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Data Article

Draft genome sequence data of T-5 like *Salmonella* bacteriophage Φ SP3 with demonstrated therapeutic potential



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ABSTRACT

This data article describes the complete draft genome sequence of *Salmonella* specific bacteriophage Φ SP3 isolated from chicken intestinal contents. The Φ SP3 genome was sequenced by paired end runs using Illumina HiSeq 2500 with 100X coverage. Phylogenetic analysis using major capsid gene and genome wide comparison were performed to understand bacteriophage evolutionary relationship. Genome sequence of bacteriophage Φ SP3 was deposited in GenBank under the accession number MG387042.

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1. Data

Bacteriophage Φ SP3 was isolated from chicken intestinal contents obtained from retail market in Cochin (10.060256 N; 76.321881E) as a therapeutic agent against Salmonella [1]. The physicochemical characteristics of the bacteriophage were studied in detail [1]. The phage significantly reduced bacteria applied on chicken cuts, especially at refrigerated conditions, making it an ideal candidate for storage applications [2]. A consortium of phages including Φ SP3 also increased the longevity of *C. elegans* infected with Salmonella [3], indicating its ability to control infections.

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Specifications Table

Subject area	Biology
More specific subject area	Genomics
Type of data	Genome sequence data
How data was acquired	shot gun method using Illumina HiSeq 2500 with paired end runs
Data format	Raw and analyzed
Experimental factors	chicken intestinal content obtained from retail market
Experimental features	Draft genome sequence of Salmonella bacteriophage Φ SP3
Data source location	Cochin, India (10.060256 N; 76.321881E)
Data accessibility	Genome sequence was deposited in GenBank under the accession number MG387042

Value of the Data

- The sequence data confirms the lytic nature and absence of toxic genes of Φ SP3 bacteriophage, fulfilling with the requirements for future commercialization.
- Data contributes to phage genomics which requires many more phages to be sequenced and analyzed, to fill the gaps of unidentified proteins as well as evolutionary relationships.
- Phage genome data can also be used for screening and Identification of novel antimicrobial proteins.

The transmission electron micrograph of Φ SP3 showed a bacteriophage with a hexagonal head (53.77 ± 0.38 nm) and a distinguishable long non-contractile tail (123.66 ± 0.32 nm), which are typical morphological features of family *Siphoviridae*, Fig. 1a [4]. The draft genome sequence of Φ SP3 genome had a size of 109,106 bp with 39.5% GC content. Many short overlapping regions between adjoining genes were frequently detected. There were 166 predicted ORFs in the genome of which 57 ORFs were with assigned functions (S1). 55 genes were transcribed in rightward direction (strand +) while 111 genes on the leftward direction (strand -). 16 tRNA encoding genes were identified. No gene related to phage lysogeny was detected, confirming the lytic nature of the phage. Moreover, the absence of genes encoding virulence and allergy inducing genes makes Φ SP3 highly innocuous for application. Single gene analysis using major capsid gene (ORF 149) was used to determine bacteriophage evolutionary relationship. It was observed that Φ SP3 clustered together with the T5 like phages (Fig. 1b). Another approach to understand the relationship is via whole genome comparison with related phages. This approach revealed that Φ SP3 was 97% similar to *Salmonella* phage SPC35. Aligning Φ SP3 with well-studied T5 genome [10] showed 95% similarity, but did not show any similarity to T4, T7 and T3 phages.

The genome map of Φ SP3 is displayed in Fig. 2. Genome annotation analysis showed that Φ SP3 genome is functionally organized into modules containing gene clusters involved in different functions viz genes required for inactivation of host genome as well as transfer of DNA, genes involved in nucleotide metabolism, lytic processes, packing and morphogenesis cluster. The DNA end structures can be predicted from the terminase amino acid sequence as the enzymes that generate the virion DNA ends are quite diverse. These different types of ends reflect differing DNA replication. Accordingly, the amino acid sequence of large terminase genes can be used to predict the packing strategy of phages [5,6]. The large terminase gene with 1316 bp (ORF 156) of phage Φ SP3 clustered with phages having known DNA termini and packing mechanisms. Phage Φ SP3 terminase gene clustered with that of T5 which shows that they have complex concatemeric packing mechanisms (Fig. 3) Moreover, the blast alignment with *Salmonella* phage SPC35 showed presence of long terminal repeats with a size of around 9000 bp.

2. Experimental design, materials, and methods

The phage morphology was determined by transmission electron microscopy (Model JOEL JEM-100X).

Phage DNA was extracted as per Sambrook *et al.* [7], and purity was determined in terms of 260/280 and 260/230 ratios. Phage genome was sequenced by shot gun method using Illumina HiSeq 2500 with



Fig. 1. a) Transmission electron micrograph image of phage Φ SP3 stained with 1% uranyl acetate (bar represents 60nm). b) Phylogenetic tree based on major capsid gene of selected bacteriophages. The gene sequences are compared using the ClustalW program, and the phylogenetic tree was generated using the neighbour-joining method and 1000 bootstrap replicates.



Fig. 2. Genome map of bacteriophage Φ SP3 (DNA plotter).



Fig. 3. a) Phylogenetic tree on based amino acid sequences of terminase gene. Φ SP3 were clustered with phages of known termini and with experimentally determined packaging mechanisms. The DNA termini structures of phages that have been experimentally determined are indicated on right side on each cluster [5] structures have been experimentally determined are indicated.

paired end runs with 100X Coverage. The whole genome sequences were assembled using IVA [8] and SEQuel for correcting errors [9]. Genes were predicted using GeneMarkS [10]. Predicted ORFs were annotated with BLASTX, Uniprot, NCBI Conserved Domain Database (CDD). DNA Plotter was used to construct phage genome map [11] and tRNA gene prediction by tRNAscan-SE [12]. Phylogenetic tree depicting the evolutionary relationship of *Salmonella* bacteriophage was generated based on terminase gene and major capsid gene by neighbour-joining method [13] using MEGA 7.0 software [14].

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Conflict of Interest

The authors declare that they have no known competing financial interests or personal relationships that could have appeared to influence the work reported in this paper.

Appendix A. Supplementary data

Supplementary data to this article can be found online at https://doi.org/10.1016/j.dib.2019.104606.

References

- J. Augustine, S.G. Bhat, Physicochemical characterization of a T5-like Salmonella phage [Phi] SP-3, J. Microbiol. Biotechnol. Food Sci. 4 (2014) pp102–pp107, https://doi.org/10.15414/jmbfs.2014.4.2.102-107.
- [2] J. Augustine, S.G. Bhat, Bio control of Salmonella Enteritidis in spiked chicken cuts by lytic bacteriophages ΦSP-1 and ΦSP-3, J. Basic Microbiol. 55 (2015) 500–503.

- [3] J. Augustine, M.V. Gopalakrishnan, S.G. Bhat, Application of ΦSP-1 and ΦSP-3 as a therapeutic strategy against Salmonella Enteritidis infection using Caenorhabditis elegans as model organism, FEMS Microbiol. Lett. 356 (2014) pp113–pp117.
- [4] M. Krupovic, B.E. Dutilh, E.M. Adriaenssens, J. Wittmann, F.K. Vogensen, M.B. Sullivan, J. Rumnieks, D. Prangishvili, R. Lavigne, A.M. Kropinski, J. Klumpp, Taxonomy of prokaryotic viruses: update from the ICTV bacterial and archaeal viruses subcommittee, Arch. Virol. 161 (2016) pp1095-pp1099.
- [5] S.R. Casjens, E.B. Gilcrease, D.A. Winn-Stapley, P. Schicklmaier, H. Schmieger, M.L. Pedulla, R.W. Hendrix, The generalized transducing *Salmonella* bacteriophage ES18: complete genome sequence and DNA packaging strategy, J. Bacteriol. 187 (2005) 1091–1104.
- [6] S.R. Casjens, E.B. Gilcrease, Determining DNA packaging strategy by analysis of the termini of the chromosomes in tailedbacteriophage virions, Bacteriophages Methods Protocols Vol. 2 Molec. Appl. Aspect. (2009) 91–111.
- [7] J. Sambrook, E. Fritsch, I. Maniatis, Molecular Cloning A Laboratory Manual, second ed., Cold Spring Harbor Laboratory, Cold Spring Harbor, 2000.
- [8] M. Hunt, A. Gall, S.H. Ong, J. Brener, B. Ferns, P. Goulder, E. Nastouli, J.A. Keane, P. Kellam, T.D. Otto, IVA: accurate de novo assembly of RNA virus genomes, Bioinformatics 31 (2015) 2374–2376.
- [9] R. Ronen, C. Boucher, H. Chitsaz, P. Pevzner, SEQuel: improving the accuracy of genome assemblies, Bioinformatics 12 (2012) i188–i196.
- [10] J. Besemer, A. Lomsadze, M. Borodovsky, GeneMarkS: a self-training method for prediction of gene starts in microbial genomes. Implications for finding sequence motifs in regulatory regions, Nucleic Acids Res. 29 (2001) 2607–2618.
- [11] T. Carver, N. Thomson, A. Bleasby, M. Berriman, J. Parkhill, DNAPlotter: circular and linear interactive genome visualization, Bioinformatics 25 (2009) 119–120.
- [12] T.M. Lowe, S.R. Eddy, tRNAscan-SE: a program for improved detection of transfer RNA genes in genomic sequence, Nucleic Acids Res. 25 (1997) 955–964.
- [13] N. Saitou, M. Nei, The neighbor-joining method: a new method for reconstructing phylogenetic trees, Mol. Biol. Evol. 4 (1987) 406-425.
- [14] S. Kumar, G. Stecher, K. Tamura, MEGA7: molecular evolutionary genetics analysis version 7.0 for bigger datasets, Mol. Biol. Evol. 33 (2016) 1870–1874.