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Article

Electrochemical Oxidative Clean Halogenation Using HX/NaX with Hydrogen Evolution

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SUMMARY

Organic halides (R-X) are prevalent structural motifs in pharmaceutical molecules and key building blocks for the synthesis of fine chemicals. Although a number of routes are available in the literature for the synthesis of organic halides, these methods often require stoichiometric additives or oxidants, metal catalysts, leaving or directing groups, or toxic halogenating agents. In addition, the necessity of employing different, often tailor-made, catalytic systems for each type of substrate also limits the applicability of these methods. Herein, we report a clean halogenation by electrochemical oxidation with NaX/HX. A series of organic halides were prepared under metal catalyst- and exogenous-oxidant-free reaction conditions. It is worth noting that this reaction has a broad substrate scope; various heteroarenes, arenes, alkenes, alkynes, and even aliphatic hydrocarbons could be applied. Most importantly, the reaction could also be performed on a 200-mmol scale with the same efficiency (86%, 50.9 g pure product).

INTRODUCTION

Organic halides (R-X) are compounds of high practical utility, which are not only important structural motifs in many pharmaceutical molecules and natural products (Hernandez et al., 2010; Jeschke, 2010; Butler and Sandy, 2009) but also key building blocks for the synthesis of fine chemicals via transition-metal-catalyzed oxidative/reductive cross-coupling reactions (Yue et al., 2018; Fairlamb, 2007; Nicolaou et al., 2005; Meijere and Diederich, 2004; Liu et al., 2017a). Consequently, practical and efficient methods to access this class of compounds are highly valuable. Extensive efforts have been made, and great achievement has been reached (Ye et al., 2018; Mo et al., 2010; Petrone et al., 2016; Rafiee et al., 2018; Liu et al., 2017b; Fu et al., 2017a; Wang et al., 2012; Wallentin et al., 2012; Liu and Groves, 2010; Murphy et al., 2007; Smith et al., 2002), such as the electrophilic aromatic substitutions (Barluenga et al., 2007; Prakash et al., 2004; David, 1976) and the directed C-H halogenations (Teskey et al., 2015; Schröder et al., 2015; Schröder et al., 2012; Bedford et al., 2011; Kakiuchi et al., 2009; Mei et al., 2008; Whitfield and Sanford, 2007; Wan, 2006). Although these methods have been widely used for the synthesis of organic halides (R-X), they still have one or more of the following limitations: (1) the use of hazardous and toxic X_2 (X = Br, Cl) as halogenating agents; (2) the need of stoichiometric amount of additives/exogenous oxidants; (3) the need of a metal salt as the catalyst; (4) the need of custom-built substrate bearing leaving or directing groups; (5) the necessity of employing different, often tailor-made, catalytic systems for each types of substrate; and (6) the harsh reaction conditions. Therefore, exploring an efficient and versatile method for the synthesis of various organic halides (R-X) with non-toxic and green halogenating agents under environmentally benign metalcatalyst-free and exogenous-oxidant-free reaction conditions would be highly desirable.

Electrochemical anodic oxidation presents the prospect of the efficient and environmentally benign synthesis of complex molecules and has attracted considerable interest (Tang et al., 2018a; Yoshida et al., 2018; Jiang et al., 2018; Yan et al., 2017; Pletcher et al., 2018; Francke and Little, 2014; Jutand, 2008; Sperry and Wright, 2006; Qiu et al., 2018; Xiong et al., 2017; Gieshoff et al., 2017; Fu et al., 2017b; Yang et al., 2017; Horn et al., 2016; Badalyan and Stahl, 2016; Kärkäs, 2018; Liu et al., 2018; Lyalin and Petrosyan, 2013; Raju et al., 2006; Kulangiappar et al., 2016; Tan et al., 2017). As part of our continuing studies in the area of electrochemical oxidative C-C and C-heteroatom bonds formation (Yuan et al., 2019; Tang et al., 2018b; Gao et al., 2018; Yuan et al., 2018a, 2018b, 2018c), we herein report a clean halogenation by exogenousoxidant-free electrochemical oxidation. A series of significant organic halides (R-X) were prepared under ¹National Research Center for Carbohydrate Synthesis, Jiangxi Normal University, Nanchang, Jiangxi 330022, P. R. China

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Entry	Variation from the Standard Conditions	Yield (%)ª
1	None	81
2	HCI (aq.) instead of NaCl	49
3	LiCl instead of NaCl	34
4	KCI instead of NaCI	75
5	MgCl ₂ instead of NaCl	44
6	CaCl ₂ instead of NaCl	51
7	6 mA, 7 h	75
8	18 mA, 2.3 h	70
9	Carbon cloth cathode	61
10	Platinum plate anode	53
11	Without H ₂ O	69
12	MeCN instead of DMF	48
13	No electric current	ND

Table 1. Optimization of Electrochemical Oxidative C-H Chlorination

Reaction conditions: carbon rod anode, platinum plate cathode, constant current = 12 mA, **1a** (0.3 mmol), **2a** (2.0 equiv.), DMF (10.5 mL), H₂O (0.5 mL), 80°C, N₂, 3.5 h (5.2 F/mol).

ND, not detected.

^alsolated yields.

metal-catalyst-free and exogenous-oxidant-free reaction conditions with commercially available, nontoxic, and atom-efficient NaX/HX (aq.). It is worth noting that this electrochemical oxidative synthetic protocol has a broad substrate scope. Various heteroarenes, arenes, alkenes, alkynes, and even aliphatic hydrocarbons were suitable for this transformation.

RESULTS AND DISCUSSION

Imidazopyridines (Dyminska, 2015; Enguehard-Gueiffier and Gueiffier, 2007), especially C-3-substituted imidazopyridines, are often used as commercially available drugs including alpidem (Okubo et al., 2004), zolpidem (Langer et al., 1990), necopidem (Depoortere and George, 1991), and saripidem (Sanger, 1995). The introduction of a halogen moiety into the C-3 position of imidazopyridines has been considered to be important because the generated C-3 halogenated imidazopyridines are key intermediates for the synthesis of these drugs. Our investigation included 2-phenylimidazo[1,2-a]pyridine (1a) and sodium chloride (2a) as the starting materials for the synthesis of these class of significant C-3 halogenated imidazopyridines. As shown in Table 1, by employing a two-electrode system with carbon rod as the anode and platinum plate as the cathode, the desired C-H chlorination product 3a was produced in 81% yield with a 12 mA constant current in an undivided cell (entry 1). A range of other chlorides were investigated, but all displayed lower effectiveness than sodium chloride (entries 2–6). Both decreasing the operating current to 8 mA led to slightly decreased reaction yields (entries 7–8). Then different electrode materials were surveyed, employing either carbon cloth as cathode or platinum plate as anode led to decreased reaction efficiency (entries 9–10). The effect of solvent was explored as





Figure 1. Substrate Scope for Electrochemical Oxidative C-H Halogenation

(A) Substrate scope of C-H chlorination.
(B) Substrate scope of C-H bromination.
(C) Gram-scale synthesis.
Reaction conditions: carbon rod anode, platinum plate cathode, constant current = 12 mA, 1 (0.3 mmol), 2a (2.0 equiv.) or 2b (4.0 equiv.), DMF (10.5 mL), H₂O (0.5 mL), 80°C, N₂, 3.5 h (5.2 F/mol), isolated yields.
^aCH₃CN (10.5 mL), H₂O (0.5 mL), 75°C.
^b1 (2.0 equiv.), 2 (0.3 mmol).
^c7.0 h.

well. When N,N-dimethylformamide was used as the sole solvent, 69% yield of **3a** could still be obtained (entry 11). However, when the reaction was performed using acetonitrile instead of N,N-dimethylformamide, an obvious loss of the yield was observed (entry 12). As was expected, no reaction could be observed in the absence of electric current (entry 13).

With the optimized reaction conditions in hand, the scope and generality of this clean halogenation was explored (Figure 1). With respect to the C-H chlorination (Figure 1A), diverse heteroarenes/arenes served as effective reaction partners with 2a to form C-Cl bond. The phenyl- and naphthyl-substituted imidazo [1,2-a]pyridines showed good reactivity and gave the corresponding products in 81% and 76% yields (Figure 1A, 3a-b), respectively. 2-Arylimidazo[1,2-a]pyridines bearing halogen substituents on the phenyl ring delivered the C-H chlorination products in good to high yields (Figure 1A, 3d-f). Delightfully, strong electron-withdrawing groups such as trifluoromethyl and cyano at the para position of the phenyl ring of 2-phenylimidazo[1,2-a]pyridines nearly did not affect the reaction efficiency (Figure 1A, 3g-h). By contrast, 2-phenylimidazo[1,2-a]pyridines bearing electron-rich group showed decreased reaction efficiency (Figures 1A, 3i). It is worth noting that the substrates bearing tert-butyl, trifluoromethyl, and -H groups at the C-2 position of imidazo[1,2-a]pyridines also reacted smoothly and delivered the desired products in moderate to good yields (Figure 1A, 3j-I). Moreover, imidazo[1,2-a]pyridines bearing various substituents such as methyl, chlorine, and trifluoromethyl groups at different positions of the pyridine ring all furnished the C-H chlorination products in high yields (Figure 1A, 3m-p). Besides various imidazo[1,2-a]pyridines, 1-phenylpyrazole, benzo[d]-imidazo[2,1-b]thiazole derivatives, and very-electronrich 1,3,5-trimethoxybenzene were also suitable substrates for this transformation, affording the desired products in 62%-90% yields (Figure 1A, 3q-t).

We subsequently turned our attention to the C-H bromination (Figure 1B). To our delight, imidazo[1,2-a] pyridines bearing various substituents such as alkyl, alkoxy, halogen, cyano, and trifluoromethyl groups at different positions of the phenyl ring or pyridine ring all underwent clean transformations to generate the C-H bromination products in good to excellent yields (Figure 1B, 4a-o). Notably, besides various imidazo[1,2-a]pyridines, other kinds of heteroarenes and arenes were also suitable for this transformation. For example, 2-aminopyridine, benzo[d]-imidazo[2,1-b]thiazole derivative, 1-phenylpyrazole, 3-phenylpyrazole, 8-aminoquinoline, and 2-aminopyridine derivatives all delivered the corresponding C-H bromination products in moderate to high yields (Figure 1B, 4p-v). It is worth noting that for 3-phenylpyrazole and 8-aminoquinoline, the double C-H bromination products were the major products (Figure 1B, 4r-s). In the case of electron-rich arenes, p-chloroaniline and p-bromoaniline afforded the corresponding C-H bromination products in good yields (Figure 1B, 4w-x). The very-electron-rich arenes, such as 1,3,5-trimethoxybenzene and 3,5-dimethoxytoluene, gave the C-Br bond formation products in 85% and 83% yields (Figure 1B, 4y-z), respectively. To examine the scalability of the exogenous-oxidant-free electrochemical oxidative C-H halogenation, reactions on the 15- and 50-mmol scale were performed (Figure 1C). The corresponding C-H halogenation products were afforded in 81% and 70% isolated yield, respectively (see Supplemental Information for details).

To shed light on the reaction mechanism for this electrochemical oxidative C-H halogenation, a series of control experiments were conducted. First, voltammograms of the substrates were recorded (see Figure S160 of the Supplemental Information for details). The oxidation peak of 1a was observed in N,N-dimethylformamide (DMF)/H₂O at 1.59 V, whereas the oxidation peak of NaCl and NaBr were observed at 1.55 V and 1.40 V, respectively, which indicated that NaCl or NaBr was likely to be first oxidized under the electrolytic conditions. Moreover, under the standard optimized conditions, no homo-coupling product of 1a was observed in either C-H chlorination or bromination (Figures 2A and 2B). These results further indicated that NaCl and NaBr are readily oxidized than 1a in this electrochemical oxidative C-H halogenation. The reaction of 1a with molecular Cl₂ and Br₂ in the absence of electricity was also investigated



Figure 2. Control Experiments

(Figures 2C-2E). When 1.0 equiv. of molecular Br_2 was added into the reaction system, the desired C-H bromination product could be obtained in high yield and H_2O did not affect the efficiency of this reaction, whereas no chlorination product was detected when molecular Cl_2 was used as the chlorinating agent. These results suggest that molecular Cl_2 might not be involved as the intermediate in C-H chlorination, whereas molecular Br_2 ought to be a key intermediate in C-H bromination. Meanwhile, the pathway in which molecular Br_2 reacted with H_2O yielding the Br^+ (HOBr), then attacked by heteroarenes (1) to form the desired product, could be completely ruled out. Last but not least, the reaction of 1a with MeOH in the absence of sodium halides was carried out (Figure 2F); 9% homo-coupling product of 1a was isolated from the reaction system, but the product of radical cation intermediate captured by MeOH was not detected. These results suggest that the pathway in which 1a is oxidized to the corresponding radical cation intermediate and then captured by nucleophile could be ruled out.



Figure 3. Proposed Mechanism of C-H Halogenation

Based on the above-mentioned experimental results, a plausible reaction mechanism for C-H halogention is depicted in Figure 3. For the C-H chlorination, the reaction begins with the anodic oxidation of chlorine ion to generate the chlorine radical. The radical intermediate A could then be formed through a radical addition of chlorine radical to 1a. Finally, further single-electron oxidation and the following deprotonation led to product 3a. Concomitant cathodic reduction of water leads to hydrogen evolution. Different from the C-H chlorination, in C-H bromination, bromide ion is directly oxidized to molecular Br₂, which then is attacked by 1a to access the intermediate C. Finally, the following deprotonation led to the product 4a.

Having successfully demonstrated electrochemical oxidative halogenation of heterocycles/arenes, we subsequently turned our attention to the other type substrates. Indeed, this versatile electrochemical oxidative synthetic protocol was not limited to the heterocycles/arenes; alkenes (5) were identified as amenable substrates as well. As shown in Figure 4A, when the ratio of alkenes to HBr (aq.) was 1:2, various styrenes and aliphatic alkenes were compatible with the reaction conditions, providing the desired C-Br double bond forming products in moderate to high yields (Figures 4A, **6a-6x**). Moreover, besides terminal alkenes, internal alkenes were also tolerated in this electrochemical system, and the *trans*-1,2-dibromides were isolated as the sole diastereomeric products (Figures 4A, **6k**, **6r**, **6s**, **6u**). This result suggests that molecular Br_2 might be the key intermediate for this transformation. To evaluate the practicability of this method, we conducted the exogenous-oxidant-free electrochemical oxidative dibromination of 1-decene on a 200-mmol scale and finally obtained 50.9 g pure product (Figure 4B; see Supplemental Information for details), which is hard to access traditionally. This indicates that our protocol could be conveniently scaled up in industry.

To develop a more general method, we also turned our attention to investigate the dibromination of alkynes (Figure 5). Delightfully, when 4-methoxyphenylacetylene (7a) and 1-phenyl-1-propyne (7b) were employed as the surrogates of alkynes, the desired dibromination products were isolated in 65% and 33% yields (Figure 5), respectively, and *E*-isomers were isolated as the sole diastereomeric products. This result suggests that molecular Br_2 might also be the key intermediate in this dibrominating reaction.

To further affirm that the alkene and alkyne dibromination involved a molecular Br_2 intermediate, the reaction of molecular Br_2 with styrene (**5a**) and 4-methoxyphenylacetylene (**7a**) in the absence of electricity was investigated (Figure 6), respectively. The reaction results indicate that these two transformations indeed involve a molecular Br_2 intermediate.

The success of the heteroarenes, arenes, alkenes, and alkynes led us to extend this method to aliphatic hydrocarbons because alkyl halides are also powerful substrates. To our delight, when ethyl 2-pyridylace-tate (9) and α -menthylstyrene (12) were employed as the surrogates of aliphatic hydrocarbons, the desired alkyl halides 10 and 13 were isolated in 54% and 32% yields (Figure 7), respectively. Moreover, for

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Figure 4. Substrate Scope for Electrochemical Oxidative Dibromination of Alkenes

(A) Substrate scope of alkenes.

(B) Gram-scale synthesis.

Reaction conditions: carbon rod anode, platinum plate cathode, constant current = 12 mA, 5 (0.5 mmol), 2c (2.0 equiv.), MeCN (10.8 mL), H₂O (0.2 mL), ⁿBu₄NBF₄ (0.1 mmol), RT, N₂, 3.0 h (2.7 F/mol), isolated yields.



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Figure 5. Substrate Scope for Electrochemical Oxidative Dibromination of Alkynes Reaction conditions: carbon rod anode, platinum plate cathode, constant current = 12 mA, **7** (0.3 mmol), **2b** (4.0 equiv.), MeCN (10.5 mL), H₂O (0.5 mL), 75°C, N₂, 3.5 h, isolated yields.

2-pyridylacetate (9), when the amount of sodium bromide (2b) was increased to 4.0 equiv. and the reaction time was extended to 3.5 h, the double C-H halogenated product 11 could be isolated in 40% yield.

Limitations of Study

Substrate scope of alkyne dibromination is limited to the electron-rich alkynes.

Conclusion

We have successfully employed constant current for clean halogenation. A series of significant organic halides (R-X) were prepared under a metal-catalyst-free and exogenous-oxidant-free reaction conditions with commercially available, nontoxic, and atom-efficient NaX/HX (aq.). Remarkably, this electrochemical oxidative synthetic protocol has a broad substrate scope. Besides, various heteroarenes/arenes, alkenes, alkynes, and aliphatic hydrocarbons were also suitable. Most importantly, the reaction could also be performed on a 200-mmol scale with the same efficiency (86%, 50.9 g pure product), which further highlighted the synthetic practicability of this electrochemical oxidative strategy.



Figure 6. Mechanism Experiments



Figure 7. Electrochemical Oxidative C-H Bromination of Aliphatic Hydrocarbons

METHODS

All methods can be found in the accompanying Transparent Methods supplemental file.

SUPPLEMENTAL INFORMATION

Supplemental Information includes Transparent Methods and 160 figures and can be found with this article online at https://doi.org/10.1016/j.isci.2019.01.017.

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AUTHOR CONTRIBUTIONS

A.L. and Y.Y. conceived the project and designed the experiments. Y.Y., A.Y., Y.Z., Z.Z., J.Q., J.H., B.Y., J.Z., and H.W. performed and analyzed the experiments. Y.Y., A.L., and M.G. wrote the manuscript. Y.Y., A.Y., and Y.Z. wrote the Supplemental Information and contributed other related materials. All the authors discussed the results and commented on the manuscript.

DECLARATION OF INTERESTS

The authors declare no competing interests.

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Supplemental Information

Electrochemical Oxidative Clean

Halogenation Using HX/NaX

with Hydrogen Evolution

Yong Yuan, Anjin Yao, Yongfu Zheng, Meng Gao, Zhilin Zhou, Jin Qiao, Jiajia Hu, Baoqin Ye, Jing Zhao, Huilai Wen, and Aiwen Lei

Supplemental Information

Electrochemical Oxidative Clean Halogenation Using HX/NaX with Hydrogen Evolution

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Copies of product NMR spectra

Figure S1. ¹H NMR (400 MHz, CDCl₃) spectrum of compound 3a, related to Figure 1



Figure S2. ¹³C NMR (100 MHz, CDCl₃) spectrum of compound 3a, related to Figure 1



Figure S3. ¹H NMR (400 MHz, DMSO-*d*₆) spectrum of compound **3b**, related to **Figure 1**



Figure S4. ¹³C NMR(100 MHz, CDCl₃) spectrum of compound 3b, related to Figure 1

143.72	139.62 133.36 133.06 133.06 129.85 128.44	128.08 127.61 126.69 126.27 126.17 125.02 124.92 124.92 124.92 125.02 117.53 112.91 117.53	77.32 77.00 76.68
1	1 -> >6		\leq





Figure S6. ¹³C NMR (100 MHz, CDCl₃) spectrum of compound 3c, related to Figure 1



Figure S7. ¹H NMR (400 MHz, CDCl₃) spectrum of compound 3d, related to Figure 1 1.00 A1.032.101.05 $3.01 \pm$ 5.0 4.5 fl (ppm) 9.5 7.5 7.0 8.0 5.5 4.0 3.5 2.5 1.5 . 1.0 9.0 8.5 6.5 6.0 3.0 2.0 0.5 0.0

Figure S8. ¹³C NMR (100 MHz, CDCl₃) spectrum of compound 3d, related to Figure 1







ii (ppii),





Figure S11. ¹³C NMR (100 MHz, CDCl₃) spectrum of compound **3e**, related to Figure 1





Figure S12. ¹H NMR (400 MHz, DMSO-*d*₆) spectrum of compound **3f**, related to Figure 1

Figure S13. ¹³C NMR (100 MHz, CDCl₃) spectrum of compound **3f**, related to Figure 1



Figure S14. ¹H NMR (400 MHz, DMSO-*d*₆) spectrum of compound 3g, related to Figure 1



Figure S15. ¹³C NMR (100 MHz, CDCl₃) spectrum of compound 3g, related to Figure 1



Figure S16. ¹⁹F NMR (376 MHz, CDCl₃) spectrum of compound **3g**, related to Figure 1



10 0 -10 -20 -30 -40 -50 -60 -70 -80 -90 -100 -110 -120 -130 -140 -150 -160 -170 -180 -190 -200 -210 fl (ρρm) Figure S17. ¹H NMR (400 MHz, CDCl₃) spectrum of compound **3h**, related to Figure 1



Figure S18. ¹³C NMR (100 MHz, CDCl₃) spectrum of compound 3h, related to Figure 1



Figure S19. ¹H NMR (400 MHz, CDCl₃) spectrum of compound 3i, related to Figure 1



Figure S20. ¹³C NMR (100 MHz, CDCl₃) spectrum of compound 3i, related to Figure 1







Figure S22. ¹³C NMR (100 MHz, CDCl₃) spectrum of compound 3j, related to Figure 1



Figure S23. ¹H NMR (400 MHz, DMSO- d_6) spectrum of compound **3k**, related to **Figure 1**



Figure S24. ¹³C NMR (100 MHz, CDCl₃) spectrum of compound 3k, related to Figure 1







10 0 -10 -20 -30 -40 -50 -50 -70 -80 -90 -100 -110 -120 -130 -140 -150 -160 -170 -180 -190 -200 -210 fl (ppm) Figure S26. ¹H NMR (400 MHz, CDCl₃) spectrum of compound 3l, related to Figure 1



Figure S27. ¹³C NMR (100 MHz, CDCl₃) spectrum of compound **3**l, related to Figure 1





Figure S28. 1 H NMR (400 MHz, CDCl₃) spectrum of compound 3m, related to Figure 1

Figure S29. ¹³C NMR (100 MHz, CDCl₃) spectrum of compound 3m, related to Figure 1



Figure S30. ¹H NMR (400 MHz, DMSO-*d*₆) spectrum of compound 3n, related to Figure 1



Figure S31. ¹³C NMR (100 MHz, CDCl₃) spectrum of compound 3n, related to Figure 1



Figure S32. ¹H NMR (400 MHz, DMSO-*d*₆) spectrum of compound 30, related to Figure 1



Figure S33. 13 C NMR (100 MHz, CDCl₃) spectrum of compound 30, related to Figure 1

143.32 141.61 131.67 128.79 128.63	127.86 127.48 124.70 122.00	121.88 121.82 121.77 121.71	120.76 120.74 120.71 120.69 119.30 118.32	117.91 117.57 1117.57 1116.89 1116.89 107.21 777.32 777.00
				$ \rightarrow $



Figure S34. ¹⁹F NMR (376 MHz, CDCl₃) spectrum of compound **30**, related to Figure 1



Figure S35. ¹H NMR (400 MHz, CDCl₃) spectrum of compound **3p**, related to Figure 1



Figure S36. ¹³C NMR (100 MHz, CDCl₃) spectrum of compound **3p**, related to Figure 1





Figure S38. ¹³C NMR (100 MHz, CDCl₃) spectrum of compound 3q, related to Figure 1



Figure S39. 1 H NMR (400 MHz, CDCl₃) spectrum of compound 3r, related to Figure 1



Figure S40. ¹³C NMR (100 MHz, CDCl₃) spectrum of compound **3r**, related to Figure 1


Figure S41. 1 H NMR (400 MHz, CDCl₃) spectrum of compound 3s, related to Figure 1



Figure S42. ¹³C NMR (100 MHz, CDCl₃) spectrum of compound 3s, related to Figure 1

147.93	$\begin{array}{c} 139.79\\ 135.43\\ 135.54\\ 122.54\\ 122.88\\ 122.88\\ 125.81\\ 125.13\\ 125.13\\ 125.08\\ 125.13\\ 125.08\\ 125.13\\ 125.08\\ 125.13\\ 125.08\\ 122.08\\$	90.78	77.32 77.00 76.68
1		1	





Figure S44. ¹³C NMR (100 MHz, CDCl₃) spectrum of compound 3t, related to Figure 1



Figure S45. ¹H NMR (400 MHz, DMSO-*d*₆) spectrum of compound 4a, related to Figure 1



Figure S46. ¹³C NMR (100 MHz, DMSO-*d*₆) spectrum of compound 4a, related to Figure 1



Figure S47. ¹H NMR (400 MHz, DMSO-*d*₆) spectrum of compound 4b, related to Figure 1



Figure S48. ¹³C NMR (100 MHz, CDCl₃) spectrum of compound 4b, related to Figure 1







Figure S50. ¹³C NMR (100 MHz, CDCl₃) spectrum of compound 4c, related to Figure 1

145.32 142.67 138.23 129.19 127.76 123.90 117.46 112.99	91.41	77.32 77.00 76.68	21.33
	1	\checkmark	1







Figure S52. ¹³C NMR (100 MHz, CDCl₃) spectrum of compound 4d, related to Figure 1





Figure S54. ¹H NMR (400 MHz, CDCl₃) spectrum of compound 4e, related to Figure 1



Figure S55. ¹³C NMR (100 MHz, CDCl₃) spectrum of compound 4e, related to Figure 1





Figure S56. ¹H NMR (400 MHz, DMSO- d_6) spectrum of compound 4f, related to Figure 1

Figure S57. ¹³C NMR (100 MHz, CDCl₃) spectrum of compound 4f, related to Figure 1



Figure S58. ¹H NMR (400 MHz, DMSO-*d*₆) spectrum of compound 4g, related to Figure 1







Figure S59. ¹³C NMR (100 MHz, DMSO-*d*₆) spectrum of compound 4g, related to Figure 1



Figure S60. ¹H NMR (400 MHz, DMSO- d_6) spectrum of compound 4h, related to Figure 1



Figure S61. ¹³C NMR (100 MHz, DMSO-*d*₆) spectrum of compound 4h, related to Figure 1



Figure S62. ¹⁹F NMR (376 MHz, CDCl₃) spectrum of compound 4h, related to Figure 1



Figure S63. ¹H NMR (400 MHz, DMSO- d_6) spectrum of compound **4**i, related to **Figure 1**





Figure S64. ¹³C NMR (100 MHz, DMSO-*d*₆) spectrum of compound 4i, related to Figure 1





Figure S66. ¹³C NMR (100 MHz, DMSO-*d*₆) spectrum of compound 4j, related to Figure 1





Figure S67. ¹H NMR (400 MHz, CDCl₃) spectrum of compound 4k, related to Figure 1

Figure S69. ¹H NMR (400 MHz, CDCl₃) spectrum of compound 4l, related to Figure 1



Figure S70. ¹³C NMR (100 MHz, CDCl₃) spectrum of compound 4l, related to Figure 1



Figure S71. ¹H NMR (400 MHz, DMSO-*d*₆) spectrum of compound 4m, related to Figure 1



Figure S72. ¹³C NMR (100 MHz, CDCl₃) spectrum of compound 4m, related to Figure 1



Figure S73. ¹H NMR (400 MHz, CDCl₃) spectrum of compound 4n, related to Figure 1



Figure S74. ¹³C NMR (100 MHz, CDCl₃) spectrum of compound 4n, related to Figure 1



Figure S75. ¹H NMR (400 MHz, DMSO-*d*₆) spectrum of compound 40, related to Figure 1



Figure S76. ¹³C NMR (100 MHz, CDCl₃) spectrum of compound 40, related to Figure 1



Figure S77. ¹H NMR (400 MHz, CDCl₃) spectrum of compound **4p**, related to **Figure 1**



Figure S78. ¹³C NMR (100 MHz, CDCl₃) spectrum of compound 4p, related to Figure 1

$\int_{125.09}^{147.89} \frac{147.89}{132.82}$	- 113.51	- 91.81	$\frac{77.32}{\sqrt{77.00}}$
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Figure S79. ¹H NMR (400 MHz, CDCl₃) spectrum of compound **4q**, related to **Figure 1**



Figure S80. ¹³C NMR (100 MHz, CDCl₃) spectrum of compound 4q, related to Figure 1

~ 141.43 ~ 139.52	earrow 129.48 $ earrow 126.98 $ $ earrow 126.95$	- 118.93	- 95.56	$\frac{17.32}{17.00}$
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Figure S81. 1 H NMR (400 MHz, CDCl₃) spectrum of compound 4r, related to Figure 1

Figure S82. ¹³C NMR (100 MHz, CDCl₃) spectrum of compound 4r, related to Figure 1







Figure S84. ¹³C NMR (100 MHz, CDCl₃) spectrum of compound 4s, related to Figure 1

148.22 141.93 138.09 135.66 133.10	126.61 122.37	106.75	77.32 77.00 76.68
1 5 5 5 5			\leq



Figure S85. ¹H NMR (400 MHz, DMSO-*d*₆) spectrum of compound 4t, related to Figure 1



Figure S86. ¹³C NMR (100 MHz, CDCl₃) spectrum of compound 4t, related to Figure 1





Figure S88. ¹³C NMR (100 MHz, CDCl₃) spectrum of compound 4u, related to Figure 1



Figure S87. ¹H NMR (400 MHz, CDCl₃) spectrum of compound 4u, related to Figure 1

Figure S89. ¹H NMR (400 MHz, DMSO-d₆) spectrum of compound 4v, related to Figure 1



Figure S90. ¹³C NMR (100 MHz, CDCl₃) spectrum of compound 4v, related to Figure 1

157.77	144.60 144.56 137.30 137.27	127.33 124.63 121.94 119.25 118.22 117.58 117.55 117.51 117.21 103.44	77.32 77.00 76.68
	\checkmark \checkmark		\leq

















Figure S95. ¹³C NMR (100 MHz, CDCl₃) spectrum of compound 4x, related to Figure 1





Figure S97. ¹³C NMR (100 MHz, CDCl₃) spectrum of compound 4y, related to Figure 1





80 90 fl (ppm) . 170 . 160 . 150 . 140 130 120 100 . 80 70 60 50 . 40 . 30 . 20 10 0 110

Figure S100. ¹H NMR (400 MHz, CDCl₃) spectrum of compound **6a**, related to Figure 4



Figure S101. ¹³C NMR (100 MHz, CDCl₃) spectrum of compound 6a, related to Figure 4





Figure S102. ¹H NMR (400 MHz, CDCl₃) spectrum of compound 6b, related to Figure 4

Figure S103. ¹³C NMR (100 MHz, CDCl₃) spectrum of compound 6b, related to Figure 4



Figure S104. ¹H NMR (400 MHz, CDCl₃) spectrum of compound 6c, related to Figure 4



Figure S105. ¹³C NMR (100 MHz, CDCl₃) spectrum of compound 6c, related to Figure 4



Figure S106. ¹⁹F NMR (376 MHz, CDCl₃) spectrum of compound 6c, related to Figure 4





Figure S107. ¹H NMR (400 MHz, CDCl₃) spectrum of compound 6d, related to Figure 4

Figure S108. ¹³C NMR (100 MHz, CDCl₃) spectrum of compound 6d, related to Figure 4




Figure S109. ¹H NMR (400 MHz, CDCl₃) spectrum of compound 6e, related to Figure 4

Figure S110. ¹³C NMR (100 MHz, CDCl₃) spectrum of compound 6e, related to Figure 4



Figure S111. ¹H NMR (400 MHz, CDCl₃) spectrum of compound **6f**, related to **Figure 4**



Figure S112. ¹³C NMR (100 MHz, CDCl₃) spectrum of compound 6f, related to Figure 4



Figure S113. ¹H NMR (400 MHz, CDCl₃) spectrum of compound **6g**, related to **Figure 4**



Figure S114. ¹³C NMR (100 MHz, CDCl₃) spectrum of compound 6g, related to Figure 4

137.53 133.24 130.29 128.13 128.13 124.32	77.32 77.00 76.68	48.32	33.70
ノノンてし		ì	Ĩ

Br Br Br Br





Figure S115. 1 H NMR (400 MHz, CDCl₃) spectrum of compound 6h, related to Figure 4





Figure S117. ¹H NMR (400 MHz, CDCl₃) spectrum of compound **6i**, related to Figure **4**









Figure S124. ¹H NMR (400 MHz, CDCl₃) spectrum of compound 6l, related to Figure 4



Figure S125. ¹³C NMR (100 MHz, CDCl₃) spectrum of compound 6l, related to Figure 4





Figure S126. ¹H NMR (400 MHz, CDCl₃) spectrum of compound 6m, related to Figure 4





Figure S131. ¹³C NMR (100 MHz, DMSO-*d*₆) spectrum of compound **60**, related to Figure 4





Figure S133. ¹³C NMR (100 MHz, CDCl₃) spectrum of compound **6p**, related to Figure 4





Figure S135. ¹³C NMR (100 MHz, CDCl₃) spectrum of compound 6q, related to Figure 4

77.32 77.00 76.68 50.9943.9537.2135.5233.7731.4826.4026.4025.83

Br





Figure S137. ¹³C NMR (100 MHz, CDCl₃) spectrum of compound 6r, related to Figure 4





Figure S139. ¹³C NMR (100 MHz, CDCl₃) spectrum of compound 6s, related to Figure 4









Figure S143. ¹³C NMR (100 MHz, CDCl₃) spectrum of compound 6u, related to Figure 4

77.32 77.00 76.68	55.95	36.10 24.63	23.29 23.08 22.53
			\sim





Figure S145. ¹³C NMR (100 MHz, CDCl₃) spectrum of compound 6v, related to Figure 4





Figure S147. ¹³C NMR (100 MHz, CDCl₃) spectrum of compound 6w, related to Figure 4

77.32 77.00 76.68	52.75 52.73	36.19 35.74 35.70 35.70 27.93 27.93 26.41 26.37
\sim	$\mathbf{\nabla}$	\checkmark

Br Br Br Br Br Br Br





Figure S149. ¹³C NMR (100 MHz, CDCl₃) spectrum of compound 6x, related to Figure 4



79





Figure S153. ¹³C NMR (100 MHz, CDCl₃) spectrum of compound 8b, related to Figure 5





Figure S154. ¹H NMR (400 MHz, CDCl₃) spectrum of compound 10, related to Figure 7

90 80 fl (ppm) .






Figure S159. ¹³C NMR (100 MHz, CDCl₃) spectrum of compound 13, related to Figure 7



Transparent Methods

The instrument for electrolysis is dual display potentiostat (DJS-292B) (made in China). The anodic electrode was graphite rod (ϕ 6 mm) and cathodic electrode was platinum plate (15 mm × 15 mm × 0.3 mm). Thin layer chromatography (TLC) employed glass 0.25 mm silica gel plates. Flash chromatography columns were packed with 200-300 mesh silica gel in petroleum (boiling point, 60 to 90 °C). NMR spectra were recorded on a Bruker spectrometer at 400 MHz (¹H NMR), 100 MHz (¹³C NMR), 376 MHz (¹⁹F NMR), respectively. All chemical shifts are reported relative to tetramethylsilane and solvent peaks. ¹H, ¹³C and ¹⁹F NMR data spectra were reported in delta (δ) units, parts per million (ppm) downfield from the internal standard. Coupling constants are reported in Hertz (Hz).

General procedure for electrochemical oxidative C–H chlorination: In an undivided three-necked bottle (25 mL) equipped with a stir bar, 1 (0.3 mmol), 2a (0.6 mmol, 35.1 mg), were combined and added. The bottle was equipped with graphite rod (ϕ 6 mm, about 18 mm immersion depth in solution) as the anode and platinum plate (15 mm × 15 mm × 0.3 mm) as the cathode and then charged with nitrogen. Under the protection of N₂, H₂O (0.5 mL) and DMF (10.5 mL) were injected respectively into the bottle via syringes. The reaction mixture was stirred and electrolyzed with a constant current of 12 mA at 80 °C for 3.5 h. When the reaction was finished, the solution was extracted with EtOAc (3×10mL) and H₂O (3×30mL). The combined organic layer was dried with Na₂SO₄, filtered. The solvent was removed with a rotary evaporator. The pure product was obtained by flash column chromatography on silica gel using petroleum ether and ethyl acetate as the eluent.

General procedure for electrochemical oxidative C–H bromination: In an undivided three-necked bottle (25 mL) equipped with a stir bar, **1** (0.3 mmol), **2b** (1.2 mmol, 123.5 mg.), were combined and added. The bottle was equipped with graphite rod (ϕ 6 mm, about 18 mm immersion depth in solution) as the anode and platinum plate (15 mm × 15 mm × 0.3 mm) as the cathode and then charged with nitrogen. Under the protection of N₂, H₂O (0.5 mL) and DMF (10.5 mL) were injected respectively into the bottle via syringes. The reaction mixture was stirred and electrolyzed with a constant current of 12 mA at 80 °C for 3.5 h. When the reaction was finished,

the solution was extracted with EtOAc (3×10 mL) and H₂O (3×30 mL). The combined organic layer was dried with Na₂SO₄, filtered. The solvent was removed with a rotary evaporator. The pure product was obtained by flash column chromatography on silica gel using petroleum ether and ethyl acetate as the eluent.

General procedure for electrochemical oxidative dibromination of alkenes: In an undivided three-necked bottle (25 mL) equipped with a stir bar, ^{*n*}Bu₄NBF₄ (0.1 mmol, 33 mg) was added. The bottle was equipped with graphite rod (ϕ 6 mm, about 18 mm immersion depth in solution) as the anode and platinum plate (15 mm × 15 mm × 0.3 mm) as the cathode and then charged with nitrogen. Under the protection of N₂, **5** (0.5 mmol), **2c** (1.0 mmol, 113 uL, 48%), H₂O (0.2 mL) and CH₃CN (10.8 mL) were injected respectively into the bottle via syringes. The reaction mixture was stirred and electrolyzed with a constant current of 12 mA at room temperature for 3 h. The pure product was obtained by flash column chromatography on silica gel using petroleum ether as the eluent.

Procedure for gram scale synthesis of electrochemical oxidative C–H bromination (15 mmol scale): In an undivided three-necked bottle equipped with a stir bar, **1a** (15 mmol), **2b** (4 equiv.) were combined and added. The bottle was equipped with graphite rod (ϕ 6 mm) as the anode and platinum plate (15 mm × 15 mm × 0.3 mm) as the cathode. Under the air, H₂O (5 mL) and DMF (105 mL) were injected respectively into the bottle via syringes. The reaction mixture was stirred and electrolyzed with a constant current of 60 mA at 80 °C for 35 h. When the reaction was finished, the solution was extracted with EtOAc (3×50 mL) and H₂O (3×150 mL). The combined organic layer was dried with Na₂SO₄, filtered. The solvent was removed with a rotary evaporator. The pure product was obtained by flash column chromatography on silica gel using petroleum ether and ethyl acetate as the eluent.

Procedure for gram scale synthesis of electrochemical oxidative C–H bromination (50 mmol scale): In an undivided three-necked bottle equipped with a stir bar, **1a** (50 mmol), **2b** (4 equiv.) were combined and added. The bottle was equipped with graphite rod (ϕ 6 mm) as the anode and platinum plate (15 mm × 15 mm × 0.3 mm) as the cathode. Under the air, H₂O (5 mL) and DMF (105 mL) were injected respectively into the bottle via syringes. The reaction mixture was stirred

and electrolyzed with a constant current of 120 mA at 80 °C for 58.4 h. When the reaction was finished, the solution was extracted with EtOAc ($3 \times 100 \text{ mL}$) and H₂O ($3 \times 300 \text{ mL}$). The combined organic layer was dried with Na₂SO₄, filtered. The solvent was removed with a rotary evaporator. The pure product was obtained by flash column chromatography on silica gel using petroleum ether and ethyl acetate as the eluent.

Procedure for gram scale synthesis of electrochemical oxidative dibromination of alkenes: In an undivided three-necked bottle equipped with a stir bar. The bottle was equipped with graphite rod (ϕ 6 mm) as the anode and platinum plate (15 mm × 15 mm × 0.3 mm) as the cathode and then charged with nitrogen. Under the N₂, **5t** (200 mmol), **2c** (400 mmol, 48%), H₂O (4 mL) and CH₃CN (210 mL) were injected respectively into the bottle via syringes. The reaction mixture was stirred and electrolyzed with a constant current of 120 mA at room temperature for 120 h. The pure product was obtained by flash column chromatography on silica gel using petroleum ether as the eluent.

Procedure for cyclic voltammetry (CV)

Cyclic voltammetry was performed in a three-electrode cell connected to a schlenk line under nitrogen at room temperature. The working electrode was a steady glassy carbon disk electrode, the counter electrode was a platinum wire. The reference was an Ag/AgCl electrode submerged in saturated aqueous KCl solution. 11 mL mix-solvent (DMF/H₂O = 10.5/0.5) containing 0.01 M n Bu₄NBF₄ were poured into the electrochemical cell in all experiments. The scan rate is 0.1 V/s, ranging from 0 V to 1.8 V.





Fig. S160. Cyclic voltammograms.

Characterization of all compounds



3-Chloro-2-phenylimidazo[1,2-a]pyridine (3a) (Xiao et al., 2015).

White solid was obtained in 81% isolated yield. ¹H NMR (400 MHz, CDCl₃) δ 8.13 (d, *J* = 8.1 Hz, 2H), 7.98 – 7.96 (m, 1H), 7.58 (d, *J* = 9.1 Hz, 1H), 7.46 (t, *J* = 7.7 Hz, 2H), 7.37 – 7.35 (m, 1H), 7.16 – 7.12 (m, 1H), 6.79 (t, *J* = 6.8 Hz, 1H); ¹³C NMR (100 MHz, CDCl₃) δ 143.60, 139.66, 132.42, 128.49, 128.20, 127.41, 124.81, 122.61, 117.56, 112.85, 105.62.



3-Chloro-2-(naphthalen-2-yl)imidazo[1,2-a]pyridine (3b).

White solid was obtained in 76% isolated yield. ¹H NMR (400 MHz, DMSO- d_6) δ 8.63 (s, 1H), 8.36 (d, J = 6.8 Hz, 1H), 8.27 – 8.25 (m, 1H), 8.02 (d, J = 8.5 Hz, 2H), 7.95 – 7.92 (m, 1H), 7.70 (d, J = 9.1 Hz, 1H), 7.57 – 7.52 (m, 2H), 7.41 – 7.37 (m, 1H), 7.10 (t, J = 6.6 Hz, 1H); ¹³C NMR (100 MHz, CDCl₃) δ 143.72, 139.62, 133.36, 133.06, 129.85, 128.44, 128.08, 127.61, 126.69, 126.27, 126.17, 125.02, 124.92, 122.61, 117.53, 112.91, 105.94. HRMS (ESI): m/z calcd for C₁₇H₁₂ClN₂ [M+H]⁺: 279.00684, found: 279.0688.



3-Chloro-2-(p-tolyl)imidazo[1,2-a]pyridine (3c) (Xiao et al., 2015).

White solid was obtained in 73% isolated yield. ¹H NMR (400 MHz, CDCl₃) δ 8.04 – 8.02 (m, 3H), 7.60 (d, J = 9.1 Hz, 1H), 7.28 – 7.266 (m, 2H), 7.19 – 7.15 (m, 1H), 6.86 – 6.82 (m, 1H), 2.39 (s, 3H); ¹³C NMR (100 MHz, CDCl₃) δ 143.50, 139.77, 138.00, 129.57, 129.14, 127.25, 124.55, 122.45, 117.37, 112.63, 105.17, 21.22.



3-Chloro-2-(4-fluorophenyl)imidazo[1,2-a]pyridine (3d).

White solid was obtained in 70% isolated yield. ¹H NMR (400 MHz, CDCl₃) δ 8.14 – 8.06 (m, 3H), 7.61 (d, *J* = 9.1 Hz, 1H), 7.26 (m, 1H), 7.19 – 7.13 (m, 2H), 6.93–6.90 (m, 1H); ¹³C NMR (100 MHz, CDCl₃) δ 163.91, 161.44, 143.59, 138.83, 129.21 (d, *J* = 8.2 Hz), 128.58 (d, *J* = 3.2 Hz), 124.98, 122.64, 117.51, 115.49 (d, *J* = 21.6 Hz), 112.96, 105.33; ¹⁹F NMR (376 MHz, CDCl₃) δ –113.20. HRMS (ESI): m/z calcd for C₁₃H₉ClFN₂ [M+H]⁺: 247.0433, found: 247.0436.



3-Chloro-2-(4-chlorophenyl)imidazo[1,2-a]pyridine (3e) (Xiao et al., 2015).

White solid was obtained in 75% isolated yield. ¹H NMR (400 MHz, CDCl₃) δ 8.06 – 8.01 (m, 3H), 7.59 (d, *J* = 9.1 Hz, 1H), 7.41 (d, *J* = 8.5 Hz, 2H), 7.23 – 7.19 (m, 1H), 6.88 (t, *J* = 6.8 Hz, 1H); ¹³C NMR (100 MHz, CDCl₃) δ 143.50, 138.40, 133.93, 130.88, 128.58, 128.46, 124.96, 122.52, 117.46, 112.91, 105.59.



3-Chloro-2-(3,4-dichlorophenyl)imidazo[1,2-a]pyridine (3f).

White solid was obtained in 73% isolated yield. ¹H NMR (400 MHz, DMSO- d_6) δ 8.36 (d, J = 6.8 Hz, 1H), 8.21 (d, J = 1.8 Hz, 1H), 8.05 – 8.02 (m, 1H), 7.74 (d, J = 8.5 Hz, 1H), 7.66 (d, J = 9.1 Hz, 1H), 7.43 – 7.39 (m, 1H), 7.12 (t, J = 6.8 Hz, 1H); ¹³C NMR (100 MHz, CDCl₃) δ 143.65, 137.25, 132.72, 132.50, 132.06, 130.39, 128.97, 126.31, 125.35, 122.66, 117.65, 113.22, 106.09. HRMS (ESI): m/z calcd for C₁₃H₈Cl₃N₂ [M+H]⁺: 296.9748, found: 296.9740.



3-Chloro-2-(4-(trifluoromethyl)phenyl)imidazo[1,2-a]pyridine (3g).

White solid was obtained in 73% isolated yield. ¹H NMR (400 MHz, DMSO-*d*₆) δ 8.34 (d, *J* = 6.9 Hz, 1H), 8.24 (d, *J* = 8.1 Hz, 1H), 7.80 (d, *J* = 8.3 Hz, 1H), 7.66 (d, *J* = 9.1 Hz, 1H), 7.40 – 7.36 (m, 1H), 7.11 – 7.07 (m, 1H); ¹³C NMR (100 MHz, CDCl₃) δ 143.77, 138.10, 135.97, 129.83 (q, *J* = 32.4 Hz), 127.45, 125.36 (q, *J* = 4.0 Hz), 125.37, 124.15 (q, *J* = 270.0 Hz), 122.69, 117.75, 113.21, 106.49; ¹⁹F NMR (376 MHz, CDCl₃) δ –62.53. HRMS (ESI): m/z calcd for C₁₄H₉ClF₃N₂ [M+H]⁺: 297.0401, found: 297.0405.



4-(3-Chloroimidazo[1,2-a]pyridin-2-yl)benzonitrile (3h).

White solid was obtained in 62% isolated yield. ¹H NMR (400 MHz, CDCl₃) δ 8.28 – 8.26 (m, 2H), 8.13 – 8.11 (m, 1H), 7.75 – 7.73 (m, 2H), 7.64 (d, *J* = 9.1 Hz, 1H), 7.33 – 7.28 (m, 1H), 6.01 – 6.97 (m, 1H); ¹³C NMR (100 MHz, CDCl₃) δ 143.80, 137.41 (d, *J* = 2 Hz), 136.89, 132.18, 127.53, 125.62, 122.73, 118.81, 117.79, 113.44, 111.27, 106.93. HRMS (ESI): m/z calcd for C₁₄H₉ClN₃ [M+H]⁺: 254.0480, found: 254.0485.



3-Chloro-2-(4-methoxyphenyl)imidazo[1,2-a]pyridine (3i) (Xiao et al., 2015).

White solid was obtained in 30% isolated yield. ¹H NMR (400 MHz, CDCl₃) δ 8.09 – 8.07 (m, 3H), 7.62 (d, *J* = 9.1 Hz, 1H), 7.24 – 7.20 (m, 1H), 7.02 (d, *J* = 8.7 Hz, 2H), 6.90 (t, *J* = 6.8 Hz, 1H), 3.86 (s, 3H); ¹³C NMR (100 MHz, CDCl₃) δ 159.62, 143.54, 139.62, 128.74, 125.03, 124.65, 122.52, 117.33, 113.93, 112.70, 104.76, 55.25.



2-(Tert-butyl)-3-chloroimidazo[1,2-a]pyridine (3j) (Li et al., 2018).

Light yellow liquid was obtained in 68% isolated yield. ¹H NMR (400 MHz, CDCl₃) δ 8.03 (d, *J* = 6.9 Hz, 1H), 7.59 (d, *J* = 9.1 Hz, 1H), 7.19 – 7.15 (m, 1H), 6.86 (t, *J* = 6.8 Hz, 1H), 1.51 (s, 9H); ¹³C NMR (100 MHz, CDCl₃) δ 149.42, 142.16, 123.69, 122.00, 117.22, 112.35, 104.33, 32.87, 29.49.



3-Chloro-2-(trifluoromethyl)imidazo[1,2-a]pyridine (3k).

White solid was obtained in 73% isolated yield. ¹H NMR (400 MHz, DMSO- d_6) δ 8.47 – 7.45 (m, 1H), 7.77 – 7.74 (m, 1H), 7.56 – 7.51 (m, 1H), 7.26 – 7.23(m, 1H); ¹³C NMR (100 MHz, CDCl₃) δ 143.40, 130.8 (q, J = 38.5 Hz), 126.57, 123.13, 121.09 (q, J = 267.0 Hz), 118.89, 114.51, 108.92; ¹⁹F NMR (376 MHz, CDCl₃) δ –61.77. HRMS (ESI): m/z calcd for C₈H₅ClF₃N₂ [M+H]⁺: 221.0088, found: 221.0091.



3-Chloroimidazo[1,2-a]pyridine (3l) (Li et al., 2018).

Light yellow oil was obtained in 37% isolated yield. ¹H NMR (400 MHz, CDCl₃) δ 8.05 (d, J = 6.9 Hz, 1H), 7.61 – 7.56 (m, 2H), 7.22 – 7.18 (m, 1H), 6.91 (t, J = 6.8 Hz, 1H); ¹³C NMR (100 MHz, CDCl₃) δ 144.29, 130.00, 124.06, 122.37, 117.88, 112.76, 109.40.



3-Chloro-6-methyl-2-phenylimidazo[1,2-a]pyridine (3m) (Xiao et al., 2015).

Light yellow liquid was obtained in 87% isolated yield. ¹H NMR (400 MHz, CDCl₃) δ 8.14 – 8.11 (m, 2H), 7.83 (s, 1H), 7.52 – 7.45 (m, 3H), 7.38 – 7.33 (m, 1H), 7.06 – 7.33 (m, 1H), 2.33 (s, 3H); ¹³C NMR (100 MHz, CDCl₃) δ 142.66, 139.37, 132.63, 128.40, 127.96, 127.91, 127.24, 122.62, 120.21, 116.81, 105.11, 18.25.



3,6-Dichloro-2-phenylimidazo[1,2-a]pyridine (**3n**) (Xiao et al., 2015).

White solid was obtained in 76% isolated yield. ¹H NMR (400 MHz, DMSO-*d*₆) δ 8.49 – 8.48 (m, 1H), 8.70 – 8.05 (m, 2H), 7.67 (d, *J* = 9.6 Hz, 1H), 7.52 – 7.48 (m, 2H), 7.42 – 7.36 (m, 2H); ¹³C NMR (100 MHz, CDCl₃) δ 141.93, 140.71, 131.96, 128.53, 128.47, 127.36, 126.26, 121.41, 120.55, 117.93, 106.08.



3-Chloro-2-phenyl-6-(trifluoromethyl)imidazo[1,2-a]pyridine (30).

White solid was obtained in 78% isolated yield. ¹H NMR (400 MHz, DMSO- d_6) δ 8.75 (s, 1H), 8.10 –8.07 (m, 2H), 7.85 (d, J = 9.5 Hz, 1H), 7.61 – 7.58 (m, 1H), 7.53 – 7.50 (m, 2H), 7.45 – 7.40 (m, 1H); ¹³C NMR (100 MHz, CDCl₃) δ 143.32, 141.61, 131.67, 128.79, 128.63, 127.48, 123.35 (q, J = 270.0 Hz), 121.8 (q, J = 5.7 Hz), 120.72 (q, J = 2.6 Hz), 118.32, 117.4 (q, J = 34.4

Hz), 107.21; ¹⁹F NMR (376 MHz, CDCl₃) δ –62.14. HRMS (ESI): m/z calcd for C₁₄H₉ClF₃N₂ [M+H]⁺: 297.0400, found: 297.0361.



3-Chloro-5-methyl-2-phenylimidazo[1,2-a]pyridine (3p).

Light yellow oil was obtained in 83% isolated yield. ¹H NMR (400 MHz, CDCl₃) δ 8.04 – 8.02 (m, 2H), 7.79 (d, *J* = 6.8 Hz, 1H), 7.36 (t, *J* = 7.6 Hz, 2H), 7.25 (t, *J* = 7.4 Hz, 1H), 6.86 (d, *J* = 6.9 Hz, 1H), 6.65 (t, *J* = 6.9 Hz, 1H), 2.53 (s, 3H); ¹³C NMR (100 MHz, CDCl₃) δ 143.99, 139.25, 132.75, 128.44, 128.00, 127.64, 127.58, 123.52, 120.45, 112.84, 105.86, 16.47. HRMS (ESI): m/z calcd for C₁₄H₁₂ClN₂ [M+H]⁺: 243.0684, found: 243.0690.



4-Chloro-1-phenyl-1H-pyrazole (3q) (Wang et al., 2016).

White solid was obtained in 90% isolated yield.¹H NMR (400 MHz, CDCl₃) δ 7.81 (s, 1H), 7.55 (s, 1H), 7.55 – 7.51 (m, 2H), 7.39 – 7.32 (m, 2H), 7.25 – 7.19 (m, 1H); ¹³C NMR (100 MHz, CDCl₃) δ 139.70, 139.46, 129.55, 127.01, 124.84, 118.96, 112.39.



3-Chloro-2-phenylbenzo[d]imidazo[2,1-b]thiazole (3r).

Yellow solid was obtained in 68% isolated yield.¹H NMR (400 MHz, CDCl₃) δ 8.01 (d, *J* = 8.2 Hz, 1H), 7.90 (d, *J* = 7.0 Hz, 2H), 7.49 (d, *J* = 8.0 Hz, 1H), 7.33 (t, *J* = 7.7 Hz, 2H), 7.26 – 7.13 (m, 3H); ¹³C NMR (100 MHz, CDCl₃) δ 145.90, 140.77, 132.24, 132.18, 129.78, 128.37, 127.65, 126.47, 125.90, 124.99, 123.95, 113.23, 108.27. HRMS (ESI): m/z calcd for C₁₅H₁₀ClN₂S[M+H]⁺: 285.0248, found:285.0258.



3-Chloro-2-(thiophen-2-yl)benzo[d]imidazo[2,1-b]thiazole (3s).

Yellow solid was obtained in 62% isolated yield. ¹H NMR (400 MHz, CDCl₃) δ 8.19 (d, J = 8.5 Hz, 1H), 7.62 (d, J = 3.7 Hz, 1H), 7.53 (d, J = 8.0 Hz, 1H), 7.31 – 7,27 (m, 1H), 7.25 – 7.18 (m, 2H), 7.02 – 7.00 (m, 1H).¹³C NMR (100 MHz, CDCl₃) δ 147.93, 139.79, 135.43, 132.54, 129.88, 127.45, 125.81, 125.13, 125.08, 124.29, 124.03, 113.26, 90.78. HRMS (ESI): m/z calcd for C₁₃H₈ClN₂S₂ [M+H]⁺: 290.9812, found: 290.9816.



2-Bromo-1,3,5-trimethoxybenzene (3t) (Tang et al., 2018).

White solid was obtained in 77% isolated yield. ¹H NMR (400 MHz, CDCl₃) δ 6.18 (s, 2H), 3.88 (s, 6H), 3.81 (s, 3H); ¹³C NMR (100 MHz, CDCl₃) δ 159.37, 156.49, 102.61, 91.55, 56.24, 55.48.



3-Bromo-2-phenylimidazo[1,2-a]pyridine (4a) (Zhou et al., 2016).

White solid was obtained in 87% isolated yield. ¹H NMR (400 MHz, DMSO-*d*₆) δ 8.35 (d, *J* = 6.9 Hz, 1H), 8.10 – 8.08 (m, 2H), 7.66 (d, *J* = 9.1 Hz, 1H), 7.51 – 7.47 (m, 2H), 7.42 – 7.35 (m, 2H), 7.09 – 7.05 (m, 1H); ¹³C NMR (100 MHz, DMSO-*d*₆) δ 144.87, 141.39, 132.81, 128.58, 128.27, 127.36, 125.87, 124.58, 117.09, 113.64, 91.65.



3-Bromo-2-(naphthalen-2-yl)imidazo[1,2-a]pyridine (4b) (Zhou et al., 2016).
White solid was obtained in 66% isolated yield. ¹H NMR (400 MHz, DMSO- d_6) δ 8.64 (s, 1H), 8.40 (d, J = 6.8 Hz, 1H), 8.27 – 7.25 (m, 1H), 8.03 – 8.00 (m, 2H), 7.95 – 7.93 (m, 1H), 7.70 (d, J = 9.0 Hz, 1H), 7.57 – 7.52 (m, 2H), 7.42 – 7.38 (m, 1H), 7.10 (t, J = 6.8 Hz, 1H); ¹³C NMR (100 MHz, CDCl₃) δ 145.37, 142.37, 133.22, 133.00, 130.17, 128.37, 127.93, 127.55, 127.01, 126.21, 126.10, 125.37, 125.06, 123.78, 117.41, 112.94, 91.91.



3-Bromo-2-(p-tolyl)imidazo[1,2-a]pyridine (4c) (Zhou et al., 2016).

White solid was obtained in 91% isolated yield. ¹H NMR (400 MHz, CDCl₃) δ 8.01 (d, *J* = 6.9 Hz, 1H), 7.92 (d, *J* = 8.2 Hz, 2H), 7.51 (d, *J* = 9.1 Hz, 1H), 7.19 – 7.15 (m, 2H), 7.13 – 7.09 (m, 1H), 6.79 – 6.75 (m, 1H), 2.29 (s, 3H); ¹³C NMR (100 MHz, CDCl₃) δ 145.32, 142.67, 138.23, 129.85, 129.19, 127.76, 125.05, 123.90, 117.46, 112.99, 91.41, 21.33.



3-Bromo-2-(4-fluorophenyl)imidazo[1,2-a]pyridine (4d) (Zhou et al., 2016).

White solid was obtained in 76% isolated yield. ¹H NMR (400 MHz, CDCl₃) δ 8.14 – 8.09 (m, 3H), 7.61 (d, J = 9.1 Hz, 1H), 7.26 – 7.22 (m, 1H), 7.19 – 7.13 (m, 2H), 6.92 – 6.89 (m, 1H); ¹³C NMR (100 MHz, CDCl₃) δ 162.68 (d, J = 248.0 H), 145.31, 141.69, 129.55 (d, J = 8.2 Hz), 128.95 (d, J = 3.2 Hz), 125.12, 123.85, 117.45, 115.36 (d, J = 21.5 Hz), 113.01, 91.32; ¹⁹F NMR (376 MHz, CDCl₃) δ –13.19.



3-Bromo-2-(4-chlorophenyl)imidazo[1,2-a]pyridine (4e) (Zhou et al., 2016).

White solid was obtained in 59% isolated yield. ¹H NMR (400 MHz, CDCl₃) δ 8.12 (d, *J* = 6.9 Hz, 1H), 8.09 – 8.05 (m, 2H), 7.61 (d, *J* = 9.1 Hz, 1H), 7.45 – 7.41 (m, 2H), 7.26 – 7.22 (m, 1H), 6.93

- 6.89 (m, 1H); ¹³C NMR (100 MHz, CDCl₃) δ 145.32, 141.34, 134.15, 131.22, 128.98, 128.61, 125.37, 123.91, 117.51, 113.20, 91.74.



3-Bromo-2-(3,4-dichlorophenyl)imidazo[1,2-a]pyridine (4f) (Zhou et al., 2016). White solid was obtained in 74% isolated yield. ¹H NMR (400 MHz, DMSO-*d*₆) δ 8.36 (d, *J* = 6.9 Hz, 1H), 8.25 (d, *J* = 2.0 Hz, 1H), 8.08 – 8.06 (m, 1H), 7.73 (d, *J* = 8.4 Hz, 1H), 7.65 (d, *J* = 9.1 Hz, 1H), 7.43 – 7.39 (m, 1H), 7.13 – 7.10 (m, 1H); ¹³C NMR (100 MHz, CDCl₃) δ 145.40, 140.10, 132.80, 132.70, 132.25, 130.39, 129.44, 126.78, 125.71, 124.01, 117.65, 113.47, 92.14.



3-Bromo-2-(4-bromophenyl)imidaz[1,2-a]pyridine (4g) (Salgado-Zamora et al., 2008).

White solid was obtained in 58% isolated yield. ¹H NMR (400 MHz, DMSO- d_6) δ 8.33 (d, J = 6.9 Hz, 1H), 8.04 – 8.02 (m, 2H), 7.68 – 7.63 (m, 3H), 7.40 – 7.36 (m, 1H), 7.08 (t, J = 6.8 Hz, 1H); ¹³C NMR (100 MHz, DMSO- d_6) δ 144.78, 140.15, 131.89, 131.34, 129.00, 125.86, 124.39, 121.40, 116.96, 113.57, 91.59.



3-Bromo-2-(4-(trifluoromethyl)phenyl)imidazo[1,2-a]pyridine (4h) (Zhou et al., 2016).

White solid was obtained in 67% isolated yield. ¹H NMR (400 MHz, DMSO- d_6) δ 8.36 – 8.34 (m, 1H), 8.27 (d, J = 8.1 Hz, 2H), 7.82 (d, J = 8.2 Hz, 2H), 7.68 – 7.65 (m, 1H), 7.41 – 7.37 (m, 1H), 7.11 – 7.07 (m, 1H); ¹³C NMR (100 MHz, DMSO- d_6) δ 145.03, 139.70, 136.79, 128.33 (q, J = 32 Hz), 127.73, 126.36, 125.48 (q, J = 3.7 Hz), 124.76, 124.35 (q, J = 271 Hz), 117.29, 113.99, 92.85; ¹⁹F NMR (376 MHz, CDCl₃) δ –62.54.



4-(3-Bromoimidazo[1,2-a]pyridin-2-yl)benzonitrile (4i).

White solid was obtained in 56% isolated yield. ¹H NMR (400 MHz, DMSO- d_6) δ 8.44 (d, J = 5.6 Hz, 1H), 8.30 (d, J = 7.1 Hz, 2H), 7.98 (d, J = 7.2 Hz, 2H), 7.71 (d, J = 8.8 Hz, 1H), 7.45 (t, J = 7.5 Hz, 1H), 7.15 (s, 1H); ¹³C NMR (100 MHz, DMSO- d_6) δ 144.98, 139.33, 137.15, 132.28, 127.55, 126.28, 124.56, 118.49, 117.15, 113.88, 110.42, 92.90. HRMS (ESI): m/z calcd for C₁₄H₉BrN₃ [M+H]⁺: 297.9974, found: 297.9967.



3-Bromo-2-(4-methoxyphenyl)imidazo[1,2-a]pyridine (4j) (Zhou et al., 2016).

White solid was obtained in 74% isolated yield. ¹H NMR (400 MHz, DMSO-*d*₆) δ 8.35 – 8.33 (m, 1H), 8.05 – 8.01 (m, 2H), 7.65 – 7.62 (m, 1H), 7.38 – 7.34 (m, 1H), 7.08 – 7.04 (m, 3H), 3.81 (s, 3H); ¹³C NMR (100 MHz, DMSO-*d*₆) δ 159.37, 144.76, 141.38, 128.70, 125.71, 125.19, 124.48, 116.85, 114.06, 113.48, 90.68, 55.25 (d, *J* = 7.3 Hz).



3-Bromo-2-(tert-butyl)imidazo[1,2-a]pyridine (4k) (Li et al., 2018).

Light yellow oil was obtained in 83% isolated yield. ¹H NMR (400 MHz, CDCl₃) δ 8.12 (d, J = 6.9 Hz, 1H), 7.60 (d, J = 9.0 Hz, 1H), 7.21 – 7.17 (m, 1H), 6.89 – 7.86 (m, 1H), 1.53 (s, 9H); ¹³C NMR (100 MHz, CDCl₃) δ 151.84, 143.75, 124.08, 123.22, 117.19, 112.59, 90.06, 33.06, 29.67.



3-Bromo-8-methyl-2-phenylimidazo[1,2-a]pyridine (4l) (Zhou et al., 2016).

Light yellow oil was obtained in 93% isolated yield. ¹H NMR (400 MHz, CDCl₃) δ 8.04 – 8.01 (m, 2H), 7.87 (d, *J* = 6.8 Hz, 1H), 7.37 (t, *J* = 7.6 Hz, 2H), 7.27 (t, *J* = 7.4 Hz, 1H), 6.89 – 6.87 (m, 1H), 6.66 (t, *J* = 6.9 Hz, 1H), 2.54 (s, 3H); ¹³C NMR (100 MHz, CDCl₃) δ 145.58, 142.00, 133.01, 128.32, 128.01, 127.92, 127.46, 123.70, 121.65, 112.86, 91.88, 16.47.



3-Bromo-6-methyl-2-phenylimidazo[1,2-a]pyridine (4m) (Zhou et al., 2016).

Light yellow oil was obtained in 83% isolated yield. ¹H NMR (400 MHz, DMSO- d_6) δ 8.16 (s, 1H), 8.09 – 8.07 (m, 2H), 7.56 (d, J = 9.2 Hz, 1H), 7.49 (t, J = 7.6 Hz, 2H), 7.39 (t, J = 7.4 Hz, 1H), 7.24 – 7.21 (m, 1H), 2.35 (s, 3H); ¹³C NMR (100 MHz, CDCl₃) δ 144.31, 142.11, 132.85, 128.32, 128.19, 128.04, 127.65, 122.79, 121.50, 116.73, 91.14, 18.24.



3-Bromoimidazo[1,2-a]pyridine (4n) (Pelleter et al., 2009).

Light yellow oil was obtained in 61% isolated yield. ¹H NMR (400 MHz, CDCl₃) δ 8.07–8.05 (m, 1H), 7.60 (d, *J* = 9.1 Hz, 1H), 7.56 (s, 1H), 7.21 – 7.17 (m, 1H), 6.90–6.86 (m, 1H); ¹³C NMR (100 MHz, CDCl₃) δ 145.55, 133.36, 124.19, 123.42, 117.68, 112.82, 94.46.



3-Bromo-6-chloro-2-phenylimidazo[1,2-a]pyridine (40).

White solid was obtained in 90% isolated yield. ¹H NMR (400 MHz, DMSO- d_6) δ 8.48 (d, J = 1.1 Hz, 1H), 8.07 (d, J = 7.6 Hz, 2H), 7.69 (d, J = 9.5 Hz, 1H), 7.50 (t, J = 7.5 Hz, 2H), 7.43 – 7.39 (m, 2H); ¹³C NMR (100 MHz, CDCl₃) δ 143.65, 143.48, 132.24, 128.51, 128.46, 127.73, 126.50, 121.84, 121.49, 117.86, 92.11. HRMS (ESI): m/z calcd for C₁₃H₉BrClN₂ [M+H]⁺: 306.9632, found: 306.9621.



3-Bromo-2-phenylbenzo[d]imidazo[2,1-b]thiazole (4p).

White solid was obtained in 44% isolated yield.¹H NMR (400 MHz, CDCl₃) δ 8.24 (d, *J* = 8.2 Hz, 1H), 7.92 (d, *J* = 7.0 Hz, 2H), 7.53 (d, *J* = 7.9 Hz, 1H), 7.35 (t, *J* = 7.6 Hz, 2H), 7.29 – 7.17 (m, 3H); ¹³C NMR (100 MHz, CDCl₃) δ 147.89, 143.89, 132.82, 132.53, 129.98, 128.33, 127.83, 127.07, 125.73, 125.09, 124.03, 113.51, 91.81.HRMS (ESI): m/z calcd for C₁₅H₁₀BrN₂S [M+H]⁺: 328.9743, found: 328.9757.



4-Bromo-1-phenyl-1H-pyrazole (4q) (Song et al., 2015).

White solid was obtained in 80% isolated yield. ¹H NMR (400 MHz, CDCl₃) δ 7.93 (s, 1H), 7.68 (s, 1H), 7.66 – 7.61 (m, 2H), 7.49 – 7.42 (m, 2H), 7.35 – 7.28 (m, 1H); ¹³C NMR (100 MHz, CDCl₃) δ 141.43, 139.52, 129.48, 126.98, 126.95, 118.93, 95.56.



4,5-Dibromo-3-phenyl-1H-pyrazole (4r) (Trofimenko et al., 2007).

White solid was obtained in 68% isolated yield. ¹H NMR (400 MHz, CDCl₃) δ 10.84 (s, 1H), 7.75 – 7.64 (m, 2H), 7.54 – 7.37 (m, 3H); ¹³C NMR (100 MHz, CDCl₃) δ 142.78, 129.65, 129.36, 128.94, 127.40, 127.25, 95.22.



5,7-Dibromoquinolin-8-amine (4s) (da Silva et al., 2007).

Yellow solid was obtained in 80% isolated yield. ¹H NMR (400 MHz, CDCl₃) δ 8.73 – 8.72 (m, 1H), 8.36 – 8.33 (m, 1H), 7.76 (s, 1H), 7.48-7.45 (m, 1H), 5.35 (s, 2H); ¹³C NMR (100 MHz, CDCl₃) δ 148.22, 141.93, 138.09, 135.66, 133.10, 126.61, 122.37, 106.75, 103.17.

5-Bromopyridin-2-amine (4t) (Li et al., 2013).

White solid was obtained in 60% isolated yield. ¹H NMR (400 MHz, DMSO- d_6) δ 7.98 – 7.90 (m, 1H), 7.50 – 7.47 (m, 1H), 6.47 – 6.37 (m, 1H), 6.14 (s, 2H); ¹³C NMR (100 MHz, CDCl₃) δ 156.96, 148.43, 140.15, 110.09, 108.16.



3-Bromo-5-chloropyridin-2-amine (4u).

Yellow solid was obtained in 49% isolated yield. ¹H NMR (400 MHz, CDCl₃) δ 7.95 (d, J = 2.2 Hz, 1H), 7.63 (d, J = 2.2 Hz, 1H), 5.19 (s, 2H); ¹³C NMR (100 MHz, CDCl₃) δ 154.16, 145.06, 139.42, 119.94, 103.95. HRMS (ESI): m/z calcd for C₅H₅BrClN₂ [M+H]⁺: 206.9319, found: 206.9330.

3-Bromo-5-(trifluoromethyl)pyridin-2-amine (4v).

White solid was obtained in 60% isolated yield. ¹H NMR (400 MHz, DMSO- d_6) δ 8.25 (d, J = 1.1 Hz, 1H), 8.00 (d, J = 2.0 Hz, 1H), 7.04 (s, 2H); ¹³C NMR (100 MHz, CDCl₃) δ 157.77, 144.58 (q, J = 4.0 Hz)), 144.56, 123.29 (q, J = 271.1 Hz), 117.71 (q, J = 33.6 Hz), 103.44; ¹⁹F NMR (376 MHz, CDCl₃) δ –61.18. HRMS (ESI): m/z calcd for C₆H₅BrF₃N₂ [M+H]⁺: 240.9583, found: 240.9584.



2-Bromo-4-chloroaniline (4w) (Li et al., 2013).

White solid was obtained in 59% isolated yield. ¹H NMR (400 MHz, CDCl₃) δ 7.36 (d, J = 2.3 Hz, 1H), 7.04 – 7.01 (m, 1H), 6.61 (d, J = 8.6 Hz, 1H), 4.01 (s, 2H); ¹³C NMR (100 MHz, CDCl₃) δ 142.73, 131.62, 128.18, 122.75, 116.09, 108.98.

2,4-Dibromoaniline (4x) (Li et al., 2013).

White solid was obtained in 57% isolated yield. ¹H NMR (400 MHz, CDCl₃) δ 7.49 (d, J = 2.1 Hz, 1H), 7.16 – 7.13 (m, 1H), 6.57 (d, J = 8.5 Hz, 1H), 3.99 (s, 2H); ¹³C NMR (100 MHz, CDCl₃) δ 143.11, 134.22, 130.98, 116.56, 109.40, 109.36.



2-Bromo-1,3,5-trimethoxybenzene (4y) (Tang et al., 2018).

White solid was obtained in 85% isolated yield. ¹H NMR (400 MHz, CDCl₃) δ 6.14 (s, 2H), 3.85 (s, 6H), 3.79 (s, 3H); ¹³C NMR (100 MHz, CDCl₃) δ 160.30, 157.24, 91.70, 91.44, 56.12, 55.31.



2-Bromo-1,5-dimethoxy-3-methylbenzene (4z) (Davis et al., 2000).

White solid was obtained in 83% isolated yield. ¹H NMR (400 MHz, CDCl₃) δ 6.41 (d, *J* = 2.7 Hz, 1H), 6.33 (d, *J* = 2.7 Hz, 1H), 3.84 (s, 3H), 3.77 (s, 3H), 2.38 (s, 3H); ¹³C NMR (100 MHz, CDCl₃) δ 159.24, 156.56, 139.68, 107.19, 105.03, 97.16, 56.14, 55.33, 23.43.



(1,2-Dibromoethyl)benzene (6a) (Martins et al., 2018).

White solid was obtained in 73% isolated yield.¹H NMR (400 MHz, CDCl₃) δ 7.42 – 7.30 (m, 5H), 5.15 – 5.11 (m, 1H), 4.08 – 3.98 (m, 2H); ¹³C NMR (100 MHz, CDCl₃) δ 138.53, 129.10, 128.78, 127.59, 50.86, 35.00.



1-(Tert-butyl)-4-(1,2-dibromoethyl)benzene (6b) (Martins et al., 2018).

Colorless oil was obtained in 50% isolated yield. ¹H NMR (400 MHz, CDCl₃) δ 7.43 – 7.37 (m, 2H), 7.36 – 7.32 (m, 2H), 5.19 – 5.15 (m, 1H), 4.13 – 3.99 (m, 2H), 1.33 (s, 9H); ¹³C NMR (100 MHz, CDCl₃) δ 152.26, 135.51, 127.26, 125.78, 51.23, 35.14, 34.69, 31.22.



1-(1,2-Dibromoethyl)-4-fluorobenzene (6c) (Wilson et al., 2018).

White solid was obtained in 74% isolated yield. ¹H NMR (400 MHz, CDCl₃) δ 7.48 – 7.31 (m, 2H), 7.15 – 7.02 (m, 2H), 5.16 – 5.12 (m, 1H), 4.10 – 4.05 (m, 1H), 4.01 – 3.95 (m, 1H); ¹³C NMR (100 MHz, CDCl₃) δ 162.77 (d, *J* = 249.1 Hz), 134.52 (d, *J* = 4Hz), 129.48 (d, *J* = 9 Hz), 115.85 (d, *J* = 22 Hz) 49.79, 34.97; ¹⁹F NMR (376 MHz, CDCl₃) δ -111.64.



1-Chloro-4-(1,2-dibromoethyl)benzene (6d) (Martins et al., 2018).

Yellow oil was obtained in 77% isolated yield. ¹H NMR (400 MHz, CDCl₃) δ 7.38 – 7.33 (m, 4H), 5.13 – 5.09 (m, 1H), 4.09 – 4.05 (m, 1H), 4.00 – 3.94 (m, 1H); ¹³C NMR (100 MHz, CDCl₃) δ 137.10, 134.89, 129.03, 128.98, 49.53, 34.65.



1-Bromo-4-(1,2-dibromoethyl)benzene (6e) (Karki et al., 2015).

Yellow oil was obtained in 80% isolated yield. ¹H NMR (400 MHz, CDCl₃) δ 7.57 – 7.48 (m, 2H), 7.35 – 7.23 (m, 2H), 5.12 – 5.08 (m, 1H), 4.08 – 4.04 (m, 1H), 3.97 (t, *J* = 10.6 Hz, 1H); ¹³C NMR (100 MHz, CDCl₃) δ 137.60, 132.00, 129.25, 123.10, 49.54, 34.57.



1-Bromo-3-(1,2-dibromoethyl)benzene (6f).

Yellow oil was obtained in 58% isolated yield. ¹H NMR (400 MHz, CDCl₃) δ 7.55 (t, J = 1.9 Hz, 1H), 7.48 – 7.46 (m, 1H), 7.34 – 7.31 (m, 1H), 7.25 – 7.22 (m, 1H), 5.07 – 5.03 (m, 1H), 4.06 – 4.02 (m, 1H), 3.95 (t, J = 10.6 Hz, 1H); ¹³C NMR (100 MHz, CDCl₃) δ 140.71, 132.18, 130.72, 130.28, 126.29, 122.65, 49.21, 34.55. HRMS (EI): m/z calcd for C₈H₇Br₃ [M]⁺: 339.8098, found: 339.8105.



1-Bromo-2-(1,2-dibromoethyl)benzene (6g).

Yellow oil was obtained in 57% isolated yield. ¹H NMR (400 MHz, CDCl₃) δ 7.61 – 7.59 (m, 1H), 7.56 – 7.53 (m, 1H), 7.41 – 7.37 (m, 1H), 7.22 – 7.18 (m, 1H), 5.74 – 5.70 (m, 1H), 4.10 – 4.07 (m, 2H); ¹³C NMR (100 MHz, CDCl₃) δ 137.53, 133.24, 130.29, 128.28, 128.13, 124.32, 48.32, 33.70. HRMS (EI): m/z calcd for C₈H₇Br₃ [M]⁺: 339.8098, found: 339.8094.



4-(1,2-Dibromoethyl)phenyl acetate (6h) (Rej et al., 2017).

White solid was obtained in 70% isolated yield. ¹H NMR (400 MHz, CDCl₃) δ 7.47 – 7.37 (m, 2H), 7.18 – 7.08 (m, 2H), 5.16 – 5.12 (m, 1H), 4.08 – 4.04 (m, 1H), 3.98 (t, *J* = 10.5 Hz, 1H), 2.29 (s, 3H); ¹³C NMR (100 MHz, CDCl₃) δ 168.97, 150.89, 135.99, 128.76, 121.87, 50.03, 34.95, 21.07.



1-(1,2-Dibromoethyl)-4-(trifluoromethyl)benzene (6i).

Yellow oil was obtained in 40% isolated yield. ¹H NMR (400 MHz, CDCl₃) δ 7.64 (d, *J* = 8.2 Hz, 2H), 7.51 (d, *J* = 8.1 Hz, 2H), 5.16 – 5.12 (m, 1H), 4.09 – 4.05 (m, 1H), 3.98 (t, *J* = 10.7 Hz, 1H); ¹³C NMR (100 MHz, CDCl₃) δ 142.45, 131.11 (q, *J* = 32.7 Hz), 128.13, 125.83 (q, *J* = 3.8 Hz), 123.73 (q, *J* = 270.6 Hz), 48.88, 34.28; ¹⁹F NMR (376 MHz, CDCl₃) δ –62.71. HRMS (EI): m/z calcd for C₉H₇Br₂F₃ [M]⁺: 329.8867, found: 329.8869.



1-(1,2-Dibromoethyl)-4-nitrobenzene (6j).

Colorless oil was obtained in 41% isolated yield. ¹H NMR (400 MHz, CDCl₃) δ 8.35 – 8.13 (m, 2H), 7.78 – 7.47 (m, 2H), 5.19 – 5.15 (m, 1H), 4.11 – 4.07 (m, 1H), 3.98 (t, *J* = 10.8 Hz, 1H); ¹³C NMR (100 MHz, CDCl₃) δ 148.01, 145.45, 128.76, 124.04, 47.77, 33.86. HRMS (ESI): m/z calcd for C₈H₇Br₂NNaO₂ [M+Na]⁺: 329.8736, found: 329.8732.



(1,2-Dibromopropyl)benzene (6k) (Kulangiappar et al., 2016).

White solid was obtained in 86% isolated yield. ¹H NMR (400 MHz, CDCl₃) δ 7.41 – 7.27 (m, 5H), 5.03 (d, J = 10.2 Hz, 1H), 4.63 – 4.55 (m, 1H), 2.03 (d, J = 6.5 Hz, 3H); ¹³C NMR (100MHz, CDCl₃) δ 140.46, 128.70, 128.55, 127.65, 59.11, 51.10, 25.75.



2-(1,2-Dibromoethyl)naphthalene (6l) (Song et al., 2015).

White solid was obtained in 43% isolated yield. ¹H NMR (400 MHz, CDCl₃) δ 7.98 – 7.81 (m, 4H), 7.55 – 7.51 (m, 3H), 5.37 – 5.33 (m, 1H), 4.19 – 4.14 (m, 2H); ¹³C NMR (100 MHz, CDCl₃) δ 135.68, 133.48, 132.88, 129.05, 128.16, 127.74, 127.39, 126.87, 126.65, 124.34, 51.34, 34.78.



(2,3-Dibromopropyl)benzene (6m) (Karki et al., 2015).

Yellow oil was obtained in 86% isolated yield. ¹H NMR (400 MHz, CDCl₃) δ 7.40 – 7.27 (m, 5H), 4.41 – 7.35 (m, 1H), 3.86 – 3.82 (m, 1H), 3.67 – 3.62 (m, 1H), 3.55 – 3.50 (m, 1H), 3.18 – 3.12 (m, 1H); ¹³C NMR (100 MHz, CDCl₃) δ 136.81, 129.46, 128.46, 127.16, 52.38, 41.97, 36.02.



(2,3-Dibromopropoxy)benzene (6n) (Song et al., 2015).

Light yellow oil was obtained in 38% isolated yield. ¹H NMR (400 MHz, CDCl₃) δ 7.43 – 7.28 (m, 2H), 7.07 – 7.00 (m, 1H), 7.00 – 6.93 (m, 2H), 4.48 – 4.42 (m, 1H), 4.40 – 4.35 (m, 2H), 4.02 – 3.87 (m, 2H); ¹³C NMR (100 MHz, CDCl₃) δ 157.90, 129.55, 121.62, 114.82, 69.03, 47.72, 32.74.



2-(3,4-Dibromobutyl)isoindoline-1,3-dione (60).

Colorless oil was obtained in 70% isolated yield. ¹H NMR (400 MHz, DMSO- d_6) δ 7.84 – 7.79 (m, 4H), 4.94 – 4.43 (m, 1H), 4.00 – 3.96 (m, 1H), 3.92 – 3.88 (m, 1H), 3.80 – 3.71 (m, 2H), 2.42 – 4.34 (m, 1H), 2.20 – 2.01 (m, 1H). ¹³C NMR (100 MHz, DMSO- d_6) δ 167.83, 134.33, 131.67, 123.00, 51.56, 38.45, 35.61, 34.99. HRMS (ESI): m/z calcd for C₁₂H₁₁Br₂NNaO₂ [M+Na]⁺: 381.9049, found: 381.9048.



(2,3-Dibromopropyl)cyclopentane (6p).

Colorless oil was obtained in 77% isolated yield. ¹H NMR (400 MHz, CDCl₃) δ 4.18 – 4.11 (m, 1H), 3.92 – 3.80 (m, 1H), 3.64 – 3.59 (m, 1H), 2.20 – 1.96 (m, 2H), 1.95 – 1.73 (m, 3H), 1.73 – 1.46 (m, 4H), 1.31 – 1.12 (m, 1H), 1.10 – 1.01 (m, 1H); ¹³C NMR (100 MHz, CDCl₃) δ 52.70, 42.68, 38.00, 37.10, 32.80, 31.44, 25.03, 24.97. HRMS (EI): m/z calcd for C₈H₁₄Br₂ [M]⁺: 267.9462, found: 267.9452.

(2,3-Dibromopropyl)cyclohexane (6q).

Colorless oil was obtained in 85% isolated yield. ¹H NMR (400 MHz, CDCl₃) δ 4.31 – 4.13 (m, 1H), 3.8 – 3.84 (m, 1H), 3.70 – 3.49 (m, 1H), 2.01 – 1.94 (m, 1H), 1.85 – 1.49 (m, 7H), 1.31 – 1.15 (m, 3H), 1.09 – 0.95 (m, 1H), 0.93 – 0.77 (m, 1H); ¹³C NMR (100 MHz, CDCl₃) δ 50.99, 43.95, 37.21, 35.52, 33.77, 31.48, 26.40, 26.12, 25.83. HRMS (EI): m/z calcd for C₉H₁₆Br₂ [M]⁺: 281.9619, found: 281.9622.



2,3-Dibromooctane (6r) (Badetti et al., 2016).

Colorless oil was obtained in 80% isolated yield. ¹H NMR (400 MHz, CDCl3) δ 4.47 – 4.41 (m, 1H), 4.25 – 4.18 (m, 1H), 2.08 – 2.01 (m, 1H), 1.77 (d, *J* = 6.7 Hz, 3H), 1.66 – 1.58 (m, 1H), 1.43 – 1.28 (m, 5H), 0.98 – 0.82 (m, 4H); ¹³C NMR (100 MHz, CDCl₃) δ 60.11, 52.37, 33.89, 30.95, 27.44, 22.42, 21.52, 13.95.



3,4-Dibromooctane (6s) (Conte et al., 1994).

Colorless oil was obtained in 71% isolated yield. ¹H NMR (400 MHz, CDCl₃) δ 4.19 – 4.10 (m, 2H), 2.22 – 2.09 (m, 2H), 2.04 – 1.90 (m, 2H), 1.65 – 1.54 (m, 1H), 1.49 – 1.26 (m, 3H), 1.08 (t, *J* = 7.2 Hz, 3H), 0.93 (t, *J* = 7.2 Hz, 3H); ¹³C NMR (100 MHz, CDCl₃) δ 61.53, 59.40, 36.69, 30.23, 29.00, 22.01, 13.89, 11.35.



1,2-Dibromodecane (6t) (Song et al., 2015).

Colorless oil was obtained in 89% isolated yield. ¹H NMR (400 MHz, CDCl₃) δ 4.22 – 4.12 (m, 1H), 3.87 – 3.83 (m, 1H), 3.63 (t, *J* = 10.0 Hz, 1H), 2.20 – 2.05 (m, 1H), 1.83 – 1.73 (m, 1H), 1.60 – 1.53 (m, 1H), 1.49 – 1.24 (m, 11H), 0.92 – 0.83 (m, 3H); ¹³C NMR (100 MHz, CDCl₃) δ 53.12, 36.33, 36.02, 31.81, 29.33, 29.17, 28.80, 26.73, 22.63, 14.08.



1,2-Dibromocyclododecane (6u).

Light yellow oil was obtained in 73% isolated yield. ¹H NMR (400 MHz, CDCl₃) δ 4.37 – 4.30 (m, 2H), 2.22 – 2.14 (m, 2H), 2.08 – 1.93 (m, 2H), 1.45 – 1.27 (m, 16H); ¹³C NMR (100 MHz, CDCl₃) δ 55.95, 36.10, 24.63, 23.29, 23.08, 22.53. HRMS (EI): m/z calcd for C₁₂H₂₂Br₂ [M]⁺: 324.0088, found: 324.0093.

1,2-Dibromooctane (6v) (Martins et al., 2018).

Colorless oil was obtained in 83% isolated yield; ¹H NMR (400 MHz, CDCl₃) δ 4.24 – 4.10 (m, 1H), 3.87 – 3.82 (m, 1H), 3.63 (t, *J* = 10.0 Hz, 1H), 2.18 – 2.09 (m, 1H), 1.88 – 1.69 (m, 1H), 1.64 – 1.47 (m, 1H), 1.48 – 1.27 (m, 7H), 0.95 – 0.82 (m, 3H); ¹³C NMR (100 MHz, CDCl₃) δ 53.13, 36.34, 36.03, 31.56, 28.46, 26.70, 22.53, 14.02.



1,2,8,9-Tetrabromononane (6w).

Colorless oil was obtained in 89% isolated yield. ¹H NMR (400 MHz, CDCl₃) δ 4.23 – 4.08 (m, 2H), 3.86 – 3.81 (m, 2H), 3.62 (t, *J* = 10.0 Hz, 2H), 2.21 – 2.05 (m, 2H), 1.88 – 1.69 (m, 2H), 1.69 – 1.52 (m, 2H), 1.51 – 1.28 (m, 4H); ¹³C NMR (100 MHz, CDCl₃) δ 52.75, 52.73, 36.19, 35.74, 35.70, 27.93, 27.85, 26.41, 26.37. HRMS (ESI): m/z calcd for C₉H₁₆Br₃ [M-Br]⁺: 360.8802, found: 360.8803.

(2-Bromoethene-1,1-diyl)dibenzene (6x) (Bi et al., 2017).

Light yellow oil was obtained in 95% isolated yield. ¹H NMR (400 MHz, CDCl₃) δ 7.41 – 7.32 (m, 3H), 7.31 – 7.23 (m, 5H), 7.22 – 7.16 (m, 2H), 6.74 (s, 1H); ¹³C NMR (100 MHz, CDCl₃) δ 146.78, 140.65, 139.02, 129.61, 128.37, 128.17, 128.05, 127.91, 127.56, 105.16.



(E)-1-(1,2-dibromovinyl)-4-methoxybenzene (8a) (Song and Li et al., 2015).

Colorless oil was obtained in 65% isolated yield. ¹H NMR (400 MHz, CDCl₃) δ 7.41 (d, J = 8.9 Hz, 2H), 6.82 (d, J = 8.9 Hz, 2H), 6.66 (s, 1H), 3.75 (s, 3H); ¹³C NMR (100 MHz, CDCl₃) δ 160.11, 130.76, 129.09, 121.44, 113.51, 101.92, 55.30.



(E)-(1,2-dibromoprop-1-en-1-yl)benzene (8b). (Kikushima et al., 2010).

Colorless oil was obtained in 33% isolated yield. ¹H NMR (400 MHz, CDCl₃) δ 7.40 – 7.30 (m, 5H), 2.62 (s, 3H); ¹³C NMR (100 MHz, CDCl₃) δ 140.75, 129.07, 128.58, 128.21, 117.22, 116.77, 29.31.



Ethyl 2-bromo-2-(pyridin-2-yl)acetate (10).

Light yellow oil was obtained in 54% isolated yield. ¹H NMR (400 MHz, CDCl₃) δ 8.64 – 8.45 (m, 1H), 7.82 – 7.56 (m, 2H), 7.24 – 7.21 (m, 1H), 5.49 (s, 1H), 4.30 – 4.17 (m, 2H), 1.25 (t, *J* = 7.1 Hz, 3H); ¹³C NMR (100 MHz, CDCl₃) δ 167.44, 155.16, 148.93, 137.12, 123.46, 123.41, 62.50, 47.43, 13.74. HRMS (ESI): m/z calcd for C₉H₁₀BrNNaO₂ [M+Na]⁺: 265.9787, found: 265.9788.



Ethyl 2,2-dibromo-2-(pyridin-2-yl)acetate (11).

Light yellow oil was obtained in 40% isolated yield. ¹H NMR (400 MHz, CDCl₃) δ 8.57 – 8.43 (m, 1H), 8.02 – 7.99 (m, 1H), 7.81 – 7.76 (m, 1H), 7.24 –7.20 (m, 1H), 4.44 – 4.29 (m, 2H), 1.30 – 1.25 (m, 3H); ¹³C NMR (100 MHz, CDCl₃) δ 165.41, 158.39, 148.05, 137.44, 123.66, 121.80,

64.24, 58.61, 13.67. HRMS (ESI): m/z calcd for $C_9H_9Br_2NNaO_2$ [M+Na]⁺: 343.8892, found: 343.8889.

Br

1-(3-Bromoprop-1-en-2-yl)-4-chlorobenzene (13) (Gonzalez-de-Castro et al., 2015). Colorless oil was obtained in 32% isolated yield.¹H NMR (400 MHz, CDCl₃) δ 7.43 (d, J = 8.6 Hz, 2H), 7.35 (d, J = 8.7 Hz, 2H), 5.54 (s, 1H), 5.50 (s, 1H), 4.35 (s, 2H); ¹³C NMR (100 MHz, CDCl₃) δ 143.17, 135.96, 134.13, 128.67, 127.42, 117.60, 33.84.

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