Precision medicine in inflammatory bowel disease

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Abstract

Inflammatory bowel disease (IBD) is an incurable disease characterized by remission-relapse cycles throughout its course. Both Crohn's disease (CD) and ulcerative colitis (UC), the two main forms of IBD, exhibit tendency to develop complications and substantial heterogeneity in terms of frequency and severity of relapse, thus posing great challenges to the clinical management for IBD. Current treatment strategies are effective in different ways in induction and maintenance therapies for IBD. Recent advances in studies of genetics, pharmacogenetics, proteomics and microbiome provide a strong driving force for identifying molecular markers of prognosis and treatment response, which should help clinicians manage IBD patients more effectively, and then, improve clinical outcomes and reduce treatment costs of patients. In this review, we summarize and discuss precision medicine in IBD, focusing on predictive markers of disease course and treatment response, and monitoring indices during therapeutic drug monitoring.

Keywords: disease course; inflammatory bowel disease; precision treatment; precision monitoring; treatment response

Introduction

Inflammatory bowel disease (IBD), including Crohn's disease (CD) and ulcerative colitis (UC), is characterized by chronicity, destructiveness, and a remission-relapse pattern [1]. The spectrum of disease symptoms is wide. Patients with UC typically present with diarrhea, bloody stool and tenesmus; while abdominal pain, diarrhea and weight loss are the common symptoms of CD [2]. Moreover, 6%-47% of IBD patients suffer from extra-intestinal manifestations (EIMs) involving organs or tissues like joints, eyes, skin, etc. [3]. The heterogeneous presentations of IBD make it difficult for physicians to diagnose the condition by clinical features alone. A large-scale, prospective and multicenter study including 1399 children demonstrated that diagnostic delay conferred risk for the development of complicated diseases and growth impairment in pediatric CD patients [4]. From this point, making a timely and accurate diagnosis is extremely important for IBD patients. Such precision diagnosis can be achieved by combining consideration of clinical manifestations, laboratory analysis, endoscopic examination, imaging tests, and histologic assessment. Furthermore, the severity of diseases and frequency of flare-ups vary substantially from one patient to another. Some patients may experience a mild disease course, while others may progress quickly. Remarkably, the phenotype of CD may vary and evolve over time. It can progress from non-stricturing/fistulizing behavior to stricturing and fistulizing behavior in a manner which is largely unpredictable. The one-year recurrence rate of IBD is approximately 10%–30%, despite achieving remission [5]. Although some markers have been identified to be useful in the prediction of disease flare-ups, relapses are always difficult to predict [5, 6]. IBD exhibits

highly heterogeneity on all levels, and its management faces great challenges.

IBD has become a global disease with the highest prevalence in Westernized countries and the greatest growing incidence in newly industrialized countries [7]. The disease puts a heavy burden not only on patients themselves and their families, but also on health care systems [8]. Substantial evidence indicates that IBD results from the interaction of genetic/epigenetic, environmental, immunological, and microbial aspects. Large-scale genetic studies provided major insights into the etiopathogenesis of IBD, and highlighted the shared and distinct genetic risk factors in CD and UC [9, 10]. However, for most identified loci, their functions remain unknown. Progress in pharmacogenetics, proteomics and microbiome also shed light on the complicated signaling pathways of IBD. Understanding these distinct signaling pathways further provides an impetus for IBD treatment. The current therapeutic goal for IBD is "treat-to-target", aiming at achieving mucosal healing (MH), avoiding permanent complications, and altering the natural history of IBD [11]. Thus, assessment of disease course and therapeutic response play key roles in IBD management. Selecting a targeted therapy for individual patient must be based on risk stratification by analyzing the determinants of disease course and treatment response, including clinical, genetic, epigenetic, serological and fecal markers (Fig. 1).

With the "Precision Medicine Initiative" put forward in 2015, precision medicine has become a hotspot in the field of health care [12]. A large number of studies have been conducted to optimize the precision diagnosis, treatment, and monitoring of IBD. Herein, we mainly discuss how research on signaling pathways

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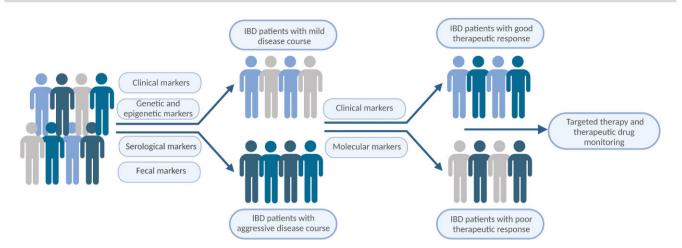


Figure 1. Precision medicine in IBD. Based on risk stratification by analyzing the determinants of disease course and treatment response including clinical, genetic, epigenetic, serological and fecal markers, physicians then select a targeted therapy and apply precision monitoring for individual patient. IBD: inflammatory bowel disease.

facilitates targeted therapy, and elaborate on precision treatment and precision monitoring in IBD.

Signaling pathways involved in IBD

Though the exact pathogenesis of remains unclear, it is believed that complicated mechanisms involving environmental triggers, luminal microbiota and host genetic susceptibility generate the disequilibrium between pro-inflammatory and antiinflammatory signaling, resulting in a chronic inflammatory state in IBD patients. Amongst numerous signaling pathways implicated in IBD, pathways related to tumor necrosis factor (TNF), leukocyte trafficking, and interleukin-12 (IL-12)/interleukin-23 (IL-23) have been intensively studied [13]. Undoubtedly, an improved understanding of these signaling pathways substantially facilitates the development of targeted treatment for IBD.

TNF has two forms: transmembrane TNF (mTNF) and soluble TNF. The former mainly binds with TNF receptor I (TNFRI), and then mediates the activation of nuclear factor kappa-B (NF- κ B) and caspase-8-dependent death signaling pathways, resulting in mucosal inflammation and intestinal epithelial barrier damage [14]. The latter often binds with TNF receptor II (TNFRII) and contributes to the activation of pro-survival and pro-inflammatory signaling pathways [15]. So far, available evidence indicates that TNF plays a central role in the pathogenesis of IBD. In order to block its pro-inflammatory action, researchers have developed some full-length anti-TNF monoclonal IgG1 antibodies such as infliximab, adalimumab and golimumab, and antibodies with Fab fragments such as certolizumab as well [14]. These antibodies exert anti-inflammatory effects by neutralizing mTNF and soluble TNF, reducing pro-inflammatory cytokines and cell adhesion molecules, and prompting T cell apoptosis, and inducing M2-type wound-healing macrophages [16–18]. Therefore, anti-TNF monoclonal antibodies showed outstanding therapeutic efficacy in the induction and maintenance of clinical, biochemical, and endoscopic remission in both animal models and patients with IBD [14]. It also has become a breakthrough in the precision treatment of IBD, encouraging further studies of other signaling pathways involved in IBD.

The IL-23/T helper cell 17 (Th17) pathway is critical in the pathophysiology of IBD. IL-23 consists of a p40 subunit and a p19 subunit. It is responsible for conferring pathogenicity to Th17 and

producing pro-inflammatory cytokines including interleukin-17A (IL-17A), interleukin-17F (IL-17F), interleukin-22 (IL-22), TNF, C-C chemokine receptor type 6 (CCR6), chemokine ligand 20 (CCL20), and others [19]. Th17 and Th17-related cytokines are acknowledged as strong inducers of inflammation. Increased levels of Th17 cells and Th17-related cytokines in IBD patients indicate that the IL-23/Th17 pathway plays an important role in IBD [20]. The association between the IL-23/Th17 pathway and IBD has been further emphasized by Genome-wide Association Studies (GWAS). Several risk genes involved in the IL-23/Th17 pathway, such as interleukin 23 receptor (IL23R), caspase recruitment domain family member 9 (CARD9), interleukin 12B (IL12B), Janus kinase 2 (JAK2), and CCR6 have been demonstrated to be associated with susceptibility to IBD [20]. Besides Th17, IL-23 also exerted effects on another T helper cell subset. Recently, a study reported that IL-23 also drove intestinal inflammation by evoking a pathogenic phenotype in Th1-like cells [21]. This finding provides a new direction for research on IBD. IL-12 is also a heterodimeric cytokine composed of a p35 and a shared p40 subunit. IL-12 can promote Th1 cytokine-mediated immune responses that is considered to be an integral part in the pathogenesis of CD. Besides, IL-12 is also involved in the activation of natural killer (NK) cells, cytotoxic T lymphocytes (CTLs), and group 1 innate lymphoid cells (ILC1s), and the production of interferon-gamma (IFN- γ) and TNF-a [22]. Therefore, new drugs targeting IL-12/IL-23 p40 (ustekinumab), and IL-23 p19 (risankizumab and briakinumab) have showed great benefit for IBD patients [23-25]. These examples further support the idea that targeting a key molecule within a signaling pathway can be an optimal option for targeted therapy in IBD.

Migration of leukocytes from the periphery to inflamed bowel tissues, and adhesion to the intestinal vasculature are two indispensable processes in the development and progression of IBD. Leukocyte-specific integrins, including alpha 4 beta 7 ($\alpha 4\beta 7$), alpha E beta 7 ($\alpha E\beta 7$), alpha 4 beta 1 ($\alpha 4\beta 1$), etc., are transmembrane glycoprotein receptors, mediating the connection between leukocytes and extracellular matrix ligands [11, 26]. The adhesive process of leukocytes to vascular endothelium can be activated by several pro-inflammatory cytokines such as TNF- α and interleukin-1 (IL-1), which are also responsible for the up-regulation of expression levels of intracellular adhesion molecules-1 (ICAM-1), mucosal adhesion cell adhesion molecule (MADCAM), and E-selectin on inflamed tissues [11]. As a guthoming receptor, $\alpha 4\beta 7$ participates in the key processes of lymphocyte homing (rolling migration and firm adhesion) by binding with MADCAM-1. Therefore, blocking the binding of $\alpha 4\beta 7$ to MADCAM-1 prevents lymphocytes from homing to the gut and thus attenuates intestinal inflammation [26]. Several antibodies to integrins such as vedolizumab, etrolizumab and abrilumab have shown great improvement in clinical outcomes in IBD patients [27, 28]. Therefore, blocking the leukocyte migration and adhesion process may become a novel direction in drug discovery in IBD.

Other signaling pathways involving sphingosine 1-phosphate (S1P)/sphingosine 1-phosphate receptors (S1PRs), JAK-signal transducer and activator of transcription (STAT), and Toll-like receptor 9 (TLR9) also show promise in targeted therapy. Numerous studies have linked S1P/S1PRs to leukocyte trafficking, a pivotal process in the development of IBD. S1P/S1PRs drive intestinal inflammation and regulate intestinal immune response by mediating the egress of lymphocytes from primary and secondary lymphoid organs [29]. Thus, S1P modulators such as ozanimod and etrasimod show some beneficial effects on patients with IBD [30, 31]. Existing evidence demonstrated that JAKs mediate the phosphorylation of the STAT family and participate in the inflammatory processes of IBD [32]. Activation of JAK-STAT may cause great changes in the level and ratio of pro-inflammatory and anti-inflammatory cytokines, as well as in the balance between immune activation and tolerance [32]. Drugs inhibiting the biological activity of the JAKs such as tofacitinib, filgotinib and upadacitinib have attracted great interest. These drugs provide therapeutic options for patients who are unresponsive to or intolerant of other-class drugs [33-35]. It is noteworthy that a novel therapeutic strategy, dual therapy, a combination of a biologic with a small molecule drug, holds great promise to help refractory IBD patients achieve remission [36]. A study of 16 biologic-refractory pediatric IBD patients showed that the dual therapy (vedolizumab/ustekinumab and tofacitinib) quickly facilitated steroid-free remission in 75% of patients with little serious safety events [37]. Several other studies also drew similar conclusions [38, 39]. The synergistic effects (preventing lymphocyte homing, neutralizing pro-inflammatory cytokines, and inhibiting downstream cytokine receptor signaling pathways) of vedolizumab/infliximab/ustekinumab and tofacitinib may explain these interesting findings. Contrary to these abovementioned signaling molecules, TLR9 shows beneficial effects on intestinal inflammation. Compared with the control group, TLR9-deficient mice with dextran sodium sulfate (DSS)-induced colitis presented more severe inflammation and delayed wound repair [40]. The protective effects of TLR9 on inflammation were further confirmed by the fact that activation of TLR9 contributed to the upregulation of mucosal IL10 and suppression of Th17 cells [41]. Correlations between mucosal TLR9 levels and severity of inflammation have also been demonstrated [41]. All these findings pave a new way to TLR9-targeted treatment in IBD. Cobitolimod, the TLR9 agonist, has been claimed to be effective in inducing clinical response in UC patients with poor response to conventional or biological treatments [42]. However, the efficacy of cobitolimod has only been evaluated in a phase II clinical trial, yet to be validated by large-sample clinical trials.

Indeed, revolutionary discoveries of different signaling pathways and major advances in IBD drug discovery have made great changes in disease management and also opened up the possibility of implementing precision treatment strategies for IBD.

Risk stratification based on clinical and molecular markers

Great heterogeneity in IBD makes it inappropriate and unreasonable for physicians to treat IBD patients with a unified therapeutic program. Disease course can substantially differ between individual patients. Some patients may undergo an aggressive disease course while others may experience a mild one. A link between a severe disease course and a poor disease outcome has been well documented [43]. Patients with an aggressive disease course need a timely and potent treatment, while a conventional stepup approach would suit a benign disease course [43]. Thus, the key of IBD management decision lies in screening out those patients with aggressive disease course at the early stage. So, doctors are advised to make risk stratification firstly according to various markers, and then select the most suitable therapy for patients [44]. Such a personalized treatment is closely correlated with better clinical outcomes, improved therapeutic efficacy and reduced risk of adverse events for IBD patients.

Clinical markers of disease course

Available data showed that patients with a diagnosis at an early age, perianal disease, complicated behaviors (structuring or penetrating lesions), and others were more likely to undergo an aggressive disease course [45]. We summarize the clinical markers in Table 1. However, onset age and disease location show some opposite effects on disease for patients with UC [46]. Different research methods and various sample sizes may explain these inconsistent results. As for the controversial factor, smoking, some studies demonstrated that it was a valuable predictor of unfavorable disease outcomes including complicated behaviors and the need for surgery, as well as the requirement for steroids/immunosuppressants and post-operative recurrence in patients with CD [47]. However, the concept that smoking cessation was linked to worse disease course for UC patients has been proposed [48]. Given that the benefits associated with smoking do not overweigh the potential risks, patients with UC are advised to give up smoking. What's more, IBD also shows strong sexual dimorphism in disease course. In comparison with male patients with CD, females frequently suffer from more severe clinical symptoms and disabilities [49]. It is noteworthy that most of these clinical markers were identified by retrospective studies, indicating a need of validating these markers in larger prospective cohort studies.

Endoscopy, a crucial tool for the assessment of mucosal inflammation and MH, is also of great importance in the prediction of disease course [64, 65]. Patients with extensive and deep ulcerations are at a higher risk of having an aggressive disease course [59]. Compared with patients exhibiting mild endoscopic lesions, the risk of colectomy was 5.43-fold higher in those with severe endoscopic lesions [59]. Conversely, endoscopic MH is associated with mild disease course [66]. Even so, endoscopic MH is not parallel to histologic remission [67]. Existing data showed that up to 40% of patients presenting with normal mucosa on endoscopy manifested mild to moderate inflammation on histopathology [67]. It is widely recognized that unresolved intestinal inflammation is associated with disease complications, colectomy, neoplasia and hospitalization, suggesting that endoscopy alone is likely inadequate to predict disease course in patients with IBD [68]. Therefore, combined analysis of endoscopic and histologic features may further reduce false negatives and increase the accuracy of prediction.

Table 1. Risk stratification based on clinical markers.

Markers	Roles in risk stratification	Sample number	Reference
Diagnosis at an early age	Predicted an aggressive disease course for CD	CD (1123)	[50]
Extensive disease	Predicted an aggressive disease course for CD and	CD (361);	[51, 52]
	UC, and medically refractory disease for UC	MR-UC (324),	
		non-MR-UC (537)	
Upper GI involvement	Predicted an aggressive disease course for CD	CD (358)	[53]
Ileal/ileocolonic involvement	Predicted an aggressive disease course for CD	CD (2105)	[54]
Perianal disease	Predicted an aggressive disease course for CD	CD (1123)	[50]
Complicated behaviors	Predicted an aggressive disease course for CD	CD (361)	[51]
Need of corticosteroids at initial	Predicted an aggressive disease course for CD and	CD (1123)	[50]
presentation	UC		
Fiber intake	Decreased risk for ileocolonic CD	CD (346), UC (456)	[48]
Older age at disease onset	Predicted an aggressive disease course for UC	UC (601)	[46]
Proximal disease location	Predicted an aggressive disease course for UC	UC (601)	[46]
Smoking	Showed bidirectional effects (protective or	CD (476), UC (630), IC	[48, 55, 56]
C	destructive) on disease course for CD and UC	(81);	
	,	CD (346), UC (456);	
		UC (6754)	
Female	Predicted more severe clinical symptoms and	CD (541)	[57]
	disabilities for CD		
Male	Predicted a high risk of developing CRC for UC	UC (4192), CD (3482)	[58]
Severe endoscopic lesions	Predicted an increased risk of penetrating	CD (102)	[59]
1	behaviors and colectomy for CD	× ,	
Endoscopic MH	Predicted of lower risk of relapse, colectomy and	UC (513), CD (227)	[60]
1	hospitalization for CD and UC		. ,
Coexisting with PSC	Increased risk of proximal disease extension,	UC (420);	[61–63]
5	dysplasia, CRC and colectomy for CD and UC	PSC-IBD (71), UC (142);	. ,
	jr, i,	IBD-neoplasia (43), IBD	
		(102)	
Co-occurrence of psoriasis	Predicted an aggressive disease course for UC	UC (420)	[61]

Abbreviations: CD: Crohn's disease; UC: ulcerative colitis; MR-UC: medically refractory-UC; GI: gastrointestinal; IC: indeterminate colitis; CRC: colorectal cancer; MH: mucosal healing; PSC: primary sclerosing cholangitis.

Given that IBD is an immune-mediated disease, IBD patients may present autoimmune comorbidities including primary sclerosing cholangitis (PSC), psoriasis and systemic lupus erythematosus (SLE). Co-occurrence of PSC or psoriasis contributes to a severe disease course in IBD patients [61, 69]. UC patients with PSC were more likely to suffer from progression of disease extension with a hazard ratio (HR) of 12.83 [61]. Recently, a close association between IBD and psoriasis has been reported in a Mendelian randomization study of 463,372 cases [70]. Given that autoimmune comorbidity always makes IBD management more difficult, special treatment and enhanced surveillance protocols in these patients are usually needed.

As it is known to all, IBD patients showed great heterogeneity in disease course. Different disease course often corresponds to different treatment strategies. Although lots of clinical markers have been identified to be associated with disease course, some of markers were not reliable or useful for the prediction, as the predictive accuracy is a little bit low [43]. In order to achieve adequate predictive accuracy, a prediction panel including clinical and other different class markers such as genetic, epigenetic, serological and fecal surrogates may be more helpful, and thus help physicians perform risk stratification and decide an appropriate treatment plan.

Genetic and epigenetic markers of disease course

In recent years, rapid progress has been made in the genetics of IBD. 320 risk alleles have been identified, some conferring susceptibility to IBD, while others related to disease course [6, 71]. We summarize genetic and epigenetic markers in Table 2.

The gene Nucleotide binding oligomerization domain containing 2 (NOD2) was the first susceptibility gene of CD, and three risk single nucleotide polymorphisms (SNPs) (R702W, G908R, and L1007finsC) have been studied extensively. A large-scale, multicenter study revealed that the three NOD2 SNPs were significantly associated with an aggressive disease course [86]. NOD2 risk SNPs conferred a 58% increase in the risk for colectomy [86]. In addition, risk genes including immunity related GTPase M (IRGM), TNF superfamily member 15 (TNFSF15), IL23R, etc. were also reported to be predictive markers of an aggressive disease course [6]. Although genetic markers are stable and heritable, they may only explain a small portion of variability. It has been shown that epigenetic markers (such as DNA methylation and non-coding RNAs) also shape the disease course of IBD patients [6, 87]. Tahara et al. claimed that higher methylation levels of protease-activated receptor2 (PAR2) and multi-drug resistance gene 1 (MDR1) were correlated with total colitis phenotypes, and the former was also identified as a potential marker in the prediction of refractory phenotypes of UC [79, 80]. In 2018, a Cambridge research team further observed that gut segment-specific DNA methylation profiles might be used as a clinically useful tool for predicting the requirement for biologics and the time to third treatment escalation [88]. Similarly, cellspecific DNA methylation signatures are also correlated with disease severity and colectomy in patients with UC [89]. One predictive model incorporating three methylation markers can predict treatment escalation with an HR of 5.19 [87]. From this point, DNA methylation markers are crucial in the evaluation of disease course. Furthermore, several studies also suggested that miRNAs are differentially expressed in IBD patients. Expression levels of the miR-29 family, miR-19-3p family and miR-200 family were

Table 2. Risk stratification based on genetic and epigenetic markers.

Markers	Roles in risk stratification	Sample number	Reference
NOD2	Predicted of stricturing/penetrating phenotype, ileal involvement and colectomy for CD	CD (316), UC (408), HC (205); CD (107)	[72, 73]
IRGM	Predicted of colectomy, stricturing phenotype, ileal involvement and perianal disease for CD	CD (263), UC (206), HC (245)	[74]
TNFSF15	Predicted of colectomy, stricturing phenotype and perianal fistula for CD, and medically refractory disease for UC	CD (906); MR-UC (324), non-MR-UC (537)	[52, 75]
IL23R	Predicted of stricturing/penetrating phenotype and ileocolonic involvement for CD	CD (1528)	[76]
PRDM1 IL12B	Predicted of penetrating phenotype for CD Predicted of medically refractory disease for UC	CD (1528) MR-UC (324), non-MR-UC (537)	[76] [52]
HLA-DRB1*0103 NFKBIL1	Predicted of extensive disease for UC Predicted of extensive disease and more severe disease for UC	UC (466), HC (2099) UC (155), HC (298)	[77] [78]
PAR2 (hypermethylation)	Predicted of extensive disease, steroid-dependent and steroid-refractory disease for UC	UC (84)	[79]
MDR1 (hypermethylation)	Predicted of extensive disease and earlier onset of disease for UC	UC (83)	[80]
RPS6KA2 (hypomethylation)	Predicted of stricturing/penetrating phenotype for CD, and extensive disease for UC	CD (121), UC (119), HC (191)	[81]
miR-29 family (low mucosa expression)	Predicted of stricturing phenotype for CD	CD (13)	[82]
miR-19-3p family (low serum expression)	Predicted of stricturing phenotype for CD	CD (108)	[83]
miR-200 family (low mucosa expression)	Predicted of stricturing phenotype for CD	CD (20), HC (16)	[84]
miR-31-5p, miR-215 and miR-223-3p (high mucosa expression)	Predicted of stricturing/penetrating phenotype for CD	CD (21), NIBD (14)	[85]
miR-149-5p and miR-203 (low mucosa expression)	Predicted of stricturing/penetrating phenotype for CD	CD (21), NIBD (14)	[85]

Abbreviations: NOD2: nucleotide binding oligomerization domain containing 2; CD: Crohn's disease; UC: ulcerative colitis; HC: healthy control; IRGM: immunity related GTPase M; TNFSF15: TNF superfamily member 15; MR-UC: medically refractory-UC; IL23R: interleukin 23 receptor; PRDM1: positive regulatory domain 1; IL12B: interleukin 12B; HLA-DRB1*0103: major histocompatibility complex, class II, DR beta 1, 0103; NFKBIL1: NFKB inhibitor like 1; PAR2: protease-activated receptor2; MDR1: multi-drug resistance gene 1; RPS6KA2: ribosomal protein S6 kinase A2; NIBD: non-IBD.

significantly decreased in patients with stricturing disease, in comparison with those with inflammatory phenotypes [90, 91]. In contrast, some other miRNAs are associated with complicated phenotypes [85]. One prospective study proposed that the expression level of miR-215 increased 4.8-fold when the disease behavior progressed from inflammatory phenotype to penetrating phenotype. In this regard, miRNAs may provide important clues in the assessment of disease course in IBD patients.

For patients with a higher risk of undergoing complicated disease and surgery, physicians are advised to make an aggressive therapeutic approach, aiming at improving disease outcomes. Although genetic and epigenetic markers show their potential role in the prediction of disease course and risk stratification, there are still some limitations. Firstly, although genetic markers are stable and heritable, their value is ethnicity-specific. Some risk loci are reliable markers in predicting disease course in one ethnic population, but may be absent in some other ethnicities, and showed no predictive value in this respect. Secondly, given that DNA methylation patterns are cell-specific, the epigenome differs substantially between sampling sites, which might result in dubious conclusions and limit their clinical application [92]. Thirdly, the association between a genetic/epigenetic marker and disease course is not always robust, therefore leaving uncertainty in its predictive value for disease course. Fourthly, the functional relevance of DNA methylation and miRNAs to intestinal stricturing/penetrating remains largely unknown [91]. Therefore, exploring DNA methylation and miRNA downstream targets is urgently required. Most importantly, considering that IBD results from the complex interplay between different contributors, a reliable disease course prediction must be based on the combined assessment of serological and fecal markers, in addition to clinical, genetic and epigenetic ones. Moreover, identified markers also should be validated and replicated in other ethnic groups, thereby generalizing them in clinical practice.

Serological markers of disease course

Existing and emerging serum markers have been studied extensively in IBD, thus providing valuable information into the prediction of disease course. Different kinds of antibodies against microbial components, neutrophils, and exocrine pancreas such as anti-*Saccharomyces cerevisiae* (ASCA), anti-outer membrane protein C (anti-OmpC), anti-neutrophil cytoplasmic antibodies (ANCA) and anti-glycoprotein 2 (anti-GP2) have been found in the serum of IBD patients. They are more likely to be detected in IBD patients in comparison with healthy controls, suggesting a possibility of differentiating IBD and controls by them [6]. More importantly, there is substantial evidence demonstrating that seropositivity to

Table 3. Risk stratification based on sero	logical	markers.
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Markers	Roles in risk stratification	Sample number	Reference
ASCA	Predicted of small bowel surgery,	CD (303);	[93–95]
	stricturing/penetrating phenotype, ileocolonic	CD (252), UC (53), HC	
	disease and perianal lesion for CD	(43);	
		CD (169), UC (102)	
AMCA	Predicted of surgery and	CD (103), CD-ITB (10),	[96–98]
	stricturing/penetrating phenotype for CD, and	ITB (9), HC (68);	
	severe disease course for UC	CD (913), UC (272), HC	
		(200)	
		NIBD (113);	
		CD (107), UC (88)	
ACCA	Predicted of steroid dependency and severe	CD (107), UC (88)	[98]
	disease course for CD and UC		
pANCA	Predicted of low risk of developing	CD (913), UC (272), HC	[97, 99]
	stricturing/penetrating phenotype and	(200)	
	receiving surgery for CD, and severe disease	NIBD (113);	
	course for UC	CC (17), UC (143), IBDU	
		(146)	
Anti-Fla2	Predicted of stricturing phenotype for CD	CD (252), UC (53), HC	[94]
		(43)	
Anti-Fla-X	Predicted of stricturing phenotype for CD	CD (252), UC (53), HC	[94]
		(43)	
Anti-CBir1	Predicted of ileal disease, surgery and	CD (796)	[100]
	stricturing/penetrating phenotype for CD		
Anti-GP2	Predicted of stricturing phenotype and perianal	CD (169), UC (102)	[95]
	disease for CD		
anti-OmpC	Predicted of small bowel surgery and	CD (303);	[93, 100]
	stricturing/penetrating phenotype for CD	CD (796)	
anti-I2	Predicted of small bowel disease, surgery,	CD (303);	[93, 101, 102]
	stricturing/penetrating phenotype and long	CD (196);	
	disease duration for CD	CD (142)	
CRP (high baseline	Predicted of intestinal surgery for CD, and the	CD (957);	[103–105]
level)	need of immunosuppressant treatment for CD	CD (313), UC (111),	
	and UC	IBDU (41);	
		CD (162)	
Albumin (low baseline	Predicted of surgery and severe postoperative	CD (957);	[103, 106, 107]
level)	complications for CD, and the need for	UC (710);	
	biologics and colectomy for UC	UC (97), CD (87), IBDU	
		(6)	

Abbreviations: ASCA: anti-Saccharomyces cerevisiae; CD: Crohn's disease; UC: ulcerative colitis; AMCA: anti-mannobioside carbohydrate IgG antibodies; ITB: intestinal tuberculosis; NIBD: non-IBD; ACCA: anti-chitobioside carbohydrate IgA; pANCA: perinuclear anti-neutrophil cytoplasmic antibodies; CC: crohn's colitis; IBDU: inflammatory bowel disease-unclassified; anti-GP2: anti-glycoprotein 2; anti-OmpC: anti-outer membrane protein C; anti-I2: anti-bacterial sequence I2; CRP: c-reactive protein.

these antibodies is associated with disease course in IBD patients. We summarize the serological markers in Table 3. tify UC patients with severe disease course with an area under curve (AUC) of 0.67 [98].

With regard to CD, several studies have identified an association between serological antibodies such as ASCA, anti-OmpC and anti-bacterial sequence I2 (anti-I2) and complicated disease and small bowel surgery [93, 94]. Furthermore, serum responses to flagellin and GP2 also help identify patients with complicated disease [94, 95]. A prospective study further suggested that increasing seropositivity to ASCA, anti-CBir1, and anti-OmpC was predictive for a faster disease progression. When patients with these three positive antibodies, they progress to penetrating and/or stricturing disease with an HR of 6.0, and receive CD-related surgery with an HR of 6.6 [100]. This is in line with the perspective of Schoepfer et al. that the risk of suffering from complicated disease and surgery was increased in patients with an increasing number of antibodies [94]. As for UC, pANCA + and ANCA-IgG levels were claimed to be associated with severe disease course [99, 108]. A French study also reported that combined analysis of anti-mannobioside carbohydrate IgG antibodies (AMCA) and anti-chitobioside carbohydrate IgA (ACCA) could correctly iden-

Of course, other conventional serological markers including Creactive protein (CRP) and albumin are also claimed to be associated with disease course in IBD [103, 106]. Combined analysis of more serum antibodies might increase the prediction accuracy to some extent, but we should also keep in mind that there is dissimilarity between association and predictivity. Only a small part of studies explored the predictive role of serum antibody markers in IBD patients. Most studies simply retrospectively analyzed associations between markers and disease course, while didn't investigate the predictive values of these markers in a prospective cohort. Besides, although predictive panels of different-class markers performed better in the disease course prediction, medical cost is another factor should be taken into account [109]. This indicated a need to do a cost-effectiveness analysis and develop a cost-effective panel for IBD patients. It is worth noting that the above serological surrogates also present in other diseases such as intestinal tuberculosis, irritable bowel syndrome (IBS), celiac disease and even healthy controls, which might render it suboptimal Table 4. Risk stratification based on fecal markers.

Markers	Roles in risk stratification	Sample number	Reference
Ruminococcus (high baseline level)	Predicted of stricturing phenotype for CD	CD (913)	[111]
Collinsella (high baseline level)	Predicted of penetrating phenotype for CD	CD (913)	[111]
Veillonella (low baseline level)	Predicted of penetrating phenotype for CD, and severe disease course for UC	CD (913); UC (48), HC (48)	[111, 112]
Rothia (low baseline level)	Predicted of stricturing phenotype for CD	CD (913)	[111]
Bacteroides (low baseline level)	Protected from severe disease course for UC	UC (48), HC (48)	[112]
F. prausnitzii (low baseline level)	Predicted of severe disease course for UC	UC (48), HC (48)	[112]
Proteobacteria (high baseline level)	Predicted of severe disease course for CD and UC	CD (72), UC (51), HC (73)	[113]
FC (high level)	Predicted of colectomy and pouchitis for UC, and postoperative recurrence for CD and UC	UC (90); CD (135); UC (60)	[114–116]
FL (high level)	Predicted of pouchitis for UC	UC (60)	[116]
Fecal BAFF (high level)	Predicted of severe disease course for UC	CD (44), UC (49), IBS (27), HC (26)	[117]
Fecal NGAL (high level)	Predicted of severe disease course for CD and UC	UC (43), CD (30), IEC (21), IBS (21), HC (23)	[118]

Abbreviations: CD: Crohn's disease; UC: ulcerative colitis; HC: healthy control; F. prausnitzii: faecalibacterium prausnitzii; HC: healthy control; FC: fecal calprotectin; FL: fecal lactoferrin; BAFF: B cell-activating factor of the TNF family; IBS: irritable bowel syndrome; IEC: infectious enterocolitis; NGAL: neutrophil gelatinase-associated lipocalin

in the prediction of disease course and discrimination of disease subtypes [110]. Now that different studies set various thresholds of serum antibodies in different cohorts, this might bring additional hurdles to explain these test results, thereby limiting the clinical application in other cohorts. So, it is absolutely a critical need to validate these results in larger, external and prospective cohorts.

Fecal markers of disease course

It has increasingly become apparent that fecal microbiome plays a critical role in the development and progression of IBD. Explorations in fecal microbiome not only cast insight into the complex pathogenesis of IBD, but also give a new perspective and way to evaluate of disease course. We summarize the fecal markers in Table 4.

In 2017, the RISK study clearly demonstrated that gut microbiota was significantly associated with disease phenotypes [111]. Ruminococcus and Collinsella are enriched in patients with stricturing/penetrating behaviors. While, the levels of Rothia and Veillonella are deceased in complicated disease [111]. As for patients with UC, different kinds of species of microbes were also claimed to be associated with severe disease course [112]. These findings provide additional information about the discriminant power of fecal bacteria between different disease phenotypes and courses. One year later, a Chinese study team also made a similar conclusion that different kinds of gut microbiota conferred risk to different phenotypes [113]. Most importantly, this study further revealed consistent microbial alteration patterns between Chinese and Western IBD patients, suggesting the possibility of using microbial markers to classify IBD patients across different ethnicities [113]. Although microbiota markers showed great potential in risk prediction, they haven't been broadly applicable in clinical practice. The following factors should be considered before application. Firstly, it is an established fact that diet, smoking, drugs, etc. markedly influence the composition and diversity of the microbiome [43, 119]. Some studies didn't take these confounding variables into consideration, which might affect the reliability and accuracy of results. Secondly, microbiota can indeed add value to the prediction of disease course, but it is not specific to IBD. Other diseases such as infective enteritis, celiac disease and IBS can also influence its form and diversity. Further work is warranted to elucidate its specific association with IBD. Thirdly, the functional consequences of most microbiota are unclear. So, conducting a metabolomics study is definitely needed. Fourthly, microbial shift in stool samples is not parallel with that in tissue samples [120]. Gevers et al. claimed that microbial imbalance was less seen in stool samples, but more often in tissue samples [120]. Therefore, additional efforts are required to further study the microbial community network in different intestinal segments. Combined analyzing microbiota markers in stool and tissues may be more reliable, but tissue samples must be collected by invasive endoscopy, which might increase medical expenses and expose patients to additional risks caused by endoscopy. Based on the above, exploring more reliable and cost-effective fecal markers is in desperate need

Besides the fecal microbiome, fecal calprotectin (FC) is now widely used as a reliable and noninvasive marker in assessing disease activity and differentiating IBD [121]. Patients with increased FC are at a higher risk of receiving colectomy and having postoperative recurrence [114, 115]. FC was superior to CRP and Crohn's Disease Activity Index (CDAI) in the reflection of the presence and severity of recurrence [115]. As for UC patients, several studies claimed that higher levels of FC were more often presented in patients with pouchitis [116]. It is important to note that the levels of FC were elevated two months before the confirmed diagnosis of pouchitis [116]. Based on these findings, FC might be a prominent marker in the prediction of postoperative recurrence. Other fecal markers including fecal lactoferrin (FL), fecal B cell-activating factor of the TNF family (BAFF), fecal neutrophil gelatinase-associated lipocalin (NGAL) also show their potential role in the evaluation of disease course [117, 118, 122]. However,

as mentioned above, we should pay attention to the difference between association and prediction. So, the predictive value should be validated further.

Combined predictive models

Analyzing one class of markers alone cannot ensure an accurate prediction of disease course, combined analysis of differentclass of markers such as clinical, genetic, epigenetic, serological and fecal surrogates may facilitate the prediction process. The RISK study developed a competing-risk model consisting of age, race, disease location, serologic markers and extracellular matrix gene profiling. It could predict complicated disease in CD patients with an AUC of 0.72 [111]. Similarly, another web-based system dynamic model incorporating disease location, serologic markers, NOD2 polymorphisms, and an interaction term between perianal disease and ASCA could correctly identify a high-risk population (developing strictures/fistulas, or receiving surgery over a three-year period) with a high concordance index [109]. More recently, a promising model including six genetic SNPs, ileal location, and three specific antibodies can predict intestinal surgery and/or complicated disease at 5 years with an AUC of 0.84 [123]. In general, the combined predictive model outperforms the single predictive model in helping physicians perform risk stratification and decide an appropriate treatment plan. But the cost of examinations and genetic heterogeneity should be taken into consideration when interpreting these results.

Available data indicate that IBD patients with an aggressive disease course are more likely to undergo frequent flares, disease complications, treatment refractory, bowel surgeries and frequent hospitalization [124]. Some severe patients even present stricturing and/or fistulizing disease and have to get abdominal surgery at the time of diagnosis. The intestinal surgery rate is as many as 80% and 30% for CD and UC, respectively [125]. Undoubtedly, physicians should do risk stratification before embarking on treatment. Any one-size-fits-all therapeutic approach is improper [43]. Given that bowel damage is progressive, accumulative and nearly irreversible, any delayed and inadequate treatment may accelerate disease progression, especially in those severe IBD patients. Early and progressive therapeutics can mitigate the disabling disease course and even alter the natural history of IBD. Therefore, patients with an aggressive disease course need a timely and potent treatment. A combined therapy of biologics and immunomodulators (even small molecule inhibitors) is recommended for these patients. For severe perianal fistulizing CD, early surgical treatments including abscess drainage, abscess setons, fistulotomy, and ligation of the intersphincteric fistula tract are also recommended [126]. With respect to intestinal stricturing/fistulizing CD, the anti-TNF biologic and ileocolic resection is the optimal pharmacotherapy and surgical treatment choice, respectively [127]. For acute severe ulcerative colitis (ASUC), besides corticosteroids and anti-TNF biologic salvage therapy, timely colectomy should be taken into consideration. While, a conventional step-up approach would suit those mild IBD patients. Conventional treatments including 5-aminosalicylate (5-ASA), corticosteroids, immunomodulators and others are recommended [43]. This personalized treatment not only reduces unnecessary expenses, but also decreases the unnecessary risk of adverse events including myelotoxicity, opportunistic infection and lymphoma for patients with an indolent disease course. Moreover, it also markedly improves clinical outcomes for patients with an aggressive disease course [90, 128]. Even so, the challenge remains to select the most suitable drugs for each individual patient, given that different patients show significant differences in drug metabolism and treatment response. Thus, physicians are advised to make an individualized therapeutic regimen based on the clinical characteristics and molecular markers for each patient.

Precision treatment with key medications

Despite many drugs showing promising potential in the treatment of IBD, unfortunately, the pharmacokinetics and pharmacogenetics vary between different patients with IBD. Some patients respond well to them with no adverse events, while others have lower response rates with serious adverse reactions. Therefore, adequate curative effects should be balanced with adverse events associated with their use before treatment. Here, this review will only discuss these well-studied drugs for IBD, namely thiopurines (azathioprine and 6-mercaptopurine), infliximab, adalimumab, vedolizumab, and ustekinumab (Table 5).

Thiopurines

Thiopurines, conventional immunosuppressants, have extremely complicated metabolic pathways. Taking azathioprine for example, azathioprine changes into 6-mercaptopurine after absorption by the GI tract. 6-mercaptopurine can then be metabolized through three competing pathways: conversion into 6-thioinosine monophosphate (6-TIMP) by hypoxanthine phosphoribosyltransferase (HGPRT); methylation by TPMT into 6-MMP that is responsible for hepatotoxicity; and conversion into 6-thiouric acid (6-TU) by xanthine oxidase (XO). 6-TIMP can then be successively metabolized into 6-thioxanthosine monophosphate (6-TXMP) and 6-thioguanine nucleotides (6-TGNs) by inosine-5-monophosphate dehydrogenase (IMPDH) and guanidine-5-monophosphate synthetase (GMPS), respectively(Fig. 2) [172]. Thiopurines play a wellestablished role in the induction and maintenance of remission, facilitation of MH, and prevention of postoperative recurrence for IBD patients. Such good therapeutic effects are directly related to their metabolites 6-TGNs, which are also responsible for the common side effect, myelosuppression [172]. Similarly, an increased concentration of another metabolite 6-methylmercaptopurine (6-MMP) is involved in hepatotoxicity [173]. Available data indicated that thiopurines had to be reduced or discontinued due to adverse effects in about 34%-35% of patients [174].

Thiopurine S-methyltransferase (TPMT), and nudix hydrolase 15 (NUDT15) gene variants can influence the activities of important enzymes implicated in the metabolism of thiopurines. Therefore, pharmacogenetics analyses may add value to treatment decisions and individualized treatment. One population frequency analysis of TPMT alleles showed that TPMT*3A is the most common allele in Caucasians, while Asian and African populations often present with TPMT*3C [175]. In the Caucasian population, approximately 11% of individuals are heterozygous carriers with intermediate TPMT activity, and only 0.3% are homozygous for TPMT variants with low/absent TPMT activity [173]. Thus, patients with TPMT variants are prone to develop myelosuppression when compared with those with wild-type genotypes [129]. Notably, the TPMT variant allele frequency is significantly lower in Asians than that in European populations [130]. The low frequency of TPMT variants in Asians limits the clinical value of predicting thiopurine-induced myelosuppression. It is also noteworthy that TPMT variants cannot explain the overall myelosuppression, suggesting other contributing factors should be explored further. Sutiman et al. reported that NUDT15 (p. Arg139Cys) conferred a 22.9-fold increased risk of leukopenia in Asian IBD patients

Drugs God response Poor response Thiopurines N/A N/A Thiopurines N/A N/A Inflixinab Concurrent immuomodulator treatment: male; non-stricturing/penetrating phenotype; IL23R N/A Inflixinab Concurrent immuomodulator treatment: male; non-stricturing/penetrating/pene	Poor response Adverse events NIA MA NIA TPMT: NUDT15 Smoking: older age at first dose: IL23R TPMT: NUDT15 Finable: decreasing variants): caspase 9 3 (CC/CT); HLA-DQA1*05 (A>-G); ATG16L1 (rs2241880, AA); PHACTR3 (rs6100556, TT); CXCL12 (rs1050884, CC/CT); p-ANCA+/ASCA-; high baseline WBC; high CATH; high FC TMA Male: Smoking: family history of IBD; infliximab failure; EIMs; perianal disease; high CDAI; high BMI; FasL (Lrs763110, CC genotype); TVF-308 (Intra1800629, AA/GA); IL17A N/A Male: Smoking: family history of IBD; infliximab failure; EIMs; perianal disease; high CDAI; high BMI; FasL (Intra36210, ACG); IU17A N/A Male: Smoking: family history of IBD; infliximab failure; EIMs; perianal disease; high CDAI; high BMI; FasL (Intra36210, CG genotype); TVF-308 (Intra800629, AA/GA); IL17A N/A infliximab failure; EIMs; perianal disease; high CDAI; high Escherichia: Shigella N/A	
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ATG1611 (rs10210302, CT/TT); TLR2 (rs3804099, TC/CC); TLR4 (rs5030728, GA/AA); TNFR5F1A (rs4149570, TT); MIF (rs755622, GG); TNFa (rs361525, GG); IRGM (rs13361189, TT); high CRP*; high mTNF+ cells; high Barnesiell; high Anaerostipes; high Tyzzerella; high Lachnoclostridium; high Lachnospiraceae_unclassified; high F. prausnitzii	Male; Smoking; family history of IBD; Male; Smoking; family history of IBD; infliximab failure; EIMs; perianal disease; high CDA1; high BMI; FasL (Lrs763110, CC genotype); TNF-308 (Lns1800629, AA/GA); IL17A (rs2275913, AA/AG); low FT3/fT4; high CD25; high IL-5; high Escherichia-Shigella	$ \begin{array}{c} CD \left(17 \right) UC \left(6 \right), BDU \\ (6), \\ CD \left(36 \right) UC \left(26 \right) \\ JIA \left(18 \right), HC \left(8 \right); \\ CD \left(100 \right); \\ CD \left(1610 \right); \\ CD \left(152 \right), UC \left(110 \right); \\ CD \left(122 \right); \\ CD \left(240 \right), UC \left(93 \right), IC \\ (7); \\ UC \left(191 \right); \\ CD \left(35 \right); \\ CD \left(483 \right); \\ CD \left(483 \right); \\ CD \left(483 \right); \\ CD \left(1610 \right); \\ CD \left(1610 \right); \\ CD \left(1610 \right); \\ CD \left(162 \right); \\ CD \left(162 \right); \\ CD \left(162 \right); \\ CD \left(121 \right); \\ CD \left(997 \right); \\ UC \left(56 \right) \\ UC \left(56 \right) \end{array} $
ATG16L1 (rs10210302, CT/TT); TLR2 (rs3804099, TC/CC); TLR4 (rs5030728, GA/AA); TNFRSF1A (rs4149570, TT); MIF (rs555622, GG); TNFa (rs361525, GG); IRGM (rs13361189, TT); high CRP*, high mTNF+ cells; high Barnesiel!, high Anaerostipes; high Tyzzerella; high Lachnoclostridium; high Lachnospiraceae_unclassified; high F. prausnitzii	Male; Smoking; family history of IBD; infliximab failure; EIMs; perianal disease; high CDAl; high BMI; FasL (Drs763110, CC genotype); TNF-308 (Drs1800629, AA/GA); IL17A (rs2275913, AA/AG); low fT3/fT4; high CD25; high IL-5; high Escherichia-Shigella	(6) (6) (6) (106) (106) (1152), UC (106) (1152), UC (110); (1152), UC (110); (1152), UC (110); (1151) (1161) (1151) (1161) (1151) (1161)
ATG16L1 (rs10210302, CT/TT); TLR2 (rs3804099, TC/CC); TLR4 (rs5030728, GA/AA); TNFRSF1A (rs4149570, TT); MIF (rs555622, GG); TNFa (rs361525, GG); IRGM (rs13361189, TT); high CRP*; high mTNF+ cells; high Barnesieli; high Anaerostipes; high Tyzzerella; high Lachnoclostridium; high Lachnospiraceae_unclassified; high F. prausnitzii	Male; Smoking; family history of IBD; N/A infliximab failure; EIMs; perianal disease; high CDAI; high BMI; FasL (□rs763110, CC genotype); TNF-308 (□rs1800629, AA/GA); IL17A (rs2275913, AA/AG); low fT3/fT4; high CD25; high Escherichia-Shigella	$\label{eq:relation} \begin{tabular}{lllllllllllllllllllllllllllllllllll$
ATG16L1 (rs10210302, CT/TT); TLR2 (rs3804099, TC/CC); TLR4 (rs5030728, GA/AA); TNFRSF1A (rs4149570, TT); MIF (rs555622, GG); TNFa (rs361525, GG); IRGM (rs13361189, TT); high CRP*; high mTNF+ cells; high Barnesiell; high Anaerostipes; high Tyzzerella; high Lachnoclostridium; high Lachnospiraceae_unclassified; high F. prausnitzii	Male; Smoking; family history of IBD; Male; Smoking; family history of IBD; infliximab failure; EIMs; perianal disease; high CDAI; high BMI; FasL (Drs763110, CC genotype); TNF-308 (Drs1800629, AA/GA); IL17A (rs2275913, AA/AG); low fT3/fT4; high CD25; high IL-5; high Escherichia-Shigella	JIA (18), HC (8); CD (160); CD (152), UC (100); CD (240), UC (93), IC (7); UC (191); CD (240), UC (93), IC (7); UC (191); CD (191); CD (102); CD (102); CD (102); CD (102); CD (102); CD (102); CD (102); CD (115); CD (256); CD (115); CD (257); CD (115); CD (297); UC (56)
ATG16L1 (rs10210302, CT/TT); TLR2 (rs3804099, TC/CC); TLR4 (rs5030728, GA/AA); TNFRSF1A (rs4149570, TT); MIF (rs555622, GG); TNFa (rs361525, GG); RGM (rs13361189, TT); high CRP*; high mTNF+ cells; high Barnesiell; high Anaerostipes; high Tyzzerella; high Lachnoclostridium; high Lachnospiraceae_unclassified; high F prausnitzii	Male; Smoking; family history of IBD; N/A infliximab failure; EIMs; perianal disease; high CDAI; high BMI; FasL (□rs763110, CC genotype); TNF-308 (□rs180629, AA/GA); IL17A (rs2275913, AA/AG); low fT3/fT4; high CD25; high IL-5; high Escherichia-Shigella	CD (1610); CD (1610); CD (152), UC (110); CD (240), UC (93), IC (7); UC (1911); CD (35); CD (35); CD (35); CD (35); CD (410); CD (410); CD (412); CD (422); UC (64); CD (115); CD (121); CD (251); CD (251); C
ATG16L1 (rs10210302, CT/TT); TLR2 (rs3804099, TC/CC); TLR4 (rs5030728, GA/AA); TNFRSF1A (rs4149570, TT); MIF (rs555622, GG); TNFa (rs361525, GG); RGM (rs13361189, TT); high CRP*; high mTNF+ cells; high Barnesiell; high Anaerostipes; high Tyzzerella; high Lachnoclostridium; high Lachnospiraceae_unclassified; high F. prausnitzii	Male; Smoking; family history of IBD; N/A infliximab failure; EIMs; perianal disease; high CDAI; high BMI; FasL (□rs763110, CC genotype); TNF-308 (□rs1800629, AA/GA); IL17A (rs2275913, AA/AG); low fT3/fT4; high CD25; high IL-5; high Escherichia-Shigella	CD (152), UC (110); CD (240), UC (93), IC (7); UC (191); CD (35); CD (35); CD (35); CD (35); CD (1610); CD (1610); CD (1610); CD (1610); CD (1610); CD (1610); CD (1610); CD (1610); CD (1610); CD (115); CD (255); CD (
ATG16L1 (rs10210302, CT/TT); TLR2 (rs3804099, TC/CC); TLR4 (rs5030728, GA/AA); TNFRSF1A (rs4149570, TT); MIF (rs555622, GG); TNFa (rs361525, GG); RGM (rs13361189, TT); high CRP*; high mTNF+ cells; high Barnesiell; high Anaerostipes; high Tyzzerella; high Lachnoclostridium; high Lachnospiraceae_unclassified; high F. prausnitzii	Male; Smoking; family history of IBD; N/A infliximab failure; EIMs; perianal disease; high CDAI; high BMI; FasL (□rs763110, CC genotype); TNF-308 (□rs1800629, AA/GA); IL17A (rs2275913, AA/AG); low fT3/fT4; high CD25; high IL-5; high Escherichia-Shigella	CD (240), UC (93), IC (7); UC (191); UC (191); CD (35); CD (35); CD (35); CD (35); CD (35); CD (410); CD (410); CD (410); CD (412); UC (64); CD (115); CD (25); CD (25); CD (115); CD (25); CD (
ATG1611 (rs10210302, CT/TT); TLR2 (rs3804090, TC/CC); TLR4 (rs5030728, GA/AA); TNFR5F1A (rs4149570, TT); MIF (rs755622, GG); TNFa (rs361525, GG); IRGM (rs13361189, TT); high CRP*; high mTNF+ cells; high Barnesiell; high Anaerostipes; high Tyzzerella; high Lachnoclostridium; high Lachnospiraceae_unclassified; high F. prausnitzii	Male; Smoking; family history of IBD; N/A infliximab failure; EIMs; perianal disease; high CDAI; high BMI; FasL (Lns763110, CC genotype); TNF-308 (Lns180629, AA/GA); IL17A (rs2275913, AA/AG; low FT3/FT4; high CD25; high IL-5; high Escherichia-Shigella	N/A (7); UC (191); CD (35); CD (35); CD (5); UC (56); CD (1610); CD (1610); CD (483); CD (115); CD (256); CD (115); CD (256); CD (115); CD (256); CD (115); CD (256); CD (115); CD (256); CD (115); CD (256); CD (256);
ATG16L1 (rs10210302, CT/TT); TLR2 (rs3804099, TC/CC); TLR4 (rs5030728, GA/AA); TNFR5F1A (rs4149570, TT); MIF (rs755622, GG); TNFa (rs361525, GG); IRGM (rs13361189, TT); high CRP*; high TJYzzerella; high Barnesiell; high Anaerostipes; high TJyzzerella; high Lachnospiraceae_unclassified; high F. prausnitzii	Male; Smoking; family history of IBD; infliximab failure; EIMs; perianal disease; high CDAI; high BMI; FasL (Drs763110, CC genotype); TDF-308 (Drs1800629, AA/GA); IL17A (rs2275913, AA/AG); low fT3/fT4; high CD25; high IL-5; high Escherichia-Shigella	UC (191); CD (5); CD (5); CD (56); CD (56); CD (1610); CD (483); CD (483); CD (483); CD (483); CD (483); CD (483); CD (483); CD (483); CD (256); CD (256); CD (256); CD (257); CD (257); C
ATG16L1 (rs10210302, CT/TT); TLR2 (rs3804099, TC/CC); TLR4 (rs5030728, GA/AA); TNFR5F1A (rs4149570, TT); MIF (rs755622, GG); TNFa (rs361525, GG); IRGM (rs13361189, TT); high CRP*, high mTNF+ cells; high Barnesiell; high Anaerostipes; high Tyzzerella; high Lachnospiraceae_unclassified; high F. prausnitzii Lachnospiraceae_unclassified; high F. prausnitzii	Male; Smoking; family history of IBD; infliximab failure; EIMs; perianal disease; high CDAI; high BMI; <i>Fast</i> (Tns763110, CC genotype); TNF-308 (Drs1800629, AA/GA); IL17A (rs2275913, AA/AG); low fT3/fT4; high CD25; high IL-5; high Escherichia-Shigella	CD (5) CD (5) CD (5) CD (56); CD (1610); CD (1610); CD (483); CD (102); CD (115); CD (1610); CD (1610);CD (1610); CD (1610); CD (1610); CD (1610); CD (1610);CD
ATG16L1 (rs10210302, CT/TT); TLR2 (rs3804099, TC/CC); TLR4 (rs5030728, GA/AA); TNFR5F1A (rs4149570, TT); MIF (rs755622, GG); TNFa (rs361525, GG); IRGM (rs13361189, TT); high CRP*; high mTNF+ cells; high Barnesiell; high Anaerostipes; high Tyzzerella; high Lachnospiraceae_unclassified; high F. prausnitzii Lachnospiraceae_unclassified; high F. prausnitzii	Male; Smoking; family history of IBD; infliximab failure; EIMs; perianal disease; high CDAI; high BMI; FasL ([Drs763110, CC genotype); TNF-308 ([Drs1800629, AA/GA]; IL17A ; (rs2275913, AA/AG); low fT3/fT4; high CD25; high IL-5; high Escherichia-Shigella	CD (5) UC (56); CD (1610); CD (1610); CD (483); CD (483); CD (483); CD (483); CD (483); CD (483); CD (483); CD (483); CD (483); CD (256); CD (102); CD (102); CD (115); CD (15); CD (
ATG16L1 (rs10210302, CT/TT); TLR2 (rs3804099, TC/CC); TLR4 (rs5030728, GA/AA); TNFR5F1A (rs4149570, TT); MIF (rs755622, GG); TNFa (rs361525, GG); IRGM (rs13361189, TT); high CRP*; high mTNF+ cells; high Barnesiell; high Anaerostipes; high Tyzzerella; high Lachnospiraceae_unclassified; high F. prausnitzii Lachnospiraceae_unclassified; high F. prausnitzii	Male; Smoking; family history of IBD; N/A infliximab failure; EIMs; perianal disease; high CDAI; high BMI; FasL (Drs763110, CC genotype); TNF-308 (Drs1800629, AA/GA); IL17A ; (rs2275913, AA/AG); low fT3/fT4; high CD25; high IL-5; high Escherichia-Shigella	N/A UC (56); CD (1610); CD (1610); CD (483); CD (483); CD (483); CD (483); CD (483); CD (483); CD (483); CD (483); CD (256); CD (256); CD (251); CD (251); CD (257); CD (257); C
	infliximab failure; EIMs: perianal disease; high TDA1; high BM1; Fast ("Drs763110, CC genotype); TNF-308 ("Drs1800629, AA/GA); IL17A (rs2275913, AA/AG); low fT3/fT4; high CD25; high IL-5; high Escherichia-Shigella	
	CDAI; high BMI; FasL ("TrSF63110, CC genotype); TNF-308 ("Trs1800629, AA/GA); IL17A ; (rs2275913, AA/AG); low fT3/fT4; high CD25; high IL-5; high Escherichia-Shigella	CC genotype); 17A high CD25;
	TNF-308 (□rs1800629, AA/GA); IL17A ; (rs2275913, AA/AG); low fT3/fT4; high CD25; high IL-5; high Escherichia-Shigella	17A high CD25;
	(rs2275913, AA/AG); low fT3/fT4; high CD25; high IL-5; high Escherichia-Shigella	high CD25;
	high IL-5; high Escherichia-Shigella	
Lachnospiraceae_unclassified; high F. prausnitzii		UC (64); CD (25); CD (115); CD (121); CD (997); UC (56)
		CD (25); CD (115); CD (121); CD (997); UC (56)
	CD (1 CD (1 CD (2 CD (5 CD (5 C) C) (5 CD (5 CD (5 C) C) C) (5 C) C) (5	CD (115); CD (121); CD (997); UC (56)
		CD (12.1); CD (997); UC (56)
		UC (56)

		Markers			
Drugs	Good response	Poor response	Adverse events	Sample number	References
Vedolizumab	Mayo score <9; CDAI score≤330; younger patients; longer disease duration*; Concurrent thiopurine treatment; high BLOC151; high TLCD1; high TMEM223; high <i>a</i> 4 <i>β</i> 7*; high Roseburia inulinivorans; high Burkholderiales	Smoking history; anti-TNF failure; active perianal lesions; \square concomitant steroid use; \square HBI score>10; high CRP*; low albumin; high PHLDA1; high OSBPL11; high CXCL3; high $\alpha 4\beta$ 1; high $\alpha E\beta7$; low $\alpha 4\beta7$ receptor saturation; absent $\alpha 4\beta7$ expressing cells	N/A	CD (161), UC (111) CD (1115); UC (620); CD (13), UC (19); CD (15), UC/IC (11); CD (15), UC (13); CD (12), UC (11); CD (212); CD (212); CD (212); CD (27), HC (148); CD (967), HC (148); CD (967), HC (148); CD (5) (5)	[153-164]
Ustekinumab	Colonic/ileocolonic disease; concurrent immunomodulator treatment; CRP>10mg/L; high TNF; high TBX21; high IL-23A; high IL-6; high FOXP3; high OSM; high OSMR; high <i>F. prausnitz</i> ii; high Baateroides; high clostridium citroniae; high Agathobaculum butyriciproducens; high Phascolarctobacterium faecium; low FC	HBI>7; structuring disease; perianal disease; intestinal resection history; current corticosteroid use; anti-TNF failure; Mayo score>6; high BMI; high albumin	N/A	CD (167); CD (104); UC (133); CD (123); CD (123); CD (102); CD (102); CD (28), UC (28), NIBD (111); CD (28), UC (77); CD (116); CD (116); CD (116); CD (1369)	[23, 165–171]
Abbreviations: TPMT: 843: Fas ligand-843; A PHACTR3: phosphatas perinuclear anti-neutr arthritis; TLR2: Toll lik factor alpha; BMI: bodi 1 subunit 1; IC: indeter Protein like 11; CCL33	Abbreviations: TPMT: thiopurine s-methyltransferase; NUDT15: nudix hydrolase 15; CD: Crohn's disease; STAT3: signal transducer and activator of transcription 3; II.23R: interleukin 23 receptor; UC: ulcerative colitis; FasL- 843: Fas ligand-843; ADAM17: ADAM metallopeptidase domain 17; SLCO1C1: solute carrier organic anion transporter family member 1C1; ATG16L1: autophagy related 16 like 1; CRP: c-reactive protein; DEF5: defensin 5; PHACTR3: phosphatase and actin regulator 3; IBDU: inflammatory bowel disease unclassified; CXCL12: C.X-C motif chemokine ligand 12; ECP: eosinophil cationic protein; F. prausnitzii: faecalibacterium prausnitzii: p-ANCA: perinclear ann-neutophil cytophasmic anti-bacicharomyces cerevisiae, WBC: while blood cell, CXTH: aehelicidin antimicrobial peptide; FC: fecal calprotectin; JF. ieral lactofernin; JA: juran i adoptathic perinclear ann-neutophil cytophasmic anti-bacicharomyces cerevisiae; WBC: while blood cell, CXTH: aehelicidin antimicrobial peptide; FC: fecal calprotectin; JF. ieral lactofernin; JA: juran necrosis farchrits; TLR2: Toll like receptor 2; TLR4: Toll like receptor 4; EIRS: extra-intestinal manie; WBC: motiv 5 disease activity index; HBI: harvey-backaw index; IL17A: interleukin 17A, fT3/fT3/fT3/fT3/fT3/fT3/fT3/fT3/fT3/fT3/	D: Crohn's disease; STAT3: signal transducer and actival arrier organic anion transporter family member 1C1; A sissified: CXCL12: C-X-C motif chemokine ligand 12; ECP: siae; WDC: white blood cell, CATH: cathelicdin antimicr iae: VMC: white blood cell, CATH: reathelicdin antimicr indestations; TNFRSFIA: TNF receptor superfamily m nanifestations; TNFRSFIA: TNF receptor superfamily m ne-to-thyroxin; CDAI: crohn's disease activity index; HB smembrane protein 223; $\alpha 4\beta7$: alpha 4 beta 7; PHLDA1: that E beta 7; TBX21: t-box transcription factor 21; NIBE	tor of transcription 3; II.23R: .TG16L1: autophagy related ' eosinophil cationic protein; F obial peptide; FC: fecal calpr ember 1A; MIF: macrophage i: harvey-bradshaw index; bl oleckstrin homology like dom): non-IBD; IL-23A: interleuk	interleukin 23 receptor; UC: u 6 like 1; CRP: c-reactive proto prausnitzii: faccalibacterium biectin; FL: fecal lactoferrin, JI migration inhibitory factor; migration inhibitory factor; 0CLS1: biogenesis of lyosoom cain family a member 1; OSBP n 23A; IL-6: interleukin 6; FO.	cerative colitis; FasL- in; DEF5: defensin 5 prausnitzii; p-ANCA A; juvenile idiopathic NFa: tumor necrosis 11 oxysterol binding (P3: forkhead box p3

Table 5. (Continued)

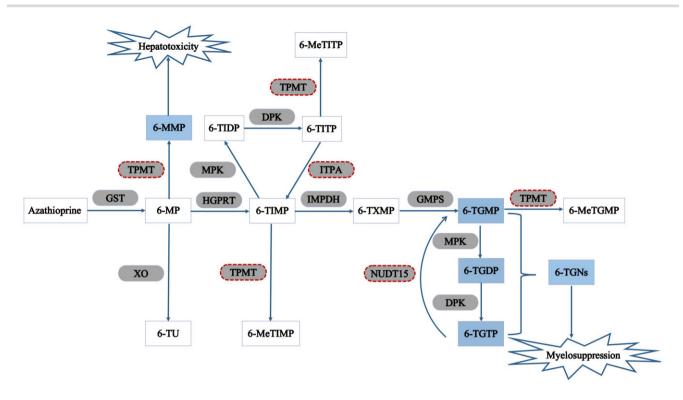


Figure 2. The metabolic pathways of azathioprine. Azathioprine changes into 6-mercaptopurine after absorption by the GI tract. 6-mercaptopurine can then be metabolized through three competing pathways: conversion into 6-TIMP by HGPRT; methylation by TPMT into 6-MMP; and conversion into 6-TU by XO. 6-TIMP can then be successively metabolized into 6-TXMP and 6-TGNs by IMPDH and GMPS, respectively. GST: glutathione s-transferase; 6-MP: 6-mercaptopurine; TPMT: thiopurine s-methyltransferase; 6-MMP: 6-methylmercaptopurine; XO: xanthine oxidase; 6-TU: 6-thiouric acid; HGPRT: hypoxanthine–guanine phosphoribosyl transferase; 6-TIMP: 6-thioinosine monophosphate; MPK: monophosphate kinase; 6-TIDP: 6-thioinosine diphosphate; DPK: diphosphate kinase; 6-TITP: 6-thioinosine triphosphate; 6-MeTITP: 6-methylthioinosine triphosphate; GMPS: inosine triphosphate synthetase; 6-TGMP: 6-thioguanine monophosphate; 6-TGTP: 6-thioguanine diphosphate; 6-TGTP: 6-thioguanine diphosphate; 6-TGTP: 6-thioguanine triphosphate; 6-TGTP: 6-thioguanine diphosphate; 6-TGTP: 6-thioguanine diphosphate; 6-TGTP: 6-thioguanine triphosphate; 6-TGTP: 6-thioguanine monophosphate; 6-TGTP: 6-thioguanine diphosphate; 6-TGTP: 6-thioguanine triphosphate; 6-TGTP: 6-thioguanine diphosphate; 6-TGTP: 6-thioguanine triphosphate; 6-TGTP: 6-thioguanine monophosphate; 6-TGTP: 6-thioguanine diphosphate; 6-TGTP: 6-thioguanine diphosphate; 6-TGTP: 6-thioguanine triphosphate; 6-TGTP: 6-thioguanine diphosphate; 6-TGTP: 6-thioguanine triphosphate; 6-TGTP: 6-thioguanine diphosphate; 6-TGTP: 6-thioguanine triphosphate; 6-TGTP: 6-thioguanine monophosphate; 6-MeTGMP: 6-methylthioguanine monophosphate.

[176]. Another European case-control study drew a similar conclusion and strongly recommended to detect NUDT15 polymorphisms before initiation of thiopurine treatment [177]. Given that NUDT15 genetic variants are more common in Asians, NUDT15 polymorphisms are claimed to be the better genetic surrogate for the prediction of thiopurine-induced myelotoxicity compared with TPMT genetic variants in Asians [130]. However, the correlation between adverse events of thiopurines and inosine triphosphate pyrophosphatase (ITPA) polymorphisms is fairly controversial [178, 179]. Further studies should be taken to elaborate on their correlations and guide treatment decisions. The question then arises whether TPMT/NUDT15 genetic testing should be systematically indicated in all patients who are going to receive thiopurines, considering that this testing is not cheap. In recent years, we proposed to foster value-based healthcare, a strategy to increase the quality and value of healthcare services by promoting the shift from volume-based payments to outcomes-based payments. So, several studies have done cost-effectiveness analyses of pretreatment screening TPMT/NUDT15 polymorphisms. The final results prove it as a cost-beneficial strategy [180, 181]. Therefore, prospective screening for TPMT and NUDT15 should be considered in principle before starting thiopurine therapy in various

Besides genetic markers, the roles of gut microbiota in predicting thiopurine treatment response should also be noted. Available data demonstrated that gut microbiota affect thiopurine biotransformation by releasing microbial enzymes [182]. Liu *et al.* found that *Bacteroides vulgatus* could encode thiopurine metabolic enzymes including GST, HGPRT, GMPS and IMPDH [183]. Besides the above enzymes, this study also suggested that *Escherichia coli* further possessed another critical enzyme, XO [183]. Several other gut bacteria including *Enterococcus faecalis*, *Bacteroides fragilis*, and *Pseudomonas aeruginosa* were also claimed to be responsible for azathioprine metabolism [184]. Based on these findings, we can conclude that gut microbiota might be a promising and novel tool for personalized thiopurine treatment of IBD. However, little study prospectively evaluates the predictive performance of baseline microbiota in thiopurine response. More studies are needed to fill this gap.

Dose reduction or even exclusion of thiopurines should be taken into account for patients with mutant genotypes. Recently, a Chinese research team conducted a randomized clinical trial and demonstrated that NUDT15 C415T-based dose optimization before treatment mitigated the risk of developing leucopenia in CD patients [185]. The predictive roles of these risk genes have been well confirmed in clinical practice. However, some subjects with wild-type genotypes still suffered from severe adverse events, indicating that other factors such as environmental, microbiota, other genetic predictors, etc. may account for the remaining toxicity. Further work is warranted to explore potential predictors and their interactions with thiopurine-induced adverse events, in order to achieve a precision selection of appropriate medication for individual patients. Other immunomodulators used for IBD including methotrexate, ciclosporin and tacrolimus are also effective in achieving steroid-free remission. Predictive markers of treatment response and adverse events have not been fully investigated, therefore more studies are needed to identify predictors for these medications.

Albeit immunomodulators are widely used in clinical practice and show acceptable efficacy in the treatment of IBD, many discontinue these treatments partly due to toxicity, intolerance, unfavorable response rate, and inconvenience of application. Even among those who continue receiving immunomodulators, a great number of patients fail to improve the aggressive disease course and poor prognosis. Therefore, appealing biological agents such as infliximab, adalimumab, vedolizumab, and ustekinumab have been added to the treatment options for those with a moderate to severe disease course. Adequate therapeutic effects make them highly acceptable for patients and physicians, while there are also plenty of primary non-responders and secondary non-responders to biological agents. Additionally, high cost, increased risk of opportunistic infections and malignant tumor, and inconvenience of parenteral application limit their routine clinical use. Current evidence suggests that clinical features, genetic surrogates, and some other predictive molecular markers can assist physicians in distinguishing responders from non-responders with good accuracy. Therefore, physicians should assess disease status carefully and make an individualized treatment plan based on existing markers, in order to minimize the risk of adverse events, maximize treatment effects, reduce medical costs, and improve the quality of life of patients to the most extent.

Infliximab

Infliximab, a chimeric monoclonal IgG1 antibody against TNF- α , shows excellent therapeutic effects in CD and UC, especially in those with moderate to severe disease and medically refractory disease [186]. However, nearly 40% of patients do not show an early response, and 23%–46% develop a secondary loss of response over time [187]. Some different classes of factors are of great value in predicting the initial and sustained response to infliximab, which can assist physicians in determining individualized therapy for individual patients.

Among these various predictive factors of response to infliximab, genetic predisposing factors are the most studied. Jürgens et al. concluded that homozygous carriers for IBD risk-increasing IL23R variants were more prone to respond to infliximab than those who are homozygous for IBD risk-decreasing IL23R variants (74.1% vs. 34.6%) [133]. In addition to alleles of IL23R, a favorable treatment response is also linked to the Fas ligand (FasL)-843 CC/CT genotype in CD [134]. Besides, a significant association between HLA-DQA1*05 and poor response in patients with IBD has been found in some studies [141, 188]. HLA-DQA1*05 carriers are at a higher risk of developing antidrug antibodies (ADAs) and losing therapeutic response [141]. However, Laserna-Mendieta et al. drew a negative result that HLA-DQA1*05 didn't affect infliximab response [135]. Different standards of treatment response may explain this opposite conclusion. In another study, an apoptotic pharmacogenetic index (API) based on genetic and clinical data for the prediction of response rates has been developed. Higher API scores implied a higher response rate to infliximab in patients with CD [189]. Other genes such as autophagy related 16 like 1 (ATG16L1), C-X-C motif chemokine ligand 12 (CXCL12), FasL-670, etc. were also claimed to be valuable markers of predicting treatment response [134, 142]. Besides single genetic predictor, gene expression signatures also add value to precision prediction. By combined expression analysis of five identified genes, the responders can be distinguished from non-responders in UC patients with a sensitivity and a specificity of 95% and 85%, respectively [190].

The predictive value of gene expression profiles in CD patients has also been investigated. Arijs *et al.* reported that differentially expressed gene profiles were capable of predicting response to infliximab with an overall accuracy of 100% based on microarray analysis [191]. For those patients carrying risk haplotypes, concomitant immunomodulator treatment or switch therapy may be the next step of treatment. Based on these findings, we can make a conclusion that pharmacogenetics paves a novel way to the prediction of treatment response. If possible, prior-to-treatment screening for risk genetic markers should be considered in routine clinical practice.

Aside from genotype testing, other clinical and serological markers may provide additional information on the prediction of infliximab response. Previous studies claimed that long disease duration, smoking and others were associated with poor response to infliximab, while concurrent immunomodulator treatment and non-stricturing/penetrating phenotype were possible predictors of favorable response rates [131, 132]. However, some patients with clinical risk factors show adequate response to infliximab and even gain MH, indicating that relying on clinical factors alone cannot guarantee an accurate prediction. Studies in serological, histologic, and fecal markers might provide more valuable and reliable information. Studies suggested that a high baseline CRP was associated with a better response rate, while p-ANCA+/ASCAwas a hopeful predictor for lower response rates to infliximab [133, 136]. Moreover, serum and mucosal proteomic profiling can also add value to a more precise prediction [137]. Pre-treatment serum infliximab-modulated immune profiling including oncostatin-M (OSM), TNFSF14 and others was demonstrated to be helpful in the prediction of clinical response [192]. Caution needs to be exercised when interpreting these results, because some results were gained from a single-center study with a small sample. Moreover, these candidate proteomic markers have not been further validated, resulting in little clinical utility.

Gut microbiota is a key factor in the pathophysiology of IBD [193]. Available data strongly support that fecal surrogates can not only assist physicians in differential diagnosis and assessment of disease activity, but also serve as clinically useful predictors of therapeutic response to infliximab. Analysis of the composition, abundance, and diversity of intestinal microbiome before and after the infliximab therapy may provide some clues about treatment response. A study demonstrated that six groups of fecal bacteria might be promising predictive markers of therapeutic response to infliximab [139]. In accordance with it, responders presented lower dysbiosis indexes and a higher number of faecalibacterium prausnitzii (F. prausnitzii) and Bifidobacteriales when compared to non-responders, suggesting that F. prausnitzii and Bifidobacteriales could be candidate markers of predicting therapeutic response of infliximab [137, 138]. Moreover, in virtue of non-invasiveness, intestinal-specificity and stability, fecal proteins such as FC and FL are also claimed to be potential markers for prediction. Although these fecal markers were reported to be associated with response rates, divergent results should also be noted [194]. Moreover, most of these findings are at the candidate discovery stage of the biomarker pipeline, more efforts are needed to qualify and verify these candidate predictors in larger populations. Therefore, larger, prospective, and independent studies should be carried out to clarify their roles in predicting treatment response, and thus achieve precise prediction and avoid exposure of non-responders to infliximab.

One class of predictors is insufficient for an accurate prediction, therefore combined analysis of different-class markers may further improve the accuracy of prediction and assist in making individualized treatment regimens. This is a matter of prime importance when making a therapeutic plan. Recently, a combined panel of genetic and clinical surrogates showed an increased accuracy in the prediction of primary nonresponse, compared with a clinical-only panel (AUC 0.87 vs. 0.57) [195]. Dubinsky and colleagues combined analyzed genetic effects, clinical markers, and serological surrogates, and built a predictive model comprising of three "pharmacogenetic" loci, a known locus, p-ANCA positivity and diagnosis of UC in pediatric patients [196]. When the risk factors were more than two, the relative risk of non-response became 15-fold higher than those who had only two or fewer risk factors, with an AUC of 0.98. Similarly, Zhou et al. claimed that combined analysis of Clostridiales abundance, FC levels and CDAI could discriminate infliximab responders from non-responders with an accuracy of 93.8% [113]. Indeed, these findings provide a possibility for physicians to use a predictive model for the prediction of infliximab response, although they must be confirmed independently, on a larger scale, in a prospective cohort and also studied in an adult cohort.

Adalimumab

Adalimumab is one full recombinant human IgG1 antibody against TNF- α and shows great effectiveness in induction and maintenance of remission in CD and UC patients [197, 198]. It is also used as a second-line therapy for moderate to severe active patients and those nonresponse or intolerance to infliximab [199]. Similar to infliximab, a great number of patients do not respond to adalimumab. About one-third of CD patients fail to respond to adalimumab in one-year follow-up [200]. More importantly, even among primary responders, 18.2% of patients suffer from secondary adalimumab failure, and 37% of cases need dose escalation [149]. Therefore, discriminating responders from non-responders prior to initial therapy becomes particularly important.

Some genetic markers might aid physicians in predicting the therapeutic response to adalimumab. Koder et al. suggested that patients with ATG16L1 (rs10210302) CT/TT genotype were more likely to achieve biological response, compared to those with CC genotype (OR: 9.44) [144]. Moreover, other candidate predictive markers including Toll like receptor 2 (TLR2), TNF receptor superfamily member 1A (TNFRSF1A), FasL etc. were also claimed to be associated with adalimumab response [142, 145, 150]. However, it is noteworthy that different standards of treatment response are set in various studies. Some investigated the clinical response rates, whereas others explored the difference in endoscopic remission and histologic remission between responders and nonresponders. Besides, these identified genetic variants show relatively small effect sizes on composite disease response scores [194]. So, more risk SNPs with large effect sizes are needed to be explored. What's more, genetic heterogeneity across ethnicities also should be noted.

In addition to genetic predictors, predictive roles of clinical, serological and fecal markers have also been identified. Available data showed that demographic and disease characteristics including smoking, primary failure to infliximab, EIMs and others are correlated with a loss of response and dose escalation [140, 149]. With respect to CRP, contradictory results have been found. Some studies suggested an association between low baseline CRP and good treatment response, while other studies claimed that high baseline levels of CRP were associated with a better therapeutic response [194, 197, 201]. Such inconsistency can be explained by the fact that CRP is not only associated with inflam-

matory phenotypes, but also predictive of more severe disease [194]. Besides the conventional inflammatory protein, a team from Switzerland further investigated the predictive role of T-cells from peripheral blood mononuclear cells (PBMCs). A serological predictive panel comprising T-cell surface receptor (CD25) and related cytokine markers (IL-5) was generated, which performed effectively with an acceptable accuracy of 91% [152]. Recent advances in endoscopy also provide a possibility for physicians to predict treatment response. in vivo molecular imaging by confocal laser endomicroscopy (CLE) and fluorescent antibodies to TNF revealed that the patients with increased baseline levels of mTNF + cells had significantly higher short-term response rates than those with decreased numbers of mTNF + cells [147]. This result could be explained by the fact that high levels of mTNF + cells indicate high numbers of targets for anti-TNF biologics. Therefore, the response rates increase. This finding does hold promise for endoscopy-based treatment prediction. Fecal markers also show promising potential in the prediction of therapeutic response to adalimumab. The abundance of protective microbiota including Barnesiella, Anaerostipes, Tyzzerella, etc. was increased in responders. Conversely, a decrease in pathogenic bacteria Escherichia-Shigella was found in adalimumab-responsive patients [148]. From this point, these changed fecal microbiota are capable of predicting the treatment response to adalimumab. It is important to note that human gut microbiome is highly dynamic and personalized, but most microbiome studies concentrate on a single time point and certain patients (small sample size and specific ethnic group). Longitudinal studies of the long-term change of microbiome in responders and non-responders across different ethnicities are therefore required.

It should be noted that a single marker seems to be inadequate for the prediction of treatment response. So, Gorenjak *et al.* used machine learning support vector machines algorithm, and developed a prediction model consisting of the expression and genotype data of four potential genes [202]. This model showed a surprisingly high accuracy of 100% in predicting adalimumab response. More recently, Busquets *et al.* developed an algorithm comprising four microbial markers and used it to differentiate responders from non-responders, with a favorable sensitivity and specificity (93.33% and 100%) [203]. Furthermore, Bouhnik *et al.* assigned a value to different variables (clinical, laboratory, and imaging parameters) and constructed a prognostic score to aid precise prediction [204]. A higher prognostic score represents a high possibility of adalimumab response at week 24.

Given that adalimumab and infliximab are both anti-TNF- α agents, most predictors used in infliximab therapy might also be used in adalimumab treatment. However, an important issue deserves our close attention. For those with a loss of response to infliximab, the response rate to adalimumab varies significantly between different individuals. Some show astonishing response rates, while others are still non-responders. There may be various underlying factors influencing the responsiveness to adalimumab and infliximab respectively. Therefore, comparative studies are required to identify specific predictors of infliximab and adalimumab with the aim of improving the accuracy of prediction and avoiding the failure of second-line anti-TNF therapy of adalimumab.

Vedolizumab

Vedolizumab is a humanized, more selective, monoclonal antibody against gut-homing $a4\beta7$ integrin [154]. Well-known, three-phase, randomized controlled trials (GEMINI) demonstrated its adequate efficacy in induction and maintenance of remission for patients with CD and UC [154, 205]. Similar to infliximab and adalimumab, vedolizumab is not always an effective treatment. Available data suggested that the clinical response rates at week 14 after vedolizumab therapy are 49%–64% in CD and 43%–57% in UC, respectively [162, 206, 207]. However, even in these initial responders, approximately 20% of patients become secondary nonresponders and stop vedolizumab due to lack or loss of effectiveness [208, 209]. Hence, identifying predictors of treatment response to vedolizumab holds the key to precision treatment.

Existing evidence demonstrated that clinical features and serological biomarkers, as well as fecal surrogates and pharmacological parameters, are correlated with the therapeutic response of vedolizumab in IBD. The association between baseline disease activity and clinical remission rates has been confirmed in several studies. GEMINI 1 and 2 trials showed that patients with baseline Mayo score < 9 and CDAI score \leq 330 had higher clinical remission rates at week 6 and week 54 [207]. Other clinical characteristics such as smoking history, anti-TNF failure, active perianal lesions, etc. are also predictors of unfavorable response rates [154, 160, 161]. Different opinions regarding the association between disease course and vedolizumab response have been expressed. Patients with longer disease duration are more prone to lose response to vedolizumab [154, 207]. However, the contradictory finding was seen in another study [155]. The former can be explained by the fact that patients with longer disease duration are prone to have a severe disease course and to be treated with anti-TNF before, thus, they are at risk of losing response to vedolizumab. However, longer disease duration also results in very chronic inflammation and T cell exhaustion, indicating a good prognosis in chronic autoimmune disease [155, 210]. These findings again need further replication studies to validate their predictive roles in vedolizumab response.

Conventional serum markers may further assist physicians in evaluating the disease state and selecting the most appropriate patients. Current evidence shows that high baseline CRP and low baseline albumin are associated with poor response [155, 163]. However, whether CRP served as a positive or negative predictor of therapeutic response remains to be determined [194, 206]. Underlying factors including different outcome definition, different observation time, and confounding variables might contribute to these paradoxical findings. Recently, vedolizumab responders were claimed to have higher baseline expression of transmembrane protein 223 (TMEM223) in PBMC Treg cells in comparison with those non-responders. On the contrary, a high expression level of CXCL3 was suggested to be a negative marker of adequate response to vedolizumab [156]. Besides PBMC, transcriptional profiles of mucosal Treg cells also provided additional information about discrimination between vedolizumab responders and nonresponders [156].

Besides traditional inflammatory markers, specific changes in integrin expression profiles are also associated with treatment response. Schneider *et al.* demonstrated that the baseline frequencies of $\alpha 4\beta 7$ -expressing T cells were statistically lower in clinical responders than that in non-responders [211]. However, other studies drew the opposite conclusion that high baseline $\alpha 4\beta 7$ expression levels of T, B and NK cells predicted good therapeutic response [157, 158]. During vedolizumab therapy, an increased expression of $\alpha 4\beta 7$ integrin was associated with good clinical presentation, while increased levels of $\alpha 4\beta 1$ and $\alpha E\beta 7$ indicated bad outcomes [158]. Such contrasting conclusions provide an impetus for further studies to clarify the relationship between baseline $\alpha 4\beta 7$ integrin levels and vedolizumab response. In addition, $\alpha 4\beta 7$ receptor saturation was also identified as a candidate predictive biomarker. Non-responders often present lower $\alpha 4\beta 7$ receptor saturation rates at trough than responders, and the saturation rates are reduced over time [157]. In 2017, Rath *et al.* used CLE to detect $\alpha 4\beta 7$ expressing cells in colonic mucosa, and further suggested that absent $\alpha 4\beta 7$ expressing cells might lead to poor therapeutic response to vedolizumab [164]. These results certainly open a new approach for identifying patients who will benefit most from vedolizumab and add value to personalized vedolizumab therapy. However, this study only included five patients with CD, highlighting the need to conduct studies with a larger sample size and validating its predictive role in the UC patients.

As mentioned previously, a central role of gut microbiota has been confirmed in the pathophysiology of IBD. A recent study assessed its relationship with vedolizumab response. CD patients in remission (at week 14) had a higher baseline α -diversity and more abundant *Roseburia inulinivorans* and *Burkholderiales* species, compared with the patients with high disease activity [159]. Thirteen microbial pathways including branched chain amino acid (BCAA) synthesis were markedly enriched in quiescent CD patients, compared with non-remission patients. With the help of gut microbiota, physicians might predict the vedolizumab response more accurately and make a personalized therapeutic regimen according to individual microbiota characteristics of each patient.

However, the one class of markers alone performs imperfectly in predicting vedolizumab response. Therefore, researchers successfully developed a mixed model consisting of clinical data, microbial taxonomy and pathway relative abundance to predict treatment response with an AUC of 0.776, which outperformed each individual model established in their study [159]. Furthermore, another two scoring systems consisting of various clinical and serological markers have also been established and validated in patients with CD and UC [155, 212]. Dulai et al. assigned different values to various variables (medication history, surgery, disease behavior, albumin and CRP) and developed a vedolizumab response scoring system [212]. It performed effectively in the prediction of clinical remission, and MH with an AUC of 0.67 and 0.72, respectively [212]. As for UC, another scoring system consisting of different parameters including medication history, disease duration, endoscopic activity and albumin was constructed. When the score is below 26 points, patients are less likely to achieve corticosteroid-free remission at week 26 (the sensitivity and specificity is 93% and 15%, respectively) [155]. It should be noted that there are few data specifically investigating the effects of genetic variants, FL and serum antibodies such as ASCA and p-ANCA on the vedolizumab response prediction. Further studies are needed to clarify the relationships clearly.

Ustekinumab

Ustekinumab is a fully human monoclonal IgG _{1k} antibody to the p40 subunit of IL-12 and IL-23 [213]. Ustekinumab binds the common p40 subunit, blocks the biological activity of IL-12 and IL-23, and finally stops the inflammatory cascade [214]. Well-established UNITI trials demonstrated that ustekinumab is a more effective treatment than placebo in induction and maintenance therapy for patients with CD [23]. It has also been approved to treat moderate to severe UC patients in the UNIFI study [24]. Similar to the above biological agents, there were also lots of primary non-responders and secondary non-responders to ustekinumab. Moreover, some patients suffer from unacceptable side effects during the course of treatment. Therefore, exploring predictors of ustekinumab response and applying them to clinical practice become an essential part of the treatment work in IBD.

Initial studies revealed that patients with higher disease activity exhibited poorer response in the long term [165, 166]. CD patients with Harvey-Bradshaw index (HBI)>7 at induction have a lower likelihood of achieving clinical response at follow-up [165]. Disease locations and phenotypes may also provide clinically useful information for the prediction process. Structuring disease is a negative predictor of good clinical response, while patients with colonic/ileocolonic disease are more prone to have clinical response at 6 months [165]. Other clinical characteristics including female, previous anti-TNF failure and others may also help physicians to predict ustekinumab response [207].

As for genetic, serological, and fecal predictors, few studies investigated the associations with ustekinumab response in patients with IBD. Most studies focused on psoriasis. Several genetic studies claimed that SNPs in IL-12B and TNFAIP3 (TNF alpha induced protein 3) influence therapeutic response in psoriasis patients [215, 216]. However, genetic studies on IBD patients are still limited. A Japanese study team analyzed the mucosal gene expression pattern and found that the baseline expression levels of IL-23A, TNF, FOXP3 and others differed between ustekinumab responders and non-responders [168]. This opens the possibility of using mucosal gene expression patterns to predict therapeutic response in IBD. As for serological data, a previous study suggested that the response rates were higher in CD patients with baseline $CRP \ge 10 \text{mg/L}$ than that in those with CRP < 10 mg/L [167]. Overall, little is known about the effects of other serum inflammatory markers and antibodies such as ESR, ANCA and ASCA on the response rates of ustekinumab. Recently, the low baseline FC level was claimed to be a valuable predictor of good response to ustekinumab [171]. With respect to intestinal microbiota, the CER-TIFI study suggested that baseline microbial signatures could predict disease remission with acceptable accuracy [169]. The baseline Faecalibacterium and Bacteroides were significantly higher in patients in remission than that in non-remission patients six weeks after ustekinumab induction [169]. Thus, a random forest prediction model including several clinical and microbiota markers has been developed. It can successfully predict clinical remission and clinical response with an AUC of 0.844 and 0.733, respectively [169]. These findings suggested that baseline microbial metacommunity could help physicians identify□patients who will benefit most from specific treatment.

Based on the above findings, Ustekinumab Clinical Decision Support Tool (UST-CDST) has been developed. The UST-CDST is calculated using five markers including anti-TNF- α exposure, bowel surgery, fistulizing disease, smoking and albumin level. Then, Park *et al.* assessed the predictive performance of UST-CDST in 130 patients with CD, and demonstrated it highly effective in predicting clinical remission at week 20 [217]. On the whole, exploration and analysis of predictors of ustekinumab do add value to personalized therapy, but available predictors need to be validated in independent and larger cohorts. More novel, accurate and feasible predictors are also required to be identified.

Indeed, biological agents become the mainstay in the treatment of IBD. They effectively help IBD patients in achieving disease remission and prevent patients from abdominal surgery and hospitalization. However, response rates are extremely different in individuals. To those primary non-responders, biologics not only expose patients to unnecessary risk of infection, allergy and even death, but also delay effective treatment and increase medical expense. Therefore, precise prediction of treatment response to biologics before giving treatment has been a pressing matter in the management of IBD. Additional new predictors with favorable sensitivity and specificity, and comprehensive panels or models of different-class predictors are required to guide the treatment.

Precision monitoring of key medications

Therapeutic drug monitoring (TDM) is the most important aspect in the field of precision monitoring. Once patients start treatment, rigorous monitoring of treatment response becomes an integral part in the management of IBD. Variations of pharmacodynamics, pharmacokinetics and pharmacogenetics between different patients lead to further study into the relationships between drug metabolites, serum drug concentrations, anti–drug antibodies, and clinical outcomes. Thiopurines and biological agents including infliximab, adalimumab, vedolizumab and ustekinumab are the most studied in the treatment of IBD.

Thiopurines

Thiopurines have extremely complicated metabolic pathways. As aforementioned, 6-TGNs are the therapeutic metabolites, and also responsible for myelosuppression [172]. Therefore, during thiopurine treatment, monitoring thiopurine metabolites become an essential part, which may assist in selecting appropriate therapeutic doses, achieving better therapeutic effectiveness, and reducing the possibility of adverse events.

Among these kinds of metabolites, measurements of 6-TGN and 6-MMP levels are applied in routine clinical practice. Several studies reported that the 6-TGN cut-off level of 230 pmol/8 \times 10⁸ red blood cells (RBCs) was associated with clinical remission [218]. Combined analysis of prior-treatment TPMT activity and posttreatment 6-TGN levels can further assist physicians in monitoring treatment response of thiopurines. Kwan and colleagues proposed that TPMT activity below 30.5 U combined with a 6-TGN level above 230 pmol/8 \times 10⁸ RBCs was significantly correlated with clinical response [219]. Another commonly used monitoring parameter is 6-MMP. Combined assessment of 6-TGN and 6-MMP further helps physicians distinguish clinical response, resistance and nonadherence, and thus guide dose and therapeutic program adjustment [220]. However, the monitoring of 6-TGN and 6-MMP levels shows an unfavorable sensitivity of 62% and a specificity of 72% for clinical response [221]. Due to different study designs, sample sizes and included groups, as well as different assays and instruments used to detect metabolite concentrations, various threshold values have been set in different studies. This caused some difficulties for physicians to make explanations for 6-TGN and 6-MMP values and monitor therapeutic effects.

As aforementioned, thiopurine metabolites are also in close association with adverse events secondary to thiopurine treatment. Thus, it is possible to minimize the risk of side effects of thiopurines by measuring 6-TGN and 6-MMP concentrations. Patients with 6-MMP levels above 5700 pmol/8 \times 10⁸ RBCs have an increased 3-fold risk of hepatotoxicity than those with lower 6-MMP levels, whereas 6-TGN steady-state levels above 490 pmol/8 \times 10⁸ RBCs are found to be significantly correlated with leukopenia [222]. The TOPIC study also revealed that not only increased 6-TGN concentrations (213 pmol/8 \times 10⁸ RBCs), but also elevated 6-MMP levels (3525 pmol/8 × 10⁸ RBCs) were in association with leukopenia with an OR of 6.2 and 5.9, respectively [173]. Given that patients with mutant genotypes of TPMT, NUDT15 and ITPA presented higher 6-TGN levels in comparison with wildtypes, the optimal cut-off value of 6-TGN should be considered to be reduced in those with mutant genotypes [223]. So,

measurements of 6-MMP and 6-TGN concentrations have been recommended as an effective strategy to maximize therapeutic efficacy and minimize adverse events. A target 6-TGN level between 230 and 450 pmol/8 \times 10⁸ RBCs is recommended by the American Gastroenterological Association Institute for IBD patients with thiopurine monotherapy [224]. Dose escalation or therapy switch should be considered when the 6-TGN concentration is below 230 pmol/8 \times 10⁸ RBCs, while dose reduction should be suggested once the 6-TGN concentration is above 450 pmol/8 \times 10⁸ RBCs. It is noteworthy that some patients with very high concentrations of 6-MMP and 6-TGN do not develop hepatotoxicity and leukopenia, while patients with relatively lower levels of 6-MMP and 6-TGN may still suffer from these adverse events. In this regard, thiopurine metabolite measurement cannot replace serial monitoring of liver enzymes and complete blood counts, but may provide useful supplemental information to therapeutic monitoring.

Infliximab

Infliximab is a highly effective treatment in both CD and UC patients, however about 20%-40% of patients become secondary non-responders over time [225]. The rationale for the lack or loss of response is complex. Multiple factors including molecular structures, pharmacodynamics, pharmacokinetics and pharmacogenetics result in different response rates. A good many nonresponders show inadequate serum drug concentrations, which are associated with increased clearance by either development of ADAs or mechanisms other than immunogenicity [220]. ADAs neutralize infliximab effects by binding to it and forming an immune complex, then cleared by the reticuloendothelial system. Smoking, a diagnosis of rheumatoid arthritis, high disease activity, treatment interval of more than 11 weeks, neutrophil CD64 ratio > 6 and starting infliximab dose < 7.5 mg/kg are claimed to be risk factors for ADAs [226, 227]. Many studies demonstrated that serum drug concentrations and ADAs are significantly correlated with clinical efficacy. The landmark study into the correlation was pioneered by Baert and colleagues [228]. They reported that a serum infliximab concentration of 12.0 μ g/ml or more at week four was associated with a longer duration of clinical response. In the same study, the concentration of ADAs (8.0 ug/ml) was claimed to be inversely correlated with the duration of response to infliximab. The following studies also confirmed the correlation in UC patients [229]. Besides clinical response, increased IFX trough levels are also associated with MH, improved radiologic outcomes and a better disease course, as well as reduced hospitalizations and surgeries [225, 230]. Therefore, monitoring of serum infliximab concentrations and ADAs during infliximab treatment is particularly important in the management of IBD.

In view of the close relationship between clinical efficacy and serum infliximab concentrations, TDM can be used to manage patients with a secondary loss of response to infliximab. Physicians can make therapy adjustments such as dose intensification, dose reduction, dose interval shortening, adding concomitant immunomodulator, or therapy switch (other anti-TNF agents or other-class biological agents) according to concentrations of infliximab and ADAs. In comparison with the empiric management of secondary non-responders, the TDM-tailored therapeutic algorithms show improved outcomes and cost-effectiveness [225]. The TAXIT study concluded that trough-level-based infliximab therapy outperforms system-based therapy in preventing flares during maintenance treatment. This study also indicated that TDMbased therapy can be proactively applied prior to loss of response [231]. Importantly, different disease phenotypes may show different optimal trough concentrations of infliximab. For example, trough levels of 10 ug/ml or more are recommended for patients with fistulizing phenotypes, while for patients with luminal CD, the recommended range is $3-7\mu g/ml$ [224]. From this point, target drug concentrations are not universal.

It should be noted that about 16%-39% of patients receiving scheduled infusion of infliximab have undetectable drug concentrations without the development of antibodies [232]. Antibodypositive subjects show similar rates of clinical remission and endoscopic improvement to antibody-negative patients, which limits its clinical utility in guiding physicians to optimize therapy outcomes [229, 232]. Moreover, studies also found that similar serum drug concentrations resulted in different effectiveness of infliximab between IBD patients, and a large number of non-responders had very high circulating drug trough levels [233]. These findings indicate that other inflammatory mediators other than TNF- α may be implicated in the ongoing inflammatory activity, and other contributing factors such as body weight, gender and unhealthy lifestyles may influence therapeutic effectiveness. Thus, monitoring drug concentrations and ADAs alone is not adequate enough for precisely monitoring therapeutic effects. Algorithms consisting of different contributing factors such as body weight, gender, disease activity, disease extent, albumin levels, CRP concentrations, etc. are needed to be explored.

Adalimumab

Adalimumab is another anti-TNF agent widely used in clinical practice. Lacking of or losing response to adalimumab is also relatively frequent in IBD patients. Undetectable concentrations of adalimumab and the development of ADAs partly account for the unfavorable response rates. Several studies demonstrated that IBD patients greatly benefit from higher adalimumab drug concentrations in clinical, endoscopic and histological remission [234]. In an American study, a cut-off value of 7.5 μ g/mL and 7.8 μ g/mL of adalimumab was best associated with endoscopic healing and histological remission, respectively [234]. Similarly, another exposure-response relationship study suggested that a cutoff value of 8.14 ug/ml correctly distinguished patients with MH from those without MH, with a sensitivity of 91.4% [235]. With respect to ADAs, the random adalimumab concentrations are notably lower in those with detectable ADAs. As a result, histological and endoscopic remission rates are lower [234]. These findings reflected that monitoring serum adalimumab concentrations and ADAs during the treatment is of great importance in disease management.

Given the vital roles of serum adalimumab concentrations and ADAs, TDM is of great help in guiding clinical decision making. For example, secondary non-responders with low adalimumab trough levels and lacking of ADAs conformation benefit most from adalimumab escalation. However, switching to other-class biologics should be considered in patients with low concentrations of adalimumab and detectable ADAs. However, one aspect should be taken into consideration is that no defined threshold has been established for guidance of therapeutic interventions. One pilot study of 78 children with CD investigated the association between proactive TDM and clinical remission. They set the treatment target of adalimumab level as 5 μ g/ml. As a result, the proportion of corticosteroid-free clinical remission in the proactive TDM group and the reactive TDM group was 82% and 48%, respectively. Although most of patients in the proactive TDM group achieved clinical remission, about 87% of subjects

underwent adalimumab escalation [236]. From this point, the optimal concentration target would be higher than 5 μ g/ml. Thus, some studies then recommend a target range of 7.2–12.0 μ g/ml [237, 238]. Moreover, currently available assay techniques used for the detection and quantification of serum drug levels and ADAs include the enzyme-linked immunosorbent assay (ELISA), fluid-phase radioimmunoassay, and homogeneous mobility shift assay (HMSA) [233]. The lack of a gold standard assay limited its routine clinical use. Different studies used different assay techniques and proposed different threshold values, which caused great difficul-

ties for disease management in daily clinical practice. Therefore, it is definitely a pressing need to establish a gold standard or optimal assay technique, and set up a universally acknowledged threshold of adalimumab to assist in optimizing dosing regimens, therefore maximizing the effectiveness and minimizing the adverse events of adalimumab in clinical practice.

Vedolizumab

Vedolizumab shows a unique function that it specifically suppresses gut inflammation without systemic immunosuppression [163]. Moreover, it also presents a better safety profile with minor infusion reaction and serious infection than anti-TNF agents, because it is a more specific antibody against gut-homing $\alpha 4\beta 7$ integrin [154]. Published data demonstrated that higher vedolizumab serum concentrations are associated with higher remission rates and better clinical response in both UC and CD patients [239]. The GEMINI trials demonstrated that the median trough concentrations at week 6 were higher in remitters than that in nonremitters [163]. In 2019, Osterman et al. proposed that a cut-off value of 37.1 μ g/ml at week 6 and 12.7 ug/ml at steady state was associated with clinical remission [240]. More recently, the target trough concentration of 32.0 μ g/ml at week 6 was claimed to be correlated with week 52 clinical remission [239]. However, other studies suggested a lower target concentration for endoscopic remission (10ug/ml) and MH (18ug/ml) [225, 241]. A cut-off value of 20.0 ug/ml at week 22 was suggested to be a predictor of achieving endoscopic remission in another study [242]. It is clear that higher trough concentrations are correlated with better outcomes. So, in view of this, monitoring serum drug levels may add value to dosing regimens in patients with insufficient response to vedolizumah

Several factors have been reported to have an effect on drug concentrations or clearance rates. A population pharmacokinetic analysis demonstrated similar clearance rates in CD and UC patients, while for patients with extremely lower albumin and higher body weight, the clearance rates would increase [243]. Given that only 3.7%-4.1% of patients develop transient ADAs and 0.4%-1.0% of patients have persistently positive ADAs in GEMINI trials, the relationship between ADAs and clinical efficacy is still uncertain [154, 205]. A randomized, double-blind, placebo-controlled study suggested similar clinical remission rates in patients with ADAs and those without ADAs (12% vs. 14%) [244]. While in the GEMINI trials, the development of ADAs was associated with a significant decrease in serum drug concentrations, and the latter was confirmed to be correlated with clinical effectiveness [154, 205]. This is in line with the results of a population pharmacokinetic analysis that vedolizumab linear clearance in those with positive ADAs was estimated to be 12% higher than that in patients with negative ADAs [243]. Therefore, more efforts are needed to elucidate the effects of ADAs on the clinical outcomes, and then optimize disease management.

With regard to these patients with concomitant immunosuppressive treatment, special attention should be paid. Available data suggested that concomitant immunomodulator is associated with decreased immunogenicity of vedolizumab, while it has no clinical effect on the pharmacokinetics of vedolizumab [163, 243]. This did not correspond to the finding in anti-TNF agents that concomitant immunosuppressant therapy is not only correlated with decreased immunogenicity, but also associated with increased clearance [228]. There might be additional modes of action of vedolizumab and some underlying factors accounting for the difference. Exploring these contributing factors and other mechanisms of action might further assist physicians in determining therapeutic strategies in patients with insufficient responses. It is important to note that the evidence for proactive/reactive TDM of vedolizumab is relatively limited. Whether TDM of vedolizumab is cost-effective also remains to be elucidated.

Ustekinumab

Ustekinumab is a new biological agent used for patients with moderate-to-severe CD and UC [23, 24]. Available data into the relationship between trough ustekinumab concentrations and treatment outcomes are relatively limited. In accordance with the above biologics, higher trough concentrations of ustekinumab indicate higher response rates. Ustekinumab target threshold of 3.7 ug/ml at week 8 was proved to be correlated with clinical response, while a trough concentration of 4.5 ug/ml at week 26 was claimed to be associated with endoscopic improvement and lower CRP levels, as well as trends toward FC normalization and endoscopic remission [245, 246]. The IM-UNITI trial proposed that the target trough concentration of ustekinumab at week 24 was 1 ug/mL, which was best associated with clinical remission [247]. More recently, a target ustekinumab concentration of 2.11 μ g/mL at week 16 was claimed to be correlated with fistula healing in CD [248]. Caution needs to be exercised when explaining these results, because different disease phenotypes, measurement time points, and desired outcomes of interest have been set in studies. As a result, different target trough concentrations of ustekinumab have been proposed.

Contrary to anti-TNF biologics, immunogenicity has less of an effect on the response rates to ustekinumab [213]. About 2.3% of patients were reported to develop ADAs during treatment in the IM-UNITI trial, while in the CERTIFI trial, only 0.7% of patients had positive results for ADAs at week 36 [23, 213]. Such a low prevalence of ADAs is not powerful enough to explain reasons for the tloss of response. Moreover, the positive effect of concomitant immunosuppressive therapy on the prevention of ADAs development seen in anti-TNF- α treatment may not be relevant to ustekinumab [249]. Therefore, further exploration of other factors influencing therapeutic response should be a research priority. Although dose optimization results in higher response rates in several other studies, whether patients with low trough ustekinumab concentrations will benefit from dose escalation is unclear. So, proactive/reactive TDM studies are needed to fill this gap. Moreover, definite thresholds and therapeutic drug concentration intervals are also required to be defined and validated in large, independent cohorts.

It is an indisputable fact that TDM plays a vital role in the monitoring and management of IBD. Based on pharmacodynamic, pharmacokinetic and pharmacogenetic properties of drugs, therapeutic targets will be achieved more easily and final outcomes of IBD patients will be improved. Indeed, measurements of drug metabolites, drug concentrations and ADAs significantly optimize IBD therapy, but it is still insufficient to achieve precise monitoring. Combined analysis of other clinical, serological, histologic and fecal factors along with TDM might further improve the precision of monitoring. Although TDM shows its great advantages in monitoring treatment response, deficiencies such as invasiveness, inconvenience and costliness make it unacceptable for some patients. Exploring markers directly or indirectly reflecting drug concentrations in saliva, sweat and feces, as well as noninvasive tests with acceptable price, sensitivity and specificity is in urgent demand. What's more, the time delay between sample collection and sample results should also be taken into consideration. Current studies mostly elucidate the influence of TDM on clinical and endoscopic outcomes. The relationship between TDM and optimal therapeutic targets such as MH, deep remission, and disease course change remains to be established in the following studies. Of note, various studies claimed an association between drug levels and disease remission. Whether this relationship is causal (high drug levels cause disease remission) or consequential (disease remission/decreased disease activity causes reduced drug clearance/high drug levels) remains to be fully clarified. Moreover, TDM for infliximab has been widely used in clinic, while TDM for new biologics such as adalimumab, vedolizumab and ustekinumab has been limited partly due to incomplete analytic techniques, undefined thresholds, and unclear pharmacokinetics. Therefore, more efforts should be put into the investigation of standard assay techniques, optimal thresholds, and exact metabolic mechanisms. With the unceasing efforts, TDM will play an increasingly key role in precision monitoring in patients with IBD

Future precision medicine in IBD

Medical therapy does play a critical role in the treatment of patients with IBD, and biological drugs such as infliximab, adalimumab, vedolizumab and ustekinumab targeting different signaling pathways have brought a revolutionary influence on the treatment of IBD. To achieve the goal of precision treatment, studies regarding new therapeutic agents, optimal therapeutic targets, different disease patterns, and patients' choices are in desperate need. With the increasing understanding of the pathogenesis of IBD, new pathophysiology has been found. Exploration of novel medicine targeting new targets with excellent therapeutic effects may further promote the development of precise treatment. Some new medicines targeting different targets such as JAK3, interleukins, chemokine receptor, cell adhesion molecule and protein kinase are developed. In recent years, IBD has been added to the expanding disease indications for some 'old' medicines that have already been applied in other immune-mediated diseases such as RA, SLE and psoriasis. This paved the way for the new use of old medicine. Therefore, exploration of the same signaling pathways implicated in IBD and other diseases may add value to the precise treatment of IBD. Combination therapy of immunosuppressants and biological agents obtained favorable therapeutic efficacy, which provides a possibility of application of various drugs targeting different pathways for IBD treatment.

Given that IBD is a progressive disease, patients with IBD present different pathophysiological characteristics in different disease stages. Therefore, physicians are advised to select different therapeutic targets in different disease stages during the entire disease course. Precision and individualized therapy will be the future medical model. Although numerous markers have been identified for precision treatment and precision monitoring in IBD, however, these available markers need external and prospective validation. Well-designed, large-scale, and well-paired phase II or phase III trials may provide more information about clinical translation. Moreover, clarifying that these identified markers merely reflect inflammation (correlation) or are part of the pathogenesis of IBD (causation) is also required. Compared studies on the above markers between unaffected siblings of IBD patients and those affected siblings will facilitate the identification of the exact roles of available markers in IBD. Besides, it is also important to determine the various roles of markers in different disease stages and the functional impacts on disease onset and progression. More importantly, intestinal damage of IBD is a progressive process, which impels doctors to carry out early and effective interventions before bowel damage. Thus, defining the terminology of the preclinical phase and exploring preclinical markers will be needed. Prospectively collecting preclinical samples and closely following up 'atrisk' family cohorts hold great promise to help precision prevention and change the natural disease course of IBD. Moreover, more importance should be attached to the environmental risk markers including prenatal and perinatal factors, drug exposure, diet and physical exercise, and imaging (such as magnetic resonance imaging and ultrasound) characteristics. Explaining the contributions of environmental and imaging risk markers in the preclinical stage might provide crucial insights into the disease pathogenesis and precision prediction of disease onset and development. What's more, considering different healthcare systems and financial structures around the world, more multidimensional prediction and monitoring tools integrating multi-omics data should be developed. Thus, an interdisciplinary collaboration between medical scientists, bioinformaticians, economists and manufacturers is encouraged. By achieving these endeavors, we are getting closer and closer to the goal of precision medicine in IBD.

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Author contributions

Writing—original draft preparation, Z.Z.; writing—review and editing, Z.Z. M. J. X.L. and J.Y.; supervision—H.Z. All authors have read and agreed to the published version of the manuscript.

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