



NEURODEGENERATION

Fly model sheds light on brain disease

Experiments on flies suggest that a gain-of-function mechanism in a protein called CSP α contributes to the progressive brain disease CLN4.

MARTIN H BERRYER, SARA G KOSMACZEWSKI AND LINDY E BARRETT

Related research article Imler E, Pyon JS, Kindelay S, Torvund M, Zhang YQ, Chandra SS, Zinsmaier KE. 2019. A *Drosophila* model of neuronal ceroid lipofuscinosis CLN4 reveals a hypermorphic gain of function mechanism. *eLife* 8:e46607. DOI: 10. 7554/eLife.46607

euronal ceroid lipofuscinosis (NCL) refers to a group of progressive brain diseases that affect between 1 and 30 per 100,000 people and are characterized by a decline in motor abilities, seizures, dementia and premature death (Nosková et al., 2011; Haltia, 2003). Most NCLs are caused by the failure of cells to recycle various proteins inside lysosomes (Wisniewski et al., 2001; Henderson et al., 2016; Sambri et al., 2017; Warrier et al., 2013). However, one type of neuronal ceroid lipofuscinosis is different: CLN4 disease occurs when a patient inherits one mutant copy of the gene that encodes for a protein called CSP α , but the precise cellular dysfunction underlying CLN4 disease remains a mystery (Nosková et al., 2011; Benitez and Sands, 2017).

Mutations in this gene have been associated with both loss and gain of function. In theory, the mutations could deplete normal CSP α , and this loss of function could contribute to disease pathology. Alternatively, the mutations could enhance the normal activity of CSP α or lead to

an additional, toxic function to drive disease pathology. Now, in eLife, Konrad Zinsmaier and colleagues at the University of Arizona and Yale University – including Elliot Imler as first author – report the generation of a new animal model to investigate the biological mechanisms underlying CLN4 disease (*Imler et al., 2019*).

Imler et al. started by expressing either the normal or mutant human forms of $CSP\alpha$ in flies and confirming that both were functional in fly cells. Next they confirmed that mutant forms of $CSP\alpha$ could mimic pathological features seen in CLN4 patients. Interestingly, they found that the severity of the disease correlated with the copy number of the mutated gene: a single copy of the mutated gene did not affect lifespan, but two copies led to more severe phenotypes and early death. To confirm that these results were not an artifact of expressing a human protein in flies, Imler et al. repeated many of their experiments using mutant versions of fly $CSP\alpha$.

The Arizona–Yale team then looked at where mutant CSP α resides in neurons. Normally, one would expect to find CSP α at nerve terminals, but the mutations resulted in lower levels of the protein at nerve terminals and higher levels in regions of the cell that contained other proteins that had been marked for degradation. By using markers of different cellular components, the researchers demonstrated that mutant CSP α was accumulating on prelysosomal endosomes. An endosome is a mini-compartment within a cell that internalizes molecules from the cell membrane: the endosome then fuses with an organelle called a lysosome, and the molecules inside it are broken down and recycled. Imler

© Copyright Berryer et al. This article is distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use and redistribution provided that the original author and source are credited



et al. hypothesize that mutant CSP α may be rerouted from nerve terminals through the endolysosomal pathway. Consistent with this, electron microscopy revealed the formation of abnormal membrane structures in cells, which may be due to mutant CSP α congesting the trafficking system.

Next, Imler et al. modulated the levels of normal and mutant CSP α in cells. Reducing the level of normal CSP α reduced the mutant phenotypes, while increasing it exacerbated the mutant phenotypes. Similarly, reducing the level of a heat shock protein that normally interacts with CSP α attenuated mutant phenotypes, suggesting that the heat shock protein may play a role in disease biology. The Arizona–Yale team suggests that the disease-causing mutations in CSP α enhance its normal activity to disrupt the function of neurons. This gain-of-function mechanism is distinct from that found in the other NCLs, which typically arise from a deficiency in the function of the mutated gene.

The results of Imler et al. demonstrate the value of the fruit fly model to study CLN4 disease pathology. However, these results also paint a complex picture of CLN4, and much work remains to be done to understand how each molecular or cellular pathology contributes to the devastating cognitive and motor deficits found in patients. Future work can now focus on how these biochemical disruptions impact neuronal function over time, taking advantage of the many tools and techniques available in fly genetics in order to interrogate pathways that have additive or compensatory mechanisms that may someday yield therapeutic potential.

Martin H Berryer is in the Stanley Center for Psychiatric Research, Broad Institute of MIT and Harvard, Cambridge, United States and the Department of Stem Cell and Regenerative Biology, Harvard University, Cambridge, United States mberryer@broadinstitute.org

Sara G Kosmaczewski is in the Stanley Center for Psychiatric Research, Broad Institute of MIT and Harvard, Cambridge, United States and the Department of Stem Cell and Regenerative Biology, Harvard University, Cambridge, United States skosma@broadinstitute.org

Lindy E Barrett is in the Stanley Center for Psychiatric Research, Broad Institute of MIT and Harvard, Cambridge, United States and the Department of Stem Cell and Regenerative Biology, Harvard University, Cambridge, United States Ibarrett@broadinstitute.org

b https://orcid.org/0000-0002-3229-4774

Competing interests: The authors declare that no competing interests exist.

Published 06 December 2019

References

Benitez BA, Sands MS. 2017. Primary fibroblasts from CSP α mutation carriers recapitulate hallmarks of the adult onset neuronal ceroid lipofuscinosis. *Scientific Reports* **7**:6332. DOI: https://doi.org/10.1038/s41598-017-06710-1

Haltia M. 2003. The neuronal ceroid-lipofuscinoses. Journal of Neuropathology & Experimental Neurology 62:1–13. DOI: https://doi.org/10.1093/jnen/62.1.1, PMID: 12528813

Henderson MX, Wirak GS, Zhang YQ, Dai F, Ginsberg SD, Dolzhanskaya N, Staropoli JF, Nijssen PC, Lam TT, Roth AF, Davis NG, Dawson G, Velinov M, Chandra SS. 2016. Neuronal ceroid lipofuscinosis with DNAJC5/CSPα mutation has PPT1 pathology and exhibit aberrant protein palmitoylation. *Acta*

Neuropathologica **131**:621–637. DOI: https://doi.org/ 10.1007/s00401-015-1512-2, PMID: 26659577

Imler E, Pyon JS, Kindelay S, Torvund M, Zhang YQ, Chandra SS, Zinsmaier KE. 2019. A *Drosophila* model of neuronal ceroid lipofuscinosis *CLN4* reveals a hypermorphic gain of function mechanism. *eLife* 8: e46607. DOI: https://doi.org/10.7554/eLife.46607, PMID: 31663851

Nosková L, Stránecký V, Hartmannová H, Přistoupilová A, Barešová V, Ivánek R, Hůlková H, Jahnová H, van der Zee J, Staropoli JF, Sims KB, Tyynelä J, Van Broeckhoven C, Nijssen PC, Mole SE, Elleder M, Kmoch S. 2011. Mutations in DNAJC5, encoding cysteine-string protein alpha, cause autosomaldominant adult-onset neuronal ceroid lipofuscinosis. American Journal of Human Genetics 89:241–252. DOI: https://doi.org/10.1016/j.ajhg.2011.07.003, PMID: 21820099

Sambri I, D'Alessio R, Ezhova Y, Giuliano T, Sorrentino NC, Cacace V, De Risi M, Cataldi M, Annunziato L, De Leonibus E, Fraldi A. 2017. Lysosomal dysfunction disrupts presynaptic maintenance and restoration of presynaptic function prevents neurodegeneration in lysosomal storage diseases. *EMBO Molecular Medicine* **9**:112–132. DOI: https://doi.org/10.15252/emmm. 201606965, PMID: 27881461

Warrier V, Vieira M, Mole SE. 2013. Genetic basis and phenotypic correlations of the neuronal ceroid lipofusinoses. *Biochimica et Biophysica Acta (BBA) - Molecular Basis of Disease* 1832:1827–1830.

DOI: https://doi.org/10.1016/j.bbadis.2013.03.017

Wisniewski KE, Zhong N, Philippart M. 2001. Pheno/ genotypic correlations of neuronal ceroid lipofuscinoses. *Neurology* 57:576–581. DOI: https://doi.org/10.1212/WNL.57.4.576, PMID: 11548735