MycoKeys 60: 17–29 (2019)
doi: 10.3897/mycokeys.60.46395
http://mycokeys.pensoft.net

RESEARCH ARTICLE

A peer-reviewed open-access journal
MycoKeys
Launched to accelerate biodiversity research

Taxonomy of two synnematal fungal species from *Rhus chinensis*, with *Flavignomonia* gen. nov. described

Ning Jiang[1*], Qin Yang[1,2*], Ying-Mei Liang[3], Cheng-Ming Tian[1]

1 *The Key Laboratory for Silviculture and Conservation of the Ministry of Education, Beijing Forestry University, Beijing 100083, China* **2** *Forestry Biotechnology Hunan Key Laboratories, Central South University of Forestry and Technology, Changsha 410004, China* **3** *Museum of Beijing Forestry University, Beijing Forestry University, Beijing 100083, China*

Corresponding author: *Cheng-Ming Tian* (chengmt@bjfu.edu.cn)

Academic editor: *G. Rambold* | Received 8 September 2019 | Accepted 23 October 2019 | Published 31 October 2019

Citation: Jiang N, Yang Q, Liang Y-M, Tian C-M (2019) Taxonomy of two synnematal fungal species from *Rhus chinensis*, with *Flavignomonia* gen. nov. described. MycoKeys 60: 17–29. https://doi.org/10.3897/mycokeys.60.46395

Abstract

Rhus chinensis represents a commercially and ecologically important tree species in China, but suffers from canker diseases in Jiangxi Province. Synnemata, pycnidia and ascomata were discovered on cankered tissues. Strains were obtained from single ascospore or conidium within the fruiting bodies and identified based on morphological comparison and the phylogenetic analyses of partial ITS, LSU, *tef1* and *rpb2* gene sequences. As a result, two species were confirmed to represent two kinds of synnemata. One of these species is described herein as *Flavignomonia rhoigena* **gen. et sp. nov.**; and *Synnemasporella aculeans* is illustrated showing ascomata, pycnidia and synnemata. *Flavignomonia* is distinguished from *Synnemasporella* by the colour of the synnematal tips. Additionally, *Flavignomonia* can be distinguished from the other gnomoniaceous genera by the formation of synnemata.

Keywords

Diaporthales, Gnomoniaceae, systematics, taxonomy

Introduction

Many Diaporthales species are important branch canker pathogens, forming acervuli or pycnidia on diseased tissues (Rossman et al. 2007, Senanayake et al. 2017, Jiang et al. 2018, 2019, Wijayawardene et al. 2018, Yang et al. 2018, Voglmayr et al. 2019).

* These authors contributed equally to this work and should be considered as co-first authors.

However, two diaporthalean species with synnemata were reported to cause cankers, namely *Synnemasporella aculeans* (syn. *Cryptodiaporthe aculeans*) and *S. toxicodendri* (Fan et al. 2018). These two species differ from the other diaporthalean taxa in conidiomata and form a distinct clade phylogenetically, which was named Synnemasporellaceae and distinguished by Fan et al. (2018).

Gnomoniaceae was initially introduced with *Gnomonia* as the type (Winter 1886). Species in Gnomoniaceae formed upright perithecia, with or without long or short necks and presence or absence of stromatic tissues (Barr 1978, Sogonov et al. 2008, Walker et al. 2012). In the recent monograph of Diaporthales, 34 genera were accepted in the family Gnomoniaceae (Senanayake et al. 2018). Subsequently, *Neognomoniopsis* and *Tenuignomonia* were added based on both molecular and morphological evidence (Crous et al. 2019, Minoshima et al. 2019).

Chinese gall (*Rhus chinensis* Mill.) has a range of uses as source of medicine, dye and oil, and has a wide distribution in China (Wang et al. 2014). However, cankers were found to be associated with different ascomata during our fungal collection trips in Jiangxi Province, China. The objectives of the present study were to identify these fungi based on morphological and phylogenetic evidence.

Materials and methods

Sample collections and isolation

We conducted our fungal collection surveys from April to October in China, and found *Rhus chinensis* to be one of the major tree species in Jiangxi Province. Twigs, branches and stems were carefully checked, and diseased tissues were cut into small pieces and packed in paper bags. Isolates were obtained by transferring the ascospores or conidial masses from ascomata to sterile PDA plates, incubating at 25 °C until spores germinated. Single germinating spores were transferred onto new PDA plates, which were kept at 25 °C in darkness. Specimens were deposited in the Museum of the Beijing Forestry University (**BJFC**) and axenic cultures maintained in the China Forestry Culture Collection Centre (**CFCC**).

Morphological analysis

Recognition and identification of the fungal species on *Rhus chinensis* was based on fruiting bodies formed on the bark. Ascomata and conidiomata were sectioned by hand using a double-edged blade, and microscopic structures were observed under a dissecting microscope. At least 10 conidiomata/ascomata, 10 asci, and 50 conidia/ascospores were measured to calculate mean and standard deviation. Measurements are reported as maxima and minima in parentheses and the range representing the mean plus and minus the standard deviation of the number of measurements given in parentheses (Voglmayr et al. 2017). Microscopy photographs were captured with a Nikon Eclipse

80i compound microscope equipped with a Nikon digital sight DS-Ri2 high definition colour camera, using differential interference contrast illumination. Nomenclatural novelties and descriptions were deposited in MycoBank (Crous et al. 2004).

DNA extraction, PCR amplification and sequencing

Genomic DNA was extracted from colonies grown on cellophane-covered PDA plates using a modified CTAB method (Doyle and Doyle 1990). PCR amplifications were performed in a DNA Engine Peltier Thermal Cycler (PTC-200; Bio-Rad Laboratories, Hercules, CA, USA). The primer sets ITS1/ITS4 (White et al. 1990) were used to amplify the ITS region. The primer pair LR0R/LR5 (Vilgalys and Hester 1990) was used to amplify the LSU region. The primer pairs EF1-688F/EF1-986R or EF1-728F/TEF1-LLErev (Carbone and Kohn 1999; Jaklitsch et al. 2005; Alves et al. 2008) were used to amplify *tef1* gene. The primer pair dRPB2-5f/dRPB2-7r (Voglmayr et al. 2016) was used to amplify the *rpb2* gene. The polymerase chain reaction (PCR) assay was conducted as described by Fan et al. (2018). PCR amplification products were assayed via electrophoresis in 2 % agarose gels. DNA sequencing was performed using an ABI PRISM 3730XL DNA Analyzer with a BigDye Terminater Kit v.3.1 (Invitrogen, USA) at the Shanghai Invitrogen Biological Technology Company Limited (Beijing, China).

Phylogenetic analyses

The preliminary identities of the isolates sequenced in this study were obtained by conducting a standard nucleotide BLAST search using the sequences generated from the above primers of the different genomic regions (ITS, LSU, *tef1* and *rpb2*). The BLAST results showed that three isolates were grouped in the family Gnomoniaceae, and five isolates in the genus *Synnemasporella*. The phylogenetic analyses for the three gnomoniaceous isolates were conducted based on Senanayake et al. (2018), supplemented by sequences of *Tenuignomonia styracis* and *Neognomoniopsis quercina* from Crous et al. (2019) and Minoshima et al. (2019). *Melanconis marginalis* (CBS 109744) in Melanconidaceae was selected as the out-group taxon. All sequences were aligned using MAFFT v. 6 (Katoh and Toh 2010) and edited manually using MEGA v. 6 (Tamura et al. 2013). Phylogenetic analyses were performed using PAUP v. 4.0b10 for maximum parsimony (MP) analysis (Swofford 2003), and PhyML v. 3.0 for Maximum Likelihood (ML) analysis (Guindon et al. 2010).

MP analysis was run using a heuristic search option of 1000 search replicates with random additions of sequences with a tree bisection and reconnection algorithm. Other calculated parsimony scores were tree length (TL), consistency index (CI), retention index (RI), and rescaled consistency (RC). ML analysis was performed using a GTR site substitution model including a gamma-distributed rate heterogeneity and a proportion of invariant sites (Guindon et al. 2010). The branch support was evaluated using a bootstrapping method of 1000 replicates (Hillis and Bull 1993). The matrix

was partitioned for the different gene regions. Phylograms were shown using FigTree v. 1.4.3 (Rambaut 2016). Novel sequences generated in the current study were deposited in GenBank (Table 1) and the aligned matrices used for phylogenetic analyses in TreeBASE (accession number: S25047).

Results

Phylogenetic analyses

The alignment based on the combined sequence dataset (ITS, LSU, *tef1*, and *rpb2*) included 42 in-group taxa and one out-group taxon, comprising 3368 characters in the aligned matrix. Of these, 2201 characters were constant, 224 variable characters were parsimony-uninformative and 943 characters were parsimony informative (282 from the ITS-LSU, 280 from *tef1*, 381 from *rpb2*). The MP analysis resulted in nine equally most parsimonious trees with identical tree backbone. The best ML tree (lnL = –20604.0384) was compatible with the MP strict consensus tree, except for unsupported clades in Fig. 1. As the trees obtained from different analytical methods were similar, only the ML tree was present in Fig. 1. The phylogram based on the four gene sequence matrix indicated that the three strains from the present study represent a novel genus in Gnomoniaceae.

Taxonomy

Flavignomonia **C.M. Tian, Q. Yang & N. Jiang, gen. nov.**
MycoBank No: 829530

Diagnosis. *Flavignomonia* is distinguished from *Synnemasporella* by the orange tips of its synnemata.

Type species. *Flavignomonia rhoigena* C.M. Tian & Q. Yang

Etymology. The generic name is derived from the colour of synnemata (flavus = yellow) and the genus name *Gnomonia*.

Description. Sexual morph: not observed. Asexual morph: Conidiomata synnematal. Synnemata long and determinate, growing from host tissue, with brown base and orange tip, straight to curved, parallel, with flat to slightly concave and dark zone of conidiogenous cells and host tissue at their bases. Conidiophores reduced to conidiogenous cells. Conidiogenous cells phialidic, aggregated, hyaline, straight to curved, cylindrical, arranged adjacent to one another at the end of the synnema, producing a single conidium. Conidia cylindrical to oblong, smooth, multiguttulate, hyaline.

Notes. *Flavignomonia* is included in Gnomoniaceae based on DNA sequences data. *Flavignomonia* is morphologically similar to *Synnemasporella* in forming synne-

Table 1. Strains used in the phylogenetic tree and their culture accession and GenBank numbers. Strains from this study are in bold.

Species	Strains	GenBank numbers			
		ITS	LSU	*tef1*	*rpb2*
Alnecium auctum	CBS 124263	KF570154	KF570154	KF570200	KF570170
Ambarignomonia petiolorum	CBS 116866	EU199193	AY818963	NA	EU199151
	CBS 121227	EU254748	EU255070	EU221898	EU219307
Amphiporthe tiliae	CBS 119289	EU199178	EU199122	NA	EU199137
Anisogramma anomala	529478	EU683064	EU683066	NA	NA
Anisogramma virgultorum	529479	EU683062	EU683065	NA	NA
Apiognomonia errabunda	AR 2813	DQ313525	NG027592	DQ313565	DQ862014
Apiognomonia veneta	MFLUCC 16-1193	MF190114	MF190056	NA	NA
Apioplagiostoma populi	858501	KP637024	NA	NA	NA
Asteroma alneum	CBS 109840	EU167609	EU167609	NA	NA
Asteroma sp.	Masuya 8Ah9-1	NA	AB669035	NA	NA
Cryptosporella hypodermia	CBS 116866	EU199181	AF408346	NA	EU199140
Discula destructiva	MD 254	AF429741	AF429721	AF429732	NA
Ditopella biseptata	MFLU 15-2661	MF190147	MF190091	NA	MF377616
Ditopella ditopa	CBS 109748	DQ323526	EU199126	NA	EU199145
Ditopellopsis sp.	CBS 121471	EU254763	EU255088	EU221936	EU219254
Flavignomonia rhoigena	**CFCC 53118**	**MK432674**	**MK429917**	NA	**MK578102**
	CFCC 53119	**MK432675**	**MK429918**	NA	**MK578103**
	CFCC 53120	**MK432676**	**MK429919**	NA	**MK578104**
Gnomonia gnomon	CBS 199.53	DQ491518	AF408361	EU221885	EU219295
	CBS 829.79	AY818957	AY818964	EU221905	NA
Gnomoniopsis alderdunensis	CBS 125680	GU320825	NA	NA	NA
Gnomoniopsis chamaemori	CBS 803.79	EU254808	EU255107	NA	NA
Gnomoniopsis racemula	AR 3892	EU254841	EU255122	EU221889	EU219241
Mamianiella coryli	BPI 877578	EU254862	NA	NA	NA
Marsupiomyces quercina	MFLUCC 13-0664	MF190116	MF190061	NA	NA
Marsupiomyces epidermoidea	MFLU 15-2921	NA	MF190058	NA	NA
Melanconis marginalis	CBS 109744	EU199197	AF408373	EU221991	EU219301
Neognomoniopsis quercina	CBS 145575	MK876399	MK876440	NA	NA
Occultocarpon ailaoshanense	LCM 524.01	JF779849	JF779853	NA	JF779856
	LCM 522.01	JF779848	JF779852	JF779862	JF779857
Ophiognomonia melanostyla	LCM 389.01	JF779850	JF779854	NA	JF779858
Ophiognomonia vasiljevae	AR 4298	EU254977	EU255162	EU221999	EU219331
Plagiostoma aesculi	AR 3640	EU254994	EU255164	NA	EU219269
Linospora capreae	CBS 372.69	NA	AF277143	NA	NA
Pleuroceras oregonense	AR 4333	EU255060	EU255196	EU221931	EU219313
Pleuroceras pleurostylum	CBS 906.79	EU255061	EU255197	EU221962	EU219311
Phragmoporthe conformis	AR 3632	NA	AF408377	NA	NA
Valsalnicola oxystoma	AR 5137	JX519561	NA	NA	NA
	AR 4833	JX519559	JX519563	NA	NA
Sirococcus tsugae	AR 4010	EF512478	EU255207	EU221928	EU219289
	CBS 119626	EU199203	EU199136	EF512534	EU199159
Synnemasporella aculeans	CFCC 52094	MG682086	MG682026	MG682066	MG682046
	CFCC 53123	**MK432679**	**MK429920**	**MK578148**	**MK578105**
	CFCC 53124	**MK432680**	**MK429921**	**MK578149**	**MK578106**
Synnemasporella aculeans	**CFCC 53125**	**MK432681**	**MK429922**	**MK578150**	**MK578107**
	CFCC 53126	**MK432682**	**MK429923**	**MK578151**	**MK578108**
	CFCC 53127	**MK432683**	**MK429924**	**MK578152**	**MK578109**
Synnemasporella toxicodendri	CFCC 52097	MG682089	MG682029	MG682069	MG682049
Tenuignomonia styracis	BPI 89278	NA	LC379288	LC379282	LC379294

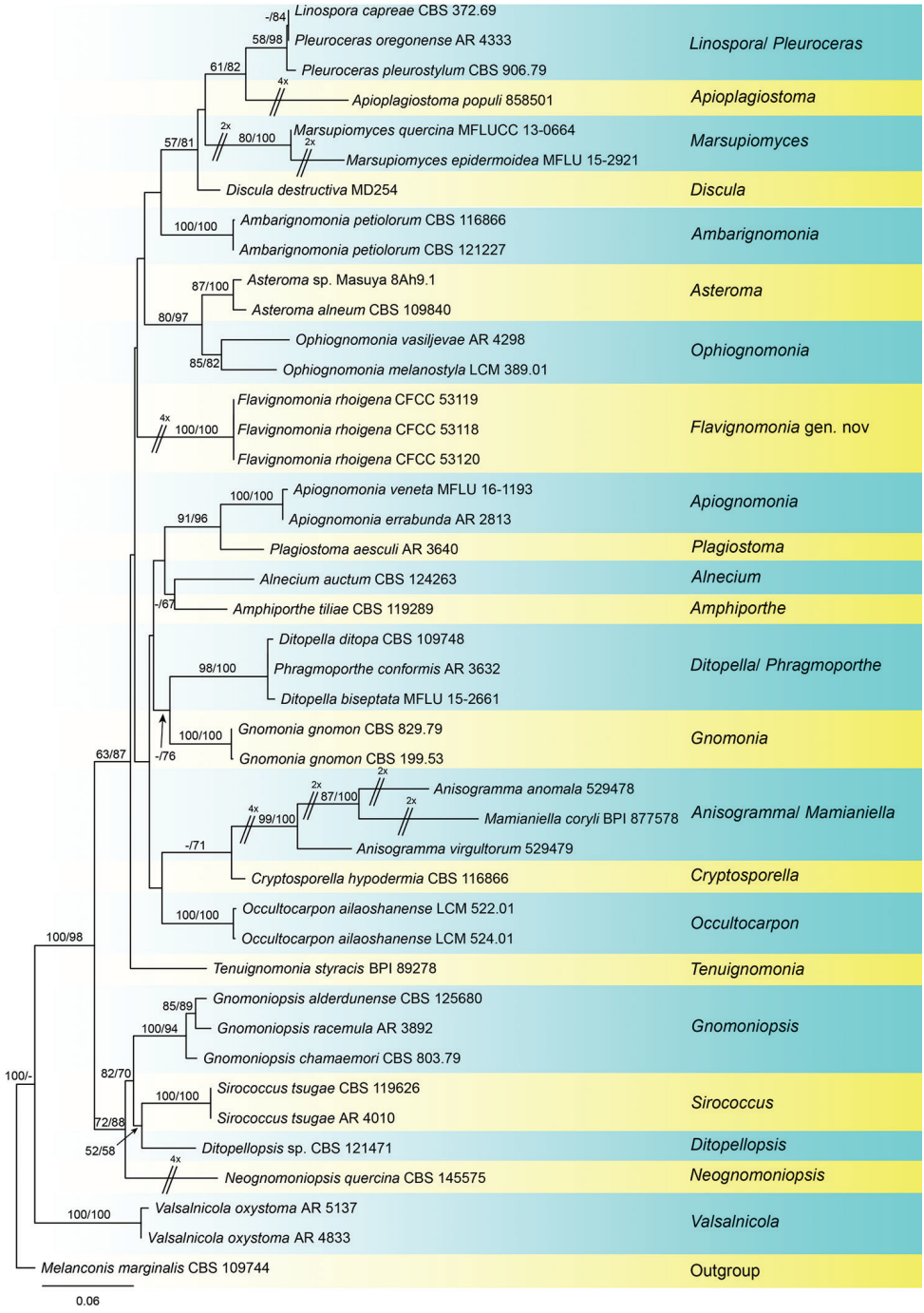

Figure 1. Phylogenetic tree based on an ML analysis of a combined DNA dataset of ITS, LSU, *tef1* and *rpb2* gene sequences for all genera with DNA data and some species of Gnomoniaceae. Bootstrap values ≥ 50 % for MP and ML analyses are presented at the branches. The scale bar represents the number of changes per site.

mata (Wehmeyer 1933, Fan et al. 2018). However, *Flavignomonia*, typified with *Flavignomonia rhoigena*, is distinguished from *Synnemasporella* species by its orange synnematal tips and hyaline conidia (Fan et al. 2018).

Flavignomonia rhoigena **C.M. Tian & Q. Yang, sp. nov.**
Figure 2
MycoBank No: 829531

Diagnosis. *Flavignomonia rhoigena* can be distinguished from other gnomoniaceous species by the formation of synnemata.

Etymology. Named after the host genus, *Rhus*.

Description. Sexual morph: not observed. Asexual morph: Conidiomata synnematal. Synnemata (650–)750–1100 µm high, 150–300 µm diam, determinate, growing from host tissue, with brown base and orange tip, straight to curved, parallel, with flat to

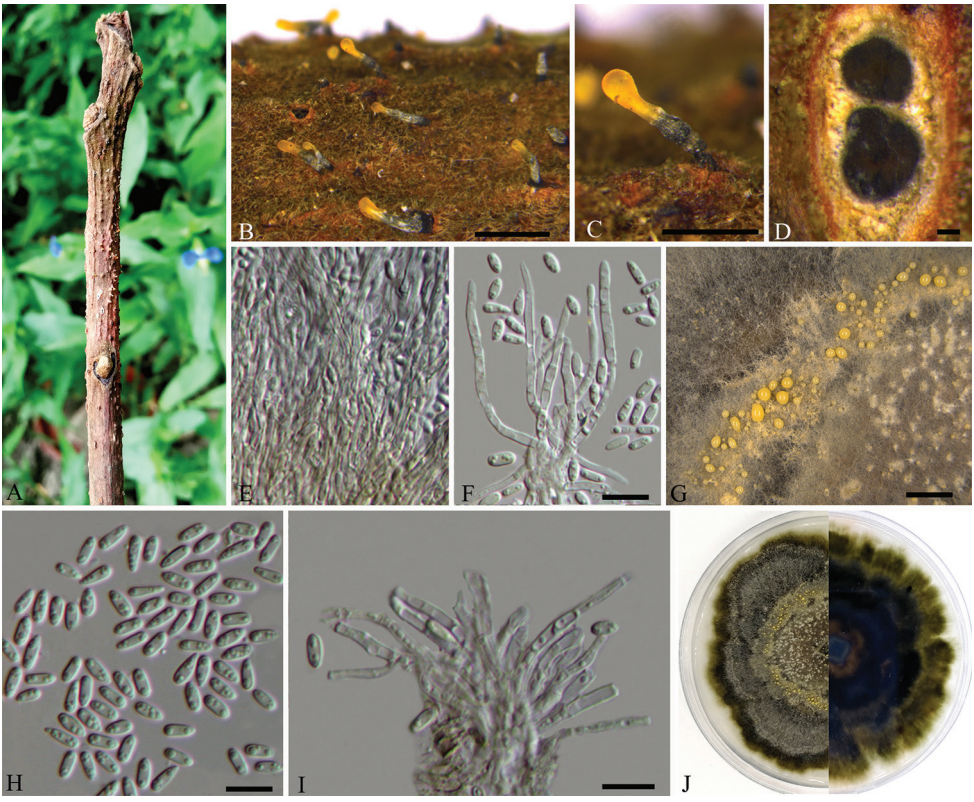

Figure 2. *Flavignomonia rhoigena* on *Rhus chinensis* (BJFC-S1766, holotype) **A–C** habit of conidiomata on twigs **D** transverse section through synnema **E** longitudinal section through synnema **F, I** conidiogenous cells attached with conidia **G** conidiomata on PDA **H** condia **J** the colony on PDA. Scale bars: 1 mm (**B**); 500 µm (**C**); 100 µm (**D**); 10 µm (**F, H–I**); 200 µm (**G**).

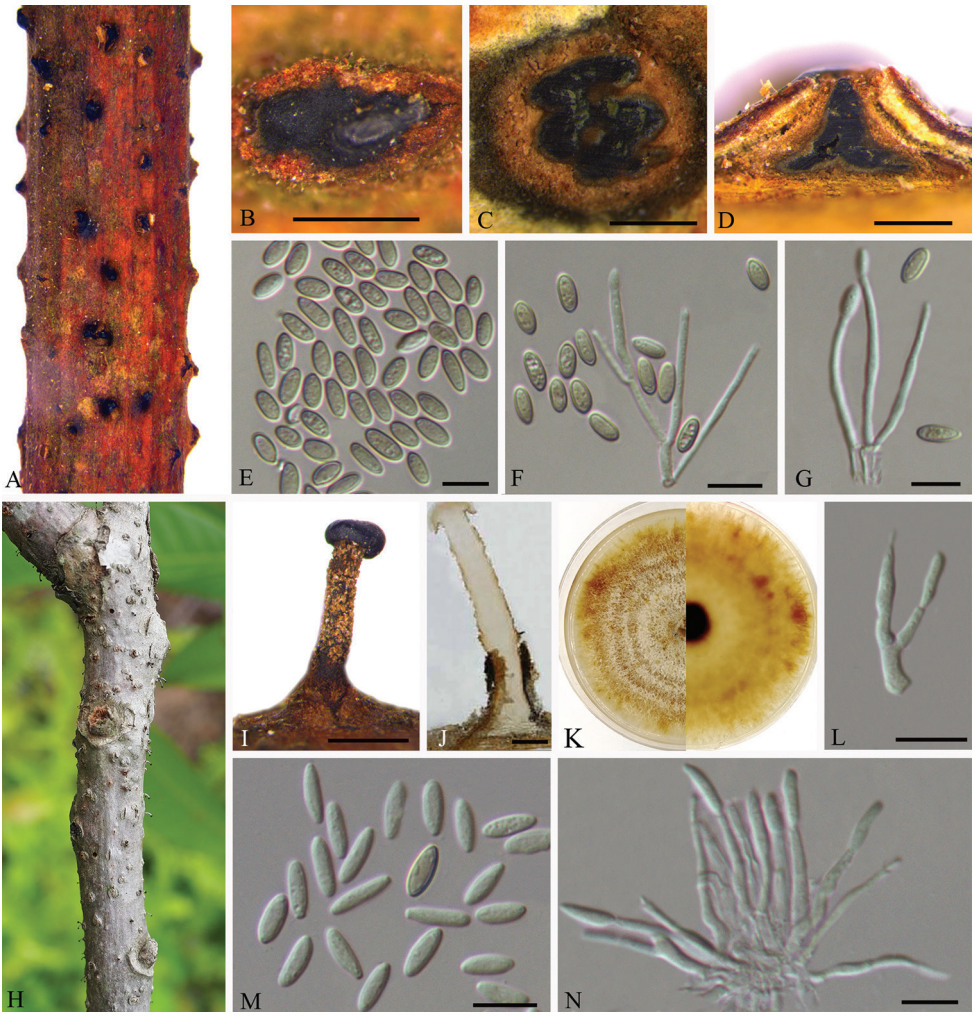

Figure 3. Asexual morphology of *Synnemasporella aculeans* on *Rhus chinensis* (BJFC-S1740) **A, B** habit of pycnidia on twigs **C** transverse section of pycnidium **D** longitudinal section through pycnidium **E** conidia **F, G** conidiogenous cells and conidia **H, I** habit of synnemata on twigs **J** longitudinal section through synnema **K** the colony on PDA **L, N** conidiogenous cells bearing conidia **M** conidia. Scale bars: 500 μm (**B–D, I, J**); 10 μm (**E–G, L–N**).

slightly concave and dark zone of conidiogenous cells and host tissue at their bases. Conidiophores reduced to conidiogenous cells. Conidiogenous cells (12.5–)16–22(–25) × 2 μm, phialidic, aggregated, hyaline, straight to curved, cylindrical, arranged adjacent to one another at the end of the synnema, producing a single conidium. Conidia cylindrical to oblong, smooth, multiguttulate, hyaline, (5–)5.5–7(–8) × 1.5–2 μm.

Culture characters. On PDA at 25 °C in darkness, initially white, becoming olivegreen to black after 3 wk, zonate with 3–4 well defined zones. Conidiomata distributed concentrically over agar surface.

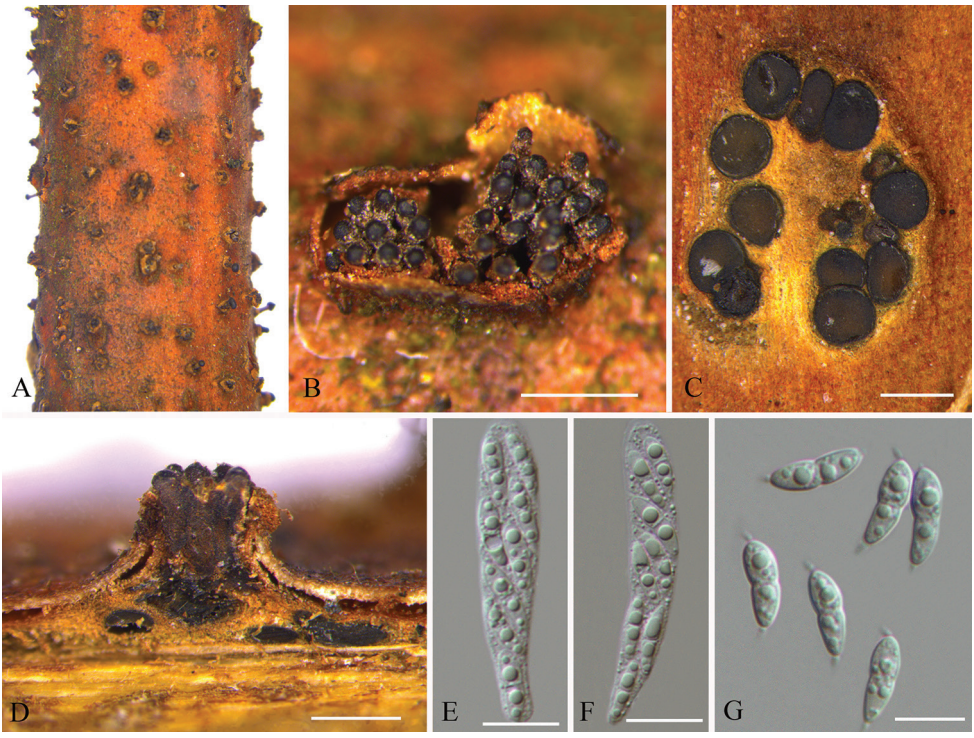

Figure 4. Sexual morphology of *Synnemasporella aculeans* on *Rhus chinensis* (BJFC-S1745) **A, B** habit of ascomata on twigs **C** transverse section of ascomata **D** longitudinal section through ascomata **E, F** asci **G** ascospores. Scale bars: 500 μm (**B–D**); 10 μm (**E–G**).

Specimen examined. CHINA, Jiangxi Province, Ganzhou City, Xunwu County, 24°52'31.34"N, 115°35'39.53"E, on branches of *Rhus chinensis*, 14 May 2018, Q. Yang, Y. Liu & Y.M. Liang (holotype BJFC-S1766, ex-type living cultures CFCC 53118, CFCC 53119 and CFCC 53120).

Notes. *Flavignomonia rhoigena* is the type species of *Flavignomonia* in the family Gnomoniaceae. It can be easily distinguished from the other gnomoniaceous genera by its unique conidiomata (Walker et al. 2004, Senanayake et al. 2018, Crous et al. 2019, Minoshima et al. 2019).

***Synnemasporella aculeans* (Schwein.) X.L. Fan & J.D.P. Bezerra, Persoonia 40: 130. 2018.**

Figure 3, 4

Description. Sexual morph: See Wehmeyer (1933) and Fan et al. (2018). Asexual morph: See Fan et al. (2018).

Specimens examined. CHINA, Jiangxi Province, Ganzhou City, Xunwu County, 24°52'31.34"N, 115°35'39.53"E, on branches of *Rhus chinensis*, 14 May 2018, Q. Yang, Y.

Liu & Y.M. Liang (BJFC-S1740, living culture CFCC 53123); Ganzhou City, Fengshan forest park, 25°44'32.14"N, 114°59'25.54"E, on branches of *Rhus chinensis*, 15 May 2018, Q. Yang, Y. Liu & Y.M. Liang (BJFC-S1753, living culture CFCC 53124 and CFCC 53125). 24°38'38.18"N, 115°33'58.45"E, on branches of *Rhus chinensis*, 16 May 2018, Q. Yang, Y. Liu & Y.M. Liang (BJFC-S1745, living culture CFCC 53126 and CFCC 53127).

Notes. *Synnemasporella aculeans* was proposed as a new combination in the new genus *Synnemasporella* based on the description of *Cryptodiaporthe aculeans* (Fan et al. 2018), which was introduced producing perithecial ascomata, and an asexual morph producing sporodochial and/or pycnidial conidiomata (Wehmeyer 1933). In the present study, five isolates from canker tissues on *Rhus chinensis* were congruent with *S. aculeans* based on morphology and DNA sequences data. This was the first time that the sexual morph of *Synnemasporella aculeans* in China had been collected.

Discussion

In this study, two diaporthalean species forming synnemata on *Rhus chinensis* were identified based on morphology and ITS, LSU, *tef1*, and *rpb2* sequence datasets. As a result, *Flavignomonia* typified with *F. rhoigena* is proposed as a new genus in Gnomoniaceae for its distinct phylogenic position and distinctive asexual fruiting body, Also, *Synnemasporella aculeans* strains were successfully isolated from perithecia, pycnidia and synnemata, which was confirmed by molecular data.

Nineteen fungal species have been recorded from the commercially and ecologically important tree species in China, including *Cladosporium cladosporioides*, *Cronartium quercuum*, *Mycosphaerella fushinoki*, *Pestalotiopsis diospyri*, *P. guepinii*, *P. mangiferae*, *P. sorbi*, *Phaeoramularia rhois*, *Phyllactinia corylea*, *Ph. rhoina*, *Pileolaria klugkistiana*, *Pi. shiraiana*, *Pseudocercospora rhoina*, *Ps. toxicodendri*, *Septoria* sp., *Tubercularia phyllophila*, *Uncinula verniciferae*, and two synnematal species from branch cankers in this study (Farr and Rossman 2019). *Flavignomonia rhoigena* and *Synnemasporella aculeans*, described and illustrated in the present study can be easily recognized by the asexual fruiting bodies, and they differ from each other in the colour of the synnematal tips.

Gnomoniaceae is a globally distributed fungal family on diverse plant hosts (Mejía et al. 2008, 2011a, 2011b, 2012, Sogonov et al. 2008, Walker et al. 2012, Senanayake et al. 2017, 2018). Host specificity of this family has been confirmed to be important in the evolution (Walker et al. 2014). Our newly discovered genus *Flavignomonia* was only found on *Rhus chinensis*, and more *Flavignomonia* species might be collected from the plant family Anacardiaceae in the future.

Acknowledgements

This study is financed by National Natural Science Foundation of China (Project No.: 31670647). We are grateful to Chungen Piao, Minwei Guo (China Forestry Culture Collection Center (CFCC), Chinese Academy of Forestry, Beijing.

References

Alves A, Crous PW, Correia A, Phillips AJL (2008) Morphological and molecular data reveal cryptic speciation in *Lasiodiplodia theobromae*. Fungal Diversity 28: 1–13.

Barr ME (1978) The Diaporthales in North America with emphasis on *Gnomonia* and its segregates. Mycologia Memoir 7: 1–232.

Carbone I, Kohn LM (1999) A method for designing primer sets for speciation studies in filamentous ascomycetes. Mycologia 3: 553–556. https://doi.org/10.2307/3761358

Crous PW, Carnegie AJ, Wingfield MJ, et al. (2019) Fungal Planet description sheets: 868–950. Persoonia 42: 291–473. https://doi.org/10.3767/persoonia.2019.42.11

Crous PW, Gams W, Stalpers JA, Robert V, Stegehuis G (2004) MycoBank: an online initiative to launch mycology into the 21st century. Studies in Mycology 50: 19–22.

Doyle JJ, Doyle JL (1990) Isolation of plant DNA from fresh tissue. Focus 12: 13–15. https://doi.org/10.2307/2419362

Fan XL, Bezerra JD, Tian CM, Crous PW (2018) Families and genera of diaporthalean fungi associated with canker and dieback of tree hosts. Persoonia 40: 119–134. https://doi.org/10.3767/persoonia.2018.40.05

Farr DF, Rossman AY (2019) Fungal Databases. US National Fungus Collections, ARS, USDA. https://nt.ars-grin.gov/fungaldatabases/ [Retrieved September 1, 2019]

Guindon S, Dufayard JF, Lefort V, Anisimova M, Hordijk W, Gascuel O (2010) New algorithms and methods to estimate maximum-likelihood phylogenies: assessing the performance of PhyML 3.0. Systematic Biology 59: 307–321. https://doi.org/10.1093/sysbio/syq010

Hillis DM, Bull JJ (1993) An empirical test of bootstrapping as a method for assessing confidence in phylogenetic analysis. Systematic Biology 42: 182–192. https://doi.org/10.1093/sysbio/42.2.182

Jaklitsch WM, Komon M, Kubicek CP, Druzhinina IS (2005) *Hypocrea voglmayrii* sp. nov. from the Austrian Alps represents a new phylogenetic clade in *Hypocrea*/*Trichoderma*. Mycologia 97: 1365–1378. https://doi.org/10.1080/15572536.2006.11832743

Jiang N, Fan XL, Tian CM (2019) Identification and pathogenicity of Cryphonectriaceae species associated with chestnut canker in China. Plant Pathology 68(6): 1132–1145. https://doi.org/10.1111/ppa.13033

Jiang N, Voglmayr H, Tian CM (2018) New species and records of *Coryneum* from China. Mycologia 110: 1172–1188. https://doi.org/10.1080/00275514.2018.1516969

Katoh K, Toh H (2010) Parallelization of the MAFFT multiple sequence alignment program. Bioinformatics 26: 1899–1900. https://doi.org/10.1093/bioinformatics/btq224

Mejía LC, Castlebury LA, Rossman AY, Sogonov MV, White JF (2008) Phylogenetic placement and taxonomic review of the genus *Cryptosporella* and its synonyms *Ophiovalsa* and *Winterella* (Gnomoniaceae, Diaporthales). Mycological Research 112(1): 23–35. https://doi.org/10.1016/j.mycres.2007.03.021

Mejía LC, Castlebury LA, Rossman AY, Sogonov MV, White JF (2011a) A systematic account of the genus *Plagiostoma* (Gnomoniaceae, Diaporthales) based on morphology, host-associations, and a four-gene phylogeny. Studies in Mycology 68: 211–235. https://doi.org/10.3114/sim.2011.68.10

Mejía LC, Rossman AY, Castlebury LA, White JF (2011b) New species, phylogeny, host-associations and geographic distribution of genus *Cryptosporella* (Gnomoniaceae, Diaporthales). Mycologia 103(2): 379–399. https://doi.org/10.3852/10-134

Mejía LC, Rossman AY, Castlebury LA, Yang ZL, White JF (2012) *Occultocarpon*, a new monotypic genus of Gnomoniaceae on *Alnus nepalensis* from China. Fungal Diversity 52(1): 99–105. https://doi.org/10.1007/s13225-011-0108-y

Minoshima A, Walker DM, Takemoto S, Hosoya T, Walker AK, Ishikawa S, Hirooka Y (2019) Pathogenicity and taxonomy of *Tenuignomonia styracis* gen. et sp. nov., a new monotypic genus of Gnomoniaceae on *Styrax obassia* in Japan. Mycoscience 60(1): 31–39. https://doi.org/10.1016/j.myc.2018.08.001

Rambaut A (2016) FigTree, version 1.4.3. University of Edinburgh, Edinburgh.

Rossman AY, Farr DF, Castlebury LA (2007) A review of the phylogeny and biology of the Diaporthales. Mycoscience 48: 135–144. https://doi.org/10.1007/S10267-007-0347-7

Senanayake IC, Crous PW, Groenewald JZ, Maharachchikumbura SSN, Jeewon R, Phillips AJL, Bhat JD, Perera RH, Li QR, Li WJ, Tangthirasunun N, Norphanphoun C, Karunarathna SC, Camporesi E, Manawasighe IS, Al-Sadi AM, Hyde KD (2017) Families of Diaporthales based on morphological and phylogenetic evidence. Studies in Mycology 86: 217–296. https://doi.org/10.1016/j.simyco.2017.07.003

Senanayake IC, Jeewon R, Chomnunti P, Wanasinghe DN, Norphanphoun C, Karunarathna A, Pem D, Perera RH, Camporesi E, McKenzie EHC, Hyde KD, Karunarathna SC (2018) Taxonomic circumscription of Diaporthales based on multigene phylogeny and morphology. Fungal Diversity 93: 241–443. https://doi.org/10.1007/s13225-018-0410-z

Sogonov MV, Castlebury LA, Rossman AY, Mejía LC, White JF (2008) Leaf-inhabiting genera of the Gnomoniaceae, Diaporthales. Studies in Mycology 62: 1–77. https://doi.org/10.3114/sim.2008.62.01

Swofford DL (2003) PAUP*: Phylogenetic Analyses Using Parsimony, * and Other Methods, Version 4.0b10. Sinauer Associates, Sunderland.

Vilgalys R, Hester M (1990) Rapid genetic identification and mapping of enzymatically amplified ribosomal DNA from several *Cryptococcus* species. Journal of Bacteriology 172: 4238–4246. https://doi.org/10.1128/jb.172.8.4238-4246.1990

Voglmayr H, Akulov OY, Jaklitsch WM (2016) Reassessment of *Allantonectria*, phylogenetic position of *Thyronectroidea*, and *Thyronectria caraganae* sp. nov. Mycological Progress 15: 921–937. https://doi.org/10.1007/s11557-016-1218-4

Voglmayr H, Castlebury LA, Jaklitsch WM (2017) *Juglanconis* gen. nov. on Juglandaceae, and the new family Juglanconidaceae (Diaporthales). Persoonia 38: 136–155. https://doi.org/10.3767/003158517X694768

Voglmayr H, Jaklitsch WM, Mohammadi H, Chakusary MK (2019) The genus *Juglanconis* (Diaporthales) on *Pterocarya*. Mycological Progress 18: 425–437. https://doi.org/10.1007/s11557-018-01464-0

Wang L, Wang N, Li T, Chen HZ (2014) Sumac (*Rhus chinensis* Mill) biomass refinery engineering. Chinese Journal of Biotechnology 30(5): 695–706. http://doi.org/10.13345/j.cjb.140058

Walker DM, Castlebury LA, Rossman AY, Mejía LC, White JF (2012) Phylogeny and taxonomy of *Ophiognomonia* (Gnomoniaceae, Diaporthales), including twenty-five new species in this highly diverse genus. Fungal Diversity 57(1): 85–147. https://doi.org/10.1007/s13225-012-0200-y

Walker DM, Castlebury LA, Rossman AY, Struwe L (2014) Host conservatism or host specialization? Patterns of fungal diversification are influenced by host plant specificity in *Ophiognomonia* (Gnomoniaceae: Diaporthales). Biological Journal of the Linnean Society 111(1): 1–6. https://doi.org/10.1111/bij.12189

Wehmeyer LE (1933) The genus *Diaporthe* Nitschke and its segregates. University of Michigan, USA. https://doi.org/10.1016/S0007-1536(33)80010-6

White TJ, Bruns T, Lee S, Taylor J (1990) Amplification and direct sequencing of fungal ribosomal RNA genes for phylogenetics. PCR Protocols: A Guide to Methods and Applications 18: 315–322. https://doi.org/10.1016/B978-0-12-372180-8.50042-1

Wijayawardene NN, Hyde KD, Lumbsch HT, Liu JK, Maharachchikumbura SSN, Ekanayaka AH, Tian Q, Phookamsak R (2018) Outline of Ascomycota: 2017. Fungal Diversity 88: 167–263. https://doi.org/10.1007/s13225-018-0394-8

Winter G (1886) Fungi Australienses. Revue Mycologique Toulouse 8: 207–213.

Yang Q, Fan XL, Guarnaccia V, Tian CM (2018) High diversity of *Diaporthe* species associated with dieback diseases in China, with twelve new species described. Mycokeys 39: 97–149. https://doi.org/10.3897/mycokeys.39.26914