

# Virtual Double-System Single-Box for Absolute Dissociation Free Energy Calculations in GROMACS

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Cite This: *J. Chem. Inf. Model.* 2021, 61, 5320–5326



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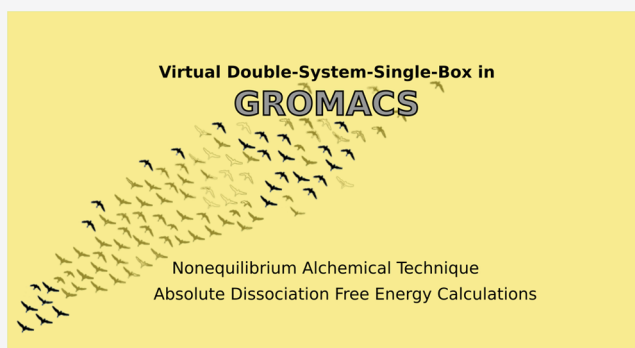


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**ABSTRACT:** We describe a step-by-step protocol for the computation of absolute dissociation free energy with GROMACS code and PLUMED library, which exploits a combination of advanced sampling techniques and nonequilibrium alchemical methodologies. The computational protocol has been automated through an open source Python middleware (HPC\_Drug) which allows one to set up the GROMACS/PLUMED input files for execution on high performing computing facilities. The proposed protocol, by exploiting its inherent parallelism and the power of the GROMACS code on graphical processing units, has the potential to afford accurate and precise estimates of the dissociation constants in drug-receptor systems described at the atomistic level. The procedure has been applied to the calculation of the absolute dissociation free energy of PF-07321332, an oral antiviral proposed by Pfizer, with the main protease (3CL<sup>pro</sup>) of SARS-CoV-2.



## 1. INTRODUCTION

In the last decades various computational methodologies, based on molecular dynamics (MD) simulations with explicit solvent, have been devised to improve (beyond the traditional docking approach) the calculation of absolute dissociation free energy in drug receptor systems. In this context, the so-called alchemical approach<sup>1</sup> emerged recently as one of the best-automated and most widely used MD-based free energy methods. The alchemical protocol relies on a stratification of nonphysical intermediate states gradually connecting the ligand-environment potential energy function of the target end states,<sup>2</sup> recovering the corresponding free energy change by a sum of free energy contributions evaluated along the stratification using free energy perturbation (FEP).<sup>3</sup> Alchemical FEP, due to well known sampling issues especially at low ligand-environment coupling,<sup>4</sup> is generally applied in industrial applications to the calculation of *relative* binding free energies (RBFE),<sup>5–8</sup> whereby a ligand is *transmuted* into a strictly congeneric compound.

As recently noted,<sup>8,9</sup> a reliable estimate of the *absolute* dissociation free energy (ADFE) via MD is a key requirement for virtual screening campaigns in industrial and academic settings in order to evaluate distant hits that are not easily amenable for RBFE calculations. Here, we present the fine-tuning of nonequilibrium (NE) alchemical transformations for ADFE calculations using the program GROMACS<sup>10</sup> on high performing computing (HPC) platforms. The approach is called NEW-vDSSB. The acronym NEW<sup>2</sup> stands for Non-Equilibrium Work. The work histograms, obtained by a swarm

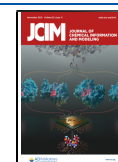
of rapid and independent NE alchemical trajectories connecting the target thermodynamic end states, are related to the corresponding free energy via the Jarzynski<sup>11</sup> and Crooks<sup>12</sup> theorems. vDSSB stands for *virtual* double-system single-box, a technique recently developed by some of us<sup>13–15</sup> as a variant of the so-called double-system single-box (DSSB) method.<sup>16,17</sup>

NEW-vDSSB has been first implemented in the ORAC molecular dynamics package<sup>18</sup> and has been successfully applied, for example, to the calculation of absolute binding free energy (ABFE) of ligands of the SARS-CoV-2 main protease.<sup>13</sup> In this note, we describe an automated protocol, managed by a Python middleware (HPC\_Drug), for implementing the NEW-vDSSB method on heterogeneous (GPU/CPU) HPC facilities using the GROMACS popular molecular dynamics package<sup>10,19,20</sup> and the PLUMED library,<sup>21,22</sup> requiring no intervention on the GROMACS source code.

The paper is organized as follows: we first provide some basic information on the fast switching alchemical technique employed for ADFE calculations. We then describe the

Received: July 28, 2021

Published: November 1, 2021



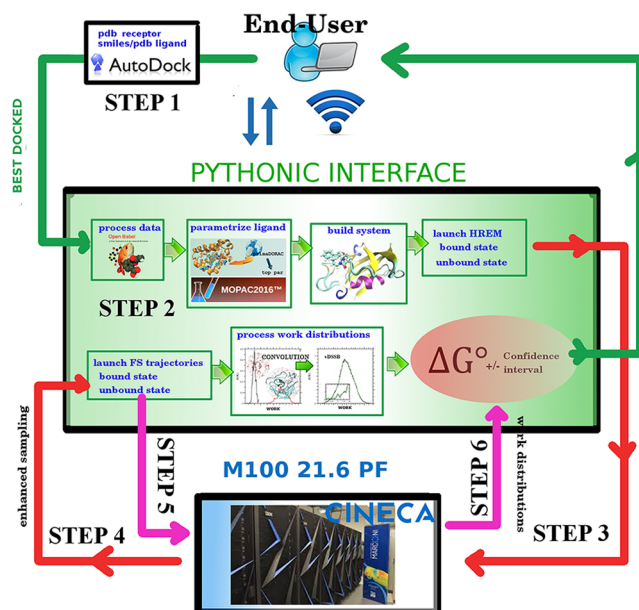
computational steps to use NEW-vDSSB methodology with GROMACS. For each step, we make available the corresponding input files locally produced by the automated HPC\_Drug Python middleware for execution on the remote HPC facility. In the last section, as a real-world example, we have calculated on the CINECA<sup>23</sup> Marconi100 CPU-GPU heterogeneous platform the ADFE of PF-07321332 against the SARS-CoV-2 main protease (3CL<sup>pro</sup>). PF-07321332 is a 3CL<sup>pro</sup> inhibitor proposed by Pfizer and exhibiting nanomolar affinity and capable of suppressing virus replication in human cells at submicromolar concentrations.<sup>24</sup>

## 2. BACKGROUND

Our protocol for ADFE calculation is based on the vDSSB approach. In brief, vDSSB is a NE variant of the alchemical technique whereby the ADFE is computed in a thermodynamic cycle as the difference of the ligand solvation free energy in the bulk solvent and in the solvated complex. vDSSB relies on the *enhanced sampling* of the unbound (decoupled ligand in bulk) and bound (coupled protein-bound ligand) end states, followed by the NE step, consisting of the production of two swarms of NE-independent trajectories where the ligand is gradually recoupled (unbound leg) and decoupled (bound leg) via a driven alchemical parameter  $\lambda$ , producing two samples of  $n_u$  growth and  $n_b$  annihilation work values. The  $n_b$  and  $n_u$  work values are combined as two independent random variables obtaining  $n_b \times n_u$  work values, referring to the process of bringing the ligand *from* the bound state *to* the bulk solvent. The convolution of the two work histograms allows us to increase significantly the resolution of the resulting work histogram with benefit for accuracy and precision of the ADFE estimate. At variance with DSSB, in vDSSB, different protocols for the bound and the unbound state can be used without violation of the Crooks<sup>12</sup> and Jarzynski<sup>11</sup> theorems, hence allowing us to choose the optimal box size and time protocol according to the physical dimension of the solute and the nature of the environment. As such, vDSSB is an inherently parallel algorithm, affording very efficient performances on massively parallel facilities. The free energy and the corresponding confidence interval are recovered, in a fast postprocessing stage, by constructing the convolution of the recoupling (unbound leg) and decoupling (bound leg) work histograms to which the Jarzynski theorem is applied and by correcting for a standard state-dependent term related to the translational binding site volume. For further details on vDSSB theory and technicalities, we refer to refs 13, 15, and 25. vDSSB, as any other MD-based technology for ADFE calculations, requires expertise and knowledge, which may deter the average end user. In the following, we describe a step-by-step protocol where the most complex tasks in the vDSSB workflow have been automated by an *ad hoc* devised Python middleware.

## 3. ADFE CALCULATION WORKFLOW

The vDSSB procedure for the HPC calculation of ADFEs (schematically described in Figure 1) is made up of six consecutive steps, starting from the 3D structure of receptor and ligand. In the following, we provide an overview of the vDSSB steps to arrive at the ADFE estimate. A companion step-by-step technical tutorial for the calculation of the ADFE in the PF-07321332–3CL<sup>pro</sup> system is provided in the Supporting Information (SI). The step-by-step tutorial can



**Figure 1.** HPC\_Drug/vDSSB layout for ADFE calculations: Dark green lines define end user minimal tasks. Light green arrows define tasks and data flow within the Python HPC\_Drug automaton. Red (HREM) and magenta lines (FSDAM) represent connections and data flow between the Python library and the underlying HPC.

be also accessed at the [https://procacci.github.io/vdssb\\_gromacs/](https://procacci.github.io/vdssb_gromacs/) site.

**3.1. Step 1: Docking (Local).** Assuming that the experimental cocrystal structure of the complex structure is not available, we generate the initial bound state pose by way of a preliminary docking calculation of the ligand–receptor system using Autodock4.<sup>26</sup> Starting from the receptor pdb code and the ligand SMILES or pdb structure, the docking step can be completed in few minutes for a typical target of 5000 atoms on a local workstation using the application script `docking.bash` for Unix platforms providing a convenient interface for Autodock4. The script is included in the SI with details on docking parameters. The lowest energy docked conformation is the “best-docked” bound state conformation for following vDSSB steps (used as complex with receptor, `complex_ligand.pdb`).

**3.2. Step 2: HPC\_Drug for HREM setup (Local).** Starting from the “best-docked” conformation generated in step 1 (`complex_ligand.pdb`), we produce the GROMACS input files to perform Hamiltonian Replica Exchange (HREM) MD simulation for HPC execution of the target bound and unbound states. vDSSB HREM for the bound state relies on a definition of the “hot zone” comprising the ligand and nearby residues where the enhanced sampling is implemented. The hot zone for the unbound state is the ligand itself. These complex steps, which are central in the vDSSB approach, are assisted by the HPC\_Drug middleware,<sup>27</sup> an effective tool for the guided submission of vDSSB job on a HPC system.

HPC\_Drug is a Python application distributed under the AGPL that can be cloned from the github repository [https://github.com/MauriceKarrenbrock/HPC\\_Drug](https://github.com/MauriceKarrenbrock/HPC_Drug). All installation instructions are detailed at the GitHub site. HPC\_Drug reads a user-defined input file with minimal information (receptor–ligand pdb (`complex_ligand.pdb`) file, force field, water model, pH, etc.). For complex operations (e.g., replica exchange setup and NE input files productions), HPC\_Drug has a solid default

behavior, which can be changed by the end user in the main HPC\_Drug input file, if necessary. Once started, HPC\_Drug performs, with no further user intervention, a series of operations consisting of (i) the generation of the GROMACS topology files using PrimaDORAC<sup>28</sup> as the default tool for the ligand force field parametrization, (ii) a preliminary minimization of the complex with the user-selected force field, (iii) the NPT equilibration in standard conditions of the resulting solvated receptor–ligand structure (using as default the TIP3P<sup>29</sup> model for water or a user-selected model) in a optimal MD box, and (iv) the production of the input files (ready for use) to perform HREM simulations for bound and unbound ligand state on the HPC platforms. The GROMACS HREM input files automatically define the hot zone for the bound state based on ligand–residue distance criteria.<sup>30</sup> The hot zone is implemented generating multiple topology files along the replica progression via PLUMED,<sup>21,22</sup> a community-wide open source library for enhanced sampling techniques and free energy methods. PLUMED is used by HPC\_Drug as a standalone tool. For HPC execution, GROMACS needs to be patched with PLUMED. Most of HPC systems for scientific applications provide modules for executing jobs with GROMACS/PLUMED.

HPC\_Drug produces the mdp GROMACS main input file for HREM execution of the bound state with a series of “pulling” directives whereby the ligand is tethered to the receptor via a weak harmonic potential acting on the centers of mass (COM) of the partners. These directives generate output files, during the HREM simulation step, containing information about the COM–COM distance distributions, allowing us to calculate the binding site volume correction for the dissociation free energy.<sup>13,15</sup>

The final output of HPC\_Drug (produced in few minutes on a local workstation for a typical system with 50,000 atoms) consists of two directories, one for the bound state and the other for the gas-phase unbound state, where all necessary data for HREM execution on a HPC (including examples of batch submission scripts) are stored. As previously stated, some advanced options (like the number of HREM replicates, number of replica, and HREM simulation time) can be changed in the input file. An example of HPC\_Drug input file is reported in the SI.

**3.3. Step 3: HREM Simulations (HPC).** Upon uploading on the remote HPC platform the two HPC\_Drug generated HREM directories, the HREM simulations can be straightforwardly executed on the HPC system. In the SI, we included the batch script used to run the bound state HREM simulation on the heterogeneous Marconi100 HPC platform (CINECA), equipped with 4 Nvidia VOLTA GPU per node. In particular we ran six batteries (replicates) each of 24 replica on 36 nodes (144 Nvidia VOLTA GPU). For the unbound state, we perform the HREM for a ligand molecule in gas phase; we ran four batteries each of eight replicas on eight nodes (32 Nvidia VOLTA GPU).

**3.4. Step 4: Selection of (Enhanced Sampled) Configurations.** This postprocessing operation is conveniently done on the HPC system. A preliminary analysis of the “pulling” output files in target state replicas might be needed to verify that the ligand did not leave the binding site despite the weak ligand–receptor tethering restraint. No less than 200 configurations files are extracted using the GROMACS tools from the target state HREM trajectories xtc files for the bound and unbound states. Simple bash scripts to harvest

configurations at regular intervals are provided in the SI. For the unbound state, the gas-phase HREM configurations of the ligand (gro files) are combined with a well-equilibrated box of water molecules.

### 3.5. Step 5: Fast Switching Alchemical Simulations (HPC).

**3.5.1. Bound State.** The ligand decoupling in the bound state, starting from the HREM-generated pdb configurations, is straightforwardly implemented via the multidir GROMACS option whereby the independent alchemical trajectories are originated from the HREM-generated pdb configurations in dedicated directories. In the HPC submission script for Marconi100 (see the SI), each independent trajectory on the MPI layer is assigned to a GPU, in a very effective hybrid parallelization scheme. The resulting embarrassingly parallel run engages as many GPUs (MPI processes) as HREM-generated pdb files. According to a consensus protocol,<sup>2</sup> the alchemical decoupling, for each independent trajectory, is done in two consecutive annihilation steps and hence using two GROMACS mdp files (provided in the SI). In the first and second steps, we switch off the electrostatic ligand–environment interactions and the Lennard–Jones (LJ) potential, respectively, using a soft-core regularization.<sup>31</sup> Separate estimates of the electrostatic and LJ contributions to the dissociation free energy are useful to gain hints on how to modify the molecule to increase the affinity. Typical annihilation times depends on the ligand size. For a typical drug size ligand (300:400 MW), we use 0.375 ns for the switching off the atomic charges and 0.750 ns for turning off the LJ ligand–environment interactions. Intra-molecular interactions are always on during the alchemical process. An example of the batch script for submitting the fast-switching parallel job for the bound state is provided in the SI.

**3.5.2. Unbound State.** The fast switching growth run is technically similar to the bound state run. In each fast-growth trajectory (MPI process), we switch on the LJ and electrostatic potential in sequence in a total time (for a typical ligand) of 0.520 ns. Further details can be found in the SI.

### 3.6. Step 6: Calculation of Dissociation Free Energy.

The main output in the preceding step 5 are the dhdl.xvg files, stored in the directories specified in the multidir option. In these files, the instantaneous values of the derivative of the Hamiltonian with respect to  $\lambda$  at each time step are recorded. For each alchemical trajectory, the work can be computed by numerical integration. This operation is done for the two contributions, electrostatic and LJ, and for the bound and unbound transformations. The resulting  $n_b$  and  $n_u$  total work values, referring to the bound and unbound work histograms,  $P_b(W)$ ,  $P_u(W)$ , can be combined yielding  $n_b \times n_u$  work data as  $W_{ij} = W_i^b + W_j^u$  in the convolution  $(P_b \times P_u)(W) = \int dw P_b(W)P_u(W - w)$ , affording the Jarzynski estimates of the free energy cost of transferring the ligand from the bound state to the bulk (dissociation free energy)

$$\Delta G_{\text{vDSSB}} = -\beta^{-1} \ln \left( \frac{1}{n_b n_u} \sum_i \sum_j^{n_b \times n_u} e^{-\beta W_{ij}} \right) \quad (1)$$

The confidence interval is evaluated by bootstrapping with resampling the bound and unbound work data before the combination of work values.

The free energy estimate can also be computed assuming that the convolution  $(P_b \times P_u)(W)$  can be represented as a Gaussian mixture using

$$\Delta G_{EM} = -RT \ln \left[ \sum_i^c w_i n(W, \mu_i, \sigma_i^2) \right] \quad (2)$$

where  $c$  is the number of components in the mixture,  $n(W, \mu_i, \sigma_i^2)$  is the normal distribution of the  $i$ th component with mean and variance  $\mu_i, \sigma_i^2$  and  $w_i$  are the  $i$ th weights such that  $\sum_i^c w_i = 1$ . The parameters in eq 2 are determined from the  $n_b \times n_u$  work data using the Expectation Maximization (EM) algorithm.<sup>32,33</sup> Equation 2 is a direct consequence of the Crooks theorem, providing an unbiased alternative estimate of the dissociation free energy if the distribution is the result of a combination of normal distributions. While  $\Delta G_{EM}$  does not suffer of the inherent<sup>34</sup> bias of the Jarzynski estimate, eq 2 can be less precise as the EM fitting process can be ill-defined and unstable if the mixture is made by overlapping components with disparate variances. A discussion about Jarzynski and EM estimates and their differences can be found in refs 13 and 15.

All these postprocessing calculations can be performed directly on the HPC using a simple script file (works.bash) detailed in the SI.

Finally, the standard dissociation free energy can be calculated by summing to  $\Delta G_{vDSSB}$  the volume correction and the finite-size correction, when a ligand is not electrically neutral.<sup>35,36</sup> The volume correction, evaluated from the variance of COM–COM distribution, is calculated as

$$\Delta G_{vol} = RT \ln(V_{site}/V_0) \quad (3)$$

with  $V_0 = 1661 \text{ \AA}^3$  and  $V_{site} = \frac{4}{3}\pi(2\sigma)^3$ , where  $\sigma^2$  is the variance of the COM–COM distribution in the HREM stage obtained starting from pullx.vvg files. The finite-size correction, necessary only in case of charged ligands, is calculated applying the following equation

$$\Delta G_{charge} = -\frac{\pi}{2\alpha^2} \left\{ \frac{[Q_p^2 - (Q_p + Q_L)^2]}{V_{BOX}^{(b)}} + \frac{Q_L^2}{V_{BOX}^{(u)}} \right\} \quad (4)$$

where  $Q_p$  and  $Q_L$  are the net charge on the receptor and ligand, respectively,  $V_{BOX}^{(b/u)}$  are the MD box volumes of the bound and unbound states, and  $\alpha$  is the Ewald convergence parameter. Summarizing, the vDSSB estimate of the standard dissociation energy is given by

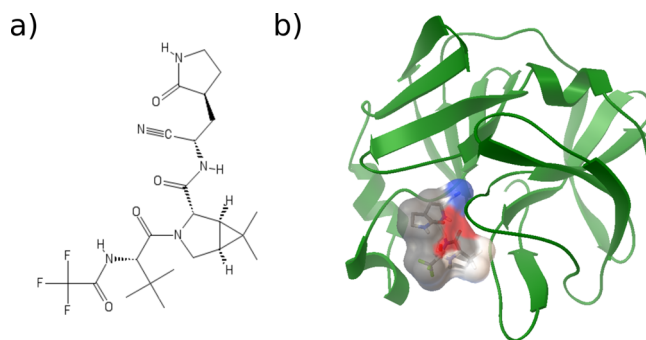
$$\Delta G_0 = \Delta G_{vDSSB} + \Delta G_{vol} + \Delta G_{charge} \quad (5)$$

For further technical details on the postprocessing stage for ADfE calculations, see the SI.

#### 4. APPLICATION

During the Covid-19 pandemic, considerable efforts have been devoted to the identification of an effective antiviral agent for SARS-CoV-2 via computational approaches.<sup>37</sup> Here, we report on the calculation of the dissociation affinity of the PF-07321332 compound (structure in Figure 2a) against the main protease (3CL<sup>pro</sup>) of SARS-CoV-2 using the vDSSB technique.

Starting from the protein X-ray structure<sup>38</sup> and the PubChem<sup>39</sup> generated SMILES code of PF-07321332, we performed the docking calculation on the catalytic domains I + II of 3CL<sup>pro</sup> obtaining as the best docking pose with a binding energy value of  $-9.07 \text{ kcal/mol}$  the structure of the ligand–receptor complex reported in Figure 2b (step 1).

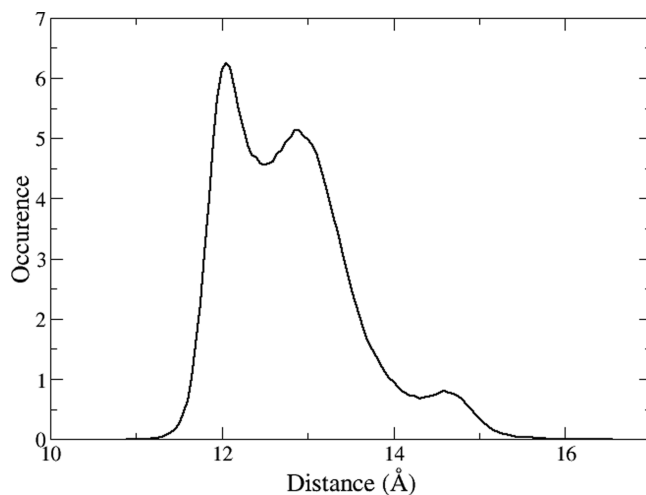


**Figure 2.** (a) Structure of PF-07321332. (b) Ligand–receptor complex with lowest docking binding free energy, “best-docked”. The catalytic dyad (HIS41-CYS145) has been considered in the neutral state.<sup>13</sup> The surface shows the interactions between PF-07321332 and 3CL<sup>pro</sup>.

Using this starting configuration, the Python application generates the two directories to perform HREM of bound and unbound states (step 2).

The two HPC\_Drug-generated directories are then transferred to the HPC system (CINECA Marconi100). On the HPC, the bound HREM was carried out using six batteries and 24 replicas on 144 GPUs collecting about 142 ns on the target state in 24 h, while for the unbound HREM we launched four batteries and eight replicas on 32 GPUs, collecting 32 ns in about 2 h (step 3).

From the “pulling” directive output in the target states of the bound HREM directory, we computed the COM receptor–ligand distribution (Figure 3) to obtain the volume correction



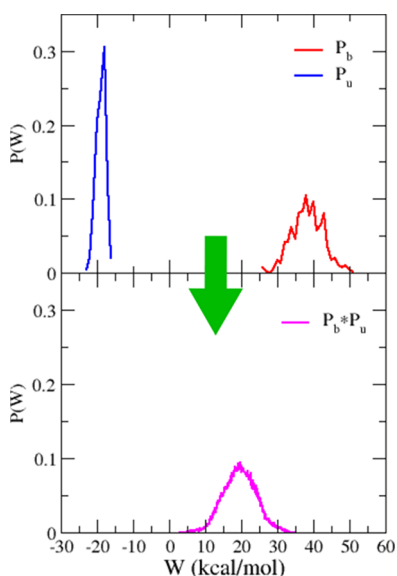
**Figure 3.** COM–COM distribution obtained from bound state HREM trajectories.

to the dissociation free energy. From the GROMACS xtc files in the bound HREM, we extracted 384 configurations of the target state sampled at regular intervals as starting points for the subsequent NE simulations corresponding to the fast annihilation of the ligand in the bound state. From the gas-phase HREM target state trajectories, we extracted 192 configurations of the ligand. These conformations were combined with a well-equilibrated water box to provide the starting structures for the ligand growth NE process in the solvent (step 4).

The fast switching stage produced 384 NE bound state trajectories lasting  $\approx 1.2$  ns and engaging as many GPUs in about 2 h and 192 NE 0.520 ns unbound state trajectories on a many GPUs in about 20 min (step 5).

The dhdl files produced in step 5 were finally postprocessed on the HPC using the work.bash script obtaining the fast switching work values for bound (annihilation) and unbound (growth) states along with the estimates of the dissociation free energy  $\Delta G_{\text{vdSSB}}$  and  $\Delta G_{\text{EM}}$  in eqs 1 and 2, respectively (step 6).

In Figure 4, we report the work distributions for bound and unbound state obtained and the convolution calculated by



**Figure 4.** Upper panel:  $P_u(W)$ ,  $P_b(W)$  work histograms obtained from NE decoupling (bound) and recoupling (unbound) runs. Lower panel:  $(P_b \times P_u)(W)$  convolution process.

combining the bound and unbound work samples. The correction volume, obtained applying eq 3 and analyzing the Figure 3, is of  $-2.81$  kcal/mol. The finite-charge correction is null since PF-07321332 is a neutral molecule. The final dissociation free energy estimate ( $\Delta G_{\text{vdSSB}}$ , eq 1) is  $5.8 \pm 0.9$  kcal/mol.

## 5. CONCLUSIONS

We have described a step-by-step protocol for the calculations of the absolute dissociation free energy calculation using the GROMACS code and the PLUMED library, relying on the end states enhanced sampling with HREM followed by the production of concurrent non equilibrium fast switching alchemical trajectories. The approach emulates a double-system single-box ligand–receptor dissociation process by effectively combining the ligand annihilation work in the bound state with the ligand growth work in the bulk solvent. The methodology, specifically tailored for massively parallel architectures, allows us to obtain a reliable estimate of the ligand–receptor ADFE on GPU-accelerated HPC architectures in about 24 h of wall clock. HPC submission of the complex HREM computational stage with GROMACS is greatly facilitated by the Python middleware HPC\_Drug, freely available at the GitHub site. A detailed tutorial is provided in the Supporting Information where the noncovalent dissociation

affinity of the PF-07321332 inhibitor of the SARS-CoV-2 main protease is evaluated.

## 6. DATA AND SOFTWARE AVAILABILITY

HPC\_Drug is publicly available at [https://github.com/MauriceKarrenbrock/HPC\\_Drug](https://github.com/MauriceKarrenbrock/HPC_Drug) under a AGPL license. The simulations have been performed using GROMACS (<https://www.gromacs.org/>) and the PLUMED library (<https://www.plumed.org/>). Autodock4 (<http://autodock.scripps.edu/>) has been adopted for preliminary docking calculations. ORAC software (<http://www1.chim.unifi.it/orac/>) has been used for ligand force field parameters. All homemade scripts used to calculate PF-07321332–3CL<sup>Pro</sup> absolute dissociation free energy are provided in the zip file in the Supporting Information and described in the pdf file in the Supporting Information. The step-by-step tutorial can be accessed at the [https://procacci.github.io/vdssb\\_gromacs/](https://procacci.github.io/vdssb_gromacs/) site. All output files are reported on <https://zenodo.org/record/5139374>.

## ■ ASSOCIATED CONTENT

### Supporting Information

The Supporting Information is available free of charge at <https://pubs.acs.org/doi/10.1021/acs.jcim.1c00909>.

Step-by-step technical tutorial for the calculation of the ADFE in the PF-07321332–3CL<sup>Pro</sup> system (PDF)

All homemade scripts used to calculate PF-07321332–3CL<sup>Pro</sup> absolute dissociation free energy (ZIP)

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### Notes

The authors declare no competing financial interest.

## ■ ACKNOWLEDGMENTS

We acknowledge the Partnership for Advanced Computing in Europe (PRACE) for awarding us access to Marconi100 at CINECA consortium (Italy) (Pra20\_CV28). We thank the CINECA staff for technical assistance on the Marconi100 HPC. We thank Giovanni Bussi for assistance in the use of the PLUMED library with GROMACS. We thank Yuriy Khalak,

Vytautas Gapsys, Matteo Aldeghi, and Bert de Groot for their self-less collaborative attitude and precious assistance in performing nonequilibrium free energy calculations with GROMACS code. The authors thank MIUR-Italy (“Progetto Dipartimenti di Eccellenza 2018-2022” allocated to Department of Chemistry “Ugo Schiff”).

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