isaac P. Thomsen isaac.thomsen@vanderbilt.edu Pediatric Infectious Diseases, D-7235 MCN 1161 21st Avenue South Nashville, Nashville, Tennessee 37232-2581, USA

Comment On: Rouha Haralld ,et al. Disarming Staphylococcus aureus from destroying human cells by simultaneously neutralizing six cytotoxins with two human monoclonal antibodies.VIRULENCE, 2018 VOL. 9, NO. 1, 231–247 https://doi.org/10.1080/21505594.2017.1391447

capacity of ASN-1 [24], itself an important discovery given the redundant nature of *S. aureus* virulence factor expression. In this report, Rouha and colleagues characterize the individual and combined effects of the mAbs in a variety of *in vitro* models using human leukocytes, an important distinction given leukocidin tropism.

Several notable findings emerge from this work. First, the authors observed marked differences in toxin production in the presence of different culture media, particularly for the leukocidins. This speaks to the difficulty of interpreting the importance of staphylococcal toxins (and many other virulence factors) from different in vitro models, as protein expression by S. aureus may vary dramatically based on factors such as pH, oxygen tension, and nutrient availability [13,25,26]. Of note, the authors found that LukAB/LukGH was the dominant toxin in the media that may best recapitulate the host environment in the setting of human infection, RPMI + Casamino acids. Second, the authors observed that toxin production also varied widely across S. aureus strains. As the pore-forming toxins are evaluated as putative targets of intervention against S. aureus, it will be important to define the critical toxin(s) in the setting of human infection and produced by widely circulating clinical isolates. The authors also found that both antibodies were required to fully prevent toxin-mediated lysis of human neutrophils in most in vitro conditions, emphasizing the apparent redundancy in this pathway, though caution must be used when extrapolating these in vitro findings to what occurs in the human host during natural infection.

Many fascinating questions remain unanswered regarding pore-forming toxin biology and the interaction between these important S. aureus virulence factors and the human host. Despite the robust in vitro characterization of antibody-mediated pore-forming toxin inhibition reported by Rouha and colleagues[23], gaps remain in our understanding of antibody-toxin interactions in the setting of serious human infections, the setting in which a putative therapeutic would be deployed. For example, our group recently reported that different human antibodies (purified from Bcells obtained from children with invasive S. *aureus* disease) neutralize LukAB/LukGH-mediated cytotoxicity by distinct mechanisms [22]. It remains unclear whether certain of these mechanisms are more biologically relevant or important in the setting of invasive human infection. Further, evidence of antibody-enhanced disease has been reported in a murine model for at least one of the leukocidins (PVL) [27], and the relevance of this in humans (both for PVL and the other leukocidins) remains largely unexplored. The authors note that the antibody combination reported in this manuscript is under investigation in a Phase II clinical trial involving mechanically ventilated patients heavily colonized with S. aureus. The findings of this and other future work will hopefully provide further insights into the potential roles of antibody-mediated neutralization of poreforming toxins in the setting of human disease.

Disclosures / conflicts of interest

The author is supported by National Institute of Allergy and Infectious Diseases (NIAID) grant 1K23AII13150 K23. The author serves as an investigator on studies funded by GlaxoSmithKline and Horizon Pharma. None of these studies conflict with the contents of this manuscript.

Funding

National Institute of Allergy and Infectious Diseases (1K23AI113150).

ORCID

Isaac P. Thomsen (b) http://orcid.org/0000-0001-9240-0164

References

- Sutter DE, Milburn E, Chukwuma U, et al. Changing Susceptibility of Staphylococcus aureus in a US pediatric population. Pediatrics. 2016;137(4). doi: 10.1542/ peds.2015-3099. PMID: 26933211.
- [2] Wright AE. Notes on the treatment of furunculosis, sycosis, and acne by the inoculation of a staphylococcus aureus vaccine. Lancet. 1902;159(4100):874-884.
- [3] Spaan AN, van Strijp JAG, Torres VJ. Leukocidins: staphylococcal bi-component pore-forming toxins find their receptors. Nat Rev Microbiol. 2017;15(7):435–447. doi: 10.1038/nrmicro.2017.27. PMID: 28420883.
- [4] Rigby KM, DeLeo FR. Neutrophils in innate host defense against Staphylococcus aureus infections. Semin Immunopathol. 2012;34(2):237–59. doi: 10.1007/s00281-011-0295-3. PMID: 22080185.
- [5] Alonzo F, 3rd, Kozhaya L, Rawlings SA, et al. CCR5 is a receptor for Staphylococcus aureus leukotoxin ED. Nature. 2013;493(7430):51–5. doi: 10.1038/nature11724. PMID: 23235831.
- [6] DuMont AL, Yoong P, Day CJ, et al. Staphylococcus aureus LukAB cytotoxin kills human neutrophils by targeting the CD11b subunit of the integrin Mac-1. Proc Natl Acad Sci U S A. 2013;110(26):10794–9. doi: 10.1073/pnas.1305121110. PMID: 23754403.
- [7] Spaan AN, Henry T, van Rooijen WJ, et al. The staphylococcal toxin Panton-Valentine Leukocidin targets human C5a receptors. Cell Host Microbe. 2013;13(5):584–94. doi: 10.1016/j.chom.2013.04.006. PMID: 23684309.
- [8] Spaan AN, Vrieling M, Wallet P, et al. The staphylococcal toxins gamma-haemolysin AB and CB differentially target phagocytes by employing specific chemokine receptors. Nat Commun. 2014;5:5438. doi: 10.1038/ ncomms6438. PMID: 25384670.
- [9] Cremieux AC, Dumitrescu O, Lina G, et al. Panton-valentine leukocidin enhances the severity of communityassociated methicillin-resistant Staphylococcus aureus

rabbit osteomyelitis. PloS One. 2009;4(9):e7204. doi: 10.1371/journal.pone.0007204. PMID: 19779608.

- [10] Diep BA, Chan L, Tattevin P, et al. Polymorphonuclear leukocytes mediate Staphylococcus aureus Panton-Valentine leukocidin-induced lung inflammation and injury. Proc Natl Acad Sci U S A. 2010;107(12):5587–92. doi: 10.1073/pnas.0912403107. PMID: 20231457.
- [11] Alonzo F, 3rd, Benson MA, Chen J, et al. Staphylococcus aureus leucocidin ED contributes to systemic infection by targeting neutrophils and promoting bacterial growth in vivo. Mol Microbiol. 2012;83(2):423–35. doi: 10.1111/ j.1365-2958.2011.07942.x. PMID: 22142035.
- [12] Dumont AL, Nygaard TK, Watkins RL, et al. Characterization of a new cytotoxin that contributes to Staphylococcus aureus pathogenesis. Mol Microbiol. 2011;79 (3):814–25. doi: 10.1111/j.1365-2958.2010.07490.x. PMID: 21255120.
- [13] DuMont AL, Yoong P, Surewaard BG, et al. Staphylococcus aureus elaborates leukocidin AB to mediate escape from within human neutrophils. Infect Immunity. 2013;81(5):1830-41. doi: 10.1128/IAI.00095-13. PMID: 23509138.
- [14] Inoshima I, Inoshima N, Wilke GA, et al. A Staphylococcus aureus pore-forming toxin subverts the activity of ADAM10 to cause lethal infection in mice. Nat Med. 2011;17 (10):1310–4. doi: 10.1038/nm.2451. PMID: 21926978.
- [15] Powers ME, Kim HK, Wang Y, et al. ADAM10 mediates vascular injury induced by Staphylococcus aureus alphahemolysin. J Infect Dis. 2012;206(3):352–6. doi: 10.1093/ infdis/jis192. PMID: 22474035.
- [16] Diep BA, Le VT, Visram ZC, et al. Improved protection in a rabbit model of community-associated methicillin-resistant staphylococcus aureus necrotizing pneumonia upon neutralization of leukocidins in addition to alpha-hemolysin. Antimicrob Agents Chemother. 2016;60(10):6333–40. doi: 10.1128/AAC.01213-16. PMID: 27527081.
- [17] Loffler B, Hussain M, Grundmeier M, et al. Staphylococcus aureus panton-valentine leukocidin is a very potent cytotoxic factor for human neutrophils. PLoS Pathog. 2010;6(1):e1000715. doi: 10.1371/journal.ppat.1000715. PMID: 20072612.
- [18] Ventura CL, Malachowa N, Hammer CH, et al. Identification of a novel Staphylococcus aureus two-component leukotoxin using cell surface proteomics. PloS One. 2010;5(7):e11634. doi: 10.1371/journal.pone.0011634. PMID: 20661294.

- [19] DuMont AL, Yoong P, Liu X, et al. Identification of a crucial residue required for Staphylococcus aureus LukAB cytotoxicity and receptor recognition. Infect Immunity. 2014;82(3):1268–76. doi: 10.1128/IAI.01444-13. PMID: 24379286.
- [20] Chadha AD, Thomsen IP, Jimenez-Truque N, et al. Host response to Staphylococcus aureus cytotoxins in children with cystic fibrosis. J Cyst Fibros. 2016;15 (5):597-604. doi: 10.1016/j.jcf.2015.12.023. PMID: 26821814.
- [21] Thomsen IP, Dumont AL, James DB, et al. Children with invasive Staphylococcus aureus disease exhibit a potently neutralizing antibody response to the cytotoxin LukAB. Infect Immunity. 2014;82(3):1234–42. doi: 10.1128/ IAI.01558-13. PMID: 24379282.
- [22] Thomsen IP, Sapparapu G, James DBA, et al. Monoclonal antibodies against the Staphylococcus aureus Bicomponent Leukotoxin AB isolated following invasive human infection reveal diverse binding and modes of action. J Infect Dis. 2017;215(7):1124–1131. doi: 10.1093/ infdis/jix071. PMID: 28186295.
- [23] Rouha H, Weber S, Janesch P, et al. Disarming Staphylococcus aureus from destroying human cells by simultaneously neutralizing six cytotoxins with two human monoclonal antibodies. Virulence. 2017.
- [24] Rouha H, Badarau A, Visram ZC, et al. Five birds, one stone: neutralization of alpha-hemolysin and 4 bi-component leukocidins of Staphylococcus aureus with a single human monoclonal antibody. MAbs. 2015;7(1):243–54. doi: 10.4161/19420862.2014.985132. PMID: 25523282.
- [25] Janesch P, Rouha H, Weber S, et al. Selective sensitization of human neutrophils to LukGH mediated cytotoxicity by Staphylococcus aureus and IL-8. J Infect. 2017;74 (5):473-483. doi: 10.1016/j.jinf.2017.02.004. PMID: 28237625.
- [26] Wilde AD, Snyder DJ, Putnam NE, et al. Bacterial hypoxic responses revealed as critical determinants of the host-pathogen outcome by TnSeq analysis of staphylococcus aureus invasive infection. PLoS Pathog. 2015;11 (12):e1005341. doi: 10.1371/journal.ppat.1005341. PMID: 26684646.
- [27] Yoong P, Pier GB. Antibody-mediated enhancement of community-acquired methicillin-resistant Staphylococcus aureus infection. Proc Natl Acad Sci U S A. 2010;107 (5):2241-6. doi: 10.1073/pnas.0910344107. PMID: 20133867.