

## Research Article

# Inhibitory Effect of *Dodonaea viscosa* var. *angustifolia* on the Virulence Properties of the Oral Pathogens *Streptococcus mutans* and *Porphyromonas gingivalis*

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**Aim.** This study investigated the effect of *Dodonaea viscosa* var. *angustifolia* (DVA) on the virulence properties of cariogenic *Streptococcus mutans* and *Porphyromonas gingivalis* implicated in periodontal diseases. **Methods.** *S. mutans* was cultured in tryptone broth containing a crude leaf extract of DVA for 16 hours, and the pH was measured after 10, 12, 14, and 16 h. Biofilms of *S. mutans* were grown on glass slides for 48 hours and exposed to plant extract for 30 minutes; the adherent cells were reincubated and the pH was measured at various time intervals. Minimum bactericidal concentration of the extracts against the four periodontal pathogens was determined. The effect of the subinhibitory concentration of plant extract on the production of proteinases by *P. gingivalis* was also evaluated. **Results.** DVA had no effect on acid production by *S. mutans* biofilms; however, it significantly inhibited acid production in planktonic cells. Periodontal pathogens were completely eliminated at low concentrations ranging from 0.09 to 0.02 mg/mL of crude plant extracts. At subinhibitory concentrations, DVA significantly reduced Arg-gingipain (24%) and Lys-gingipain (53%) production by *P. gingivalis* ( $P \leq 0.01$ ). **Conclusions.** These results suggest that DVA has the potential to be used to control oral infections including dental caries and periodontal diseases.

## 1. Introduction

The oral cavity is a complex ecosystem comprising many surfaces coated with a wide variety of species, and an opportunistic microflora which exists in the form of a biofilm may vary due to dietary constituents, systemic illnesses, poor saliva flow, and oral hygiene resulting in the alteration of the microbial communities which leads to the development of oral diseases.

*Streptococcus mutans* occurs in the oral cavity and has the ability to ferment dietary carbohydrates rapidly and produce acids which are responsible for the demineralization of enamel. The acidic environment also promotes the growth and virulence of the opportunistic pathogen *Candida* in the oral environment [1, 2]. In addition, it is implicated in root canal infections, odontogenic abscesses, and endocarditis.

Soft tissue infections including periodontal disease are caused by obligate oral anaerobic gram negative bacteria, categorized as red and orange complexes of *Porphyromonas gingivalis*, *Fusobacterium nucleatum*, *Prevotella intermedia*, and others [3]. Among these opportunistic pathogens, *P. gingivalis* is the most aggressive organism mainly because it has the ability to produce proteases, for example, Arg-gingipain and Lys-gingipain as well as lipopolysaccharides, collagenases, and haemagglutinin and is protected by a capsule [4, 5]. In addition, *P. gingivalis* is linked to systemic diseases such as aortic atherosclerosis, myocarditis, myocardial infarction, and rheumatoid arthritis [6–8].

Oral hygiene products containing antimicrobial agents such as fluoride and chlorhexidine have been used to control biofilms containing *S. mutans* and *P. gingivalis*. In addition, the antimicrobial effect of many natural products has been

tested on the virulence properties of *S. mutans*, that is, biofilm formation and acid production [9, 10].

Inhibitors of proteases produced by *P. gingivalis* are potentially new therapeutic agents that could control these organisms and related diseases [11]. *Dodonaea viscosa* var. *angustifolia* (DVA) also called hopbush, which is traditionally used for many ailments including oral infections [12, 13], is known to inhibit biofilm formation by *S. mutans* and reduce the virulence of *Candida albicans* [14, 15]. Furthermore, its cytotoxicity has also been established [16].

This study investigated the effect of a plant extract of the leaves of *Dodonaea viscosa* var. *angustifolia* on acid production by *S. mutans* and protease production by *P. gingivalis* and the antibacterial effect against *P. gingivalis*, *P. intermedia*, *F. nucleatum*, and *Capnocytophaga* species.

## 2. Materials and Methods

**2.1. Cultures.** Saliva and periodontal pocket debris samples from patients attending the Dental Clinic at Charlotte Maxeke Johannesburg Academic Hospital were collected and cultured on Mutans Bacitracin agar to isolate *Streptococcus mutans*. Blood agar supplemented with haem and menadione was used to isolate *P. gingivalis*, *Prevotella intermedia*, *Fusobacterium nucleatum*, and *Capnocytophaga* species. *S. mutans* cultures were identified using API 20 Strep auxanogram (bioMérieux), and additional biochemical reactions. *S. mutans* NCTC 10919 and 4 clinical isolates of *S. mutans* were used in the study. *P. gingivalis*, *P. intermedia*, and *F. nucleatum* were identified using API 32A and PCR technique [17]. *Capnocytophaga* spp. was identified using colony morphology and gram stain. Ethical clearance was obtained from The Committee for Research on Human Subjects (Medical), University of the Witwatersrand. Written consent was obtained from the subjects. Cultures were stored at  $-70^{\circ}\text{C}$  until required. For each experiment, fresh inoculums containing approximately  $10^6$  organisms per millilitre were prepared [14].

**2.2. Plant Material and Extract Preparation.** Plant material was collected from the Pipeklipberg, Mkhunyanne Eco Reserve, Mpumalanga province of South Africa, verified (Voucher Specimens no. J 94882) as described previously [15].

Leaves of *Dodonaea viscosa* var. *angustifolia* were dried in the shade and milled to a fine powder. Thereafter, 1.0 g was extracted in 10 mL methanol with vigorous shaking and then centrifuged. The procedure was repeated three times [18]. The solvents were removed under a cold air stream, and a yield of 0.15 g dried extract was obtained. The crude dry extract was weighed and dissolved in DMSO to yield a solution containing 50 mg of crude extract per mL of DMSO. Similarly, extracts were prepared using acetone and ethanol as solvents. Fresh plant extracts were prepared for each experiment.

**2.3. Minimum Bactericidal Concentration (MBC).** MBC tests were performed to determine the subinhibitory concentrations for the subsequent experiments. Twofold dilutions of

crude plant extract were prepared in microtitre plates using appropriate medium; *P. gingivalis* and *P. intermedia*, Tryptone Soy broth containing haem and menadione; *Capnocytophaga* spp., Tryptone Soy broth; and *F. nucleatum*, Fusiform media. One hundred microlitres of each of the diluted plant extract was added to each of the 96-well round bottom microtitre plate. Fresh inocula with optical density of 0.2 (405 nm) containing approximately  $10^5$ - $10^6$  organisms per millilitre were prepared, and each well was inoculated with 100  $\mu\text{L}$  of inocula. Microtitre plates containing *Capnocytophaga* spp. were incubated under  $\text{CO}_2$  at  $37^{\circ}\text{C}$  for 4 days. Plates containing *P. gingivalis*, *P. intermedia*, and *F. nucleatum* were incubated anaerobically at  $37^{\circ}\text{C}$  for 7 days. Chlorhexidine gluconate was used as a positive control and water as a negative control. Effect of DMSO was also measured. After incubation, each well was subcultured on blood agar. The lowest concentration that had no growth was recorded as MBC for that test organism. Each experiment was repeated in triplicate. Based on the MBC for *P. gingivalis*, subinhibitory concentrations were selected for the protease inhibition experiments. With *S. mutans*, the MBC of a previous study was used [14].

**2.4. Acid Production by *S. mutans*.** Methanol extract was used for the experiment. The effect of crude plant extract on the acid production by *S. mutans* grown in biofilms was studied using a technique described by Kim et al. [19], with modifications [19]. Fresh inocula of *S. mutans* with optical density of 0.2 (405 nm) containing approximately  $10^5$ - $10^6$  organisms per millilitre were prepared. *S. mutans* biofilms were allowed to grow on glass slides in tryptone broth containing approximately  $10^5$  cfu/mL *S. mutans* for 48 h with a change of the media after 24 h. Biofilms were then exposed to a 0.78 mg/mL subinhibitory concentration of crude plant extract [14] for 30 min and rinsed with phosphate-buffered saline (PBS). Thereafter, the adherent cells were transferred to 30 mL tryptone broth and incubated in  $\text{CO}_2$  at  $37^{\circ}\text{C}$ . The pH of the media was measured after 2, 4, 6, 8, 13, and 22 h. The effect of crude plant extract on the acid production by planktonic cells of *S. mutans* was tested by inoculating 30 mL of tryptone broth containing 0.78 mg/mL crude plant extract with 1 mL of inoculum containing  $10^5$ - $10^6$  cfu/mL of *S. mutans* and incubated for 16 hours. The pH of the media was measured at 0, 10, 12, 14, and 16 h., and the *S. mutans* count was determined after 0, 12, and 16 hours using the microdilution technique [20].

In both of these experiments water instead of plant extract was used as a control. The effect of DMSO was also measured. Each experiment was repeated in triplicate for the 5 strains of *S. mutans* that were tested. The results were analysed using the Wilcoxon rank-sum test (Mann-Whitney).

**2.5. Protease Inhibition.** The inhibitory activity of the methanol plant extract against production of proteinases Arg-gingipain and Lys-gingipain by *P. gingivalis* was evaluated using the technique of Yamanaka et al. [21] with modifications [21]. *P. gingivalis* was harvested by centrifugation at 5 000 g for 20 minutes, washed 3 times, resuspended in

TABLE 1: Minimum bactericidal concentrations of *Dodonaea viscosa* var. *angustifolia* against periodontal pathogens.

Cultures	Median MBC in mg/mL ( $n = 3$ )		
	Acetone	Ethanol	Methanol
<i>P. intermedia</i>	0.04	0.09	0.04
<i>P. gingivalis</i>	0.04	0.09	0.09
<i>F. nucleatum</i>	0.09	0.09	0.04
<i>Capnocytophaga</i>	0.04	0.02	0.02

Lowest MBC of DMSO was 6.125% and chlorhexidine gluconate killed all the test organisms.

50 mM phosphate-buffered saline (pH 7.4), and adjusted to an optical density of 2.0 at 660 nm. Benzoyl-arginine-*p*-nitroanilide and N-(*p*-Tosyl)-Gly-Pro-Lys 4-nitroanilide acetate salts in 0.1 M Tris-HCL (pH 8.0) containing 1 mM dithiothreitol were used as substrates for Arg-gingipain and Lys-gingipain, respectively.

One hundred microliters of the substrates was dispensed into the wells of a microtitre plate. Different concentrations of plant extract (including water as a control) and bacterial cell suspensions were added and incubated at 37°C for 20 min. Controls without substrate were included in each plate. Adsorption was measured at a wavelength of 405 nm. Relative enzymatic activity was determined as follows:  $[(A_{405} \text{ with bacterial cells and plant extract} - A_{405} \text{ of control}) / (A_{405} \text{ with bacterial cells} - A_{405} \text{ of control})] \times 100$ . Each experiment was repeated 10 times. The results were compared to the control, that is, water using Student's *t*-test.

### 3. Results

**3.1. MBC.** Periodontal pathogens were completely eliminated at low concentrations ranging from 0.09 to 0.02 mg/mL of crude plant extracts (Table 1). All the solvents gave similar results. Based on these results, subinhibitory concentrations ( $\leq 0.02$  mg/mL) were selected for the virulence study of *P. gingivalis*. The antimicrobial effect of DMSO was lost at a concentration of 6.125% whereas chlorhexidine gluconate killed all the test organisms.

**3.2. Acid Production.** DVA had no effect on the acid production ( $P \geq 05$ ) by *S. mutans* biofilms (Figure 1); however, it significantly inhibited acid production ( $P \leq 0.01$ ) in planktonic cells (Figure 2). Bacterial counts in these cultures were performed to eliminate the possible antimicrobial effect of DVA at this subinhibitory concentration. The results showed that there was no significant difference in the bacterial counts of the control and the test cultures ( $P \geq 0.05$ ). This suggests that reduced acid production was due to the effect of DVA rather than the bacterial growth.

**3.3. Proteinase Inhibition.** DVA significantly ( $P \leq 0.01$ ) reduced the Arg-gingipain and Lys-gingipain production by *P. gingivalis* (Figure 3). The highest reduction of Arg-gingipain was at 0.02 mg/mL DVA (24%) whereas 53% reduction in the production of Lys-gingipain was achieved at 0.01 mg/mL.

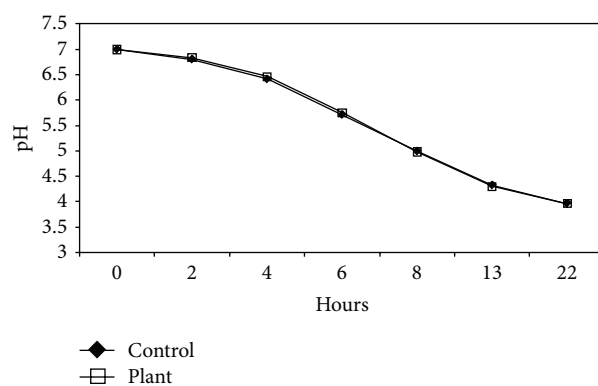


FIGURE 1: Effect of *Dodonaea viscosa* var. *angustifolia* on the acid production by biofilm of *S. mutans*.

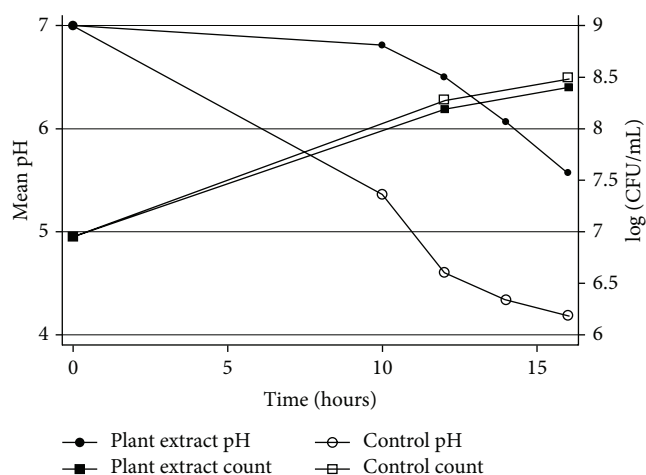


FIGURE 2: Effect of *Dodonaea viscosa* var. *angustifolia* on the acid production by planktonic *S. mutans*.

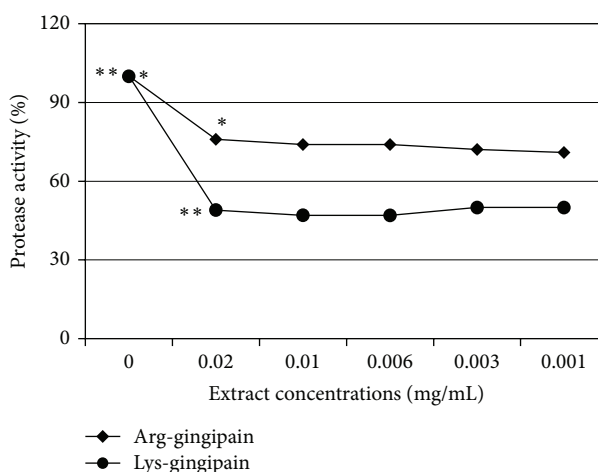


FIGURE 3: Effect of *Dodonaea viscosa* var. *angustifolia* on the production of Arg-gingipain and Lys-gingipain by *Porphyromonas gingivalis*. \* and \*\*:  $P \leq 0.01$ .

#### 4. Discussion

In the last decade due to the escalation in the development of drug resistance in microorganisms, a novel approach has been proposed whereby the therapeutic agent can target the virulence of causative organisms rather than the number of infectious agents [22, 23]. This approach can be applied to commensals that cause opportunistic infections as they are often not eliminated completely.

This study showed that a subinhibitory concentration of the crude extract of *D. viscosa* var. *angustifolia* significantly inhibits acid production by *S. mutans*, which is an important virulence factor of this organism. *S. mutans* ferments dietary carbohydrates and produces acids which can cause demineralization of teeth [24]. As demonstrated in this study, the presence of plant extract will not allow *S. mutans* to produce acid, thereby preventing dental caries. This effect was not dependent on the number of *S. mutans* because the cfu/mL of *S. mutans* was not significantly different from the controls. This suggests that the phosphotransferase (PTS) uptake system that assists the transport of glucose could not have been affected by the plant extract. However, conversion of glucose to pyruvate by EMP pathway may have been affected. Polyphenols and tannins are known to interact with cellular enzymes that are responsible for the metabolic pathways, and these compounds have been identified from DVA [14, 25–27].

Our results also showed that acid production by *S. mutans* in the biofilm form is not affected by DVA, though it is known to prevent biofilm formation [14]. If DVA is added to oral hygiene products, regular use will inhibit the plaque formation as well as acid production by planktonic cells of *S. mutans*. The unfavorable results regarding acid production by the biofilms may have been due to the extracellular polysaccharides that are normally produced in the biofilm by *S. mutans*. They are tenacious and may have prevented the penetration of many large molecules including chemicals present in the plant extract.

DVA inhibited the growth of some of the anaerobic gram negative bacteria including *P. gingivalis* that are implicated in soft tissue infections. In addition, it reduced the production of Arg-gingipain by 24% and Lys-gingipain by 53%, which are the most important virulence factors of *P. gingivalis*. These proteinases downregulate polymorphonuclear neutrophils, modulate host cytokine networks, and degrade proteins which are present in the gingival tissues [21, 28]. Gingipains are essential to *P. gingivalis* as they provide iron, peptides, and amino acids from environmental proteins [29]. In addition, they are responsible for the cleavage of human transferrin which promotes growth and formation of hydroxyl radicals that play an important role in tissue destruction during infection [30]. Therefore, inhibition of gingipains by DVA will also deprive *P. gingivalis* of available iron and prevent some tissue destruction. It has been suggested that polyphenols and catechins may be responsible for the antibacterial and antiproteolytic effects against *P. gingivalis* [21, 30, 31] which are also present in DVA [14].

The results of this study have shown that if DVA is incorporated into a mouth rinse, gel or toothpaste at high

concentrations, it will eliminate four of the major periodontal pathogens including *P. gingivalis*. Even though the concentration of the plant extract is reduced in the oral cavity due to salivary flow, it will render *S. mutans* and *P. gingivalis* avirulent. However, further research is needed to identify the active ingredient responsible for the beneficial effects.

#### 5. Conclusions

At subinhibitory concentrations a crude extract of DVA renders *S. mutans* and *P. gingivalis* avirulent by preventing acid and proteinase production, respectively. In addition, it kills *P. gingivalis*, *P. intermedia*, *F. nucleatum*, and *Capnocytophaga* species at low concentrations. This supports the suggestion that the extract of this plant has the potential to be used as preventive and therapeutic agent for the treatment of infections of the mouth.

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