

Description of *Collinsella avium* sp. nov., a new member of the *Collinsella* genus isolated from the ceacum of feral chicken

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Abstract

An obligately anaerobic, non-motile and Gram-positive rod bacterium, strain SW219 was isolated from ceacum of feral chickens. Based on 16S rRNA sequence analysis, the strain SW219 exhibited 97.88% similarity to *Collinsella massiliensis* strain GD3 strain, the closest valid species. The genome size of SW219 was 2.58 Mbp with 64.5 mol% of G+C content. The phenotypic and genotypic analysis suggested that the strain SW219 is a new species belonging to the family *Coriobacteriaceae* within the *Actinobacteria* phylum, which the name *Collinsella avium* sp. nov. is proposed. The type strain of *Collinsella avium* is SW219 (= DSM 109235^T and = CCOS 1884^T).

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Keywords: Chicken, *Collinsella*, gut health, microbiome, new species, taxonogenomics, whole genome

Original Submission: 21 August 2020; **Revised Submission:** 19 April 2021; **Accepted:** 9 May 2021

Article published online: 18 May 2021

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Introduction

Animal gut harbours a high diversity of microbial community which plays a significant role in host metabolism and immune system, including pathogen protection [1,2]. Although gut microbiota composition and diversity has been accessed using metagenomics analysis, huge number of bacterial species remains uncultured [3–5]. To study interaction and function of gut bacteria, isolation and cultivation of uncultured bacteria is an important step. Taxonomical characterisation of new bacterial species allows the study of their phenotype and genotype, which may potentially impact host health. Here we report the isolation of new strain SW219 from the pooled ceacal contents of feral chicken. We further characterised the strain using taxonogenomics approach and identified it as a new species.

Methods

Isolation and growth conditions

The strain SW219 was isolated from ceacum of feral chicken by culturing in strict anaerobic conditions using a Coy Lab anaerobic chamber containing 85% nitrogen, 10% hydrogen and 5% carbon dioxide. Modified brain heart infusion (BHI-M) medium, which contained 37 g/L of BHI, 5 g/L of yeast extract, 1 mL of 1 mg/mL menadione, 0.3 g of L-cysteine, 1 mL of 0.25 mg/L of resazurin, 1 mL of 0.5 mg/mL hemin, 10 mL of vitamin and mineral mixture, 1.7 mL of 30 mM acetic acid, 2 mL of 8 mM propionic acid, 2 mL of 4 mM butyric acid, 100 µl of 1 mM isovaleric acid, and 1% of pectin and inulin was used for strain culturing and maintenance.

Strain identification and phylogenetic analysis

The strain SW219 initially was identified using MALDI-TOF (Bruker, Germany). Later, genomic DNA of the strain SW219 was extracted using DNeasy Blood & Tissue kit (Qiagen), in accordance with the manufacturer's instructions. 16S rRNA gene sequences was amplified using universal primer set 27F (5'-AGAGTTTGATCMTGGCTCAG-3') and 1492R (5'-ACCTTGTTACGACTT-3') [6,7]. The PCR amplicon was sequenced using Sanger dideoxy method (ABI 3730XL; Applied

Biosystems). The full-length of 16S rRNA gene of strain SW219 was assembled using Geneious software [8]. The 16S rRNA gene sequence of strain SW219 was compared with closely related strains in the GenBank (www.ncbi.nlm.nih.gov/genbank/) and EZBioCloud databases (www.ezbiocloud.net/eztaxon) [9]. The 16S rRNA sequences of valid related species were obtained from EZBioCloud databases for phylogenetic analysis using MEGA7 software [10]. Multiple alignments were generated using the CLUSTAL-W algorithm [11]. Reconstruction of phylogenetic tree was built using the maximum-likelihood [12], maximum-parsimony [13] and neighbour-joining [14] methods. The distance matrices were generated using Kimura's two-parameter model. Bootstrap

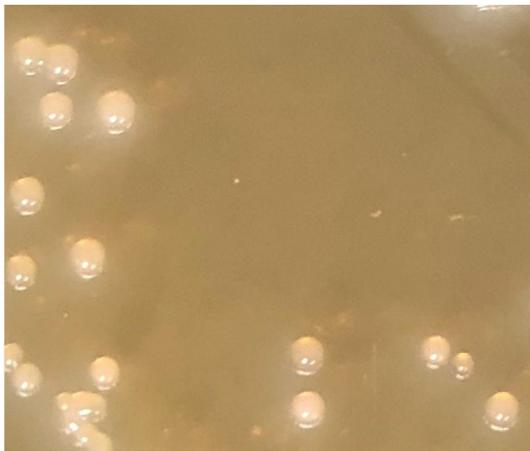


FIG. 1. Colony morphology of *Collinsella avium* sp. nov. strain SW219. Cells were cultured for 48–72 hours at 37° C in BHI-M medium.

resampling analysis of 1000 replicates was performed to estimate the confidence of tree topologies.

Phenotypic tests

Colony morphology of strain SW219 was determined after 2–3 days of incubation on BHI-M agar plates. Gram-staining was performed using a Gram-Staining kit set (Difco), in accordance with the manufacturer's instructions. Cell morphologies of cultures during exponential growth were examined by scanning electron microscopy. Aerotolerance was examined by incubating cultures for 2 days under aerobic and anaerobic conditions. Motility of this microorganism was verified using motility medium with triphenyltetrazolium chloride [15]. The growth was indicated by the presence of red colour, reduced form of triphenyltetrazolium chloride after it is absorbed into bacterial cell wall. Other biochemical tests including utilisation of various substrates and enzyme activities were investigated using the AN MicroPlate (Biolog) and API ZYM (bioMérieux) in accordance with the manufacturer's instructions. Cellular fatty acids were determined following Microbial Identification System (MIDI) protocols from cell biomass using gas chromatography (Agilent 7890A) [16].

Genome sequencing and analysis

The library for genome sequencing was prepared using Nextera XT kit (Illumina Inc). The library was sequenced on a MiSeq platform using Illumina 2 × 250 paired end chemistry. The reads were assembled using Unicycler that builds an initial assembly graph from short reads using the *de novo* assembler SPAdes 3.11.1 [17]. The quality assessment for the assembly was performed using QUAST [18]. The circular map of SW219 genome was generated using DNAPlotter software [19]. Genome

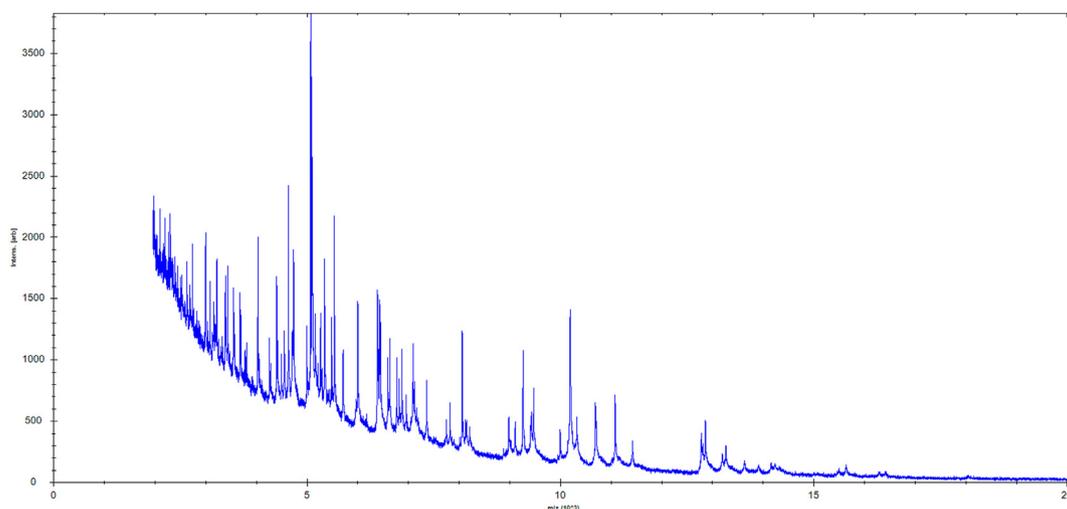


FIG. 2. MALDI-TOF MS reference spectrum of *Collinsella avium* sp. nov. strain SW219. The reference spectrum was generated by comparison of spectra from 12 individual colonies.

annotation was performed using Rapid Annotation using Sub-system Technology server [20]. The analysis of functional categories of strain SW219 and other members of the genus *Collinsella* was analysed by Rapid Annotation using Subsystem Technology and was presented as a heatmap generated using Explicet software [21]. Furthermore, the average nucleotide identity (ANI) was calculated between SW219 and the related genomes using OrthoANI software [22]. The digital DNA-DNA hybridisation (dDDH) was performed between strain SW219 and the closet phylogenetic neighbour using Genome-to Genome Distance Calculator (GGDC 2.1) web server (<http://ggdc.dsmz.de>) [23].

Results

Strain SW219 was isolated from cecal content of feral chicken. Colonies of strain SW219 were white convex, translucent and entire circular margin with a mean diameter of 0.1-0.3 mm after 2-3 days of incubation on BHI-M agar (Fig. 1). For an initial identification, the score of MALDI-TOF MS was less than 1.7 suggesting no match organism in the database (Fig. 2). Based on 16S rRNA phylogenetic analysis, strain SW219 was clustered within the *Collinsella* genus. It exhibited 97.88% similarity to *Collinsella massiliensis* strain GD3 (GenBank accession number

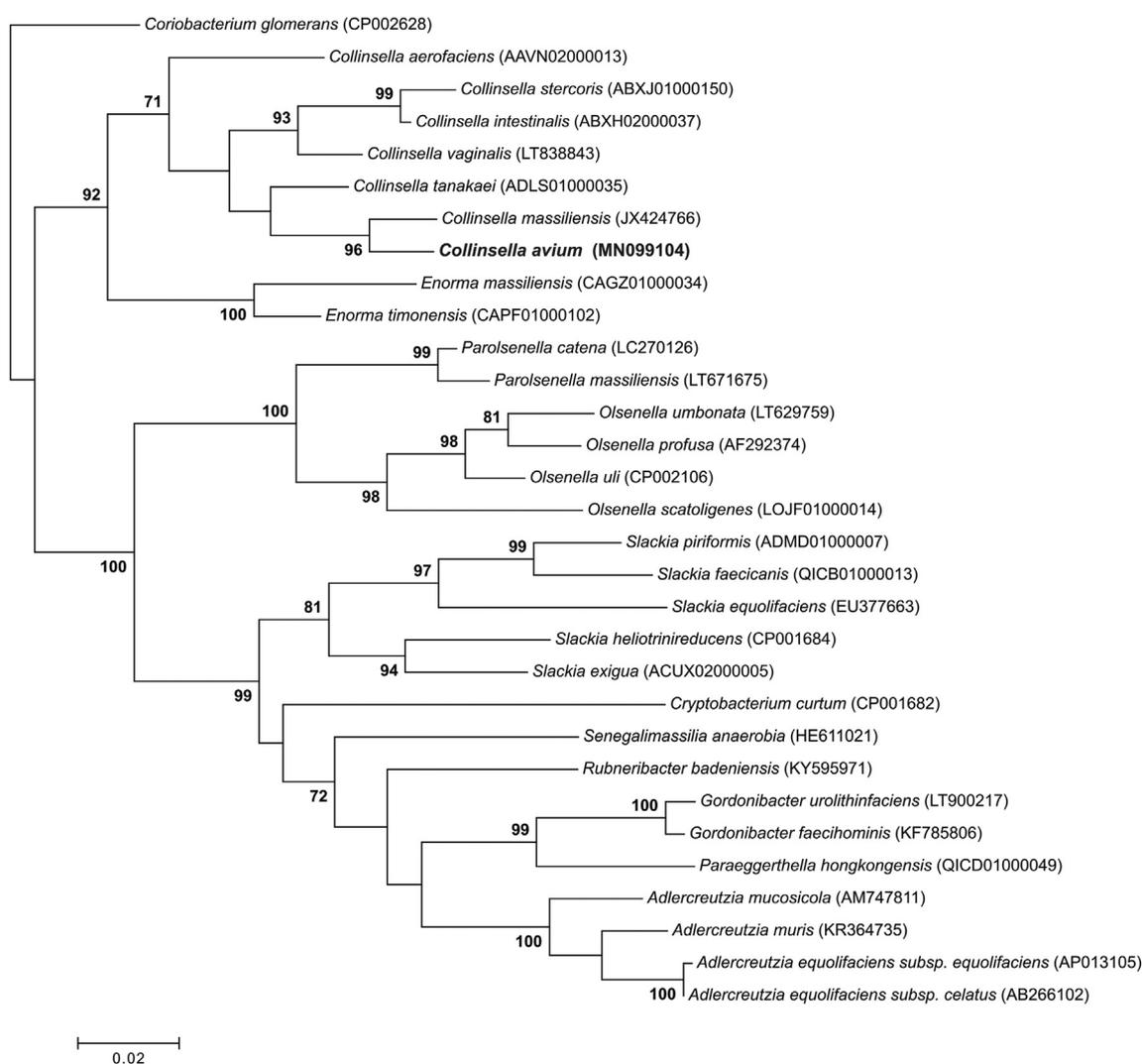


FIG. 3. Phylogenetic tree showing position of *Collinsella avium* sp. nov. strain SW219 within closely related taxa in the genus *Collinsella* of *Coriobacteriaceae* family. GenBank accession numbers of the 16S rRNA gene sequences are given in parentheses. Sequences were aligned using CLUSTALW. The phylogenetic tree was obtained using the maximum-likelihood method and generated using MEGA 7 software. Bootstrap values obtained by repeating the analysis 1000 times to generate a majority consensus tree are indicated at the nodes. The scale bar indicates a 2% nucleotide sequence divergence.

JX424766.1), the closest species with standing in nomenclature (Fig. 3). The similarity less than 98% of 16S rRNA sequences suggested that it is a new species. The characteristics of this strain were further investigated using multiple tests and analyses.

The strain SW219 grew in BHI-M medium at 37°C for 24 h under anaerobic condition. Cells were Gram-positive bacilli and non-motile with a diameter of 0.5-2.0 µm (Fig. 4). General

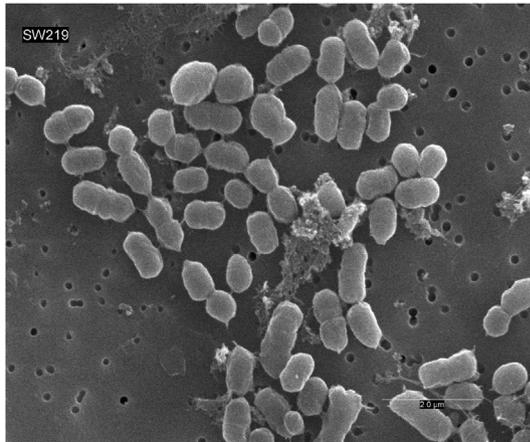


FIG. 4. Scanning electron microscopy (SEM) of *Collinsella avium* strain SW219. Cell were cultured for 24 hours at 37°C in BHI-M medium. Bar, 2.0 µm.

features and biochemical characteristics of the strain SW219 were distinct from other members of the genus *Collinsella* (Table 1). The major cellular fatty acids included C_{16:0} DMA, C_{16:0} aldehyde, C_{16:0} and C_{12:0} (Table 2). The genome of strain SW219 is 2.58 Mbp with 64.5 mol% of G+C content (Fig. 5). A total of 2162 coding sequences (CDS) and 56 RNAs were identified in the genome. Based on the analysis of functional

TABLE 2. Cellular fatty acid compositions of *Collinsella avium* strain SW219

Fatty acid	Mean relative %
Straight chain	
12:00	10.98
14:00	14.62
16:00	12.26
18:0	2.12
16:0 aldehyde	15.36
18:0 aldehyde	2.21
Demethylacetal (DMA)	
12:0 DMA	7.01
14:0 DMA	2.58
16:0 DMA	22.05
18:0 DMA	5.13
Unsaturated	
13:1 at 12-13	2.11
18:1 w9c	2.45
18:2 w6,9c	1.13
Summed feature I	2.11

*Summed features are fatty acids that could not be separated using the MIDI System. Summed feature I contains C_{13:1} and/or C_{14:0} aldehyde.

TABLE 1. Characteristics of strain SW219 and other members of the genus *Collinsella*

Characteristic	SW219	<i>C. massiliensis</i>	<i>C. intestinalis</i>	<i>C. aerofaciens</i>	<i>C. tanakei</i>	<i>C. stercoris</i>
Habitat	chicken gut	human gut	human gut	human gut	human gut	NA
Cell morphology	rods	rods	rods	cocci to rods	rods	rods
Cell diameter (µm)	0.5-2.0	0.57	0.3-0.5	0.3-0.7	0.5-1.0	0.3-0.5
Gram stain	+	+	+	+	+	+
Motility	—	—	—	—	—	—
DNA G+C content (mol%)	64.5	65.8	64.4	61	60.8	61.2
API test						
Alkaline phosphatase	+	+	+	—	+	+
Indirect ELISA	+	—	NA	NA	NA	NA
Leucine arylamidase	—	—	—	—	—	—
Valine arylamidase	+	+	NA	NA	NA	NA
Cystine arylamidase	+	—	NA	NA	NA	NA
Trypsin	—	—	NA	NA	NA	NA
Acid phosphatase	+	+	+	—	+	+
α-galactosidase	+	NA	NA	NA	NA	NA
β-galactosidase	—	—	—	+	—	+
β-glucuronidase	—	—	—	—	+	—
α-glucuronidase	+	—	—	+	—	—
β-glucosidase	—	+	—	—	+	+
N-acetyl-β-glucosaminidase	+	—	+	—	—	+
Utilisation of						
Glucose	+	—	+	+	+	+
Mannose	+	—	+	+	+	+
Galactose	+	—	+	+	NA	+
Fructose	+	—	+	+	NA	+
Maltose	—	—	—	+	+	+
Sucrose	+	NA	NA	NA	NA	NA
Maltotriose	+	NA	NA	NA	NA	NA
Cellulbiase	—	—	—	—	+	+
Lactose	—	—	—	+	+	+
Rhamnase	—	—	—	NA	—	—
Ribose	NA	—	—	+	+	+
Raffinose	—	—	—	NA	NA	—
Sorbitol	+	+	—	—	—	—

NA, data not available; +/-, depending on tests used.

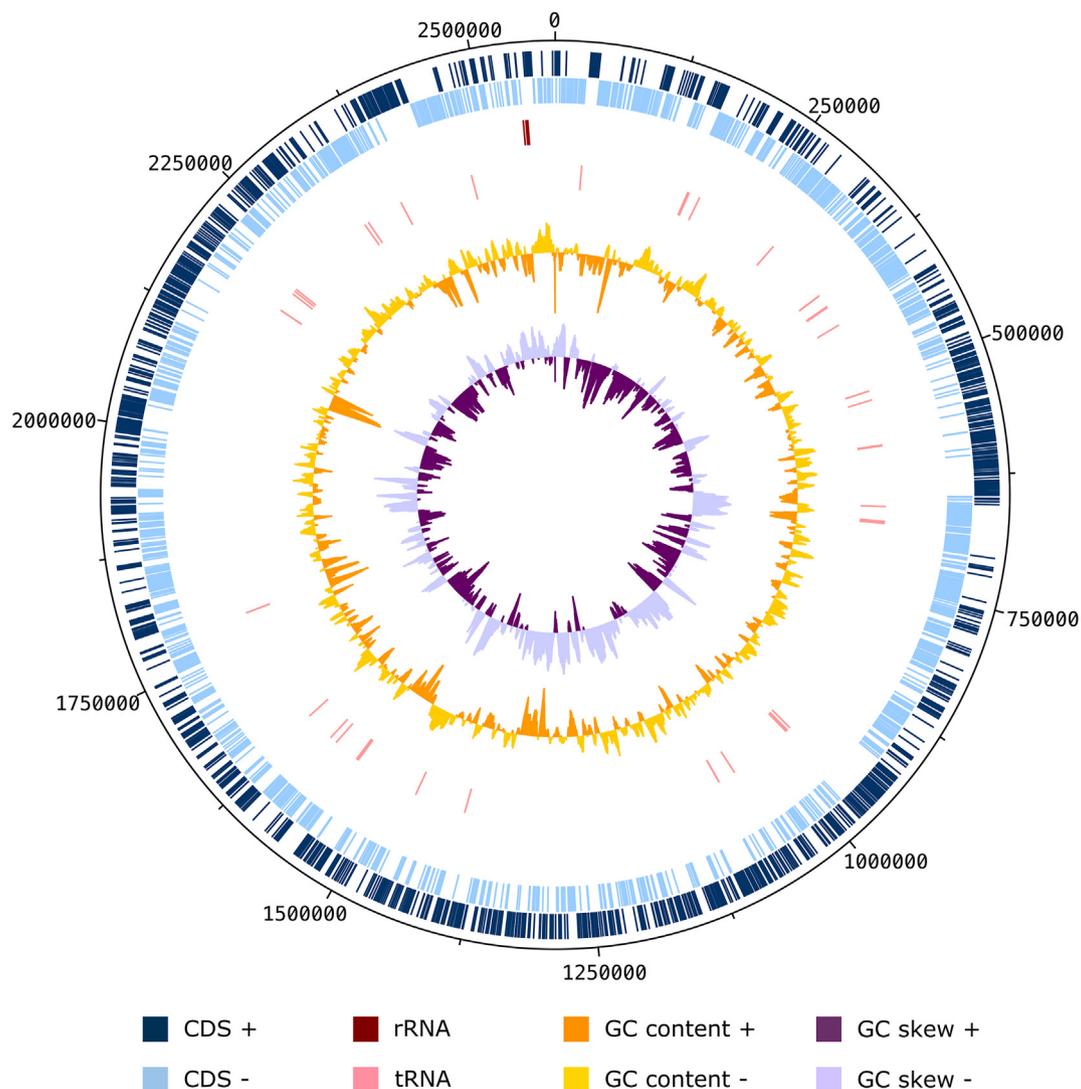


FIG. 5. Graphical circular map of *Collinsella avium* strain SW219 genome. From outside to the centre: coding sequences on the forward strand (CDS +), coding sequences on the reverse strand (CDS -), tRNAs, rRNAs, GC content and GD skew. This map was generated using DNAPlotter software.

categories (COGs), the dominant categories were amino acids and derivatives, carbohydrate and protein mechanism (Fig. 6). Overall, the COGs pattern of the strain SW219 was very similar to other species of *Collinsella*. The stain SW219 showed the similar level of multiple categories such as amino acids and derivatives, phosphorus metabolism, and stress response to the closest relative, *C. massiliensis* while other categories were similar to other members of the genus *Collinsella*. For example, DNA metabolism and nucleosides and nucleotides categories were similar to those of *C. tanakaei* and *C. stercoris*, respectively. In addition, stain SW219 contained the same percentage of protein metabolism genes as *C. aerofaciens*, *C. stercoris*, *C. tanakaei*, and *C. vaginalis*. The ANI values of genome sequences between the stain SW219 and other members of the genus *Collinsella* ranged

from 75–77% (Fig. 6). These values were significantly less than the proposed ANI cutoff of 95–96% for the identical species [24]. Species cutoff value for a new species is 70% based dDDH analysis. When the stain SW219 is compared with its closely related species, all values were less than 70%, which clearly shows the need for the designation of this strain as a new species (Table 3) (Fig. 7).

Discussion

Recently, the high throughput culturing of gut microbiota known as “culturomics” have been developed to characterise and identify unculturable bacteria, allowing isolation of several new

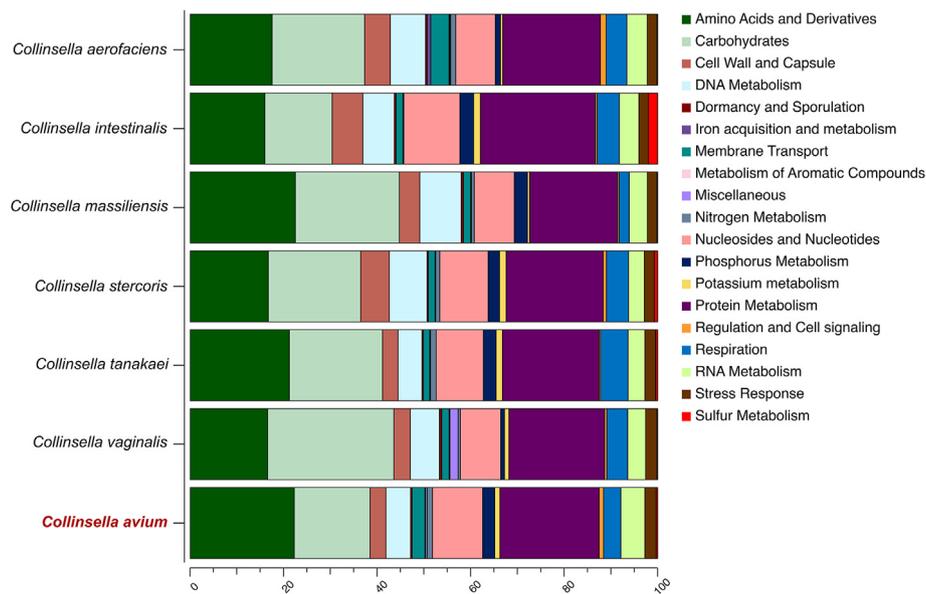


FIG. 6. Distribution of functional features of predicted coding sequences of *Collinsella avium* strain SW219 and closely related genomes. The functional features were predicted based on the clusters of orthologous groups. Heatmap was generated from genome annotation of individual species by RAST using Explicet software.

TABLE 3. Digital DNA-DNA hybridisation comparing the genomic similarity between members of the genus *Collinsella*

% DDH estimate	<i>C. massiliensis</i>	<i>C. tanakaei</i>	<i>C. vaginalis</i>	<i>C. intestinalis</i>	<i>C. stercoris</i>	<i>C. aerofaciens</i>
HSP length/total length	18.30	15.8	15.6	16.60	16.5	16
Identities/HSP length	2.32	2.27	22.3	22.90	2.36	22.3
Identities/total length	2.32	2.27	15.7	22.90	16.5	16.1
Difference in % G+C	1.35	4.23	0.1	2	1.27	3.92

DDH,

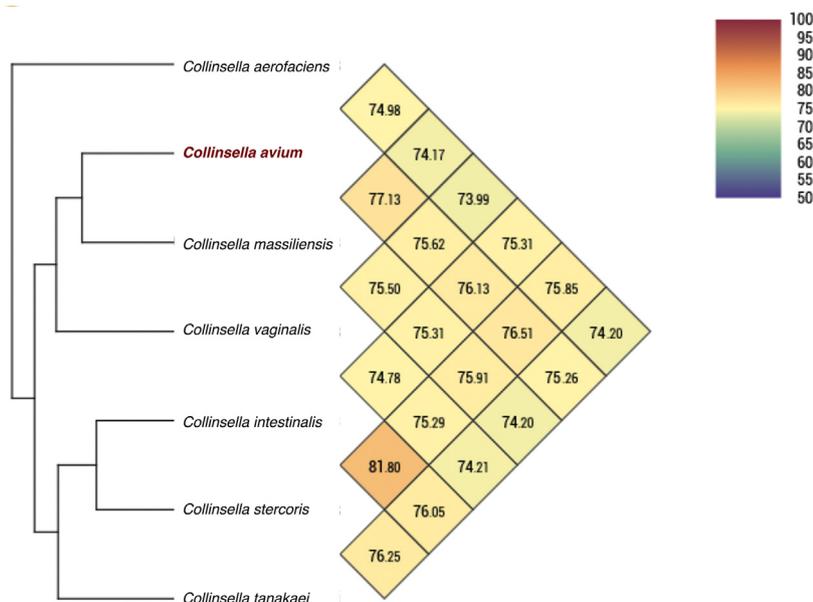


FIG. 7. Average nucleotide identity comparison of *Collinsella avium* strain SW219 and related species with a valid taxonomy. Heatmap generated with OrthoANI values calculated using the OAT software between *Collinsella avium* sp. nov. and the closely related species.

species from the gut microbiota [25,26]. Although metagenomics is the primary strategy for the study of gut microbiota [27], the large portion of gut microbiota are uncultured organisms [5]. Culturing of bacteria is important to understand their functions, and contributions in health and diseases [28,29]. In this study, culturomics based isolation recovered a previously uncultured bacterium SW219 from the caecum of feral chickens.

Generally, 16S rRNA gene sequence similarity is primarily used to determine the taxonomic threshold for determining new species [24]. The 16S rRNA-based phylogenetic analysis showed that *C. massiliensis* was the closest related species of the strain SW219 with 97.88% similarity. A 16S rRNA identity cutoff value of up to 98.6% can be used to designate new isolate as a distinct species [30]. Furthermore, the ANI and dDDH values of the strain SW219 is markedly different when compared to its closest genomes (Figs. 5 and 6). In addition, strain SW219 exhibited distinct phenotypic properties to its closest valid strain, including carbon source utilisation, enzymatic activity and cellular fatty acid components (Tables 1 and 2).

Therefore, strain SW219 could be classified as a new species within the genus *Collinsella*.

Conclusion

Based on the genotypic and phenotypic features of the strain SW219, this organism identified herein should be described as a new species of the genus *Collinsella*. This organism is proposed as the type strain of the new species, for which the name is *Collinsella avium* is proposed.

Description of *Collinsella avium* sp. nov

Collinsella avium sp. nov. (a'vi.um. L. gen. pl. n. avium, of birds). *Collinsella avium* is an obligately anaerobic, non-motile and Gram-positive rod bacterium. The optimal growth of this strain is at 37° C and pH 7 in BHI-M medium. The average size of each cell is 0.5-2.0 µm. Colonies are visible on BHI-M agar after 2-3 days and are approximately 0.1-0.3 cm in diameter, with white translucent, round and convex with entire circular margin. Positive enzyme activity of this organism includes alkaline phosphatase, esterase (C4), esterase lipase (C8), leucine arylamidase, valine arylamidase, cystine arylamidase, acid phosphatase, naphthol-As-Bi-phosphorydrolase, α-galactosidase, α-glucosidase and N-acetyl-β-glucosaminidase. The major fatty acids are C_{16:0} DMA, C_{16:0} aldehyde, C_{16:0} and C_{12:0}. The genome of strain SW219 is 2.58 Mbp with 64.5 mol% of G+C content. The type strain is SW219 (= DSM 109235^T) isolated from caecum of feral chickens.

Nucleotide sequence accession number

The 16S rRNA gene was deposited in GenBank under accession number MN099104. Whole genome sequence was deposited in NCBI Sequence Read Archive under the accession number SRR9967483.

Deposit in culture collection

Strain SW219 was deposited in The Leibniz Institute DSMZ—German Collection of Microorganisms and Cell Cultures GmbH and The national Culture Collection of Switzerland (CCOS) under numbers DSM 109235 and CCOS 1884, respectively.

Transparency declaration

None to declare.

This work was supported in part by the USDA National Institute of Food and Agriculture, Hatch projects SD00H532-14 and SD00R540-15, and a grant from the South Dakota Governor's Office of Economic Development awarded to JS.

Acknowledgements

SW would like to acknowledge the Science Achievement Scholarship of Thailand. The authors acknowledge Electron Microscopy Core Facility at the Bowling Green State University, Ohio, USA, for assistance with scanning electron microscopy. The authors would like to thank Linto Antony and Roshan Kumar, Veterinary and Biomedical Science Department, South Dakota State University, USA, for their assistance in genome sequencing and analysis.

References

- [1] Brestoff JR, Artis D. Commensal bacteria at the interface of host metabolism and the immune system. *Nat Immunol* 2013;14(7):676–84.
- [2] Khosravi A, Yanez A, Price JG, Chow A, Merad M, Goodridge HS, et al. Gut microbiota promote hematopoiesis to control bacterial infection. *Cell Host Microbe* 2014;15(3):374–81.
- [3] Morgan XC, Huttenhower C. Meta'omic analytic techniques for studying the intestinal microbiome. *Gastroenterology* 2014;146(6):1437–14348 e1.
- [4] Greub G. Culturomics: a new approach to study the human microbiome. *Clin Microbiol Infect* 2012;18(12):1157–9.
- [5] Lagier JC, Khelaifa S, Alou MT, Ndongo S, Dione N, Hugon P, et al. Culture of previously uncultured members of the human gut microbiota by culturomics. *Nat Microbiol* 2016;1:16203.
- [6] Lane DJ. 16S/23S rRNA sequencing. In: Stackebrandt E, Goodfellow M, editors. *Nucleic acid techniques in bacterial systematic*. New York: John Wiley and Sons; 1991.

- [7] Stackebrandt E, Liesack W. Nucleic acids and classification. In: Goodfellow M, O'Donnell AG, editors. Handbook of new bacterial systematics. London, England: Academic Press; 1993.
- [8] Kearsse M, Moir R, Wilson A, Stones-Havas S, Cheung M, Sturrock S, et al. Geneious Basic: an integrated and extendable desktop software platform for the organization and analysis of sequence data. *Bioinformatics* 2012;28(12):1647–9.
- [9] Kim OS, Cho YJ, Lee K, Yoon SH, Kim M, Na H, et al. Introducing EzTaxon-e: a prokaryotic 16S rRNA gene sequence database with phylotypes that represent uncultured species. *Int J Syst Evol Microbiol* 2012;62(Pt 3):716–21.
- [10] Kumar S, Stecher G, Tamura K. MEGA7: molecular evolutionary genetics analysis Version 7.0 for bigger datasets. *Mol Biol Evol* 2016;33(7):1870–4.
- [11] Thompson JD, Higgins DG, Gibson TJ, Clustal W. Improving the sensitivity of progressive multiple sequence alignment through sequence weighting, position-specific gap penalties and weight matrix choice. *Nucleic Acids Res* 1994;22(22):4673–80.
- [12] Felsenstein J. Evolutionary trees from DNA sequences: a maximum likelihood approach. *J Mol Evol* 1981;17(6):368–76.
- [13] Fitch WM. Toward defining the course of evolution: minimum change for a specific tree topology. *Syst Zool* 1971;20(10):406–16.
- [14] Saitou N, Nei M. The neighbor-joining method: a new method for reconstructing phylogenetic trees. *Mol Biol Evol* 1987;4(4):406–25.
- [15] Tittsler RP, Sandholzer LA. The use of semi-solid agar for the detection of bacterial motility. *J Bacteriol* 1936;31:575–80.
- [16] Sasser M. Identification of bacteria by gas chromatography of cellular fatty acids. In: MIDI technical note 101 Newark. DE: MIDI Inc; 1990.
- [17] Wick RR, Judd LM, Gorrie CL, Holt KE. Unicycler: resolving bacterial genome assemblies from short and long sequencing reads. *PLoS Comput Biol* 2017;13(6):e1005595.
- [18] Gurevich A, Saveliev V, Vyahhi N, Tesler G. QUAST: quality assessment tool for genome assemblies. *Bioinformatics* 2013;29(8):1072–5.
- [19] Carver T, Thomson N, Bleasby A, Berriman M, Parkhill J. DNAPlotter: circular and linear interactive genome visualization. *Bioinformatics* 2009;25(1):119–20.
- [20] Overbeek R, Olson R, Pusch GD, Olsen GJ, Davis JJ, Disz T, et al. The SEED and the Rapid annotation of microbial genomes using subsystems Technology (RAST). *Nucleic Acids Res* 2014;42(Database issue):D206–14.
- [21] Robertson CE, Harris JK, Wagner BD, Granger D, Browne K, Tatem B, et al. Explicet: graphical user interface software for metadata-driven management, analysis and visualization of microbiome data. *Bioinformatics* 2013;29(23):3100–1.
- [22] Lee I, Ouk Kim Y, Park SC, Chun J. OrthoANI: an improved algorithm and software for calculating average nucleotide identity. *Int J Syst Evol Microbiol* 2016;66(2):1100–3.
- [23] Meier-Kolthoff JP, Auch AF, Klenk HP, Goker M. Genome sequence-based species delimitation with confidence intervals and improved distance functions. *BMC Bioinform* 2013;14:60.
- [24] Kim M, Oh HS, Park SC, Chun J. Towards a taxonomic coherence between average nucleotide identity and 16S rRNA gene sequence similarity for species demarcation of prokaryotes. *Int J Syst Evol Microbiol* 2014;64(Pt 2):346–51.
- [25] Lagier JC, Hugon P, Khelaifia S, Fournier PE, La Scola B, Raoult D. The rebirth of culture in microbiology through the example of culturomics to study human gut microbiota. *Clin Microbiol Rev* 2015;28(1):237–64.
- [26] Kaeberlein T, Lewis K, Epstein SS. Isolating "uncultivable" microorganisms in pure culture in a simulated natural environment. *Science* 2002;296(5570):1127–9.
- [27] Dhakan DB, Maji A, Sharma AK, Saxena R, Pulikkan J, Grace T, et al. The unique composition of Indian gut microbiome, gene catalogue, and associated fecal metabolome deciphered using multi-omics approaches. *Gigascience* 2019;8(3).
- [28] Goodman AL, Kallstrom G, Faith JJ, Reyes A, Moore A, Dantas G, et al. Extensive personal human gut microbiota culture collections characterized and manipulated in gnotobiotic mice. *Proc Natl Acad Sci U S A* 2011;108(15):6252–7.
- [29] Rettedal EA, Gumpert H, Sommer MO. Cultivation-based multiplex phenotyping of human gut microbiota allows targeted recovery of previously uncultured bacteria. *Nat Commun* 2014;5:4714.
- [30] Yarza P, Yilmaz P, Pruesse E, Glockner FO, Ludwig W, Schleifer KH, et al. Uniting the classification of cultured and uncultured bacteria and archaea using 16S rRNA gene sequences. *Nat Rev Microbiol* 2014;12(9):635–45.