MITOGENOME ANNOUNCEMENT

OPEN ACCESS Check for updates

Taylor & Francis

Taylor & Francis Group

The complete mitochondrial genome of a marine triclad *Miroplana shenzhensis* (Platyhelminthes, Tricladida, Maricola)

Jia-Jie Huang^a (D), Yuan-Yuan Liao^a (D), Wei-Xuan Li^b (D), Jun-Yu Li^a, An-Tai Wang^a and Yu Zhang^{a,c} (D)

^aShenzhen Key Laboratory of Marine Bioresource and Eco-environmental Science, College of Life Sciences and Oceanography, Shenzhen University, Shenzhen, Guangdong, P. R. China; ^bState Key Laboratory of Protein and Plant Gene Research, Center for Bioinformatics, School of Life Sciences, Peking University, Beijing, P. R. China; ^cGuangdong Engineering Research Center for Marine Algal Biotechnology, College of Life Sciences and Oceanography, Shenzhen University, Shenzhen, Guangdong, P. R. China

ABSTRACT

The complete mitochondrial genome (mitogenome) of *Miroplana shenzhensis* Yu & Wang, 2013 is reported in the present study, representing the second mitogenome recorded in the suborder Maricola. The circular mitogenome is 14,344 bp in length, containing 12 protein-coding genes, 2 ribosomal RNAs and 22 transfer RNAs. Comparative analysis on mitochondrial gene order reveals a rearrangement in the suborder Maricola, indicating that mitochondrial gene order is conserved only in Continenticola, and is divergent across Tricladida. Phylogenetic analysis shows *M. shenzhensis* is clustered with an another marine triclad, forming a well-supported monophyletic group of Maricolan.

ARTICLE HISTORY Received 19 October 2021 Accepted 12 May 2022

KEYWORDS Mitogenome; gene order; molecular phylogeny

Miroplana shenzhensis Yu & Wang, 2013 was classified into the genus Miroplana (Platyhelminthes, Tricladida, Maricola) according to its morphological features (Yu et al. 2013), while a more recent 18S and 28S rDNA based phylogenetic study demonstrated the phylogenetic position of the genus Miroplana (Li et al. 2019). Intriguingly, Miroplana exhibits good adaptation to both brackish water and freshwater habitats, which is similar to Sluysia triapertura (Souza et al. 2018) and species of the genus Paucumara (Li et al. 2021, Chen et al. 2019, Sluys 1989) and Pentacoelum (Sluys et al. 2015), but stands out of the majority of suborder Maricola species. Therefore, it will be important to further resolve the phylogenetic position of *M. shenzhensis* among triclad species using molecular markers other than 18S and 28S rDNA. However, complete mitogenome is only available for one marine triclad, namely Obrimoposthia wandeli (Yang et al. 2019). In this study, we present the mitogenome of M. shenzhensis, representing the second mitogenome belonging to the suborder Maricola.

The specimen of *M. shenzhensis* was collected on 20 May, 2018 from mangrove wetlands of Shenzhen Waterfront Ecological Park, Guangdong, China (22°52.27′N, 114°00.34′E). The genomic DNA was extracted by Li et al (2019) and currently deposited at College of Life Sciences and Oceanography, Shenzhen University (Zhang Yu, biozy@szu. edu.cn) under the voucher number SW001. We used the REPLI-g Midi Kit (QIAGEN, Hilden, Germany) to amplify the genomic DNA. Paired-end sequencing was conducted on the Illumina Hiseq 2500 platform (Novogene, Beijing, China). The mitogenome sequences were assembled using MitoFlex

v0.2.9 (Li et al. 2021). MITOS web server was used for gene annotation (Bernt et al. 2013), while the start and stop codons and the functional regions of the genes were verified by BLASTN (Altschul et al. 1997) with the guery sequences from the closely related species O. wandeli (Yang et al. 2019). Multiple sequences alignment (MSA) was performed using MACSE v2.03 (Ranwez et al. 2018). MSAs were subsequently trimmed using Gblocks v0.91b (Talavera and Castresana 2007). Substitution saturation test (Xia et al. 2003; Xia and Lemey 2009) for each protein-coding gene (PCG) was carried out in DAMBE6 (Xia 2017), while the third positions of all PCGs were excluded for downstream analysis due to nucleotide substitution saturation. Best-fit evolution model for each PCG was selected by PartitionFinder2 (Lanfear et al. 2017). The phylogenetic trees were constructed by Maximum Likelihood (ML) and Bayesian Inference (BI) methods, respectively. For ML, standard bootstrap analysis with 10,000 replications was performed by IQ-TREE v2.1.2 (Chernomor et al. 2016; Minh et al. 2020). While for BI, MrBayes v3.2.6 (Ronguist et al. 2012) was applied with 5,000,000 generations, sampling every 5,000 generations.

The circular mitogenome of *M. shenzhensis* is 14,344 bp in length and contains 12 PCGs, two rRNAs and 22 tRNAs. The nucleotide base composition is 28.7% A, 10.8% C, 15.8% G, and 44.7% T, with a total A + T content of 73.4%. *ATP8* gene, through automatic annotation, was noted as missing in the mitogenome of *M. shenzhensis. rrnL* is situated at 5' upstream of *rrnS* in the mitogenome of *M. shenzhensis* and *O. wandeli*, which is opposite to the unique arrangement of ribosomal genes in the suborder Continenticola (Solà et al. 2015).

© 2022 The Author(s). Published by Informa UK Limited, trading as Taylor & Francis Group.

CONTACT Yu Zhang 🔯 biozy@szu.edu.cn 🗈 College of Life Sciences and Oceanography, Shenzhen University, Shenzhen, Guangdong, P. R. China

This is an Open Access article distributed under the terms of the Creative Commons Attribution License (http://creativecommons.org/licenses/by/4.0/), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.



Figure 1. Bayesian inference phylogenetic tree topology inferred from the concatenated sequences of 12 PCGs. Nodal values indicate support values (bootstrap/ posterior probability).

Unexpectedly, except a few gene groups, namely *ND4-ND4L* and *ND3-ND2*, which are conserved in their arrangement in mitogenome, the PCG arrangement of *M. shenzhensis* differs from those of other known triclads. The current data show that the gene order is highly conserved across the suborder Continenticola while being divergent not only within Maricola, but also between Continenticola and Maricola. As such, our data provides a valuable addition to the previous perspective that gene order is strikingly conserved among Tricladida (cf. Ross et al. 2016).

The phylogeny was inferred from 12 concatenated PCG sequences of *M. shenzhensis* and 12 species of Tricladida, while two species of Polycladida were included as outgroups. The phylogenetic trees obtained from both BI and ML analysis share identical topologies, and being well-supported in most nodes (Figure 1). *M. shenzhensis* is more closely related to a marine triclad *O. wandeli* than other freshwater and terrestrial Continenticolans, forming a monophyletic group consists of Maricolans with high supporting values.

In conclusion, our study provides baseline information for future research regarding the origin and evolution in the suborder Maricola, suggesting the possible existence of the divergence of mitochondrial gene order in Tricladida. To better understand the phylogenetic relationships among species of Tricladida, it will be important to expand the mitogenome analysis within the order.

Acknowledgement

We thank Ms. Xiao-Yu Song and Ms. Ying Yang for their constructive comments on this work.

Ethical approval

This study does not involve Endangered or protected species according to IUZN (2021). The approval of sample collection is not required

according to the Animal Ethical and Welfare Committee of Shenzhen University (+86-755-86671906).

Disclosure statement

No potential conflict of interest was reported by the author(s).

Funding

This study was supported by Special Funds for the Cultivation of Guangdong College Students' Scientific and Technological Innovation ("Climbing Program" Special Funds; [grant no. pdjh2021b0429, pdjh2021b0430]; a China Undergraduate Training Program for Innovation and Entrepreneurship [grant no. S202010590038], as well as grants from Scientific and Technical Innovation Council of Shenzhen and Guangdong Natural Science Foundation [JCYJ20210324093412035 and 2020A1515011117].

Author contributions

YZ and ATW conceived the project. JJH, WXL and YZ designed the experiments. JJH, YYL and WXL collected the samples. JJH and YYL extracted the DNA. JJH, YYL and JYL performed the bioinformatics analysis. JJH and YZ drafted the paper. All of the authors have read and edited the paper and approved its submission.

ORCID

Jia-Jie Huang () http://orcid.org/0000-0001-9209-9754 Yuan-Yuan Liao () http://orcid.org/0000-0003-4002-7294 Wei-Xuan Li () http://orcid.org/0000-0001-8853-3589 Yu Zhang () http://orcid.org/0000-0001-7378-6946

Data availability statement

The genome sequence data that support the findings of this study are openly available in Genbank of NCBI at https://www.ncbi.nih.gov under the accession no. MW703985. The associated BioProject, SRA, and BioSample number are PRJNA777831, SRR16770661, and SAMN22883523 respectively.

References

- Altschul SF, Madden TL, Schaffer AA, Zhang JH, Zhang Z, Miller W, Lipman DJ. 1997. Gapped BLAST and PSI-BLAST: a new generation of protein database search programs. Nucleic Acids Res. 25(17):927–3402.
- Bernt M, Donath A, Juhling F, Externbrink F, Florentz C, Fritzsch G, Putz J, Middendorf M, Stadler PF. 2013. MITOS: improved de novo metazoan mitochondrial genome annotation. Mol Phylogenet Evol. 69(2):313–319.
- Chen JJ, Li WX, Sluys R, Wu MQ, Wang L, Li SF, Wang AT. 2019. Two new species of marine flatworm from southern China facilitate determination of the phylogenetic position of the genus *Nerpa* Marcus, 1948 and the histochemical structure of the nervous system in the genus *Paucumara* Sluys, 1989 (Platyhelminthes. Tricladida, Maricola). Zootaxa. 4568(1):zootaxa.4568.1.9–167.
- Chernomor O, Haeseler AV, Minh BQ. 2016. Terrace aware data structure for phylogenomic inference from supermatrices. Syst Biol. 65(6): 997–1008.
- Lanfear R, Frandsen PB, Wright AM, Senfeld T, Calcott B. 2017. PartitionFinder 2: new methods for selecting partitioned models of evolution for molecular and morphological phylogenetic Analyses. Mol Biol Evol. 34(3):772–773.
- Li JY, Li WX, Wang AT, Zhang Y. 2021. MitoFlex: an efficient, high-performance toolkit for animal mitogenome assembly, annotation and visualization. Bioinformatics. 37(18):3001–3003.
- Li M-Y, Ma X-Y, Li W-X, Yang Y, Sluys R, Chen J-J, Li S-F, Wang A-T. 2021. A new species of *Pacifides* from the Western Pacific Coast and the first fully freshwater species of the Maricolan planarian genus *Pacumara* (Platyhelminthes, Tricladida, Maricola. Syst Biodivers. 19(5):488–506.)
- Li W-X, Sluys R, Vila-Farré M, Chen J-J, Yang Y, Li S-F, Wang A-T. 2019. A new continent in the geographic distribution of the genus *oregoniplana* (Platyhelminthes: Tricladida: Maricola), its rediscovery in South Africa and its molecular phylogenetic position. Zool J Linn Soc. 187(1):82–99.
- Minh BQ, Schmidt HA, Chernomor O, Schrempf D, Woodhams MD, Haeseler A, Von Lanfear R. 2020. IQ-TREE 2: new models and efficient methods for phylogenetic inference in the genomic era. Mol Biol Evol. 37(5):1530–1534.
- Ranwez V, Douzery EJP, Cambon C, Chantret N, Delsuc F. 2018. MACSE v2: Toolkit for the alignment of coding sequences accounting for frameshifts and stop codons. Mol Biol Evol. 35(10):2582–2584.

- Ronquist F, Teslenko M, van der Mark P, Ayres DL, Darling A, Höhna S, Larget B, Liu L, Suchard MA, Huelsenbeck JP. 2012. MrBayes 3.2: efficient Bayesian phylogenetic inference and model choice across a large model space. Syst Biol. 61(3):539–542.
- Ross E, Blair D, Guerrero-Hernandez C, Alvarado AS. 2016. Comparative and transcriptome analyses uncover key aspects of coding- and long noncoding RNAs in flatworm mitochondrial genomes. G3 (Bethesda). 6(5):1191–1200.
- Sluys R. 1989. In: A. A. Balkema, editor. A monograph of the marine triclads. Rotterdam & Brookfield.
- Sluys R, Vila-Farré M, Álvarez-Presas M, Riutort M, Kawakatsu M, Tulp AS. 2015. The diet and distribution of *Pentacoelum kazukolinda* (Kawakatsu & Mitchell, 1984), a Maricolan planarian with a freshwater ecology. Zool Scr. 44(1):72–91.
- Solà E, álvarez-Presas M, Frías-López C, Littlewood DTJ, Rozas J, Riutort M. 2015. Evolutionary analysis of mitogenomes from parasitic and free-living flatworms. PLOS One. 10(3):e0120081.
- Souza S, Riutort M, Ferreira RL, Leal-Zanchet A. 2018. An integrative taxonomic approach reveals the first marine triclad (Platyhelminthes) trapped in a cave from a semiarid neotropical environment. Invert Systematics. 32(3):627–638.
- Talavera G, Castresana J. 2007. Improvement of phylogenies after removing divergent and ambiguously aligned blocks from protein sequence alignments. Syst Biol. 56(4):564–577.
- Xia X. 2017. DAMBE6: new tools for microbial genomics, phylogenetics, and molecular evolution. J Hered. 108(4):431–437.
- Xia X, Lemey P. 2009. Assessing substitution saturation with DAMBE. In: Lemey P, Salemi M, Vandamme A, editors. The phylogenetic handbook: a practical approach to phylogenetic analysis and hypothesis testing. Cambridge: University Press; p. 615–630.
- Xia X, Zheng X, Salemi M, Chen L, Wang Y. 2003. An index of substitution saturation and its application. Mol Phylogenet Evol. 26(1):1–7.
- Yang HM, Ji SJ, Min GS. 2019. The complete mitochondrial genome of the Antarctic marine triclad, *Obrimoposthia wandeli* (platyhelminthes, tricladida, maricola). Mitochondrial DNA B Resour. 4(2):2515–2516.
- Yu AQ, AT W, Lai XT. 2013. A first recorded family Uteriporidae with a new species of the genus *Miroplana* (Platyhelminthes, Tricladida) from China. Acta Zootaxonomica Sinica. 38(2):257–266.