

Research Article

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An ensemble method to predict target genes and pathways in uveal melanoma

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Abstract: Objective: This work proposes to predict target genes and pathways for uveal melanoma (UM) based on an ensemble method and pathway analyses. **Methods:** The ensemble method integrated a correlation method (Pearson correlation coefficient, PCC), a causal inference method (IDA) and a regression method (Lasso) utilizing the Borda count election method. Subsequently, to validate the performance of PIL method, comparisons between confirmed database and predicted miRNA targets were performed. Ultimately, pathway enrichment analysis was conducted on target genes in top 1000 miRNA-mRNA interactions to identify target pathways for UM patients. **Results:** Thirty eight of the predicted interactions were matched with the confirmed interactions, indicating that the ensemble method was a suitable and feasible approach to predict miRNA targets. We obtained 50 seed miRNA-mRNA interactions of UM patients and extracted target genes from these interactions, such as *ASPG*, *BSDC1* and *C4BP*. The 601 target genes in top 1,000 miRNA-mRNA interactions were enriched in 12 target pathways, of which Phototransduction was the most significant one. **Conclusion:** The target genes and pathways might provide a new way to reveal the molecular mechanism of UM and give hand for target treatments and preventions of this malignant tumor.

Keywords: uveal melanoma; miRNA; mRNA; target; gene; pathway

1 Introduction

Uveal melanoma (UM) is the most frequent and aggressive ocular primary tumor that arises from neural crest-derived melanocytes of the uveal tract of the eye in adults [1], with an incidence rate of up to 8 per 1,000,000 person years in Europe [2, 3]. The fatality rate of UM is high, since patients are at risk of developing metastases up to 20 years after the initial diagnosis, and 80% of metastatic patients die within one year and 92% within 2 years of the diagnosis of metastases [4, 5]. However, no effective adjuvant therapy is available to prevent metastases, neither is there any effective treatment once metastases have developed at present [3]. With the development of gene expression related analyses, target treatments could provide new insights for effective therapy to large extent and potentially improve patient survival [6]. Besides, understanding the molecular characteristics and mechanisms of UM is critical for the creation of a treatment for this tumor.

It has been demonstrated that intratumoral discordance in gene expression profile is associated with intratumoral heterogeneity based upon histopathologic features in UM [7]. Furthermore, several gene signatures underlying UM have been uncovered, such as $G\alpha_q$ stimulatory subunit *GNAQ* and *BAP1* [8, 9]. However, mutated genes do not play roles individually and similar genes often work together to complete certain biological functions. What's more, those correlated genes might be regulated by one microRNA (miRNA) whose signatures may be promising biomarkers for the classification or outcome prediction of large number of human cancers [10]. Therefore, investigating miRNAs offers an excellent way to elucidate the complex pathological mechanisms underlying malignant tumors, and gives a hand to the design of drugs for treatments.

In the present study, we proposed to predict targets of miRNAs in UM based on an ensemble method produced by Le et al. [11]. It could solve the inconsistent results problem resulting from individual methods by including complementary results [12]. Specifically, it merged a correlation method (Pearson correlation coefficient,

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PCC), a causal inference method (IDA) and a regression method (Lasso) utilizing the Borda count election method. Subsequently, the predicted miRNA targets were validated by matching them with the known confirmed databases. Ultimately, pathway enrichment analysis was conducted on target genes to identify target pathways for UM patients. The target genes and pathways might light a new lamp for revealing molecular mechanism of UM and give a hand for target treatments and preventions of this malignant tumor.

2 Materials and methods

2.1 Preparation of miRNA and mRNA data

MiRNA and mRNA expression data for UM patients were downloaded from the Cancer Genome Atlas (TCGA) (<http://cancergenome.nih.gov/>), respectively. Only 80 samples which were existed in both miRNA and mRNA expression data were reserved for the following analysis. Subsequently, the miRNAs or mRNAs with expression values = 0 were removed. Then the residual expression values were converted into log₂ forms and normalized using a Global Variance Stabilizing Normalization (VSN) method [13]. Consequently, 793 miRNAs and 19,511 mRNAs were obtained in the expression data. For purpose of making the data more confident and reliable, the PCC method was utilized to compute the correlations between miRNA and mRNA. If the absolute PCC value of a pair of miRNA and mRNA was more than 0.7, it would be remained. Finally, a total of 107 miRNAs and 904 mRNAs were obtained for subsequent analyses.

Ethical approval: The conducted research is not related to either human or animals use.

2.2 Prediction of miRNA targets

Using the miRNA and mRNA data, the ensemble method which integrated three methods (PCC, IDA and Lasso) based on Borda count election method, was applied to predict miRNA targets for UM. This process was comprised of three steps:

Firstly, the PCC, IDA and Lasso method was used to predict miRNA targets on the basis of miRNA and mRNA data, and then these miRNA targets were ranked, respectively. Only the top k ($k = 100$) ranked targets were left to perform the followed analysis. Secondly, Borda rank election method was employed to integrate top k

ranked genes from the merged list as the final output, i.e. the potential target genes for the given miRNA of UM.

2.3 Validations of predicted miRNA targets

To validate the feasibility and confidence of the predicted miRNA targets in UM patients, we compared our results with the union of four popular databases, miRTarbase v4.5 [15], Tarbase v6.0 [16], miRecords v2013 [17] and miRWalk v2.0 [18]. Briefly, miRTarbase provides the most current and comprehensive information of experimentally validated miRNA-mRNA target interactions [19]. While TarBase is the first resource to provide experimentally verified miRNA target interactions by surveying pertinent literature [20]. As for miRecords, it accumulates experimentally validated miRNA targets and computationally predicts miRNA targets [17]. Last but not least, miRWalk is an available comprehensive resource that hosts the predicted as well as experimentally validated miRNA target interaction pairs [18]. After removing the duplicated interactions, we could obtain a union of known interactions and referred them to confirmed interactions in the paper. If a miRNA target interaction was involved in confirmed interactions, we thought that the predicted miRNA target was validated.

2.4 Pathway enrichment analysis

In order to investigate biological functions of miRNA targets enriched in the top k miRNA-mRNA interactions, Kyoto Encyclopedia of Genes and Genomes (KEGG) pathway enrichment analysis was carried out based on the Database for Annotation, Visualization, and Integrated Discovery (DAVID, <https://david.ncifcrf.gov/>) tool [21]. Here, the KEGG database (<http://www.genome.jp/kegg/>) is a collection of manually drawn pathway maps for metabolism, genetic information processing, environmental information processing [22]. Besides, the Fisher's exact test was employed to identify significant pathways between UM patients and normal controls [23]. The threshold of significance was defined as $P < 0.01$

which were adjusted by false discovery rate (FDR) based on Benjamini & Hochberg method [24].

3 Results

3.1 Predicted miRNA targets

In the current study, a total of 107 miRNAs and 904 mRNAs of UM were prepared from the TCGA database for the subsequent analyses. Based on these expression data, miRNA targets were predicted by PCC, IDA and Lasso method respectively, and the top 100 targets from the three individual methods were integrated by the Borda rank election method. For each miRNA, only its top 100 targets were computed. During this process, a *z*-score was calculated for each miRNA-mRNA interaction. All interactions were ordered in descending order of *z*-scores, and the top 50 interactions were regarded as seed miRNA-mRNA interactions for UM patients, as displayed in Table 1.

We found that among the 50 interactions, 10 of them had *z*-score > 2,000, especially 3 ones with *z*-score > 3,000, while the *z*-score of 12 interactions ranged from 1,000 to 2,000. In details, the pair of hsa-mir-203-*ASPG* obtained the highest *z*-score of 3,204. The other two interactions with *z*-score > 3,000 were hsa-mir-195-*BSDC1* (*z*-score = 3,179), and hsa-mir-3915-*C4BPA* (*z*-score = 3,007). The followed two miRNA-mRNA interactions were hsa-mir-30a-*C6orf155* (*z*-score = 2972), and hsa-mir-1253-*C6orf191* (*z*-score = 2748). Interestingly, *HOXA10* was regulated by two miRNAs (hsa-mir-196b and hsa-mir-196a-1) at the same time.

3.2 Validations of predicted miRNA targets

With an attempt to validate miRNA targets predicted by the ensemble method, we took a comparison of our results with confirmed miRTarBase, Tarbase, miRecords and miRWalk database. In short, miRTarbasev4.5 contains

Table 1 Seed miRNA-mRNA interactions for UM patients

ID	miRNA	mRNA	<i>z</i> -score	ID	miRNA	mRNA	<i>z</i> -score
1	hsa-mir-203	<i>ASPG</i>	3204	26	hsa-mir-3166	<i>LMAN1</i>	873
2	hsa-mir-195	<i>BSDC1</i>	3179	27	hsa-mir-3612	<i>MC2R</i>	851
3	hsa-mir-3915	<i>C4BPA</i>	3007	28	hsa-mir-335	<i>MEST</i>	822
4	hsa-mir-30a	<i>C6orf155</i>	2972	29	hsa-mir-155	<i>MIR155HG</i>	809
5	hsa-mir-1253	<i>C6orf191</i>	2748	30	hsa-mir-186	<i>MKNK1</i>	774
6	hsa-mir-511-2	<i>CD209</i>	2530	31	hsa-mir-92b	<i>MMP11</i>	748
7	hsa-mir-150	<i>CD96</i>	2484	32	hsa-mir-501	<i>NEDD9</i>	729
8	hsa-mir-3927	<i>DEFB109P1B</i>	2218	33	hsa-mir-142	<i>NLRP1</i>	713
9	hsa-mir-1247	<i>DIO3</i>	2104	34	hsa-mir-708	<i>ODZ4</i>	710
10	hsa-mir-221	<i>EXTL1</i>	2007	35	hsa-mir-935	<i>OGG1</i>	705
11	hsa-mir-887	<i>FBXL7</i>	1986	36	hsa-mir-143	<i>OR51E1</i>	703
12	hsa-mir-504	<i>FGF13</i>	1863	37	hsa-mir-3200	<i>OSBP2</i>	701
13	hsa-mir-105-1	<i>GABRA3</i>	1853	38	hsa-mir-139	<i>PDE2A</i>	700
14	hsa-mir-1185-2	<i>GPX5</i>	1794	39	hsa-let-7b	<i>SEC22C</i>	697
15	hsa-mir-1185-1	<i>HECW1</i>	1766	40	hsa-mir-383	<i>SGCZ</i>	693
16	hsa-mir-196b	<i>HOXA10</i>	1735	41	hsa-mir-584	<i>SH3TC2</i>	689
17	hsa-mir-196a-1	<i>HOXC10</i>	1684	42	hsa-mir-134	<i>SLIT3</i>	682
18	hsa-mir-196a-2	<i>HOXC11</i>	1507	43	hsa-mir-181a-1	<i>SORBS2</i>	680
19	hsa-mir-10b	<i>HOXD8</i>	1436	44	hsa-mir-513b	<i>TBC1D22B</i>	679
20	hsa-mir-3614	<i>ISG15</i>	1332	45	hsa-mir-199a-1	<i>TGFB1</i>	679
21	hsa-mir-874	<i>KLHL3</i>	1105	46	hsa-mir-140	<i>NFATC4</i>	678
22	hsa-mir-2861	<i>KRT39</i>	1082	47	hsa-mir-24-2	<i>PAIP2B</i>	672
23	hsa-mir-511-1	<i>LILRB5</i>	973	48	hsa-mir-532	<i>PCBP4</i>	670
24	hsa-mir-618	<i>LIN7A</i>	927	49	hsa-mir-216b	<i>PDC</i>	669
25	hsa-mir-873	<i>LINGO2</i>	904	50	hsa-mir-151	<i>PYCR1</i>	668

37,372 miRNA-mRNA interactions (covering 576 miRNAs). There were 20,095 interactions with 228 miRNAs in Tarbase v6.0. A total of 21,590 interactions representing 195 miRNAs were found in miRecords v2013. And miRWalk v2.0 covers 1,710 miRNA-mRNA interactions involved 226 miRNAs. By removing the duplicated interactions, we obtained total 62,858 confirmed interactions for validations. When comparing our predicted miRNA-mRNA interactions with confirmed interactions, 38 interactions were matched, which further indicated that our method was an available and valuable method for predicting miRNA targets.

3.3 Pathway enrichment analysis

After prediction and validation for miRNA targets obtained from the ensemble method, we aimed to identify significant functional gene sets of miRNA targets. Due to the too large scale of miRNA targets, we selected genes enriched in the top 1,000 ranked interactions which might be more important than the others for UM as study objects. Thus, KEGG pathway enrichment analysis was conducted on 601 targets in the top 1,000 miRNA-mRNA interaction based on the DAVID tool. When setting the cut-off as p-value < 0.05 (adjusted by Benjamini-Hochberg (BH) method), a total of 12 target pathways were detected (Table 2). The

top five significant pathways were Phototransduction ($P = 1.85E-06$), Chemokine signaling pathway ($P = 4.36E-05$), Ribosome ($P = 7.13E-04$), Phenylalanine metabolism ($P = 2.25E-03$), and Cytokine-cytokine receptor interaction ($P = 5.02E-03$). Particularly, Phototransduction was comprised of 9 targets including *CNGB1*, *GNAT1*, *GNAT2*, *GNGT1*, *GUCA1A*, *GUCY2F*, *RCVRN*, *RHO* and *GUCA1C*. Meanwhile, the Chemokine signaling pathway consisted of 21 targets (*ADCY1*, *GNB3*, *GNGT1*, *HCK*, *ITK*, *PRKCD*, *CCL4*, *CCL5*, *CXCL11*, *VAV2*, *CXCL14*, *CXCR6*, *GNG13*, *RPL10A*, *RPL3*, *RPL11*, *RPL22*, *RPL35A*, *RPS8*, *RPS23* and *RPS27A*).

4 Discussion

MiRNAs, a family of small non-coding RNA molecules, regulate expressions of genes by promoting mRNA degradation and repressing translation [25]. Their roles and functions in tumors have attracted more and more attentions from researchers, and the possible inferences are that miRNA participate in cancer-related processes, including proliferation, metabolism, differentiation, apoptosis and even cancer development and progression [26]. But there have been few studies to uncover miRNA targets in UM systemically. Hence, in this paper, we predicted target genes and pathways for UM patients based on the ensemble method that was an integration of

Table 2 Target pathways in top 1000 miRNA-mRNA interactions

ID	Pathway	miRNA targets	P value
1	Phototransduction	<i>CNGB1;GNAT1;GNAT2;GNGT1;GUCA1A;GUCY2F;RCVRN;RHO;GUCA1C</i>	1.85E-06
2	Chemokine signaling pathway	<i>ADCY1;GNB3;GNGT1;HCK;ITK;PRKCD;CCL4;CCL5;CXCL11;VAV2;CXCL14;CXCR6;GNG13;RPL10A;RPL3;RPL11;RPL22;RPL35A;RPS8;RPS23;RPS27A</i>	4.36E-05
3	Ribosome	<i>DDC;HPD;MAOB</i>	7.13E-04
4	Phenylalanine metabolism	<i>DDC;HPD;MAOB</i>	2.25E-03
5	Cytokine-cytokine receptor interaction	<i>TNFRSF8;CSF2RB;CTF1;IL2RB;IL12RB1;LTB;NGFR;CCL4;CCL5;CXCL11;TNFRSF1B;CXCL14;CXCR6;TNFRSF19;RELT</i>	5.02E-03
6	Long-term depression	<i>GRIA1;GRIA3;GRID2;GRM5;IGF1;RYR1</i>	2.33E-02
7	Primary immunodeficiency	<i>LCK;PTPRC;TAP1;ZAP70</i>	3.74E-02
8	Cell adhesion molecules (CAMs)	<i>HLA-F;PECAM1;PTPRC;SDC2;SIGLEC1;CNTNAP1;CADM1;CNTNAP2;CADM3</i>	3.85E-02
9	Amyotrophic lateral sclerosis (ALS)	<i>DAXX;GRIA1;MAPK12;TNFRSF1B;DERL1</i>	3.91E-02
10	Tyrosine metabolism	<i>DDC;HPD;MAOB;HEMK1</i>	4.77E-02
11	Glycosaminoglycan biosynthesis - heparan sulfate / heparin	<i>EXT1;EXTL1;NDST4</i>	4.79E-02
12	Neuroactive ligand-receptor interaction	<i>CHRNA3;CHRNA4;CHRN3;EDNRB;GABRA1;GABRA3;GABRG2;GRIA1;GRIA3;GRID2;GRIK1;GRM5;HTR2B;MC2R</i>	4.91E-02

The p-values have been corrected based on Benjamini & Hochberg method. $P < 0.01$ was considered as the threshold of significance.

PCC, IDA and Lasso methods.

Briefly, PCC is the commonly used correlation method for the strength between a pair of variables [27]. But it often leads to negative rank of miRNA-mRNA correlations due to down-regulation of miRNAs for mRNAs [11]. In addition, the PCC would not be greatly reduced if the data were in the non-linear distribution [28]. Meanwhile, IDA is a causal inference method that counts the causal effects between two variables [29, 30]. And the miRNA-mRNA correlations predicted by the IDA method have parts of overlap with outcomes of the follow-up gene knockdown experiments [31]. As for the Lasso, it minimizes the usual sum of squared errors, with a bound on the sum of the absolute values of the coefficients [32]. Like the limitation of PCC method, the miRNA-mRNA pairs identified by Lasso have negative effects are ranked at the top of the ranking list to favor the down regulation. Moreover, the ensemble method captured confirmed interactions in the incomplete ground truth that existing individual methods fail to discover, although there is no complete ground truth of miRNA target prediction [11].

Therefore, we employed Borda count election method to integrate the above three methods together, and obtained the ensemble method. Generally speaking, great challenges have been occurred on validating our predicted results, because the amount of experimentally confirmed miRNA targets is still limited and there is no complete authority for accessing and comparing different computational methods [33]. Hence the feasibility of our predicted results has been validated by comparing them with confirmed interactions. Results of the ensemble method showed that hsa-mir-203-*ASPG*, hsa-mir-195-*BSDC1* and hsa-mir-3915-*C4BPA* were the most important miRNA-mRNA interactions, and consequently *ASPG*, *BSDC1* and *C4BPA* were more critical target genes for UM than the others predicted. However, there have still been no studies to investigate the regulatory mechanisms of hsa-mir-203-*ASPG*, hsa-mir-195-*BSDC1*, and hsa-mir-3915-*C4BPA*. miR-203 has been reported to be overexpressed in pancreatic adenocarcinoma cells [34], while it also has been suggested as a tumor-inhibitory miRNA in hepatocellular carcinoma [35]. The abnormal of miR-195 in many cancers has also been reported by many researchers. It increased in breast cancer and chronic lymphocytic leukemia while decreased in gastric cancer, hepatocellular carcinoma, colorectal carcinoma and bladder cancer [36]. So far, study on miR-3915 was still limited. *ASPG* (asparaginase, also known as 60-kDa lysophospholipase) catalyzes the hydrolysis of L-asparagine to L-aspartate and ammonia [37]. It is used for remission induction and intensification treatment in all pediatric regimens and in the majority

of adult treatment protocols [38]. *C4BPA* (complement component 4 binding protein alpha) a member of a super-family of proteins composed predominantly of tandemly arrayed short consensus repeats of approximately 60 amino acids [39]. It had been reported that the *C4BPA* locus was a new susceptibility locus for venous thrombosis via protein S regulation, opening a new research area focusing on *C4BP* regulatory pathway [40]. It is the first time to uncover the relations between the target genes and UM, and further experimental validations would be finished as soon as possible.

As mentioned above, KEGG pathway enrichment analysis for 601 target genes in top 1,000 miRNA-mRNA interactions were performed, and 12 target pathways with $P < 0.05$ were identified. Importantly, Phototransduction and Chemokine signaling pathway were the most ones for UM compared with normal controls. The definition for Phototransduction in KEGG pathway database is a biochemical process by which the photoreceptor cells generate electrical signals in response to captured photons. Aguila et al revealed that heat shock protein 90 inhibition on visual function are likely to relate to essential its client proteins in the phototransduction pathway in the retina and potentially elsewhere in the eye [41]. Hence target pathway Phototransduction was related to UM tightly.

In conclusion, we have successfully predicted miRNA target genes and pathways for UM patients based on the ensemble method. The findings in this study might shed new light on uncovering the molecular mechanism underlying UM, and provide potential target signatures for prevention and treatment of this tumor. Moreover, whether the predicted miRNA targets are indeed involved in the development of UM, need to be confirmed by experiments urgently.

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Reference

- [1] Harbour J.W., Onken M.D., Roberson E.D., Duan S., Cao L., Worley L.A., et al., Frequent mutation of BAP1 in metastasizing uveal melanomas, *Science*, 2010, 330, 1410-1413
- [2] Mallone S., De V.E., Guzzo M., Midena E., Verne J., Coebergh J.W., et al., Descriptive epidemiology of malignant mucosal

- and uveal melanomas and adnexal skin carcinomas in Europe, *European Journal of Cancer*, 2012, 48, 1167-1175
- [3] Amirouchene-Angelozzi N., Nemati F., Gentien D., Nicolas A., Dumont A., Carita G., et al., Establishment of novel cell lines recapitulating the genetic landscape of uveal melanoma and preclinical validation of mTOR as a therapeutic target, *Molecular Oncology*, 2014, 8, 1508-1520
- [4] Schmidt-Pokrzywniak A., Kalbitz S., Kuss O., Jöckel K.-H., Bornfeld N., Stang A., Assessment of the effect of iris colour and having children on 5-year risk of death after diagnosis of uveal melanoma: a follow-up study, *BMC Ophthalmology*, 2014, 14, 42
- [5] Singh A.D., Turell M.E., Topham A.K., Uveal melanoma: trends in incidence, treatment, and survival, *Ophthalmology*, 2011, 118, 1881-1885
- [6] Itahana Y., Han R., Barbier S., Lei Z., Rozen S., Itahana K., The uric acid transporter SLC2A9 is a direct target gene of the tumor suppressor p53 contributing to antioxidant defense, *Oncogene*, 2015, 34, 1799-1810
- [7] Miller A.K., Benage M.J., Wilson D.J., Skalet A.H., Uveal Melanoma with Histopathologic Intratumoral Heterogeneity Associated with Gene Expression Profile Discordance, *Ocular Oncology and Pathology*, 2017, 3, 156-160
- [8] Field M.G., Harbour J.W., GNAQ/11 Mutations in Uveal Melanoma: Is YAP the Key to Targeted Therapy?, *Cancer cell*, 2014, 25, 714-715
- [9] Harbour J.W., The genetics of uveal melanoma: an emerging framework for targeted therapy, *Pigment cell & melanoma research*, 2012, 25, 171-181
- [10] Jay C., Nemunaitis J., Chen P., Fulgham P., Tong A.W., miRNA profiling for diagnosis and prognosis of human cancer, *DNA and cell biology*, 2007, 26, 293-300
- [11] Le T.D., Zhang J., Liu L., Li J., Ensemble Methods for miRNA Target Prediction from Expression Data, *PLoS ONE*, 2015, 10, e0131627
- [12] Marbach D., Costello J.C., Küffner R., Vega N.M., Prill R.J., Camacho D.M., et al., Wisdom of crowds for robust gene network inference, *Nature Methods*, 2012, 9, 796-804
- [13] Huber W., von Heydebreck A., Sultmann H., Poustka A., Vingron M., Variance stabilization applied to microarray data calibration and to the quantification of differential expression, *Bioinformatics*, 2002, 18 Suppl 1, S96-104
- [14] Russell N., Complexity of control of Borda count elections, 2007,
- [15] Chou C.-H., Chang N.-W., Shrestha S., Hsu S.-D., Lin Y.-L., Lee W.-H., et al., miRTarBase 2016: updates to the experimentally validated miRNA-target interactions database, *Nucleic acids research*, 2015, gkv1258
- [16] Vergoulis T., Vlachos I.S., Alexiou P., Georgakilas G., Maragkakis M., Reczko M., et al., TarBase 6.0: capturing the exponential growth of miRNA targets with experimental support, *Nucleic acids research*, 2012, 40, D222-D229
- [17] Xiao F., Zuo Z., Cai G., Kang S., Gao X., Li T., miRecords: an integrated resource for microRNA-target interactions, *Nucleic acids research*, 2009, 37, D105-D110
- [18] Dweep H., Gretz N., Sticht C., miWalk Database for miRNA-Target Interactions, *RNA Mapping: Methods and Protocols*, 2014, 289-305
- [19] Hsu S.-D., Tseng Y.-T., Shrestha S., Lin Y.-L., Khaleel A., Chou C.-H., et al., miRTarBase update 2014: an information resource for experimentally validated miRNA-target interactions, *Nucleic acids research*, 2014, 42, D78-D85
- [20] Papadopoulos G.L., Reczko M., Simossis V.A., Sethupathy P., Hatzigeorgiou A.G., The database of experimentally supported targets: a functional update of TarBase, *Nucleic acids research*, 2009, 37, D155-D158
- [21] Huang D.W., Sherman B.T., Lempicki R.A., Systematic and integrative analysis of large gene lists using DAVID bioinformatics resources, *Nat. Protoc.*, 2008, 4, 44-57
- [22] Kanehisa M., KEGG: Kyoto Encyclopedia of Genes and Genomes, *Nucleic Acids Research*, 2000, 27, 29-34(26)
- [23] Routledge R., Fisher's Exact Test. John Wiley & Sons, Ltd, 2005
- [24] Benjamini Y., Drai D., Elmer G., Kafkafi N., Golani I., Controlling the false discovery rate in behavior genetics research, *Behavioural brain research*, 2001, 125, 279-284
- [25] Bartel D., MicroRNAs: genomics, biogenesis, mechanism, and function. *cell*. 2004; 116: 281-297. In: PubMed Abstract| Publisher Full Text OpenURL. 2014
- [26] Ha M., Kim V.N., Regulation of microRNA biogenesis, *Nature Reviews Molecular Cell Biology*, 2014, 15, 509-524
- [27] Nahler G., Pearson correlation coefficient, *Dictionary of Pharmaceutical Medicine*, 2009, 132-132
- [28] Speed T., A correlation for the 21st century, *Science*, 2011, 334, 1502-1503
- [29] Maathuis M.H., Kalisch M., Bühlmann P., Estimating high-dimensional intervention effects from observational data, *The Annals of Statistics*, 2009, 37, 3133-3164
- [30] Maathuis M.H., Colombo D., Kalisch M., Bühlmann P., Predicting causal effects in large-scale systems from observational data, *Nature Methods*, 2010, 7, 247-248
- [31] Le T.D., Liu L., Tsykin A., Goodall G.J., Liu B., Sun B.-Y., et al., Inferring microRNA-mRNA causal regulatory relationships from expression data, *Bioinformatics*, 2013, btt048
- [32] Friedman J., Hastie T., Tibshirani R., glmnet: Lasso and elastic-net regularized generalized linear models. R package version 1.9-5. R Foundation for Statistical Computing Vienna. 2013
- [33] Le T.D., Liu L., Zhang J., Liu B., Li J., From miRNA regulation to miRNA-TF co-regulation: computational approaches and challenges, *Briefings in bioinformatics*, 2015, 16, 475-496
- [34] Greither T., Grochola L.F., Udelnow A., Lautenschlager C., Wurl P., Taubert H., Elevated expression of microRNAs 155, 203, 210 and 222 in pancreatic tumors is associated with poorer survival, *International journal of cancer*, 2010, 126, 73-80
- [35] Furuta M., Kozaki K.I., Tanaka S., Arii S., Imoto I., Inazawa J., miR-124 and miR-203 are epigenetically silenced tumor-suppressive microRNAs in hepatocellular carcinoma, *Carcinogenesis*, 2010, 31, 766-776
- [36] Yang B., Tan Z., Song Y., Study on the molecular regulatory mechanism of MicroRNA-195 in the invasion and metastasis of colorectal carcinoma, *International journal of clinical and experimental medicine*, 2015, 8, 3793-3800
- [37] Karamitros C.S., Konrad M., Human 60-kDa lysophospholipase contains an N-terminal L-asparaginase domain that is allosterically regulated by L-asparagine, *Journal of Biological Chemistry*, 2014, 289, 12962-12975
- [38] Pieters R., Hunger S.P., Boos J., Rizzari C., et al., L-asparaginase treatment in acute lymphoblastic leukemia †, *Cancer*, 2011, 117, 238-249

- [39] Cai X.-W., Shedden K.A., Yuan S.-H., Davis M.A., Xu L.-Y., Xie C.-Y., et al., Baseline Plasma Proteomic Analysis to Identify Biomarkers that Predict Radiation-Induced Lung Toxicity in Patients Receiving Radiation for Non-small Cell Lung Cancer, *Journal of Thoracic Oncology*, 6, 1073-1078
- [40] Buil A., Trégouët D.A., Souto J.C., Saut N., Germain M., Rotival M., et al., C4BPB/C4BPA is a new susceptibility locus for venous thrombosis with unknown protein S-independent mechanism: results from genome-wide association and gene expression analyses followed by case-control studies, *Blood*, 2010, 115, 4644-4650
- [41] Aguilà M., Cheetham M.E., Hsp90 as a Potential Therapeutic Target in Retinal Disease. In: *Retinal Degenerative Diseases: Mechanisms and Experimental Therapy*. Bowes Rickman C., LaVail M.M., Anderson R.E., Grimm C., Hollyfield J. and Ash J. (eds.) Springer International Publishing. 2016; Cham, pp 161-167