

## Article

# Application of Silica Nanoparticles for Improving Growth, Yield, and Enzymatic Antioxidant for the Hybrid Rice EHR1 Growing under Water Regime Conditions

Omnia M. Elshayb <sup>1</sup>, Abdelwahed M Nada <sup>1</sup> , Heba M. Ibrahim <sup>2</sup>, Heba E. Amin <sup>3</sup> and Ayman M. Atta <sup>4,\*</sup> 

<sup>1</sup> Rice Research and Training Center, Field Crops Research Institute, Agricultural Research Center (ARC), Giza 588, Egypt; omniaelshayb3434@yahoo.com (O.M.E.); nadaabdelwahed456@gmail.com (A.M.N.)

<sup>2</sup> Botany Department, Faculty of Agriculture, Mansoura University, Mansoura 35516, Egypt; hebaho@mans.edu.eg

<sup>3</sup> Department of Food Industry, Faculty of Agriculture, Mansoura University, Mansoura 35516, Egypt; heba\_emad1127@hotmail.com

<sup>4</sup> Chemistry Department, College of Science, King Saud University, Riyadh 11451, Saudi Arabia

\* Correspondence: aatta@ksu.edu.sa

**Abstract:** The current study was designed to assess the effect of different concentrations of silica oxide nanoparticles (SiO<sub>2</sub>NPs) (0, 30, 60, and 90 ppm) as foliar applications under three irrigation regimes i.e., irrigation every 3 days (IR3, control), irrigation every 6 days (IR6), and irrigation every 9 days (IR9) on growth, yield and certain metabolites of rice (*Oryza sativa* L. cv. EHR1). To achieve such a goal, 2 field experiments were conducted during the 2018 and 2019 seasons at the Experimental Farm of Rice Research and Training Center (RRTC), Sakha Agricultural Station, Kafr El-sheik, Egypt. Firstly, the as-prepared nanoparticles of SiO<sub>2</sub> were prepared from useless materials (RHs) which are considered as one of the bio burdens on the environment via treating with HCl and followed by drying and calcination. Consequently, the synthesis was examined by making use of advanced tools such as X-ray diffraction (XRD), transmission electron microscopy (TEM), dynamic light scattering (DLS) for illustrating the hydrodynamic particle size of SiO<sub>2</sub>NPs and scanning electron microscopy (SEM). The nanoparticles were formed with nearly spherical shape and small size. The results indicated that leaf area index, dry matter production, the number of panicles/m<sup>2</sup>, the number of filled grains/ panicles, 1000 grain weight, grain yield, and biological yield as well as chlorophyll content have witnessed a significant increase under irrigated application every 3 and 6 days. Whilst a prolonged irrigation regime up to 9 days recorded a remarkable decline in the aforementioned characteristics except for the number of unfilled grains/panicle which increased considerably in both seasons. On the other hand, proline concentration and the activity of the antioxidant enzymes were increased in both irrigated treatments every 6 and 9 days compared with control treatment (irrigation every 3 days). The foliar supplementations of (SiO<sub>2</sub>NPs) contributed to ameliorating all the aforementioned characteristics progressively up to the dosage of 90 ppm compared to control treatment (no Si/NPS application) in both seasons. Invariably, growth and yield parameters in water-stressed plants treated with SiO<sub>2</sub>NPs were higher than those in water-stressed plants without SiO<sub>2</sub>NPs addition. Based on that, it could be concluded that the foliar application of SiO<sub>2</sub>NPs can mitigate the adverse effect of water stress on rice plants.

**Keywords:** agriculture domains; egyptian rice; physical characterization; silica nanoparticles



**Citation:** Elshayb, O.M.; Nada, A.M.; Ibrahim, H.M.; Amin, H.E.; Atta, A.M. Application of Silica Nanoparticles for Improving Growth, Yield, and Enzymatic Antioxidant for the Hybrid Rice EHR1 Growing under Water Regime Conditions. *Materials* **2021**, *14*, 1150. <https://doi.org/10.3390/ma14051150>

Academic Editor: Montserrat Colilla

Received: 29 December 2020

Accepted: 23 February 2021

Published: 28 February 2021

**Publisher's Note:** MDPI stays neutral with regard to jurisdictional claims in published maps and institutional affiliations.



**Copyright:** © 2021 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (<https://creativecommons.org/licenses/by/4.0/>).

## 1. Introduction

Rice (*Oryza Sativa* L.) is a crucial cereal crop that supplies around half of the Earth's people within 6% of the daily calorie intake [1]. It is worth noting that to meet global needs in 2035, rice productivity must be increased by 26% [2]. Various drawbacks restrict the yield of the rice crop and have a devastating impact on rice cultivation including

pest infestation, unfavorable environmental conditions accompanied by climate change, salinity stress, insufficient nutrients, and deficit irrigation (DI) as a result of an emerging shortage of water resources. The annual rice production shortfall due to DI approximated 18 million tons [2,3]. In comparison with other cereals, the role of combating and diluting the adverse effect of DI is much more complex in rice due to its higher sensitivity to DI [4]. The adverse impact of DI on rice was manifested by reducing root volume, plant height, leaf dimension, leaf greenness, tillering, panicle exertion, flowering, and fertility, all of which have a negative impact reflected in yield loss [5,6]. Moreover, DI resulted in chlorophyll breakdown and enhanced generation of reactive oxygen species (ROS), which attack cell biological components, thereby affecting adversely and in a deadly way the integrity and functions of plant cells [7]. Correspondingly, the influence of water-conserving water irrigation regimes mostly on the growth and yield of *Oryza sativa* L. cv. EHR1 should be examined. Silicon is a major contributor to the Earth's crust after oxygen. It is not considered a vital nutrient for most terrestrial plants, but it plays an important role in improving the quality, quantity, and protection of some plants, especially rice and wheat. It also has the ability for providing resistance towards harsh conditions in plants [8]. The reduction of silicon-induced stress as mentioned in previous studies is attributed to structural as well as biochemical effects, such as enhancing photosynthetic parameters [9], improving activities of antioxidant enzymes as well as increasing soluble protein content, and preserved water relations of the plant cells [10].

The utilization of nanomaterials has produced several notable results in improving growth, yield production, and alleviate the influence of different stresses on plants. Among elegant and unique nanomaterials, silica nanoparticles ( $\text{SiO}_2\text{NPs}$ ) which have benign efficacy in remediating the negative effect of heavy metals [11,12], salinity [13], ultraviolet (UV-B) radiation [14], pathogens, and insects [15–17] was reported. Besides, the drought stress-mitigation effects of  $\text{SiO}_2\text{NPs}$  on cereals plants are well-documented [18–20]. Nevertheless, its drought stress mitigation effectiveness on a semi-hydrophyte like rice is less investigated and poorly understood. Various effects were reported as the basis of  $\text{SiO}_2\text{NP}$ -induced alleviation of drought stress. Nanoparticles of  $\text{SiO}_2$  at 400 mg/L, 2000 mg/L, and 4000 mg/L increased the content of photosynthetic pigments (chlorophyll a, b and carotenoids) in *Z. mays* compared with control [21]. Furthermore, the application of  $\text{SiO}_2\text{NPs}$  caused a significant increase in the content of soluble sugar and the activities of catalase (CAT) and peroxidase (POD) in faba bean leaves [22]. Whether growth, yield, and stress-related metabolites in a high water-demanding crop like rice will be similarly affected by  $\text{SiO}_2\text{NPs}$  fortification needs to be ascertained. This investigation was conducted to assess the effect of silica nanoparticles ( $\text{SiO}_2\text{NPs}$ ) at different concentrations (30 ppm, 60 ppm, and 90 ppm) on growth, yield, and some key stress-related metabolites of the Egyptian hybrid rice variety EHR1 plants subjected to moderate and severe water stress. The work was aimed to prepare the valuable  $\text{SiO}_2\text{NPs}$  from useless wastes aiding to remove one of the issues that cause problems for the environment. Moreover, the prepared nanoparticles can be scaled up on a large scale without any noticeable cost.

## 2. Experimental Details

### 2.1. Synthesis of Silica Nanoparticles ( $\text{SiO}_2\text{NPs}$ ) from Useless Materials (RH Silica)

Silica nanoparticles were prepared after useless materials (RHs) were treated with HCl, then calcination. In brief, running tap water was used to wash RHs, then washing with deionized water was important to remove any dirt or other contaminants stacked on RHs. The RHs were dried at 90 °C along the night. After drying, the portion of RHs was refluxed with 5 wt.% HCl solution for 2 h, then washed with deionized water and dried at 90 °C for at least 12 h. The produced  $\text{SiO}_2\text{NP}$  with a diameter of ca. 60–70 nm was obtained by calcining HCl-treated RHs in a furnace at 700 °C for 2 h. The yield of  $\text{SiO}_2\text{NPs}$  was about 17 wt.%. Then the as-synthesized  $\text{SiO}_2\text{NP}$  was characterized as follows: The  $\text{SiO}_2\text{NPs}$  sample was deposited on a carbon-coated copper grid and left for drying at room temperature to be examined with a transmission electron microscope

(TEM, JEOL 200 kV, Jeol, Tokyo, Japan). Selected area diffraction (SAED) was utilized to investigate the nature of the prepared nanoparticles in terms of their amorphous state. For the evaluation of the particle size and size distribution of the as-prepared SiO<sub>2</sub>NPs, dynamic light scattering (DLS, Nano-Sizer SZ90, Malvern instruments Ltd., Malvern, Worcestershire, UK) was used at pH = 7 and 25 °C. Scanning electron microscopy was scanned using a field emission scanning electron microscope (FESEM, ZEISS GeminiSEM 360; Mladá Boleslav, Czech Republic). X-ray diffraction pattern (XRD-X'Pert Pro, PANalytical, Almelo, The Netherlands) was used to investigate the crystalline lattice structure.

## 2.2. Cultivation and Experimental Conditions

Two field experiments were conducted during the two cropping seasons of 2018 and 2019 at the Experimental Farm of Rice Research and Training Center (RRTC), Sakha, Kafr-Elsheikh, Egypt. The previous cultivated crop was wheat during both seasons of the study. Egyptian hybrid rice (EHR1) variety was used in this study. Pregerminated seeds (healthy hybrid rice grains at the rate of 24 kg/ha were soaked more than water for 24 h and further incubated for another 48 h to enhance germination) were broadcasted in the nursery on 11 and 14 of May in the 2018 and 2019 seasons, respectively. Three seedlings per hill at 25 days old were transplanted in the permanent field's experimental plots at a 20 cm × 20 cm distance between hills and rows in 15 m<sup>2</sup> (5 m × 3 m) size plots. Weeds were controlled chemically using Saturn 50% at the rate of 5 L/ha at five days after transplant. Nitrogen in the form of urea (46% N) at the rate of 165 kg/ha was applied as recommended in two doses; 2/3 basal application + 1/3 at panicle initiation. The recommended phosphorous and Potassium fertilizers in the form of calcium superphosphate (15% P<sub>2</sub>O<sub>5</sub>) at a rate of 37 kg P<sub>2</sub>O<sub>5</sub>/ha and potassium sulfate (48% K<sub>2</sub>O) at the rate of 50 kg K<sub>2</sub>O kg/ha were applied. Zinc fertilizer, applied at the rate of 24 kg/ha ZnSO<sub>4</sub>, was mixed with sand, and manually broadcasted before transplanting. During both seasons, representative soil samples were taken from the experimental site at the depth of 0–30 cm. The physical and chemical characteristics of the experimental soil were analyzed according to [23] and presented in Table 1.

**Table 1.** Physical and chemical analysis of the experimental soil during the 2018 and 2019 seasons.

Seasons	Texture	Sand (%)	Silt (%)	Clay (%)	pH (1:2.5 Soil Extract)	E.C. (dSm <sup>-1</sup> )	Organic Matter %	Available N (ppm)	Available P (ppm)	Available K (ppm)	Available Zn (ppm)	Available Mn (ppm)	Available Fe (ppm)
2018	Clayey	12.8	31.5	55.7	8.33	3.12	1.45	18.4	14.7	322	0.78	3.44	3.12
2019	Clayey	12.0	32.0	56.0	8.40	3.48	1.50	19.1	15.2	347	0.95	3.22	3.70

The experiment was laid out in a split-plot design with four replicates. The main plots were devoted to three water regimes; irrigation every 3, 6, and 9 days which denoted as IR3, IR6, and IR9 respectively. The subplots were assigned to four treatments of SiO<sub>2</sub>NPs namely, SiO<sub>2</sub>NPs 0, control; SiO<sub>2</sub>NPs 30, foliar application of SiO<sub>2</sub>NPs at the rate of 30 ppm; SiO<sub>2</sub>NPs 60, foliar application of SiO<sub>2</sub>NPs at the rate of 60 ppm and SiO<sub>2</sub>NPs-90, foliar application of SiO<sub>2</sub>NPs at the rate of 90 ppm. In the SiO<sub>2</sub>NPs 0 treatment, the foliar spray was done with distilled water. Treatments of SiO<sub>2</sub>NPs had been applied thrice, at mid-tillering, panicle initiation, and full heading. Main plots containing irrigation treatments were tightly separated by ditches, 2 m wide and 1 m depth. The irrigation regime treatments, applied as mentioned above, were commenced after 10 days from transplanting.

## 2.3. Studied Traits

### 2.3.1. Plant Growth Characteristics

At the heading stage, plants of five hills were randomly taken from each plot to estimate leaf area index (LAI) and dry matter production. Leaf area index is the ratio between the leaves areas (cm<sup>2</sup>) of the plant divided by the ground area occupied by the plant. Dry matter production (g/m<sup>2</sup>) was estimated as described by [24].

At the harvest stage, panicles of five random hills from each plot were counted then converted to the number of panicles/m<sup>2</sup>. Ten panicles were randomly collected from each plot to determine the number of filled grains/panicles, and unfilled grains/panicle and 1000 grain weight (g). The biological yield (both grain and strawweight) was measured from an area of 12 m<sup>2</sup> (3 m × 4 m) which was harvested from each plot at random avoiding the border effects. Grain yield was adjusted to 14% moisture content as described by [25].

### 2.3.2. Determination of Chlorophyll and Proline Content

Chlorophyll content: total chlorophyll content was determined as SPAD (Soil Plant Analysis Development) value in ten flag leaves using chlorophyll meter (Model-SPAD 502) Minolta, Japan.

Determination of proline: proline concentration was determined according to the method of [26]. About 300 mg of leaf tissue was homogenized in 10 mL of 3% (*w/v*) aqueous sulfosalicylic acid and filtered. To 2 mL of the filtrate was added 2 mL of ninhydrin acid, then 2 mL of glacial acetic acid was added, and the mixture was boiled for 60 min. The mixture was extracted with toluene and the free proline was quantified by spectrophotometry at 520 nm.

### 2.3.3. Determination of the Activity of the Antioxidant Enzymes

Leaf tissues were homogenized (1:5 *w/v*) in an ice mortar using 50 mM sodium phosphate buffer, pH 7.0, containing 1 M NaCl, 1% polyvinyl pyrrolidone and 1 mM ethylenediaminetriacetic acid (EDTA). After centrifugation (20,000× *g*, 15 min), the supernatant (crude leaf extract) was used to determine enzyme activity; and hereinafter referred to as the enzymatic extract (EE).

CAT activity (EC 1.11.1.6) was evaluated by the decomposition of H<sub>2</sub>O<sub>2</sub> according to [27]. At 200 μL, 1 aliquot of EE was added to 1.8 mL of the reaction mixture (RM) containing 50 mM K-P buffer (pH 7.0) and 30 mM H<sub>2</sub>O<sub>2</sub>. The decrease in H<sub>2</sub>O<sub>2</sub> was accompanied by a decrease in absorbance at 240 nm. One unit of CAT activity is defined as the amount of enzyme that breaks down 1 M H<sub>2</sub>O<sub>2</sub> in one minute.

The POD activity (EC 1.11.1.7) was analyzed according to the method [28] by adding 25 μL of EE to 2 mL of a solution containing 50 mM of K-P buffer (pH 6.8), 20 mM of guaiacol, and 20 mM H<sub>2</sub>O<sub>2</sub>. After 10 min of incubation, the reaction was stopped by adding 0.5 mL of 5% (*v/v*) H<sub>2</sub>SO<sub>4</sub>, and the optical density was measured at 480 nm. One unit of POD activity is defined as the amount of substrate transformed by the enzyme in 1 min.

The SOD activity (EC 1.15.1.1) was analyzed according to the method of Van [29]. An aliquot of 50 μL of the enzyme extract was mixed with a solution containing L-methionine (13 mM), nitroblue tetrazolium chloride (75 μM), EDTA (100 μM) and riboflavin (2 μM) in a 50 mM potassium-phosphate-buffer (pH 7.8). The assay was performed in a chamber illuminated with 30 W fluorescent lamps. The reaction was started by turning the lamp on and terminated 5 min later by turning it off. The substrate formed as a result of NBT's (nitro blue tetrazolium) photoreduction (the blue formazan) was estimated as the increase in absorption at 560 nm. The control reaction mixture had no enzyme extract. The blank solution contained the same constituents included in the complete reaction mixture but was kept in the dark. The amount of enzyme required to inhibit 50% of the NBT photoreduction in comparison with tubes lacking the enzyme extract was considered as one unit of SOD activity.

## 2.4. Statistical Analysis

All statistical analyses used analysis of variance technique employing the "COSTATC" computer software package. Treatment means were compared by Duncan's Multiple Range Test [30].

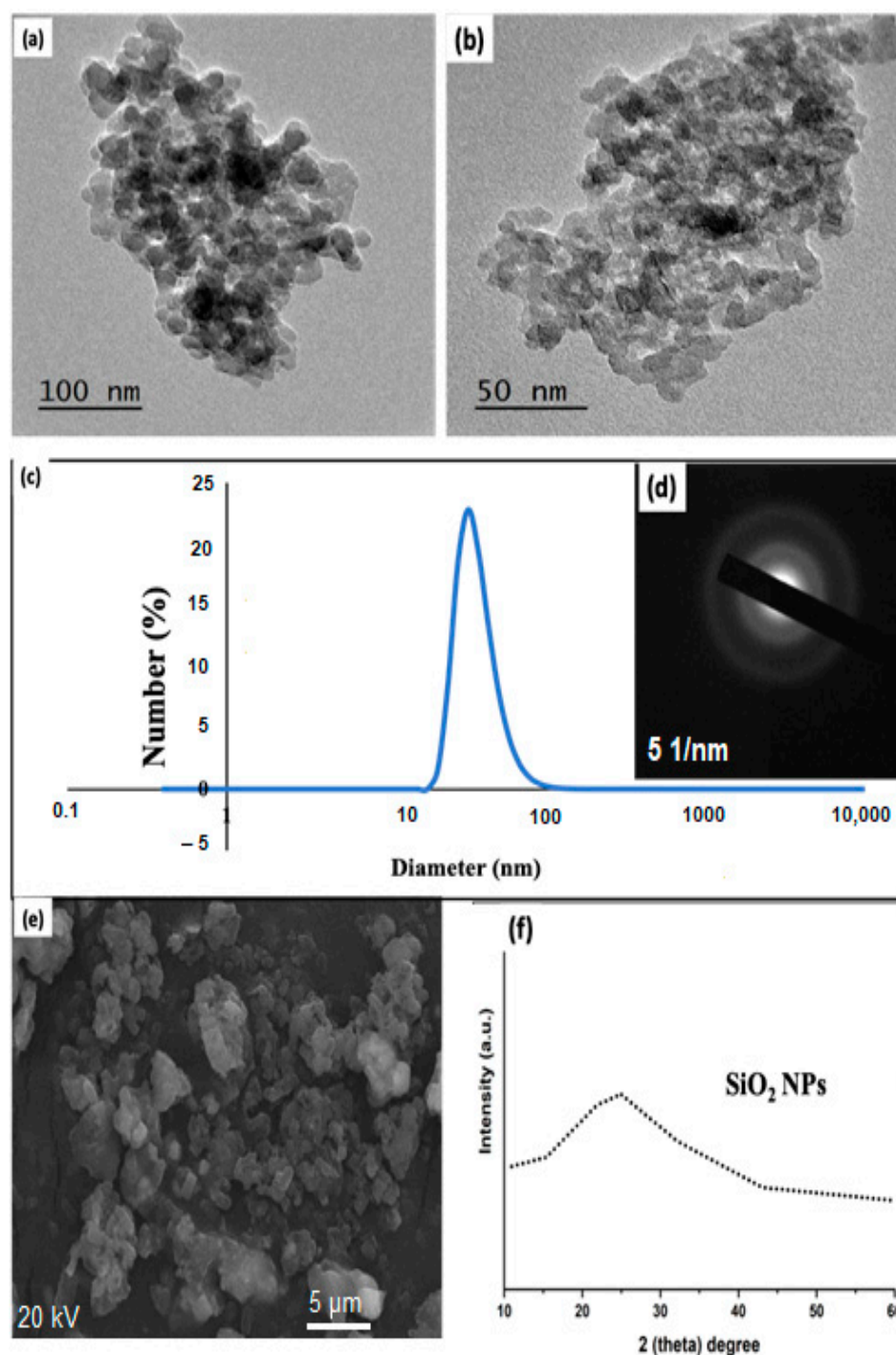
### 3. Results and Discussion

The first target of our current research work was designed to achieve two promising goals. The first one is to prepare SiO<sub>2</sub>NP from useless materials and that caused a hazardous effect on the environment. This target was initially characterized using TEM, SEM, DLS for determining the hydrodynamic particle size, and XRD. As mentioned in the experiment part for the preparation of SiO<sub>2</sub>NPs, the process for the preparation is very cost-effective and no need for extra chemical or advanced instruments for preparation, just washing, drying, and calcination, which, in turn, is a promising method to be called up to obtain large production of SiO<sub>2</sub>NPs suitable for many huge industrial and agricultural domains. Then, the characterized SiO<sub>2</sub>NP was used as a foliar treatment at different concentrations (30, 60 and 90 ppm) as will be discussed in the second part of our current research work. Below are the characteristics data that confirm the preparation of SiO<sub>2</sub>NPs.

#### *3.1. Utilized Advanced Tools in Terms of Transmission Electron Microscopy (TEM), Dynamic Light Scattering (DLS), Selected Area Diffraction (SAED), Scanning Electron Microscopy (SEM), and X-ray Diffraction (XRD) for Affirming the Preparation of Silica Oxide Nanoparticles (SiO<sub>2</sub>NPs)*

First of all, to describe the shape and prediction of the particle size of the resultant SiO<sub>2</sub>NPs, TEM was used. SiO<sub>2</sub>NPs (Figure 1a) has appeared clearly spherical and nearly as uniform particle size. It is noted that the spherical particle's average diameter is about 30 nm. For clarification of the particle shape and size distribution of SiO<sub>2</sub>NPs, the image (Figure 1b) is taken at high magnification. It can be assumed that SiO<sub>2</sub>NPs have a structure with a spherical shape. The insignificant agglomeration for these very small particles could be attributed to the absence of a stabilizing agent and the high surface area which facilitates the tendency of particles to be agglomerated in cluster form. However, these cluster agglomerated particles are still in small size and do not exceed 30 nm in all conditions of agglomerations.

Next, hydrodynamic average particle size is very important to be measured using DLS to stand on the actual size of the prepared SiO<sub>2</sub>NPs. The obtained particle size from DLS is set in Figure 1c. Note that the average particle size is 43 nm. As observed in Figure 1c, the image contains one sharp peak which confirmed the particles are formed in the monodisperse form with a polydispersity index equal to 0.107. The difference in the formed diameter between and TEM and DLS could be related to the swelling properties of SiO<sub>2</sub>NPs in a solution. As known, DLS needs during the measurement that the particles are dispersed in water and the analysis was carried for more than 18 times. All these runs make these particles swell and form a large particle. Nonetheless, the particle evaluated by both TEM and DLS is still below 50 nm which confirmed the small size and the large surface area (active materials) of these nanoparticles that are expected to have a very important role and efficiency in the agriculture field as will be presented in the second target of our current research work. Returning to our discussion and for further confirmation, the SAED pattern was conducted (Figure 1d). As clearly remarkable that, the particles are formed in the amorphous state with no spots or light points which affirms that the calcination step is very critical in our work to convert the useless compound into nanoparticles of SiO<sub>2</sub> in nanoparticle form. The surface structure and morphology of SiO<sub>2</sub>NPs was further investigated using SEM. Figure 1e displays the morphological structure of SiO<sub>2</sub>NPs. SiO<sub>2</sub>NPs as fairly uniform spherical particles but the particles are remarkably formed with sufficient aggregation which could be attributed to the absence of the extra stabilizing agent. Therefore, spherical SiO<sub>2</sub>NPs are formed due to isotropic growth. Briefly speaking, the anisotropic structures of SiO<sub>2</sub>NPs could be induced by growth rates in different directions and as such, due to isotropic growth, nearly spherical SiO<sub>2</sub>NPs are formed. In brief, the calcination step (high temperatures) has resulted in the development of this shape (nearly spherical particles) which has led to extreme anisotropic growth, leading to the formation of aggregated particles due to the collision between a particle that causes the production of large particles at high temperature. Ultimately, the data of XRD (Figure 1f) agree with the data of SAED in which SiO<sub>2</sub>NPs in the amorphous state have no peaks for the crystallinity.



**Figure 1.** Transmission electron microscopy (TEM) of  $\text{SiO}_2$ NPs at different magnification (a,b), (c) average hydrodynamic size, (d) selected area diffraction (SAED), (e) scanning electron microscopy (SEM) and (f) X-ray diffraction (XRD) of silica oxide nanoparticles ( $\text{SiO}_2$ NPs).

### 3.2. Application of $\text{SiO}_2$ NPs as a Foliar for Rice Plants

#### 3.2.1. Plant Growth Characteristics

Compared with IR3 (normal irrigation, no stress) leaf area index (LAI) and dry matter (DM) accumulation in plants irrigated every 9 days (IR9 plants) are decreased whereas those parameters, in plants irrigated every 6 days (IR6 plants), are not significantly affected in both seasons (Table 2). On the other hand,  $\text{SiO}_2$ NPs with different concentrations increased both LAI and DM accumulation, in a concentration-dependent manner, i.e., both parameters are increased with increasing  $\text{SiO}_2$ NPs concentration. Foliar spray of

SiO<sub>2</sub>NPs at 90 ppm increased the leaf area index by 8.1% and 9.7%, and dry matter production by 6.0% and 6.5% in both the first and second seasons, respectively. The effects of both IRs and SiO<sub>2</sub>NPs were consistent during the two experimental seasons. The interaction effect between IRs and SiO<sub>2</sub>NPs levels on LAI was significant only during the first experimental season whereas the corresponding effect on DM was significant only during the second experimental season. During the experimental season of 2018, the highest LAI in water-stressed plants was recorded in plants that were irrigated every 6 days and treated with SiO<sub>2</sub>NPs at 90 ppm (IR6-SiO<sub>2</sub>NPs-90 plants). Moreover, in IR6 plants, there was no significant difference between LAI in SiO<sub>2</sub>NPs-0 plants and either SiO<sub>2</sub>NPs-30 or SiO<sub>2</sub>NPs-60 plants. Noticeably, SiO<sub>2</sub>NPs treatments did not affect LAI in IR9 plants (Table 3). Considering the impact of the interaction between the two experimental factors on DM during the experimental season of 2019 (Table 4), it is obvious that there was a dramatic increase in DM only attributable to the treatment of SiO<sub>2</sub>NPs at 90 ppm in IR3 plants, owing to both treatments of SiO<sub>2</sub>NPs at 60 ppm and 90 ppm in IR6 plants, and according to all concentrations of SiO<sub>2</sub>NPs in IR9 plants, with the highest adopted SiO<sub>2</sub>NPs level (90 ppm) being the most effective treatment in this regard.

**Table 2.** Growth characteristics of EHR1 rice variety as affected by water regimes and SiO<sub>2</sub>NPs concentrations during the 2018 and 2019 seasons.

Treatments	Leaf Area Index (LAI)		Dry Matter Production (g/m <sup>2</sup> )	
	2018	2019	2018	2019
Water regimes	–	–	–	–
IR3	7.07 a	6.94 a	1575.1 a	1562.2 a
IR6	6.98 a	6.87 a	1566.0 a	1551.7 a
IR9	5.44 b	5.29 b	1231.2 b	1224.1 b
F test	**	**	**	**
SiO <sub>2</sub> NPs concentrations	–	–	–	–
SiO <sub>2</sub> NPs-0	6.26 d	6.06 d	1415.6 d	1399.5 d
SiO <sub>2</sub> NPs-30	6.41 c	6.28 c	1440.3 c	1428.5 c
SiO <sub>2</sub> NPs-60	6.56 b	6.49 b	1462.2 b	1465.7 b
SiO <sub>2</sub> NPs-90	6.77 a	6.65 a	1501.6 a	1491.6 a
F test	**	**	**	**
IRs X SiO <sub>2</sub> NPs	**	N.S	N.S	**

\*\* and N.S indicate significant at 0.01 level and not significant respectively. IR3, irrigation every 3 days; IR6, irrigation every 6 days; IR9: irrigation every 9 days. SiO<sub>2</sub>NPs-0, without SiO<sub>2</sub>NPs foliar application; SiO<sub>2</sub>NPs-30, foliar application of SiO<sub>2</sub>NPs at 30 ppm; SiO<sub>2</sub>NPs-60, foliar application of SiO<sub>2</sub>NPs at 60 ppm; SiO<sub>2</sub>NPs-90, foliar application of SiO<sub>2</sub>NPs at 90 ppm.

**Table 3.** Influence of the interaction between irrigation treatments (IRs) and SiO<sub>2</sub>NPs concentrations on the leaf area index (LAI) of EHR1 rice variety during the 2018 season.

SiO <sub>2</sub> NPs Concentrations	Water Regimes		
	2018		
	IR3	IR6	IR9
SiO <sub>2</sub> NPs-0	6.71 c	6.75 c	5.32 d
SiO <sub>2</sub> NPs-30	6.96 bc	6.77 bc	5.49 d
SiO <sub>2</sub> NPs-60	7.21 ab	7.06 abc	5.38 d
SiO <sub>2</sub> NPs-90	7.38 a	7.34 a	5.58 d

IR3, irrigation every 3 days; IR6, irrigation every 6 days; IR9: irrigation every 9 days. SiO<sub>2</sub>NPs-0, without SiO<sub>2</sub>NPs foliar application; SiO<sub>2</sub>NPs-30, foliar application of SiO<sub>2</sub>NPs at 30 ppm; SiO<sub>2</sub>NPs-60, foliar application of SiO<sub>2</sub>NPs at 60 ppm; SiO<sub>2</sub>NPs-90, foliar application of SiO<sub>2</sub>NPs at 90 ppm.

**Table 4.** Influence of the interaction between IRs and SiO<sub>2</sub>NPs concentrations on dry matter production of EHR1 rice variety during the 2019 season.

SiO <sub>2</sub> NPs Concentrations	Water Regimes		
	2019		
	IR3	IR6	IR9
SiO <sub>2</sub> NPs-0	1536.2 bc	1518.0 d	1192.3 g
SiO <sub>2</sub> NPs-30	1570.0 b	1546.4 cd	1235.0 f
SiO <sub>2</sub> NPs-60	1580.1 b	1579.2 b	1231.2 f
SiO <sub>2</sub> NPs-90	1623.2 a	1620.4 a	1266.6 e

IR3, irrigation every 3 days; IR6, irrigation every 6 days; IR9: irrigation every 9 days. SiO<sub>2</sub>NPs-0, without SiO<sub>2</sub>NPs foliar application; SiO<sub>2</sub>NPs-30, foliar application of SiO<sub>2</sub>NPs at 30 ppm; SiO<sub>2</sub>NPs-60, foliar application of SiO<sub>2</sub>NPs at 60 ppm; SiO<sub>2</sub>NPs-90, foliar application of SiO<sub>2</sub>NPs at 90 ppm.

Water stress-induced growth inhibition recorded in the current research work is in line with the findings of [31]. Water shortage is linked with nutrients malabsorption especially K<sup>+</sup> which has a tremendous role in the water relations of plant cells [32]. Subsequently, losses in leaf chlorophyll, defects in both cell division and enlargement have been accrued thereby hindering dry matter production. Moreover, the generation and accumulation of abscisic acid (ABA) in the plant cell resulted in stomatal closure, decrement of both gas exchange, and CO<sub>2</sub> assimilation which hurts the cell physiological pathway [19]. Contrariwise, treatment with SiO<sub>2</sub>NPs rendered a positive response in the estimated growth-indicative parameters, with the superiority of the higher adopted level (90 ppm). In conformity with the results of the present study, [33] demonstrated that Si enhanced both root and shoot biomass of rice under either normal or stressed conditions. Similar results are reported in wheat [18] and barley [20]. In this context, [9] described the benefits of silicon to rice plant tissues. He explained that Si deposits in the cell wall of rice leaves and stems, resulting in erected leaves especially under mutual shading. Accordingly, rice plants probably take higher advantage of light interception, thereby activating photosynthetic rate, enhancing chlorophyll content, leaf area index, and dry matter production. As indicated by [34], Si-induced cytokinin synthesis and may be reflected in increased chlorophyll content and an enhanced photosynthesis process.

### 3.2.2. Yield and Yield Attributes

Data in Tables 5 and 6 reveal significant and different responses on rice yield attributes (the number of panicles/m<sup>2</sup>, the number of filled grains/panicles, and unfilled grains/panicle and 1000 grain weight) as well as on both grain and biological yield under various treatments. The imposition of DI reduced all recorded yield parameters of rice whereas the number of unfilled grains is significantly increased, proportionately with increasing the stress magnitude. Nevertheless, the decrease recorded in all the estimated yield components in IR6 plants is not significant. These results are consistent during both experimental seasons. In IR9 plants (irrigated every 9 days), the number of panicles/m<sup>2</sup> is decreased by 27.7% and 28.4%; the number of filled grains/panicles is decreased by 22.9 and 24.4%; 1000 grain weight is decreased by 5.9% and 6.2 %; grain yield is decreased by 27.3% and 27.4% and biological yield is decreased by 20.3% and 20.7% in the first and second experimental season, respectively, relative to the values in IR3 plants. The aforementioned data are supported by those reported by [31,33]. The reduction in the number of panicles/m<sup>2</sup> and the number of filled grains/panicle in IR9 plants may be due to decreased chlorophyll content and reduced leaf area, and as a consequence disrupted leaf carbohydrate metabolism, thereby reduced assimilates transported to the sink organ (grains) and increasing reproductive abortion [31]. In this regard, it was stated that DI caused a marked decline in nutrient uptake and, consequently, the plant became under nourished, which probably decreased reproductive tillers, number of filled grains/panicle, and grain weight and grain yield outcome [32].



**Table 5.** The number of panicles/m<sup>2</sup>, filled grains/panicle, and unfilled grains/panicle of EHR1 rice variety as affected by water regimes and SiO<sub>2</sub>NPs concentrations.

Treatments	The Number of Panicles/m <sup>2</sup>		The Number of Filled Grains/Panicle		The Number of Unfilled Grains/Panicles	
	2018	2019	2018	2019	2018	2019
Water regimes	–	–	–	–	–	–
IR3	616.7 a	613.4 a	154.1 a	150.3 a	6.3 c	7.1 c
IR6	612.9 a	609.2 a	151.7 a	146.5 a	7.4 b	8.4 b
IR9	447.5 b	438.1 b	118.8 b	113.6 b	15.4 a	17.1 a
F test	**	**	**	**	**	**
SiO <sub>2</sub> NPs concentrations	–	–	–	–	–	–
SiO <sub>2</sub> NPs-0	538.0 d	530.4 d	137.3 c	131.7 c	10.9 a	12.7 a
SiO <sub>2</sub> NPs-30	554.7 c	548.4 c	139.3 b	135.8 b	10.2 b	11.4 b
SiO <sub>2</sub> NPs-60	565.3 b	560.3 b	144.1 a	138.0 ab	9.1 c	10.3 c
SiO <sub>2</sub> NPs-90	578.1 a	575.2 a	145.4 a	140.5 a	8.4 d	9.8 c
F test	**	**	**	**	**	**
IRs X SiO <sub>2</sub> NPs	**	**	N.S	N.S	N.S	N.S

\*\* and N.S indicate significant at 0.01 level and not significant respectively. IR3, irrigation every 3 days; IR6, irrigation every 6 days; IR9: irrigation every 9 days. SiO<sub>2</sub>NPs-0, without SiO<sub>2</sub>NPs foliar application; SiO<sub>2</sub>NPs-30, foliar application of SiO<sub>2</sub>NPs at 30 ppm; SiO<sub>2</sub>NPs-60, foliar application of SiO<sub>2</sub>NPs at 60 ppm; SiO<sub>2</sub>NPs-90, foliar application of SiO<sub>2</sub>NPs at 90 ppm.

**Table 6.** The 1000 grain weight (g), grain yield (t/ha), and biological yield (t/ha) of EHR1 rice variety as affected by water regimes and SiO<sub>2</sub>NPs concentrations during the 2018 and 2019 seasons.

Treatments	1000 Grain Weight (g)		Grain Yield (t/ha)		Biological Yield (t/ha)	
	2018	2019	2018	2019	2018	2019
Water regimes	–	–	–	–	–	–
IR3	26.17 a	26.11 a	12.01 a	11.83 a	27.42 a	27.01 a
IR6	26.10 a	26.06 a	11.89 a	11.74 a	27.18 a	26.85 a
IR9	24.62 b	24.48 b	8.62 b	8.58 b	21.84 b	21.40 b
F test	**	**	**	**	**	**
SiO <sub>2</sub> NPs concentrations	–	–	–	–	–	–
SiO <sub>2</sub> NPs-0	25.34 d	25.27 d	10.41 d	10.23 d	24.30 d	23.90 d
SiO <sub>2</sub> NPs-30	25.54 c	25.37 c	10.75 c	10.56 c	25.13 c	24.67 c
SiO <sub>2</sub> NPs-60	25.68 b	25.62 b	10.92 b	10.93 b	25.86 b	25.48 b
SiO <sub>2</sub> NPs-90	25.95 a	25.93 a	11.29 a	11.15 a	26.62 a	26.30 a
F test	**	**	**	**	**	**
IRs X SiO <sub>2</sub> NPs	**	**	**	**	**	**

\*\* and N.S indicate significant at 0.01 level and not significant respectively. IR3, irrigation every 3 days; IR6, irrigation every 6 days; IR9: irrigation every 9 days. SiO<sub>2</sub>NPs-0, without SiO<sub>2</sub>NPs foliar application; SiO<sub>2</sub>NPs-30, foliar application of SiO<sub>2</sub>NPs at 30 ppm; SiO<sub>2</sub>NPs-60, foliar application of SiO<sub>2</sub>NPs at 60 ppm; SiO<sub>2</sub>NPs-90, foliar application of SiO<sub>2</sub>NPs at 90 ppm.

Concerning SiO<sub>2</sub>NPs effects, both yield attributes and grain yield increased progressively until the highest adopted level, i.e., to the treatment of SiO<sub>2</sub>NPs-90 (Tables 5 and 6). Relative to control treatment (SiO<sub>2</sub>NPs-0), treatment with SiO<sub>2</sub>NPs at 90 ppm increased the number of panicles/m<sup>2</sup> by 7.4% and 8.4%, the number of filled grains/panicle by 5.8 and 6.6%, 1000-grain weight by 2.4% and 2.6%, grain yield by 8.4% and 9.0% and biological yield by 9.5% and 10.0% whereas unfilled grains/panicle decreased by 22.9% and 22.8% in the first and second experimental season, respectively. Si-induced water stress mitigation was explained by indicating that Si involves cell signaling because it has an immense function in binding the hydroxyl group of proteins [34,35]. Under DI conditions, silicic acid (the form in which the plant uptake Si) is polymerized and converted to silica gel. Silica gel concentrated on the surface of shoot parts as a double layer causing a remarkable decrease in water loss by leaf transpiration [36,37].

Induction of rice fertility, which is reflected in enhancing yield, in response to SiO<sub>2</sub>NPs, has been previously reported [18]. Furthermore, [38] described that the addition of SiO<sub>2</sub>NPs at the rate of 50 ppm at early tillering, mid tillering, panicle initiation, and full heading caused a significant increase in fertile tillers/hill, filled grain per panicle, grain yield, and straw yield compared with control. The beneficial effect of SiO<sub>2</sub>NPs on grain yield may be ascribed to enhancing panicle fertility during grain filling [39]. Moreover, it is suggested that supplementation of Si in the nano form resulted in elevating Zn content in plant tissues, which lead to enhanced productivity [40]. It is worth noting that Si causes amendments of C/N homeostasis by remobilization of amino acids manifested by an enhancement of N needs during grain development [41].

The interactive treatments between IRs and SiO<sub>2</sub>NPs concentrations significantly affected the number of panicles/m<sup>2</sup> and 1000 grain weight, as well as both grain and biological yield in both seasons (Tables 7–10). The highest values for all these parameters are recorded in IR3 plants that are treated with SiO<sub>2</sub>NPs at 90 ppm, whereas the lowest is recorded in IR9 plants that are not treated with SiO<sub>2</sub>NPs in both experimental seasons. However, IR9 plants treated with SiO<sub>2</sub>NPs at 90 ppm gave an increase in grain productivity by 7.4% and 10.8% compared to IR9 plants with no SiO<sub>2</sub>NPs application. All these parameters in IR6 plants that are treated with SiO<sub>2</sub>NPs at 90 ppm are not significantly different than those in IR3 plants which are treated with the same level of SiO<sub>2</sub>NPs.

**Table 7.** Influence of the interaction between water regimes and SiO<sub>2</sub>NPs concentrations on panicles /m<sup>2</sup> of EHR1 rice variety during the 2018 and 2019 seasons.

SiO <sub>2</sub> NPs Concentrations	Water Regimes					
	2018			2019		
	IR3	IR6	IR9	IR3	IR6	IR9
SiO <sub>2</sub> NPs-0	595.1 c	589.2 d	430.0 g	600.0 bc	584.0 c	407.3 f
SiO <sub>2</sub> NPs-30	618.0 b	605.3 c	441.0 fg	606.3 b	605.0 b	434.0 e
SiO <sub>2</sub> NPs-60	620.6 b	623.6 ab	452.3 f	614.7 ab	619.3 ab	447.1 de
SiO <sub>2</sub> NPs-90	634.0 a	633.7 a	466.6 e	632.6 a	628.6 a	464.3 d

IR3, irrigation every 3 days; IR6, irrigation every 6 days; IR9: irrigation every 9 days. SiO<sub>2</sub>NPs-0, without SiO<sub>2</sub>NPs foliar application; SiO<sub>2</sub>NPs-30, foliar application of SiO<sub>2</sub>NPs at 30 ppm; SiO<sub>2</sub>NPs-60, foliar application of SiO<sub>2</sub>NPs at 60 ppm; SiO<sub>2</sub>NPs-90, foliar application of SiO<sub>2</sub>NPs at 90 ppm.

**Table 8.** Influence of the interaction between water regimes and SiO<sub>2</sub>NPs concentrations on the 1000-grains weight of EHR1 rice variety during the 2018 and 2019 seasons.

SiO <sub>2</sub> NPs Concentrations	Water Regimes					
	2018			2019		
	IR3	IR6	IR9	IR3	IR6	IR9
SiO <sub>2</sub> NPs-0	25.76 d	25.90 cd	24.37 f	25.88 bc	25.71 c	24.22 e
SiO <sub>2</sub> NPs-30	26.15 bc	25.99 cd	24.51 f	25.91 bc	25.90 bc	24.30 e
SiO <sub>2</sub> NPs-60	26.32 ab	26.13 bc	26.69 f	26.25 ab	26.26 ab	24.37 e
SiO <sub>2</sub> NPs-90	26.46 a	26.37 ab	25.02 e	26.39 a	26.38 a	25.03 d

IR3, irrigation every 3 days; IR6, irrigation every 6 days; IR9: irrigation every 9 days. SiO<sub>2</sub>NPs-0, without SiO<sub>2</sub>NPs foliar application; SiO<sub>2</sub>NPs-30, foliar application of SiO<sub>2</sub>NPs at 30 ppm; SiO<sub>2</sub>NPs-60, foliar application of SiO<sub>2</sub>NPs at 60 ppm; SiO<sub>2</sub>NPs-90, foliar application of SiO<sub>2</sub>NPs at 90 ppm.

The positive impact of SiO<sub>2</sub>NPs supplementation on the yield of water-stressed plants, as noted in the current study, is in line with the findings of [18] who recorded a 25% and 17.81% increase in wheat grain yield of water-stressed plants that are treated with SiO<sub>2</sub>NPs as foliar and soil application, respectively. It should be noted that [12] claimed that supplementation with SiO<sub>2</sub>NPs at 100 mg kg<sup>-1</sup> enhanced biomass and yield outcome of wheat plants grown under two deficit irrigation regimes; 70% and 35% of water-holding capacity. Application of SiO<sub>2</sub>NPs alleviated water stress resulted not only from drought but also from salinity stress. In salt-stressed rice, the application of SiO<sub>2</sub>NPs at the rate of

25 mgL<sup>-1</sup> enhanced the number of grains/panicle, panicle length, and 1000 grain weight as well as both grain and straw yield [42]. In this regard, it has been reported that the treatment with SiO<sub>2</sub>NPs caused a noticeable increase in leaf lignin content and net C assimilation of rice plants, which reinforces the plants for combating against different harsh stresses [16].

**Table 9.** Influence of the interaction between water regimes and SiO<sub>2</sub>NPs concentrations on grain yield of EHR1 rice variety during the 2018 and 2019 seasons.

SiO <sub>2</sub> NPs Concentrations	Water Regimes					
	2018			2019		
	IR9	IR6	IR9	IR9	IR6	IR9
SiO <sub>2</sub> NPs-0	11.42 d	11.40 d	8.35 f	11.25 e	11.29 e	8.14 h
SiO <sub>2</sub> NPs-30	11.94 bc	11.70 cd	8.56 ef	11.73 c	11.52 d	8.40 g
SiO <sub>2</sub> NPs-60	12.21 ab	12.01 bc	8.61 e	12.08 ab	11.98 b	8.73 g
SiO <sub>2</sub> NPs-90	12.49 a	12.41 a	8.97 e	12.24 a	12.18 a	9.02 f

IR3, irrigation every 3 days; IR6, irrigation every 6 days; IR9: irrigation every 9 days. SiO<sub>2</sub>NPs-0, without SiO<sub>2</sub>NPs foliar application; SiO<sub>2</sub>NPs-30, foliar application of SiO<sub>2</sub>NPs at 30 ppm; SiO<sub>2</sub>NPs-60, foliar application of SiO<sub>2</sub>NPs at 60 ppm; SiO<sub>2</sub>NPs-90, foliar application of SiO<sub>2</sub>NPs at 90 ppm.

**Table 10.** Influence of the interaction between water regimes and SiO<sub>2</sub>NPs concentrations on the biological yield of EHR1 rice variety during the 2018 and 2019 seasons.

SiO <sub>2</sub> NPs Concentrations	Water Regimes					
	2018			2019		
	IR3	IR6	IR9	IR3	IR6	IR9
SiO <sub>2</sub> NPs-0	26.30 de	25.94 e	20.71 h	25.87 d	25.65 d	20.17 h
SiO <sub>2</sub> NPs-30	27.04 bcd	26.70 cd	21.67 g	26.59 c	26.30 c	21.12 g
SiO <sub>2</sub> NPs-60	27.77 b	27.42 bc	22.40 fg	27.46 b	27.15 b	21.82 f
SiO <sub>2</sub> NPs-90	28.60 a	28.67 a	22.58 f	28.11 a	28.28 a	22.52 e

IR3, irrigation every 3 days; IR6, irrigation every 6 days; IR9: irrigation every 9 days. SiO<sub>2</sub>NPs-0, without SiO<sub>2</sub>NPs foliar application; SiO<sub>2</sub>NPs-30, foliar application of SiO<sub>2</sub>NPs at 30 ppm; SiO<sub>2</sub>NPs-60, foliar application of SiO<sub>2</sub>NPs at 60 ppm; SiO<sub>2</sub>NPs-90, foliar application of SiO<sub>2</sub>NPs at 90 ppm.

### 3.2.3. Effects of Water Regimes, SiO<sub>2</sub>NPs Concentrations, and Their Interaction on Chlorophyll and Proline Content

In comparison with IR3 plants, chlorophyll content, determined as SPAD value, is decreased in IR9 plants but did not significantly affect IR6 plants (Table 11). On the other hand, all tested SiO<sub>2</sub>NPs concentrations increased chlorophyll content in a concentration-dependent manner. The effect of the interaction between IRs and SiO<sub>2</sub>NPs concentrations is significant during both experimental seasons (Table 12). During both seasons, the highest chlorophyll content is recorded in the leaves of IR3 X SiO<sub>2</sub>NPs-90 plants, whereas the lowest is recorded in those of IR9 X SiO<sub>2</sub>NPs plants.

**Table 11.** The chlorophyll content is affected by water regimes and SiO<sub>2</sub>NPs concentrations of the EHR1 rice variety during the 2018 and 2019 seasons.

Seasons	Water Regimes				SiO <sub>2</sub> NPs Concentrations				IRs X SiO <sub>2</sub> NPs	
	IR3	IR6	IR9	F Test	SiO <sub>2</sub> NPs-0	SiO <sub>2</sub> NPs-30	SiO <sub>2</sub> NPs-60	SiO <sub>2</sub> NPs-90	F Test	F Test
2018	44.67 a	44.32 a	36.68 b	**	40.98 d	41.56 c	42.00 b	43.04 a	**	**
2019	43.87a	43.36 a	36.27 b	**	39.86 d	40.59 c	41.63 b	42.59 a	**	**

\*\* Indicates significant at 0.01 level. IR3, irrigation every 3 days; IR6, irrigation every 6 days; IR9: irrigation every 9 days. SiO<sub>2</sub>NPs-0, without SiO<sub>2</sub>NPs foliar application; SiO<sub>2</sub>NPs-30, foliar application of SiO<sub>2</sub>NPs at 30 ppm; SiO<sub>2</sub>NPs-60, foliar application of SiO<sub>2</sub>NPs at 60 ppm; SiO<sub>2</sub>NPs-90, foliar application of SiO<sub>2</sub>NPs at 90 ppm.

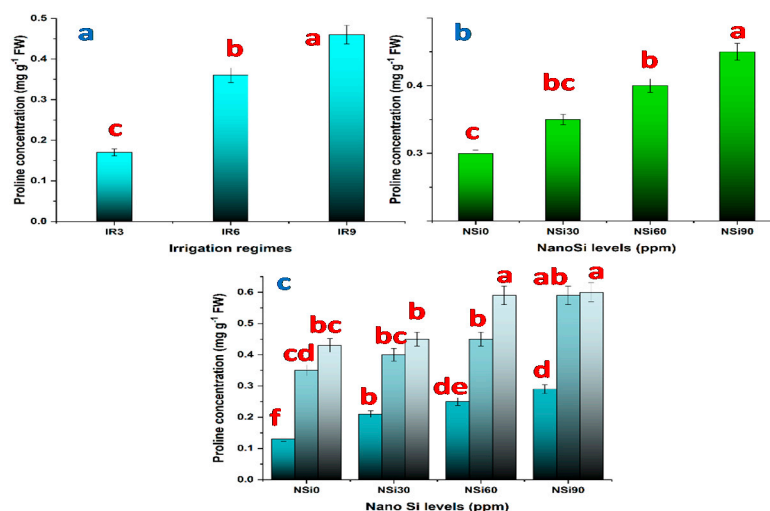
**Table 12.** The chlorophyll content is affected by the interaction between water regimes and SiO<sub>2</sub>NPs concentrations during the 2018 and 2019 seasons.

SiO <sub>2</sub> NPs Applications	Water Regimes					
	2018			2019		
	IR3	IR6	IR9	IR3	IR6	IR9
SiO <sub>2</sub> NPs-0	43.85 cd	43.67 d	35.43 f	42.34 d	41.95 e	35.30 h
SiO <sub>2</sub> NPs-30	44.24 cd	44.23 cd	36.23 f	43.33 cd	42.70 de	35.75 gh
SiO <sub>2</sub> NPs-60	44.76 bc	44.11 cd	37.15 e	44.70 ab	43.91 bc	36.30 g
SiO <sub>2</sub> NPs-90	45.83 a	45.35 ab	37.93 e	45.11 a	44.90 a	37.76 f

IR3, irrigation every 3 days; IR6, irrigation every 6 days; IR9: irrigation every 9 days. SiO<sub>2</sub>NPs-0, without SiO<sub>2</sub>NPs foliar application; SiO<sub>2</sub>NPs-30, foliar application of SiO<sub>2</sub>NPs at 30 ppm; SiO<sub>2</sub>NPs-60, foliar application of SiO<sub>2</sub>NPs at 60 ppm; SiO<sub>2</sub>NPs-90, foliar application of SiO<sub>2</sub>NPs at 90 ppm.

The depressive effect of water stress on chlorophyll content is recorded in the current study and has also been previously reported by [20,31]. Additionally, [18] documented an increase in chlorophyll concentration under various stressful water regimes when the wheat plants have been treated with SiO<sub>2</sub>NPs using different concentrations (30, 60, and 90 ppm) under various stressful water regimes. Since the production of reactive oxygen species is mostly caused by excess energy absorption in the photosynthetic apparatus, such an issue can be prevented by destroying or degrading the absorbing pigments [43]. Moreover, water stress-induced K<sup>+</sup> deficiency may cause losses in leaf chlorophyll [32]. Also, in alignment with the gained results of the current research study [20] reported an increase in chlorophyll and carotenoid content in barely seedling in response to plants treated with SiO<sub>2</sub>NPs at the rate of 125 mL/L in moderate conditions of water-stress.

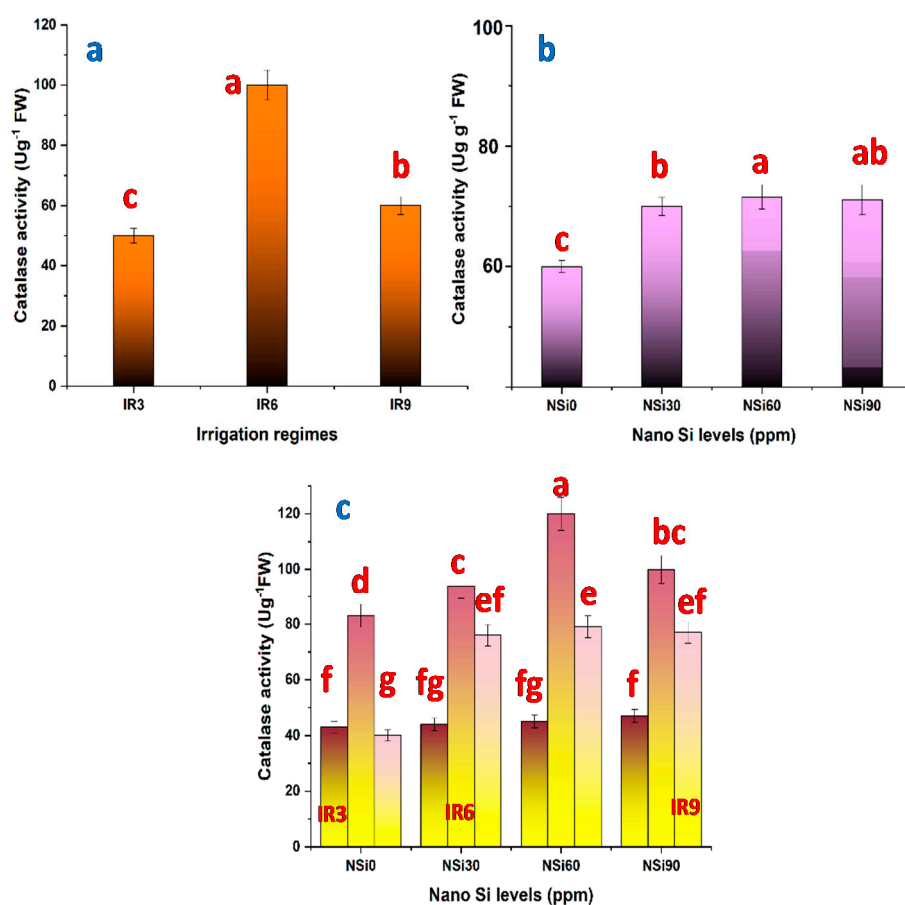
On the other hand, proline concentration is increased significantly under water-stressed plants compared with unstressed plants (Figure 2a). Relative to the concentration in IR3 (unstressed plants), proline concentration is increased by 86.9% and 117.3% in response to IR6 and IR9, respectively. Moreover, SiO<sub>2</sub>NPs treatments increased proline concentration; however, the increase recorded in response to 30 ppm SiO<sub>2</sub>NPs is insignificant (Figure 2b). At 60, 90 ppm SiO<sub>2</sub>NPs, proline concentration increased by 33.3%, 56.6%, respectively. The interaction between IRs and SiO<sub>2</sub>NPs levels is significant. The highest proline concentration is recorded in plants that are irrigated every 9 days and treated with SiO<sub>2</sub>NPs at 90 ppm, followed by that in plants exposed to the same level of water stress and treated with SiO<sub>2</sub>NPs at 60 ppm (Figure 2c).

**Figure 2.** Proline concentration in rice leaves as affected by (a) irrigation regimes, (b) nano Si levels, and (c) their interactions.

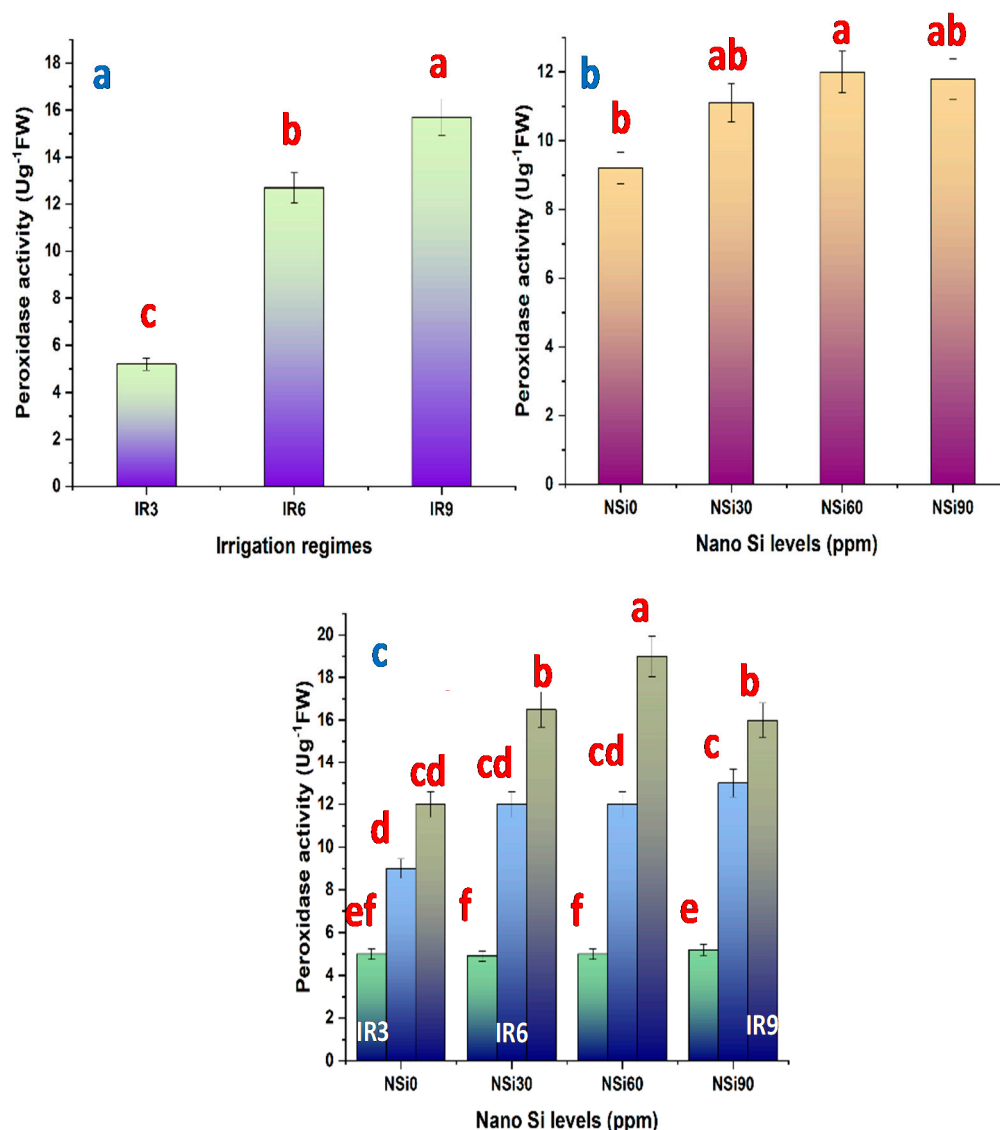
The response of plants to drought stress involves several changes in physiology and metabolism that displayed remarkable differences in their morphology. The result of the present study implied that deficit irrigation increased proline content in rice plants (Figure 2a). Following the findings of our current study that the content of proline increased in response to water stress in rice. Besides that, the content of proline for all upland rice varieties increases dramatically to the highest drought level [44]. Proline accumulation in plants under water stress protects the cell by balancing the osmotic potential of cytosol with that of vacuole and the external environment [45]. Accumulated proline could, therefore, serve as a compatible solute that controls and reduces plant cell water loss during water deficit and plays an important role in osmotic balance [46]. Moreover, accumulated proline under stress also provides energy for survival and growth, thereby, enabling the plants to resist stress conditions with regard to these phenomena, proline content is considered a good indicator for screening drought-tolerant genotypes in water stressful environments [4,47].

### 3.2.4. Antioxidant Enzymes Activity

Prolonging the irrigation interval from 3 to 6 days increased the activity of CAT (Figure 3a), POD (Figure 4a), and SOD (Figure 5a). With increasing the magnitude of water stress further by extending the irrigation interval to every 9 days (IR9), the activity of POD tended to be increased further, whereas those of CAT and SOD tended to be decreased, but still higher compared with control plants. However, the increase recorded regarding SOD activity in IR9 plants was not significant. Compared with the activity of POD in control plants, this activity was increased by 129.4% and 211.7% in IR6- and IR9-plants, respectively.



**Figure 3.** Catalase activity in rice leaves as affected by (a) irrigation regimes, (b) nano Si levels, and (c) their interactions.



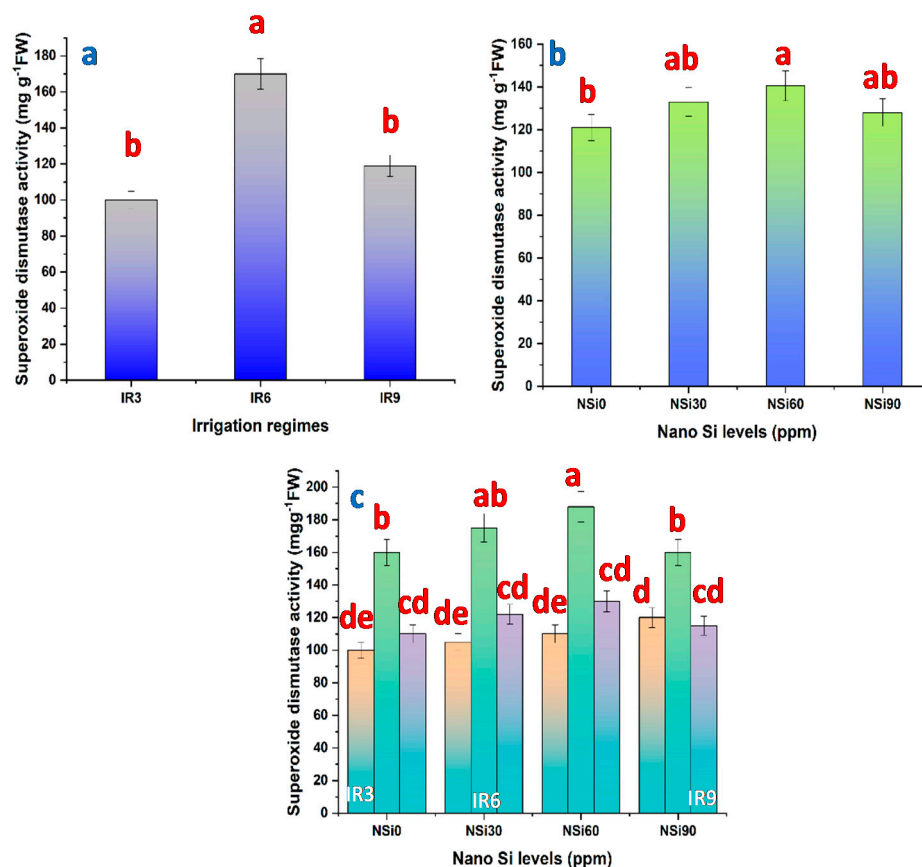
**Figure 4.** Peroxidase activity in rice leaves as affected by (a) irrigation regimes, (b) nano Si levels, and (c) their interactions.

All levels of SiO<sub>2</sub>NPs significantly increased the activity of CAT (Figure 3b), and the highest increase was recorded in response to SiO<sub>2</sub>NPs at 60 ppm. Although all tested levels of SiO<sub>2</sub>NPs increased the activity of POD (Figure 4b) and SOD (Figure 5b) compared with control plants, the increase was significant only in plants treated with SiO<sub>2</sub>NPs at 60 ppm. So, when plants were treated with SiO<sub>2</sub>NPs at 30 ppm, the activity of both POD and SOD was elevated, and further increased when the level of SiO<sub>2</sub>NPs was increased to 60 ppm, then decreased by increasing the level of SiO<sub>2</sub>NPs to 90 ppm.

The interaction between irrigation intervals and SiO<sub>2</sub>NPs levels affected significantly the activity of CAT (Figure 3c), POD (Figure 4c), and SOD (Figure 5c). The highest increase in CAT and SOD activity was recorded in plants irrigated every 6 days and treated with SiO<sub>2</sub>NPs at 60 ppm. On the other hand, the highest POD activity was recorded in plants irrigated every 9 days and treated with SiO<sub>2</sub>NPs at 60 ppm.

Drought stress contributes to the oxidative stress induced by the accumulation of reactive oxygen species (ROS), created mostly in chloroplast and to some extent in mitochondria. Plants could eventually create various types of antioxidants to detoxify ROS that reduces oxidative damage and confer resistance to drought. The ROS scavengers may be enzymatic including superoxide dismutase, peroxidase, and catalase, or non-enzymatic e.g., ascorbate, glutathione, and tocopherols. By extending irrigation duration, from “every

three days” to “every six days”, the activities of the antioxidant enzymes, CAT, POD, SOD are increased (Figures 3–5).



**Figure 5.** Superoxide dismutase in rice leaves as affected by (a) irrigation regimes, (b) nano Si levels, and (c) their interactions.

In water-stressed rice, higher ROS-scavenging enzyme activities have been identified and it has been implied that in water-stressed that the antioxidant system’s component plays a key role in plant protection against water stress rice [48]. In compliance with [49], since plants have established enzymatic and non-enzymatic mechanisms to scavenge ROS, sensitivity to drought-stress in plants corresponds to the levels of antioxidant systems and the substrate is higher, which implies that plants will produce more CAT, SOD, and POD under conditions of drought to remove the extra ROS in cells. Catalase has the potential to directly dismutase H<sub>2</sub>O<sub>2</sub> into H<sub>2</sub>O and O<sub>2</sub> and is indispensable for ROS detoxification in peroxisomes during stress conditions [50]. Superoxide dismutase detoxifies superoxide anion (O<sub>2</sub><sup>-</sup>) by forming H<sub>2</sub>O<sub>2</sub>, and then the H<sub>2</sub>O<sub>2</sub> can be eliminated by CA and POD [51]. Moreover, POD is also involved in various plant processes, including lignification [52], oxidation of phenolics [53], regulation of cell elongation, and detoxification of toxic compounds such as H<sub>2</sub>O<sub>2</sub> [54]. Additionally, stress-tolerant genotypes have been distinguished by the higher activities of antioxidant enzymes. In particular, the drought-tolerant genotypes of *Triticum aestivum* [51,55] have higher activities of SOD, POD, and CAT than the drought-sensitive species.

According to the results of the present study, SiO<sub>2</sub>NPs increased proline content and enhanced the activity of the antioxidant enzymes in a concentration-dependent manner (Figures 2–5). Similar results are reported with regard to proline and antioxidant enzymes. It seems that SiO<sub>2</sub>NPs could alter the activity of antioxidative enzymes in plant organs to improve stress tolerance, suggesting that the treated plants possess a better scavenging ability [18,20,48]. Moreover, it has been reported that the highest activities of superoxide

dismutase, catalase, and peroxidase in Changbai larch (*Larix olgensis*) and soybean plants are recorded in response to SiO<sub>2</sub>NPs [56].

#### 4. Conclusions

It could be concluded based on the results obtained that the Egyptian rice cultivar EHR1 is not highly water-demanding so that irrigation every 6 days did not cause harm either to its growth or to its yield. Moreover, treatment with SiO<sub>2</sub>NPs can reduce water stress effects on growth and reproductive structures, thereby avoiding substantial yield loss. According to our obtained data, SiO<sub>2</sub>NPs with a concentration of 90 ppm is the optimal and best concentration for achieving our promising aim. Since this concentration is the highest used, there is still a probability that a higher level leads to more gains, and thereby there is a need for more studies in this regard to continue to explore and examine the alleviate potential of SiO<sub>2</sub>NPs on rice plants growing in water-stressed conditions.

**Author Contributions:** Conceptualization, O.M.E., H.M.I., H.E.A. and A.MN.; methodology, O.M.E., H.M.I., H.E.A. and A.MN.; software, O.M.E., H.M.I., H.E.A. and A.MN.; validation, O.M.E., H.M.I., H.E.A. and A.MN.; formal analysis, O.M.E., H.M.I., H.E.A. and A.MN.; investigation, O.M.E.; resources, A.M.A.; data curation, O.M.E., H.M.I., H.E.A., A.MN. and A.M.A.; writing—original draft preparation, O.M.E. and A.M.A.; writing—review and editing O.M.E., H.M.I., H.E.A., A.MN. and A.M.A.; visualization, A.M.A.; supervision, A.M.A.; project administration, O.M.E. and A.M.A.; funding acquisition, A.M.A. All authors have read and agreed to the published version of the manuscript.

**Funding:** King Saud University, researchers supporting project number (RSP-2020/63), King Saud University, Riyadh, Saudi Arabia.

**Institutional Review Board Statement:** Not applicable.

**Informed Consent Statement:** Not applicable.

**Data Availability Statement:** The data presented in this study are available on request from the corresponding author.

**Acknowledgments:** The authors acknowledge King Saud University, researchers supporting project number (RSP-2020/63), King Saud University, Riyadh, Saudi Arabia for funding support.

**Conflicts of Interest:** The authors declare no conflict of interest.

#### References

- Hasan, S.A. Price hike of staple food, nutritional impact and consumption adjustment: Evidence from the 2005–2010 rice price increase in rural Bangladesh. *Appl. Econ.* **2019**, *51*, 743–761. [[CrossRef](#)]
- Seck, P.A.; Diagne, A.; Mohanty, S.; Wopereis, M.C.S. Crops that feed the world 7: Rice. *Food Secur.* **2012**, *4*, 7–24. [[CrossRef](#)]
- O'Toole, J.C. *Rice and Water: The Final Frontier*; Rockefeller Foundation: Bangkok, Thailand, 2004.
- Kumar, S.; Dwivedi, S.K.; Basu, S.; Kumar, G.; Mishra, J.; Koley, T.K.; Rao, K.; Choudhary, A.; Mondal, S.; Kumar, S.; et al. Anatomical, agro-morphological and physiological changes in rice under cumulative and stage specific drought conditions prevailed in eastern region of India. *Field Crops Res.* **2020**, *245*, 107658. [[CrossRef](#)]
- Ndjiondjop, M.-N.; Cisse, F.; Futakuchi, K.; Lorieux, M.; Manneh, B.; Bocco, R.; Fatondji, B. Effect of drought on rice (*Oryza* spp.) genotypes according to their drought tolerance level. In Proceedings of the Second Africa Rice Congress, Bamako, Mali, 22–26 March 2010; pp. 1–15.
- Bocco, R.; Lorieux, M.; Seck, P.; Futakuchi, K.; Manneh, B.; Baimey, H.; Ndjiondjop, M. Agro-morphological characterization of a population of introgression lines derived from crosses between IR 64 (*Oryza sativa indica*) and TOG 5681 (*Oryza glaberrima*) for drought tolerance. *Plant Sci.* **2012**, *183*, 65–76. [[CrossRef](#)] [[PubMed](#)]
- Cruz de Carvalho, M.H. Drought stress and reactive oxygen species: Production, scavenging and signaling. *Plant Signal. Behav.* **2008**, *3*, 156–165. [[CrossRef](#)] [[PubMed](#)]
- Datnoff, L.E.; Snyder, G.H.; Korndörfer, G.H. *Silicon in Agriculture*; Elsevier: Amsterdam, The Netherlands, 2001.
- Ma, J.F. Silicon uptake and translocation in plants. In Proceedings of the International Plant Nutrition Colloquium XVI, Davis, CA, USA, 26–30 August 2009.
- Mehrabanjoubani, P.; Abdolzadeh, A.; Sadeghipour, H.R.; Aghdasi, M. Silicon affects transcellular and apoplastic uptake of some nutrients in plants. *Pedosphere* **2015**, *25*, 192–201. [[CrossRef](#)]
- Cui, J.; Liu, T.; Li, F.; Yi, J.; Liu, C.; Yu, H. Silica nanoparticles alleviate cadmium toxicity in rice cells: Mechanisms and size effects. *Environ. Pollut.* **2017**, *228*, 363–369. [[CrossRef](#)] [[PubMed](#)]



12. Khan, Z.S.; Rizwan, M.; Hafeez, M.; Ali, S.; Adrees, M.; Qayyum, M.F.; Khalid, S.; Rehman, M.Z.U.; Sarwar, M.A. Effects of silicon nanoparticles on growth and physiology of wheat in cadmium contaminated soil under different soil moisture levels. *Environ. Sci. Pollut. Res.* **2020**, *27*, 4958–4968. [[CrossRef](#)]
13. Abdel-Halim, M.E.; Hegazy, H.S.; Hassan, N.S.; Naguib, D.M. Effect of silica ions and nano silica on rice plants under salinity stress. *Ecol. Eng.* **2017**, *99*, 282–289. [[CrossRef](#)]
14. Tripathi, D.K.; Singh, S.; Singh, V.P.; Prasad, S.M.; Dubey, N.K.; Chauhan, D.K. Silicon nanoparticles more effectively alleviated UV-B stress than silicon in wheat (*Triticum aestivum*) seedlings. *Plant Physiol. Biochem.* **2017**, *110*, 70–81. [[CrossRef](#)]
15. Li, L.; Huang, J.; Sun, L.; Liu, L.; Wang, L.; Hou, Y.; Wang, A.; Wang, G.; Huang, S. The effects of nanomaterial treated water on the pathogens of rice diseases and fungicides. *Nanosci. Nanotechnol. Lett.* **2017**, *9*, 957–963. [[CrossRef](#)]
16. Alvarez, R.d.C.F.; de Mello Prado, R.; Felisberto, G.; Deus, A.C.F.; de Oliveira, R.L.L. Effects of soluble silicate and nanosilica application on rice nutrition in an Oxisol. *Pedosphere* **2018**, *28*, 597–606. [[CrossRef](#)]
17. Cáceres, M.; Vassena, C.V.; Garcerá, M.D.; Santo-Orihuela, P.L. Silica nanoparticles for insect pest control. *Curr. Pharm. Des.* **2019**, *25*, 4030–4038. [[CrossRef](#)]
18. Behboudi, F.; Tahmasebi Sarvestani, Z.; Kassae, M.Z.; Modares Sanavi, S.A.M.; Sorooshzadeh, A. Improving growth and yield of wheat under drought stress via application of SiO<sub>2</sub> nanoparticles. *J. Agric. Sci. Technol.* **2018**, *20*, 1479–1492.
19. Moonmoon, S.; Fakir, M.; Islam, M. Effect of drought stress on grain dry weight, photosynthesis and chlorophyll in six rice genotypes. *Sch. J. Agric. Vet. Sci.* **2017**, *4*, 13–17.
20. Ghorbanpour, M.; Mohammadi, H.; Kariman, K. Nanosilicon-based recovery of barley (*Hordeum vulgare*) plants subjected to drought stress. *Environ. Sci. Nano* **2020**, *7*, 443–461. [[CrossRef](#)]
21. Sharifi Rad, J.; Karimi, J.; Mohsenzadeh, S.; Sharifi Rad, M.; Moradgholi, J. Evaluating SiO<sub>2</sub> nanoparticles effects on developmental characteristic and photosynthetic pigment contents of *Zeamays* L. *Bull. Environ. Pharm. Life Sci.* **2014**, *3*, 194–201.
22. Qados, A.M.S.A. Mechanism of nanosilicon-mediated alleviation of salinity stress in Faba bean (*Vicia faba* L.) Plants. *Am. J. Exp. Agric.* **2015**, *7*, 78–95. [[CrossRef](#)]
23. Bottomley, P.J.; Angle, J.S.; Weaver, R. *Methods of Soil Analysis, Part 2: Microbiological and Biochemical Properties*; John Wiley & Sons: Hoboken, NJ, USA, 2020; Volume 12.
24. Yoshida, S.; Forno, D.A.; Cock, J.H.; Gomez, K.A. *Laboratory Manual for Physiological Studies of Rice*; The International Rice Research Institute: Los Baños, Philippines, 1971.
25. Yoshida, S. *Fundamentals of Rice Crop Science*; The International Rice Research Institute: Los Baños, Philippines, 1981.
26. Bates, L.S.; Waldren, R.P.; Teare, I.D. Rapid determination of free proline for water-stress studies. *Plant Soil* **1973**, *39*, 205–207. [[CrossRef](#)]
27. Aebi, H. Catalase in vitro. In *Methods in Enzymology*; Elsevier: Amsterdam, The Netherlands, 1984; Volume 105, pp. 121–126.
28. Urbanek, H.; Kuzniak-Gebrowska, E.; Herka, K. Elicitation of defence responses in bean leaves by *Botrytis cinerea polygalacturonase*. *Acta Physiol. Plant.* **1991**, *13*, 43–50.
29. Van Rossum, M.W.; Alberda, M.; van der Plas, L.H. Role of oxidative damage in tulip bulb scale micropropagation. *Plant Sci.* **1997**, *130*, 207–216. [[CrossRef](#)]
30. Duncan, D.B. Multiple range and multiple F tests. *Biometrics* **1955**, *11*, 1–42. [[CrossRef](#)]
31. Emam, M.M.; Khattab, H.E.; Helal, N.M.; Deraz, A.E. Effect of selenium and silicon on yield quality of rice plant grown under drought stress. *Aust. J. Crop Sci.* **2014**, *8*, 596.
32. Hamoud, Y.A.; Wang, Z.; Guo, X.; Shaghaleh, H.; Sheteiwy, M.; Chen, S.; Qiu, R.; Elbashier, M.M.A. Effect of irrigation regimes and soil texture on the potassium utilization efficiency of rice. *Agronomy* **2019**, *9*, 100. [[CrossRef](#)]
33. Yang, R.; Howe, J.A.; Golden, B.R. Calcium silicate slag reduces drought stress in rice (*Oryza sativa* L.). *J. Agron. Crop Sci.* **2019**, *205*, 353–361. [[CrossRef](#)]
34. Liang, Y. Effect of silicon on leaf ultrastructure, chlorophyll content and photosynthetic activity of barley under salt stress. *Pedosphere* **1998**, *8*, 289–296.
35. Fauteux, F.; Rémus-Borel, W.; Menzies, J.G.; Bélanger, R.R. Silicon and plant disease resistance against pathogenic fungi. *FEMS Microbiol. Lett.* **2005**, *249*, 1–6. [[CrossRef](#)]
36. Sangster, A.G.; Hodson, M.J.; Tubb, H.J. Chapter 5. Silicon deposition in higher plants. In *Silicon in Agriculture*; Elsevier BV: Amsterdam, The Netherlands, 2001; pp. 85–113.
37. Maghsoudi, K.; Emam, Y.; Pessaraki, M. Effect of silicon on photosynthetic gas exchange, photosynthetic pigments, cell membrane stability and relative water content of different wheat cultivars under drought stress conditions. *J. Plant. Nutr.* **2016**, *39*, 1001–1015. [[CrossRef](#)]
38. Wang, S.; Wang, F.; Gao, S. Foliar application with nano-silicon alleviates Cd toxicity in rice seedlings. *Environ. Sci. Pollut. Res.* **2014**, *22*, 2837–2845. [[CrossRef](#)]
39. Tamai, K.; Ma, J.F. Reexamination of silicon effects on rice growth and production under field conditions using a low silicon mutant. *Plant Soil* **2008**, *307*, 21–27. [[CrossRef](#)]
40. Kheyri, N.; Norouzi, H.A.; Mobasser, H.R.; Torabi, B. Effects of silicon and zinc nanoparticles on growth, yield, and biochemical characteristics of rice. *Agron. J.* **2019**, *111*, 3084–3090. [[CrossRef](#)]
41. Detmann, K.C.; Araújo, W.L.; Martins, S.C.; Fernie, A.R.; DaMatta, F.M. Metabolic alterations triggered by silicon nutrition: Is there a signaling role for silicon? *Plant Signal. Behav.* **2013**, *8*, e22523. [[CrossRef](#)]

42. Kheir, A.M.S.; Abouelsoud, H.M.; Hafez, E.M.; Ali, O.A.M. Integrated effect of nano-Zn, nano-Si, and drainage using crop straw-filled ditches on saline sodic soil properties and rice productivity. *Arab. J. Geosci.* **2019**, *12*, 471. [[CrossRef](#)]
43. Herbinger, K.; Tausz, M.; Wonisch, A.; Soja, G.; Sorger, A.; Grill, D. Complex interactive effects of drought and ozone stress on the antioxidant defence systems of two wheat cultivars. *Plant Physiol. Biochem.* **2002**, *40*, 691–696. [[CrossRef](#)]
44. Saha, S.; Begum, H.H.; Nasrin, S. Effects of drought stress on growth and accumulation of proline in five rice varieties (*Oryza Sativa* L.). *J. Asiat. Soc. Bangladesh Sci.* **2019**, *45*, 241–247. [[CrossRef](#)]
45. Johari-Pireivatlou, M. Effect of soil water stress on yield and proline content of four wheat lines. *Afr. J. Biotechnol.* **2010**, *9*, 36–40.
46. Lum, M.; Hanafi, M.; Rafii, Y.; Akmar, A. Effect of drought stress on growth, proline and antioxidant enzyme activities of upland rice. *J. Anim. Plant Sci.* **2014**, *24*, 1487–1493.
47. Rahdari, P.; Hosseini, S.M.; Tavakoli, S. The studying effect of drought stress on germination, proline, sugar, lipid, protein and chlorophyll content in purslane (*Portulaca oleracea* L.) leaves. *J. Med. Plants Res.* **2012**, *6*, 1539–1547.
48. Hegazy, H.; Hassan, N.; Abdel-Haliem, M.; Naguib, D. biochemical response of rice plant to biotic and abiotic stress under silica ions and nanoparticles application. *Egypt. J. Bot.* **2015**, *55*, 79–103.
49. Athar, H.-U.-R.; Khan, A.; Ashraf, M. Exogenously applied ascorbic acid alleviates salt-induced oxidative stress in wheat. *Environ. Exp. Bot.* **2008**, *63*, 224–231. [[CrossRef](#)]
50. Sairam, R.K.; Rao, K.; Srivastava, G. Differential response of wheat genotypes to long term salinity stress in relation to oxidative stress, antioxidant activity and osmolyte concentration. *Plant Sci.* **2002**, *163*, 1037–1046. [[CrossRef](#)]
51. Hasheminasab, H.; Assad, M.T.; Aliakbari, A.; Sahhafi, S.R. Influence of drought stress on oxidative damage and antioxidant defense systems in tolerant and susceptible wheat genotypes. *J. Agric. Sci.* **2012**, *4*, 20. [[CrossRef](#)]
52. Hendriks, T.; Wijsman, H.J.W.; Loon, L.C. Petunia peroxidase a: Isolation, purification and characteristics. *JBIC J. Biol. Inorg. Chem.* **1991**, *199*, 139–146. [[CrossRef](#)]
53. Lagrimini, L.M. Wound-induced deposition of polyphenols in transgenic plants overexpressing peroxidase. *Plant Physiol.* **1991**, *96*, 577–583. [[CrossRef](#)]
54. Mohammadkhani, N.; Heidari, R. Drought-induced accumulation of soluble sugars and proline in two maize varieties. *World Appl. Sci. J.* **2008**, *3*, 448–453.
55. Almaghrabi, O.A. Impact of drought stress on germination and seedling growth parameters of some wheat cultivars. *Life Sci. J.* **2012**, *9*, 590–598.
56. Lin, B.-S.; Diao, S.-Q.; Li, C.-H.; Fang, L.-J.; Qiao, S.-C.; Yu, M. Effect of TMS (nanostructured silicon dioxide) on growth of Changbai larch seedlings. *J. For. Res.* **2004**, *15*, 138–140. [[CrossRef](#)]