Therapeutic Interventions Targeting Beta Amyloid Pathogenesis in an Aging Dog Model

Sarah B. Martin^a, Amy L.S. Dowling^a and Elizabeth Head^{a,b,*}

^aSanders Brown Center on Aging, University of Kentucky, Lexington KY, USA; ^bDepartment of Molecular and Biomedical Pharmacology, University of Kentucky, Lexington KY, USA

Abstract: Aged dogs and humans share complex cognitive and pathological responses to aging. Specifically, dogs develop Alzheimer's Disease (AD) like beta-amyloid (A β) that are associated with cognitive deficits. Currently, therapeutic approaches to prevent AD are targeted towards reduced production, aggregation and increased clearance of A β . The current review discusses cognition and neuropathology of the aging canine model and how it has and continues to be useful in further understanding the safety and efficacy of potential AD prevention therapies targeting A β .

Keywords: Alzheimer's disease, canine, statins, BACE-1 inhibitors, metal-chelators, A-beta vaccination.

INTRODUCTION

Alzheimer's disease (AD) is the most common cause of dementia in the elderly and currently affects 5.5 million people in the United States alone. AD is characterized by the presence of senile plaques and neurofibrillary tangles (NFTs) in the brain [1-3]. Given that the greatest risk factor for developing AD is age and that our elderly population is rapidly rising, it is critical to identify preventative measures and early interventions to slow or stop the disease [4]. The current review provides an overview of how the dog model of human aging and AD has been and continues to be a beneficial preclinical model to test the efficacy and safety of certain types of disease modifying treatments for AD.

Beta-Amyloid (AB)

Senile plaques, which are primarily composed of β amyloid (A β), are one of two neuropathological characteristics of AD [5]. The A β peptide is produced by the sequential cleavage of the amyloid precursor protein (APP) by β - and γ secretases [6, 7]. Cleavage by γ -secretase results in differing lengths of A β , with the 42 amino acid form, A β_{42} , making up most of the insoluble deposits found in the AD brain [8]. A β is largely thought to play a role in the disease process because it is a unifying pathological feature of the genetically diverse forms of AD.

The gene encoding APP is located on chromosome 21. Because three copies of this chromosome are present in individuals with Down Syndrome (DS), DS research has been particularly important in elucidating the role of A β in AD [9, 10]. Individuals with DS show A β deposition as early as 8 years of age [11] and have classical neuropathological features of AD by their early forties [12-15]. The localization of the APP gene to chromosome 21 initiated a search for genetic linkages in families with autosomal dominant AD [7],

leading to the association of missense mutations in the APP gene with familial AD [16-19]. Furthermore, duplication of the APP gene causes autosomal dominant AD in the absence of DS [20]. Past studies of DS and familial AD have established that A β is a pathological feature of both disorders and that A β production and decreased clearance [21, 22] is at the root of disease progression. The importance of A β in the pathogenesis of AD has led to the search for therapeutic interventions targeting A β .

Animal Models for A^β Therapeutics: Beyond the Mouse

Potential AD therapeutics are generally screened first in transgenic mouse models of AD. As well as being inexpensive and easy to house, mouse models are ideal because they age quickly and their entire genome is mapped. However, mice do not produce human sequence A^β naturally [23]. Transgenic mouse models of AD are produced by over expressing mutant human APP alone or combined with transgenic presenilin 1 (PS1) and presenilin2 (PS2) genes in mice, which leads to $A\beta$ plaque formation [24]. Importantly, unlike humans, transgenic mice show cellular and behavioral resilience to $A\beta$ pathology and thus do not develop the extensive neuronal loss observed in the AD brain. While transgenic mouse models continue to get more complex and more closely replicate human AD pathology, there are several components of the disease process that mice cannot model. For example, most AD patients neither have the genetic mutations present in transgenic mice nor overexpress these mutations if present at such high levels. While transgenic mouse models have been and will continue to be instrumental in the development of therapeutics, the basic evolutionary differences between the two species makes taking potential therapeutics from mice to humans less direct.

The successful development of therapeutic interventions targeting $A\beta$ benefits from testing in animal models that naturally recapitulate critical aspects of human disease. Diseases associated with brain aging are especially challenging, due to the time required to track both the aging process and the interplay between pathology and cognition. An ideal

^{*}Address correspondence to this author at the Sanders Brown Center on Aging, University of Kentucky, Lexington KY, USA; Tel: 859-257-1412x481; Fax: 859-323-2866; E-mail: elizabeth.head@uky.edu

animal model has several key features in addition to the natural development of AD pathology. For example, it is beneficial for an animal model of AD to have a naturally diverse response to aging, given that humans show individual variations with age [25-27]. While some people show mild cognitive decline with age, others develop severe cognitive decline. Still others develop no visible signs of decline, the definition of "successful aging". This spectrum of cognition in aging is suggested to result from both genetic and environmental factors [28, 29], another key feature of an ideal animal model of AD. Although increased variability presents statistical challenges, requiring additional animals and experimental studies, natural diversity offers certain unique advantages. The ability to compare similarly aged animals with differing cognitive functions allows researchers to distinguish pathological underpinnings associated with cognitive function.

Not all cognitive domains are equally vulnerable to aging or AD in humans. For example, the first sign of AD in humans, often referred to as amnestic mild cognitive impairment (MCI), is defined by a gradual decline in episodic memory function with preserved general cognitive and functional abilities, combined with no evidence of dementia [27, 30]. As the disease progresses into clinical AD, patients show a disturbance in at least one of the following cognitive functions: language, motor skills, visual processing or executive function [31]. More specifically, AD patients show a decline in declarative learning, while procedural abilities remain intact. Animal models that capture these cognitive characteristics are well suited to develop and test therapeutic interventions to translate into human clinical trials in AD patients. Unfortunately, no single animal model can replicate all aspects of AD, but each can provide unique strengths that advance our knowledge.

CANINE MODEL OF AGING

The canine model has a rich literature in psychological and neurobiological research, dating back to the 1800s. Canines are useful for aging research, have moderate lifespans of 12 to 20 years, depending on the breed [32, 33], and are easy to handle due to a long history of domestication [34]. Furthermore, canines are highly motivated to perform consistently on cognitive tests using simple food rewards, making food deprivation paradigms unnecessary. In contrast, mice are not readily cooperative in performing behavioral tasks, so physiological stressors including food restriction, water deprivation and immersion in water are often used [35]. Therefore measures of cognition may be engaging other processes involved in stress response, confounding performance scores. Importantly, the cognitive decline and progressive age-associated neuropathology observed in dogs parallels that of humans.

Cognition and Aging in Dogs

Cognitive testing procedures for canines were initially developed by modifying non-human primate cognitive tests [36]. A variety of tests have been developed to assess cognitive function in various domains and corresponding brain localization in dogs. Table **1** [originally published in 37] outlines cognitive domains assessed in dog aging and

how they compare with assessment tasks for non-human primate and humans. Many of these tests are analogous to cognitive tests used for nonhuman primates and humans. As in humans, canine individual variability and domain-specific cognitive vulnerabilities are key features of decline with age [36]. Beginning in middle age, individual variability in cognitive scores begins to increase, with the largest variability seen in aged dogs [38]. Additionally, vulnerability to decline with age varies as a function of cognitive domain and the cortical circuits engaged. For example, size discrimination learning is sensitive to age, while simple object discrimination is not [36, 38, 39], similar to monkey models of aging [40-44]. Further, prefrontal dependent reversal learning is more age sensitive than discrimination learning [36, 38, 39].

Efficacy of a therapeutic intervention for AD is ultimately measured by its ability to stabilize or improve function in cognitive domains compromised by AD. The canine parallels features of human cognitive decline and has well established measures that are domain specific [reviewed in 37]. Use of the canine model allows researchers to study the complex cognitive implications of therapeutic interventions, an important measure preceding clinical trials.

Neuropathology in Aging Dogs

In addition to the cognitive features of aging, dogs naturally develop brain changes similar to human brain aging [reviewed in 37]. Importantly, a key feature that makes dogs useful as a model of human A β pathogenesis was the observation in 1956, by Braumühl, who reported "Alzheimer'slike" senile plaques in aged dogs [reviewed by 34]. Dogs initially develop plaques between the ages of 8 and 9 years [37, 45], a relatively young age, compared to many nonhuman primate models with naturally occurring AD-like pathology [46, 47]. Deposition of human-like A β accumulation in the aged canine brain has been well-described [34, 48-52]. There are significant homologies between canines and humans in the APP protein sequence, as well as in many enzymes involved in the processing of APP to form A β (http://www.ensembl.org/Canis_familiaris/).

Diffuse plaques are the predominant subtype of A β in the canine model [49, 52-56], whereas more compact neuritic plaques (i.e. $A\beta$ associated with tau positive neurites) are predominant in AD Fig. (1A, B). However, evidence suggests that diffuse plaques are extensive in humans and appear early in AD progression [57, 58]. As observed in humans, A β pathology in canines shows specific brain regions being differentially vulnerable to A β [1, 2, 50, 52, 59-61]. Modeling early stages of AD is critical, given that AD therapies are likely to be most effective early in disease progression. Importantly, unlike humans, canines do not develop NFTs [47, 53, 55, 56], the second neuropathological characteristic of AD. However, the lack of NFTs in canines is a feature of the model that can be used to differentiate between the effects of $A\beta$ and NFTs in the disease process, an ongoing subject of AD research.

Both humans and canines develop cerebrovascular amyloid angiopathy (CAA), the deposition of A β in association with the cerebrovasculature Fig. (**1C**, **D**). The presence of CAA in canines enhances the utility of this model in developing

Table 1.	Cognitive Domains Assessed in Dog Aging and Comparison with Nonhuman Primate Tasks and A	nalogous Tasl	s Used in
	Human Neuropsychological Testing		

Cognitive Domain	Dog Task	Localization in Dog Brain	Nonhuman Primate Tasks	Examples of Human Neuropsychological Tasks**	
	Visual discrimination learning	Medial temporal lobe/parietal lobe*	Visual discrimination learning [40, 164]	digit copy, rotary pursuit, face discrimination [165], object discrimination [166, 167]	
Learning	Reward and object approach learning	Nigrostriatal and motor cortex*	Food pickup task, fine motor learning [168, 169]		
	delayed nonmatching to sample acquisition	Rhinal cortex [170]	Object recognition memory task [171]	Delayed recognition and recall, digit span [172]	
Manager	delayed nonmatching to sample memory	Rhinal cortex [170]	Object recognition memory task [171]		
Memory	spatial delayed nonmatch to sample acquisition	Dorsolateral prefrontal cortex [170]	Delayed Response Task [43, 173]		
	spatial delayed nonmatch to sample memory	Hippocampus [174]	Delayed Response Task [43, 173]		
	Visual reversal learning	Prefrontal cortex/medial temporal lobe [175]	Visual reversal learning [40, 164]	card or object sorting tasks, set shifting, response inhibi- tion [176]	
	Oddity discrimination	Prefrontal cortex/medial temporal lobe*	N/A		
Function	Egocentric spatial reversal learning	Hippocampal/prefrontal cortex*	Spatial reversal [164]		
	Size concept learning	Prefrontal cortex/medial temporal lobe*	Hierarchical/Relational learning [177]		
Visuospatial	Landmark discrimination	Prefrontal cortex/parietal cortex*	Landmark discrimination [178]	Visual construction, block design, spatial learning [166, 167]	
Function	Egocentric spatial learning	Hippocampus/medial temporal lobe*	Spatial learning [164]		

* Proposed localization - not confirmed in lesion studies in dogs

** Neuropsychological tasks for humans that assess function in similar cognitive domains.

and testing therapeutic interventions to target A β . In both canines and humans with CAA, the blood vessels of the brain typically contain the shorter, 40 amino acid-long species of A β [62-64]. Further, the distribution of CAA is similar in both species, with the occipital cortex being predominantly susceptible. Vascular A β may compromise the blood-brain barrier, disrupt vessel wall viability [65] and cause microhemorrhages [66, 67].

Recently, the focus in AD pathogenesis has shifted from A β plaques to consider smaller, soluble forms of A β assemblies called A β oligomers. Oligomers are highly toxic and impair synaptic function [68]. Furthermore, elevated oligomer levels are strongly associated with cognitive dysfunction [69, 70]. A recent study by Pop *et al.*, examined the accumulation of oligomeric A β in the temporal lobe of canines. These results provide evidence that canines, like humans, experience an increase in toxic oligomers with age [71].

Another key feature of AD that is also found in dogs is neuronal loss in the presence of cognitive deficits. A study by Pugliese *et al.*, (2007) demonstrated that cognitive deficits correlated with loss of purkinje cells [72]. More recently, a study by Insua *et al.*, (2010) examined neuroadrenergic neurons in the locus ceruleus of aged canines, a group of neurons that are vulnerable to AD in humans [73, 74]. They found that cognitively impaired dogs exhibited significant reduction in noradrenergic neurons [75].



Fig. (1). Immunoreactivity for $A\beta$ 1-42 in frontal cortex brain tissue of (**A**) an aged canine and (**B**) an aged human. Compact $A\beta$ deposits are similar in humans and canines (arrow head). The outline of an intact neuron enveloped by a diffuse plaque is visible (arrow). $A\beta$ 1-40 immunoreactivity of cerebral amyloid angiopathy is similar in aged canine occipital cortex (**C**) and aged human occipital cortex (**D**).

The efficacy of $A\beta$ modifying therapies are measured by neuropathological and cognitive improvement. Studies have

found an association between $A\beta$ load and cognitive dysfunction in aging dogs, similar to humans with AD [48]. As with humans, there is not a perfect link between the extent of $A\beta$ pathology and the severity of cognitive decline, suggesting that other neuropathological cascades are involved in neurodegeneration. The natural presence of cognitive decline and $A\beta$ neuropathology make the dog a valuable model in developing therapeutics for AD.

POTENTIAL TARGETS FOR AB MODIFICATION

A β deposition is thought to result from one or more of the following mechanisms: 1) increased production of A β , 2) decreased A β clearance and/or 3) enhanced aggregation of A β Fig. (2). Strategies to decrease A β deposition have focused on modulating all of these mechanisms individually. For example, studies focused on preventing AB production inhibit the enzymatic step required to cleave APP into an $A\beta$ peptide. However, the monomeric A β peptide alone is easily cleared from the brain via the blood-brain barrier and is not considered a toxic form of A β [76]. The A β peptide becomes more toxic as it aggregates to the oligomer structural state [77-81] and subsequently further assembles into insoluble fibrils [reviewed in 82]. Therefore, a considerable amount of research is also focused on therapeutic interventions that prevent A β aggregation or enhance clearance of A β and A β aggregates. Here we will review some promising therapies that target production (antioxidant diet, behavioral enrichment, BACE1 inhibition, statins), aggregation (metal chelators) and clearance (A β immunotherapy) of A β Fig. (2). The dog model has been or may be useful for determining the efficacy of each potential therapy.

Antioxidants, Behavioral Enrichment and Prevention of $A\beta$ Deposition

Lifestyle factors are poorly understood, yet are potentially effective at preventing $A\beta$ production. Specifically, nutrition and lifestyle enrichment (e.g. exercise, education and social interaction) effectively prevent cognitive decline

and $A\beta$ production [83]. Results from mouse models of AD suggest that an antioxidant enriched diet [84, 85] and/or environmental enrichment [86] decreases production of $A\beta$ peptides. Human studies have determined that nutrition and other lifestyle factors offer cognitive benefits and reduce the risk of developing AD [87-93]. However, the genetic and environmental variability of the human population makes it difficult to determine the efficacy of these complex interventions and their mechanisms of action.

To measure cognitive and neurobiological benefits of lifestyle factors in aged dogs, diet and environment have been evaluated singularly and in combination, using a variety of learning and memory tasks. In the first study, aged beagles were treated with either a standard senior dog diet or an antioxidant-rich senior dog diet for 6 months [88, 94]. The formulation differences between control and enriched diets were as follows: D,L-alpha-tocopherol acetate (120 ppm vs. 1050 ppm), L-carnitine (<20ppm vs. 260ppm), D,Lalpha-lipoic acid (<20ppm vs. 128ppm), ascorbic acid as Stay-C (<30ppm vs. 80ppm) and 1% inclusion of each of the following (1 to 1 exchange for corn): spinach flakes, tomato pomace, grape pomace, carrot granules and citrus pulp. Oddity discrimination learning was assessed after 6 months of treatment. Aged dogs fed an antioxidant-enriched diet had significantly less age-dependent cognitive impairment than aged dogs fed the control diet.

In the follow up study, cognitive benefits of the antioxidant diet were evaluated after two years of treatment in the aged dogs [95]. The antioxidant-enriched diet was as described above. Further, the cognitive effects of the antioxidant diet either alone or in combination with behavioral enrichment were measured. Behavioral enrichment consisted of group exercise for 15 minute intervals twice a week, exposure to toys that were alternated weekly and housing with kennel mates. After two years of treatment, results suggested an attenuation of age-related cognitive decline that was particularly striking with the combined treatment of



Fig. (2). A β -modifying therapies can target the modulation of production and clearance. 1) Lifestyle interventions (e.g. nutrition and lifestyle enrichment) increase α -secretase activity. α -secretase cleaves APP in the middle of the A β region (denoted in yellow), releasing a non-amyloidogenic, soluble peptide. 2) BACE inhibitors and statins decrease β -secretase activity or BACE1 protein levels, which may result in decreased production of A β from APP. 3) Metal chelators prevent metal ions from inducing A β aggregation, thereby allowing A β peptides to be cleared from the brain. 4) Immunization therapies use anti-amyloid antibodies to bind and remove A β deposits.

behavioral and antioxidant enrichment. At the end of the study, A β neuropathology was measured in the prefrontal cortex and the extent of $A\beta$ was significantly decreased in animals receiving the antioxidant enrichment [96]. Further, A β pathology was most robustly decreased in animals that received both antioxidant and behavioral enrichment, suggesting that the combination of treatments may affect the production and/or clearance of AB. An increase in alpha-secretase enzyme activity in aged dogs treated with the antioxidant and behavioral enrichment protocols, suggests a shift towards non-amyloidogenic APP processing. These canine studies provide support for the combination of antioxidant and behavioral enrichment during the aging process in order to maintain cognition and inhibit AB production. Further, these results emphasize the importance of therapeutic interventions that target α -secretase activity. The canine model has lead to an increased understanding of the mechanisms and efficacy of these interventions on cognition and pathology.

BACE1 Inhibition and Prevention of Aβ Production

Two sequential enzymatic cleavages of APP are necessary to produce A β . The first is cleavage of APP by β secretase APP-cleaving enzyme 1 (BACE1). BACE1 cleaves APP, generating the N-terminus of A β . In AD brains, levels of BACE1 protein and activity are approximately twice that of controls [97-99], indicating enhanced production of Aβ. BACE1 is a promising target for the development of therapeutic interventions to lower A β production. However, it is important to note that BACE1 has substrates other than APP [100] and toxic side effects could result when BACE1 is fully inhibited. One possible side effect identified in a BACE1 knockout mouse (BACE1^{-/-}) involved NRG1, a substrate for BACE1 and a protein implicated in schizophrenia [101]. BACE1^{-/-}mice exhibit endophenotypes typically seen in schizophrenia [101, 102], leading researchers to test whether partial BACE1 inhibition decreases AB load while limiting unwanted side effects. Using a heterozygous mouse (BACE1^{+/-}) to impart partial BACE1 inhibition, McConlogue et al., found decreased AB burden in 13 and 18 month old BACE1^{+/-}APP transgenic mice [103]. Thus, BACE1 inhibition remains a potential therapeutic target.

The use of mouse models in designing BACE1 inhibitors has been instrumental. When inhibiting an enzyme that plays critical physiological roles in brain chemistry, it is increasingly important to test the putative therapy in several different models of AD prior to clinical trial. Humans and canines share a 98% homology in BACE1 (http://www. ensembl.org/Canis_familiaris/). Prior to clinical trials, testing therapeutic interventions that target BACE1 inhibition in a dog model may provide additional information regarding the dose requirements, safety and efficacy of these AD therapies.

Statins, Cholesterol and the Prevention of A_β Production

Several cross-sectional or case-control epidemiological studies have revealed a striking link between cholesterollowering drugs (e.g. statins and others) and a 20-70% reduction in risk of developing AD [104-111]. However some [112-114], but not all [115], prospective studies have reported no link between statin use and protection against dementia. Differential reports of the positive effects of statins on the development of AD may be due to the cohort studied, confounds by indication, type of statin used, age group studied, and type of study conducted (e.g. crosssectional, case-control or prospective study) [116, 117]. Further, in preliminary AD clinical trials with simvastatin [118] and atorvastatin [119-122], modest cognitive benefits have been reported. In particular, AD patients with mild to moderate dementia who were treated with 80 mg/day atorvastatin had significantly improved scores on one measure of cognition (ADAS-Cog) at 6 months of treatment, with smaller non-significant benefits at 12 months [120].

Recent studies strongly suggest once the signs of AD are evident that the use of statins might reflect a preventative approach rather than a treatment. For example, several studies suggest that mid-life cholesterol levels and statin use impact the risk of developing AD [123, 124]. Further, in a 5-year prospective study of individuals who were not demented at the start of the trial, statin use resulted in a 50% reduction in the risk of developing either dementia or cognitive impairments without dementia (CIND), as compared to placebo/control [125]. Thus, modulating mid-life cholesterol and/or statin administration and/or implementing statin use prior to CIND or dementia may be more beneficial than treatment approaches in patients with AD. Prevention studies are long, costly and challenging to conduct, but can be greatly facilitated by appropriate preclinical testing. Studies in animal models can provide preliminary information regarding potential benefits of statins in the prevention of cognitive decline and AD neuropathology.

Statins may reduce the risk of incident AD through the prevention of A β production [126, 127]. In rodent models, treatment with inhibitors of 3-hydroxy-3-methylglutaryl coenzyme A (HMG-CoA) or statins reduces A β [128]. However, rodents respond to statin treatment by massively upregulating HMG-CoA reductase levels [129-132]. To compensate, long-term studies in rodent often employ physiologically excessive doses, making it difficult to translate the results of these studies into human trials.

In contrast to rodent models, the dog model is a useful model for studies of chronic statin treatment, given similarities with humans in terms of dose requirements, responsiveness, drug handling and metabolism [129, 133]. For example, in a study of 12 animals, dogs were treated with 80 mg/day of atorvastatin for 14.5 months (Head, unpublished data). Peripheral levels of cholesterol, low density lipoproteins, triglycerides and high density lipoproteins were reduced in treated dogs. Surprisingly, a transient impairment in reversal learning was observed, suggesting prefrontal dysfunction. Spatial memory remained unchanged up to over a year of treatment. The lack of cognitive benefits of treatment was also reflected by a lack of reduction in plasma, cerebrospinal fluid and brain A β . Interestingly, BACE1 protein level was decreased in the brains of atorvastatin-treated dogs. This intriguing outcome may suggest that statins might be more useful to prevent the production of $A\beta$ through lowering BACE1 if started in animals in middle age, consistent with human studies indicating that middle-aged individuals using statins are protected from AD. Aged dogs are a unique model that may provide novel insights and translational data to predict outcomes of statin use in human clinical trials.

Metal Chelators and Reducing Aβ Aggregation

Preventing $A\beta$ aggregation may also be a promising approach to AD prevention. Metal ions such as zinc (Zn^{2+}) , Copper (Cu^{2+}) and iron (Fe³⁺) rapidly induce A β aggregation [134-137]. Further, Fe³⁺, Zn²⁺ and Cu²⁺ abnormally accumulate in the brains of AD patients [138]. Accumulation of these metals increases with disease progression, and high levels are found in AB plaques [139]. Additionally, studies suggest that the presence of Cu²⁺ is required for beta-amyloid aggregation and neurotoxicity [140]. A promising therapeutic approach to AD prevention involves the administration of agents to chelate metal ions and remove them from the blood and brain, which may result in A β disaggregation and clearance. For example, in a study of transgenic AD mice (Tg2576), the Cu^{2+} and Zn²⁺chelator clioquinol reduced metal ion accumulation in the brain, reduced A β burden by 49% [141] and improved cognitive performance [142]. The use of clioquinol as a treatment for AD has gone to phase II clinical trials [143], with inconclusive results [144, 145]. Following cliquinol administration, patients with mild AD had decreased plasma $A\beta_{42}$ with no cognitive benefit, while patients with severe AD showed cognitive benefit with no decrease in plasma A β_{42} . Questions of efficacy and safety remain unanswered, in part due to the small sample size and incomplete understanding of the mechanism of clioquinol in decreasing $A\beta$.

Metal ion chelators have potential in AD therapy, but remain controversial. Manipulating essential metals in the CNS can have neurotoxic effects and it is important to examine the efficacy and pathological mechanism of a potential therapy in more than one animal model prior to clinical trials. Additionally, it is important to test chelation therapies in an animal model that naturally exhibits $A\beta$ pathology, $A\beta$ associated cognitive decline and metal dyshomeostasis with age, in order to offer insight into the mechanisms, efficacy and safety of these interventions. Importantly, dogs exhibit these characteristics and have increased metal ions in the brain with age [146].

Aβ Vaccination and Increased Clearance of Aβ

As mentioned previously, another approach to reducing A β and potential effects on cognition is to clear pre-existing or new deposits. In transgenic mouse models of AD, deposition of A β was prevented or significantly reduced by vaccination with fibrillar A $\beta_{1.42}$ [147-150]. In parallel with A β reduction, learning and memory was improved by either active [148, 149, 151] or passive immunization [152-155].

Aged dogs may be very useful for assessing Aβ-targeted immunization therapies. Using an active immunization approach, dogs were injected with fibrillar A $\beta_{1.42}$ formulated with aluminum hydroxide, an adjuvant safe for use in humans [156]. Over a 2 year period of monthly vaccinations, surprisingly little cognitive improvement was observed on multiple measures of learning and memory, using tasks that are both age and intervention-sensitive. Interestingly, after 22 months of treatment, a significant improvement on reversal learning in treated animals was observed. Overall, error scores in control dogs increased over time, reflecting both an increase in task difficulty and the aging process. However, error scores were differentially affected by treatment, given that animals immunized with fibrillar $A\beta$ showed maintenance of reversal learning ability over time, suggesting maintained frontal lobe function. These results suggest that immunization with fibrillar $A\beta_{1-42}$ leads to improved and maintained executive function in aged dogs, when both pre-existing $A\beta$ and cognitive deficits are typically present. At the end of the study, $A\beta$ plaque accumulation, soluble and insoluble $A\beta_{1-40}$ and $A\beta_{1-42}$ were measured. Reductions in $A\beta$ were observed in multiple brain regions that are essential for learning and memory, including the prefrontal, parietal, occipital and entorhinal cortex.

Studies in higher mammalian species with disease characteristics that naturally parallel those in human AD may be a critically important step in the process of determining whether a drug should be taken to a clinical trial. For example, the promise of A β vaccination was derived from research in the transgenic mouse models of AD and rapidly translated into a human clinical trial. However, the clinical trial in patients with AD using a similar formulation of immunizing with fibrillar A β_{1-42} (AN1792 study) lead to an unexpected adverse event and an early halt to the study [157], which was not predicted from the mouse studies. Further, the clinical outcomes were not as robust as observed in transgenic mice. In the Swiss cohort of the trial, function was maintained on a global test of cognition (Mini-Mental State Examination) and on a hippocampal-dependent task (visual paired associated test) in vaccinated individuals who developed antibodies capable of binding to plaques [158]. However, in a second, larger study and in contrast to the predicted outcomes based on work in transgenic mice, no differences were observed on several cognitive and disability scales between treated and untreated patients [159]. One promising outcome was that 12 months after treatment, the composite score from a battery of neuropsychological tests indicated less severe memory decline [159]. Eight patients enrolled in the AN1792 study have been autopsied and show A β plaque reduction without any effect on the extent of NFTs or CAA [160-162]. Interestingly, in a case report by Masliah et al., (2005), the frontal cortex showed the largest response to immunotherapy [162]. Notably, the decreased A β pathology persisted 5 years after the last vaccination [163], although reduced brain AB did not slow AD progression and 7 these patients had severe end stage dementia prior to death. These findings parallel observations in the dog vaccination study, further emphasizing the need to test therapeutics in natural models of aging and AD.

Although there were several important differences between the dog and human vaccination studies [reviewed in 156]), dog studies suggest that reducing plaque accumulation or total A β in dogs with pre-existing pathology may be insufficient to restore neuronal function without directly targeting neuron health. Evidence from the previously discussed antioxidant and behavior enrichment study suggest alternative pathways that might be used in combination with A β vaccination to improve neuronal health. For example, significant improvements in cognition may be achieved by combining A β immunotherapy with either behavioral enrichment or an antioxidant diet to restore neuron health after A β removal.

SUMMARY

AD is a complex disease that remains a challenge in terms of developing therapeutics for clinical trials. To date, no disease pathology modifying therapies are commercially available for AD. Although mice and other rodent models are invaluable in learning about the mechanistic pathways involved in AD pathogenesis and identifying therapeutic targets, these studies should be extended to natural models to design safe and effective therapeutic strategies.

Because AD affects multiple pathways, therapeutic strategies may need to target the disease using parallel approaches. As discussed above, this can be achieved by combining therapeutics. For example, it may be beneficial to pair therapies that increase A β clearance with those that repair neuron health and attenuate oxidative stress. The canine model complements other animal models of AD and continues to be a beneficial system in which to test the efficacy and safety of therapeutics or preventative approaches for AD.

ACKNOWLEDGEMENTS

Funding provided by NIH/NIA AG031764 and AG032550.

REFERENCES

- Braak, H.; Braak, E. Neuropathological stageing of Alzheimerrelated changes. *Acta Neuropathol.*, **1991**, 82(4), 239-259.
- [2] Braak, H.; Braak, E.; Bohl, J. Staging of Alzheimer-related cortical destruction. *Eur. Neurol.*, **1993**, *33*(6), 403-408.
- [3] Khachaturian, Z.S. Diagnosis of Alzheimer's disease. Arch. Neurol., 1985, 42(11), 1097-1105.
- [4] Evans, D.A.; Funkenstein, H.H.; Albert, M.S.; Scherr, P.A.; Cook, N.R.; Chown, M.J.; Hebert, L.E.; Hennekens, C.H.; Taylor, J.O. Prevalence of Alzheimer's disease in a community population of older persons. Higher than previously reported. *JAMA*, **1989**, 262(18), 2551-2556.
- [5] Mirra, S.S.; Heyman, A.; McKeel, D.; Sumi, S.M.; Crain, B.J.; Brownlee, L.M.; Vogel, F.S.; Hughes, J.P.; van Belle, G.; Berg, L. The Consortium to Establish a Registry for Alzheimer's Disease (CERAD). Part II. Standardization of the neuropathologic assessment of Alzheimer's disease. *Neurology*, **1991**, *41*(4), 479-486.
- [6] Murphy, M.P.; LeVine, H., 3rd. Alzheimer's disease and the amyloid-beta peptide. J. Alzheimers Dis., 2010, 19(1), 311-323.
- [7] Selkoe, D.J. Amyloid beta-protein and the genetics of Alzheimer's disease. J. Biol. Chem., 1996, 271(31), 18295-18298.
- [8] Selkoe, D.J. Alzheimer's disease: genes, proteins, and therapy. *Physiol. Rev.*, 2001, 81(2), 741-766.
- [9] Antonarakis, S.E.; Lyle, R.; Dermitzakis, E.T.; Reymond, A.; Deutsch, S. Chromosome 21 and down syndrome: from genomics to pathophysiology. *Nat. Rev. Genet.*, 2004, 5(10), 725-738.
- [10] Lott, I.T.; Head, E.; Doran, E.; Busciglio, J. Beta-amyloid, oxidative stress and down syndrome. *Curr. Alzheimer Res.*, 2006, 3(5), 521-528.
- [11] Leverenz, J.B.; Raskind, M.A. Early amyloid deposition in the medial temporal lobe of young Down syndrome patients: a regional quantitative analysis. *Exp. Neurol.*, **1998**, *150*(2), 296-304.
- [12] Giaccone, G.; Tagliavini, F.; Linoli, G.; Bouras, C.; Frigerio, L.; Frangione, B.; Bugiani, O. Down patients: extracellular preamyloid deposits precede neuritic degeneration and senile plaques. *Neurosci. Lett.*, **1989**, 97(1-2), 232-238.
- [13] Mann, D.M. Cerebral amyloidosis, ageing and Alzheimer's disease; a contribution from studies on Down's syndrome. *Neurobiol. Aging*, **1989**, *10*(5), 397-399; discussion 412-394.
- [14] Iwatsubo, T.; Mann, D.M.; Odaka, A.; Suzuki, N.; Ihara, Y. Amyloid beta protein (A beta) deposition: A beta 42(43) precedes A beta 40 in Down syndrome. *Ann. Neurol.*, **1995**, *37*(3), 294-299.
- [15] Lemere, C.A.; Blusztajn, J.K.; Yamaguchi, H.; Wisniewski, T.; Saido, T.C.; Selkoe, D.J. Sequence of deposition of heterogeneous

amyloid beta-peptides and APO E in Down syndrome: implications for initial events in amyloid plaque formation. *Neurobiol. Dis.*, **1996**, 3(1), 16-32.

- [16] Goate, A.; Chartier-Harlin, M.C.; Mullan, M.; Brown, J.; Crawford, F.; Fidani, L.; Giuffra, L.; Haynes, A.; Irving, N.; James, L.; Mant, R.; Newton, P.; Rooke, K.; Roques, P.; Talbot, C.; Pericak-Vance, M.; Roses, A.; Williamson, R.; Rossor, M.; Owen, M.; Hardy, J. Segregation of a missense mutation in the amyloid precursor protein gene with familial Alzheimer's disease. *Nature*, **1991**, *349*(6311), 704-706.
- [17] Chartier-Harlin, M.C.; Crawford, F.; Houlden, H.; Warren, A.; Hughes, D.; Fidani, L.; Goate, A.; Rossor, M.; Roques, P.; Hardy, J.; Mullan, M. Early-onset Alzheimer's disease caused by mutations at codon 717 of the beta-amyloid precursor protein gene. *Nature*, **1991**, *353*(6347), 844-846.
- [18] Murrell, J.; Farlow, M.; Ghetti, B.; Benson, M.D. A mutation in the amyloid precursor protein associated with hereditary Alzheimer's disease. *Science*, **1991**, *254*(5028), 97-99.
- [19] Hendriks, L.; van Duijn, C.M.; Cras, P.; Cruts, M.; Van Hul, W.; van Harskamp, F.; Warren, A.; McInnis, M.G.; Antonarakis, S.E.; Martin, J.J.; Hofman, A.; van Broeckhoven, C. Presenile dementia and cerebral haemorrhage linked to a mutation at codon 692 of the beta-amyloid precursor protein gene. *Nat Genet*, **1992**, *1*(3), 218-221.
- [20] Rovelet-Lecrux, A.; Hannequin, D.; Raux, G.; Le Meur, N.; Laquerriere, A.; Vital, A.; Dumanchin, C.; Feuillette, S.; Brice, A.; Vercelletto, M.; Dubas, F.; Frebourg, T.; Campion, D. APP locus duplication causes autosomal dominant early-onset Alzheimer disease with cerebral amyloid angiopathy. *Nat. Genet.*, **2006**, *38*(1), 24-26.
- [21] Mawuenyega, K.G.; Sigurdson, W.; Ovod, V.; Munsell, L.; Kasten, T.; Morris, J.C.; Yarasheski, K.E.; Bateman, R.J. Decreased clearance of CNS beta-amyloid in Alzheimer's disease. *Science*, 2010, 330(6012), 1774.
- [22] Hardy, J.A.; Higgins, G.A. Alzheimer's disease: the amyloid cascade hypothesis. *Science*, **1992**, 256(5054), 184-185.
- [23] Elder, G.A.; Gama Sosa, M.A.; De Gasperi, R. Transgenic mouse models of Alzheimer's disease. *Mt Sinai J. Med.*, 2010, 77(1), 69-81.
- [24] Gotz, J.; Ittner, L.M. Animal models of Alzheimer's disease and frontotemporal dementia. *Nat. Rev. Neurosci.*, 2008, 9(7), 532-544.
- [25] Albert, M.S.; Funkenstein, H.H. Disorders of the Nervous System. In: *Nervous System, 2nd ed.* Asburg, A.K., McKhanney, G.M., McDonald, W.I., Eds.; Saunders Inc.: Philadelphia; **1992**, pp. 598-611.
- [26] Petersen, R.C.; Smith, G.E.; Waring, S.C.; Ivnik, R.J.; Kokmen, E.; Tangelos, E.G. Aging, memory, and mild cognitive impairment. *Int. Psychogeriatr.*, **1997**, 9(Suppl 1), 65-69.
- [27] Petersen, R.C.; Smith, G.E.; Waring, S.C.; Ivnik, R.J.; Tangalos, E.G.; Kokmen, E. Mild cognitive impairment: clinical characterization and outcome. *Arch. Neurol.*, **1999**, *56*(3), 303-308.
- [28] Joseph, J.; Cole, G.; Head, E.; Ingram, D. Nutrition, brain aging, and neurodegeneration. J. Neurosci., 2009, 29(41), 12795-12801.
- [29] Zawia, N.H.; Lahiri, D.K.; Cardozo-Pelaez, F. Epigenetics, oxidative stress, and Alzheimer disease. *Free Radic. Biol. Med.*, 2009, 46(9), 1241-1249.
- [30] Morris, J.C.; Storandt, M.; Miller, J.P.; McKeel, D.W.; Price, J.L.; Rubin, E.H.; Berg, L. Mild cognitive impairment represents earlystage Alzheimer disease. *Arch. Neurol.*, 2001, 58(3), 397-405.
- [31] Knopman, D.S. The initial recognition and diagnosis of dementia. *Am. J. Med.*, **1998**, *104*(4A), 2S-12S; discussion 39S-42S.
- [32] Brizzee, K.R.; Ordy, J.M.; Hofer, H.; Kaack, B. Animal models for the study of brain disease and aging changes in teh brain. In: *Alzheimer's disease: Senile dementia and related disorders*, Katzman, R.; Terry, R.D.; Bick, K.L.; Eds.; Raven: New York; 1978, pp. 515-554.
- [33] Mosier, J.E. Effect of aging on body systems of the dog. Vet. Clin. North Am. Small Anim. Pract., 1989, 19(1), 1-12.
- [34] Cummings, B.J.; Head, E.; Ruehl, W.; Milgram, N.W.; Cotman, C.W. The canine as an animal model of human aging and dementia. *Neurobiol. Aging*, **1996**, *17*(2), 259-268.
- [35] Blizard, D.A.; Klein, L.C.; Cohen, R.; McClearn, G.E. A novel mouse-friendly cognitive task suitable for use in aging studies. *Behav. Genet.*, 2003, 33(2), 181-189.
- [36] Milgram, N.W.; Head, E.; Weiner, E.; Thomas, E. Cognitive functions and aging in the dog: acquisition of nonspatial visual tasks. *Behav. Neurosci.*, **1994**, *108*(1), 57-68.

- [37] Cotman, C.W.; Head, E. The canine (dog) model of human aging and disease: dietary, environmental and immunotherapy approaches. J. Alzheimers Dis., 2008, 15(4), 685-707.
- [38] Adams, B.; Chan, A.; Callahan, H.; Milgram, N.W. The canine as a model of human cognitive aging: recent developments. *Prog. Neuropsychopharmacol. Biol. Psychiatry*, 2000, 24(5), 675-692.
- [39] Head, E.; Mehta, R.; Hartley, J.; Kameka, M.; Cummings, B.J.; Cotman, C.W.; Ruehl, W.W.; Milgram, N.W. Spatial learning and memory as a function of age in the dog. *Behav. Neurosci.*, **1995**, *109*(5), 851-858.
- [40] Rapp, P.R. Visual discrimination and reversal learning in the aged monkey (Macaca mulatta). *Behav. Neurosci.*, **1990**, *104*(6), 876-884.
- [41] Rapp, P.R.; Amaral, D.G. Evidence for task-dependent memory dysfunction in the aged monkey. J. Neurosci., 1989, 9(10), 3568-3576.
- [42] Rapp, P.R.; Amaral, D.G. Recognition memory deficits in a subpopulation of aged monkeys resemble the effects of medial temporal lobe damage. *Neurobiol. Aging*, **1991**, *12*(5), 481-486.
- [43] Walker, L.C.; Kitt, C.A.; Struble, R.G.; Wagster, M.V.; Price, D.L.; Cork, L.C. The neural basis of memory decline in aged monkeys. *Neurobiol. Aging*, **1988**, 9(5-6), 657-666.
- [44] Davis, R.T. Old monkey behavior. Exp. Gerontol., 1978, 13(3-4), 237-250.
- [45] Russell, M.J.; Bobik, M.; White, R.G.; Hou, Y.; Benjamin, S.A.; Geddes, J.W. Age-specific onset of beta-amyloid in beagle brains. *Neurobiol. Aging*, **1996**, *17*(2), 269-273.
- [46] Bons, N.; Mestre, N.; Petter, A. Senile plaques and neurofibrillary changes in the brain of an aged lemurian primate, Microcebus murinus. *Neurobiol. Aging*, 1992, 13(1), 99-105.
- [47] Selkoe, D.J.; Bell, D.S.; Podlisny, M.B.; Price, D.L.; Cork, L.C. Conservation of brain amyloid proteins in aged mammals and humans with Alzheimer's disease. *Science*, **1987**, 235(4791), 873-877.
- [48] Cummings, B.J.; Head, E.; Afagh, A.J.; Milgram, N.W.; Cotman, C.W. Beta-amyloid accumulation correlates with cognitive dysfunction in the aged canine. *Neurobiol. Learn Mem.*, **1996**, 66(1), 11-23.
- [49] Cummings, B.J.; Su, J.H.; Cotman, C.W.; White, R.; Russell, M.J. Beta-amyloid accumulation in aged canine brain: a model of early plaque formation in Alzheimer's disease. *Neurobiol. Aging*, **1993**, *14*(6), 547-560.
- [50] Head, E.; McCleary, R.; Hahn, F.F.; Milgram, N.W.; Cotman, C.W. Region-specific age at onset of beta-amyloid in dogs. *Neurobiol. Aging*, 2000, 21(1), 89-96.
- [51] Head, E.; Torp, R. Insights into Abeta and presenilin from a canine model of human brain aging. *Neurobiol. Dis.*, 2002, 9(1), 1-10.
- [52] Giaccone, G.; Verga, L.; Finazzi, M.; Pollo, B.; Tagliavini, F.; Frangione, B.; Bugiani, O. Cerebral preamyloid deposits and congophilic angiopathy in aged dogs. *Neurosci. Lett.*, **1990**, *114*(2), 178-183.
- [53] Morys, J.; Narkiewicz, O.; Maciejewska, B.; Wegiel, J.; Wisniewski, H.M. Amyloid deposits and loss of neurones in the claustrum of the aged dog. *Neuroreport*, **1994**, 5(14), 1825-1828.
- [54] Okuda, R.; Uchida, K.; Tateyama, S.; Yamaguchi, R.; Nakayama, H.; Goto, N. The distribution of amyloid beta precursor protein in canine brain. *Acta Neuropathol.*, **1994**, 87(2), 161-167.
- [55] Russell, M.J.; White, R.; Patel, E.; Markesbery, W.R.; Watson, C.R.; Geddes, J.W. Familial influence on plaque formation in the beagle brain. *Neuroreport*, **1992**, *3*(12), 1093-1096.
- [56] Uchida, K.; Tani, Y.; Uetsuka, K.; Nakayama, H.; Goto, N. Immunohistochemical studies on canine cerebral amyloid angiopathy and senile plaques. J. Vet. Med. Sci, 1992, 54(4), 659-667.
- [57] Markesbery, W.R.; Schmitt, F.A.; Kryscio, R.J.; Davis, D.G.; Smith, C.D.; Wekstein, D.R. Neuropathologic substrate of mild cognitive impairment. *Arch. Neurol.*, **2006**, *63*(1), 38-46.
- [58] Morris, J.C.; Storandt, M.; McKeel, D.W., Jr.; Rubin, E.H.; Price, J.L.; Grant, E.A.; Berg, L. Cerebral amyloid deposition and diffuse plaques in "normal" aging: Evidence for presymptomatic and very mild Alzheimer's disease. *Neurology*, **1996**, *46*(3), 707-719.
- [59] Ishihara, T.; Gondo, T.; Takahashi, M.; Uchino, F.; Ikeda, S.; Allsop, D.; Imai, K. Immunohistochemical and immunoelectron microscopical characterization of cerebrovascular and senile plaque amyloid in aged dogs' brains. *Brain Res.*, **1991**, *548*(1-2), 196-205.

- [60] Thal, D.R.; Rub, U.; Orantes, M.; Braak, H. Phases of A betadeposition in the human brain and its relevance for the development of AD. *Neurology*, 2002, 58(12), 1791-1800.
- [61] Wisniewski, H.; Johnson, A.B.; Raine, C.S.; Kay, W.J.; Terry, R.D. Senile plaques and cerebral amyloidosis in aged dogs. A histochemical and ultrastructural study. *Lab. Invest.*, **1970**, *23*(3), 287-296.
- [62] Attems, J. Sporadic cerebral amyloid angiopathy: pathology, clinical implications, and possible pathomechanisms. Acta Neuropathol., 2005, 110(4), 345-359.
- [63] Attems, J.; Jellinger, K.A.; Lintner, F. Alzheimer's disease pathology influences severity and topographical distribution of cerebral amyloid angiopathy. *Acta Neuropathol.*, 2005, 110(3), 222-231.
- [64] Herzig, M.C.; Van Nostrand, W.E.; Jucker, M. Mechanism of cerebral beta-amyloid angiopathy: murine and cellular models. *Brain Pathol.*, 2006, 16(1), 40-54.
- [65] Prior, R.; D'Urso, D.; Frank, R.; Prikulis, I.; Pavlakovic, G. Loss of vessel wall viability in cerebral amyloid angiopathy. *Neuroreport*, 1996, 7(2), 562-564.
- [66] Tian, J.; Shi, J.; Mann, D.M. Cerebral amyloid angiopathy and dementia. *Panminerva Med.*, 2004, 46(4), 253-264.
- [67] Uchida, K.; Nakayama, H.; Goto, N. Pathological studies on cerebral amyloid angiopathy, senile plaques and amyloid deposition in visceral organs in aged dogs. J. Vet. Med. Sci., 1991, 53(6), 1037-1042.
- [68] Selkoe, D.J. Soluble oligomers of the amyloid beta-protein impair synaptic plasticity and behavior. *Behav. Brain Res.*, 2008, 192(1), 106-113.
- [69] Tomic, J.L.; Pensalfini, A.; Head, E.; Glabe, C.G. Soluble fibrillar oligomer levels are elevated in Alzheimer's disease brain and correlate with cognitive dysfunction. *Neurobiol. Dis.*, 2009, 35(3), 352-358.
- [70] Mc Donald, J.M.; Savva, G.M.; Brayne, C.; Welzel, A.T.; Forster, G.; Shankar, G.M.; Selkoe, D.J.; Ince, P.G.; Walsh, D.M. The presence of sodium dodecyl sulphate-stable Abeta dimers is strongly associated with Alzheimer-type dementia. *Brain*, 2010, *133*(Pt 5), 1328-1341.
- [71] Pop, V.; Head, E.; Berchtold, N.C.; Glabe, C.G.; Studzinski, C.M.; Weidner, A.M.; Murphy, M.P.; Cotman, C.W. Abeta aggregation profiles and shifts in APP processing favor amyloidogenesis in canines. *Neurobiol. Aging*, **2010** [Epub a head of print]. doi:10. 1016/j.neurobiolaging.2010.02.008
- [72] Pugliese, M.; Gangitano, C.; Ceccariglia, S.; Carrasco, J.L.; Del Fa, A.; Rodriguez, M.J.; Michetti, F.; Mascort, J.; Mahy, N. Canine cognitive dysfunction and the cerebellum: acetylcholinesterase reduction, neuronal and glial changes. *Brain Res.*, **2007**, *1139*, 85-94.
- [73] Dringenberg, H.C. Alzheimer's disease: more than a 'cholinergic disorder' - evidence that cholinergic-monoaminergic interactions contribute to EEG slowing and dementia. *Behav. Brain Res.*, 2000, 115(2), 235-249.
- [74] Grudzien, A.; Shaw, P.; Weintraub, S.; Bigio, E.; Mash, D.C.; Mesulam, M.M. Locus coeruleus neurofibrillary degeneration in aging, mild cognitive impairment and early Alzheimer's disease. *Neurobiol. Aging*, 2007, 28(3), 327-335.
- [75] Insua, D.; Suarez, M.L.; Santamarina, G.; Sarasa, M.; Pesini, P. Dogs with canine counterpart of Alzheimer's disease lose noradrenergic neurons. *Neurobiol. Aging*, 2010, 31(4), 625-635.
- [76] Wegiel, J.; Kuchna, I.; Nowicki, K.; Frackowiak, J.; Mazur-Kolecka, B.; Imaki, H.; Mehta, P.D.; Silverman, W.P.; Reisberg, B.; Deleon, M.; Wisniewski, T.; Pirttilla, T.; Frey, H.; Lehtimaki, T.; Kivimaki, T.; Visser, F.E.; Kamphorst, W.; Potempska, A.; Bolton, D.; Currie, J.R.; Miller, D.L. Intraneuronal Abeta immunoreactivity is not a predictor of brain amyloidosis-beta or neurofibrillary degeneration. *Acta Neuropathol.*, **2007**, *113*(4), 389-402.
- [77] Hartley, D.M.; Walsh, D.M.; Ye, C.P.; Diehl, T.; Vasquez, S.; Vassilev, P.M.; Teplow, D.B.; Selkoe, D.J. Protofibrillar intermediates of amyloid beta-protein induce acute electrophysiological changes and progressive neurotoxicity in cortical neurons. J. Neurosci., 1999, 19(20), 8876-8884.
- [78] Walsh, D.M.; Hartley, D.M.; Kusumoto, Y.; Fezoui, Y.; Condron, M.M.; Lomakin, A.; Benedek, G.B.; Selkoe, D.J.; Teplow, D.B. Amyloid beta-protein fibrillogenesis. Structure and biological

activity of protofibrillar intermediates. J. Biol. Chem., 1999, 274(36), 25945-25952.

- [79] Lambert, M.P.; Barlow, A.K.; Chromy, B.A.; Edwards, C.; Freed, R.; Liosatos, M.; Morgan, T.E.; Rozovsky, I.; Trommer, B.; Viola, K.L.; Wals, P.; Zhang, C.; Finch, C.E.; Krafft, G.A.; Klein, W.L. Diffusible, nonfibrillar ligands derived from Abeta1-42 are potent central nervous system neurotoxins. *Proc. Natl. Acad. Sci. U. S. A.*, **1998**, *95*(11), 6448-6453.
- [80] Lesne, S.; Koh, M.T.; Kotilinek, L.; Kayed, R.; Glabe, C.G.; Yang, A.; Gallagher, M.; Ashe, K.H. A specific amyloid-beta protein assembly in the brain impairs memory. *Nature*, **2006**, *440*(7082), 352-357.
- [81] Roher, A.E.; Chaney, M.O.; Kuo, Y.M.; Webster, S.D.; Stine, W.B.; Haverkamp, L.J.; Woods, A.S.; Cotter, R.J.; Tuohy, J.M.; Krafft, G.A.; Bonnell, B.S.; Emmerling, M.R. Morphology and toxicity of Abeta-(1-42) dimer derived from neuritic and vascular amyloid deposits of Alzheimer's disease. J. Biol. Chem., 1996, 271(34), 20631-20635.
- [82] LaFerla, F.M.; Green, K.N.; Oddo, S. Intracellular amyloid-beta in Alzheimer's disease. *Nat. Rev. Neurosci.*, 2007, 8(7), 499-509.
- [83] Middleton, L.E.; Yaffe, K. Promising strategies for the prevention of dementia. Arch. Neurol., 2009, 66(10), 1210-1215.
- [84] Lee, J.W.; Lee, Y.K.; Ban, J.O.; Ha, T.Y.; Yun, Y.P.; Han, S.B.; Oh, K.W.; Hong, J.T. Green tea (-)-epigallocatechin-3-gallate inhibits beta-amyloid-induced cognitive dysfunction through modification of secretase activity via inhibition of ERK and NFkappaB pathways in mice. *J. Nutr.*, 2009, *139*(10), 1987-1993.
- [85] Lim, G.P.; Calon, F.; Morihara, T.; Yang, F.; Teter, B.; Ubeda, O.; Salem, N., Jr.; Frautschy, S.A.; Cole, G.M. A diet enriched with the omega-3 fatty acid docosahexaenoic acid reduces amyloid burden in an aged Alzheimer mouse model. *J. Neurosci.*, **2005**, *25*(12), 3032-3040.
- [86] Lazarov, O.; Robinson, J.; Tang, Y.P.; Hairston, I.S.; Korade-Mirnics, Z.; Lee, V.M.; Hersh, L.B.; Sapolsky, R.M.; Mirnics, K.; Sisodia, S.S. Environmental enrichment reduces Abeta levels and amyloid deposition in transgenic mice. *Cell*, 2005, *120*(5), 701-713.
- [87] Churchill, J.D.; Galvez, R.; Colcombe, S.; Swain, R.A.; Kramer, A.F.; Greenough, W.T. Exercise, experience and the aging brain. *Neurobiol. Aging*, 2002, 23(5), 941-955.
- [88] Cotman, C.W.; Berchtold, N.C. Exercise: a behavioral intervention to enhance brain health and plasticity. *Trends Neurosci.*, 2002, 25(6), 295-301.
- [89] Engelhart, M.J.; Geerlings, M.I.; Ruitenberg, A.; van Swieten, J.C.; Hofman, A.; Witteman, J.C.; Breteler, M.M. Dietary intake of antioxidants and risk of Alzheimer disease. *JAMA*, 2002, 287(24), 3223-3229.
- [90] Morris, M.C.; Evans, D.A.; Bienias, J.L.; Tangney, C.C.; Wilson, R.S. Vitamin E and cognitive decline in older persons. Arch. Neurol., 2002, 59(7), 1125-1132.
- [91] Erickson, K.I.; Kramer, A.F. Aerobic exercise effects on cognitive and neural plasticity in older adults. *Br. J. Sports Med.*, 2009, 43(1), 22-24.
- [92] Willis, S.L.; Tennstedt, S.L.; Marsiske, M.; Ball, K.; Elias, J.; Koepke, K.M.; Morris, J.N.; Rebok, G.W.; Unverzagt, F.W.; Stoddard, A.M.; Wright, E. Long-term effects of cognitive training on everyday functional outcomes in older adults. *JAMA*, 2006, 296(23), 2805-2814.
- [93] Ball, K.; Berch, D.B.; Helmers, K.F.; Jobe, J.B.; Leveck, M.D.; Marsiske, M.; Morris, J.N.; Rebok, G.W.; Smith, D.M.; Tennstedt, S.L.; Unverzagt, F.W.; Willis, S.L. Effects of cognitive training interventions with older adults: a randomized controlled trial. *JAMA*, 2002, 288(18), 2271-2281.
- [94] Milgram, N.W.; Zicker, S.C.; Head, E.; Muggenburg, B.A.; Murphey, H.; Ikeda-Douglas, C.J.; Cotman, C.W. Dietary enrichment counteracts age-associated cognitive dysfunction in canines. *Neurobiol. Aging*, **2002**, *23*(5), 737-745.
- [95] Milgram, N.W.; Head, E.; Zicker, S.C.; Ikeda-Douglas, C.J.; Murphey, H.; Muggenburg, B.; Siwak, C.; Tapp, D.; Cotman, C.W. Learning ability in aged beagle dogs is preserved by behavioral enrichment and dietary fortification: a two-year longitudinal study. *Neurobiol. Aging*, **2005**, 26(1), 77-90.
- [96] Pop, V.; Head, E.; Berchtold, N.C.; Glabe, C.; Studzinski, C.M.; Weidner, A.M.; Murphy, M.P.; Cotman, C.W. Beta-amyloid aggregation profiles and shifts in APP processing favor

amyloidogenesis in canines. *Neurobiol. Aging*, **2010**. doi:10.1016/j.neurobiolaging.2010.02.008

- [97] Fukumoto, H.; Cheung, B.S.; Hyman, B.T.; Irizarry, M.C. Betasecretase protein and activity are increased in the neocortex in Alzheimer disease. *Arch Neurol*, **2002**, *59*(9), 1381-1389.
- [98] Yang, L.B.; Lindholm, K.; Yan, R.; Citron, M.; Xia, W.; Yang, X.L.; Beach, T.; Sue, L.; Wong, P.; Price, D.; Li, R.; Shen, Y. Elevated beta-secretase expression and enzymatic activity detected in sporadic Alzheimer disease. *Nat. Med.*, **2003**, *9*(1), 3-4.
- [99] Li, R.; Lindholm, K.; Yang, L.B.; Yue, X.; Citron, M.; Yan, R.; Beach, T.; Sue, L.; Sabbagh, M.; Cai, H.; Wong, P.; Price, D.; Shen, Y. Amyloid beta peptide load is correlated with increased beta-secretase activity in sporadic Alzheimer's disease patients. *Proc. Natl. Acad. Sci. U. S. A.*, **2004**, *101*(10), 3632-3637.
- [100] Vassar, R.; Kovacs, D.M.; Yan, R.; Wong, P.C. The beta-secretase enzyme BACE in health and Alzheimer's disease: regulation, cell biology, function, and therapeutic potential. *J. Neurosci.*, 2009, 29(41), 12787-12794.
- [101] Stefansson, H.; Sigurdsson, E.; Steinthorsdottir, V.; Bjornsdottir, S.; Sigmundsson, T.; Ghosh, S.; Brynjolfsson, J.; Gunnarsdottir, S.; Ivarsson, O.; Chou, T.T.; Hjaltason, O.; Birgisdottir, B.; Jonsson, H.; Gudnadottir, V.G.; Gudmundsdottir, E.; Bjornsson, A.; Ingvarsson, B.; Ingason, A.; Sigfusson, S.; Hardardottir, H.; Harvey, R.P.; Lai, D.; Zhou, M.; Brunner, D.; Mutel, V.; Gonzalo, A.; Lemke, G.; Sainz, J.; Johannesson, G.; Andresson, T.; Gudbjartsson, D.; Manolescu, A.; Frigge, M.L.; Gurney, M.E.; Kong, A.; Gulcher, J.R.; Petursson, H.; Stefansson, K. Neuregulin 1 and susceptibility to schizophrenia. Am. J. Hum. Genet., 2002, 71(4), 877-892.
- [102] Gerlai, R.; Pisacane, P.; Erickson, S. Heregulin, but not ErbB2 or ErbB3, heterozygous mutant mice exhibit hyperactivity in multiple behavioral tasks. *Behav. Brain Res.*, 2000, 109(2), 219-227.
- [103] McConlogue, L.; Buttini, M.; Anderson, J.P.; Brigham, E.F.; Chen, K.S.; Freedman, S.B.; Games, D.; Johnson-Wood, K.; Lee, M.; Zeller, M.; Liu, W.; Motter, R.; Sinha, S. Partial reduction of BACE1 has dramatic effects on Alzheimer plaque and synaptic pathology in APP Transgenic Mice. J. Biol. Chem., 2007, 282(36), 26326-26334.
- [104] Jick, H.; Zornberg, G.L.; Jick, S.S.; Seshadri, S.; Drachman, D.A. Statins and the risk of dementia. *Lancet*, **2000**, *356*(9242), 1627-1631.
- [105] Wolozin, B.; Kellman, W.; Ruosseau, P.; Celesia, G.G.; Siegel, G. Decreased prevalence of Alzheimer disease associated with 3hydroxy-3-methyglutaryl coenzyme A reductase inhibitors. *Arch. Neurol.*, 2000, 57(10), 1439-1443.
- [106] Rockwood, K.; Kirkland, S.; Hogan, D.B.; MacKnight, C.; Merry, H.; Verreault, R.; Wolfson, C.; McDowell, I. Use of lipid-lowering agents, indication bias, and the risk of dementia in communitydwelling elderly people. *Arch. Neurol.*, **2002**, *59*(2), 223-227.
- [107] Hajjar, I.; Schumpert, J.; Hirth, V.; Wieland, D.; Eleazer, G.P. The impact of the use of statins on the prevalence of dementia and the progression of cognitive impairment. J. Gerontol. A Biol. Sci. Med. Sci., 2002, 57(7), M414-418.
- [108] Rodriguez, E.G.; Dodge, H.H.; Birzescu, M.A.; Stoehr, G.P.; Ganguli, M. Use of lipid-lowering drugs in older adults with and without dementia: a community-based epidemiological study. J. Am. Geriatr. Soc., 2002, 50(11), 1852-1856.
- [109] Dufouil, C.; Richard, F.; Fievet, N.; Dartigues, J.F.; Ritchie, K.; Tzourio, C.; Amouyel, P.; Alperovitch, A. APOE genotype, cholesterol level, lipid-lowering treatment, and dementia: the Three-City Study. *Neurology*, **2005**, *64*(9), 1531-1538.
- [110] Zamrini, E.; McGwin, G.; Roseman, J.M. Association between statin use and Alzheimer's disease. *Neuroepidemiology*, 2004, 23(1-2), 94-98.
- [111] Wolozin, B.; Wang, S.W.; Li, N.C.; Lee, A.; Lee, T.A.; Kazis, L.E. Simvastatin is associated with a reduced incidence of dementia and Parkinson's disease. *BMC Med*, **2007**, *5*, 20.
- [112] Li, G.; Higdon, R.; Kukull, W.A.; Peskind, E.; Van Valen Moore, K.; Tsuang, D.; van Belle, G.; McCormick, W.; Bowen, J.D.; Teri, L.; Schellenberg, G.D.; Larson, E.B. Statin therapy and risk of dementia in the elderly: a community-based prospective cohort study. *Neurology*, **2004**, *63*(9), 1624-1628.
- [113] Rea, T.D.; Breitner, J.C.; Psaty, B.M.; Fitzpatrick, A.L.; Lopez, O.L.; Newman, A.B.; Hazzard, W.R.; Zandi, P.P.; Burke, G.L.; Lyketsos, C.G.; Bernick, C.; Kuller, L.H. Statin use and the risk of

incident dementia: the Cardiovascular Health Study. Arch. Neurol., 2005, 62(7), 1047-1051.

- [114] Zandi, P.P.; Sparks, D.L.; Khachaturian, A.S.; Tschanz, J.; Norton, M.; Steinberg, M.; Welsh-Bohmer, K.A.; Breitner, J.C. Do statins reduce risk of incident dementia and Alzheimer disease? The Cache County Study. Arch. Gen. Psychiatry, 2005, 62(2), 217-224.
- [115] Haag, M.D.; Hofman, A.; Koudstaal, P.J.; Stricker, B.H.; Breteler, M.M. Statins are associated with a reduced risk of Alzheimer disease regardless of lipophilicity. The Rotterdam Study. J. Neurol. Neurosurg. Psychiatry, 2009, 80(1), 13-17.
- [116] Rockwood, K. Epidemiological and clinical trials evidence about a preventive role for statins in Alzheimer's disease. Acta Neurol. Scand Suppl., 2006, 185, 71-77.
- [117] Sparks, L. Statins and cognitive function. J. Neurol. Neurosurg. Psychiatry, 2009, 80(1), 1-2.
- [118] Simons, M.; Schwarzler, F.; Lutjohann, D.; von Bergmann, K.; Beyreuther, K.; Dichgans, J.; Wormstall, H.; Hartmann, T.; Schulz, J.B. Treatment with simvastatin in normocholesterolemic patients with Alzheimer's disease: A 26-week randomized, placebocontrolled, double-blind trial. *Ann. Neurol.*, **2002**, *52*(3), 346-350.
- [119] Sparks, D.L.; Connor, D.J.; Sabbagh, M.N.; Petersen, R.B.; Lopez, J.; Browne, P. Circulating cholesterol levels, apolipoprotein E genotype and dementia severity influence the benefit of atorvastatin treatment in Alzheimer's disease: results of the Alzheimer's Disease Cholesterol-Lowering Treatment (ADCLT) trial. Acta Neurol. Scand Suppl, 2006, 185, 3-7.
- [120] Sparks, D.L.; Sabbagh, M.N.; Connor, D.J.; Lopez, J.; Launer, L.J.; Petanceska, S.; Browne, P.; Wassar, D.; Johnson-Traver, S.; Lochhead, J.; Ziolkowski, C. Atorvastatin therapy lowers circulating cholesterol but not free radical activity in advance of identifiable clinical benefit in the treatment of mild-to-moderate AD. Curr. Alzheimer Res., 2005, 2(3), 343-353.
- [121] Sparks, D.L.; Sabbagh, M.N.; Connor, D.J.; Lopez, J.; Launer, L.J.; Browne, P.; Wasser, D.; Johnson-Traver, S.; Lochhead, J.; Ziolwolski, C. Atorvastatin for the treatment of mild to moderate Alzheimer disease: preliminary results. *Arch. Neurol.*, **2005**, *62*(5), 753-757.
- [122] Sparks, D.L.; Sabbagh, M.; Connor, D.; Soares, H.; Lopez, J.; Stankovic, G.; Johnson-Traver, S.; Ziolkowski, C.; Browne, P. Statin therapy in Alzheimer's disease. *Acta Neurol. Scand. Suppl*, 2006, 185, 78-86.
- [123] Pappolla, M.A. Statins, incident Alzheimer disease, change in cognitive function, and neuropathology. *Neurology*, **2008**, 71(24), **2020**; author reply 2020-2021.
- [124] Solomon, A.; Kareholt, I.; Ngandu, T.; Wolozin, B.; Macdonald, S.W.; Winblad, B.; Nissinen, A.; Tuomilehto, J.; Soininen, H.; Kivipelto, M. Serum total cholesterol, statins and cognition in nondemented elderly. *Neurobiol. Aging*, **2009**, *30*(6), 1006-1009.
- [125] Cramer, C.; Haan, M.N.; Galea, S.; Langa, K.M.; Kalbfleisch, J.D. Use of statins and incidence of dementia and cognitive impairment without dementia in a cohort study. *Neurology*, **2008**, *71*(5), 344-350.
- [126] Simons, M.; Keller, P.; Strooper, B.D.; Beyreuther, K.; Dotti, C.G.; Simons, K. Cholesterol depletion inhibits the generation of betaamyloid in hippocampal neurons. *Proc. Natl. Acad. Sci. USA*, 1998, 95, 6460-6464.
- [127] Hartmann, T. Cholesterol, A beta and Alzheimer's disease. *Trends Neurosci.*, 2001, 24(11 Suppl), S45-48.
- [128] Petanceska, S.S.; DeRosa, S.; Olm, V.; Diaz, N.; Sharma, A.; Thomas-Bryant, T.; Duff, K.; Pappolla, M.; Refolo, L.M. Statin therapy for Alzheimer's disease: will it work? *J. Mol. Neurosci.*, 2002, 19(1-2), 155-161.
- [129] Alberts, A.W. Lovastatin and simvastatin inhibitors of HMG CoA reductase and cholesterol biosynthesis. *Cardiology*, **1990**, 77(4), 14-21.
- [130] Fears, R.; Richards, D.H.; Ferres, H. The effect of compactin, a potent inhibitor of 3-hydroxy-3-methylgutaryl co-enzyme-A reductase activity, on cholesterogenesis and serum cholesterol levels in rats and chicks. *Atherosclerosis*, **1980**, *35*(4), 439-449.
- [131] Todd, P.A.; Goa, K.L. Simvastatin. A review of its pharmacological properties and therapeutic potential in hypercholesterolaemia. *Drugs*, **1990**, 40(4), 583-607.
- [132] Thelen, K.M.; Rentsch, K.M.; Gutteck, U.; Heverin, M.; Olin, M.; Andersson, U.; von Eckardstein, A.; Bjorkhem, I.; Lutjohann, D. Brain cholesterol synthesis in mice is affected by high dose of

simvastatin but not of pravastatin. J. Pharmacol. Exp. Ther., 2006, 316(3), 1146-1152.

- [133] Gerson, R.J.; MacDonald, J.S.; Alberts, A.W.; Kornbrust, D.J.; Majka, J.A.; Stubbs, R.J.; Bokelman, D.L. Animal safety and toxicology of simvastatin and related hydroxy-methylglutarylcoenzyme A reductase inhibitors. *Am. J. Med.*, **1989**, 87(4A), 28S-38S.
- [134] Bush, A.I.; Pettingell, W.H.; Multhaup, G.; d Paradis, M.; Vonsattel, J.P.; Gusella, J.F.; Beyreuther, K.; Masters, C.L.; Tanzi, R.E. Rapid induction of Alzheimer A beta amyloid formation by zinc. *Science*, **1994**, 265(5177), 1464-1467.
- [135] Bush, A.I.; Pettingell, W.H., Jr.; Paradis, M.D.; Tanzi, R.E. Modulation of A beta adhesiveness and secretase site cleavage by zinc. J. Biol. Chem., 1994, 269(16), 12152-12158.
- [136] Ha, C.; Ryu, J.; Park, C.B. Metal ions differentially influence the aggregation and deposition of Alzheimer's beta-amyloid on a solid template. *Biochemistry*, 2007, 46(20), 6118-6125.
- [137] Noy, D.; Solomonov, I.; Sinkevich, O.; Arad, T.; Kjaer, K.; Sagi, I. Zinc-amyloid beta interactions on a millisecond time-scale stabilize non-fibrillar Alzheimer-related species. J. Am. Chem. Soc., 2008, 130(4), 1376-1383.
- [138] Sensi, S.L.; Paoletti, P.; Bush, A.I.; Sekler, I. Zinc in the physiology and pathology of the CNS. *Nat. Rev. Neurosci.*, 2009, 10(11), 780-791.
- [139] Grundke-Iqbal, I.; Fleming, J.; Tung, Y.C.; Lassmann, H.; Iqbal, K.; Joshi, J.G. Ferritin is a component of the neuritic (senile) plaque in Alzheimer dementia. *Acta Neuropathol.*, **1990**, *81*(2), 105-110.
- [140] Quinn, J.F.; Crane, S.; Harris, C.; Wadsworth, T.L. Copper in Alzheimer's disease: too much or too little? *Expert Rev. Neurother.*, 2009, 9(5), 631-637.
- [141] Cherny, R.A.; Atwood, C.S.; Xilinas, M.E.; Gray, D.N.; Jones, W.D.; McLean, C.A.; Barnham, K.J.; Volitakis, I.; Fraser, F.W.; Kim, Y.; Huang, X.; Goldstein, L.E.; Moir, R.D.; Lim, J.T.; Beyreuther, K.; Zheng, H.; Tanzi, R.E.; Masters, C.L.; Bush, A.I. Treatment with a copper-zinc chelator markedly and rapidly inhibits beta-amyloid accumulation in Alzheimer's disease transgenic mice. *Neuron*, 2001, 30(3), 665-676.
- [142] Adlard, P.A.; Cherny, R.A.; Finkelstein, D.I.; Gautier, E.; Robb, E.; Cortes, M.; Volitakis, I.; Liu, X.; Smith, J.P.; Perez, K.; Laughton, K.; Li, Q.X.; Charman, S.A.; Nicolazzo, J.A.; Wilkins, S.; Deleva, K.; Lynch, T.; Kok, G.; Ritchie, C.W.; Tanzi, R.E.; Cappai, R.; Masters, C.L.; Barnham, K.J.; Bush, A.I. Rapid restoration of cognition in Alzheimer's transgenic mice with 8-hydroxy quinoline analogs is associated with decreased interstitial Abeta. *Neuron*, 2008, 59(1), 43-55.
- [143] Ritchie, C.W.; Bush, A.I.; Mackinnon, A.; Macfarlane, S.; Mastwyk, M.; MacGregor, L.; Kiers, L.; Cherny, R.; Li, Q.X.; Tammer, A.; Carrington, D.; Mavros, C.; Volitakis, I.; Xilinas, M.; Ames, D.; Davis, S.; Beyreuther, K.; Tanzi, R.E.; Masters, C.L. Metal-protein attenuation with iodochlorhydroxyquin (clioquinol) targeting Abeta amyloid deposition and toxicity in Alzheimer disease: a pilot phase 2 clinical trial. Arch. Neurol., 2003, 60(12), 1685-1691.
- [144] Jenagaratnam, L.; McShane, R. Clioquinol for the treatment of Alzheimer's Disease. *Cochrane Database Syst. Rev.*, 2006, (1), CD005380.
- [145] Sampson, E.; Jenagaratnam, L.; McShane, R. Metal protein attenuating compounds for the treatment of Alzheimer's disease. *Cochrane Database Syst. Rev.*, 2008, (1), CD005380.
- [146] Barden, H. The histochemical distribution and localization of copper, iron, neuromelanin and lysosomal enzyme activity in the brain of aging rhesus monkey and the dog. J. Neuropathol. Exp. Neurol., 1971, 30(4), 650-667.
- [147] Das, P.; Howard, V.; Loosbrock, N.; Dickson, D.; Murphy, M.P.; Golde, T.E. Amyloid-beta immunization effectively reduces amyloid deposition in FcRgamma-/- knock-out mice. *J. Neurosci.*, 2003, 23(24), 8532-8538.
- [148] Janus, C.; Pearson, J.; McLaurin, J.; Mathews, P.M.; Jiang, Y.; Schmidt, S.D.; Chishti, M.A.; Horne, P.; Heslin, D.; French, J.; Mount, H.T.; Nixon, R.A.; Mercken, M.; Bergeron, C.; Fraser, P.E.; St George-Hyslop, P.; Westaway, D. A beta peptide immunization reduces behavioural impairment and plaques in a model of Alzheimer's disease. *Nature*, **2000**, 408(6815), 979-982.

- [149] Morgan, D.; Diamond, D.M.; Gottschall, P.E.; Ugen, K.E.; Dickey, C.; Hardy, J.; Duff, K.; Jantzen, P.; DiCarlo, G.; Wilcock, D.; Connor, K.; Hatcher, J.; Hope, C.; Gordon, M.; Arendash, G.W. A beta peptide vaccination prevents memory loss in an animal model of Alzheimer's disease. *Nature*, **2000**, *408*(6815), 982-985.
- [150] Schenk, D., Barbour, R., Dunn, W., Gordon, G., Grajeda, H., Guido, T., Hu, K., Huang, J., Johnson-Wood, K., Khan, K., Kholodenko, D., Lee, M., Liao, Z., Lieberburg, I., Motter, R., Mutter, L., Soriano, F., Shopp, G., Vasquez, N., Vandervert, C., Walker, S., Wogulis, M., Yednock, T., Games, D., and Seubert, P. Immunization with amyloid-β attentuates Alzheimer-disease-like pathology in the PDAPP mouse. *Nature*, **1999**, *400*, 173-177.
- [151] Sigurdsson, E.M.; Knudsen, E.; Asuni, A.; Fitzer-Attas, C.; Sage, D.; Quartermain, D.; Goni, F.; Frangione, B.; Wisniewski, T. An attenuated immune response is sufficient to enhance cognition in an Alzheimer's disease mouse model immunized with amyloid-beta derivatives. J. Neurosci, 2004, 24(28), 6277-6282.
- [152] Dodart, J.C.; Bales, K.R.; Gannon, K.S.; Greene, S.J.; DeMattos, R.B.; Mathis, C.; DeLong, C.A.; Wu, S.; Wu, X.; Holtzman, D.M.; Paul, S.M. Immunization reverses memory deficits without reducing brain Abeta burden in Alzheimer's disease model. *Nat. Neurosci.*, 2002, 5(5), 452-457.
- [153] Kotilinek, L.A.; Bacskai, B.; Westerman, M.; Kawarabayashi, T.; Younkin, L.; Hyman, B.T.; Younkin, S.; Ashe, K.H. Reversible memory loss in a mouse transgenic model of Alzheimer's disease. *J. Neurosci.*, 2002, 22(15), 6331-6335.
- [154] Morley, J.E.; Farr, S.A.; Flood, J.F. Antibody to amyloid beta protein alleviates impaired acquisition, retention, and memory processing in SAMP8 mice. *Neurobiol. Learn Mem.*, 2002, 78(1), 125-138.
- [155] Wilcock, D.M.; Rojiani, A.; Rosenthal, A.; Levkowitz, G.; Subbarao, S.; Alamed, J.; Wilson, D.; Wilson, N.; Freeman, M.J.; Gordon, M.N.; Morgan, D. Passive amyloid immunotherapy clears amyloid and transiently activates microglia in a transgenic mouse model of amyloid deposition. *J. Neurosci.*, **2004**, *24*(27), 6144-6151.
- [156] Head, E.; Pop, V.; Vasilevko, V.; Hill, M.; Saing, T.; Sarsoza, F.; Nistor, M.; Christie, L.A.; Milton, S.; Glabe, C.; Barrett, E.; Cribbs, D. A two-year study with fibrillar beta-amyloid (Abeta) immunization in aged canines: effects on cognitive function and brain Abeta. J. Neurosci., 2008, 28(14), 3555-3566.
- [157] Orgogozo, J.M.; Gilman, S.; Dartigues, J.F.; Laurent, B.; Puel, M.; Kirby, L.C.; Jouanny, P.; Dubois, B.; Eisner, L.; Flitman, S.; Michel, B.F.; Boada, M.; Frank, A.; Hock, C. Subacute meningoencephalitis in a subset of patients with AD after Abeta42 immunization. *Neurology*, **2003**, *61*(1), 46-54.
- [158] Hock, C.; Konietzko, U.; Streffer, J.R.; Tracy, J.; Signorell, A.; Muller-Tillmanns, B.; Lemke, U.; Henke, K.; Moritz, E.; Garcia, E.; Wollmer, M.A.; Umbricht, D.; de Quervain, D.J.; Hofmann, M.; Maddalena, A.; Papassotiropoulos, A.; Nitsch, R.M. Antibodies against beta-amyloid slow cognitive decline in Alzheimer's disease. *Neuron*, **2003**, *38*(4), 547-554.
- [159] Gilman, S.; Koller, M.; Black, R.S.; Jenkins, L.; Griffith, S.G.; Fox, N.C.; Eisner, L.; Kirby, L.; Rovira, M.B.; Forette, F.; Orgogozo, J.M. Clinical effects of Abeta immunization (AN1792) in patients with AD in an interrupted trial. *Neurology*, **2005**, *64*(9), 1553-1562.
- [160] Ferrer, I.; Boada Rovira, M.; Sanchez Guerra, M.L.; Rey, M.J.; Costa-Jussa, F. Neuropathology and pathogenesis of encephalitis following amyloid-beta immunization in Alzheimer's disease. *Brain Pathol.*, 2004, 14(1), 11-20.
- [161] Nicoll, J.A.; Wilkinson, D.; Holmes, C.; Steart, P.; Markham, H.; Weller, R.O. Neuropathology of human Alzheimer disease after

Received: March 05, 2010

immunization with amyloid-beta peptide: a case report. *Nat. Med.*, **2003**, *9*(4), 448-452.

- [162] Masliah, E.; Hansen, L.; Adame, A.; Crews, L.; Bard, F.; Lee, C.; Seubert, P.; Games, D.; Kirby, L.; Schenk, D. Abeta vaccination effects on plaque pathology in the absence of encephalitis in Alzheimer disease. *Neurology*, **2005**, *64*(1), 129-131.
- [163] Holmes, C.; Boche, D.; Wilkinson, D.; Yadegarfar, G.; Hopkins, V.; Bayer, A.; Jones, R.W.; Bullock, R.; Love, S.; Neal, J.W.; Zotova, E.; Nicoll, J.A. Long-term effects of Abeta42 immunisation in Alzheimer's disease: follow-up of a randomised, placebo-controlled phase I trial. *Lancet*, **2008**, *372*(9634), 216-223.
- [164] Lai, Z.C.; Moss, M.B.; Killiany, R.J.; Rosene, D.L.; Herndon, J.G. Executive system dysfunction in the aged monkey: spatial and object reversal learning. *Neurobiol. Aging*, **1995**, *16*, 947-954.
- [165] Cronin-Golomb, A. Color vision, object recognition, and spatial localization in aging and Alzheimer's disease. In: *Functional Neurobiology of Aging*, Hof, P.R., Mobbs, C.V., Eds.; Academic Press: San Diego; **2001**, pp. 517-529.
- [166] Boutet, I.; Milgram, N.W.; Freedman, M. Cognitive decline and human (Homo sapiens) aging: an investigation using a comparative neuropsychological approach. J. Comp. Psychol., 2007, 121(3), 270-281.
- [167] Freedman, M.; Oscar-Berman, M. Spatial and visual learning deficits in Alzheimer's disease and Parkinson's disease. *Brain Cognit.*, 1989, 11, 114-126.
- [168] Emborg, M.E.; Ma, S.Y.; Mufson, E.J.; Levey, A.I.; Taylor, M.D.; Brown, W.D.; Holden, J.E.; Kordower, J.H. Age-related declines in nigral neuronal function correlate with motor impairments in rhesus monkeys. J. Comp. Neurol., 1998, 401(2), 253-265.
- [169] Kordower, J.H.; Liu, Y.T.; Winn, S.; Emerich, D.F. Encapsulated PC12 cell transplants into hemiparkinsonian monkeys: a behavioral, neuroanatomical, and neurochemical analysis. *Cell Transplant.*, **1995**, 4(2), 155-171.
- [170] Christie, L.A.; Studzinski, C.M.; Araujo, J.A.; Leung, C.S.; Ikeda-Douglas, C.J.; Head, E.; Cotman, C.W.; Milgram, N.W. A comparison of egocentric and allocentric age-dependent spatial learning in the beagle dog. *Prog. Neuropsychopharmacol. Biol. Psychiatry*, 2005, 29(3), 361-369.
- [171] Mishkin, M.; Delacour, J. An analysis of short-term visual memory in the monkey. J. Exper. Psychol: Anim. Behav. Proc., 1975, 1, 326-334.
- [172] Lezak, M.D.; Howieson, D.B.; Loring, D.W. Neuropsychological Assessment. Oxford University Press: New York, 2004.
- [173] Arnsten, A.F.T., and Goldman-Rakic, P.S. Alpha 2-adrenergic mechanisms in prefrontal cortex associated with cognitive decline in aged nonhuman primates. *Science*, **1985**, *230*, 1273-1276.
- [174] Kowalska, D.M. Effects of hippocampal lesions on spatial delayed responses in dog. *Hippocampus*, **1995**, *5*(4), 363-370.
- [175] Warren, J.M. The behavior of carnivores and primates with lesions in the prefrontal cortex. In: *The Frontal Granular Cortex and Behavior*, Warren, J.M., Akert, K., Eds.; McGraw-Hill Book Company: New York; **1964**, pp. 168-191.
- [176] Kramer, J.H.; Quitania, L. Bedside Frontal Lobe Testing. In: *The Human Frontal Lobes*, Miller, B.L., Cummings, J.L., Ed.; The Guilford Press: New York; 2007, pp. 279-291.
- [177] Rapp, P.R.; Kansky, M.T.; Eichenbaum, H. Learning and memory for hierarchical relationships in the monkey: effects of aging. *Behav. Neurosci.*, **1996**, *110*(5), 887-897.
- [178] Pohl, W. Dissociation of spatial discrimination deficits following frontal and parietal lesions in monkeys. J. Comp. Physiol. Psychol., 1973, 82, 227-239.

Accepted: January 13, 2011