



Complete and Circularized Genome Assemblies of the *Kroppenstedtia eburnea* Genus Type Strain and the *Kroppenstedtia pulmonis* Species Type Strain with MiSeq and MinION Sequence Data

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ABSTRACT *Kroppenstedtia eburnea* DSM 45196^T and *Kroppenstedtia pulmonis* W9323^T are aerobic, Gram-positive, filamentous, chemoorganotrophic thermoactinomycetes. Here, we report on the complete and circular genome assemblies generated using Illumina MiSeq and Oxford Nanopore Technologies MinION reads. Putative gene clusters predicted to be involved in the production of secondary metabolites were also identified.

Isolates within the *Kroppenstedtia* genus are characterized as Gram-positive, non-motile, aerobic, filamentous chemotrophs capable of producing heat-resistant endospores (1–3). The four species of the genus include *Kroppenstedtia eburnea* (1), *Kroppenstedtia guangzhouensis* (2), *Kroppenstedtia pulmonis* (3), and *Kroppenstedtia sanguinis* (3). *K. eburnea* DSM 45196^T was isolated from a plastic surface in Germany; subsequently, clinical isolates of the same species were identified in the United States (1, 4). *K. pulmonis* W9323^T was isolated from a lung biopsy sample from a 78-year-old male patient from the United States (3). The genomes for *Kroppenstedtia eburnea* DSM 45196^T and *Kroppenstedtia pulmonis* W9323^T described here were sequenced because of their potential sources of genes encoding secondary metabolites, as well as adaptation of an environmental thermoactinomycete isolated from soil to clinical sources.

Kroppenstedtia eburnea DSM 45196^T was purchased from the German Collection of Microorganisms and Cell Cultures (catalogue number DSM45196), and *Kroppenstedtia pulmonis* W9323^T was obtained from the Special Bacteriology Reference Laboratory, Centers for Disease Control and Prevention (CDC) (Atlanta, GA) (3). Cells were grown in Trypticase soy broth from single colonies, and genomic DNA used for both libraries was purified using the MasterPure DNA purification kit (Epicentre, Madison, WI) according to the manufacturer's protocol (5). MinION libraries were made with the rapid barcoding kit (Oxford Nanopore Technologies), and sequences were generated with R9.4.1 flow cells and Guppy v3.2.8+bd97289. The numbers of raw reads for *K. eburnea* from the MiSeq and MinION instruments were 13,310,694 and 148,000, respectively, and those for *K. pulmonis* from the MiSeq and MinION instruments were 2,248,184 and 404,000, respectively. The MinION sequence N_{50} values were 5,681 bp and 5,607 bp for *K. eburnea* and *K. pulmonis*, respectively. Default parameters were used for all software unless otherwise specified. Flye v2.6 with the setting “-g 4m” formed initial assemblies (6). Three sequential rounds of assembly corrections were performed with minimap

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v2.17-r941, using the setting “-x map-ont,” and racon v1.3.2 (7, 8). A final long-read correction was accomplished with medaka v0.11.1, with 131× and 362× coverages for *K. eburnea* and *K. pulmonis*, respectively. Illumina MiSeq sequences (2 × 250 bp) that had been filtered with Trimmomatic v0.35 to scores of ≥Q30 were used with the Unicycler v0.4.8 polishing function, which used Bowtie v2.3.4.3, SAMtools v1.3.1, and Pilon v1.23 (9–13), with 642× and 89× coverages for *K. eburnea* and *K. pulmonis*, respectively. Polishing continued until the assembly likelihood scores no longer improved according to ALE v20180904 (four rounds for *K. eburnea* fixed 20,779 variants, and two rounds for *K. pulmonis* fixed 22,201 variants) (14). The 3,564,999-bp (54% GC content) and 3,345,811-bp (46% GC content) circular chromosomes for *K. eburnea* and *K. pulmonis*, respectively, were reoriented to start with *dnaA*, which was located with BLAST 2.9.0+ (15) using locus tag D1G38_003660 in *Kroppenstedtia sanguinis* (5). Pseudogenes were inferred by comparing proteins aligned with DIAMOND v0.9.22 to the UniProtKB/TrEMBL database v2019_10 (16–20). The numbers of best-match (based on bit scores) alignments with >10% deviation in length were 88 in *K. eburnea* and 298 in *K. pulmonis*. CheckM v1.0.13 estimated both assemblies as 100.00% complete (21). The genome assemblies were annotated using PGAP v4.11, which predicted 4 and 6 CRISPR sequences in *K. eburnea* and *K. pulmonis*, respectively (22). AntiSMASH v5.1.0 and Prism v4.4.3 predicted ectoine biosynthesis capability in both genomes (23, 24). AntiSMASH found 10 additional putative biosynthetic gene clusters (BGCs) of interest (5 nonribosomal peptide synthetases [NRPSs] and putative genes associated with siderophore production, which may enhance pathogenicity [25]) for *K. pulmonis* and 6 additional BGCs of interest (2 NRPSs) for *K. eburnea*. Thermonucleases can serve as virulence factors (26, 27); 2 were predicted for *K. eburnea* (protein accession numbers [QKI81670.1](#) and [QKI83414.1](#)) and 1 was predicted for *K. pulmonis* (protein accession number [QKG85853.1](#)). These complete type strain genomes will be valuable for taxonomic assignments and will aid in biosynthesis and natural product research.

Data availability. The whole-genome sequences of *Kroppenstedtia eburnea* DSM 45196^T and *Kroppenstedtia pulmonis* W9323^T have been deposited in DDBJ/ENA/GenBank under the accession numbers [CP048103](#) and [CP048104](#), respectively. The BioProject accession number for *Kroppenstedtia eburnea* DSM 45196^T and *Kroppenstedtia pulmonis* W9323^T is [PRJNA602730](#), and the associated BioSample accession numbers are [SAMN13905711](#) and [SAMN13905715](#), respectively. The *Kroppenstedtia eburnea* DSM 45196^T Illumina MiSeq reads and Nanopore reads are available in the NCBI Sequence Read Archive (SRA) under the accession numbers [SRX7624950](#) and [SRX7624951](#), respectively. The *Kroppenstedtia pulmonis* W9323^T Illumina MiSeq reads and Nanopore reads are available in the NCBI SRA under the accession numbers [SRX7624960](#) and [SRX7624961](#), respectively.

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REFERENCES

- von Jan M, Riegger N, Pötter G, Schumann P, Verburg S, Spröer C, Rohde M, Lauer B, Labeda DP, Klenk H-P. 2011. *Kroppenstedtia eburnea* gen. nov., sp. nov., a thermoactinomycete isolated by environmental screening, and emended description of the family *Thermoactinomycetaceae* Matsuo et al. 2006 emend. Yassin et al. 2011. *Int J Syst Evol Microbiol* 61:2304–2310. <https://doi.org/10.1099/ijs.0.026179-0>.
- Yang G, Qin D, Wu C, Yuan Y, Zhou S, Cai Y. 2013. *Kroppenstedtia guangzhouensis* sp. nov., a thermoactinomycete isolated from soil. *Int J Syst Evol Microbiol* 63:4077–4080. <https://doi.org/10.1099/ijs.0.051011-0>.
- Bell ME, Lasker BL, Klenk H-P, Hoyles L, Spröer C, Schumann P, Brown JM. 2016. *Kroppenstedtia pulmonis* sp. nov. and *Kroppenstedtia sanguinis* sp. nov., isolated from human patients. *Antonie Van Leeuwenhoek* 109: 603–610. <https://doi.org/10.1007/s10482-016-0663-z>.
- Barker AP, Simmon KE, Cohen S, Slechta ES, Fisher MA, Schlaberg R. 2012. Isolation and identification of *Kroppenstedtia eburnea* isolates from multiple patient samples. *J Clin Microbiol* 50:3391–3394. <https://doi.org/10.1128/JCM.01186-12>.
- Arthur RA, Nicholson AC, Humrighouse BW, McQuiston JR, Lasker BL. 2019. Draft genome sequence of *Kroppenstedtia sanguinis* X0209^T, a clinical isolate recovered from human blood. *Microbiol Resour Announc* 8:e00354-19. <https://doi.org/10.1128/MRA.00354-19>.
- Kolmogorov M, Yuan J, Lin Y, Pevzner P. 2019. Assembly of long, error-prone reads using repeat graphs. *Nat Biotechnol* 37:540–546. <https://doi.org/10.1038/s41587-019-0072-8>.
- Li H. 2018. Minimap2: pairwise alignment for nucleotide sequences. *Bioinformatics* 34:3094–3100. <https://doi.org/10.1093/bioinformatics/bty191>.
- Vaser R, Sović I, Nagarajan N, Šikić M. 2017. Fast and accurate de novo genome assembly from long uncorrected reads. *Genome Res* 27:737–746. <https://doi.org/10.1101/gr.214270.116>.

9. Bolger AM, Lohse M, Usadel B. 2014. Trimmomatic: a flexible trimmer for Illumina sequence data. *Bioinformatics* 30:2114–2120. <https://doi.org/10.1093/bioinformatics/btu170>.
10. Wick RR, Judd LM, Gorrie CL, Holt KE. 2017. Unicycler: resolving bacterial genome assemblies from short and long sequencing reads. *PLoS Comput Biol* 13:e1005595. <https://doi.org/10.1371/journal.pcbi.1005595>.
11. Langmead B, Salzberg S. 2012. Fast gapped-read alignment with Bowtie 2. *Nat Methods* 9:357–359. <https://doi.org/10.1038/nmeth.1923>.
12. Li H, Handsaker B, Wysoker A, Fennell T, Ruan J, Homer N, Marth G, Abecasis G, Durbin R, 1000 Genome Project Data Processing Subgroup. 2009. The Sequence Alignment/Map format and SAMtools. *Bioinformatics* 25:2078–2079. <https://doi.org/10.1093/bioinformatics/btp352>.
13. Walker BJ, Abeel T, Shea T, Priest M, Abouelliel A, Sakthikumar S, Cuomo CA, Zeng Q, Wortman J, Young SK, Earl AM. 2014. Pilon: an integrated tool for comprehensive microbial variant detection and genome assembly improvement. *PLoS One* 9:e112963. <https://doi.org/10.1371/journal.pone.0112963>.
14. Clark SC, Egan R, Frazier PI, Wang Z. 2013. ALE: a generic assembly likelihood evaluation framework for assessing the accuracy of genome and metagenome assemblies. *Bioinformatics* 29:435–443. <https://doi.org/10.1093/bioinformatics/bts723>.
15. Camacho C, Coulouris G, Avagyan V, Ma N, Papadopoulos J, Bealer K, Madden TL. 2009. BLAST+: architecture and applications. *BMC Bioinformatics* 10:421. <https://doi.org/10.1186/1471-2105-10-421>.
16. Watson M, Warr A. 2019. Errors in long-read assemblies can critically affect protein prediction. *Nat Biotechnol* 37:124–126. <https://doi.org/10.1038/s41587-018-0004-z>.
17. Seemann T. 2014. Prokka: rapid prokaryotic genome annotation. *Bioinformatics* 30:2068–2069. <https://doi.org/10.1093/bioinformatics/btu153>.
18. Hyatt D, Chen GL, Locascio PF, Land ML, Larimer FW, Hauser LJ. 2010. Prodigal: prokaryotic gene recognition and translation initiation site identification. *BMC Bioinformatics* 11:119. <https://doi.org/10.1186/1471-2105-11-119>.
19. Buchfink B, Xie C, Huson DH. 2015. Fast and sensitive protein alignment using DIAMOND. *Nat Methods* 12:59–60. <https://doi.org/10.1038/nmeth.3176>.
20. UniProt Consortium. 2019. UniProt: a worldwide hub of protein knowledge. *Nucleic Acids Res* 47:D506–D515. <https://doi.org/10.1093/nar/gky1049>.
21. Parks DH, Imelfort M, Skennerton CT, Hugenholtz P, Tyson GW. 2015. CheckM: assessing the quality of microbial genomes recovered from isolates, single cells, and metagenomes. *Genome Res* 25:1043–1055. <https://doi.org/10.1101/gr.186072.114>.
22. Tatusova T, DiCuccio M, Badretdin A, Chetvernin V, Nawrocki EP, Zaslavsky L, Lomsadze A, Pruitt KD, Borodovsky M, Ostell J. 2016. NCBI Prokaryotic Genome Annotation Pipeline. *Nucleic Acids Res* 44:6614–6624. <https://doi.org/10.1093/nar/gkw569>.
23. Blin K, Shaw S, Steinke K, Villebro R, Ziemert N, Lee SY, Medema MH, Weber T. 2019. antiSMASH 5.0: updates to the secondary metabolite genome mining pipeline. *Nucleic Acids Res* 47:W81–W87. <https://doi.org/10.1093/nar/gkz310>.
24. Skinnider MA, Merwin NJ, Johnston CW, Magarvey NA. 2017. PRISM 3: expanded prediction of natural product chemical structures from microbial genomes. *Nucleic Acids Res* 45:W49–W54. <https://doi.org/10.1093/nar/gkx320>.
25. Miethke M, Marahiel MA. 2007. Siderophore-based iron acquisition and pathogen control. *Microbiol Mol Biol Rev* 71:413–451. <https://doi.org/10.1128/MMBR.00012-07>.
26. Hu Y, Meng J, Shi C, Hervein K, Fratamico PM, Shi X. 2013. Characterization and comparative analysis of a second thermonuclease from *Staphylococcus aureus*. *Microbiol Res* 168:174–182. <https://doi.org/10.1016/j.micres.2012.09.003>.
27. Juneau RA, Stevens JS, Apicella MA, Criss AK. 2015. A thermonuclease of *Neisseria gonorrhoeae* enhances bacterial escape from killing by neutrophil extracellular traps. *J Infect Dis* 212:316–324. <https://doi.org/10.1093/infdis/jiv031>.