

SCIENTIFIC REPORTS



OPEN

Photoelectrochemical detection of alpha-fetoprotein based on ZnO inverse opals structure electrodes modified by Ag₂S nanoparticles

Received: 02 August 2016
Accepted: 08 November 2016
Published: 06 December 2016

Yandong Jiang¹, Dali Liu¹, Yudan Yang², Ru Xu¹, Tianxiang Zhang¹, Kuang Sheng¹ & Hongwei Song¹

In this work, a new photoelectrochemical biosensor based on Ag₂S nanoparticles (NPs) modified macroporous ZnO inverse opals structure (IOs) was developed for sensitive and rapid detection of alpha fetal protein (AFP). Small size and uniformly dispersed Ag₂S NPs were prepared using the Successive Ionic Layer Adsorption And Reaction (SILAR) method, which were adsorbed on ZnO IOs surface and frame work as matrix for immobilization of AFP. The composite structure of ZnO/Ag₂S expanded the scope of light absorption to long wavelength, which can make full use of the light energy. Meanwhile, an effective matching of energy levels between the conduction bands of Ag₂S and ZnO are beneficial to the photo-generated electrons transfer. The biosensors based on FTO (fluorine-doped tin oxide) ZnO/Ag₂S electrode showed enough sensitivity and a wide linear range from 0.05 ng/mL to 200 ng/mL with a low detection limit of 8 pg/mL for the detection of AFP. It also exhibited high reproducibility, specificity and stability. The proposed method was potentially attractive for achieving excellent photoelectrochemical biosensor for detection of other proteins.

Primary liver cancer is known as malignant tumor, which is a serious threat to health and has a high mortality rate in the world^{1,2}. Fast and accurate early detection of cancer biomarker is vital for clinical diagnosis³, thus, specific biomarkers are highly needed⁴. AFP is an oncogenic glycoprotein which is normally expressed during gestation and originally identified in the human fetus in 1956⁵, but an elevated AFP concentration in adult plasma may be an early symptom of malignant tumor. AFP can act as the most important biomarkers in the diagnosis and targeting of liver cancer.

In the past few years, many efforts had been made to detect AFP, such as enzyme-linked immunosorbent assay^{6,7}, electrochemiluminescence⁸, fluorescence biosensor⁹, surface plasmon resonance immunoassays¹⁰ and electrochemical immunoassay^{11,12}. Although some results were obtained, sophisticated instruments, significant sample volume, limited sensitivity, and long detection time limited the clinical application¹³. To develop the clinical detection, a novel, highly sensitive and alternative detection method of AFP is desired.

Due to simple structure and easily operation allowing rapid, high-throughput biological assay, PEC immunosensors were widely used in the analytical methods^{14,15}. Immunochemical reactions at an electrode surface alter photocurrent generation and thus provide information about the respective biological process. Conventional immunoassays require antibody or antigen labelling with biomarkers for signal generation¹⁶. Enzyme-based PEC biosensors display high sensitivity, selectivity, simplicity, low cost, and minimal sample consumption^{17,18}, but the process of introducing the enzyme is complicated and enzyme inherent instability at the same time makes it easy inactivation in the external environment¹⁶. Therefore, to develop non-enzymatic biosensor with high sensitivity, stability and selectivity is the requirement of the science and technology¹⁹. The prominent advantage of PEC-based non-enzymatic biosensors is the possibility of utilizing photo-holes in the VB of a semiconductor to facilitate oxidation of chemical and/or biological components in a liquid or gas phase²⁰. Compared with enzyme-based PEC biosensing, non-enzymatic PEC detection is a more promising method, which need not the

¹State Key Laboratory on Integrated Optoelectronics, College of Electronic Science and Engineering, Jilin University, 2699 Qianjin Street, Changchun 130012, China. ²China-Japan Union Hospital of Jilin University, Changchun 130012, China. Correspondence and requests for materials should be addressed to D.L. (email: ldl@jlu.edu.cn) or H.S. (email: songhw@jlu.edu.cn)

sample to be labeled and has higher stability and durability against the external environment. Considerable effort has been invested in developing non-enzymatic PEC biosensors^{15,21}.

ZnO is one of the most extensively studied semiconductor oxides due to remarkable physical and chemical properties, presenting the most promising candidate in different applications, such as photocatalysis^{22,23}, solar cells^{24,25}, PEC water splitting^{26,27}, and sensing applications^{28,29}. Furthermore, its excellent thermal, chemical, low density, good biological compatibility and excellent photochemical stability^{30,31} make it attractive in PEC bioanalysis^{32,33}. Many ZnO nanostructures, such as nanorods²⁹, nanotubes³⁴, nanowires¹¹, nanosheets³⁵ and nanoflowers³⁶, have been applied in biosensor. Despite of this, the usage of ZnO without modification in PEC-based bioanalysis has some limitations because of its inherent wide band gap which results in a strong absorption in the UV region. It is noting that most biomolecules are very unstable under UV irradiation, the high activity of photo-holes produced in the VB of ZnO upon light illumination is disadvantageous to the biological molecules³⁷. The current problem for ZnO electrode is the efficient utilization of the visible light³⁸. As an important narrow band gap semiconductor material, Ag₂S has a large absorption coefficient and a direct band gap of Eg~1.1 eV³⁹, which has been successfully used for photocatalysis⁴⁰ and photovoltaic cells^{41,42}. Besides, Ag₂S possesses an ultra-low solubility product constant (Ksp = 6.3 × 10⁻⁵⁰), which guarantees that the least amount of Ag⁺ ion is released into the biological surroundings, Ag₂S possesses negligible toxicity compared to other commonly used narrow band gap materials^{23,43,44}, which is advantageous to bioanalysis. So far, little work was carried out to use Ag₂S NPs in PEC biosensor. It is a new method to composite ZnO with Ag₂S to improve visible absorption and promote the effective separation of photo-generated charges.

In this work, we report on the synthesis of Ag₂S NPs modified ZnO IOs photoelectrode used for immunosensor of AFP. The immunosensor with enhanced photocurrent intensity and less electron-hole recombination is desirable. As a 3D macropore structure, IOs possess a large surface area, which is advantageous to the electronic transmission and biomolecule immobilization³³. Coupling of Ag₂S with ZnO IOs could facilitate charges separation due to the quick electron transfer from the conduction band of the small band gap semiconductor to the conduction band of the large one⁴⁵. Our results showed that the photocurrent of the composite electrodes was significantly enhanced due to the formation of ZnO/Ag₂S composite electrodes. The electrodes also demonstrated good sensitivity and repeatability.

Results and Discussion

Characterizations of the FTO/ZnO/Ag₂S composited electrode. Figure 1A shows the fabrication procedure of the immunosensor. ZnO/Ag₂S hybrid modified electrodes were obtained by successive Ag⁺ and S²⁻ adsorption on ZnO IOs electrodes, which combines the excellent charge transport property with absorption property of the ZnO/Ag₂S⁴⁶. As a biocompatible material with high permeability, CS was fixed on FTO/ZnO/Ag₂S electrode for further immobilization of Anti-AFP antibody (Ab), then AFP was detected based on the specificity binding of antigen-antibody. Figure 1B shows photocurrent generation principle of ZnO/Ag₂S modified electrodes. Due to matching of energy levels between ZnO and Ag₂S, The loading of Ag₂S NPs can lead to more efficient light absorption and consequently increase the photocurrent response by more electron injection from the excited Ag₂S to the conduction band of ZnO.

ZnO IOs were fabricated by the sol-gel method according to our previous report with a slight modification (See in Supplementary information). Figure 2A shows the field emission scanning electron microscope (SEM) image of the surface morphology and microstructure of the synthesized ZnO IOs. ZnO IOs display an ordered pore structure of three dimensional space, with lattice constant of ~251 nm. Figure 2B shows that some Ag₂S NPs were deposited on frame work and outer surfaces after 3 SILAR cycles, and the average size of the Ag₂S NPs was about 15 nm. This indicates that the Ag⁻ and S²⁻ ions were easily diffused into the pores of the IOs without obvious aggregation and pore clogging to form Ag₂S nanocrystallites. In order to further verify compound ZnO/Ag₂S, the corresponding energy dispersive x-ray (EDX) spectrum of the FTO/ZnO/Ag₂S was carried out, as shown in Fig. 2C. The observed peaks for Zn, O, Ag and S further confirmed that the substance was composed of ZnO/Ag₂S. In order to investigate the structure of the ZnO/Ag₂S, the crystalline phases of the ZnO and ZnO/Ag₂S were characterized by XRD, as shown in Fig. 2D. The XRD pattern of ZnO showed good hexagonal matching (JCPDS, card no. 36-1451), no peaks of impurity were observed, indicating that the ZnO IOs sample was pure in hexagonal phase. After Ag₂S deposition, peaks corresponding to α-Ag₂S (JCPDS card no. 14-0072) were observed, further indicating the formation of ZnO IOs/Ag₂S NPs composites.

Optical and photoelectrochemical properties of the Ag₂S NPs modified ZnO IOs. Combining Ag₂S NPs with ZnO IOs could increase the optical absorption, accelerate charge separation and suppress photo-generated carriers recombination. In order to find the best cycles times of Ag₂S, electrodes with different cycles of Ag₂S coatings were studied. Figure 3A shows the UV-vis absorption spectra changes of ZnO/Ag₂S electrodes with various SILAR cycles. The ZnO IOs demonstrated photonic stop band (PSB) around 490 nm. The PSB in face centered cubic (fcc) photonic crystals could be described by Bragg's law of diffraction⁴⁷:

$$\lambda = \frac{2d_{hkl}}{m} \sqrt{n_{eff}^2 - \sin^2 \theta}, \quad (1)$$

where λ is the central wavelength of PSB, m is the order of the Bragg diffraction, d_{hkl} is the hkl plane distance, n_{eff} is the average refractive index, and θ is the angle from the incident light to the normal of the substrate surface. For the ZnO IOs, n_{eff} can be expressed as

$$n_{eff} = xn_{ZnO} + (1 - x)n_{air}, \quad (2)$$

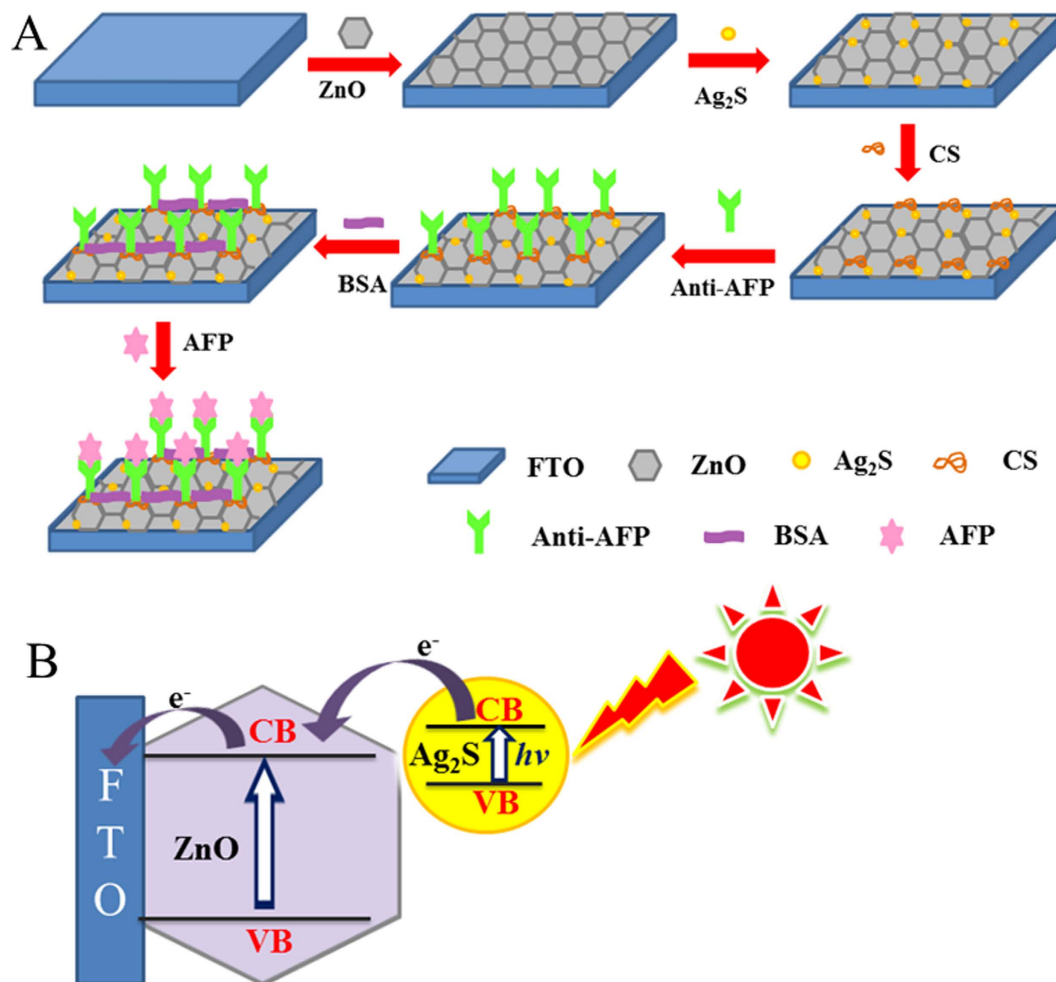


Figure 1. (A) The fabrication procedure of the immunosensor. (B) The photocurrent generation principle of ZnO/Ag₂S modified electrode.

where x is the volume ratio of ZnO IOs. Based on Eqs 1 and 2, x was deduced to be 0.22 ($n_{\text{ZnO}} = 1.9$), which was a little bit smaller than the ideal value (0.26). After the deposition of Ag₂S, the PSB of ZnO IOs was gradually covered, and the absorption in the visible and near-infrared range gradually increased. It should be noted that with increasing SILAR cycles, the color of the electrode changed from light yellow to brown, which indicated that the amount deposition of Ag₂S NPs on the ZnO IOs gradually increased, resulting in more light absorption³⁷. Figure 3B shows the photocurrent of ZnO/Ag₂S electrodes with various SILAR cycles. At the beginning, the photocurrent intensity increased with increasing SILAR cycles, and three SILAR cycles of ZnO/Ag₂S electrodes possessed the optimum, which was attributed to the improved light absorption due to Ag₂S loading. As the cycle number further increased, photocurrent gradually decreased, because effective surface area with the electrolyte solution decreased due to the excess deposition of Ag₂S, which blocked the pores of ZnO IOs. Besides, the extra Ag₂S increased the diffusion resistance to block electron transfer and offered more surface recombination centers^{33,48}. Thus, three cycle numbers of ZnO/Ag₂S electrodes were used in the following experience.

Characterizing the construction process of photoelectrochemical immunosensor.

Electrochemical impedance spectroscopy (EIS) was used to analyze the biosensor construction process which is a simple and useful tool for monitoring change of electrode. Figure 3C shows the Nyquist diagrams of electrodes fabricated in each step, with the frequency range 0.1 Hz–100 KHz in a KCl, K₃[Fe(CN)₆] and K₄[Fe(CN)₆] mixture solution. The electron transfer resistance (Ret) equals semicircle diameter. For the FTO/ZnO electrode, the impedance spectrum was obtained with a very small semicircle, indicating a very small Ret. After Ag₂S NPs was modified onto the FTO/ZnO electrode, the Ret increased owing to low conductivity of semiconductors. While CS, Ab, BSA and AFP were dropped on the electrodes step by step, Ret increased further correspondingly. This is because of insulating effect of organic molecules, which affects the electronic transfer to electrode surface. EIS indicated that the stepwise fabrication process of immunosensor was successfully designed. It should be noted that the Ret of immunosensor decreases with Xe-lamp illumination due to the improvement in the carrier concentration of the photoelectrodes⁴⁹.

The fabrication of the immunosensor through photocurrent can be also examined. Figure 3D shows photocurrent response of each step modified FTO/ZnO electrodes. Thanks to the sensitization effect of Ag₂S, the

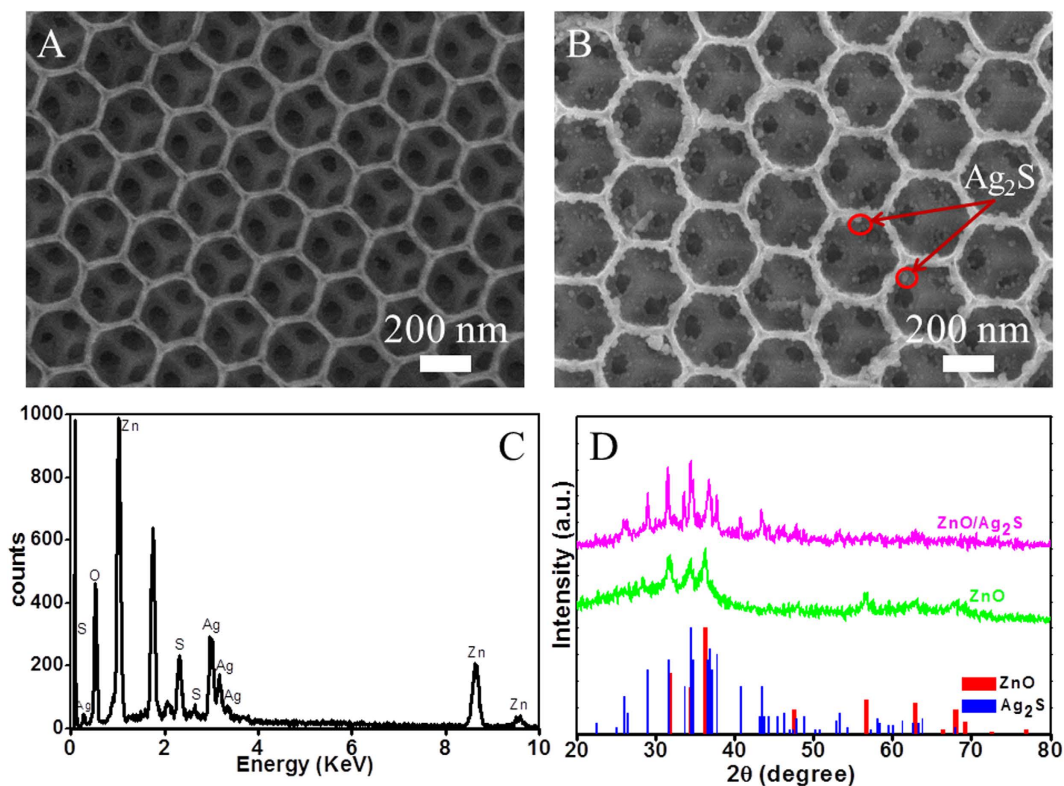


Figure 2. SEM image of ZnO IOs (A), ZnO modified with Ag₂S NPs (B), (C) EDX analysis of Ag₂S modified ZnO IOs of 3 SILAR cycle and (D) XRD patterns of ZnO IOs and ZnO/Ag₂S composited electrode.

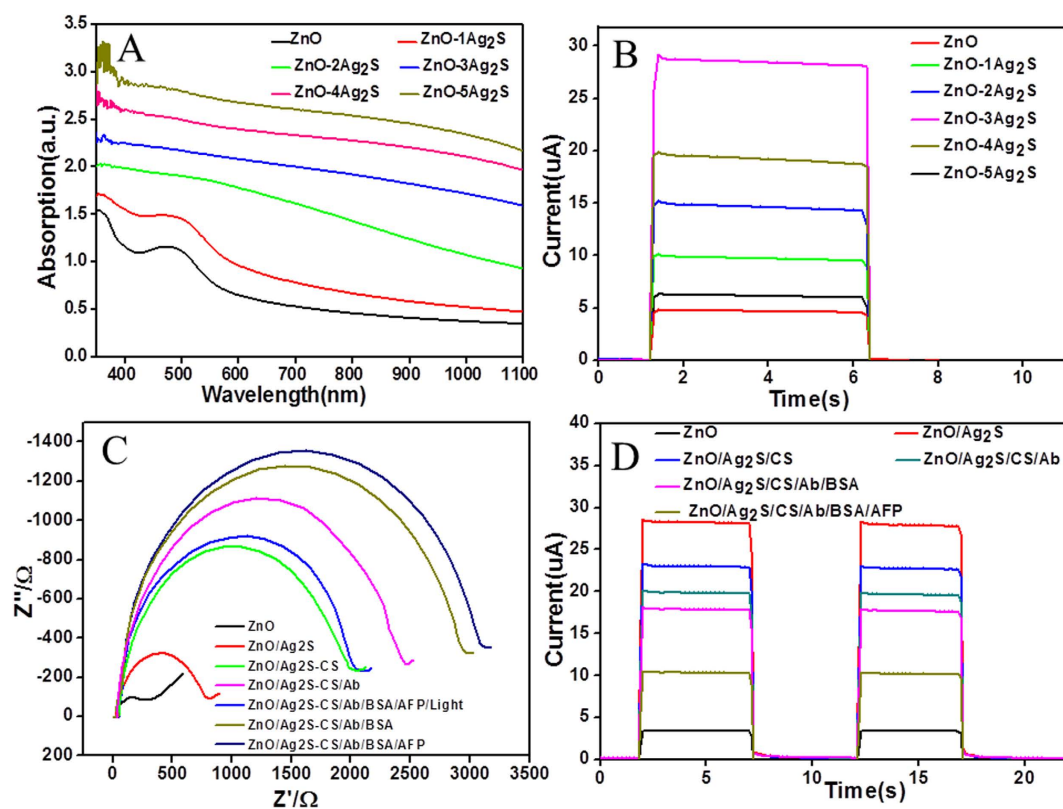


Figure 3. (A) Absorption spectra and (B) Photocurrent response of ZnO IOs electrode modified with different SILAR cycles of Ag₂S, (C) Electrochemical impedance Nyquist plot and (D) photocurrent response of modified ZnO IOs electrodes.

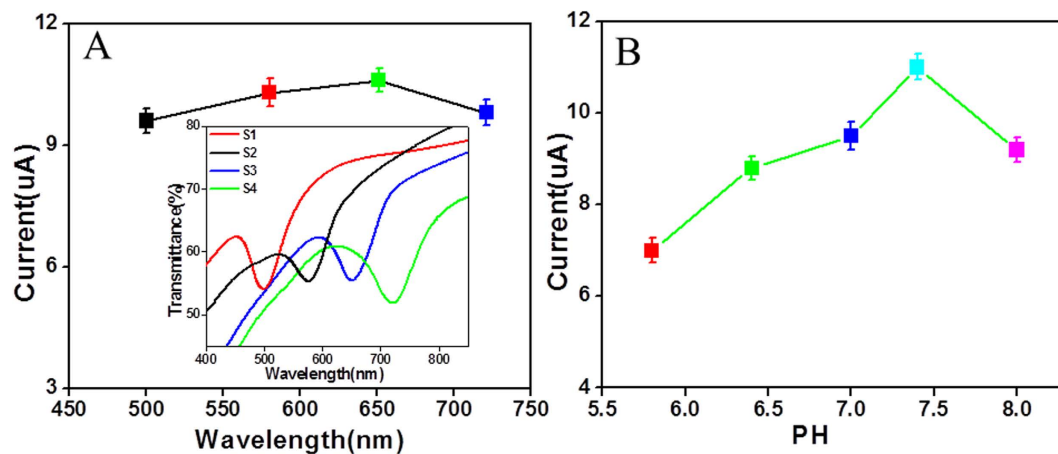


Figure 4. (A) The photocurrent of AFP detection (100 ng/ml) to different ZnO IOs electrodes (the inset figure is the transmission spectra of the four samples), (B) pH of the detection solution on the photocurrent response of the immunosensor toward 100 ng/ml AFP in 0.1 M PBS.

photocurrent significantly increased by 6.2 times than that of FTO/ZnO electrode after the deposition of Ag₂S NPs on the FTO/ZnO modified electrode⁴¹. After the successive immobilization of the CS, anti-AFP, BSA and AFP on the FTO/ZnO/Ag₂S modified electrode, the photocurrent intensity decreased. The fact is that the immobilization of these on the FTO/ZnO/Ag₂S modified electrodes hindered electronic transmission and increased steric hindrances in electrode/solution interface⁵⁰. Therefore a label-free photoelectrochemical immunosensor was achieved by monitoring the photocurrent change. Figure 1S displays cyclic voltammograms (CVs). When the FTO/ZnO electrode was modified by Ag₂S NPs, Current response increased over the FTO/ZnO IOs electrode due to the increased surface active sites. Then the CS was dropped on the FTO/ZnO/Ag₂S electrode, which could form an electron-blocking element and hinder the efficiency electron transfer resulting in current response decrease. After the electrode modification with Ab, BSA and AFP, the current further decreased. CVs also show the successful fabrication process of the electrode.

Effect of experimental conditions on photocurrent response. In order to find an effective macroporous structure, four electrodes with PSB at 500, 580, 652 nm and 721 nm were fabricated. The inset picture of Fig. 4A shows the transmission spectra of the four electrodes, showing the same good structure of IOs sample. The ZnO/Ag₂S composited electrodes were selected to detect AFP (100 ng/mL). The photocurrent response of composited electrodes to AFP with different PSB positions was shown in Fig. 4A. The photocurrents were 9.6, 10.3, 10.6 and 9.8 μA, respectively. The photocurrents had no obvious change with the location of PSB as we previously reported³³. Considering the stability of the electrode and the activity of Ab, AFP, the PH value in the process of biological detection is also very significant⁵¹. We tested the photocurrent response of different PH (5.8–8.0) detection solution in order to achieve the optimal effect. As shown in Fig. 4B, the photocurrent obtained at pH = 7.4 was optimal. Therefore, we used detection solution with a pH of 7.4 in the following experiments.

Photoelectrochemical detection of the immunosensor to AFP. Photoelectrochemical detection of the immunosensor to AFP was carried out under the optimal immunoassay conditions. Different concentrations AFP (15 uL) were immobilized on optimal electrode after blocking with BSA and the photocurrent responses were obtained. In order to check the influence of Ag₂S on the sensor performance, immunosensors based on ZnO and ZnO/Ag₂S composited electrodes were compared. Figure 5A shows the calibration curve of the developed ZnO IOs electrode and ZnO/Ag₂S composited electrode immunosensor used for the determination of the concentration of AFP. The photocurrent decrement was proportional to the logarithmic value of AFP concentration for ZnO/Ag₂S electrode. The regression equation was $\Delta I_1 = -1.60 \log C_{AFP} + 14.09$, ranging from 0.05 ng/mL to 200 ng/mL with a correlation coefficient of 0.999 and a low detection limit of 8 pg/ml. Here, ΔI_1 was the photocurrent of FTO/ZnO/Ag₂S/CS/anti-AFP/BSA electrode incubated with 15 μL different concentrations of AFP. FTO/ZnO/CS/anti-AFP/BSA electrode was incubated with 15 μL different concentrations of AFP to obtain ΔI_2 of ZnO IOs electrode, the regression equation was $\Delta I_2 = -0.327 \log C_{AFP} + 1.38$ for immunosensor without Ag₂S NPs in the range from 0.5 to 50 ng/mL. It is clear that ZnO/Ag₂S electrode showed a higher photocurrent, better linearity and sensitivity than that of ZnO electrode. The composited electrode has a wider linear range and lower detection limit than that of ZnO electrode, which was significant for the detection of AFP. The results showed that the performance of composite structure immunosensor was acceptable and promising. Compared with previous reports shown in Table 1, the proposed photoelectrochemical immunoassay exhibits enough sensitivity for the detection of AFP. Moreover, it also proved that the proposed label-free sensitization strategy of the detection biomarkers for early diagnosis and disease surveillance was particularly promising.

Reproducibility, specificity and stability of the immunosensor. The reproducibility of five immunosensors was evaluated towards to 100 ng/mL of AFP, and the relative standard deviation (RSD) of the five

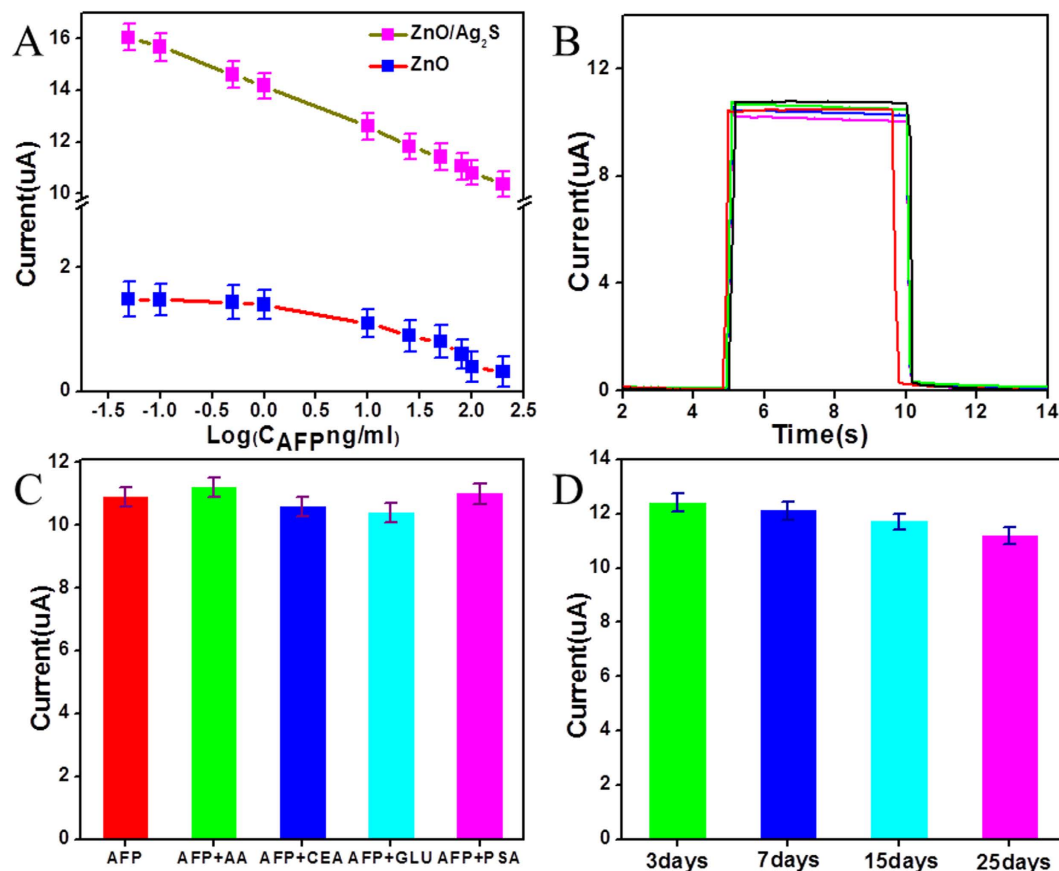


Figure 5. (A) The calibration curve of ZnO and ZnO/Ag₂S composited electrodes for different concentrations of AFP, (B) The reproducibility of the immunoassay by detecting 100 ng/mL AFP samples with five electrodes, (C) specificity of the immunoassay with 100 ng/mL of AFP without or with 500 ng/ml of (AA), 500 ng/ml of (CEA), 500 ng/ml of (GLU) and 500 ng/ml of (PSA), (D) The long-term stability of the immunosensor, detection in PBS solution (0.1 M, PH = 7.4) at potential of 0.6 V.

Method	Linear range [ng/mL]	Detection limit [pg/mL]	Ref.
Photoelectrochemical immunosensor	0.05–200	8	This work
Electrochemical immunosensor	0.05–150	20	11
Fluorescent immunosensor	0.025–5	12	9
Electrochemiluminescence immunosensor	0.5–600	480	8
Photoelectrochemical immunosensor	0.1–500	10	32

Table 1. Analytical performance of various methods for AFP immunoassays.

independent assay systems was 2.6%. As shown in Fig. 5B, no obvious changes could be found, showing good precision and acceptable reproducibility.

Specificity is vital to immunoassay, since the nonspecific adsorption can influence the sensitivity. To survey the photocurrent response originated from specific binding, the photocurrent of five electrodes with 100 ng/mL of AFP, without or with 500 ng/ml of ascorbic acid (AA), 500 ng/ml of carcinoembryonic antigen (CEA), 500 ng/ml of glucose (GLU) and 500 ng/ml of prostate specific antigen (PSA) was investigated, as seen in Fig. 5C. No obvious photocurrent change was observed, suggesting that the photocurrent responses arose from the interaction of AFP and anti-AFP, which were specific without much interference from nonspecific adsorption. The immunosensor possessed a satisfactory specificity.

The long-term storage stability of the immunosensor was investigated. When the immunosensor was stored in a refrigerator at 4 °C, photocurrent response was got in the detection of 10 ng/mL AFP after different storage time (3, 7, 15, 25 days), as shown in Fig. 5D. The immunosensor still remained more than 90% photocurrent after 25 days, which showed good long-term storage stability.

Conclusions

A simple and effective method was proposed to fabricate low-toxicity Ag₂S NPs modified ZnO electrodes. ZnO IOs composited with Ag₂S NPs could not only increase the effective utilization and absorption of light but also accelerate the electron transfer and restrain recombination of charge for ZnO/Ag₂S structure upon irradiation due to the effective matching of energy levels between ZnO and Ag₂S. The optimal cycle numbers of Ag₂S deposited on ZnO IOs were studied and the three cycles ZnO/Ag₂S composited electrode showed higher photocurrent response, wider linear range and lower detection limit. The designed immunosensor based on composited electrode for quantitative detection of AFP exhibited high sensitivity, good reproducibility, and long-term stability. This proposed photoelectrochemical method can be expanded readily for detecting other cancer biomarkers and pathogens.

References

1. Yuen, M. F. *et al.* Early detection of hepatocellular carcinoma increases the chance of treatment: Hong Kong experience. *Hepatology* **31**, 330–335 (2000).
2. Lee, J. U., Nguyen, A. H. & Sim, S. J. A nanoplasmonic biosensor for label-free multiplex detection of cancer biomarkers. *Biosensors & Bioelectronics* **74**, 341–346 (2015).
3. Jie, G.-F., Liu, P. & Zhang, S.-S. Highly enhanced electrochemiluminescence of novel gold/silica/CdSe-CdS nanostructures for ultrasensitive immunoassay of protein tumor marker. *Chemical Communications* **46**, 1323–1325 (2010).
4. Li, W., Jiang, X., Xue, J., Zhou, Z. & Zhou, J. Antibody modified gold nano-mushroom arrays for rapid detection of alpha-fetoprotein. *Biosensors & Bioelectronics* **68**, 468–474 (2015).
5. Sun, W. *et al.* AFP (alpha fetoprotein): Who are you in gastrology? *Cancer Letters* **357**, 43–46 (2015).
6. Zhao, B. *et al.* Carbon Nanotubes Multifunctionalized by Rolling Circle Amplification and Their Application for Highly Sensitive Detection of Cancer Markers. *Small* **9**, 2595–2601 (2013).
7. Nagasaki, Y., Kobayashi, H., Katsuyama, Y., Jomura, T. & Sakura, T. Enhanced immunoresponse of antibody/mixed-PEG co-immobilized surface construction of high-performance immunomagnetic ELISA system. *Journal Of Colloid And Interface Science* **309**, 524–530 (2007).
8. Zhang, J. X., Liu, S. L., Bao, J. C., Tu, W. W. & Dai, Z. H. Dual signal amplification of zinc oxide nanoparticles and quantum dots-functionalized zinc oxide nanoparticles for highly sensitive electrochemiluminescence immunosensing. *Analyst* **138**, 5396–5403 (2013).
9. Xie, Q. *et al.* A sensitive fluorescent sensor for quantification of alpha-fetoprotein based on immunosorbent assay and click chemistry. *Biosensors and Bioelectronics* **77**, 46–50 (2016).
10. Yang, X., Yu, Y. & Gao, Z. A Highly Sensitive Plasmonic DNA Assay Based on Triangular Silver Nanoprism Etching. *Acs Nano* **8**, 4902–4907 (2014).
11. Li, Q. *et al.* Wire-in-Tube IrOx Architectures: Alternative Label-Free Immunosensor for Amperometric Immunoassay toward alpha-Fetoprotein. *Acs Applied Materials & Interfaces* **7**, 22719–22726 (2015).
12. Li, L., Zhang, L., Yu, J., Ge, S. & Song, X. All-graphene composite materials for signal amplification toward ultrasensitive electrochemical immunosensing of tumor marker. *Biosensors & Bioelectronics* **71**, 108–114 (2015).
13. Wen, G. & Ju, H. Ultrasensitive photoelectrochemical immunoassay through tag induced exciton trapping. *Talanta* **134**, 496–500 (2015).
14. Zhao, W.-W., Xu, J.-J. & Chen, H.-Y. Photoelectrochemical bioanalysis: the state of the art. *Chemical Society Reviews* **44**, 729–741 (2015).
15. Wang, G.-L., Yu, P.-P., Xu, J.-J. & Chen, H.-Y. A Label-Free Photoelectrochemical Immunosensor Based on Water-Soluble CdS Quantum Dots. *Journal Of Physical Chemistry C* **113**, 11142–11148 (2009).
16. Devadoss, A., Sudhagar, P., Terashima, C., Nakata, K. & Fujishima, A. Photoelectrochemical biosensors: New insights into promising photoelectrodes and signal amplification strategies. *Journal Of Photochemistry And Photobiology C-Photochemistry Reviews* **24**, 43–63 (2015).
17. Zhao, W.-W. *et al.* A General Strategy for Photoelectrochemical Immunoassay Using an Enzyme Label Combined with a CdS Quantum Dot/TiO₂ Nanoparticle Composite Electrode. *Analytical Chemistry* **86**, 11513–11516 (2014).
18. Chen, D., Zhang, H., Li, X. & Li, J. Biofunctional Titania Nanotubes for Visible-Light-Activated Photoelectrochemical Biosensing. *Analytical Chemistry* **82**, 2253–2261 (2010).
19. Zhang, X. *et al.* WO₃ nanoparticles decorated core-shell TiC-C nanofiber arrays for high sensitive and non-enzymatic photoelectrochemical biosensing. *Chemical Communications* **49**, 7091–7093 (2013).
20. Fujishima, A. & Honda, K. Electrochemical photolysis of water at a semiconductor electrode. *Nature* **238**, 37–38 (1972).
21. Wang, Y. *et al.* A novel self-cleaning, non-enzymatic glucose sensor working under a very low applied potential based on a Pt nanoparticle-decorated TiO₂ nanotube array electrode. *Electrochimica Acta* **115**, 269–276 (2014).
22. Yang, J. L., An, S. J., Park, W. I., Yi, G. C. & Choi, W. Photocatalysis using ZnO thin films and nanoneedles grown by metal-organic chemical vapor deposition. *Advanced Materials* **16**, 1661–+ (2004).
23. Xu, T., Zhang, L., Cheng, H. & Zhu, Y. Significantly enhanced photocatalytic performance of ZnO via graphene hybridization and the mechanism study. *Applied Catalysis B-Environmental* **101**, 382–387 (2011).
24. Law, M., Greene, L. E., Johnson, J. C., Saykally, R. & Yang, P. D. Nanowire dye-sensitized solar cells. *Nature Materials* **4**, 455–459 (2005).
25. Sun, Y., Seo, J. H., Takacs, C. J., Seifert, J. & Heeger, A. J. Inverted Polymer Solar Cells Integrated with a Low-Temperature-Annealed Sol-Gel-Derived ZnO Film as an Electron Transport Layer. *Advanced Materials* **23**, 1679–+ (2011).
26. Yang, X. *et al.* Nitrogen-Doped ZnO Nanowire Arrays for Photoelectrochemical Water Splitting. *Nano Letters* **9**, 2331–2336 (2009).
27. Wang, G., Yang, X., Qian, F., Zhang, J. Z. & Li, Y. Double-Sided CdS and CdSe Quantum Dot Co-Sensitized ZnO Nanowire Arrays for Photoelectrochemical Hydrogen Generation. *Nano Letters* **10**, 1088–1092 (2010).
28. Liu, F. *et al.* Application of ZnO/graphene and S6 aptamers for sensitive photoelectrochemical detection of SK-BR-3 breast cancer cells based on a disposable indium tin oxide device. *Biosensors & Bioelectronics* **51**, 413–420 (2014).
29. Ge, S., Li, W., Yan, M., Song, X. & Yu, J. Photoelectrochemical detection of tumor markers based on a CdS quantum dot/ZnO nanorod/Au@Pt-paper electrode 3D origami immunodevice. *Journal Of Materials Chemistry B* **3**, 2426–2432 (2015).
30. Ozgur, U. *et al.* A comprehensive review of ZnO materials and devices. *Journal Of Applied Physics* **98** (2005).
31. Zhang, Y. *et al.* Scanning Probe Study on the Piezotronic Effect in ZnO Nanomaterials and Nanodevices. *Advanced Materials* **24**, 4647–4655 (2012).
32. Xu, R. *et al.* A sensitive photoelectrochemical biosensor for AFP detection based on ZnO inverse opal electrodes with signal amplification of CdS-QDs. *Biosensors & Bioelectronics* **74**, 411–417 (2015).
33. Xia, L. *et al.* Zinc oxide inverse opal electrodes modified by glucose oxidase for electrochemical and photoelectrochemical biosensor. *Biosensors & Bioelectronics* **59**, 350–357 (2014).
34. Vayssieres, L., Keis, K., Hagfeldt, A. & Lindquist, S. E. Three-dimensional array of highly oriented crystalline ZnO microtubes. *Chemistry Of Materials* **13**, 4395–+ (2001).

35. Li, W. *et al.* Stable Core/Shell CdTe/Mn-CdS Quantum Dots Sensitized Three-Dimensional, Macroporous ZnO Nanosheet Photoelectrode and Their Photoelectrochemical Properties. *Acs Applied Materials & Interfaces* **6**, 12353–12362 (2014).
36. Zhang, B., Lu, L., Hu, Q., Huang, F. & Lin, Z. ZnO nanoflower-based photoelectrochemical DNAzyme sensor for the detection of Pb²⁺. *Biosensors & Bioelectronics* **56**, 243–249 (2014).
37. Wang, G.-L., Xu, J.-J., Chen, H.-Y. & Fu, S.-Z. Label-free photoelectrochemical immunoassay for alpha-fetoprotein detection based on TiO₂/CdS hybrid. *Biosensors & Bioelectronics* **25**, 791–796 (2009).
38. Fan, G.-C., Ren, X.-L., Zhu, C., Zhang, J.-R. & Zhu, J.-J. A new signal amplification strategy of photoelectrochemical immunoassay for highly sensitive interleukin-6 detection based on TiO₂/CdS/CdSe dual co-sensitized structure. *Biosensors & Bioelectronics* **59**, 45–53 (2014).
39. Schaaff, T. G. & Rodinone, A. J. Preparation and Characterization of Silver Sulfide Nanocrystals Generated from Silver(I)-Thiolate Polymers. *The Journal of Physical Chemistry B* **107**, 10416–10422 (2003).
40. Zhao, Q. *et al.* Synthesis of biocompatible AuAgS/Ag₂S nanoclusters and their applications in photocatalysis and mercury detection. *Journal Of Nanoparticle Research* **16**, (2014).
41. Vogel, R., Hoyer, P. & Weller, H. Quantum-Sized PbS, CdS, Ag₂S, Sb₂S₃, and Bi₂S₃ Particles as Sensitizers for Various Nanoporous Wide-Bandgap Semiconductors. *The Journal of Physical Chemistry* **98**, 3183–3188 (1994).
42. Cheng, L., Ding, H., Chen, C. & Wang, N. Ag₂S/Bi₂S₃ co-sensitized TiO₂ nanorod arrays prepared on conductive glass as a photoanode for solar cells. *Journal Of Materials Science-Materials In Electronics* **27**, 3234–3239 (2016).
43. Zhang, X., Liu, M., Liu, H. & Zhang, S. Low-toxic Ag₂S quantum dots for photoelectrochemical detection glucose and cancer cells. *Biosensors & Bioelectronics* **56**, 307–312 (2014).
44. Maji, S. K., Sreejith, S., Mandal, A. K., Dutta, A. K. & Zhao, Y. Synthesis of Ag₂S quantum dots by a single-source precursor: an efficient electrode material for rapid detection of phenol. *Analytical Methods* **6**, 2059–2065 (2014).
45. Sant, P. A. & Kamat, P. V. Interparticle electron transfer between size-quantized CdS and TiO₂ semiconductor nanoclusters. *Physical Chemistry Chemical Physics* **4**, 198–203 (2002).
46. Khanchandani, S., Srivastava, P. K., Kumar, S., Ghosh, S. & Ganguli, A. K. Band Gap Engineering of ZnO using Core/Shell Morphology with Environmentally Benign Ag₂S Sensitizer for Efficient Light Harvesting and Enhanced Visible-Light Photocatalysis. *Inorganic Chemistry* **53**, 8902–8912 (2014).
47. Zhu, Y. *et al.* Inhibited Long-Scale Energy Transfer in Dysprosium Doped Yttrium Vanadate Inverse Opal. *Journal Of Physical Chemistry C* **116**, 2297–2302 (2012).
48. Li, W., Sheng, P., Cai, J., Feng, H. & Cai, Q. Highly sensitive and selective photoelectrochemical biosensor platform for polybrominated diphenyl ether detection using the quantum dots sensitized three-dimensional, macroporous ZnO nanosheet photoelectrode. *Biosensors & Bioelectronics* **61**, 209–214 (2014).
49. Bai, Z. *et al.* 3D-Branched ZnO/CdS Nanowire Arrays for Solar Water Splitting and the Service Safety Research. *Advanced Energy Materials* **6**, (2016).
50. Wang, G.-L., Shu, J.-X., Dong, Y.-M., Wu, X.-M. & Li, Z.-J. An ultrasensitive and universal photoelectrochemical immunoassay based on enzyme mimetics enhanced signal amplification. *Biosensors & Bioelectronics* **66**, 283–289 (2015).
51. Gao, J. *et al.* Ultrasensitive enzyme-free immunoassay for squamous cell carcinoma antigen using carbon supported Pd-Au as electrocatalytic labels. *Analytica Chimica Acta* **833**, 9–14 (2014).

Acknowledgements

This work was supported by NSFC (Grant Nos 61204015, 81301289, 61177042), The Jilin Province Natural Science Foundation of China (Nos 20150520090JH, 20140101171JC), Jilin Provincial Economic Structure Strategic Adjustment Fund Special Projects (No. 2014Y082).

Author Contributions

Yandong Jiang, Dali Liu, Hongwei Song wrote the main manuscript text and Yudan Yang, Ru Xu, Tianxiang Zhang, Kuang Sheng prepared Figures 2–4. All authors reviewed the manuscript.

Additional Information

Supplementary information accompanies this paper at <http://www.nature.com/srep>

Competing financial interests: The authors declare no competing financial interests.

How to cite this article: Jiang, Y. *et al.* Photoelectrochemical detection of alpha-fetoprotein based on ZnO inverse opals structure electrodes modified by Ag₂S nanoparticles. *Sci. Rep.* **6**, 38400; doi: 10.1038/srep38400 (2016).

Publisher's note: Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.



This work is licensed under a Creative Commons Attribution 4.0 International License. The images or other third party material in this article are included in the article's Creative Commons license, unless indicated otherwise in the credit line; if the material is not included under the Creative Commons license, users will need to obtain permission from the license holder to reproduce the material. To view a copy of this license, visit <http://creativecommons.org/licenses/by/4.0/>

© The Author(s) 2016