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#### CASE REPORT

# Vitamin D and COVID-19 in an immunocompromised patient with multiple comorbidities—A Case Report

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# Abstract

Routine 25-OH-Vitamin D3 measurement in COVID-19 patients could be of great importance, either for clinical course estimation or deciding on supplementation.

#### **KEYWORDS**

COVID-19, methotrexate, risk factors, SARS-CoV-2, severity, vitamin D3

# **1** | INTRODUCTION

We report mild COVID-19 manifestation in a high-risk patient with sufficient plasma 25-OH-Vitamin D3 level. Given the global pandemic of vitamin D deficiency, as well as its likely beneficial effects during SARS-CoV-2 infection, our report highlights importance of routine 25-OH-Vitamin D3 measurement, either for clinical course prediction or deciding on supplementation.

During the recent COVID-19 pandemic, several welldefined conditions have been recognized as risk factors worsening disease course and outcome.<sup>1,2</sup> The most important risk factors which can lead to severe COVID-19 symptomatology are obesity, diabetes mellitus, hypertension, age over 65 years, and immunosuppressive therapy.<sup>34</sup> These conditions are thought to affect host responses to infection by various mechanisms, enhancing, and accelerating the harmful pathophysiological processes.<sup>5</sup> Crucial events during COVID-19 pathogenesis are initial viral immune evasion and subsequent hyperinflammatory response resulting from excessive and undirected immune activation.<sup>6</sup> Patients with risk factors more likely react with extremely excessive systemic inflammation resulting with cytokine storm which is considered as an executive pathophysiological mechanism responsible for severe disease manifestations, such as acute respiratory distress syndrome (ARDS), diffuse endothelial damage with accelerated thrombogenesis, and multiple organ dysfunction syndrome (MODS).<sup>6</sup> In most groups of risk patients, common mechanism favoring development of excessive inflammation and cytokine storm is persistently present proinflammatory milieu.<sup>7</sup> Such chronic proinflammatory state can arise either due to hypersecretion of proinflammatory adipokines in obese patients,<sup>7</sup> lack of anti-inflammatory signaling in diabetic, and insulin-resistant patients<sup>7,8</sup> or dysregulation of renin-angiotensin system (RAS) with increased proinflammatory angiotensin II (Ang II) production in hypertensive patients.<sup>6,9,10</sup> Patients on immunosuppressive therapy are considered to be at risk due to the facilitated viral multiplication and dissemination during the initial phase, enabling in turn more vigorous ignition of hyperinflammatory phase.<sup>6,10,11</sup> In the other hand, hyperactive inflammatory processes could be kept under control to some extent by immunosuppressants, but conclusions and recommendations on their use in COVID-19 patients are inconsistent and unclear.<sup>12</sup> Moreover, most intriguing obscurity during COVID-19 pandemic-enormous variability of clinical presentation and unpredictability of outcome, either in previously healthy or in at-risk patients, still remains enigmatic.

In effort to prevent adverse outcomes of SARS-CoV-2 infection, growing attention is paid to potentially protective

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factors alleviating disease severity, among which vitamin D3 is currently being in the spotlight. Namely, many "precorona" studies have found antiviral, immunomodulatory, and anti-inflammatory properties of vitamin D3,<sup>13</sup> suggesting this data could be extrapolated to SARS-CoV-2 infection.<sup>14,15</sup> Assumptions on beneficial effects of vitamin D3 in COVID-19 patients have recently been verified by several convincing clinical and interventional studies.<sup>16-24</sup> The importance of this topic is highlighted by data showing that the global pandemic of hypovitaminosis D is being ongoing concomitantly with the COVID-19 pandemic.<sup>25-28</sup>

Here, we report mild clinical COVID-19 presentation in high-risk patient with the satisfactory vitamin D3 level upon ultraviolet B (UVB) phototherapy treatment, pointing to the vitamin D3 as a likely positive modulator of disease course, as well as to potential value of routine 25-OH-Vitamin D3 measurement in COVID-19 patients.

# 2 | CASE REPORT

A 66-year-old Caucasian man sought medical advice due to slightly elevated body temperature (up to 37.5°C), nausea without vomiting and occasional, mild, and dry cough during the last 2 days. He did not complain of dyspnea, shortness of breath, myalgias, or arthralgias during that period. He did not notice any loss of sense of smell or taste. He denied recent traveling or contacts with symptomatic persons. On admission, patient was alert, conscious, oriented, and independently movable and he was seeming to be in good general condition. Physical examination revealed pulse rate of 100 bpm, respiratory rate of 16 bpm, arterial blood pressure of 147/75 mm Hg, and axillary measured body temperature of 37.5°C. Chest auscultation disclosed clear breath sounds without obvious moist/dry rales or crackles, as well as normal heart sounds without audible murmurs. Abdomen was soft and painless on palpation, and no discomfort was elicited. Neither organomegaly nor palpable pathological masses were detected. The patient was obese, with a body weight of 99.5 kg and body height of 167 cm, resulting in a body mass index (BMI) of 35.7. He had been passionate smoker for a long period of time, and he stopped smoking 6 years ago. Before 25 years, patient has been diagnosed with psoriasis. Severe psoriatic arthritis has occurred 6 years ago and since that time he has been continuously receiving methotrexate at the dose of 15 mg/wk subcutaneously. Folic acid in the dose of 5 mg twice weekly was prescribed to prevent folate deficiency. Three months before the actual presentation, cutaneous psoriatic symptomatology exacerbated, and the patient was undergoing to UVB narrow band phototherapy lasting for two months.

Patient was diagnosed with arterial hypertension 8 years ago, and diagnosis of type 2 diabetes mellitus (T2DM) has

been established 6 years ago. To treat these conditions, he has been regularly taking losartan with hydrochlorothiazide (50 + 12.5 mg once a day) and metformin (500 mg three times a day).

The patient was referred to COVID-19 testing, and nasopharyngeal swab was taken. Detection of viral RNA by GeneXpert SARS-CoV-2 RT-PCR assay (Cepheid) resulted in a positive finding. Therefore, a chest X-ray was done and finding was unremarkable, without any opacity indicating infiltration or consolidation. Blood oxygen saturation (SpO2) measured by pulse oximeter was 95%. Subsequent routine laboratory tests found slightly elevated C-reactive protein (7 mg/L); increased erythrocyte sedimentation rate (50 mm/hr); normal white blood cell count ( $4.8 \times 10^9/L$ ) with lymphopenia  $(1.02 \times 10^9/L)$ ; decreased red blood cell count  $(4.06 \times 10^{12}/L)$ , hematocrit (0.387), and hemoglobin concentration (132 g/L); hyperglycemia (10.1 mmol/L), hypercreatininemia (134 µL/L), and hypertriglyceridemia (1.8 mmol/L). Semiquantitative dipstick urinalysis indicated mild proteinuria (1+). Further laboratory investigations revealed normal plasma 25-OH-vitamin D3 concentration of 92.2 nmol/L (normal range 50.0 - 125.0 nmol/L). All laboratory parameters, alongside with corresponding normal ranges, are shown in the Table 1.

Considering the mild clinical picture, no therapy was introduced, and all previous medications were continued. Given the occupancy of hospital facilities with severe cases, the patient was monitored on an outpatient basis. His condition did not worse and presenting symptoms completely subsided within 3 days. Twenty days after the initial presentation, SARS-CoV-2 RT-PCR test was negative, and serological enzyme-linked fluorescence assay (ELFA) showed presence of SARS-CoV-2 IgG (33.16) and IgM (5.09) in the patient's serum. At this time point, laboratory findings on complete blood count, blood differential test, and inflammatory indicators were within normal ranges, except for erythrocyte sedimentation rate, which had been often found to be elevated in patient even before infection given his underlying chronic inflammatory diseases. All laboratory findings obtained twenty days after the initial presentation (twentythree days after the symptoms onset) are given in the Table 2. During the next month, the patient felt well, not complaining of fatigue or exhaustion, and dermatological, rheumatological, and internistic symptoms remained stationary.

# **3** | **DISCUSSION**

Almost immediately after the pandemic outbreak in China, greater vulnerability and higher incidence of severe clinical presentation and fatal outcome were observed in patients with particular preexisting conditions, especially in persons aged >65.<sup>1,2</sup> Numerous studies have shown that the greatest

**TABLE 1** Laboratory findings upon detection of SARS-CoV-2 positivity (3 days after the symptoms appeared = median time of symptoms duration)

Total white blood cell (×10 <sup>9</sup> /L)     4.8     3.4-9.7       Lymphocytes (×10 <sup>9</sup> /L)     1.02     1.19-3.35       Neutrophils (×10 <sup>9</sup> /L)     3.10     2.06-6.49       Monocytes (×10 <sup>9</sup> /L)     0.50     0.12-0.84       Eosinophils (×10 <sup>9</sup> /L)     0.15     0.00-0.43       Basophils (×10 <sup>9</sup> /L)     0.04     0.00-0.06       Red blood cell (×10 <sup>12</sup> /L)     4.06     4.34-5.72       Hematocrit (%)     0.387     0.415-0.530       Hemoglobin (g/L)     132     138-175       MCV (fL)     95.3     83.0-97.2       MCHC (g/L)     341     320-345       Platelets (×10 <sup>9</sup> /L)     240     158-424       MPV (fL)     9.7     6.8-10.4       Prothrombin time     0.96     >0.7       Prothrombin time-INR     1.0     0.8-1.1       C-reactive protein (mg/L)     7     0-5       Erythrocyte sedimentation rate-1 h (mm/3.6 ks)     3-23       Glucose (mmol/L)     1.8     <1.7       Total cholesterol (mmol/L)     3.6     <5.0       LDL cholesterol (mmol/L)     1.1     >1.0       Urea (carbamide) (mmol/L)     1.34     64-104	Parameter (unit)	Result	Reference range
Neutrophils (×10 <sup>9</sup> /L)     3.10     2.06-6.49       Monocytes (×10 <sup>9</sup> /L)     0.50     0.12-0.84       Eosinophils (×10 <sup>9</sup> /L)     0.04     0.00-0.43       Basophils (×10 <sup>9</sup> /L)     0.04     0.00-0.06       Red blood cell (×10 <sup>12</sup> /L)     4.06     4.34-5.72       Hematocrit (%)     0.387     0.415-0.530       Hemoglobin (g/L)     132     138-175       MCV (fL)     95.3     83.0-97.2       MCH (pg)     32.5     27.4-33.9       MCHC (g/L)     341     320-345       Platelets (×10 <sup>9</sup> /L)     240     158-424       MPV (fL)     9.7     6.8-10.4       Prothrombin time     0.96     >0.7       Prothrombin time-INR     1.0     0.8-1.1       C-reactive protein (mg/L)     7     0-5       Erythrocyte sedimentation rate-1 h (mm/3.6 ks)     50     3-23       Glucose (mmol/L)     1.8     <1.7		4.8	3.4-9.7
Monocytes (×10 <sup>9</sup> /L)       0.50       0.12-0.84         Eosinophils (×10 <sup>9</sup> /L)       0.15       0.00-0.43         Basophils (×10 <sup>9</sup> /L)       0.04       0.00-0.06         Red blood cell (×10 <sup>12</sup> /L)       4.06       4.34-5.72         Hematocrit (%)       0.387       0.415-0.530         Hemoglobin (g/L)       132       138-175         MCV (fL)       95.3       83.0-97.2         MCH (pg)       32.5       27.4-33.9         MCHC (g/L)       341       320-345         Platelets (×10 <sup>9</sup> /L)       240       158-424         MPV (fL)       9.7       6.8-10.4         Prothrombin time       0.96       >0.7         Prothrombin time-INR       1.0       0.8-1.1         C-reactive protein (mg/L)       7       0-5         Erythrocyte sedimentation rate-1 h (mm/3.6 ks)       50       3-23         Glucose (mmol/L)       1.8       <1.7	Lymphocytes (×10 <sup>9</sup> /L)	1.02	1.19-3.35
Eosinophils (×10 <sup>9</sup> /L)     0.15     0.00-0.43       Basophils (×10 <sup>9</sup> /L)     0.04     0.00-0.06       Red blood cell (×10 <sup>12</sup> /L)     4.06     4.34-5.72       Hematocrit (%)     0.387     0.415-0.530       Hemoglobin (g/L)     132     138-175       MCV (fL)     95.3     83.0-97.2       MCH (pg)     32.5     27.4-33.9       MCHC (g/L)     341     320-345       Platelets (×10 <sup>9</sup> /L)     240     158-424       MPV (fL)     9.7     6.8-10.4       Prothrombin time     0.96     >0.7       Prothrombin time-INR     1.0     0.8-1.1       C-reactive protein (mg/L)     7     0-5       Erythrocyte sedimentation rate-1 h (mm/3.6 ks)     50     3-23       Glucose (mmol/L)     1.8     <1.7	Neutrophils (×10 <sup>9</sup> /L)	3.10	2.06-6.49
Basophils (×10 <sup>9</sup> /L)       0.04       0.00-0.06         Red blood cell (×10 <sup>12</sup> /L)       4.06       4.34-5.72         Hematocrit (%)       0.387       0.415-0.530         Hemoglobin (g/L)       132       138-175         MCV (fL)       95.3       83.0-97.2         MCH (pg)       32.5       27.4-33.9         MCHC (g/L)       341       320-345         Platelets (×10 <sup>9</sup> /L)       240       158-424         MPV (fL)       9.7       6.8-10.4         Prothrombin time       0.96       >0.7         Prothrombin time-INR       1.0       0.8-1.1         C-reactive protein (mg/L)       7       0-5         Erythrocyte sedimentation rate-1 h (mm/3.6 ks)       50       3-23         Glucose (mmol/L)       1.8       <1.7	Monocytes (×10 <sup>9</sup> /L)	0.50	0.12-0.84
Red blood cell (×10 <sup>12</sup> /L)     4.06     4.34-5.72       Hematocrit (%)     0.387     0.415-0.530       Hemoglobin (g/L)     132     138-175       MCV (fL)     95.3     83.0-97.2       MCH (pg)     32.5     27.4-33.9       MCHC (g/L)     341     320-345       Platelets (×10°/L)     240     158-424       MPV (fL)     9.7     6.8-10.4       Prothrombin time     0.96     >0.7       Prothrombin time-INR     1.0     0.8-1.1       C-reactive protein (mg/L)     7     0-5       Frythrocyte sedimentation rate-1 h (mm/3.6 ks)     50     3-23       Glucose (mmol/L)     1.8     <1.7	Eosinophils (×10 <sup>9</sup> /L)	0.15	0.00-0.43
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Hemoglobin (g/L)132138-175MCV (fL)95.383.0-97.2MCH (pg)32.527.4-33.9MCHC (g/L)341320-345Platelets (x10°/L)240158-424MPV (fL)9.76.8-10.4Prothrombin time0.96>0.7Prothrombin time-INR1.00.8-1.1C-reactive protein (mg/L)70-5Erythrocyte sedimentation rate-1 h (mm/3.6 ks)503-23Glucose (mmol/L)1.04.4-6.4Triglycerides (mmol/L)3.6<5.0	Red blood cell ( $\times 10^{12}/L$ )	4.06	4.34-5.72
MCV (fL)95.383.0-97.2MCH (pg)32.527.4-33.9MCHC (g/L)341320-345Platelets (×10 <sup>9</sup> /L)240158-424MPV (fL)9.76.8-10.4Prothrombin time0.96>0.7Prothrombin time-INR1.00.8-1.1C-reactive protein (mg/L)70-5Erythrocyte sedimentation rate-1 h (mm/3.6 ks)503-23Glucose (mmol/L)10.14.4-6.4Triglycerides (mmol/L)3.6<5.0	Hematocrit (%)	0.387	0.415-0.530
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MPV (fL)     9.7     6.8-10.4       Prothrombin time     0.96     >0.7       Prothrombin time-INR     1.0     0.8-1.1       C-reactive protein (mg/L)     7     0-5       Erythrocyte sedimentation rate-1 h (mm/3.6 ks)     50     3-23       Glucose (mmol/L)     10.1     4.4-6.4       Triglycerides (mmol/L)     1.8     <1.7	MCHC (g/L)	341	320-345
Prothrombin time     0.96     >0.7       Prothrombin time-INR     1.0     0.8-1.1       C-reactive protein (mg/L)     7     0-5       Erythrocyte sedimentation rate-1 h (mm/3.6 ks)     50     3-23       Glucose (mmol/L)     10.1     4.4-6.4       Triglycerides (mmol/L)     1.8     <1.7	Platelets ( $\times 10^9$ /L)	240	158-424
Prothrombin time-INR     1.0     0.8-1.1       C-reactive protein (mg/L)     7     0-5       Erythrocyte sedimentation rate-1 h (mm/3.6 ks)     50     3-23       Glucose (mmol/L)     10.1     4.4-6.4       Triglycerides (mmol/L)     1.8     <1.7	MPV (fL)	9.7	6.8-10.4
C-reactive protein (mg/L)     7     0-5       Erythrocyte sedimentation rate-1 h (mm/3.6 ks)     50     3-23       Glucose (mmol/L)     10.1     4.4-6.4       Triglycerides (mmol/L)     1.8     <1.7	Prothrombin time	0.96	>0.7
Erythrocyte sedimentation rate-1 h (mm/3.6 ks)     50     3-23       Glucose (mmol/L)     10.1     4.4-6.4       Triglycerides (mmol/L)     1.8     <1.7	Prothrombin time-INR	1.0	0.8-1.1
rate-1 h (mm/3.6 ks)       Glucose (mmol/L)     10.1     4.4-6.4       Triglycerides (mmol/L)     1.8     <1.7	C-reactive protein (mg/L)	7	0-5
Triglycerides (mmol/L)     1.8     <1.7		50	3-23
Total cholesterol (mmol/L)     3.6     <5.0	Glucose (mmol/L)	10.1	4.4-6.4
LDL cholesterol (mmol/L) 2.1 <3.0 HDL cholesterol (mmol/L) 1.1 >1.0 Urea (carbamide) (mmol/L) 7.5 2.8-8.3 Creatinine (µmol/L) 134 64-104 Total bilirubin (µmol/L) 10 3-20 25-OH-vitamin D3 (nmol/L) 92.2 50.0-125.0	Triglycerides (mmol/L)	1.8	<1.7
HDL cholesterol (mmol/L)     1.1     >1.0       Urea (carbamide) (mmol/L)     7.5     2.8-8.3       Creatinine (µmol/L)     134     64-104       Total bilirubin (µmol/L)     10     3-20       25-OH-vitamin D3 (nmol/L)     92.2     50.0-125.0	Total cholesterol (mmol/L)	3.6	<5.0
Urea (carbamide) (mmol/L)     7.5     2.8-8.3       Creatinine (µmol/L)     134     64-104       Total bilirubin (µmol/L)     10     3-20       25-OH-vitamin D3 (nmol/L)     92.2     50.0-125.0	LDL cholesterol (mmol/L)	2.1	<3.0
Creatinine (µmol/L)     134     64-104       Total bilirubin (µmol/L)     10     3-20       25-OH-vitamin D3 (nmol/L)     92.2     50.0-125.0	HDL cholesterol (mmol/L)	1.1	>1.0
Total bilirubin (µmol/L)       10       3-20         25-OH-vitamin D3 (nmol/L)       92.2       50.0-125.0	Urea (carbamide) (mmol/L)	7.5	2.8-8.3
25-OH-vitamin D3 (nmol/L) 92.2 50.0-125.0	Creatinine (µmol/L)	134	64-104
	Total bilirubin (µmol/L)	10	3-20
Urinary proteins (dipstick) 1+ Negative	25-OH-vitamin D3 (nmol/L)	92.2	50.0-125.0
	Urinary proteins (dipstick)	1+	Negative
Urinary glucose (dipstick) negative Negative	Urinary glucose (dipstick)	negative	Negative
SpO <sub>2</sub> by pulse oximeter (%) 95 >94	SpO <sub>2</sub> by pulse oximeter (%)	95	>94

risk of severe COVID-19 course/manifestation is posed by the presence of either diabetes mellitus, obesity, arterial hypertension, or immunosuppression / immunodeficiency.<sup>3-8</sup> Severe COVID-19 manifestations include development of extensive bilateral pneumonia, ARDS, and endothelial damage accompanied with accelerated thrombogenesis, which can all lead to the respiratory and multiorgan failure, being major clinical concern due to intensive care units overcrowding and high mortality rate.<sup>4,6</sup>

According to the current knowledge, the basic pathogenetic mechanisms responsible for the propensity to severe \_Clinical Case Reports

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**TABLE 2** Laboratory findings obtained 20 d after the initial presentation (23 d after the symptoms appeared)

Parameter (unit)	Result	Reference range
Total white blood cell $(\times 10^{9}/L)$	5.8	3.4-9.7
Lymphocytes (×10 <sup>9</sup> /L)	1.34	1.19-3.35
Neutrophils (×10 <sup>9</sup> /L)	3.56	2.06-6.49
Monocytes (×10 <sup>9</sup> /L)	0.72	0.12-0.84
Eosinophils (×10 <sup>9</sup> /L)	0.10	0.00-0.43
Basophils (×10 <sup>9</sup> /L)	0.03	0.00-0.06
Red blood cell ( $\times 10^{12}/L$ )	4.50	4.34-5.72
Hematocrit (%)	0.423	0.415-0.530
Hemoglobin (g/L)	142	138-175
MCV (fL)	94.0	83.0-97.2
MCH (pg)	31.6	27.4-33.9
MCHC (g/L)	336	320-345
Platelets ( $\times 10^9/L$ )	191	158-424
MPV (fL)	9.6	6.8-10.4
C-reactive protein (mg/L)	4.4	0-5
Erythrocyte sedimentation rate-1 h (mm/3.6 ks)	42	3-23
Glucose (mmol/L)	9.8	4.4-6.4
Urea (carbamide) (mmol/L)	7.8	2.8-8.3
Creatinine (µmol/L)	141	64-104
25-OH-vitamin D3 (nmol/L)	88.4	50.0-125.0
Urinary proteins (dipstick)	1+	Negative
Urinary glucose (dipstick)	negative	Negative
SpO <sub>2</sub> by pulse oximeter (%)	96	>94
SARS-CoV-2 IgG (index)	33.16	$\geq 1.00 = \text{positive}$
SARS-CoV-2 IgM (index)	5.09	$\geq 1.00 = \text{positive}$

COVID-19 course in the patients at risk could be reduced to a common denominator relating to facilitation and reinforcement of inflammatory processes. Inappropriately excessive and uncontrolled inflammation more likely can result with the development of autodestrucitive cytokine storm, which is considered to be the final effector mechanism of tissue damage.<sup>4,6,10,11</sup> Obesity, insulin resistance and diabetes mellitus, as well as hypertension, are characterized by the presence of persistent low-intensity inflammation, which may be decisive underlying factor amplifying and perpetuating viral-induced inflammatory response.<sup>4-11</sup> Some authors also suggest pathogenetic involvement of immunogenic damage-associated molecular patterns (DAMPs), released as a consequence of preexisting disease.<sup>29</sup> Immunosuppression is thought to favor more severe disease development by permitting faster viral replication resulting in extensive direct viral-mediated tissue -WILEY\_Clinical Case Reports

damage.<sup>4,6,11</sup> In the other hand, immunosuppression can mitigate excessive inflammation. There are studies showing no increased risk of severe COVID-19 manifestation in patients on immunosuppressive therapy, including methotrexate,<sup>30,31</sup> as well as reports that emphasize severe COVID-19 course in such patients.<sup>32,33</sup> Accordingly. available data on the impact of variety of immunosuppressant therapy regarding COVID-19 clinical course are inconclusive yet.<sup>30,34</sup> Both, insufficient directed immune response and unpurposeful inflammatory overactivity are assumed to increase possibility of severe COVID-19 manifestations in the elderly patients, as aging is known to impair considerably the efficacy of immunoregulatory mechanisms.<sup>35,36</sup> The probability of severe COVID-19 increases with age and number of preexisting pathological conditions and is particularly high in patients with older age and more risk comorbidities,<sup>37</sup> indicating a cumulative effect of the risk factors.

Here, we described a mild COVID-19 course, without any concerning symptoms or clinical signs, and with a favorable outcome in the patient with multiple conditions considered as risk factors: obesity, T2DM, hypertension, age >65, and immunosuppressive therapy. Laboratory findings of the proteinuria and hypercreatininemia also suggest renal function impairment, probably due to diabetic nephropathy development. Chronic renal failure was also found to be independently associated with poor clinical outcome.<sup>38</sup> Besides, given the long-standing history of psoriasis in our patient (25 years), we also point out that several studies have found increased risk of serious infection (especially cutaneous and respiratory) in psoriatic patients independently on the therapy and other comorbidities.<sup>30,39,40</sup> The risk of serious infection in psoriatic patients was found to increase further with the severity of the disease<sup>39</sup> and the presence of either diabetes mellitus, obesity, age >60 years,  $^{41,42}$  or history of smoking.<sup>42</sup> Taking into account the recent cutaneous exacerbation (3 months before the actual presentation; treated by UVB phototherapy), our patient had all of the above conditions that may additionally increase the risk of severe infection in the context of psoriasis per se.

Laboratory findings in our patients, showing normal leukocyte count and only slightly elevated inflammatory markers, are in line with a mild clinical picture. The exception is lymphopenia, which has been found to be associated with severe COVID-19 cases,<sup>37,38</sup> but in presented patient lymphopenia very likely may be influenced by long-term treatment with methotrexate, which is known to inhibit DNA synthesis and cause bone marrow toxicity.<sup>43,44</sup> Such causality can also be attributed to the anemia, found in our patient. Although there is a paucity of data on clear relationship between anemia and COVID-19 severity, one meta-analysis found an association of anemia with severe manifestation of COVID19, but not with mortality rate.<sup>45</sup>

We would like to emphasize the sufficient plasma 25-OH-vitamin D3 (calcifediol, precursor of an active form of the vitamin D3) level in the presented patient, which could have had, at least in part, positive impact on the disease course and outcome. Namely, since time when the possible favorable effects of vitamin D3 in COVID-19 patients have been proposed,<sup>14,15</sup> several well-designed and convincing studies have clearly shown association between vitamin D deficiency and more severe COVID-19 clinical manifestations.<sup>16-20,46</sup> As a recent meta-analysis shows, the need for hospitalization and mortality due to COVID-19 is significantly higher in people with insufficient levels of vitamin D.<sup>16</sup> Study by Jain and colleagues found that prevalence of vitamin D deficiency among patients with severe COVID-19 manifestations treated in intensive care units was as high as 96.82%, in contrast to 32.96% in completely asymptomatic patients.<sup>17</sup> Besides, serum IL-6 and TNF $\alpha$  levels negatively correlated with 25-OH-vitamin D3 levels, and most importantly, overall fatality rate was almost sevenfold higher in patients with vitamin D deficiency.<sup>17</sup> An ecological study that included analysis of association between vitamin D deficiency and COVID-19 incidence, complications, and case fatality in 46 countries found positive and statistically significant correlation between vitamin D deficiency and all COVID-19 variables analyzed.<sup>18</sup> Recent interventional studies provide further evidence on beneficial and protective impact of vitamin D3 among COVID-19 patients.<sup>21-24</sup> Among hospitalized COVID-19 patients recruited for the randomized clinical study, those who were treated with oral 25-OH-vitamin D3 (0.532 mg initially, followed by 0.266 mg on day 3 and 7, and then same dose once a week) as an adjuvant to standard therapy were being admitted to the intensive care unit (ICU) considerably less frequently than a control group receiving only standard therapy (2% vs 50%; P < .001).<sup>21</sup> High-dose booster application of vitamin D (in a total dose  $\geq$  280 000 IU applied by different regimens in a period of up to 7 weeks and started immediately upon COVID-19 diagnosis) also significantly reduced mortality among COVID-19 patients comparing with a control group that was not undergone such intervention.<sup>22</sup> Protective vitamin D3 mechanisms in COVID-19 patients are presumed to be multiple, including potent immunomodulatory activity, antiviral peptide induction, enhancement of physical barrier integrity, and even direct interference with viral replication.<sup>13-15,47</sup> Vitamin D has been found to induce defensins and LL-37 (the only human member of the cathelicidin family), enzymatic peptides with potent ability to disturb viral envelopes and cleave virion proteins.<sup>15</sup> Furthermore, through the induction of cell-cell adhesion molecules, mostly those from the cadherin superfamily, vitamin D promotes cellular junction integrity, which are usually disrupted by the viruses.<sup>15</sup> Weakening of intercellular junctions, in turn, allows more rapid viral invasion of the mucosal barriers and subsequent spreading through tissues.<sup>15</sup> Moreover, such "firming" effect

of vitamin D on the intercellular integrity could also mitigate capillary leakage, thereby decreasing risk of pulmonary edema and ARDS development.

Vitamin D-mediated suppression of excessive T helper cell type 1 (Th1) response and proinflammatory cytokine hypersecretion is considered as one of the most important effect that reduces the likelihood of cytokine storm development, major pathogenic event concerning severe COVID-19 manifestation.<sup>14,15,47-49</sup> Promoting induction of the T regulatory cells (Tregs), vitamin D may further enhance these immunomodulatory and anti-inflammatory effects.<sup>15</sup> Ability of vitamin D3 to induce production of type I interferons (IFNs) is also of great importance during SARS-CoV-2 infection, since type I IFNs has been known as the most powerful natural mediators of antiviral activity in humans, keeping viral replication under control and enabling effective viral clearance without excessive inflammatory response.<sup>50</sup> In addition, impaired or delayed type I IFNs response during early stage of SARS-CoV-2 infection have been found to play a major role in the cytokine storm development,<sup>51,52</sup> probably by allowing accelerated viral replication, which at a later stage leads to excessive inflammation. It should also be noted that several preclinical and clinical studies have indicated the ability of vitamin D to promote antithrombin and thrombomodulin gene expression and to inhibit tissue factor activity, thus reducing a thrombogenic microenvironment.<sup>15-17</sup> The antithrombotic activity of vitamin D also may be related to the above-mentioned inhibition of proinflammatory cytokines favoring increased thrombogenicity. Such effects of vitamin D could provide important protection during Sars-CoV-2 infection, since hypercoagulability and accelerated thrombogenesis with consequent increase in thrombotic episodes and D-dimer formation are commonly observed in COVID-19 patients, especially in those who due to intensive care treatment acquire all components of Virchow's triad.<sup>53</sup> Another beneficial effect of vitamin D is achieved by modulation of the renin-angiotensin system (RAS).<sup>47</sup> Namely, Sars-CoV-2 infection disturbs RAS activity by downregulation of angiotensin-converting enzyme 2 (ACE2), "expending" it as a viral receptor for cellular entry.<sup>6,47</sup> Resulting reduced cleavage of Angiotensin II (Ang II) leads to the excessive accumulation of this noxious molecule exerting proinflammatory and profibrotic activity, thus aggravating lung injury.<sup>6,47</sup> Vitamin D has been found to reduce renin secretion and promote ACE2 expression, preventing dysregulation of the RAS in favor of harmful Ang II.47

Considering the available data on the protective vitamin D3 effects, but also the "pandemic" of vitamin D deficiency,<sup>25-28</sup> we can assume that in our patient, burdened by multiple risk factors, sufficient plasma 25-OH-vitamin D3 curbed detrimental immunopathogenic mechanisms and enabled favorable outcome. Since there are high prevalence of vitamin D3 deficiency in general population, with even more frequent occurrence among patients with obesity, T2DM and hypertension,<sup>27,28</sup> and our patient had not been supplemented per os, we can presume that sufficient level of 25-OH-vitamin D3 had been achieved by UVB phototherapy. This assumption complies to observed seasonal changes in COVID-19 symptom severity and incidence, which are the lesser during the periods with the longer sunlight duration.<sup>15,54</sup> However, such scenario with a complex pathophysiological constellation does not allow us to rule out the possible impact of the methotrexate therapy, as well as the influence of patient's genetic makeup on the course and outcome of the disease. It is also noteworthy that patient gained appropriate antiviral immunoglobulin levels despite prolonged methotrexate treatment, since it is known that methotrexate can suppress the production of antibodies to neoantigens.<sup>55-57</sup> As vitamin D3 has been found to promote T helper type 2 (Th2) response, IL-4 production and Tregs generation,<sup>15,58</sup> such humoral outcome could also be supported by sufficient 25-OH-vitamin D3 status in described patient. Interestingly, an increase in the frequency of naïve B cells and Tregs after narrowband UVB phototherapy was previously shown.<sup>59</sup> These observations may be of great importance for improving COVID-19 vaccination efficacy, but further thorough studies are needed on this topic.

Described patient presented with typical (cough, fever), but very mild COVID-19 symptoms without any complication despite multiple risk factors. Such cases should not be confused with atypical COVID-19 cases, which can present with a wide variety of symptoms, some of which can become severe and life-threatening.<sup>60</sup>

In conclusion, we point to likely possible role of vitamin D as a positive modifier of COVID-19 course and outcome and suggest that routine determination of 25-OH-vitamin D3 status could be considered as useful tool for, at least rough, estimation of COVID-19 outcome, as well as for deciding on vitamin D3 supplementation.

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Published with written consent of the patient.

# **CONFLICT OF INTEREST**

None declared.

#### AUTHOR CONTRIBUTIONS

Martina Kralj managed the patient, collected the data, and contributed to the manuscript drafting. Hrvoje Jakovac interpreted the findings, reviewed the literature, drafted, and wrote the manuscript. Both authors have read and approved the final manuscript.

# ETHICAL CONSIDERATIONS

The patient was informed in detail and he provided written consent.

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# DATA AVAILABILITY STATEMENT

The data are not publicly available due to the protection of patient privacy and adherence to ethical principles. Clinical and laboratory data may be provided upon reasonable request to corresponding author.

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