Increased bone formation in a rabbit long-bone defect model after single local and single systemic application of erythropoietin

Georg W OMLOR¹, Kerstin KLEINSCHMIDT², Simone GANTZ¹, Anja SPEICHER², Thorsten GUEHRING³, and Wiltrud RICHTER²

Background and purpose — Delayed bone healing with nonunion is a common problem. Further options to increase bone healing together with surgery are needed. We therefore evaluated a 1-dose single application of erythropoietin (EPO), applied either locally to the defect or systemically during surgery, in a criticalsize rabbit long-bone defect.

Material and methods — 19 New Zealand White rabbits received a 15-mm defect in the radius diaphysis. An absorbable gelatin sponge was soaked with saline (control group and systemic treatment group) or EPO (local treatment group) and implanted into the gap. The systemic treatment group received EPO subcutaneously. In vivo micro-CT analysis was performed 4, 8, and 12 weeks postoperatively. Vascularization was evaluated histologically.

Results — Semiquantitative histomorphometric and radiological evaluation showed increased bone formation (2.3- to 2.5-fold) in both treatment groups after 12 weeks compared to the controls. Quantitative determination of bone volume and tissue volume showed superior bone healing after EPO treatment at all followup time points, with the highest values after 12 weeks in locally treated animals (3.0- to 3.4-fold). More vascularization was found in both EPO treatment groups.

Interpretation — Initial single dosing with EPO was sufficient to increase bone healing substantially after 12 weeks of follow-up. Local application inside the defect was most effective, and it can be administered directly during surgery. Apart from effects on ossification, systemic and local EPO treatment leads to increased callus vascularization.

Delayed or insufficient bone healing with development of non-union is a major clinical problem. Several groups have described increased bone healing by vascular-endothelial growth factor- (VEGF-) induced effects on angiogenesis and enchondral bone formation (Geiger et al. 2005, 2007). Structural and functional conformities between VEGF and the glycoprotein erythropoietin (EPO) (Jelkmann 1992) have caused growing interest in EPO to improve bone healing. EPO is routinely used to treat anemia, especially in patients with chronic renal failure. It is also known as a doping substance for athletes, due to erythropoiesis stimulation. It is a pleiotropic substance with osteogenic and angiogenic potency, anti-inflammatory effects by antagonization of TNFa, mitogenic and chemotactic effects, and neuro-, renal-, and cardioprotective functions (Anagnostou et al. 1990, Jaquet et al. 2002, Heeschen et al. 2003, Prunier et al. 2007, Brines and Cerami 2008, Arcasoy 2010, Chateauvieux et al. 2011, Rölfing 2014).

Based on bone-related pleiotropic effects, EPO appears attractive for enhancement of bone healing. Despite increased osteoclast stimulation (Hiram-Bab et al. 2015), several other EPO-induced mechanisms have been found to result in increased bone formation (McGee et al. 2012). However, only a few studies have been done to date that have evaluated the effects of EPO on in vivo bone healing in animal models (Bozlar et al. 2006, Holstein et al. 2007, 2011, Shiozawa et al. 2010, Garcia et al. 2011, Rölfing et al. 2012, Sun et al. 2012). According to the studies that have been done, relevant EPOderived effects on bone healing require daily EPO dosing, but with serious side effects related to high concomitant hemoglobin stimulation. Whether single EPO dosing, which facilitates clinical application and reduces side effects, might also

© 2016 The Author(s). Published by Taylor & Francis on behalf of the Nordic Orthopedic Federation. This is an Open Access article distributed under the terms of the Creative Commons Attribution-Non-Commercial License (https://creativecommons.org/licenses/by-nc/3.0) DOI 10.1080/17453674.2016.1198200

¹ Center for Orthopedics, Trauma Surgery and Spinal Cord Injury, Heidelberg University Hospital; ² Research Center for Experimental Orthopedics, Heidelberg University Hospital; ³ BG Trauma Hospital Ludwigshafen, University of Heidelberg, Germany. Correspondence: wiltrud.richter@med.uni-heidelberg.de

Submitted 2015-11-09. Accepted 2016-04-07.

be sufficient to improve bone healing is the subject of debate (Rölfing et al. 2014a).

We therefore analyzed bone healing after single local or systemic EPO application in an established in vivo rabbit longbone model based on a 15-mm-radius critical-size defect.

Material and methods

Animals and surgical procedure

19 skeletally mature 6-month-old female New Zealand White rabbits with closed epiphyseal plates were randomly assigned to 1 control group and 2 treatment groups. Mean animal weight was 4.2 (3.6-4.8) kg. A detailed description of the animal model has already been published (Kleinschmidt et al. 2013, 2014). Briefly, a 15-mm defect was prepared in the middle of the diaphysis of the radius using a handsaw. Due to physiological membrane fixation of the proximal and distal endings of the radius to the ulna, no additional fixation of the defect was needed to stabilize the radius. The resulting gap in the radius was filled with an absorbable gelatin sponge (Gelita-Spon; Gelita Medical BV, Amsterdam, the Netherlands). Gelita-Spon is a common, regularly available hemostat without intrinsic hemostatic action, which induces hemostasis through its intensely porous structure. It is pH-neutral, and therefore suitable as a local drug carrier. Depending on application and environment, it is completely biodegraded in less than 4 weeks (Goncalves et al. 2015). The sponge was cut into pieces of 2×0.5 cm and soaked in sterile saline (control group and systemic treatment group) or EPO (local treatment group).

6 rabbits served as controls that did not receive EPO. The remaining 13 rabbits were divided into 2 EPO treatment groups. 6 animals received 1 single high dose (4,900 IU per metabolic body weight in kg (metkg)) of EPO systemically (subcutaneously; injection site: upper back) during surgery after the defect was prepared and the saline-soaked gelatin sponge was placed. 7 animals received 1 single high dose (4,900 IU/metkg) of EPO locally. Here, the gelatin sponge was manually soaked with the individual animal EPO dose during surgery, immediately before the sponge was placed in the defect. The rationale for this setup was that it was an easy-to-use application process that could be directly transferred to the clinical situation, since all components and substances are routinely available without further modifications.

The rabbits were kept in single cages in the institutional animal laboratory according to animal care regulations (automatic light-dark cycle, pain medication with carprofen (4 mg/ kg body weight), with water and food ad libitum). Follow-up was done by in vivo micro-CT analysis 4, 8, and 12 weeks postoperatively. The rabbits were then killed by pentobarbital injection and specimens were resected for additional histological evaluation.



Figure 1. Bone formation score from 0–3 at endpoint. No relevant bone growth (bone formation thickness < 200% of ulna corticalis; bone formation width < 50% of the 15-mm-long defect) was assigned a score value of 0 (panel A). Bone formation thickness > 200% and width > 50% without significant separation from the ulna was assigned a score value of 1 (B). Bone growth around the radial aspect of the ulna with significant separation from the ulna over a distance of > 50% of the 15-mm-long callus was assigned a score value of 2 (C). Bone growth with radius corticalis and marrow cavity formation was assigned a score value of 3 (D). Here, long bone corticalis and marrow cavity should be visible over a distance of > 50% of the 15-mm-long callus.

EPO dosage

We used Epoetin alfa (Erypo FS; Centocor BV, Leiden, the Netherlands). 1 IU contained 0.0084 µg Epoetin alfa, which is genetically engineered from ovarian cells of the Chinese hamster (cell-line CHO-K1). The clinically available, ready-to-use standard injection fluid also contained polysorbat 80, glycine, sodium chloride, disodium phosphate dihydrate, sodium dihydrogen phosphate dihydrate, and water.

For dosage, the metabolic body weight was applied and calculated as follows: metabolic body weight of the animal = individual body weight of the animal to the power of 0.75. Based on our own unpublished pilot experiments on the New

Zealand White rabbit and clinical data on anemic cancer patients (Shasha et a. 2003; Gabrilove et al. 2001), a single EPO dosage of 4,900 IU/metkg was chosen as a feasible single high dose, avoiding any side effects related to increase in hematocrit.

Semiquantitative histomorphometric and radiological evaluation of bone formation after 12 weeks

Micro-CT scan reconstructions in all planes and sagittal histological slides from each rabbit were manually analyzed independently by 2 observers (GO and AS) who were blinded regarding the treatment group. They judged bone growth semiquantitatively according to a scoring system ranging from 0 to 3 points (Figure 1), where 0 points meant no relevant bone growth (bone formation thickness less than 200% of ulna corticalis; width of bone growth less than 50% of the 15-mm-long defect); where 1 point meant isolated bone growth around the radial aspect of the ulna (more than 200% thickness; more than 50% width); where 2 points meant bone growth around the radial aspect of the ulna with separation from the ulna (separation should be visible in more than 50% of the 15-mmlong callus); and 3 points meant bone growth with radius corticalis and marrow cavity formation (new long-bone corticalis and marrow cavity should be visible in more than 50% of the 15-mm-long callus). As measures of reliability, Cohen's kappa indicated a good (control animals: $\kappa = 0.7$, p = 0.4) to very good (systemically or locally treated animals: $\kappa = 1$, p = 0.01) level of agreement between the 2 raters.

Quantitative micro-CT analysis

In vivo scanning of the rabbit forelegs was done 4, 8, and 12 weeks postoperatively. The procedure was performed as previously reported (Kleinschmidt et al. 2013). Briefly, a micro-CT scanner (1076; Skyscan, Antwerp, Belgium) was used together with a rabbit bedding device customized by RJL Micro & Analytic (Karlsdorf-Neuthard, Germany). This allowed positioning of the animal beyond the system while only the foreleg was fixed in an aluminum tube that moved the region of interest horizontally into the course of the X-ray beam. Proximate scans were performed using a 0.5-mm aluminum filter with the following settings: 48 kV, 200 mA, 320 ms, spatial resolution 17.7 mm3/voxel, and rotation step 0.6°. Parameters were analyzed (using Skyscan software) with a lower gray level set at 40 and an upper gray level set at 255 to define callus tissue. The volume of interest (VOI) was determined in its proximal and distal dimensions by choosing the middle of the defect and taking the same number of 200 slices, which corresponded to 7 mm in each direction. Then interpolated VOIs that circumscribed the radius and whole callus excluding the ulna were drawn manually and automatically shrunk to the outer boundaries of the callus using the shrink-wrap function of the software. As objective parameters for quantitative evaluation of new bone formation inside the radius defect, volume of the whole callus tissue including non-mineralized marrow areas

(tissue volume (TV)) and volume of the mineralized callus tissue (bone volume (BV)) were calculated.

Histology

After 12 weeks, complete foreleg specimens were fixed, decalcified, and paraffin-embedded. Then, 5-µm-thick longitudinal sections were cut and stained with Masson-Goldner trichrome, hematoxylin and eosin, and Safranin-O, following standard protocols (Kleinschmidt et al. 2013). Aligned overview microphotographs of total slices were made to assess the adequacy of micro-CT-based analysis. For further evaluation, magnified images were taken from the area of the defect.

Quantitative histomorphometric analysis of blood vessels

HE-stained tissue sections were analyzed for capillaries and blood vessels that could be easily identified morphologically. For a quantitative evaluation, capillaries and vessels were encircled to calculate the surface area, which was divided by the total area of the tissue section according to Image J software protocols. Examinations were performed by 2 independent and blinded examiners (GO and AS), with good to very good inter-observer reliability (absolute agreement: 0.856– 0.999). Data were expressed as the percentage of vascular surface area within the callus.

Statistics

The inter-rater reliability for the semiquantitative score (0-3) was tested using Cohen's kappa. The inter-rater reliability for the quantitative analysis of blood vessels in histological slides was tested using intraclass correlation coefficients. EPO blood concentration was compared across 4 different points in time (day 0, 7, 14, and 21), using non-parametric Friedman tests with post hoc Wilcoxon tests. Differences between the animal groups were assessed with Kruskal-Wallis test and post hoc Mann-Whitney tests. Bonferroni correction for multiple comparisons was also performed, changing the significance level from p < 0.05 to p < 0.017, so statistical significance was only assumed for p < 0.017. All analyses were performed with SPSS version 21.0.

Ethics

The study was approved by the committee for animal experimentation of Baden-Wuerttemberg, Germany (reference number AZ35-9185.81/G-109/06).

Results

Animals

All animals completed the study to the endpoint. EPO therapy in humans has been rarely associated with diarrhea, fever, and local exanthema or edema at the injection site (Henry 2005). None of these effects and no other complications were found.

Table 1. Effect of EPO on hematocrit. The initial increase in hematocrit was followed by a decrease to initial values after 21 days. Hematocrit is presented as mean [SD] (range)

	Control group	Local group	Systemic group
Day 0	43 [6.9] (35–52)	40 [5.3] (33–44)	38 [5.2] (31–43)
Day 7	38 [5.1] (32–45)	50 [2.8] (46–53)	48 [5.4] (41–54)
Day 14	42 [9.3] (34–57)	44 [5.2] (39–51)	48 [6.2] (40–55)
Day 21	39 [3.1] (36–44)	42 [2.7] (39–44)	43 [3.0] (40–46)

The evaluation of hematocrit showed no statistically significant change in the control animals (p = 0.7) and in the animals treated locally (p = 0.09), but a relevant increase was seen in systemically treated animals (p = 0.02) until day 14. For all groups, there was a tendency of a decrease in initial hematocrit after 21 days (Table 1).

Semiquantitative evaluation of bone formation

Radiologically increased bone formation (Figure 1) was found in locally treated animals (p = 0.005) and systemically treated animals (p = 0.02) compared to the controls (overall Kruskal-Wallis test, p = 0.01).

The control animals showed no bone growth (n = 1) or minor bone growth around the radial aspect of the ulna (n = 5) without any signs of radial bone separation from the ulna, or corticalis and marrow cavity formation (mean score: 0.8).

Locally treated animals showed bone growth around the radial aspect of the ulna (n = 2), bone growth around the radial aspect of the ulna with separation from the ulna (n = 3), and bone growth with radius corticalis and marrow cavity formation (n = 2) (mean score: 2.0).

Systemically treated animals showed bone growth around the radial aspect of the ulna (n = 2), bone growth around the radial aspect of the ulna with separation from the ulna (n = 3), and bone growth with radius corticalis and marrow cavity formation (n = 1) (mean score: 1.8).

Quantitative micro-CT analysis of bone formation (tissue volume and bone volume)

Compared to control animals, both locally treated and systemically treated animals showed increased bone formation 4, 8, and 12 weeks after single EPO application (Figure 2). Quantitative analysis of bone formation with TV as a parameter defining the whole new callus volume and BV defining the volume of mineralized parts of the new callus showed higher values compared to control animals without EPO treatment at all time points (Figure 3).

Quantitative analysis of blood vessels

Significantly higher callus vascularization was found in both the local group (21.1% vessel surface area; 3.1-fold; p = 0.006) and the systemic group (22.3% vessel surface area; 3.3-fold;



Figure 2. Callus growth in untreated controls and in rabbits with a single EPO dose applied locally or systemically. During the 12 weeks of follow-up, statistically significantly higher bone growth was found in both treatment groups. The highest degree of bone growth was found in locally treated animals.

p = 0.004) compared to the control group with 6.8% vessel surface area (Figure 4).

Discussion

Our study clearly demonstrates the potency of regularly available EPO as an additional treatment option to increase bone healing. However, improvements in the efficiency of EPO on bone formation with less hematopoietic side effects could be achieved by using EPO-derived substances that bind specifically to the heterodimeric EPOR/CD131 receptor for pleiotropic effects instead of EPOR-mediated hematopoietic effects (Brines et al. 2004, Leist et al. 2004, Brines and Cerami 2008, Bohr et al. 2013). Further studies on EPO-derived substances are needed, to find optimal treatment modalities due to possible differences in the effectiveness and potency on bone healing. Longer-lasting EPO molecules especially offer theoretical advantages.

The risk of relevant side effects appears to be low if EPO is only used in a single dose, which was evident in the present study even with a very high single dose of EPO. Mean values of hematocrit, as a main parameter of the side effects of EPO, only increased in systemically treated animals until day 14. The pharmacological and pharmacokinetic characteristics of subcutaneously administered EPO with dosing once a week have been reported (Henry 2005) but these are hardly comparable to the situation in our animal model. Our finding that locally administered EPO resulted in less red blood cell production in the bone marrow may have different explanations. Although we used Gelita-Spon as a carrier substance in the radius defect, local application to the surgically opened defect carries a higher risk of loss of EPO during application,

Calcified callus bone volume (BV; mm³)



Figure 3. A. Quantitative micro-CT results for calcified callus, defined as bone volume (BV). BV was analyzed within the 15-mm radius defect in controls (squares) and animals with local EPO treatment (triangles) or systemic EPO treatment (circles). EPO was applied in one single dose at day 0 and in vivo micro-CT was performed at 4, 8, and 12 weeks after surgery and treatment. Data are mean (SD). B. Quantitative micro-CT results for total callus, defined as tissue volume (TV). TV was analyzed within the 15-mm radius defect in controls (squares) and animals with local EPO treatment (triangles) or systemic EPO treatment (circles). EPO was applied in one single dose at day 0 and in vivo micro-CT results for total callus, defined as tissue volume (TV). TV was analyzed within the 15-mm radius defect in controls (squares) and animals with local EPO treatment (triangles) or systemic EPO treatment (circles). EPO was applied in one single dose at day 0 and in vivo micro-CT was performed at 4, 8, and 12 weeks after surgery and treatment. Data are mean (SD). ^a p < 0.02

compared to use of a subcutaneous needle in tissue that has not been surgically opened. Secondly, EPO might be transported more quickly after local application in a bone defect with higher initial peak serum concentration but shorter elimination half-life.

Apart from the direct influence of EPO on bone, EPOinduced neovascularization is another important mechanism for promotion of bone healing. Our study revealed effects on neovascularization with significantly increased blood vessel formation after either local of systemic single-dose EPO treatment, and with a follow-up of 12 weeks. Considering the positive effect on bone formation expressed in the increase in BV and TV in quantitative in vivo micro-CT analysis, EPO treatment appears to have a rather immediate influence on bone



Figure 4. HE-stained tissue sections with a higher degree of vascularization after EPO treatment. The arrows show blood vessels in the callus after 12 weeks of follow-up.

formation, which is sustained over time during the 12 weeks of follow-up. Angiogenesis, however, was only evaluated by histology after 12 weeks, at final follow-up. One limitation of the study was that we did not analyze vascularization in the initial phase.

Possible negative effects of EPO on bone healing have been proposed by Singbrant et al. (2011), but this hypothesis has been dismissed by others (Shiozawa et al. 2010, Mc Gee et al. 2012, Sun et al. 2012), and we also found clear evidence of positive effects on bone formation.

It is a matter for debate whether EPO should be given on a daily or a weekly basis to have an effect on bone healing, or whether an initial single dosage is sufficient. The effect of EPO on bone healing appears to be dose-dependent, but the ideal dose has not yet been defined. Our findings only serve as a proof of principle, and it is not yet possible to draw conclusions on exact dosing. We used a very high single dose of EPO to prove that single application—either systemically or locally—to the defect can have an overall effect on bone formation.

Our results support the findings of other studies (Holstein et al. 2007, 2011, Garcia et al. 2011, Rölfing 2014, Rölfing et al. 2012, 2014a, 2014b) that EPO improves the early phase of bone healing in particular. Thus, single application of EPO at an early time point appears to be most effective—and from a clinical point of view, the intraoperative administration as a single dose appears most attractive to improve bone healing. Further studies will have to determine whether single-dose local application to the defect would allow further dose reduction compared to single-dose systemic treatment.

Continuous daily application of EPO has shown significant stimulation of erythropoiesis irrespective of whether highdose or low-dose therapy was used (Sun et al. 2012), so this would not be feasible for future clinical use due to erythropoiesis-related side effects.

The limitations of our study include the differences in pharmacodynamics, with higher metabolism in small animals such as rabbits and faster clearance of EPO than in humans. The dosage must therefore be adjusted, and this was addressed by using the metabolic body weight of the rabbits for calculation of the individual EPO dose. In principle, however, the same qualitative results would be expected in humans. Although far removed from the situation in humans, our rabbit study offered the advantage of larger tissues with longer diffusion distances for cell nutrition and higher biomechanical forces than if it had been based on small rodents.

In summary, our results suggest that an initial single dose of EPO, especially if applied locally, is sufficient to give increased bone healing with a reduced risk of side effects compared to repetitive EPO dosing. Further studies will be needed to evaluate the long-term outcome in animal models, and the first controlled pilot studies should now be initiated for evaluation of EPO-triggered bone healing in humans, since safety and tolerability data are already available and since EPO is well established in the treatment of other diseases.

- Anagnostou A, Lee ES, Kessimian N, Levinson R, Steiner M. Erythropoietin has a mitogenic and positive chemotactic effect on endothelial cells. Proc Natl Acad Sci U S A 1990; 87(15): 5978-82.
- Arcasoy M O. Non-erythroid effects of erythropoietin. Haematologica 2010; 95(11): 1803-5.
- Bozlar M, Kalaci A, Aslan B, Baktiroglu L, Yanat A N, Tasci A. Effects of erythropoietin on fracture healing in rats. Saudi Medical Journal 2006; 27(8): 1267-9.
- Bohr S, Patel S J, Shen K, Vitalo A G, Brines M, Cerami A, Berthiaume F, Yarmush M L. Alternative erythropoietin-mediated signaling prevents secondary microvascular throm-bosis and inflammation within cutaneous burns. Proc Natl Acad Sci U S A 2013; 110(9): 3513-8.
- Brines M, Cerami A. Erythropoietin-mediated tissue protection: reducing collateral damage from the primary injury response. J Intern Med 2008; 264(5): 405-32.
- Brines M, Grasso G, Fiordaliso F, Sfacteria A, Ghezzi P, Fratelli M, Latini R, Xie Q W, Smart J, Su-Rick C J, Pobre E, Diaz D, Gomez D, Hand C, Coleman T, Cerami A. Erythropoietin mediates tissue protection through an erythropoietin and common beta-subunit heteroreceptor. Proc Natl Acad Sci USA 2004; 101(41): 14907-12.
- Chateauvieux S, Grigorakaki C, Morceau F, Dicato M, Diederich M. Erythropoietin, erythropoiesis and beyond. Biochem Pharmacol 2011; 82(10):1291-303.
- Garcia P, Speidel V, Scheuer C, Laschke M W, Holstein J H, Histing T, Pohlemann T, Menger M D. Low dose erythropoietin stimulates bone healing in mice. J Orthop Res 2011; 29(2): 165–72.
- Geiger F, Bertram H, Berger I, Lorenz H, Wall O, Eckhardt C, Simank H G, Richter W. Vascular endothelial growth factor gene-activated matrix (VEGF165-GAM) enhances osteogenesis and angiogenesis in large segmental bone defects. J Bone Miner Res 2005; 20(11): 2028-35.
- Geiger F, Lorenz H, Xu W, Szalay K, Kasten P, Claes L, Augat P, Richter W. VEGF producing bone marrow stromal cells (BMSC) enhance vascularization and resorption of a natural coral bone substitute. Bone 2007; 41(4): 516-22.
- Goncalves S, Chiossone-Kerdel JA, Bianco AS, Ercolino J M, Hernandez-Rojas J. Effect of absorbable gelatin sponge in the middle ear: in vitro and in vivo animal model. Acta Otolaryngol 2015; 135(1): 14-25.
- Heeschen C, Aicher A, Lehmann R, Fichtlscherer S, Vasa M, Urbich C, Mildner-Rihm C, Martin H, Zeiher A M, Dimmeler S. Erythropoietin is a potent physiologic stimulus for endothelial progenitor cell mobilization. Blood 2003; 102(4): 1340-6.
- Henry D H. Epoetin alfa for the treatment of cancer- and chemotherapyrelated anaemia: product review and update. Expert Opin Pharmacother 2005; 6(2): 295-310.
- Hiram-Bab S, Liron T, Deshet-Unger N, Mittelman M, Gassmann M, Rauner M, Franke K, Wielockx B, Neumann D, Gabet Y. Erythropoietin directly stimulates osteoclast precursors and induces bone loss. FASEB J 2015; 29(5): 1890-900.
- Holstein J H, Menger M D, Scheuer C, Meier C, Culemann U, Wirbel R J, Garcia P, Pohlemann T. Erythropoietin (EPO): EPO-receptor signaling improves early endochondral ossification and mechanical strength in fracture healing. Life Sci 2007; 80(10): 893-900.
- Holstein J H, Orth M, Scheuer C, Tami A, Becker S C, Garcia P, Histing T, Mörsdorf P, Klein M, Pohlemann T, Menger M D. Erythropoietin stimulates bone formation, cell proliferation, and angiogenesis in a femoral segmental defect model in mice. Bone 2011; 49(5): 1037-45.
- Jaquet K, Krause K, Tawakol-Khodai M, Geidel S, Kuck K-H. Erythropoietin and VEGF exhibit equal angiogenic potential. Microvasc Res 2002; 64(2): 326-33.
- Jelkmann W. Erythropoietin: structure, control of production, and function. Physiol Rev 1992; 72(2): 449-89.
- Kleinschmidt K, Ploeger F, Nickel J, Glockenmeier J, Kunz P, Richter W. Enhanced reconstruction of long bone architecture by a growth factor mutant combining positive features of GDF-5 and BMP-2. Biomaterials 2013; 34(24): 5926-36.

GO: funding, planning, execution, analysis, interpretation of results, and writing of the manuscript. KK: execution, and analysis and interpretation of data, focusing on the micro-CT data. SG: planning and statistical analysis. AS: data acquisition, execution, and analysis of the data. TG: planning, analysis, and interpretation of the data. WR: planning, interpretation of data, and coordination of the study.

To support this study, institutional research funds were provided by the orthopaedic university hospital, Heidelberg, Germany. We thank technician Kathrin Brohm for her support during the whole study, especially regarding animal care, histology, and micro-CT analysis.

- Kleinschmidt K, Wagner-Ecker M, Bartek B, Holschbach J, Richter W. Superior angiogenic potential of GDF-5 and GDF-5(V453/V456) compared with BMP-2 in a rabbit long-bone defect model. J Bone Joint Surg Am 2014; 96(20):
- Leist M, Ghezzi P, Grasso G, Bianchi R, Villa P, Fratelli M, Savino C, Bianchi M, Nielsen J, Gerwien J, Kallunki P, Larsen A K, Helboe L, Christensen S, Pedersen L O, Nielsen M, Torup L, Sager T, Sfacteria A, Erbayraktar S, Erbayraktar Z, Gokmen N, Yilmaz O, Cerami-Hand C, Xie Q W, Coleman T, Cerami A, Brines M. Derivatives of erythropoietin that are tissue protective but not erythropoietic. Science 2004; 305(5681): 239-42.
- McGee SJ, Havens AM, Shiozawa Y, Jung Y, Taichman RS. Effects of erythropoietin on the bone microenvironment. Growth Factors 2012; 30(1): 22-8.
- Prunier F, Pfister O, Hadri L, Liang L, Del Monte F, Liao R, Hajjar RJ. Delayed erythropoietin therapy reduces post-MI cardiac remodeling only at a dose that mobilizes endothelial progenitor cells. Am J Physiol Heart Circ Physiol 2007; 292(1): H522-9.
- Rölfing J H D. The effect of erythropoietin on bone. Acta Orthop 2014; 85 (Suppl 353):1-27.

- Rölfing J H D, Bendtsen M, Jensen J, Stiehler M, Foldager CB, Hellfritzsch M B, Bünger C. Erythropoietin augments bone formation in a rabbit posterolateral spinal fusion model. J Orthop Res 2012; 30(7): 1083-8.
- Rölfing J H D, Jensen J, Jensen J N, Greve A-S, Lysdahl H, Chen M, Rejnmark L, Bünger C. A single topical dose of erythropoietin applied on a collagen carrier enhances calvarial bone healing in pigs. Acta Orthop 2014a; 85(2): 201-9.
- Rölfing J H D, Baatrup A, Stiehler M, Jensen J, Lysdahl H, Bünger C. The osteogenic effect of erythropoietin on human mesenchymal stromal cells is dose-dependent and involves non-hematopoietic receptors and multiple intracellular signaling pathways. Stem Cell Rev 2014b; 10(1): 69-78.
- Shiozawa Y, Jung Y, Ziegler AM, Pedersen EA, Wang J, Wang Z, Song J, Wang J, Lee CH, Sud S, Pienta KJ, Krebsbach PH, Taichman RS. Erythropoietin couples hematopoiesis with bone formation. PLoS ONE 2010; 5(5): e10853.
- Sun H, Jung Y, Shiozawa Y, Taichman R S, Krebsbach P H. Erythropoietin modulates the structure of bone morphogenetic protein 2-engineered cranial bone. Tissue Eng Part A 2012; 18(19-20): 2095-105.