



## First Complete Genome Sequence of the Dutch Veterinary *Coxiella burnetii* Strain NL3262, Originating from the Largest Global Q Fever Outbreak, and Draft Genome Sequence of Its Epidemiologically Linked Chronic Human Isolate NLhu3345937

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The largest global Q fever outbreak occurred in The Netherlands during 2007 to 2010. Goats and sheep were identified as the major sources of disease. Here, we report the first complete genome sequence of *Coxiella burnetii* goat outbreak strain NL3262 and that of an epidemiologically linked chronic human strain, both having the outbreak-related *CbNL01* multilocus variable-number tandem-repeat analysis (MLVA) genotype.

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fever is a zoonotic disease caused by Coxiella burnetii. Starting in 2007, The Netherlands has been confronted with the largest global Q fever outbreak ever, involving 4,026 human cases. Based on epidemiological and genotyping studies, dairy goats and sheep were identified as the main sources of the human Q fever outbreak. Special attention was given to the Dutch outbreakspecific strain of the CbNL01 multilocus variable-number tandem-repeat analysis (MLVA) genotype, which was identified in abortive dairy goats and in humans (1-6). The current project was performed with C. burnetii strains of this predominant genotype isolated from an aborted goat placenta (1, 7) and from a heart valve of a chronic Q fever patient during the outbreak period. Genome determination of the veterinary NL3262 strain is crucial for understanding the large outbreak and biology of this highly virulent strain. The genomes of virulent veterinary (NL3262) and related human (NLhu3345937) outbreak strains were sequenced, and their reconstructed genomes were compared to the examine similarities and differences in their genome structures.

Strains NL3262 and NLhu3345937 were cultivated axenically in acidified citrate cysteine medium (ACCM-2) and BGM cells, respectively (7, 8). Genomic DNA was isolated using the phenolchloroform method (9). A prior DNase treatment was performed to the bacterial pellet of NLhu3345937 to eliminate most hostderived DNA. The genome of NL3262 was *de novo* reconstructed from mixed assemblies using a combination of PacBio RS, Roche 454XL, and Illumina PE250 MiSeq reads. Reconstructed genomes were improved with different Illumina read sets, with a total average coverage of  $600 \times$  using Pilon-1.8 (10). The NLhu3345937 genome was *de novo* reconstructed with Illumina PE250 MiSeq reads using SPA des-3.6.2 (11), for a total average coverage of  $285\times.$ 

The complete NL3262 sequence and draft genome sequence of NLhu3345937 contain 2,093,477 and 2,088,566 bp, with G+C contents of 42.9 and 42.6%, respectively. Genome comparisons of NL3262, NLhu3345937, and the draft NL-Limburg outbreak strains (12) using Nucmer (13), MUMi-BioNJ tree (14), and visualized using Artemis Comparison Tool (ACT) (15) show that these strains are closely related, with differences mainly based on single-base-pair mutations. Our veterinary and human strains both contain the ~37-kb pQpH1 plasmid similar to that in the reference NM-RSA493 strain. Genome annotation of NL3262 by NCBI Prokaryotic Genome Annotation Pipeline (http://www .ncbi.nlm.nih.gov/genome/annotation\_prok) showed 2,101 coding sequences (CDSs), 180 pseudogenes, 42 tRNAs, 3 rRNAs, and 1 noncoding RNA (ncRNA). Compared to C. burnetii NM (accession no. NC\_002971.3), the majority of genes (84%) are orthologs found in both strains, with only 16% of the genes specific to NL3262 comprising mainly hypothetical products. Genome reconstruction was cumbersome between different passages of each strain due to high genome complexity, as described before for other isolates (many transposons, repetitive IS1111 elements, and rearrangements) (16). The observed high similarity between veterinary (NL3262) and human (NLhu3345937 and NL-Limburg) strains confirms the previous epidemiological and genotypic studies linking the goat and human infections during the Q fever outbreak in The Netherlands. A detailed comparative genome analysis of several sequenced C. burnetii isolates from different origins is ongoing.

Nucleotide sequence accession numbers. *C. burnetii* NL3262 and NLhu3345937 genome and plasmid sequences have been deposited in GenBank under accession numbers CP013667, CP013668, CP014354, and CP014355.

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#### REFERENCES

- Roest HIJ, Ruuls RC, Tilburg JJHC, Nabuurs-Franssen MH, Klaassen CHW, Vellema P, van den Brom R, Dercksen D, Wouda W, Spierenburg MAH, van der Spek AN, Buijs R, de Boer AG, Willemsen PTJ, van Zijderveld FG. 2011. Molecular epidemiology of *Coxiella burnetii* from ruminants in Q fever outbreak, the Netherlands. Emerg Infect Dis 17: 668–675. http://dx.doi.org/10.3201/eid1704.101562.
- 2. Van Steenbergen JE, Morroy G, Groot CA, Ruikes FGH, Marcelis JH, Speelman P. 2007. An outbreak of Q fever in The Netherlands—possible link to goats. Ned Tijdschr Geneeskd 151:1998–2003. (In Dutch.).
- Wouda W, Dercksen DP. 2007. Abortion and stillbirth among dairy goats as a consequence of *Coxiella burnetii*. Tijdschr Diergeneeskd 132: 908–911. (In Dutch.).
- Karagiannis I, Schimmer B, Van Lier A, Timen A, Schneeberger P, Van Rotterdam B, De Bruin A, Wijkmans C, Rietveld A, Van Duynhoven Y. 2009. Investigation of a Q fever outbreak in a rural area of The Netherlands. Epidemiol Infect 137:1283–1294. http://dx.doi.org/10.1017/ S0950268808001908.
- Karagiannis I, Morroy G, Rietveld A, Horrevorts AM, Hamans M, Francken P, Schimmer B. 2007. Q fever outbreak in the Netherlands: a preliminary report. Euro Surveill 12:pii=3247. http://www.eurosurveillance.org/ViewArticle .aspx?ArticleId=3247.
- 6. Tilburg JJHC, Roest HIJ, Buffet S, Nabuurs-Franssen MH, Horrevorts

AM, Raoult D, Klaassen CHW. 2012. Epidemic genotype of *Coxiella burnetii* among goats, sheep, and humans in the Netherlands. Emerg Infect Dis 18:887–889. http://dx.doi.org/10.3201/eid1805.111907.

- Roest H-J, van Gelderen B, Dinkla A, Frangoulidis D, van Zijderveld F, Rebel J, van Keulen L. 2012. Q fever in pregnant goats: pathogenesis and excretion of *Coxiella burnetii*. PLoS One 7:e48949. http://dx.doi.org/ 10.1371/journal.pone.0048949.
- Omsland A, Beare PA, Hill J, Cockrell DC, Howe D, Hansen B, Samuel JE, Heinzen RA. 2011. Isolation from animal tissue and genetic transformation of *Coxiella burnetii* are facilitated by an improved axenic growth medium. Appl Environ Microbiol 77:3720–3725. http://dx.doi.org/ 10.1128/AEM.02826-10.
- Kuley R, Smith HE, Frangoulidis D, Smits MA, Jan Roest HI, Bossers A. 2015. Cell-free propagation of *Coxiella burnetii* does not affect its relative virulence. PLoS One 10:e0121661. http://dx.doi.org/10.1371/ journal.pone.0121661.
- Walker BJ, Abeel T, Shea T, Priest M, Abouelliel A, Sakthikumar S, Cuomo CA, Zeng Q, Wortman J, Young SK, Earl AM. 2014. Pilon: an integrated tool for comprehensive microbial variant detection and genome assembly improvement. PLoS One 9:e112963. http://dx.doi.org/ 10.1371/journal.pone.0112963.
- Bankevich A, Nurk S, Antipov D, Gurevich AA, Dvorkin M, Kulikov AS, Lesin VM, Nikolenko SI, Pham S, Prjibelski AD, Pyshkin AV, Sirotkin AV, Vyahhi N, Tesler G, Alekseyev MA, Pevzner PA. 2012. SPAdes: a new genome assembly algorithm and its applications to singlecell sequencing. J Comput Biol 19:455–477. http://dx.doi.org/10.1089/ cmb.2012.0021.
- Hammerl JA, Mertens K, Sprague LD, Hackert VH, Buijs J, Hoebe CJ, Henning K, Neubauer H, Al Dahouk S. 2015. First draft genome sequence of a human *Coxiella burnetii* isolate, originating from the largest Q fever outbreak ever reported, the Netherlands, 2007 to 2010. Genome Announc 3(3):e00445-15. http://dx.doi.org/10.1128/genomeA.00445-15.
- Kurtz S, Phillippy A, Delcher AL, Smoot M, Shumway M, Antonescu C, Salzberg SL. 2004. Versatile and open software for comparing large genomes. Genome Biol 5:R12. http://dx.doi.org/10.1186/gb-2004-5-2-r12.
- Gascuel O. 1997. BIONJ: an improved version of the NJ algorithm based on a simple model of sequence data. Mol Biol Evol 14:685–695. http:// dx.doi.org/10.1093/oxfordjournals.molbev.a025808.
- Carver TJ, Rutherford KM, Berriman M, Rajandream M-A, Barrell BG, Parkhill J. 2005. ACT: the Artemis Comparison Tool. Bioinformatics 21: 3422–3423. http://dx.doi.org/10.1093/bioinformatics/bti553.
- Beare PA, Unsworth N, Andoh M, Voth DE, Omsland A, Gilk SD, Williams KP, Sobral BW, Kupko JJ, Porcella SF, Samuel JE, Heinzen RA. 2009. Comparative genomics reveal extensive transposon-mediated genomic plasticity and diversity among potential effector proteins within the genus *Coxiella*. Infect Immun 77:642–656. http://dx.doi.org/10.1128/ IAI.01141-08.