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Update of the list of QPS-recommended biological agents intentionally added to food or feed as notified to EFSA 10: Suitability of taxonomic units notified to EFSA until March 2019

EFSA Panel on Biological Hazards (BIOHAZ),

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Abstract

The qualified presumption of safety (QPS) procedure was developed to provide a harmonised generic preevaluation to support safety risk assessments of biological agents performed by EFSA's Scientific Panels. The taxonomic identity, body of knowledge, safety concerns and antimicrobial resistance were assessed. Safety concerns identified for a taxonomic unit (TU) are, where possible and reasonable in number, reflected by 'qualifications' which should be assessed at the strain level by the EFSA's Scientific Panels. During the current assessment, no new information was found that would change the previously recommended OPS TUs and their qualifications. The list of microorganisms notified to EFSA from applications for market authorisation was updated with 47 biological agents, received between October 2018 and March 2019. Of these, 19 already had QPS status, 20 were excluded from the QPS exercise by the previous QPS mandate (11 filamentous fungi) or from further evaluations within the current mandate (9 notifications of Escherichia coli). Sphingomonas elodea, Gluconobacter frateurii, Corynebacterium ammoniagenes, Corynebacterium casei, Burkholderia ubonensis, Phaeodactylum tricornutum, Microbacterium foliorum and Euglena gracilis were evaluated for the first time. Sphingomonas elodea cannot be assessed for a possible OPS recommendation because it is not a valid species. Corvnebacterium ammoniagenes and Euglena gracilis can be recommended for the QPS list with the qualification 'for production purposes only'. The following TUs cannot be recommended for the QPS list: Burkholderia ubonensis, due to its potential and confirmed ability to generate biologically active compounds and limited of body of knowledge; Corynebacterium casei, Gluconobacter frateurii and Microbacterium foliorum, due to lack of body of knowledge; Phaeodactylum tricornutum, based on the lack of a safe history of use in the food chain and limited knowledge on its potential production of bioactive compounds with possible toxic effects.

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Keywords: safety, QPS, bacteria, yeast, *Corynebacterium ammoniagenes*, *Gluconobacter frateurii*, *Sphingomonas elodea*, *Burkholderia ubonensis*, *Phaeodactylum tricornutum*, *Euglena gracilis*, *Microbacterium foliorum*, *Corynebacterium casei*

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Amendment: By error, the name of an author has been omitted in the previous version and has been added in this version on pages 1 and 2 – in the 'Suggested citation' paragraph. These editorial corrections do not materially affect the contents or outcome of this scientific output. To avoid confusion, the older version has been removed from the EFSA Journal, but is available on request, as is a version showing all the changes made.

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Summary

The European Food Safety Authority (EFSA) asked the Panel on Biological Hazards (BIOHAZ) to deliver a Scientific Opinion on the maintenance of the list of qualified presumption of safety (QPS) biological agents intentionally added to food or feed. The request included three specific tasks as mentioned in the Terms of Reference (ToR).

The QPS process was developed to provide a harmonised generic pre-evaluation procedure to support safety risk assessments of biological agents performed by EFSA's scientific Panels and Units. The taxonomic identity, body of knowledge and safety of biological agents are assessed. Safety concerns identified for a taxonomic unit (TU) are, where possible and reasonable in number, reflected as 'qualifications' that should be assessed at the strain level by the EFSA's scientific Panels. A generic qualification for all QPS bacterial TUs applies in relation to the absence of acquired genes conferring resistance to clinically relevant antimicrobials (EFSA, 2008).

The overall summary of the re-evaluation of TUs previously recommended for the QPS list is undertaken every 3 years in a scientific Opinion of the BIOHAZ Panel. Meanwhile, the list of microorganisms is maintained and re-evaluated approximately every 6 months in a Panel Statement. If new information is retrieved from extended literature searches that would change the QPS status of a microbial species or its qualifications, this is published in the Panel Statement. The Panel Statement also includes the evaluation of microbiological agents newly notified to EFSA within the 6-month period for an assessment for use as feed additives, food enzymes, food additives and flavourings, novel foods or plant protection products (PPPs). The main results of the assessments completed from 2017 onwards will be included in the scientific Opinion of the BIOHAZ Panel to be published by the end of the current mandate in December 2019. In the interim, as a result of each Panel Statement, the '2016 updated list of QPS status recommended biological agents for safety risk assessments carried out by EFSA scientific Panels and Units' is extended by the inclusion of new recommendations for QPS status, and appended to the Opinion adopted in December 2016 (Appendix E).

The *first ToR* requires ongoing updates of the list of biological agents notified to EFSA, in the context of a technical dossier, for intentional use in food and/or feed or as sources of food and feed additives, enzymes and PPPs for safety assessment. The list was updated with the notifications received since the latest review in October 2018. Within this period, 47 notifications were received by EFSA, of which 32 were for feed additives, two for food enzymes, food additives and flavourings, 10 for novel foods and 3 for PPPs. The new notifications, received between October 2018 and March 2019, are included in a table appended to the current Statement (Appendix F).

The *second ToR* concerns the revision of the TUs previously recommended for the QPS list and their qualifications when new information has become available, and the updating of the information provided in the previous Opinion adopted in December 2016. According to the articles retrieved through an extensive literature search (ELS), for articles published from October 2018 to March 2019 no new information was found that would affect the QPS status of those TUs and their qualifications.

The *third ToR* requires a (re)assessment of TUs notified to EFSA, but not present in the current QPS list, for their suitability for inclusion in the updated list. The current Statement focuses on the assessments of the TUs that were notified to EFSA between October 2018 and March 2019. Of the 47 notifications received, 19 biological agents already had QPS status and did not require further evaluation in this Statement and 20 were not included because: 11 were notifications of filamentous fungi that were excluded from the QPS exercise; nine were notifications of *Escherichia coli* that were excluded from further QPS evaluations within the current QPS mandate. Eight new TUs were considered for the QPS assessment within this Statement: *Sphingomonas elodea, Corynebacterium ammoniagenes, Corynebacterium casei, Gluconobacter frateurii, Burkholderia ubonensis, Phaeodactylum tricornutum, Euglena gracilis and Microbacterium foliorum* which were evaluated for the first time.

Sphingomonas elodea could not be assessed for a possible QPS recommendation because it is currently not a valid species. Corynebacterium ammoniagenes and Euglena gracilis can be recommended for the QPS list with the qualification 'for production purposes only'. The following TUs cannot be recommended for the QPS list: Burkholderia ubonensis, due to its potential ability to generate biologically active compounds and limited body of knowledge; Corynebacterium casei, Gluconobacter frateurii and Microbacterium foliorum, due to lack of body of knowledge; Phaeodactylum tricornutum, based on the lack of a safe history of use in the food chain and limited knowledge on its potential production of bioactive compounds with toxic effects.

Pediococcus dextrinicus (Coster and White, 1964) Back, 1978 *species* has been changed to *Lactobacillus dextrinicus* (Coster and White, 1964) Haakensen et al., 2009, comb. nov. It will be updated in the QPS list.



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1. Introduction

The qualified presumption of safety (QPS) approach was developed by the EFSA Scientific Committee to provide a generic concept to prioritise and to harmonise risk assessment within the European Food Safety Authority (EFSA) of microorganisms intentionally introduced into the food chain, in support of the respective Scientific Panels and Units in the frame of market authorisations (EFSA, 2007). The list, first established in 2007, has been continuously revised and updated. The publication of the overall assessment of the taxonomic units (TUS) previously recommended for the QPS list is to be evaluated every 3 years through a scientific Opinion by the Panel on Biological Hazards (BIOHAZ). Intermediate deliverables in the form of a Panel Statement are produced and published for periods of around 6 months, should an assessment for a QPS classification of a microbiological agent notified to EFSA be requested by the Units dealing with feed additives, food enzymes, food additives and flavourings, novel foods, or plant protection products. These Panel Statements also include the results of the assessment of the relevant new papers related to the TUs with QPS status.

1.1. Background and Terms of Reference as provided by EFSA

1.1.1. Background as provided by EFSA

A wide variety of microorganisms are intentionally added at different stages into the food and feed chain. In the context of applications for market authorisation of these biological agents, used either directly or as sources of food and feed additives, food enzymes and plant protection products, EFSA is requested to assess their safety.

Several taxonomic units (usually species for bacteria and yeasts, families for viruses) have been included in the qualified presumption of safety (QPS) list either following notifications to EFSA or proposals made initially by stakeholders during a public consultation in 2005, even if they were not yet notified to EFSA (EFSA, 2005).¹ The EFSA Scientific Committee reviewed the range and numbers of microorganisms likely to be the subject of an EFSA Opinion and published in 2007 a list of microorganisms recommended for the QPS list.²

In 2007, the Scientific Committee recommended that a QPS approach should provide a generic concept to prioritise and to harmonise safety risk assessment of microorganisms intentionally introduced into the food chain, in support of the respective Scientific Panels and EFSA Units in the frame of the market authorisations. The same Committee recognised that there would have to be continuing provision for reviewing and modifying the QPS list and in line with this recommendation, the EFSA Scientific Panel on Biological Hazards (BIOHAZ) took the prime responsibility for this and started reviewing annually the existing QPS list. The first annual QPS update³ was published in 2008 and EFSA's initial experience in applying the QPS approach was included. The potential application of the QPS approach to microbial plant protection products was discussed in the 2009 update.⁴ Also in 2009, bacteriophages were assessed and were not considered appropriate for the QPS list. After consecutive years of reviewing the existing scientific information, the filamentous fungi (2008 to 2013 updates) and enterococci (2010 to 2013 updates) were not recommended for the QPS list. The 2013 update⁵ of the recommended QPS list included 53 species of Gram-positive non-spore-forming bacteria, 13 Gram-positive spore forming bacteria (*Bacillus* species), one Gram-negative bacterium (*Gluconobacter oxydans*), 13 yeast species, and three virus families.

In 2014 the BIOHAZ Panel, in consultation with the Scientific Committee, decided to change the revision procedure: the overall assessment of the taxonomic units previously recommended for the QPS list is no longer carried out annually but over 3-year periods. From 2017, the search and revision of the possible safety concerns linked to those taxonomic units start to be done every 6 months period. The revision of the 2013 update (EFSA Biohaz Panel, 2013) was updated in 2016 (EFSA BIOHAZ Panel, 2017a)

¹ Opinion of the Scientific Committee on a request from EFSA related to a generic approach to the safety assessment by EFSA of microorganisms used in food/feed and the production of food/feed additives. The EFSA Journal 2005, 226, 1–12.

² Introduction of a Qualified Presumption of Safety (QPS) approach for assessment of selected microorganisms referred to EFSA – Opinion of the Scientific Committee. The EFSA Journal 2007, 293, 1–85.

³ Scientific Opinion of the Panel on Biological Hazards on a request from EFSA on the maintenance of the list of QPS microorganisms intentionally added to food or feed. The EFSA Journal 2008, 923, 1–48.

⁴ Scientific Opinion of the Panel on Biological Hazards (BIOHAZ) on the maintenance of the list of QPS microorganisms intentionally added to food or feed (2009 update). EFSA Journal 2009; 7(12):1431, 92 pp. https://doi.org/10.2903/j.efsa.2009.1431

⁵ EFSA BIOHAZ Panel (EFSA Panel on Biological Hazards), 2013. Scientific Opinion on the maintenance of the list of QPS biological agents intentionally added to food and feed (2013 update). EFSA Journal 2013;11(11):3449, 107 pp. https://doi.org/ 10.2903/j.efsa.2013.3449

and the next update will be published in a scientific Opinion of the BIOHAZ Panel after its adoption in December 2019.⁶ The QPS list of microorganisms has been maintained and frequently checked, based on the evaluation of extensive literature searches. In the meantime and every 6 months, a Panel Statement, compiling the assessments for a QPS status of the microbiological agents notified to EFSA requested by the Feed Unit, the Food Ingredients and Packaging (FIP) Unit, the Nutrition Unit or by the Pesticides Unit, has been produced and published. In the follow up of the 2013 update⁵ the Scientific Committee agreed to exclude some biological groups (filamentous fungi, bacteriophages and *Enterococcus faecium*⁷) notified to EFSA from the QPS assessment because it was considered unlikely that any taxonomical units within these groups would be granted QPS status in the foreseeable future. Thus, the assessment of members of these biological groups needs to be done at a strain level, on a case-by-case basis, by the relevant EFSA Unit.

The QPS provides a generic safety pre-assessment approach for use within EFSA that covers risks for human, animals and the environment. In the QPS concept a safety assessment of a defined taxonomic unit is considered independently of any particular specific notification in the course of an authorisation process. The OPS concept does not address hazards linked to the formulation or other processing of the products containing the microbial agents and added into the food or feed chain. Although general human safety is part of the evaluation, specific issues connected to type and level of exposure of users handling the product (e.g. dermal, inhalation, ingestion) are not addressed. In the case Genetically Modified Microorganisms (GMMs) for which the species of the recipient strain qualifies for the QPS status, and for which the genetically modified state does not give rise to safety concerns, the QPS approach can be extended to genetically modified production strains (EFSA BIOHAZ Panel, 2018a).⁸ Assessment of potential allergenicity to microbial residual components is beyond the QPS remit; if there is however, science-based evidence for some microbial species it is reported. Where applicable these aspects are assessed, separately by the EFSA Panel responsible for assessing the notification. Antimicrobial resistance was introduced as a possible safety concern for the assessment of the inclusion of bacterial species in the QPS list published in 2008 QPS Opinion (EFSA, 2008)³. In the 2009 QPS Opinion (EFSA BIOHAZ Panel, 2009)⁴ a qualification regarding the absence of antimycotic resistance for yeasts was introduced.

1.1.2. Terms of Reference as provided by EFSA

The Terms of Reference, as provided by EFSA are as follows:

ToR 1: Keep updated the list of biological agents being notified in the context of a technical dossier to EFSA Units such as Feed, Pesticides, Food Ingredients and Packaging (FIP) and Nutrition, for intentional use directly or as sources of food and feed additives, food enzymes and plant protection products for safety assessment.

ToR 2: Review taxonomic units previously recommended for the QPS list and their qualifications when new information has become available. The latter is based on a review of the updated literature aiming at verifying if any new safety concern has arisen that could require the removal of the taxonomic unit from the list, and to verify if the qualifications still efficiently exclude safety concerns.

ToR 3: (Re) assess the suitability of new taxonomic units notified to EFSA for their inclusion in the QPS list. These microbiological agents are notified to EFSA and requested by the Feed Unit, the FIP Unit, the Nutrition Unit or by the Pesticides Unit.

1.2. Interpretation of the Terms of Reference

The absence of acquired genes conferring resistance to clinically relevant antimicrobials is a qualification⁹ applied to all QPS bacterial TUs. The verification of such qualification is under the remit of the Unit conducting the safety assessment of the organism notified to EFSA for market authorisation, and is therefore done at strain level (EFSA BIOHAZ Panel, 2017a).

⁶ References updated from the original self-task mandate.

⁷ The taxonomic unit was corrected from the original mandate: 'enterococci'. It is only referred to *Enterococcus faecium*, the only species which was evaluated for a possible QSP status.

⁸ Sentence included, correcting the previous sentence from the original self-task mandate: 'Genetically modified microorganisms are similarly not taken into account'.

⁹ Identified safety concerns, including acquired antimicrobial resistance genes, for a certain TU can be, where reasonable in number and not universally present, reflected as 'qualifications'.

In June 2017 (EFSA BIOHAZ Panel, 2017b), the BIOHAZ Panel has agreed to exclude *Escherichia coli* and any species of the genus *Streptomyces* from QPS evaluation within this mandate.

In June 2018 (EFSA BIOHAZ Panel, 2018b), the BIOHAZ Panel clarified that the qualification 'for production purpose only' implies the absence of viable cells of the production organism in the final product and can also be applied for food and feed products based on microbial biomass.

2. Data and methodologies

2.1. Data

Only valid TUs covered by the relevant international committees on the nomenclature for microorganisms are considered for the QPS assessment.

In reply to ToR 3, (re)assessment of the suitability of TUs notified within the time period covered by this Statement (from October 2018 to March 2019) is carried out. The literature review considered the identification, the body of knowledge, the potential safety concerns, and the knowledge on acquired antimicrobial resistance (AMR). Relevant databases, such as PubMed, Web of Science, Cases Database, CAB Abstracts or Food Science Technology Abstracts (FSTA) and Scopus, were searched. More details on the search strategy, search keys, and approach are described in Appendix A.

In reply to ToR 2, concerning the revision of the TUs previously recommended for the QPS list and their qualifications, an extensive literature search (ELS) was conducted as described in Appendices B and C.

2.2. Methodologies

2.2.1. Evaluation of a QPS recommendation for taxonomic units notified to EFSA

In response to ToR 1, the EFSA Units were asked to update the list of biological agents being notified to EFSA. A total of 47 notifications were received between October 2018 and March 2019, of which 32 were for a feed additive, 2 for food enzymes, 10 for novel foods and 3 for plant protection products (Table 1).

In response to ToR 3, out of the 47 notifications, 19 were related to TUs that already had QPS status and did not require further evaluation. Of the remaining 26 notifications, 20 were related to TUs not evaluated for a QPS status for the following reasons:

- eleven notifications related to filamentous fungi, which were excluded from QPS evaluations in the follow-up of a recommendation of the QPS 2013 and 2016 updates (EFSA BIOHAZ Panel, 2013, 2014, 2016);
- nine notifications related to *E. coli*, which were recently excluded from the current mandate by the BIOHAZ Panel.

The TUs corresponding to the remaining eight notifications evaluated for a possible QPS recommendation:

• Sphingomonas elodea, Gluconobacter frateurii, Corynebacterium ammoniagenes, Corynebacterium casei, Burkholderia ubonensis, Phaeodactylum tricornutum, Microbacterium foliorum and Euglena gracilis, all evaluated for the first time.

The notifications received by EFSA, per risk assessment area, by biological group from October 2018 to March 2019, are presented in Table 1.

Table 1:	Notifications received by EFSA, per risk assessment area and by biological group, from	
	October 2018 and March 2019	

Risk assessment area		luated in this atement	Evaluated in this		
Biological group	Already QPS	Excluded in QPS ^(a)	Statement	Total	
Feed additives	15	13	4	32	
Bacteria	9	4	4	17	
Filamentous fungi	0	9	0	9	
Yeasts	6	0	0	6	



Risk assessment area		luated in this atement	Evaluated in this		
Biological group	Already QPS	Excluded in QPS ^(a)	Statement	Total	
Novel foods	2	5	3	10	
Bacteria	0	5	1	4	
Yeasts	2	0	0	2	
Algae	0	0	2	2	
Plant protection products	2	1	0	3	
Bacteria	1	0	0	1	
Filamentous fungi	0	1	0	1	
Viruses	1	0	0	1	
Food enzymes, food additives and flavourings	0	1	1	2	
Bacteria	0	0	1	1	
Filamentous fungi	0	1	0	1	
Total	19	20	8	47	

QPS: qualified presumption of safety.

(a): The number includes 11 notifications of filamentous fungi excluded from QPS evaluation in the 2013 QPS Opinion and 9 notifications of *E. coli* (bacterium) already excluded in the Panel Statement adopted in December 2016 (EFSA BIOHAZ Panel et al., 2017a).

2.2.2. Use of MLT in the context of the yeasts and Bacillus taxonomic units

To explore the potential application of a machine learning technique (MLT) for screening papers in the context of the QPS project the performances of such technique were assessed against the previous batch of papers retrieved for the Bacillus and Yeasts taxonomic units.

To that purpose, the DistillerAI Toolkit included in the DistillerSR online software was used.

DistillerAI 'Preview and Rank' function was used mapping the papers from 'Title screening' to 'Article evaluation'. The SVM algorithm with 100% training set and 100% references to preview was used and the references were subsequently tagged. The algorithm was trained on the combined results of the 2 reviewers in the QPS rounds from 1 June 2016 to 31 December 2017. This is considered a conservative approach since, in the case of conflicts among the experts, the algorithm considers the paper as relevant.

The MLT predicted screening results on the batch of papers corresponding to the period January– June 2018 were obtained and compared with the results obtained by the two reviewers in the real exercise.

The results of the exercise showed that, in the case of yeasts, MLT had around 88% and 80% of sensitivity and specificity, respectively, while, in the case of *Bacillus*, MLT had 100% and about 82% of sensitivity and specificity, respectively. Moreover, it was found that in case of using the MLT algorithm as a reviewer in parallel with a human reviewer, in both projects no information relevant for the QPS status would have been missed.

On the basis of these results and considering the high number of papers retrieved for both yeasts and *Bacillus* in the context of the QPS exercise, it was decided to use the MLT in parallel with a human reviewer to screen the current batch of papers in these two TUs.

As expected, considering its specificity, the application of the MLT algorithm resulted in a high number of potentially relevant papers at the end of the screening phase. On the other hand, the algorithm did not miss any paper identified as potentially relevant by the human reviewer.

2.2.3. Monitoring of new safety concerns related to the QPS list

The aim of the ELS carried out in response to ToR 2 (review of the recommendations for the QPS list and specific qualifications) was to identify any publicly available studies reporting on safety concerns for humans, animals or the environment caused by QPS organisms since the previous QPS review (i.e. publications from July to December 2018). For a detailed protocol of the process and search strategies, refer to Appendices B and C.



After removal of duplicates, 3,710 records were submitted to the *title screening* step, which led to the exclusion of 3,179 of them. The remaining 531 records were found eligible for the *Title and abstract screening* step, which led to the exclusion of 39 of these. Of the 492 articles that finally reached the *Article evaluation step* (full text), 85 were considered to be relevant for the QPS project.

The flow of records from their identification by the different search strategies (as reported in Appendix C) to their consideration as potentially relevant papers for QPS is shown in Table 2.

Table 2: Flow of records by search strategy

	No of papers						
Species	Title screening step	Title/ abstract screening step	Article evaluation step (screening for potential relevance) ^(a)	Article evaluation step (identification of potential safety concerns)			
Bacteria	2,244	258	224	11			
Bacillus spp.	804	199	199 ^(b)	2			
Geobacillus stearothermophilus			0	0			
Bifidobacterium spp.	206	21	3	1			
Carnobacterium divergens			0	0			
Corynebacterium glutamicum	45	0	0	0			
Gluconobacter oxydans	115	1	0	0			
Xanthomonas campestris			0	0			
Lactobacillus spp.	555	22	12	3			
Lactococcus lactis	173	6	4	3			
Leuconostoc spp.	68	5	4	1			
Microbacterium imperiale			0	0			
Oenococcus oeni	24	1	0	0			
Pasteuria nishizawae			0	0			
Pediococcus spp.	146	2	2	1			
Propionibacterium spp.	27	0	0	0			
Streptococcus thermophilus	81	1	0	0			
Viruses	108	0	0	0			
Alphaflexiviridae	39	0	0	0			
Potyviridae				0			
Baculoviridae	69	0	0	0			
Yeasts	1,358	273	268 ^(b)	30 ^(c)			
<i>Candida famata</i> (teleomorph = <i>Debaryomyces hansenii</i>)	1,358	273	268	10			
<i>Candida kefyr</i> (teleomorph = <i>Kluyveromyces marxianus</i>)				16			
Candida pelliculosa (synonymus = Pichia anomala) (teleomorph = Wickerhamomyces anomalus)				6			
<i>Candida utilis (</i> teleomorph = <i>Lindnera</i> <i>jadinii)</i>				1			
Hanseniaspora uvarum				1			
Saccharomyces cerevisiae including Saccharomyces boulardii				8			
Kluyveromyces lactis				1			
Schizosaccharomyces pombe				1			
Total	3,710	531	492	85			
Excluded	3,179	39	407				

(a): Relevant references in Appendix D.

(b): The relatively high number of hits is linked to the procedure followed for the screening of references using machine learning techniques (MLT) in parallel with experts' selection.

(c): 30 articles describing 45 studies related to different yeast species. In one case, there was no reference to any particular species.



3. Assessment

3.1. Taxonomic units evaluated during the previous QPS mandate and re-evaluated in the current Statement

None.

3.2. Taxonomic units to be evaluated for the first time

3.2.1. Bacteria

3.2.1.1. Burkholderia ubonensis

Identity

B. ubonensis is a species with Standing in Nomenclature. *B. ubonensis* belongs to the *Burkholderia cepacia* complex (Martina et al., 2018) that was first described as *Burkholderia uboniae* (Yabuuchi et al., 2000) and later on renamed as *B. ubonensis* (Anonymous, 2000).

Body of knowledge

There are only a limited number of scientific papers published on this TU. *B. ubonensis* as well as other *Burkholderia* species, produce lipases, which are used in a broad range of industrial applications.

Safety concerns

B. ubonensis is considered to be non-pathogenic, although its virulence potential has not been extensively tested. It appears to be innocuous after a subcutaneous challenge to mice (Price et al., 2017).

Whole genome characterisation of strains within the species revealed the presence of numerous genes encoding secondary, biologically active, metabolites such as polyketides, non-ribosomal peptides, quinolines and pyrrolnitrins (Loveridge et al., 2017). Some produce monobactams (Imada et al., 1981), bulgecins (Horsman et al., 2017), which enhance the bactericidal effect of β -lactams on Gram negative bacteria, and bacteriocin-like compounds (Marshall et al., 2010).

Antimicrobial resistance aspects

Different features, namely the external membrane structure and an inducible class A β -lactamase contributes to the intrinsic resistance to diverse antibiotics (Rhodes and Schweizer, 2016)

Conclusions on a recommendation for the QPS list

B. ubonensis cannot be recommended for the QPS list due to its ability to generate biologically active compounds and limited body of knowledge.

3.2.1.2. Corynebacterium ammoniagenes

Identity

C. ammoniagenes is a species with Standing in Nomenclature. It was first described by Cooke and Keith (1927) as *Brevibacterium ammoniagenes*, an urea-splitting bacterium isolated from the human intestinal tract. It was transferred to the genus *Corynebacterium* by Collins (1987).

Body of knowledge

C. ammoniagenes is used for the industrial production of nucleotides, nucleosides and riboflavin (Koizumi et al., 2000; Serrano et al., 2017).

C. ammoniagenes derived single-cell protein¹⁰ can also be used as a non-conventional protein source in animal diet such as in broilers (An et al., 2018) and growing pigs (Wang et al., 2013), without any negative effects on blood, bone characteristics or meat quality (An et al., 2018).

¹⁰ Single-cell protein (SCP) typically refers to mixed protein extracted from pure and mixed cultures of microorganisms like yeasts, bacteria, fungi and algae, grown in large scale culture systems (Nasseri et al., 2011). SCP can be produced using a variety of raw substrates from inexpensive agro-industrial by-products and can be used in poultry feed as an alternative protein source to soya bean meal or fishmeal (El-Deek et al., 2009).



Safety concerns

No information was found in relation to pathogenicity of the organism, including when used for production of SCP for feed (Oliveira et al., 2017).

Antimicrobial resistance aspects

No information was found in relation to the AMR.

Conclusions on a recommendation for the QPS list

C. ammoniagenes can be recommended for QPS list with the qualification 'for production purposes only'.

3.2.1.3. Corynebacterium casei

Identity

C. casei was first isolated from the surface of a smear-ripened cheese (Brennan et al., 2001) and is a valid species according to the list of prokaryotic names with Standing in Nomenclature.

Body of knowledge

There are only a limited number of articles published on this TU. Most of the relevant articles described the microbial communities and activities in ripened cheese where *C. casei* contributes as a spontaneous contaminant to the production of the desired organoleptic properties (Brennan et al., 2002; Hannon et al., 2004; Mounier et al., 2005; Rea et al., 2007; Bockelmann, 2011; Cogan et al., 2014; Bertuzzi et al., 2017).

Safety concerns

One article described the identification of *C. casei* among 57 bacterial strains isolated during the screening of blood samples of patients with cardiovascular diseases and not having any active infection (Dinakaran et al., 2012), so no proven connection with the ongoing diseases was identified.

Antimicrobial resistance aspects

No information was found in the literature.

Conclusions on a recommendation for the QPS list

C. casei cannot be recommended to the QPS list due to lack of body of knowledge.

3.2.1.4. Gluconobacter frateurii

Identity

It is a species with Standing in Nomenclature (Mason and Claus, 1989). It was separated from *Acetobacter* in 1989. It has been classified as a separate cluster in the *Gluconobacter* genus according to 16S rRNA gene sequence analysis (Sievers et al., 1995) and to restriction analysis of 16S-23S rDNA internal transcribed spacer regions (Malimas et al., 2006).

Body of knowledge

There are only a limited number of articles published on this TU. It can produce high amounts of fructans and can efficiently produce glyceric acid from raw glycerol (Poljungreed and Boonyarattanakalin, 2018). It is an obligate aerobic acetic acid bacterium that is found as a spontaneous contaminant as part of the microbial community in some food processes (e.g. vinegar production and kefir, cocoa and coconut water fermentation) (Korsak et al., 2015; Ho et al., 2018). Although these foods are consumed and exposure of humans and animals to this microorganism is expected, the scientific knowledge on this species is limited.

Safety concerns

The literature search retrieved no hits on virulence, pathogenicity or disease related to this TU.

Antimicrobial resistance aspects

No information is available in the scientific literature.

Conclusions on a recommendation for the QPS list

Gl. frateurii cannot be recommended for the QPS list due to lack of body of knowledge.

3.2.1.5. Microbacterium foliorum

Identity

M. foliorum is a valid species name according to the List of Prokaryotic Names with Standing in Nomenclature. It was first isolated as a plant associated bacterium (Behrendt et al., 2001).

Body of knowledge

There are only a limited number of articles published on this TU. *M. foliorum* is a saprophytic organism that may become endophytic and allow cruciferous plants to grow on oil seep soils, due to its capacity to process hydrocarbons such as catechol, toluene, naphthalene, octanol and others (Lumactud et al., 2016, 2017). In addition, it is, as a spontaneous contaminant, part of the microbiota involved in production of flavour and aroma in surface-ripened cheeses, such as Limburger and Munster, due to its caseinolytic, aminopeptidase and deaminase activities (Deetae et al., 2007, 2009). Although these foods are consumed and exposure of humans to this microorganism is expected, the scientific knowledge on this species is still limited.

Safety concerns

M. foliorum has been associated with clinical samples. Laffineur et al. (2003) reported the isolation of 30 *Microbacterium* spp. specimens 'during the past decades' one of which was classified as *M. foliorum*, with no further indication of its source or the conditions of the patient. Gneiding et al. (2008) identified 50 isolates as belonging to the genus *Microbacterium* in a survey of bacteria obtained during the previous 5 years from pathological samples and processed in a German Coryneform reference laboratory. Unfortunately, only the origin of the samples was reported, with no indication of the patient conditions or the procedures to which they were subjected. Of these isolates, seven were ascribed to *M. foliorum*, their origin being wound swabs (5), pleural fluid (1) and blood (1).

A toxicological study was made on lysed cultures from a single strain without adverse effects, but the results cannot be extrapolated to the species (Kim et al., 2018a).

Antimicrobial resistance aspects

One paper (Gneiding et al., 2008) reported susceptibility profiles of *M. foliorum* but without reference to transmissibility of resistances.

Conclusions on a recommendation for the QPS list

M. foliorum cannot be recommended for the QPS list due to lack of body of knowledge.

3.2.1.6. *Sphingomonas elodea*

Identity

S. elodea was described as a Gram-negative bacterium, initially named as *Pseudomonas elodea* (Kang et al., 1982). This species has not been taxonomically validated according to the List of Prokaryotic Names with Standing in Nomenclature (LPSN) (Euzéby, 2013) (http://www.bacterio.net/-allnamesdl. html) and the modifications that appear in the International Journal of Systematic and Evolutionary Microbiology (IJSEM) (Oren and Garrity, 2015).

Body of knowledge

Not applicable.

Safety concerns

Not applicable.

Antimicrobial resistance aspects

Not applicable.

Conclusions on a recommendation for the QPS list

S. elodea could not be assessed for a possible QPS recommendation because it is not a valid species.

3.2.2. Algae

3.2.2.1. Euglena gracilis

Identity

E. gracilis belongs to the genus *Euglena,* phototrophic euglenoid flagellates possessing complex chloroplasts. The taxonomy of *E. gracilis* and closely related species has not been amended so far based on molecular phylogenetic analyses (Zakryś et al., 2017). The whole genome of the *E. gracilis* strain Z1 was recently sequenced (Ebenezer et al., 2019).

Body of knowledge

There are an extended amount of scientific papers published on this TU. *E. gracilis* is found in many freshwater habitats, especially in shallow eutrophic ponds. The species is able to synthesise biotechnologically relevant compounds such as polyunsaturated fatty acids, vitamins, β -glucans and tyrosine used in cosmetics and food supplements. *E. gracilis* is also used for bioremediation of heavy metals in contaminated water and as a toxicity bioindicator (Krajčovič et al., 2015).

E. gracilis biomass, generally based on dried cells, is used as a feed additive in aquaculture and in animal feed as well as in human food (Suzuki, 2017). Food products containing *E. gracilis* are marketed in Japan as cookies, cereal bars and nutritional drinks.

Safety concerns

Using dried preparations of non-viable *E. gracilis*, no genotoxicity was observed in bacterial reverse mutation and mammalian micronucleus tests. Moreover, subchronic toxicity tests in rats did not show any adverse effect and a no-observed-adverse-effect-level (NOAEL) of 50,000 ppm was determined. (Simon et al., 2016).

Literature searches did not provide any evidence for a safety concern for human or animal health for any use of *E. gracilis*.

Antimicrobial resistance aspects

Not applicable.

Conclusions on a recommendation for the QPS list

E. gracilis may be recommended for the QPS list with the qualification 'for production purposes only'.

3.2.2.2. Phaeodactylum tricornutum

Identity

The unicellular, photosynthetic alga *P. tricornutum* was first isolated and described as a new species by the end of the 19th century (Bohlin, 1897). It is a pennate diatom (class Bacillariophyceae) and the only known species in the genus. It is an unusual diatom since it is pleiomorphic (fusiform, oval or triradiate morphology) and production of a silica frustule is facultative. Identification has recently been made by 18S rRNA gene sequencing (Demirel, 2016; Haro et al., 2017). The genome of one isolate has been sequenced (Bowler et al., 2008; Rastogi et al., 2018).

Body of knowledge

There are an extended amount of scientific papers published on the TU. *P. tricornutum* has been isolated from marine or brackish locations, often in benthic habitats, and appears to have a global distribution (De Martino et al., 2007). However, it is not a dominant species of algal communities and relatively little is known about its ecology.

It is of biotechnological interest, for studies within algal physiology and metabolism and as a model test species in toxicology. The potential of *P. tricornutum* as a source of biomass for biofuels, antimicrobial agents, health-promoting substances and food or feed components in general is subject to intensive study (Bajpai, 2016; Garcia et al., 2017; Haro et al., 2017; Shah et al., 2018). For example, the alga produces many compounds of high nutritional value, such as essential fatty acids and carotenoids (Zhang et al., 2015; Garcia et al., 2017). It has also been shown that *P. tricornutum* biomass can be used as a feed supplement (Skrede et al., 2011; Cerezuela et al., 2012; Sørensen et al., 2016).



Although several scientific studies are available, there appears to be no history of use of *P. tricornutum* in food or feed in practice, since no evidence was found that *P. tricornutum*, or substances produced by it, have yet been commercialised and/or widely consumed.

Safety concerns

The literature search did not reveal any studies reporting infection or intoxication by *P. tricornutum*. Niccolai et al. (2017) tested *in vitro* toxicity of water- or methanol extracts of *P. tricornutum* (strain F&M-M40) and found them to be toxic to fibroblasts. Neumann et al. (2018a) showed cytotoxic effects on human peripheral mononuclear cells.

In another study, Neumann et al. (2018b) investigated the bioavailability and toxicity of disrupted and freeze-dried cells of *P. tricornutum* in a mouse model and no adverse effect was reported.

Recent studies suggest that *P. tricornutum* can produce β -*N*-methylamino-L-alanine (BMAA) (Réveillon et al., 2016; Lage et al., 2019), which is a neurotoxin produced by certain cyanobacteria, diatoms and dinoflagellates. BMAA occurs ubiquitously in marine environments and can be transferred in food-webs to fish and seafood (Salomonsson et al., 2015; Lance et al., 2018).

Antimicrobial resistance aspects

Not applicable.

Conclusions on a recommendation for the QPS list

P. tricornutum is not recommended for QPS status based on the lack of a safe history of use in the food chain and on its potential for production of bioactive compounds with toxic effects.

3.3. Monitoring of new safety concerns related to organisms on the QPS list

The summaries of the evaluation of the possible safety concerns for humans, animals or the environment caused by QPS organisms described and published since the previous ELS (i.e. between January and June 2018, as described in Appendices B and C) and the references selected as potentially relevant for the QPS exercise (Appendix D) for each of the TUs or groups of TUs that are part of the QPS list (Appendix E) are presented below.

3.3.1. Gram-positive non-sporulating bacteria

3.3.1.1. *Bifidobacterium* spp.

A search for papers potentially relevant for the QPS consideration of *Bifidobacterium* spp. and *Carnobacterium divergens*¹¹ provided 206 references. The analysis of their title left 21 articles; the rest were discarded because they did not deal with safety concerns. Three articles were found relevant for the QPS consideration of *Bifidobacterium* spp. at the level of title and abstract screening. For one article, the full text was not available (conference proceedings) (Magistrelli et al., 2018). For one of the two other articles (Suzuki et al., 2018), the study was not considered because of study design shortcomings in relation to QPS assessment. For the third article (Kim et al., 2018a,b), no safety concern was identified for both *Bifidobacterium bifidum* and *Bifidobacterium longum*.

Based on the available evidence as described above, the QPS status of *Bifidobacterium* spp. is not changed.

3.3.1.2. Carnobacterium divergens

A search for papers potentially relevant for the QPS consideration of *Bifidobacterium* spp. and *Carnobacterium divergens* provided 206 references. The analysis of their title/abstracts left 21 articles; the rest were discarded because they did not deal with safety concerns. No article arrived to the final stage for this TU. Consequently, the QPS status of *C. divergens* is not changed.

3.3.1.3. Corynebacterium glutamicum

A search for papers potentially relevant for the QPS consideration of *Corynebacterium glutamicum* provided 45 references. No paper reached the final selection phase, therefore no new safety concerns were identified.

¹¹ These 2 TUs were searched together for practical reasons.

3.3.1.4. *Lactobacillus* spp.

A search for papers potentially relevant for the QPS consideration of any of the 35 *Lactobacillus* species included in the list provided 555 references. Analysis of their title left 22 articles; the rest were discarded because they did not deal with safety concerns. Inspection of their abstracts allowed the selection of 12 papers that might raise safety concerns. After analysing the full texts, nine were considered to be not relevant, while the other three described safety concerns linked to *L. paracasei* (Kao et al., 2018) and *L. rhamnosus* (Naqvi et al., 2018; Zeba et al., 2018). In one case (Kao et al., 2018), no indication is provided on how the identification of the pathogen was done, while in Zeba et al. (2018) an automated phenotypical method was used, known not to reliably identify lactobacilli. Patients in the three studies (Kao et al., 2018; Naqvi et al., 2018; Zeba et al., 2018) presented with clear predisposing conditions.

Based on the available evidence as described above, the QPS status of the lactobacilli involved in the reported cases and, by extension, of all others included in the QPS list, is not changed.

3.3.1.5. Lactococcus lactis

A search for papers potentially relevant for the QPS consideration of *Lactococcus lactis* provided 173 references. Analysis of their title/abstracts left six articles; the rest were discarded because they did not deal with safety concerns. Analysis of the abstracts allowed selection of four papers that might raise safety concerns. After analysing the full texts, one was found not to deal with safety concerns (Kato et al., 2018) while the other three (Chen et al., 2018; Kaboré et al., 2018; Shimizu et al., 2019) did. In two of them (Chen et al., 2018; Kaboré et al., 2018) phenotypical methods for identification of the etiological agent were used, which are known not to be reliable for this species. In addition, two of the papers (Chen et al., 2018; Shimizu et al., 2019) described cases in which clear predisposing conditions were identified. Shimizu et al. (2019) describes a patient suffering from a cholangiocarcinoma that blocked bile evacuation, reason why a bilioduodenal catheter was inserted. This might have been the source of the *L. lactis* cholangitis (predisposing condition) that appeared just 2 days later. The last reference (Kaboré et al., 2018) described the microbiota associated to 125 cases of endodontitis, out of which five contained *L. lactis* as part of a polymicrobial community, which is suggestive of colonisation of the oral cavity rather than aetiology of the infections.

Based on the available evidence as described above, the QPS status of *Lactococcus lactis* is not changed.

3.3.1.6. Leuconostoc citreum, Leuconostoc lactis, Leuconostoc mesenteroides, Leuconostoc pseudomesenteroides

A search for papers potentially relevant for the QPS consideration of *Leuconostoc* spp. and *Microbacterium imperiale*¹¹ provided 68 references. The analysis of their title left five articles; the rest were discarded because they did not deal with safety concerns. One paper lacked information on the identification procedures used to identify the infectious agents. Four papers arrived to the full text phase. Three were immediately excluded as they were not dealing with a safety concern or with these TUs. In the article of Menegueti et al. (2018), a 67-year-old female patient with chronic Chagas disease was submitted to several surgical interventions, and presented complications such as hypotension and hypoxemia. The identification of cultures was performed using automated phenotypical method known not to reliably identify *Leuconostoc* spp.

The safety concern identified was linked to severe underlying health conditions and the identification of strain was not reliable due to the use of phenotypic tests. Consequently the QPS status of *Leuconostoc* spp. is not changed.

3.3.1.7. Microbacterium imperiale

A search for papers potentially relevant for the QPS consideration of *Leuconostoc* and *Microbacterium imperiale*¹¹ provided 68 references. The analysis of their title left five articles; one paper lacked information on the identification procedures used to identify the infectious agents. The rest were discarded because they did not deal with safety concerns. None of the remaining four articles dealt with this TU. Consequently, the QPS status of *M. imperiale* is not changed.

3.3.1.8. Oenococcus oeni

A search for papers potentially relevant for the QPS consideration of *Oenococcus oeni* and *Pasteuria nishizawae*¹¹ provided 24 references. The analysis of their title/abstracts left one article for

consideration. No paper reached the final selection phase, no new safety concern was found. Consequently, the QPS status of *O. oeni* is not changed.

3.3.1.9. Pasteuria nishizawae

A search for papers potentially relevant for the QPS consideration of *Oenococcus oeni* and *Pasteuria nishizawae*¹¹ provided 24 references. The analysis of their title/abstracts left one article for consideration. No paper reached the final selection phase, no new safety concern was found. Consequently, the QPS status of *P. nishizawae* is not changed.

3.3.1.10. Pediococcus spp.

A search for papers potentially relevant for the QPS consideration of *Pediococcus* spp. provided 146 references. The analysis of their title left two articles (Chen et al., 2018; Singla et al., 2018). The latter does not describe a safety concern. Chen et al. (2018) describes an infectious endocarditis caused by *L. lactis* subsp. *Lactis* and *Pediococcus pentosaceus*. *P. pentosaceus* was identified after levofloxacin treatment in blood cultures, but the phenotypical method used for its identification is known not to be reliable for this species. In addition, the papers describe a case in which clear predisposing conditions were identified.

Consequently, the QPS status of *Pediococcus* spp. is not changed.

Pediococcus dextrinicus (Coster and White, 1964) Back, 1978, species, the name was changed to *Lactobacillus dextrinicus* (Coster and White, 1964) Haakensen et al., 2009, comb. nov. It will be updated in the QPS list.

3.3.1.11. *Propionibacterium* spp.

A search for papers potentially relevant for the QPS consideration of *Propionibacterium* spp. provided 27 references. Following the analysis of their title/abstracts, no articles were selected for the final selection phase, thus no new safety concerns were identified. Consequently, the QPS status of *Propionibacterium* spp. is not changed.

3.3.1.12. Streptococcus thermophilus

A search for papers potentially relevant for the QPS consideration of *Streptococcus thermophilus* provided 81 references. The analysis of their title/abstracts left one article that did not reach the final selection phase; thus, no new safety concern was found. Therefore, the QPS status of *S. thermophilus* is not changed.

3.3.2. Gram-positive spore-forming bacteria

3.3.2.1. Bacillus spp.

A search for papers potentially relevant for the QPS consideration of *Bacillus* spp. and *Geobacillus* stearothermophilus¹¹ provided 804 references. The analysis of their titles left 199 articles. A first round has been conducted using the MLT as a co-assistant; the second round of analysis of the 199 articles by two experts left two articles for more in-depth analysis. The remaining articles were discarded because they did not deal with safety concerns. The paper of Osman et al. (2018) concerns the enterotoxinogenic potential of *Bacillus pumilus* strains, a topic covered by the current qualification for *Bacillus* subtilis in an immunocompetent patient. For both papers there were methodological shortcomings on the identification methods of the bacterial strains and therefore the data presented were not included for further assessment.

The ELS did not come up with any information that would change the status of the *Bacillus* species included in the QPS list.

3.3.2.2. *Geobacillus stearothermophilus*

A search for papers potentially relevant for the QPS consideration of *Bacillus* spp. and *Geobacillus stearothermophilus*¹¹ provided 804 references. The analysis of their titles/abstract left 199 articles. None was dealing with this species. Consequently, the QPS status *Geobacillus stearothermophilus* is not changed.

3.3.3. Gram-negative bacteria

3.3.3.1. *Gluconobacter oxydans*

A search for papers potentially relevant for the QPS consideration of *Gluconobacter oxidans* and *Xanthomonas campestris*¹¹ provided 115 references. The analysis of their titles left one article; the rest were discarded because they did not deal with safety concerns. No paper reached the final selection phase for this TU. Consequently, the QPS status of *G. oxydans* is not changed.

3.3.3.2. Xanthomonas campestris

A search for papers potentially relevant for the QPS consideration of *Gluconobacter oxidans* and *Xanthomonas campestris*¹¹ provided 115 references. The analysis of their titles left one article; the rest were discarded because they did not deal with safety concerns. No paper reached the final selection phase for this TU. Consequently, the QPS status of *X. campestris* is not changed.

3.3.4. Yeasts

A search for papers potentially relevant for the QPS consideration of the yeasts' species included in the QPS list provided 1,358 references. The analysis of their titles left 268 articles. A first round has been conducted using the MLT as a co-assistant; 229 of these were immediately excluded because they were not in English or because they were not dealing with safety concerns. Thus, the ELS identified 30 articles relevant for different yeast species with QPS status (please refer to Appendix D for the complete list of references).

These 30 articles included description of 45 studies related to different yeast species with OPS status, of which 16 referred to Candida kefyr (teleomorph = Kluyveromyces marxianus) (Afsarian et al., 2018; Awad et al., 2018; Bharathi, 2018; Diba et al., 2018; García-Agudo et al., 2018; Hasan and Yassein, 2018; Kaur et al., 2018; Kesmen et al., 2018; Khedri et al., 2018; Nagy et al., 2018; Okmen et al., 2018; Omran and Mansori, 2018; de Paula et al., 2018; Sadrossadati et al., 2018; Shokohi et al., 2018; Simi et al., 2019), 8 to Saccharomyces cerevisiae including Saccharomyces boulardii (corresponding to 7 articles: Lazo-Vélez et al., 2018; Ruelle et al., 2018; Hasan and Yassein, 2018; Kara et al., 2018; Kesmen et al., 2018; Ochiai et al., 2018; Teblick et al., 2018), 10 to Candida famata (teleomorph = Debaryomyces hansenii) (Afsarian et al., 2018; Alobaid and Khan, 2018; Awad et al., 2018; Cen et al., 2018; Das et al., 2018; Diba et al., 2018; Hasan and Yassein, 2018; Kesmen et al., 2018; de Paula et al., 2018; Simi et al., 2019), 6 to Candida pelliculosa (synonymus = Pichia anomala, teleomorph = Wickerhamomyces anomalus) (Ahmadsah et al., 2018; Arendrup et al., 2018; Cen et al., 2018; Jung et al., 2018; Kesmen et al., 2018; Soman et al., 2018), 1 to Candida utilis (teleomorph = Lindnera jadinii) (Treguier et al., 2018), 1 to Hanseniaspora uvarum (Kesmen et al., 2018), 1 Kluyveromyces lactis (Kesmen et al., 2018) and 1 to Schizosaccharomyces pombe (Kesmen et al., 2018). One of these articles was considered relevant (Pfaller et al., 2018) to evaluate since it presented new data on antimycotic MIC breakpoints for azoles but it is not associated to any specific yeast TU.

For the other yeast species with QPS status, no relevant studies were identified through the ELS.

Methodological problems were identified in 30 out of those 45 studies (corresponding to 22 articles – see Table 3). In 17 of those 30 studies (corresponding to 11 articles: Alobaid and Khan, 2018; Awad et al., 2018; Bharathi, 2018; de Paula et al., 2018; Hasan and Yassein, 2018; Kaur et al., 2018; Okmen et al., 2018; Ruelle et al., 2018; Soman et al., 2018; Treguier et al., 2018; Simi et al., 2019), a problem was found in respect to the methodology used for identity confirmation of the microorganism, therefore the value of these results and conclusions were very limited. In 2 of those 30 studies (corresponding to 1 article, Awad et al., 2018), the problem was due to a lack of information regarding the source attribution. In 19 studies (corresponding to 15 articles: Afsarian et al., 2018; Alobaid and Khan, 2018; Bharathi, 2018; Diba et al., 2018; Das et al., 2018; Hasan and Yassein, 2018; Jung et al., 2018; Kara et al., 2018; Khedri et al., 2018; Kaur et al., 2018; Nagy et al., 2018; Omran and Mansori, 2018; Sadrossadati et al., 2018; Shokohi et al., 2018; Teblick et al., 2018), the problem was due to predisposing factors in the exposed subject.

For 32 of those 45 studies, no potential safety concern was reported, while a potential safety concern was described in the other 13 studies (corresponding to 11 articles: Diba et al., 2018; Das et al., 2018; Jung et al., 2018; Kara et al., 2018; Khedri et al., 2018; Nagy et al., 2018; Omran and Mansori, 2018; Sadrossadati et al., 2018; Afsarian et al., 2018; Shokohi et al., 2018; Teblick et al., 2018): 3 for *C. famata* (Das et al., 2018; Diba et al., 2018; Shokohi et al., 2018), 7 for *C. kefyr* (Afsarian et al., 2018; Diba et al., 2018; Khedri et al., 2018; Comran and Mansori, 201

Shokohi et al., 2018), 1 for *C. pelliculosa* (Jung et al., 2018) and 2 for *Saccharomyces cerevisiae* (Kara et al., 2018; Teblick et al., 2018). All these studies reported isolation of the QPS yeasts from opportunistic infections in patients with serious predisposing factors.

In short, the ELS did not identify any information that would change the status for the yeast species included in the QPS list.



Relevant to	Articles not	19 articles	Any	Yes	11 articles	Methodology used	11 articles	Alobaid and Khan (2018); Awad et al. (2018);
the QPS exercise ^{(a),(b)}	describing safety concerns	(32 studies)	methodological problem identified?		(17 studies)	for identity confirmation of the microorganism	(17 studies)	Bharathi (2018); de Paula et al. (2018); Hasan and Yassein (2018); Kaur et al. (2018); Okmen et al. (2018); Ruelle et al. (2018); Simi et al. (2019); Soman et al. (2018); Treguier et al. (2018)
						Reliability of the source attribution	1 article (2 studies)	Awad et al. (2018)
						Misuse of the microorganism	None	
						Predisposing factors in the exposed subjects	4 articles (6 studies)	Alobaid and Khan (2018); Bharathi (2018); Hasan and Yassein (2018); Kaur et al. (2018)
						Other reasons	1 article (2 studies)	de Paula et al. (2018)
				No	8 articles (15 studies)			Arendrup et al. (2018); Ahmadsah et al. (2018); Cen et al. (2018); García-Agudo et al. (2018); Lazo-Vélez et al. (2018); Kesmen et al. (2018); Ochiai et al. (2018); Pfaller et al. (2018)
	Articles dealing with safety concerns	11 articles (13 studies)	Any methodological problem identified?	Yes	11 articles (13 studies)	Methodology used for identity confirmation of the microorganism	None	
					Reliability of the source attribution	None		
					Misuse of the microorganism	None		
						Predisposing factors in the exposed subjects	11 articles (13 studies)	Afsarian et al. (2018); Diba et al. (2018); Das et al. (2018); Jung et al. (2018); Kara et al. (2018); Khedri et al. (2018); Nagy et al. (2018); Omran and Mansori (2018); Sadrossadati et al. (2018); Shokohi et al. (2018); Teblick et al. (2018)
						Other reasons	None	
				No	None			

Table 3:	Articles that arrived to the evaluation	phase (final st	tep of the extensive literature search)) for the OPS statu	us yeasts group (30 articles with 45 studi	es)
		prido e (inital o) 101 and Q1 0 0 add	so years group (be articles mar is staal	,

(a): Please refer to Appendix D for the complete list of references.(b): Number of references (articles and studies) indicated for each step.

3.3.5. Viruses used for plant protection

3.3.5.1. Alphaflexiviridae

A search for papers potentially relevant for the QPS consideration of *Alphaflexiviridae* and *Potyviridae*¹¹ provided 39 references. No paper reached the final selection phase, thus no new safety concern was found.

3.3.5.2. Potyviridae

A search for papers potentially relevant for the QPS consideration of *Alphaflexiviridae* and *Potyviridae*¹¹ provided 39 references. No paper reached the final selection phase, thus no new safety concern was found.

3.3.5.3. Baculoviridae

A search for papers potentially relevant for the QPS consideration of *Baculoviridae* provided 69 references. No article reached the final selection phase, thus no new safety concern was found.

The ELS did not come up with any information that would change the current QPS status of any of the above virus families.

4. Conclusions

ToR 1: Keep updated the list of biological agents being notified, in the context of a technical dossier to EFSA Units (such as Feed, Food Ingredients and Packaging (FIP), Nutrition Unit and Pesticides Unit), for intentional use in feed and/or food or as sources of food and feed additives, enzymes and plant protection products for safety assessment:

• Between October 2018 and March 2019, the list was updated with 47 notifications that were received by EFSA, of which 32 were for feed additives, 2 for food enzymes, food additives and flavourings, 10 for novel foods and 3 for plant protection products.

ToR 2: Review taxonomic units previously recommended for the QPS list and their qualifications when new information has become available:

• In relation to the results of the monitoring of possible new safety concerns related to the QPS list, there were no results that justify removal of any TU from the QPS list or changes in their respective qualifications.

ToR 3: (*Re*)assess the suitability of taxonomic units notified to EFSA not present in the current QPS list for their inclusion in that list:

- The TUs corresponding to 19 out of the 47 notifications received, already had a QPS status.
- Of the 28 notifications without a QPS status, 11 notifications related to filamentous fungi which were excluded from QPS activities in the follow-up of a recommendation of the QPS 2013 update (EFSA BIOHAZ Panel, 2013, 2014, 2016), 9 notifications related to *E. coli*, which was recently excluded from the current mandate by the BIOHAZ Panel (EFSA BIOHAZ Panel, 2018a).
- The remaining eight TUs, Burkholderia ubonensis, Corynebacterium ammoniagenes, Corynebacterium casei, Euglena gracilis, Gluconobacter frateurii, Microbacterium foliorum, Phaeodactylum tricornutum and Sphingomonas elodea, were evaluated for potential QPS recommendation for the first time.

5. Recommendations

- *Burkholderia ubonensis* cannot be recommended for the QPS list due to its ability to generate biologically active compounds and limited of body of knowledge.
- *Corynebacterium ammoniagenes* can be recommended for QPS list status with the qualification 'for production purposes only'.
- Corynebacterium casei cannot be recommended to the QPS list due to lack of body of knowledge.
- *Euglena gracilis* may be recommended for the QPS list with the qualification 'for production purposes only'.



- *Gluconobacter frateurii* cannot be recommended to the QPS list due to lack of body of knowledge.
- *Microbacterium foliorum* cannot be recommended to the QPS list due to lack of body of knowledge.
- *Phaeodactylum tricornutum* cannot be recommended for QPS status, based on the lack of a safe history of use in the food chain and on its potential production of bioactive compounds with toxic effects based on the lack of a safe history of use in the food chain and a limited knowledge on its potential production of bioactive compounds with toxic effects.
- *Sphingomonas elodea* could not be assessed for a possible QPS recommendation because it is not a valid species.

Pediococcus dextrinicus (Coster and White, 1964) Back, 1978 species has been changed to *Lactobacillus dextrinicus* (Coster and White, 1964) Haakensen et al., 2009, comb. nov.

This new QPS recommendation will be included as an addition to the list of QPS status recommended biological agents (EFSA BIOHAZ Panel, 2016), published both as an update to the Scientific Opinion (EFSA BIOHAZ Panel, 2016) and as supporting information available on the Knowledge Junction at https://doi.org/10.5281/zenodo.1146566.

References

- Afsarian M, Reza Ataollahi M, Badram L, Moradi N, Badali H and Shokohi T, 2018. Molecular Identification of Clinically Common and Uncommon Yeast Species. Jundishapur. Journal of Microbiology, e66240, https://doi. org/10.5812/jjm.66240
- Ahmadsah LSF, Kim E, Jung YS and Kim HY, 2018. Identification of LAB and Fungi in Laru, a Fermentation Starter, by PCR-DGGE, SDS-PAGE, and MALDI-TOF MS. Journal of Microbiology and Biotechnology, 28, 32–39. https:// doi.org/10.4014/jmb.1705.05044
- Alobaid K and Khan Z, 2018. Epidemiologic characteristics of adult candidemic patients in a secondary hospital in Kuwait: A retrospective study. Journal de Mycologie Medicale, 29, 35–38. https://doi.org/10.1016/j.mycmed. 2018.12.001
- An B-K, Choi Y-I, Kang C-W and Lee K-W, 2018. Effects of dietary *Corynebacterium ammoniagenes*-derived single cell protein on growth performance, blood and tibia bone characteristics, and meat quality of broiler chickens. Journal of Animal and Feed Sciences, 27, 140–147. https://doi.org/10.22358/jafs/91966/2018
- Anonymous , 2000. Validation of publication of new names and new combinations previously effectively published outside the IJSEM. List No. 75. International Journal of Systematic and Evolutionary Microbiology, 50, 1415–1417.
- Arendrup MC, Chowdhary A, Astvad KMT and Jørgensen KM, 2018. APX001A *In Vitro* Activity against Contemporary Blood Isolates and *Candida auris* Determined by the EUCAST Reference Method. Antimicrobial Agents and Chemotherapy, 62, https://doi.org/10.1128/aac.01225-18
- Awad L, Tamim H, Abdallah D, Salameh M, Mugharbil A, Jisr T, Zahran K, Droubi N, Ibrahim A and Moghnieh R, 2018. Correlation between antifungal consumption and the distribution of *Candida* species in different hospital departments of a Lebanese medical Centre. BMC Infectious Diseases, 18, 589. https://doi.org/10.1186/s12879-018-3512-z
- Back W, 1978. Elevation of *Pediococcus cerevisiae* subsp. *dextrinicus* Coster and White to Species Status [*Pediococcus dextrinicus* (Coster and White) comb. nov.]. International Journal of Systematic and Evolutionary Microbiology, 28, 523–527. https://doi.org/10.1099/00207713-28-4-523
- Bajpai VK, 2016. Antimicrobial bioactive compounds from marine algae: A mini review. Indian Journal of Geo-Marine Sciences, 45, 1076–1085.
- Behrendt U, Ulrich A and Schumann P, 2001. Description of Microbacterium foliorum sp. nov. and Microbacterium phyllosphaerae sp. nov., isolated from the phyllosphere of grasses and the surface litter after mulching the sward, and reclassification of Aureobacterium resistens (Funke et al. 1998) as Microbacterium resistens comb. nov. International Journal of Systematic and Evolutionary Microbiology, 51, 1267–1276. https://doi.org/10. 1099/00207713-51-4-1267
- Bertuzzi AS, Kilcawley KN, Sheehan JJ, O'Sullivan MG, Kennedy D, McSweeney PLH and Rea MC, 2017. Use of smear bacteria and yeasts to modify flavour and appearance of Cheddar cheese. International Dairy Journal, 72, 44–54. https://doi.org/10.1016/j.idairyj.2017.04.001
- Bharathi R, 2018. Comparison of Chromogenic Media with the Corn Meal Agar for Speciation of *Candida*. Journal of Pure and Applied Microbiology, 12, 1617–1622.
- Bockelmann W, 2011. Cheese | Smear-Ripened Cheeses. In: Fuquay JW, Fox PF and McSweeney PLH (eds.). Encyclopedia of Dairy Sciences. 2nd edition, Elsevier Ltd. pp. 753-766.
- Bohlin K, 1897. Zur Morphologie und Biologie einzelliger Algen. Öfversigt af Kongliga [Svenska] Vetenskadademiens Förhanligar. Stockholm, 54, 507–529.

- Bowler C, Allen AE, Badger JH, Grimwood J, Jabbari K, Kuo A, Maheswari U, Martens C, Maumus F, Otillar RP, Rayko E, Salamov A, Vandepoele K, Beszteri B, Gruber A, Heijde M, Katinka M, Mock T, Valentin K, Verret F, Berges JA, Brownlee C, Cadoret J-P, Chiovitti A, Choi CJ, Coesel S, De Martino A, Detter JC, Durkin C, Falciatore A, Fournet J, Haruta M, Huysman MJJ, Jenkins BD, Jiroutova K, Jorgensen RE, Joubert Y, Kaplan A, Kröger N, Kroth PG, La Roche J, Lindquist E, Lommer M, Martin–Jézéquel V, Lopez PJ, Lucas S, Mangogna M, McGinnis K, Medlin LK, Montsant A, Secq M-POL, Napoli C, Obornik M, Parker MS, Petit J-L, Porcel BM, Poulsen N, Robison M, Rychlewski L, Rynearson TA, Schmutz J, Shapiro H, Siaut M, Stanley M, Sussman MR, Taylor AR, Vardi A, von Dassow P, Vyverman W, Willis A, Wyrwicz LS, Rokhsar DS, Weissenbach J, Armbrust EV, Green BR, Van de Peer Y and Grigoriev IV, 2008. The *Phaeodactylum* genome reveals the evolutionary history of diatom genomes. Nature, 456, 239. https://doi.org/10.1038/nature07410, https://www.nature.com/articles/nature 07410#supplementary-information
- Brennan NM, Brown R, Goodfellow M, Ward AC, Beresford TP, Simpson PJ, Fox PF and Cogan TM, 2001. *Corynebacterium mooreparkense* sp. nov. and *Corynebacterium casei* sp. nov., isolated from the surface of a smear-ripened cheese. International Journal of Systematic and Evolutionary Microbiology, 51, 843–852. https://doi.org/10.1099/00207713-51-3-843
- Brennan NM, Ward AC, Beresford TP, Fox PF, Goodfellow M and Cogan TM, 2002. Biodiversity of the bacterial flora on the surface of a smear cheese. Applied and Environmental Microbiology, 68, 820–830. https://doi.org/10. 1128/aem.68.2.820-830.2002
- Cen QW, Chen T, Zheng W, Zhang Y, Ying RF, Tang ZX and Shi LE, 2018. Isolation, identification and partial characterization of film-forming microorganisms from Chinese homemade pickle, a traditional fermented vegetable product. Chiang Mai Journal of Science, 45, 2283–2293.
- Cerezuela R, Guardiola FA, Meseguer J and Esteban MA, 2012. Enrichment of gilthead seabream (*Sparus aurata* L.) diet with microalgae: effects on the immune system. Fish Physiology and Biochemistry, 38, 1729–1739. https://doi.org/10.1007/s10695-012-9670-9
- Chen F, Zhang Z and Chen J, 2018. Infective endocarditis caused by *Lactococcus lactis* subsp. *lactis* and *Pediococcus pentosaceus*: A case report and literature review. Medicine (Baltimore), 97, e13658. https://doi.org/10.1097/md.00000000013658
- Cogan TM, Goerges S, Gelsomino R, Larpin S, Hohenegger M, Bora N, Jamet E, Rea MC, Mounier J, Vancanneyt M, Guéguen M, Desmasures N, Swings J, Goodfellow M, Ward AC, Sebastiani H, Irlinger F, Chamba J-F, Beduhn R and Scherer S, 2014. Biodiversity of the Surface Microbial Consortia from Limburger, Reblochon, Livarot, Tilsit, and Gubbeen Cheeses. Microbiology Spectrum, 2, https://doi.org/10.1128/microbiolspec.CM-0010-2012
- Collins MD, 1987. Transfer of *Brevibacterium ammoniagenes* (Cooke and Keith) to the Genus *Corynebacterium as Corynebacterium ammoniagenes* comb. nov. International Journal of Systematic and Evolutionary Microbiology, 37, 442–443. https://doi.org/10.1099/00207713-37-4-442
- Cooke JV and Keith HR, 1927. A type of urea-splitting bacterium found in the human intestinal tract. Journal of Bacteriology, 13, 315–319.
- Coster E and White HR, 1964. Further Studies of the Genus Pediococcus., 37, 15–31. https://doi.org/10.1099/ 00221287-37-1-15
- Das S, Rai G, Tigga RA, Srivastava S, Singh PK, Sharma R, Datt S, Singh NP and Dar SA, 2018. *Candida auris* in critically ill patients: Emerging threat in intensive care unit of hospitals. J Mycol Med, 28, 514–518. https://doi.org/10.1016/j.mycmed.2018.06.005
- De Martino A, Meichenin A, Shi J, Pan K and Bowler C, 2007. Genetic and phenotypic characterization of *Phaeodactylum tricornutum* (Bacillariophyceae) accessions. Journal of Phycology, 43, 992–1009. https://doi.org/10.1111/j.1529-8817.2007.00384.x
- Deetae P, Bonnarme P, Spinnler HE and Helinck S, 2007. Production of volatile aroma compounds by bacterial strains isolated from different surface-ripened French cheeses. Applied Microbiology and Biotechnology, 76, 1161–1171. https://doi.org/10.1007/s00253-007-1095-5
- Deetae P, Spinnler HE, Bonnarme P and Helinck S, 2009. Growth and aroma contribution of *Microbacterium foliorum, Proteus vulgaris* and *Psychrobacter* sp. during ripening in a cheese model medium. Applied Microbiology and Biotechnology, 82, 169–177. https://doi.org/10.1007/s00253-008-1805-7
- Demirel Z, 2016. Identification and fatty acid composition of coccolithophore and diatom species isolated from Aegean sea. Romanian Biotechnological Letters, 21, 11761–11768.
- Diba K, Makhdoomi K, Nasri E, Vaezi A, Javidnia J, Gharabagh DJ, Jazani NH, Reza Chavshin A, Badiee P, Badali H and Fakhim H, 2018. Emerging *Candida* species isolated from renal transplant recipients: Species distribution and susceptibility profiles. Microbial Pathogenesis, 125, 240–245. https://doi.org/10.1016/j.micpath.2018.09.026
- Dinakaran V, John L, Rathinavel A, Gunasekaran P and Rajendhran J, 2012. Prevalence of bacteria in the circulation of cardiovascular disease patients, Madurai, India. Heart, Lung & Circulation, 21, 281–283. https://doi.org/10.1016/j.hlc.2012.02.007
- Ebenezer TE, Zoltner M, Burrell A, Nenarokova A, Novák Vanclová AMG, Prasad B, Soukal P, Santana-Molina C, O'Neill E, Nankissoor NN, Vadakedath N, Daiker V, Obado S, Silva-Pereira S, Jackson AP, Devos DP, Lukeš J, Lebert M, Vaughan S, Hampl V, Carrington M, Ginger ML, Dacks JB, Kelly S and Field MC, 2019. Transcriptome, proteome and draft genome of *Euglena gracilis*. BMC Biology, 17, 11. https://doi.org/10.1186/s12915-019-0626-8



- EFSA (European Food Safety Authority), 2005. Opinion of the Scientific Committee on a request from EFSA related to a generic approach to the safety assessment by EFSA of microorganisms used in food/feed and the production of food/feed additives. EFSA Journal 2005;3(6):226, 12 pp. https://doi.org/10.2903/j.efsa.2005.226
- EFSA (European Food Safety Authority), 2007. Introduction of a Qualified Presumption of Safety (QPS) approach for assessment of selected microorganisms referred to EFSA Opinion of the Scientific Committee. EFSA Journal 2007;5(12):587, 16 pp. https://doi.org/10.2903/j.efsa.2007.587
- EFSA (European Food Safety Authority), 2008. The maintenance of the list of QPS microorganisms intentionally added to food or feed Scientific Opinion of the Panel on Biological Hazards EFSA Journal 2008;6(12):923, 48 pp. https://doi.org/10.2903/j.efsa.2008.923
- EFSA (European Food Safety Authority) BIOHAZ Panel, 2009. Scientific Opinion on the maintenance of the list of QPS microorganisms intentionally added to food or feed (2009 update). EFSA Journal 2009;7(12):1431, 92 pp. https://doi.org/10.2903/j.efsa.2009.1431
- EFSA BIOHAZ Panel (EFSA Panel on Biological Hazards), 2013. Scientific Opinion on the maintenance of the list of QPS biological agents intentionally added to food and feed (2013 update). EFSA Journal 2013;11(11):3449, 106 pp. https://doi.org/10.2903/j.efsa.2013.3449
- EFSA BIOHAZ Panel (EFSA Panel on Biological Hazards), 2014. Statement on the update of the list of QPSrecommended biological agents intentionally added to food or feed as notified to EFSA 1: Suitability of taxonomic units notified to EFSA until October 2014. EFSA Journal 2014;12(12):3938, 41 pp. https://doi.org/ 10.2903/j.efsa.2014.3938
- EFSA BIOHAZ Panel(EFSA Panel on Biological Hazards), 2016. Update of the list of QPS-recommended biological agents intentionally added to food or feed as notified to EFSA 4: suitability of taxonomic units notified to EFSA until March 2016. EFSA Journal 2016;14(7):4522, 37 pp. https://doi.org/10.2903/j.efsa.2016.4522
- EFSA BIOHAZ Panel (EFSA Panel on Biological Hazards), Ricci A, Allende A, Bolton D, Chemaly M, Davies R, Girones R, Herman L, Koutsoumanis K, Lindqvist R, Nørrung B, Robertson L, Ru G, Sanaa M, Simmons M, Skandamis P, Snary E, Speybroeck N, Ter Kuile B, Threlfall J, Wahlström H, Cocconcelli PS, Klein G, Prieto Maradona M, Querol A, Peixe L, Suarez JE, Sundh I, Vlak JM, Aguilera-Gómez M, Barizzone F, Brozzi R, Correia S, Heng L, Istace F, Lythgo C and Fernández Escámez PS, 2017a. Scientific Opinion on the update of the list of QPS-recommended biological agents intentionally added to food or feed as notified to EFSA. EFSA Journal 2017;15(3):4664, 177 pp. https://doi.org/10.2903/j.efsa.2017.4664
- EFSA BIOHAZ Panel (EFSA Panel on Biological Hazards), Ricci A, Allende A, Bolton D, Chemaly M, Davies R, Girones R, Koutsoumanis K, Lindqvist R, Nørrung B, Robertson L, Ru G, Fernandez Escamez PS, Sanaa M, Simmons M, Skandamis P, Snary E, Speybroeck N, Ter Kuile B, Threlfall J, Wahlström H, Cocconcelli PS, Peixe L, Maradona MP, Querol A, Suarez JE, Sundh I, Vlak J, Correia S and Herman L, 2017b. Update of the list of QPS-recommended biological agents intentionally added to food or feed as notified to EFSA 6: suitability of taxonomic units notified to EFSA until March 2017. EFSA Journal 2017;15(7):4884, 32 pp. https://doi.org/10. 2903/j.efsa.2017.4884
- EFSA BIOHAZ Panel(EFSA Panel on Biological Hazards), Ricci A, Allende A, Bolton D, Chemaly M, Davies R, Girones R, Koutsoumanis K, Lindqvist R, Nørrung B, Robertson L, Ru G, Fernández Escámez PS, Sanaa M, Simmons M, Skandamis P, Snary E, Speybroeck N, Ter Kuile B, Threlfall J, Wahlström H, Cocconcelli PS, Peixe L, Maradona MP, Querol A, Suarez JE, Sundh I, Vlak J, Barizzone F, Correia S and Herman L, 2018a. Update of the list of QPS-recommended biological agents intentionally added to food or feed as notified to EFSA 7: suitability of taxonomic units notified to EFSA until September 2017. EFSA Journal 2018;16(1):5131, 43 pp. https://doi.org/ 10.2903/j.efsa.2018.5131
- EFSA BIOHAZ Panel (EFSA Panel on Biological Hazards), Ricci A, Allende A, Bolton D, Chemaly M, Davies R, Fernández Escámez PS, Girones R, Koutsoumanis K, Lindqvist R, Nørrung B, Robertson L, Ru G, Sanaa M, Simmons M, Skandamis P, Snary E, Speybroeck N, Ter Kuile B, Threlfall J, Wahlström H, Cocconcelli PS, Peixe L, Maradona MP, Querol A, Suarez JE, Sundh I, Vlak J, Barizzone F, Correia S and Herman L, 2018b. Update of the list of QPS-recommended biological agents intentionally added to food or feed as notified to EFSA 8: suitability of taxonomic units notified to EFSA until March 2018. EFSA Journal 2018;16(7):5315, 42 pp. https://doi.org/10.2903/j.efsa.2018.5315
- El-Deek AA, Ghonem KM, Hamdy SM, Aser MA, Fahad A and Osman MM, 2009. Producing Single Cell Protein from Poultry Manure and Evaluation for Broiler Chickens Diets. International Journal of Poultry Science, 8, 1062–1077. https://doi.org/10.3923/ijps.2009.1062.1077
- Euzéby JP, 2013. List of prokaryotic names with standing nomenclature (LPSN). Available online: http://www.bacterio.net/-allnamesdl.html [Accessed: 17 June 2019]
- Garcia JL, de Vicente M and Galán B, 2017. Microalgae, old sustainable food and fashion nutraceuticals. Microbial Biotechnology, 10, 1017–1024. https://doi.org/10.1111/1751-7915.12800
- García-Agudo L, Rodríguez-Iglesias M and Carranza-González R, 2018. Nosocomial Candiduria in the Elderly: Microbiological Diagnosis. Mycopathologia, 183, 591–596. https://doi.org/10.1007/s11046-017-0232-7
- Gneiding K, Frodl R and Funke G, 2008. Identities of *Microbacterium* spp. encountered in human clinical specimens. Journal of Clinical Microbiology, 46, 3646–3652. https://doi.org/10.1128/jcm.01202-08



- Haakensen M, Dobson CM, Hill JE and Ziola B, 2009. Reclassification of *Pediococcus dextrinicus* (Coster and White 1964) Back 1978 (Approved Lists 1980) as *Lactobacillus dextrinicus* comb. nov., and emended description of the genus *Lactobacillus*. International Journal of Systematic and Evolutionary Microbiology, 59, 615–621. https://doi.org/10.1099/ijs.0.65779-0
- Hannon J, Sousa-Gallagher MJ, Lillevang S, Sepulchre A, Bockelmann W and McSweeney PLH, 2004. Effect of defined-strain surface starters on the ripening of Tilsit cheese. International Dairy Journal, 14, 871–880. https://doi.org/10.1016/j.idairyj.2004.03.001
- Haro P, Sáez K and Gómez PI, 2017. Physiological plasticity of a Chilean strain of the diatom *Phaeodactylum tricornutum*: the effect of culture conditions on the quantity and quality of lipid production. Journal of Applied Phycology, 29, 2771–2782. https://doi.org/10.1007/s10811-017-1212-5
- Hasan KAM and Yassein SN, 2018. Prevalence and type of fungi in milk from goats with sub clinical mastitis. Online Journal of Veterinary Research, 22, 669–674.
- Ho VTT, Fleet GH and Zhao J, 2018. Unravelling the contribution of lactic acid bacteria and acetic acid bacteria to cocoa fermentation using inoculated organisms. International Journal of Food Microbiology, 279, 43–56. https://doi.org/10.1016/j.ijfoodmicro.2018.04.040
- Horsman ME, Marous DR, Li R, Oliver RA, Byun B, Emrich SJ, Boggess B, Townsend CA and Mobashery S, 2017. Whole-Genome Shotgun Sequencing of Two beta-Proteobacterial Species in Search of the Bulgecin Biosynthetic Cluster. ACS Chemical Biology, 12, 2552–2557. https://doi.org/10.1021/acschembio.7b00687
- Imada A, Kitano K, Kintaka K, Muroi M and Asai M, 1981. Sulfazecin and isosulfazecin, novel beta-lactam antibiotics of bacterial origin. Nature, 289, 590–591.
- Jung J, Moon YS, Yoo JA, Lim JH, Jeong J and Jun JB, 2018. Investigation of a nosocomial outbreak of fungemia caused by *Candida pelliculosa* (*Pichia anomala*) in a Korean tertiary care center. Journal of Microbiology, Immunology, and Infection, 51, 794–801. https://doi.org/10.1016/j.jmii.2017.05.005
- Kaboré AW, Dembélé R, Bagré ST, Konaté A, Boisramé S, Chevalier V, Konsem T, Traoré SA and Barro N, 2018. Characterization and Antimicrobial Susceptibility of *Lactococcus lactis* Isolated from Endodontic Infections in Ouagadougou. Burkina Faso. Dentistry Journal, 6, https://doi.org/10.3390/dj6040069
- Kang KS, Veeder GT, Mirrasoul PJ, Kaneko T and Cottrell IW, 1982. Agar-like polysaccharide produced by a pseudomonas species: production and basic properties. Applied and Environmental Microbiology, 43, 1086–1091.
- Kao B-Z, Lin H-J, Chen M-Y, Wu C-S, Lin S-T, Lee M-H, Lai Y-X and Hu P-J, 2018. Lactobacillus paracasei as cause of liver abscess: Case report Proceedings of the Asian Pacific Digestive Week (APDW). Connecting Excellence on Gastroenterology and Hepatology in Asia-Pacific, Seoul, Korea, 15–18 November 2018s, OE-0708 (PE-0448) pp.
- Kara I, Yıldırım F, Özgen Ö, Erganiş S, Aydoğdu M, Dizbay M, Gürsel G and Kalkanci A, 2018. *Saccharomyces cerevisiae* fungemia after probiotic treatment in an intensive care unit patient. J Mycol Med, 28, 218–221. https://doi.org/10.1016/j.mycmed.2017.09.003
- Kato Y, Kanayama M, Yanai S, Nozawa H, Kanauchi O and Suzuki S, 2018. Safety Evaluation of Excessive Intake of Lactococcus lactis subsp. lactis JCM 5805: A Randomized, Double-Blind, Placebo-Controlled. Parallel-Group Trial. Food and Nutrition Sciences, 09, 403–419. https://doi.org/10.4236/fns.2018.94032
- Kaur R, Mehra B, Dhakad MS, Goyal R, Bhalla P and Dewan R, 2018. Clinico-mycological analysis and antifungal resistance pattern in human immunodeficiency virus-associated candidiasis: An Indian perspective. Indian J Sex Transm Dis AIDS, 39, 111–119. https://doi.org/10.4103/ijstd.IJSTD_89_17
- Kesmen Z, Büyükkiraz ME, Özbekar E, Çelik M, Özkök FÖ, Kilıç Ö, Çetin B and Yetim H, 2018. Assessment of Multi Fragment Melting Analysis System (MFMAS) for the Identification of Food-Borne Yeasts. Current Microbiology, 75, 716–725. https://doi.org/10.1007/s00284-018-1437-9
- Khedri S, Santos ALS, Roudbary M, Hadighi R, Falahati M, Farahyar S, Khoshmirsafa M and Kalantari S, 2018. Iranian HIV/AIDS patients with oropharyngeal candidiasis: identification, prevalence and antifungal susceptibility of *Candida species*. Letters in Applied Microbiology, 67, 392–399. https://doi.org/10.1111/lam. 13052
- Kim HJ, Lee AW and Park C, 2018a. Toxicological evaluation of *Microbacterium foliorum* SYG27B-MF. Regulatory Toxicology and Pharmacology, 100, 16–24. https://doi.org/10.1016/j.yrtph.2018.09.022
- Kim JM, Ku S, Kim YS, Lee HH, Jin H, Kang S, Li R, Johnston VT, Park SM and Ji EG, 2018b. Safety Evaluations of Bifidobacterium bifidum BGN4 and Bifidobacterium longum BORI. International Journal of Molecular Sciences, 19, https://doi.org/10.3390/ijms19051422
- Koizumi S, Yonetani Y, Maruyama A and Teshiba S, 2000. Production of riboflavin by metabolically engineered *Corynebacterium ammoniagenes*. Applied Microbiology and Biotechnology, 53, 674–679.
- Korsak N, Taminiau B, Leclercq M, Nezer C, Crevecoeur S, Ferauche C, Detry E, Delcenserie V and Daube G, 2015. Short communication: Evaluation of the microbiota of kefir samples using metagenetic analysis targeting the 16S and 26S ribosomal DNA fragments. Journal of Dairy Science, 98, 3684–3689. https://doi.org/10.3168/jds. 2014-9065
- Krajčovič J, Matej V and Schwartzbach SD, 2015. Euglenoid flagellates: a multifaceted biotechnology platform. Journal of Biotechnology, 202, 135–145. https://doi.org/10.1016/j.jbiotec.2014.11.035
- Laffineur K, Avesani V, Cornu G, Charlier J, Janssens M, Wauters G and Delmée M, 2003. Bacteremia due to a novel *Microbacterium* species in a patient with leukemia and description of *Microbacterium paraoxydans* sp. nov. Journal of Clinical Microbiology, 41, 2242–2246. https://doi.org/10.1128/jcm.41.5.2242-2246.2003

- Lage S, Ström L, Godhe A and Rydberg S, 2019. Kinetics of β-N-methylamino-L-alanine (BMAA) and 2, 4diaminobutyric acid (DAB) production by diatoms: the effect of nitrogen. European Journal of Phycology, 54, 115–125. https://doi.org/10.1080/09670262.2018.1508755
- Lance E, Arnich N, Maignien T and Biré R, 2018. Occurrence of beta-N-methylamino-l-alanine (BMAA) and Isomers in Aquatic Environments and Aquatic Food Sources for Humans. Toxins (Basel), 10, https://doi.org/10. 3390/toxins10020083
- Lazo-Vélez MA, Serna-Saldívar SO, Rosales-Medina MF, Tinoco-Alvear M and Briones-García M, 2018. Application of *Saccharomyces cerevisiae var. boulardii* in food processing: a review. Journal of Applied Microbiology, 125, 943–951. https://doi.org/10.1111/jam.14037
- Loveridge EJ, Jones C, Bull MJ, Moody SC, Kahl MW, Khan Z, Neilson L, Tomeva M, Adams SE, Wood AC, Rodriguez-Martin D, Pinel I, Parkhill J, Mahenthiralingam E and Crosby J, 2017. Reclassification of the Specialized Metabolite Producer *Pseudomonas mesoacidophila* ATCC 31433 as a Member of the *Burkholderia cepacia* Complex. Journal of Bacteriology, 199, e00125–00117. https://doi.org/10.1128/JB.00125-17
- Lumactud R, Shen SY, Lau M and Fulthorpe R, 2016. Bacterial Endophytes Isolated from Plants in Natural Oil Seep Soils with Chronic Hydrocarbon Contamination. Frontiers in Microbiology, 7, 755. https://doi.org/10.3389/fmicb. 2016.00755
- Lumactud R, Fulthorpe R, Sentchilo V and van der Meer JR, 2017. Draft Genome Sequence of *Microbacterium foliorum* Strain 122 Isolated from a Plant Growing in a Chronically Hydrocarbon-Contaminated Site. Genome Announc, 5, https://doi.org/10.1128/genomeA.00434-17
- Magistrelli L, Amoruso A, Milner AV, Mogna L, Cantello R, Pane M and Comi C, 2018. Effects of probiotic bacterial strains on peripheral inflammation in Parkinson's disease (EPR2096). European Journal of Neurology, 25 (Suppl, S2):428–428.
- Malimas T, Yukphan P, Takahashi M, Potacharoen W, Tanasupawat S, Nakagawa Y, Tanticharoen M and Yamada Y, 2006. Heterogeneity of strains assigned to *Gluconobacter frateurii* Mason and Claus 1989 based on restriction analysis of 16S-23S rDNA internal transcribed spacer regions. Bioscience, Biotechnology, and Biochemistry, 70, 684–690. https://doi.org/10.1271/bbb.70.684
- Marshall K, Shakya S, Greenhill AR, Padill G, Baker A and Warner JM, 2010. Antibiosis of *Burkholderia ubonensis* against *Burkholderia pseudomallei*, the causative agent for melioidosis. Southeast Asian Journal of Tropical Medicine and Public Health, 41, 904–912.
- Martina P, Leguizamon M, Prieto CI, Sousa SA, Montanaro P, Draghi WO, Stämmler M, Bettiol M, de Carvalho C, Palau J, Figoli C, Alvarez F, Benetti S, Lejona S, Vescina C, Ferreras J, Lasch P, Lagares A, Zorreguieta A, Leitão JH, Yantorno OM and Bosch A, 2018. *Burkholderia puraquae* sp. nov., a novel species of the *Burkholderia cepacia* complex isolated from hospital settings and agricultural soils. International Journal of Systematic and Evolutionary Microbiology, 68, 14–20. https://doi.org/10.1099/ijsem.0.002293
- Mason LM and Claus GW, 1989. Phenotypic Characteristics Correlated with Deoxyribonucleic Acid Sequence Similarities for Three Species of *Gluconobacter*: *G. oxydans* (Henneberg 1897) De Ley 1961, *G. frateurii* sp. nov., and *G. asaii* sp. nov. International Journal of Systematic Bacteriology, 39, https://doi.org/10.1099/00207713-39-2-174
- Menegueti MG, Gaspar GG, Laus AM, Basile-Filho A, Bellissimo-Rodrigues F and Auxiliadora-Martins M, 2018. Bacteremia by *Leuconostoc mesenteroides* in an immunocompetent patient with chronic Chagas disease: a case report. BMC Infectious Diseases, 18, 547. https://doi.org/10.1186/s12879-018-3452-7
- Moher D, Liberati A, Tetzlaff J, Altman DG and The PG, 2009. Preferred Reporting Items for Systematic Reviews and Meta-Analyses: The PRISMA Statement. PLOS Medicine, 6, e1000097. https://doi.org/10.1371/journal. pmed.1000097
- Mounier J, Gelsomino R, Goerges S, Vancanneyt M, Vandemeulebroecke K, Hoste B, Scherer S, Swings J, Fitzgerald GF and Cogan TM, 2005. Surface microflora of four smear-ripened cheeses. Applied and Environmental Microbiology, 71, 6489–6500. https://doi.org/10.1128/aem.71.11.6489-6500.2005
- Nagy F, Bozó A, Tóth Z, Daróczi L, Majoros L and Kovács R, 2018. *In vitro* antifungal susceptibility patterns of planktonic and sessile *Candida kefyr* clinical isolates. Medical Mycology, 56, 493–500. https://doi.org/10.1093/mmy/myx062
- Naqvi SSB, Nagendra V and Hofmeyr A, 2018. Probiotic related *Lactobacillus rhamnosus endocarditis* in a patient with liver cirrhosis. IDCases, 13, e00439. https://doi.org/10.1016/j.idcr.2018.e00439
- Nasseri AT, Rasoul-Amini S, Morowvat MH and Younes G, 2011. Single Cell Protein: Production and Process. American Journal of Food Technology, 6, 1–13. https://doi.org/10.3923/ajft.2011.103.116
- Neumann U, Louis S, Gille A, Derwenskus F, Schmid-Staiger U, Briviba K and Bischoff SC, 2018a. Anti-inflammatory effects of *Phaeodactylum tricornutum* extracts on human blood mononuclear cells and murine macrophages. Journal of Applied Phycology, 30, 2837–2846. https://doi.org/10.1007/s10811-017-1352-7
- Neumann U, Derwenskus F, Gille A, Louis S, Schmid-Staiger U, Briviba K and Bischoff SC, 2018b. Bioavailability and Safety of Nutrients from the Microalgae *Chlorella vulgaris, Nannochloropsis oceanica* and *Phaeodactylum tricornutum* in C57BL/6 Mice. Nutrients, 10, https://doi.org/10.3390/nu10080965
- Niccolai A, Bigagli E, Biondi N, Rodolfi L, Cinci L, Luceri C and Tredici MR, 2017. In vitro toxicity of microalgal and cyanobacterial strains of interest as food source. Journal of Applied Phycology, 29, 199–209. https://doi.org/10. 1007/s10811-016-0924-2



- Ochiai A, Ogawa K, Fukuda M, Ohori M, Kanaoka T, Tanaka T, Taniguchi M and Sagehashi Y, 2018. Rice Defensin OsAFP1 is a New Drug Candidate against Human Pathogenic Fungi. Scientific Reports, 8, 11434. https://doi.org/10.1038/s41598-018-29715-w
- Okmen F, Ekici H and Ari SA, 2018. Case Report of a Tubo-ovarian Abscess Caused by *Candida kefyr*. J Obstet Gynaecol Can, 40, 1466–1467. https://doi.org/10.1016/j.jogc.2018.04.025
- Oliveira A, Oliveira LC, Aburjaile F, Benevides L, Tiwari S, Jamal SB, Silva A, Figueiredo HCP, Ghosh P, Portela RW, De Carvalho Azevedo VA and Wattam AR, 2017. Insight of Genus *Corynebacterium*: Ascertaining the Role of Pathogenic and Non-pathogenic Species. Frontiers in Microbiology, 8, 1937. https://doi.org/10.3389/fmicb. 2017.01937
- Omran AN and Mansori AG, 2018. Pathogenic Yeasts Recovered From Acne Vulgaris: Molecular Characterization and Antifungal Susceptibility Pattern. Indian Journal of Dermatology, 63, 386–390. https://doi.org/10.4103/ijd. IJD_351_17
- Oren A and Garrity GM, 2015. List of new names and new combinations previously effectively, but not validly, published. International Journal of Systematic and Evolutionary Microbiology, 65, 2017–2025. https://doi.org/10.1099/ijs.0.000317
- Osman KM, Kappell AD, Orabi A, Al-Maary KS, Mubarak AS, Dawoud TM, Hemeg HA, Moussa IMI, Hessain AM, Yousef HMY and Hristova KR, 2018. Poultry and beef meat as potential seedbeds for antimicrobial resistant enterotoxigenic Bacillus species: a materializing epidemiological and potential severe health hazard. Scientific Reports, 8, 11600. https://doi.org/10.1038/s41598-018-29932-3
- de Paula Menezes R, Silva FF, Melo SGO, Alves PGV, Brito MO, de Souza Bessa MA, Amante Penatti MP, Pedroso RS, Abdallah VOS and Röder DVDB, 2018. Characterization of *Candida* species isolated from the hands of the healthcare workers in the neonatal intensive care unit. Medical Mycology, https://doi.org/10.1093/mmy/ myy101
- Pfaller MA, Rhomberg PR, Wiederhold NP, Gibas C, Sanders C, Fan H, Mele J, Kovanda LL and Castanheira M, 2018. In vitro activity of isavuconazole against opportunistic fungal pathogens from two mycology reference laboratories. Antimicrobial Agents and Chemotherapy, 62. pii: e01230-18. https://doi.org/10.1128/aac.01230-18
- Poljungreed I and Boonyarattanakalin S, 2018. Low-cost biotransformation of glycerol to 1,3-dihydroxyacetone through *Gluconobacter frateurii* in medium with inorganic salts only. Letters in Applied Microbiology, 67, 39–46. https://doi.org/10.1111/lam.12881
- Price EP, Sarovich DS, Webb JR, Hall CM, Jaramillo SA, Sahl JW, Kaestli M, Mayo M, Harrington G, Baker AL, Sidak-Loftis LC, Settles EW, Lummis M, Schupp JM, Gillece JD, Tuanyok A, Warner J, Busch JD, Keim P, Currie BJ and Wagner DM, 2017. Phylogeographic, genomic, and meropenem susceptibility analysis of *Burkholderia ubonensis*. PLoS Neglected Tropical Diseases, 11, e0005928. https://doi.org/10.1371/journal.pntd.0005928
- Rastogi A, Maheswari U, Dorrell RG, Vieira FRJ, Maumus F, Kustka A, McCarthy J, Allen AE, Kersey P, Bowler C and Tirichine L, 2018. Integrative analysis of large scale transcriptome data draws a comprehensive landscape of *Phaeodactylum tricornutum* genome and evolutionary origin of diatoms. Scientific Reports, 8, 4834. https://doi. org/10.1038/s41598-018-23106-x
- Rea MC, Gorges S, Gelsomino R, Brennan NM, Mounier J, Vancanneyt M, Scherer S, Swings J and Cogan TM, 2007. Stability of the biodiversity of the surface consortia of Gubbeen, a red-smear cheese. Journal of Dairy Science, 90, 2200–2210. https://doi.org/10.3168/jds.2006-377
- Réveillon D, Séchet V, Hess P and Amzil Z, 2016. Production of BMAA and DAB by diatoms (Phaeodactylum tricornutum, Chaetoceros sp., Chaetoceros calcitrans and Thalassiosira pseudonana) and bacteria isolated from a diatom culture. Harmful Algae, 58, 45–50. https://doi.org/10.1016/j.hal.2016.07.008
- Rhodes KA and Schweizer HP, 2016. Antibiotic resistance in *Burkholderia* species. Drug Resistance Updates, 28, 82–90. https://doi.org/10.1016/j.drup.2016.07.003
- Ruelle L, Deyi VYM, Konopnicki D and Dauby N, 2018. *Saccharomyces cerevisiae* fungemia: risk factors, outcome and link with Enterol (R) administration. Acta Clinica Belgica, 73, 8–8.
- Sadrossadati SZ, Ghahri M, Imani Fooladi AA, Sayyahfar S, Beyraghi S and Baseri Z, 2018. Phenotypic and genotypic characterization of *Candida species* isolated from candideamia in Iran. Curr Med Mycol, 4, 14–20. https://doi.org/10.18502/cmm.4.2.64
- Salomonsson ML, Fredriksson E, Alfjorden A, Hedeland M and Bondesson U, 2015. Seafood sold in Sweden contains BMAA: A study of free and total concentrations with UHPLC-MS/MS and dansyl chloride derivatization. Toxicol Rep, 2, 1473–1481. https://doi.org/10.1016/j.toxrep.2015.11.002
- Serrano A, Sebastián M, Arilla-Luna S, Baquedano S, Herguedas B, Velázquez-Campoy A, Martínez-Júlvez M and Medina M, 2017. The trimer interface in the quaternary structure of the bifunctional prokaryotic FAD synthetase from *Corynebacterium ammoniagenes*. Scientific Reports, 7, 404–404. https://doi.org/10.1038/s41598-017-00402-6
- Shah MR, Lutzu GA, Alam A, Sarker P, Kabir Chowdhury MA, Parsaeimehr A, Liang Y and Daroch M, 2018. Microalgae in aquafeeds for a sustainable aquaculture industry. Journal of Applied Phycology, 30, 197–213. https://doi.org/10.1007/s10811-017-1234-z



- Shimizu A, Hase R, Suzuki D, Toguchi A, Otsuka Y, Hirata N and Hosokawa N, 2019. *Lactococcus lactis* cholangitis and bacteremia identified by MALDI-TOF mass spectrometry: A case report and review of the literature on *Lactococcus lactis* infection. Journal of Infection and Chemotherapy: Official Journal of the Japan Society of Chemotherapy, 25, 141–146. https://doi.org/10.1016/j.jiac.2018.07.010
- Shokohi T, Aslani N, Ahangarkani F, Meyabadi MF, Hagen F, Meis JF, Boekhout T, Kolecka A and Badali H, 2018. *Candida infanticola* and *Candida spencermartinsiae* yeasts: Possible emerging species in cancer patients. Microbial Pathogenesis, 115, 353–357. https://doi.org/10.1016/j.micpath.2017.12.069
- Sievers M, Gaberthüel C, Boesch C, Ludwig W and Teuber M, 1995. Phylogenetic position of *Gluconobacter* species as a coherent cluster separated from all *Acetobacter* species on the basis of 16S ribosomal RNA sequences. FEMS Microbiology Letters, 126, 123–126. https://doi.org/10.1111/j.1574-6968.1995.tb07404.x
- Simi WB, Leite-Jr DP, Paula CR, Hoffmann-Santos HD, Takahara DT and Hahn RC, 2019. Yeasts and filamentous fungi in psittacidae and birds of prey droppings in midwest region of Brazil: a potential hazard to human health. Brazilian Journal of Biology, 79, 414–422.
- Simon RR, Vo TD and Levine R, 2016. Genotoxicity and subchronic toxicity evaluation of dried *Euglena gracilis* ATCC PTA-123017. Regulatory Toxicology and Pharmacology, 80, 71–81. https://doi.org/10.1016/j.yrtph.2016. 06.007
- Singla V, Mandal S, Sharma P, Anand S and Tomar SK, 2018. Antibiotic susceptibility profile of *Pediococcus* spp. from diverse sources. 3. Biotech, 8, 489. https://doi.org/10.1007/s13205-018-1514-6
- Skrede A, Mydland LT, Ahlstrøm Ø, Reitan KI, Gislerød HR and Øverland M, 2011. Evaluation of microalgae as sources of digestible nutrients for monogastric animals. Journal of Animal and Feed Sciences, 20, 131–142. https://doi.org/10.22358/jafs/66164/2011
- Soman R, Gupta N, Chaudhari P, Sunavala A, Shetty A and Rodrigues C, 2018. Implications for Diagnosis and Treatment of Infective Endocarditis: Eight year Experience of an Infectious Disease Team in a Private Tertiary Care Centre. Journal of the Association of Physicians of India, 66, 22–25.
- Sørensen M, Berge GM, Reitan KI and Ruyter B, 2016. Microalga *Phaeodactylum tricornutum* in feed for Atlantic salmon (*Salmo salar*) —Effect on nutrient digestibility, growth and utilization of feed. Aquaculture, 460, 116–123. https://doi.org/10.1016/j.aquaculture.2016.04.010
- Suzuki S, 2017. Large-Scale Cultivation of Euglena. Advances in Experimental Medicine and Biology, 979, 285–293. https://doi.org/10.1007/978-3-319-54910-1_14
- Suzuki S, Campos-Alberto E, Morita Y, Yamaguchi M, Toshimitsu T, Kimura K, Ikegami S, Katsuki T, Kohno Y and Shimojo N, 2018. Low Interleukin 10 Production at Birth Is a Risk Factor for Atopic Dermatitis in Neonates with Bifidobacterium Colonization. International Archives of Allergy and Immunology, 177, 342–349. https://doi.org/ 10.1159/000492130
- Teblick A, Jansens H, Dams K, Somville FJ and Jorens PG, 2018. Boerhaave's syndrome complicated by a *Saccharomyces cerevisiae* pleural empyema. Case report and review of the literature. Acta Clinica Belgica, 73, 377–381. https://doi.org/10.1080/17843286.2017.1398439
- Treguier P, David M, Gargala G, Camus V, Stamatoullas A, Menard AL, Lenain P, Contentin N, Lemasle E, Lanic H, Tilly H, Jardin F and Lepretre S, 2018. *Cyberlindnera jadinii* (teleomorph *Candida utilis*) candidaemia in a patient with aplastic anaemia: a case report. JMM Case Rep, 5, e005160. https://doi.org/10.1099/jmmcr.0. 005160
- Tsonis I, Karamani L, Xaplanteri P, Kolonitsiou F, Zampakis P, Gatzounis G, Marangos M and Assimakopoulos SF, 2018. Spontaneous cerebral abscess due to *Bacillus subtilis* in an immunocompetent male patient: a case report and review of literature. World Journal of Clinical Cases, 6, 1169–1174. https://doi.org/10.12998/wjcc. v6.i16.1169
- Wang JP, Kim JD, Kim JE and Kim IH, 2013. Amino acid digestibility of single cell protein from *Corynebacterium ammoniagenes* in growing pigs. Animal Feed Science and Technology, 180, 111–114. https://doi.org/10.1016/j.anifeedsci.2012.12.006
- Yabuuchi E, Kawamura Y, Ezaki T, Ikedo M, Dejsirilert S, Fujiwara N, Naka T and Kobayashi K, 2000. *Burkholderia uboniae* sp. nov., L-arabinose-assimilating but different from *Burkholderia thailandensis* and *Burkholderia vietnamiensis*. Microbiology and Immunology, 44, 307–317.
- Zakryś B, Milanowski R and Karnkowska A, 2017. Evolutionary Origin of *Euglena*. Advances in Experimental Medicine and Biology, 979, 3–17. https://doi.org/10.1007/978-3-319-54910-1_1
- Zeba F, Yirerong J, Assali M, Tewary G and Noska A, 2018. A Double Whammy: *Lactobacillus acidophilus* Bacteremia and Subsequent *Lactobacillus rhamnosus* Prosthetic Valve Infective Endocarditis in an Elderly Diabetic Patient. Rhode Island Medical Journal, 2013, 32–35.
- Zhang H, Tang Y, Zhang Y, Zhang S, Qu J, Wang X, Kong R, Han C and Liu Z, 2015. Fucoxanthin: A Promising Medicinal and Nutritional Ingredient. Evidence-Based Complementary and Alternative Medicine, 2015, 10. https://doi.org/10.1155/2015/723515

Glossary and Abbreviations

Antimicrobial compounds Antibiotics, bacteriocins and/or small peptides with antimicrobial activity antimicrobial resistance

BIOHAZ	EFSA Panel on Biological Hazards
BMAA	β-N-methylamino-L-alanine
ELS	Extensive Literature Search
FIP	EFSA Food Ingredients and Packaging Unit
FSTA	Food Science Technology Abstracts
GMM	genetically modified microorganism
IJSEM	International Journal of Systematic and Evolutionary Microbiology
LPSN	List of Prokaryotic Names with Standing in Nomenclature
MLT	machine learning technique
NOAEL	no-observed-adverse-effect-level
QPS	qualified presumption of safety
PPP	plant protection product
SCP	single cell protein
ToR	Terms of Reference
TU	taxonomic unit
WG	Working Group



Appendix A – Search strategy followed for the (re)assessment of the suitability of TUs notified to EFSA not present in the current QPS list for their inclusion in the updated list (reply to ToR 3)

Burkholderia ubonensis

A literature search was performed in PubMed database using the search term: "Burkholderia ubonensis": 18 citations were identified and screened.

Corynebacterium ammoniagenes

A literature search was performed in PubMed database using the search term: "Corynebacterium ammoniagenes": 112 citations were identified and screened.

Corynebacterium casei

A literature search was performed in Scopus database using the search term: "Corynebacterium casei": 125 citations were identified and screened from which, 40 were relevant for the assessment.

Euglena gracilis

Pub Med search; "Euglena gracilis", taxonomy; 88 references screened for relevance; two selected: Zakryś et al. (2017) for taxonomy and Krajčovič et al. (2015) for body of knowledge as review article

A literature search was performed in PubMed database using the search term:

- "E. gracilis" biotechnology 49 references.
- "E. gracilis" biomass 26 references.
- "E. gracilis" and safety, 5 references, Simon et al. (2016) as relevant.
- "E. gracilis" and illness: no references.
- "E. gracilis" and hospitalization: no references
- "E. gracilis" and infectivity: 6 references, Ebenezer et al. (2019) as relevant.
- "E. gracilis" and tox: 1 reference, not relevant.
- "E. gracilis" and health: 30 references, none relevant.
- "E. gracilis" and environment and concern: 1 reference, not relevant.
- "E. gracilis" and environment: 245 references, Suzuki et al. (2018) as relevant.

Gluconobacter frateurii

A literature search was performed in Web of Science core collection and in Pubmed using the search term: "Gluconobacter frateurii": 74 and 45 citations were identified and screened respectively.

Microbacterium foliorum

A search using the species name "Microbacterium foliorum" in the Web of Science core collection provided 29 hits.

Phaeodactylum tricornutum

Literature searches were performed in the Web of Science Core Collection. Using only the species name "Phaeodactylum tricornutum" gave 2,917 references. Using refined search terms gave the following numbers of references, which were all scanned at title level:

- "Phaeodactylum tricornutum" and (species characterization): 55 references.
- "Phaeodactylum tricornutum" and taxonom*: 55 references.
- "Phaeodactylum tricornutum" and ecology: 50 references.
- "Phaeodactylum tricornutum" and (secondary metabolit*): 17 references.
- "Phaeodactylum tricornutum" and infect*: 14 references.
- "Phaeodactylum tricornutum" and toxi*: 299 references.
- "Phaeodactylum tricornutum" and safety: 14 references.
- "Phaeodactylum tricornutum" and BMAA: 3 references.

Sphingomonas elodea

A literature search was performed in PubMed database using the search term: "Sphingomonas elodea": 26 hits were identified and screened, mainly dealing with the gellan gun production and the molecular characterisation of the enzymes and genes involved in this production.

Appendix B – Protocol for Extensive literature search (ELS), relevance screening, and article evaluation for the maintenance and update of list of QPS-recommended biological agents (reply to ToR 2)

The following protocol for extensive literature search (ELS) will be used in the context of the EFSA self-task mandate on the list of QPS-recommended biological agents intentionally added to the food or feed (EFSA-Q-2016-00684).

B.1. Description of the process

An ELS of studies related to safety concerns for humans, animals, plants and/or the environment of microorganisms recommended for the Qualified Presumption of Safety (QPS) 2019 list will be performed.

The process will be performed according to the following main steps:

- ELS for potentially relevant citations;
- Relevance screening to select the citations identified by the literature search, based on titles and abstract and then full-text;
- Evaluation of articles according to pre-specified categories of possible safety concerns;
- Discussion between experts to come to collective expert evaluation of the outcome, reflected in the QPS Opinion and Panel Statements.

Considering the purpose of the QPS approach, a broad search will be performed. The review questions will be broken down into key elements using the PECO conceptual model:

- Population of interest (P);
- Exposure of interest (E);
- Comparator (C);
- Outcomes of interest (O).

B.1.1. Objective

The aim is to identify any publicly available studies reporting on safety concerns for humans, animals or the environment caused by microorganisms on the QPS recommended list (see Appendix E).

B.1.2. Target population

The populations of interest are humans, animals, plants and the environment.

B.1.3. Exposure

Citations must report on at least one species included in one of the five groups of named species specified in the EFSA QPS recommended list of the QPS 2016 update ((see Table A.1 in Appendix A to (EFSA BIOHAZ Panel, 2017a)):

- a) Gram-positive non-spore-forming bacteria;
- b) Gram-positive spore-forming bacteria;
- c) Gram-negative bacteria;
- d) Viruses used for plant protection;
- e) Yeasts.

In more detail:

a) Gram-positive non-spore forming bacteria:

Bifidobacterium adolescentis, Bifidobacterium animalis, Bifidobacterium bifidum, Bifidobacterium breve, Bifidobacterium longum, Carnobacterium divergens, Corynebacterium glutamicum, Lactobacillus acidophilus, Lactobacillus amylolyticus, Lactobacillus animalis, Lactobacillus amylovorus, Lactobacillus alimentarius, Lactobacillus aviaries, Lactobacillus brevis, Lactobacillus buchneri, Lactobacillus casei, Lactobacillus cellobiosus, Lactobacillus collinoides, Lactobacillus coryniformis, Lactobacillus crispatus, Lactobacillus curvatus, Lactobacillus delbrueckii, Lactobacillus diolivorans Lactobacillus farciminis, Lactobacillus fermentum, Lactobacillus gallinarum, Lactobacillus gasseri, Lactobacillus helveticus, Lactobacillus hilgardii, Lactobacillus johnsonii, Lactobacillus kefiranofaciens, Lactobacillus kefiri, Lactobacillus mucosae, Lactobacillus panis, Lactobacillus paracasei, Lactobacillus paraplantarum, Lactobacillus pentosus, Lactobacillus plantarum, Lactobacillus pontis, Lactobacillus reuteri, Lactobacillus rhamnosus, Lactobacillus sakei, Lactobacillus salivarius, Lactobacillus sanfranciscensis, Lactococcus lactis, Leuconostoc citreum, Leuconostoc lactis, Leuconostoc mesenteroides, Leuconostoc pseudomesenteroides, Microbacterium imperiale, Oenococcus oeni, Pasteuria nishizawae, Pediococcus acidilactici, Pediococcus dextrinicus, Pediococcus parvulus, Pediococcus pentosaceus, Propionibacterium freudenreichii, Propionibacterium acidopropionici, Streptococcus thermophilus;

b) Gram-positive spore-forming bacteria:

Bacillus amyloliquefaciens, Bacillus atrophaeus, Bacillus clausii, Bacillus coagulans, Bacillus flexus, Bacillus fusiformis, Bacillus lentus, Bacillus licheniformis, Bacillus megaterium, Bacillus mojavensis, Bacillus pumilus, Bacillus smithii, Bacillus subtilis, Bacillus vallismortis, Geobacillus stearothermophilus;

c) Gram-negative bacteria:

Gluconobacter oxydans; Xanthomonas campestris;

d) Viruses used for plant protection:

Plant viruses (Family): *Alphaflexiviridae*, *Potyviridae*; Insect viruses (Family): *Baculoviridae*;

e) Yeasts:

Candida cylindracea, Debaryomyces hansenii, Hanseniaspora uvarum, Kluyveromyces lactis, Kluyveromyces marxianus, Komagataella pastoris, Lindnera jadinii, Ogataea angusta, Saccharomyces bayanus, Saccharomyces cerevisiae, Saccharomyces pastorianus, Schizosaccharomyces pombe, Wickerhamomyces anomalus, Xanthophyllomyces dendrorhous.

For the yeast species, as previously, the name of the teleomorphic form is used in the list of QPS species, when available. Important synonyms and older names were also included in the searches. For instance, names of the anamorphic growth forms were included, when such a form is known:

- Debaryomyces hansenii: anamorph Candida famata;
- Hanseniaspora uvarum: anamorph Kloeckera apiculata;
- Kluyveromyces lactis: anamorph Candida spherica;
- Kluyveromyces marxianus: anamorph Candida kefyr;
- Komagataella pastoris: synonym Pichia pastoris;
- Lindnera jadinii: synonyms Pichia jadinii, Hansenula jadinii, Torulopsis utilis, anamorph Candida utilis;
- Ogataea angusta: synonym Pichia angusta;
- Saccharomyces cerevisiae: synonym Saccharomyces boulardii;
- Saccharomyces pastorianus: synonym Saccharomyces carlsbergensis;
- Wickerhamomyces anomalus: synonyms Hansenula anomala, Pichia anomala, Saccharomyces anomalus, anamorph Candida pelliculosa;
- Xanthophyllomyces dendrorhous: anamorph Phaffia rhodozyma.

B.1.4. Comparator

It is expected that the prevalent study designs will be case reports or case series and studies based on surveys or isolate collections. The remaining study designs may include: studies using laboratory isolates; randomised controlled trials, field trials or experimental designs in the laboratory; experimental designs in live animals with a deliberate disease challenge; observational study designs; animal or insect models; investigations to identify or to understand the causes of safety concerns (e.g. identification, characterisation of toxic factors, virulence mechanisms); studies to demonstrate beneficial effects but with reporting of unwanted side-effects.

Since it is expected that in the majority of the study designs relevant for the review question, the comparator will not be available, the latter will not be included as a key element in the search strategy.



B.1.5. Outcomes of interest

The outcomes of interest to this ELS are:

Question 1:

- potential harms;
- safety issues;
- virulence or infectivity;
- intoxication.

Question 2:

• (acquired/intrinsic) antimicrobial resistance (AMR) covering phenotypic and genotypic aspects.

The QPS concept does not address hazards linked to the formulation or processing of the products based on biological agents added into the food or feed chain. Neither the safety of users handling the product nor the genetic modifications are taken into account.

B.1.6. Identification of the review questions

The following research questions will be addressed:

- Is there evidence of any safety concerns, including virulence features and toxin production, for humans, animals, plants and/or the environment associated with microbial species currently recommended for the QPS list since the previous QPS review (i.e. published from June 2016 until June 2019)?
- Is there evidence related to the presence or absence of antimicrobial resistance or antimicrobial resistance genes for the same microbial species published during the same time period?

B.2. Eligibility criteria for study selection

The selection of studies relevant to questions 1 and 2 will be performed applying the eligibility criteria described in Table B.1 below.

_	Criteria
Study design	No specific type of study design will be used to include/exclude relevant studies, although it is expected that the prevalent study designs will be case reports or case series and studies based on surveys or isolate collections
Study characteristics:	No exclusion will be based on study characteristics
Population	Humans, animals, plants, environment
Exposure Studies must report on at least one TU as identified in Section B.1.3	
Outcome of interest	Outcomes as listed in Section B.1.5
Language	English
Time	From June 2016 until end June 2019
Publication type	Primary research studies and secondary studies reporting previously unpublished primary studies

Table B.1:Eligibility criteria for questions 1 and 2

B.3. Literature searches

Searches will be conducted in a range of relevant information sources to identify any evidence of safety concerns and AMR regarding the target microbial species.

Considering the results of the previous QPS exercise, to handle the high number of studies identified in each group, 20 search strategies were prepared: three for yeasts, one for insect viruses, one for plant viruses, 13 for Gram-positive bacteria and two for Gram-negative bacteria according to named species specified by EFSA in the QPS recommended list of the QPS 2016 update (see Table A.1 in Appendix A to (EFSA BIOHAZ Panel, 2017a)).

The 20 subgroups of target microbial species will be searched separately.



Each search strategy will comprise two elements: the search terms (Section B.3.1) and the information sources (Section B.3.2) to be searched.

B.3.1. Search terms

The search strategies used to identify studies are given in Appendix C. Each strategy will comprise two key elements:

- Target microbial species as described in Section B.1.3 ('Exposure')
- Safety issues as described in Section B.1.5 ('Outcomes').

In order to maximise the sensitivity of the search for the species for which the number of overall publications in the relevant time period is expected to be low, the search strategy will not include outcome-related terms.

The population of interest (humans, animals, plants or the environment) will not be included as a key element in the search strategies, as it is often not explicitly described within a title or abstract. It would also have been difficult to describe adequately such a broad population using title/abstract words and/or subject headings. Population information will be captured at the time of evaluating the articles (see Section B.1 above).

Search terms for safety issues were identified in close collaboration with the information specialist; example of such terms, are the following: 'toxin*', 'disease*', 'infection*', 'clinical*', 'virulen*', 'antimicrobial resistan*', 'endocarditis'.

The 20 subgroups of target microbial species will be entered on separate search lines. The search line for each group will be combined with the safety terms individually.

The searches will not be limited by language or study design.

The review period will be from June 2016 to June 2019.

B.3.2. Information sources searched

The same information sources used for the previous QPS exercise (EFSA BIOHAZ Panel et al., 2017a) will be searched for studies reporting safety concerns regarding the target microbial species (see Table B.2 below).

Information source	Interface
Web of Science Core Collection	Web of Science, Thomson Reuters 2018
CAB Abstracts	Web of Science, Thomson Reuters 2018
BIOSIS Citation Index	Web of Science, Thomson Reuters 2018
MEDLINE	Web of Science, Thomson Reuters 2018
Food Science Technology Abstracts (FSTA)	Web of Science, Thomson Reuters 2018

Table B.2: Information sources to be searched to identify relevant studies

Search results will be downloaded from the information sources and imported into EndNote[®] X8 bibliographic management software. For each of the 20 species groups, within-group removal of duplicate entries will be done in EndNote[®] X8. Following uploading of the species groups into the DistillerSR¹² online software, removal of duplicates will again be undertaken, using the Duplicate Detection feature.

B.4. Study selection and article evaluation

To identify potentially relevant studies to be included in the review the studies will be selected by a three -step procedure using the DistillerSR online software.

The results of the different phases of the study selection process will be reported in a flowchart as recommended in the PRISMA statement on preferred reporting items for systematic reviews and metaanalyses (Moher et al., 2009).

¹² DistillerSR, Evidence Partners, Ottawa, Canada. https://www.evidencepartners.com/products/distillersr-systematic-review-sof tware/



B.4.1. Screening for potential relevance at title level

Articles will initially be screened at title level in parallel by two Working Group (WG) expert reviewers and, if needed, EFSA staff.

If the information in the title is not relevant for the research objectives, the article will not proceed to the next step (Section B.4.2).

Articles that will be excluded during screening at this step will be stored in Distiller SR. In case of doubts or divergences between the reviewers, the paper will proceed to step 2.

B.4.2. Screening for potential relevance at title and abstract level

The articles passing the first step will undergo a screening at abstract level in parallel by two experts. If the information in title and abstract is not relevant for the research objectives, the article will not proceed to the next step (Section B.4.3).

Articles that will be excluded during screening at this step will be stored in Distiller SR.

In case of doubts or divergences between the reviewers, the paper will proceed to step 3.

B.4.3. Article evaluation

The aim of this step will be to confirm that the article is relevant for the QPS project and, in case it is, to evaluate it. It will be carried out at full text level.

The articles passing the second step will undergo a validation procedure carried out by two experts. One reviewer will initially be tasked with the evaluation of a paper. The evaluation will be then forwarded to another reviewer for the validation of the appraisal received.

In case of disagreement with the initial appraisal, the second reviewer will write down their comments. The reviewers will initially try to solve the disagreement. In case this will not be possible, the conflicting information will be presented for collective expert evaluation of the ELS outcome (see Section B.5).

If the information contained in the article is not relevant for the research objectives, the article will not be evaluated. Articles that will not be considered relevant will be stored in Distiller SR.

B.4.3.1. Questions for study selection and article evaluation

STEP 1 (Screening for potential relevance):

Question 1: Is the full-text available, in English and dealing with safety concerns?

- Yes: Include and continue to Article evaluation form;
- Full text not available: Exclude;
- Full text not in English: Exclude;
- Full text in English but not dealing with safety concerns: Exclude.

STEP 2 (Article evaluation):

Question 2: Identification of the microorganisms

 The article will be characterised in terms of the microorganisms involved; Single choice question: the Experts will identify the microorganism/s described in the article. In case more than one microorganism is described in the paper, the form will be repeated for each microorganism.

Question 3: Is there any "methodological" problem identified in the paper under consideration?

- No problems identified;
- Yes some problems were identified.

Question 4: Which "methodological" problems were identified in the paper under consideration? (this question will appear in case in question 3 the option "Yes some problems were identified" will be selected)

- Methodology used for identity confirmation of the microorganism;
- Reliability of the source attribution;
- Misuse of the microorganism (e.g. parenteral exposure);
- Predisposing factors in the exposed subjects;
- Others.



When one of the above options will be selected a dedicated free text box will appear to describe the problem identified.

Question 5: Is there any safety concern identified? (this question will appear in case in question 3 the option "No problems identified" will be selected)

- No safety concerns identified;
- Yes some safety concerns were identified.

Question 6: Which safety concerns were identified? (this question will appear in case in question 5 the option "Yes some safety concerns were identified" will be selected)

- On human health;
- On animal health;
- On the environment;
- On AMR;
- On other aspects.

When one of the above options will be selected a dedicated free text box will appear to describe the safety concern identified.

Question 7: Overall, is there any information that could potentially lead to a change in the QPS status of the microorganism? (this question will appear in case in question 5 the option "Yes some safety concerns were identified" will be selected)

- No;
- Yes.

In case the option "Yes" will be selected a dedicated free text box will appear to describe the information that could potentially lead to a change in the QPS status of the microorganism.

B.5. Collective expert evaluation of the ELS outcome and presentation in the QPS opinion

The overall results of the searches and evaluations of individual articles will be presented in tabular format for each group/subgroup and species. These results will be further evaluated collectively by the working group and the outcome will be reflected in the QPS opinion.

B.6. Update of the process

The literature search, study selection and collective expert evaluation will be repeated every 6 months.

References

EFSA BIOHAZ Panel (EFSA Panel on Biological Hazards), 2017. Scientific Opinion on the update of the list of QPS-recommended biological agents intentionally added to food or feed as notified to EFSA. EFSA Journal 2017;15(1):4664, 177 pp. https://doi.org/10.2903/j.efsa.2017.4664

Moher D, Liberati A, Tetzlaff J, Altman DG and the PRISMA Group, 2009. Preferred reporting items for systematic reviews and meta-analyses: the PRISMA statement. PLoS Med, 6, e1000097.



Appendix C – Search strategies for the maintenance and update of list of QPS-recommended biological agents (reply to ToR 2)

Gram-Positive Non-Spore-forming Bacteria

Bifidobacterium spp.

String for species	
"Bifidobacterium adolescentis" OR "Bifidobacterium animalis" OR "Bifidobacterium bifidum" OR "Bifidobacterium breve" OR "Bifidobacterium longum" OR "B adolescentis" OR "B animalis" OR "B bifidum" OR "B breve" OR "B longum"	
OUTCOME	String
1. Antimicrobial/Antibiotic/Antimycotic	antimicrobial resistan* OR antibiotic resistan* OR antimicrobial susceptibil*
2. Infection/Bacteremia/Fungemia/Sepsis	infection* OR abscess* OR sepsis* or septic* OR bacteremia OR bacteraemia OR toxin*
3. Type of disease	endocarditis OR abscess OR meningitis
4. Mortality/Morbidity	clinical* OR death* OR morbidit* OR mortalit* OR disease* OR illness*
5. Disease Risk	opportunistic OR virulen*

Carnobacterium divergens

String for species	
"Carnobacterium divergens" OR "C divergens"	
OUTCOME	String
6. Antimicrobial/Antibiotic/Antimycotic	Not applied
7. Infection/Bacteremia/Fungemia/Sepsis	Not applied
8. Type of disease	Not applied
9. Mortality/Morbidity	Not applied
10. Disease Risk	Not applied

Corynebacterium glutamicum

String for species	
"Corynebacterium glutamicum" OR "C glutamicum" OR "Brevibacterium lactofermentum" OR "B lactofermentum"	
OUTCOME	String
1. Antimicrobial/Antibiotic/Antimycotic	antimicrobial resistan* OR antibiotic resistan* OR antimicrobial susceptibil*
2. Infection/Bacteremia/Fungemia/Sepsis	infection* OR abscess* OR sepsis* or septic* OR bacteremia OR bacteraemia OR toxin* OR pathogen*
3. Type of disease	Not applied
4. Mortality/Morbidity	clinical* OR death* OR morbidit* OR mortalit* OR disease* OR illness*
5. Disease Risk	opportunistic OR virulen*



Lactobacillus spp.

String for species	
"Lactobacillus acidophilus" OR "Lactobacillus amylolyticus"	
OR "Lactobacillus amylovorus" OR "Lactobacillus	
alimentarius" OR "Lactobacillus animalis" OR "Lactobacillus	
aviaries" OR "Lactobacillus brevis" OR "Lactobacillus	
buchneri" OR "Lactobacillus casei" OR "Lactobacillus zeae"	
OR "Lactobacillus cellobiosus" OR "Lactobacillus	
coryniformis" OR "Lactobacillus crispatus" OR "Lactobacillus	
curvatus" OR "Lactobacillus delbrueckii" OR "Lactobacillus	
diolivorans" OR "Lactobacillus farciminis" OR "Lactobacillus	
fermentum" OR "Lactobacillus gallinarum" OR	
"Lactobacillus gasseri" OR "Lactobacillus helveticus" OR	
"Lactobacillus hilgardii" OR "Lactobacillus johnsonii" OR	
"Lactobacillus kefiranofaciens" OR "Lactobacillus kefiri" OR	
"Lactobacillus mucosae" OR "Lactobacillus panis" OR	
"Lactobacillus collinoides" OR "Lactobacillus paracasei" OR	
"Lactobacillus paraplantarum" OR "Lactobacillus pentosus"	
OR "Lactobacillus plantarum" OR "Lactobacillus pontis" OR	
"Lactobacillus reuteri" OR "Lactobacillus rhamnosus" OR	
"Lactobacillus sakei" OR "Lactobacillus salivarius" OR	
"Lactobacillus sanfranciscensis" OR "L acidophilus" OR "L	
amylolyticus" OR "L amylovorus" OR "L alimentarius" OR "L	
animalis" OR "L aviaries" OR "L brevis" OR "L buchneri" OR	
"L casei" OR "L zeae" OR "L cellobiosus" OR "L coryniformis"	
OR "L crispatus" OR "L curvatus" OR "L delbrueckii" OR "L	
diolivorans" OR "L farciminis" OR "L fermentum" OR "L	
gallinarum" OR "L gasseri" OR "L helveticus" OR "L hilgardii"	
OR "L johnsonii" OR "L kefiranofaciens" OR "L kefiri" OR "L	
mucosae" OR "L panis" OR "L collinoides" OR "L paracasei"	
OR "L paraplantarum" OR "L pentosus" OR "L plantarum"	
OR "L pontis" OR "L reuteri" OR "L rhamnosus" OR "L sakei"	
OR "L salivarius" OR "L sanfranciscensis"	
OUTCOME	String
1. Antimicrobial/Antibiotic/Antimycotic	antimicrobial resistan* OR antibiotic resistan* OR
	antimicrobial susceptibil*
2. Infection/Bacteremia/Fungemia/Sepsis	infection* OR abscess* OR sepsis* or septic* OR
	bacteremia OR bacteraemia OR toxin*
3. Type of disease	endocarditis OR abscess OR meningitis
4. Mortality/Morbidity	Not applied
5. Disease Risk	opportunistic OR virulen*

Lactococcus lactis

String for species	
"Lactococcus lactis" OR "L lactis"	
OUTCOME	String
1. Antimicrobial/Antibiotic/Antimycotic	antimicrobial resistan* OR antibiotic resistan* OR antimicrobial susceptibil*
2. Infection/Bacteremia/Fungemia/Sepsis	infection* OR abscess* OR sepsis* or septic* OR bacteremia OR bacteraemia OR toxin*
3. Type of disease	endocarditis OR abscess OR meningitis
4. Mortality/Morbidity	clinical* OR death* OR morbidit* OR mortalit* OR disease* OR illness*
5. Disease Risk	opportunistic OR virulen*



Leuconostoc spp.

String for species	
"Leuconostoc mesenteroides" OR "Leuconostoc lactis" OR "Leuconostoc pseudomesenteroides" OR "Leuconostoc citreum" OR "L mesenteroides" OR "L lactis" OR "L pseudomesenteroides" OR "L citreum"	
OUTCOME	String
1. Antimicrobial/Antibiotic/Antimycotic	antimicrobial resistan* OR antibiotic resistan* OR antimicrobial susceptibil*
2. Infection/Bacteremia/Fungemia/Sepsis	infection* OR abscess* OR sepsis* or septic* OR bacteremia OR bacteraemia OR toxin*
3. Type of disease	Not applied
4. Mortality/Morbidity	clinical* OR death* OR morbidit* OR mortalit* OR disease* OR illness*
5. Disease Risk	opportunistic OR virulen*

Microbacterium imperiale

String for species	
"Microbacterium imperiale" OR "M imperiale"	
OUTCOME	String
6. Antimicrobial/Antibiotic/Antimycotic	Not applied
7. Infection/Bacteremia/Fungemia/Sepsis	Not applied
8. Type of disease	Not applied
9. Mortality/Morbidity	Not applied
10. Disease Risk	Not applied

Oenococcus spp.

String for species	
"Oenococcus oeni" OR "O oeni"	
OUTCOME	String
1. Antimicrobial/Antibiotic/Antimycotic	Not applied
2. Infection/Bacteremia/Fungemia/Sepsis	Not applied
3. Type of disease	Not applied
4. Mortality/Morbidity	Not applied
5. Disease Risk	Not applied

Pasteuria nishizawae

String for species	
"Pasteuria nishizawae" OR "P nishizawae"	
OUTCOME	String
11. Antimicrobial/Antibiotic/Antimycotic	Not applied
12. Infection/Bacteremia/Fungemia/Sepsis	Not applied
13. Type of disease	Not applied
14. Mortality/Morbidity	Not applied
15. Disease Risk	Not applied



Pediococcus spp.

String for species	
"Pediococcus pentosaceus" OR "Pediococcus dextrinicus" OR "Pediococcus acidilactici" OR "Pediococcus parvulus" OR "P pentosaceus" OR "P dextrinicus" OR "P acidilactici" OR "P parvulus"	
OUTCOME	String
1. Antimicrobial/Antibiotic/Antimycotic	Not applied
2. Infection/Bacteremia/Fungemia/Sepsis	Not applied
3. Type of disease	Not applied
4. Mortality/Morbidity	Not applied
5. Disease Risk	Not applied

Propionibacterium spp.

String for species	Number papers retrieved and notes
"Propionibacterium acidipropionici" OR "Propionibacterium freudenreichii" OR "P acidipropionici" OR "P freudenreichii"	176
OUTCOME	String
1. Antimicrobial/Antibiotic/Antimycotic	Not applied
2. Infection/Bacteremia/Fungemia/Sepsis	Not applied
3. Type of disease	Not applied
4. Mortality/Morbidity	Not applied
5. Disease Risk	Not applied

Streptococcus thermophilus

String for species	
"Streptococcus thermophilus" OR "S thermophilus" "Streptococcus thermophilus" OR "S thermophilus"	
OUTCOME	String
1. Antimicrobial/Antibiotic/Antimycotic	antimicrobial resistan* OR antibiotic resistan* OR antimicrobial susceptibil*
2. Infection/Bacteremia/Fungemia/Sepsis	infection* OR abscess* OR sepsis* or septic* OR bacteremia OR bacteraemia OR toxin*
3. Type of disease	Not applied
4. Mortality/Morbidity	clinical* OR death* OR morbidit* OR mortalit* OR disease* OR illness*
5. Disease Risk	opportunistic OR virulen*



Gram-positive spore-forming bacteria

Bacillus spp.

String for species	
"Bacillus amyloliquefaciens" OR "Bacillus coagulans" OR "Bacillus clausii" OR "Bacillus atrophaeus" OR "Bacillus flexus" OR "Bacillus fusiformis" OR "Lysinibacillus fusiformis" OR "Bacillus licheniformis" OR "Bacillus lentus" OR "Bacillus mojavensis" OR "Bacillus megaterium" OR "Bacillus vallismortis" OR "Bacillus smithii" OR "Bacillus subtilis" OR "Bacillus pumilus" OR "Geobacillus stearothermophilus" OR "B amyloliquefaciens" OR "B flexus" OR "B clausii" OR "B atrophaeus" OR "B flexus" OR "B flexus" OR "B itusiformis" OR "B licheniformis" OR "B lentus" OR "B smithii" OR "B megaterium" OR "B vallismortis" OR "B smithii" OR "B subtilis" OR "B pumilus" OR "B smithii" OR "B subtilis" OR "B pumilus" OR "B stearothermophilus"	
OUTCOME	String
1. Antimicrobial/Antibiotic/Antimycotic	antimicrobial resistan* OR antibiotic resistan* OR antimicrobial susceptibil*
2. Infection/Bacteremia/Fungemia/Sepsis	infection* OR abscess* OR sepsis* or septic* OR bacteremia OR bacteraemia OR toxin*
3. Type of disease	endocarditis OR abscess OR meningitis
4. Mortality/Morbidity	Not applied
5. Disease Risk	opportunistic OR virulen*

Gram-negative bacteria

Gluconobacter oxydans

String for species	
"Gluconobacter oxydans" OR "G oxydans"	
OUTCOME	String
1. Antimicrobial/Antibiotic/Antimycotic	Not applied
2. Infection/Bacteremia/Fungemia/Sepsis	Not applied
3. Type of disease	Not applied
4. Mortality/Morbidity	Not applied
5. Disease Risk	Not applied

Xanthomonas campestris

String for species	
"Xanthomonas campestris" OR "X campestris"	
OUTCOME	String
1. Antimicrobial/Antibiotic/Antimycotic	Not applied
2. Infection/Bacteremia/Fungemia/Sepsis	Not applied
3. Type of disease	Not applied
4. Mortality/Morbidity	Not applied
5. Disease Risk	Not applied



Yeasts

TUs without keywords for OUTCOME

String for species	
"Candida cylindracea" OR "Debaryomyces hansenii" OR "Candida famata" OR "Hanseniaspora uvarum" OR "Kloeckera apiculata" OR "Ogataea angusta" OR "Pichia angusta" OR "Saccharomyces bayanus" OR "Saccharomyces pastorianus" OR "Saccharomyces carlsbergensis" OR "Wickerhamomyces anomalus" OR "Hansenula anomala" OR "Pichia anomala" OR "Saccharomyces anomalus" OR "Candida pelliculosa" OR "Xanthophyllomyces dendrorhous" OR "Phaffia rhodozyma" OR "C cylindracea" OR "D hansenii" OR "C famata" OR "H uvarum" OR "K apiculata" OR "O angusta" OR "P angusta" OR "S bayanus" OR "S pastorianus" OR "S carlsbergensis" OR "W anomalus" OR "H anomala" OR "P anomala" OR "S anomalus" OR "C pelliculosa" OR "X dendrorhous" OR "P rhodozyma"	
OUTCOME	String
1. Antimicrobial/Antibiotic/Antimycotic	Not applied
2. Infection/Bacteremia/Fungemia/Sepsis	Not applied
3. Type of disease	Not applied
4. Mortality/Morbidity	Not applied
5. Disease Risk	Not applied

TUs with keywords for OUTCOME except for type of disease and morbility/mortality

String for species	
"Kluyveromyces lactis" OR "Candida spherica" OR "Kluyveromyces marxianus" OR "Candida kefyr" OR "Komagataella pastoris" OR "Pichia pastoris" OR "Lindnera jadinii" OR "Pichia jadinii" OR "Hansenula jadinii" OR "Torulopsis utilis" OR "Candida utilis" OR "Schizosaccharomyces pombe" OR "K lactis" OR "C spherica" OR "K marxianus" OR "C kefyr" OR "K pastoris" OR "P pastoris" OR "L jadinii" OR "P jadinii" OR "H jadinii" OR "T utilis" OR "C utilis" OR "S pombe"	
OUTCOME	String
1. Antimicrobial/Antibiotic/Antimycotic	antimicrobial resistan* OR antimycotic resistan* OR antimicrobial susceptibil*
2. Infection/Bacteremia/Fungemia/Sepsis	infection* OR abscess* OR sepsis* or septic* OR fungemia OR fungaemia OR mycos*
3. Type of disease	Not applied
4. Mortality/Morbidity	Not applied
5. Disease Risk	opportunistic OR virulen*

TUs with keywords for OUTCOME except for type of disease

String for species	
"Saccharomyces cerevisiae" OR "Saccharomyces boulardii" OR "Scerevisiae" OR "Sboulardii"	
OUTCOME	String
1. Antimicrobial/Antibiotic/Antimycotic	antimicrobial resistan* OR antimycotic resistan* OR antimicrobial susceptibil*

2. Infection/Bacteremia/Fungemia/Sepsis	infection* OR abscess* OR sepsis* or septic* OR fungemia OR fungaemia OR mycos*
3. Type of disease	Not applied
4. Mortality/Morbidity	clinical* OR death* OR morbidit* OR mortalit* OR disease* OR illness*
5. Disease Risk	opportunistic OR virulen*

Viruses used for plant protection

Alphaflexiviridae

String for species	
"Alphaflexiviridae" OR "Potyviridae"	
OUTCOME	String
1. Antimicrobial/Antibiotic/Antimycotic	Not applied
2. Infection/Bacteremia/Fungemia/Sepsis	necros*
3. Type of disease	Not applied
4. Mortality/Morbidity	mortalit* OR safety concern* OR "health hazard"
5. Disease Risk	virulen*

Baculoviridae

String for species	
"Nuclear polyhedrosis virus" OR "granulovirus" OR "baculoviridae"	
OUTCOME	String
1. Antimicrobial/Antibiotic/Antimycotic	Not applied
2. Infection/Bacteremia/Fungemia/Sepsis	Not applied
3. Type of disease	"nuclear polyhedrosis" OR granulosis
4. Mortality/Morbidity	mortalit* OR safety concern* OR "health hazard"
5. Disease Risk	Not applied



Appendix D – References selected from the ELS exercise as relevant for the QPS for searches from July to December 2018 (reply to ToR 2)

Gram-Positive Non-Sporulating Bacteria

Bifidobacterium spp.

- Kim MJ, Seockmo K, Sun Young K, Hyun Ha L, Hui J, Sini K, Rui L, Johnston TV, Myeong Soo P and Geun Eog J, 2018. Safety evaluations of *Bifidobacterium bifidum* BGN4 and *Bifidobacterium longum* BORI. International Journal of Molecular Sciences, 19, 1422–1422.
- Magistrelli L, Amoruso A, Milner AV, Mogna L, Cantello R, Pane M and Comi C, 2018. Effects of probiotic bacterial strains on peripheral inflammation in Parkinson's disease. European Journal of Neurology, 25, 428–428.
- Suzuki S, Campos-Alberto E, Morita Y, Yamaguchi M, Toshimitsu T, Kimura K, Ikegami S, Katsuki T, Kohno Y and Shimojo N, 2018. Low Interleukin 10 Production at Birth Is a Risk Factor for Atopic Dermatitis in Neonates with *Bifidobacterium* Colonization. International Archives of Allergy and Immunology, 177, 342–349.

Carnobacterium divergens

None

Corynebacterium glutamicum

None

Lactobacilli spp.

- Arokiyaraj S, Seo SS, Kwon M, Lee JK and Kim MK, 2018. Association of cervical microbial community with persistence, clearance and negativity of Human Papillomavirus in Korean women: a longitudinal study. Scientific Reports, 8, 15479.
- Costa RL, Moreira J, Lorenzo A and Lamas CC, 2018. Infectious complications following probiotic ingestion: a potentially underestimated problem? A systematic review of reports and case series. Bmc Complementary and Alternative Medicine, 18, 329.
- Esaiassen E, Hjerde E, Cavanagh JP, Pedersen T, Andresen JH, Rettedal SI, Stoen R, Nakstad B, Willassen NP and Klingenberg C, 2018. Effects of Probiotic Supplementation on the Gut Microbiota and Antibiotic Resistome Development in Preterm Infants. Frontiers in Pediatrics, 6, 347.
- Griff PM, Schleper A, Lynch CA, Sun Y and Butler TR, 2018. CHRONIC ADMINISTRATION OF PROBIOTIC L. RHAMNOSUS AFFECTS ANXIETY-LIKE BEHAVIOR IN A MODEL OF ALCOHOL USE DISORDER VULNERABILITY. Alcoholism-Clinical and Experimental Research, 42, 35A–35A.
- Hojsak I, Fabiano V, Pop TL, Goulet O, Zuccotti GV, Cokugras FC, Pettoello-Mantovani M and Kolacek S, 2018. Guidance on the use of probiotics in clinical practice in children with selected clinical conditions and in specific vulnerable groups. Acta Paediatrica, 107, 927–937.
- Kao, B-Z, Lin H-J, Chen M-Y, Wu C-S, Lin S-T, Lee M-H, Lai Y-X and Hu P-J, 2018. *Lactobacillus paracasei* as cause of liver abscess: Case report. Journal of Gastroenterology and Hepatology, 33, 433–433.
- Kawai K, Kamochi R, Oiki S, Murata K and Hashimoto W, 2018. Probiotics in human gut microbiota can degrade host glycosaminoglycans. Scientific Reports, 8, 10674.
- Maillet F, Passeron A, Podglajen I, Ranque B and Pouchot J, 2018. *Lactobacillus delbrueckii* urinary tract infection in a male patient. Medecine et maladies infectieuses.
- Naqvi SSB, Nagendra V and Hofmeyr A, 2018. Probiotic related *Lactobacillus rhamnosus* endocarditis in a patient with liver cirrhosis. IDCases, 13, e00439–e00439.
- Okba A, Shahin R, Attallah A, Sheha D, Mkawy M and Farag M, 2018. ROLE OF INTESTINAL MICROBIOTA IN CARDIOVASCULAR DISEASE RISK IN END STAGE RENAL DISEASE PATIENTS. Nephrology Dialysis Transplantation, 33, 573–574.
- Zawistowska-Rojek A and Tyski S, 2018. Are Probiotic Really Safe for Humans? Polish Journal of Microbiology, 67, 251–258.
- Zeba F, Yirerong J, Assali M, Tewary G and Noska A, 2018. A Double Whammy: *Lactobacillus acidophilus* Bacteremia and Subsequent *Lactobacillus rhamnosus* Prosthetic Valve Infective Endocarditis in an Elderly Diabetic Patient. Rhode Island Medical Journal (2013), 101, 32–35.

Lactococcus lactis

- Chen F, Zhang Z and Chen J, 2018. Infective endocarditis caused by *Lactococcus lactis* subsp. *lactis* and *Pediococcus pentosaceus*: A case report and literature review. Medicine, 97, e13658–e13658.
- Kabore WAD, Dembele R, Bagre TS, Konate A, Boisrame S, Chevalier V, Konsem T, Traore AS and Barro N, 2018. Characterization and Antimicrobial Susceptibility of *Lactococcus lactis* Isolated from Endodontic Infections in Ouagadougou, Burkina Faso." Dentistry Journal, 6.
- Kato Y, Kanayama M, Yanai S, Nozawa H, Kanauchi O and Suzuki S, 2018. Safety evaluation of excessive intake of *Lactococcus lactis* subsp. *lactis* JCM 5805: a randomized, double-blind, placebo-controlled, parallel-group trial. Food and Nutrition Sciences, 9, 403–419.
- Shimizu A, Hase R, Suzuki D, Toguchi A, Otsuka Y, Hirata N and Hosokawa N, 2019. *Lactococcus lactis* cholangitis and bacteremia identified by MALDI-TOF mass spectrometry: A case report and review of the literature on *Lactococcus lactis* infection. Journal of infection and chemotherapy : official journal of the Japan Society of Chemotherapy, 25, 141–146.

Leuconostoc spp.

- Chen F, Zhang Z and Chen J, 2018. Infective endocarditis caused by *Lactococcus lactis* subsp. *lactis* and *Pediococcus pentosaceus*: A case report and literature review. Medicine, 97, e13658–e13658.
- Coton M, Lebreton M, Salas ML, Garnier L, Navarri M, Pawtowski A, Le Blay G, Valence F, Coton E and Mounier J, 2018. Biogenic amine and antibiotic resistance profiles determined for lactic acid bacteria and a propionibacterium prior to use as antifungal bioprotective cultures. International Dairy Journal, 85, 21–26.
- Menegueti MG, Gaspar GG, Laus AM, Basile-Filho A, Bellissimo-Rodrigues F and Auxiliadora-Martins M, 2018. Bacteremia by *Leuconostoc mesenteroides* in an immunocompetent patient with chronic Chagas disease: a case report. BMC Infectious Diseases, 18.
- Mussano F, Ferrocino I, Gavrilova N, Genova T, Dell'Acqua A, Cocolin L and Carossa S, 2018. Apical periodontitis: preliminary assessment of microbiota by 16S rRNA high throughput amplicon target sequencing. BMC Oral Health, 18.
- Shimizu A, Hase R, Suzuki D, Toguchi A, Otsuka Y, Hirata N and Hosokawa N, 2019. *Lactococcus lactis* cholangitis and bacteremia identified by MALDI-TOF mass spectrometry: a case report and review of the literature on *Lactococcus lactis* infection. Journal of infection and chemotherapy : official journal of the Japan Society of Chemotherapy, 25, 141–146.
- Ting C, Qianwen L, Wenliang X, Qing Z, Qisheng Z, Gong C and Yimin C, 2017. Antibiotic resistance evaluation and resistance gene profile of epibiotic lactic acid bacteria on red bell peppers used for Sichuan pickle fermentation. Food Science, China, 38, 27–33.

Microbacterium imperiale

None

Oenococcus oeni

None

Pasteuria nishizawae

None

Pediococci spp.

Chen F, Zhang Z and Chen J, 2018. Infective endocarditis caused by *Lactococcus lactis* subsp. *lactis* and *Pediococcus pentosaceus*: A case report and literature review. Medicine, 97, e13658–e13658.

Han A, Mehta J and Pauly RR, 2016. Septic shock secondary to a urinary tract infection with *Pediococcus pentosaceus*. Missouri Medicine, 113, 179–181.

Propionibacterium spp.

None

Streptococcus thermophilus

None

Gram-Positive Spore-forming Bacteria

Bacillus spp.

- (2018). Selection of Potential Probiotic Lactic Acid Bacteria Isolated from Palm Sap (*Borassus flabellifer* Linn.) Origin Kupang, East Nusa Tenggara. Inventing Prosperous Future through Biological Research and Tropical Biodiversity Management 2002.
- Abdelgani AM, Rukayadi Y, Shaari K and Safinar I, 2018. Antibacterial and sporicidal activities of methanolic *Sygygium polyanthum* L. Leaf extract against vegetative cells and spores of *Bacillus pumilus* and Bacillus *megaterium*. Journal of Pure and Applied Microbiology, 12, 1047-1053.
- Abreo E, Valle D, Mujica V and Altier N, 2018. Pathogenicity and virulence factors of *Lysinibacillus xylanilyticus* and *Bacillus* spp. towards *Argyrotaenia sphaleropa* larvae (Lepidoptera). Journal of Applied Entomology, 142, 882–892.
- Aditya K, Babu PPS and Roy SD, 2018. Assessment of immune response in post-probiotic treated *Litopenaeus vannamei* challenged with *Vibrio harveyi*. Proceedings of the National Academy of Sciences India. Section B, Biological Sciences, 88, 797–802.
- Agerso Y, Stuer-Lauridsen B, Bjerre K, Jensen MG, Johansen E, Bennedsen M, Brockmann E and Nielsen B, 2018. Antimicrobial susceptibility testing and tentative epidemiological cutoff values for five *Bacillus* species relevant for use as animal feed additives or for plant protection. Applied and Environmental Microbiology, 84.
- Ali M, Boonerjee S, Islam MN, Saha ML, Hoque MI and Sarker RH, 2018. Endogenous bacterial contamination of plant tissue culture materials: identification and control strategy. Plant Tissue Culture & Biotechnology, 28, 99–108.
- Al-Khalaifah HS, 2018. Benefits of probiotics and/or prebiotics for antibiotic-reduced poultry. Poultry Science, 97, 3807–3815.
- Al-Marzooq F, Al Bayat S, Sayyar F, Ishaq H, Nasralla H, Koutaich R and Al Kawas S, 2018. Can probiotic cleaning solutions replace chemical disinfectants in dental clinics? European Journal of Dentistry, 12, 532-539.
- Alonso Lucero-Velasco E, Judith Molina-Garza Z and Galaviz-Silva L, 2018. First survey of cultivable bacteria from *Rhipicephalus sanguineus sensu lato* and assessment of the antagonism against five microorganisms of clinical importance. International Journal of Acarology, 44, 204–209.
- Alsanie WF, Felemban EM, Farid MA, Hassan MM, Sabry A and Gaber A, 2018. Molecular identification and phylogenetic analysis of multidrug-resistant bacteria using 16S rDNA sequencing. Journal of Pure and Applied Microbiology, 12, 489–496.
- Amman F, D'Halluin A, Antoine R, Huot L, Bibova I, Keidel K, Slupek S, Bouquet P, Coutte L, Caboche S, Locht C, Vecerek B and Hot D, 2018. Primary transcriptome analysis reveals importance of IS elements for the shaping of the transcriptional landscape of *Bordetella pertussis*. RNA Biology, 15, 967–975.
- Ansari A, Zohra RR, Tarar OM, UI Qader SA and Aman A, 2018. Screening, purification and characterization of thermostable, protease resistant Bacteriocin active against methicillin resistant *Staphylococcus aureus* (MRSA). BMC Microbiology, 18.
- Bao L, Fang Q, Lou B, Wu D, Chen Y and Liu J, 2018. Diagnostic value of inflammatory factors in patients with sepsis caused by infections. Chinese Journal of Nosocomiology, 28, 547–550.
- Ben Abdallah D, Frikha-Gargouri O and Tounsi S, 2018. Rizhospheric competence, plant growth promotion and biocontrol efficacy of *Bacillus amyloliquefaciens* subsp plantarum strain 32a. Biological Control, 124, 61–67.
- Berger P, Messner MJ, Crosby J, Renwick DV and Heinrich A, 2018. On the use of total aerobic spore bacteria to make treatment decisions due to *Cryptosporidium* risk at public water system wells. International Journal of Hygiene and Environmental Health, 221, 704–711.
- Beric T, Biocanin M, Stankovic S, Dimkic I, Janakiev T, Fira D and Lozo J, 2018. Identification and antibiotic resistance of *Bacillus* spp. isolates from natural samples. Archives of Biological Sciences, 70, 581–588.
- Bjelic D, Ignjatov M, Marinkovic J, Milosevic D, Nikolic Z, Gvozdanovic-Varga J and Karaman M, 2018. *Bacillus* isolates as potential biocontrol agents of *Fusarium* clove rot of garlic. Zemdirbyste-Agriculture, 105, 369–376.
- Carles L, Joly M, Bonnemoy F, Leremboure M, Donnadieu F, Batisson I and Besse-Hoggan P, 2018. Biodegradation and toxicity of a maize herbicide mixture: mesotrione, nicosulfuron and Smetolachlor. Journal of Hazardous Materials, 354, 42–53.

- Caselli E, Pancaldi S, Baldisserotto C, Petrucci F, Impallaria A, Volpe L, D'Accolti M, Soffritti I, Coccagna M, Sassu G, Bevilacqua F, Volta A, Bisi M, Lanzoni L and Mazzacane S, 2018. Characterization of biodegradation in a 17th century easel painting and potential for a biological approach. Plos One, 13.
- Cerioli MF, Moliva MV, Cariddi LN and Reinoso EB, 2018. Effect of the essential oil of *Minthostachys verticillata* (Griseb.) epling and limonene on biofilm production in pathogens causing Bovine Mastitis. Frontiers in Veterinary Science, 5.
- Chen B-C, Lin C-X, Chen N-P, Gao C-X, Zhao Y-J and Qian C-D, 2018. Phenanthrene antibiotic targets bacterial membranes and kills *Staphylococcus* aureus with a low propensity for resistance development. Frontiers in Microbiology, 9.
- Chen Q, He W, Yan X, Zhang T, Jiang B, Stressler T, Fischer L and Mu W, 2018. Construction of an enzymatic route using a food-grade recombinant *Bacillus subtilis* for the production and purification of epilactose from lactose. Journal of Dairy Science, 101, 1872–1882.
- Cheng Y-H, Zhang N, Han J-C, Chang C-W, Hsiao FS-H and Yu Y-H, 2018. Optimization of surfactin production from *Bacillus subtilis* in fermentation and its effects on *Clostridium perfringens*-induced necrotic enteritis and growth performance in broilers. Journal of Animal Physiology and Animal Nutrition, 102, 1232–1244.
- Cho BC, Hardies SC, Jang GI and Hwang CY, 2018. Complete genome of streamlined marine actinobacterium *Pontimonas salivibrio* strain CL-TW6(T) adapted to coastal planktonic lifestyle. BMC Genomics, 19.
- Chomwong S, Charoensapsri W, Amparyup P and Tassanakajon A, 2018. Two host gut-derived lactic acid bacteria activate the proPO system and increase resistance to an AHPND-causing strain of Vibrio parahaemolyticus in the shrimp *Litopenaeus vannarnei*. Developmental and Comparative Immunology, 89, 54–65.
- Costa EM, Silva S, Veiga M, Tavaria FK and Pintado MM, 2018. Chitosan's biological activity upon skinrelated microorganisms and its potential textile applications. World Journal of Microbiology & Biotechnology, 34.
- Coullon H, Rifflet A, Wheeler R, Janoir C, Boneca IG and Candela T, 2018. N-Deacetylases required for muramic–lactam production are involved in *Clostridium difficile* sporulation, germination, and heat resistance. Journal of Biological Chemistry, 293, 18040–18054.
- Crowe-McAuliffe C, Graf M, Huter P, Takada H, Abdelshahid M, Novacek J, Murina V, Atkinson GC, Hauryliuk V and Wilson DN, 2018. Structural basis for antibiotic resistance mediated by the *Bacillus subtilis* ABCF ATPase VmlR. Proceedings of the National Academy of Sciences of the United States of America, 115, 8978–8983.
- Dalvand LF, Hosseini F, Dehaghi SM and Torbati ES, 2018. Inhibitory effect of bismuth oxide nanoparticles produced by *Bacillus licheniformis* on methicillin-resistant *Staphylococcus aureus* strains (MRSA). Iranian Journal of Biotechnology, 16, 279–286.
- Dasgupta D, Jasmine J and Mukherj S, 2018. Characterization, phylogenetic distribution and evolutionary trajectories of diverse hydrocarbon degrading microorganisms isolated from refinery sludge. 3 Biotech, 8.
- Demirhan B, Guragac FT, Er-Demirhan B, Tastan H and Kupeli-Akkol E, 2018. A PRECLINICAL STUDY: EFFECT OF NEW PROBIOTIC PRODUCTS, "*BACILLUS SUBTILIS* AND *BACILLUS AMYLOLIQUEFACIENS*", ON DIARRHEA. Fresenius Environmental Bulletin, 27, 4139–4148.
- Doroteo AM, Pedroso FL, Lopez JDM and Apines-Amar MJS, 2018. Evaluation of potential probiotics isolated from saline tilapia in shrimp aquaculture. Aquaculture International, 26, 1095–1107.
- De Melo Pereira GV, Coelho BdO, Magalhaes Junior AI, Thomaz-Soccol V and Soccol CR, 2018. How to select a probiotic? A review and update of methods and criteria. Biotechnology Advances, 36, 2060–2076.
- Efimenko TA, Efremenkova OV, Demkina EV, Petrova MA, Sumarukova IG, Vasilyeva BF and El'-Registan GI, 2018. Bacteria isolated from antarctic permafrost are efficient antibiotic producers. Microbiology, 87, 692–698.
- Erler S, Lewkowski O, Poehlein A and Forsgren E, 2018. The curious case of achromobacter eurydice, a gram-variable pleomorphic bacterium associated with European foulbrood disease in honeybees. Microbial Ecology, 75, 1–6.
- Escribano-Viana R, Portu J, Garijo P, Rosa Gutierrez A, Santamaria P, Lopez-Alfaro I, Lopez R and Gonzalez-Arenzana L, 2018. Evaluating a preventive biological control agent applied on grapevines against *Botrytis cinerea* and its influence on winemaking. Journal of the Science of Food and Agriculture, 98, 4517–4526.

- Etminani F and Harighi B, 2018. Isolation and identification of endophytic bacteria with plant growth promoting activity and biocontrol potential from wild pistachio trees. Plant Pathology Journal, 34, 208–217.
- Farlow J, Bolkvadze D, Leshkasheli L, Kusradze I, Kotorashvili A, Kotaria N, Balarjishvili N, Kvachadze L, Nikolich M and Kutateladze M, 2018. Genomic characterization of three novel Basilisk-like phages infecting *Bacillus anthracis*. BMC Genomics, 19.
- Fernanda Villarreal-Delgado M, Daniel Villa-Rodriguez E, Alberto Cira-Chavez L, Isabel Estrada-Alvarado M, Isela Parra-Cota F and e los Santos-Villalobos S, 2018. The genus *Bacillus* as a biological control agent and its implications in the agricultural biosecurity. Revista Mexicana de Fitopatologia, 36, 95–130.
- Fu G, Wang L, Li L, Liu J, Liu S and Zhao X, 2018. *Bacillus licheniformis* CK1 alleviates the toxic effects of zearalenone in feed on weaned female Tibetan piglets. Journal of Animal Science, 96, 4471–4480.
- Fuchs S, Mehlan H, Bernhardt J, Hennig A, Michalik S, Surmann K, Pane-Farre J, Giese A, Weiss S, Backert L, Herbig A, Nieselt K, Hecker M, Voelker U and Maeder U, 2018. AureoWiki-The repository of the *Staphylococcus aureus* research and annotation community. International Journal of Medical Microbiology, 308, 558–568.
- Galaviz-Silva L, Mario Iracheta-Villarreal J and Judith Molina-Garza Z, 2018. *Bacillus* and *Virgibacillus* strains isolated from three Mexican coasts antagonize *Staphylococcus aureus* and *Vibrio parahaemolyticus*. Fems Microbiology Letters, 365.
- Galvan DD and Yu Q, 2018. Surface-enhanced raman scattering for rapid detection and characterization of antibiotic-resistant bacteria. Advanced Healthcare Materials, 7.
- Gangola S, Sharma A, Bhatt P, Khati P and Chaudhary P, 2018. Presence of esterase and laccase in *Bacillus subtilis* facilitates biodegradation and detoxification of cypermethrin. Scientific Reports, 8.
- Garcia-Ramon DC, Berry C, Tse C, Fernandez-Fernandez A, Osuna A and Vilchez S, 2018. The parasporal crystals of *Bacillus pumilus* strain 15.1: a potential virulence factor? Microbial Biotechnology, 11, 302–316.
- Ghosh K, Kang HS, Hyun WB and Kim K-P, 2018. High prevalence of *Bacillus subtilis*-infecting bacteriophages in soybean-based fermented foods and its detrimental effects on the process and quality of Cheonggukjang. Food Microbiology, 76, 196–203.
- Ghosh K, Senevirathne A, Kang HS, Hyun WB, Kim JE and Kim K-P, 2018. Complete nucleotide sequence analysis of a Novel *Bacillus subtilis*-infecting bacteriophage BSP10 and its effect on polygamma-glutamic acid degradation. Viruses-Basel, 10.
- Goldner NK, Bulow C, Cho K, Wallace M, Hsu F-F, Patti GJ, Burnham C-A, Schlesinger P and Dantas G, 2018. Mechanism of high-level daptomycin resistance in *Corynebacterium striatum*. Msphere, 3.
- Golkar T, Zielinski M and Berghuis AM, 2018. Look and outlook on enzyme-mediated macrolide resistance. Frontiers in Microbiology, 9.
- Gosavi SM, Kharat SS, Kumkar P and Navarange SS, 2018. Interplay between behavior, morphology and physiology supports lepidophagy in the catfish *Pachypterus khavalchor* (Siluriformes: Horabagridae). Zoology, 126, 185–191.
- Goto E, Haga Y, Kubo M, Itoh T, Kasai C, Shoji O, Yamamoto K, Matsumura C, Nakano T and Inui H, 2018. Metabolic enhancement of 2,3 ',4,4 ',5-pentachlorobiphenyl (CB118) using cytochrome P450 monooxygenase isolated from soil bacterium under the presence of perfluorocarboxylic acids (PFCAs) and the structural basis of its metabolism. Chemosphere, 210, 376–383.
- Gotzmann G, Portillo J, Wronski S, Kohl Y, Gorjup E, Schuck H, Roegner FH, Mueller M, Chaberny IF, Schoenfelder J and Wetzel C, 2018. Low-energy electron-beam treatment as alternative for on-site sterilization of highly functionalized medical products A feasibility study. Radiation Physics and Chemistry, 150, 9–19.
- Grohmann E, Keller W and Muth G, 2018. Mechanisms of conjugative transfer and type IV secretionmediated effector transport in gram-positive bacteria. Type Iv Secretion in Gram-Negative and Gram-Positive Bacteria, 413, 115–141.
- Haditomo AHC and Prayitno SB, 2018. Probiotic candidates from fish pond water in central java Indonesia. 3rd International Conference on Tropical and Coastal Region Eco Development 2017 116.
- Halami PM, 2018. Sublichenin, a new subtilin-like lantibiotics of probiotic bacterium *Bacillus licheniformis* MCC 2512T with antibacterial activity. Microbial Pathogenesis, 128, 139–146.

- Hamdache A, Ezziyyani M and Lamarti A, 2018. Effect of preventive and simultaneous inoculations of *Bacillus amyloliquefaciens* (Fukumoto) strains on conidial germination of *Botrytis cinerea* Pers.:Fr. Anales de Biologia, 40, 65–72.
- Hamzah TNT, Lee SY, Hidayat A, Terhem R, Faridah-Hanum I and Mohamed R, 2018. Diversity and characterization of endophytic fungi isolated from the tropical mangrove species, *Rhizophora mucronata*, and identification of potential antagonists against the soil-borne fungus, *Fusarium solani*. Frontiers in Microbiology, 9.
- Hao X, Zhang X, Duan B, Huo S, Lin W, Xia X and Liu K, 2018. Screening and genome sequencing of deltamethrin-degrading bacterium ZJ6. Current Microbiology, 75, 1468–1476.
- Hasan SMZ, Hossain F, Zaoti ZF, Hasan F, Islam A and Sikdar B, 2018. PCR amplification of DNA sequence related to the hrpD gene of *Xanthomonas cucurbitae* in leaf spot disease of pumpkin and their antagonism by soil bacteria. Archives of Phytopathology and Plant Protection, 51, 252–266.
- He S, Feng K, Ding T, Huang K, Yan H, Liu X and Zhang Z, 2018. Complete genome sequence *Bacillus licheniformis* BL-010. Microbial Pathogenesis, 118, 199–201.
- Hill AJ, Leys JE, Bryan D, Erdman FM, Malone KS, Russell GN, Applegate RD, Fenton H, Niedringhaus K, Miller AN, Allender MC and Walker DM, 2018. Common cutaneous bacteria isolated from snakes inhibit growth of *Ophidiomyces ophiodiicola*. Ecohealth, 15, 109–120.
- Ho K, Huo W, Pas S, Dao R and Palmer KL, 2018. Loss-of-function mutations in epaR confer resistance to phi NPV1 infection in *Enterococcus faecalis* OG1RF. Antimicrobial Agents and Chemotherapy, 62.
- Hoelzer K, Bielke L, Blake DP, Cox E, Cutting SM, Devriendt B, Erlacher-Vindel E, Goossens E, Karaca K, Lemiere S, Metzner M, Raicek M, Surinach MC, Wong NM, Gay C and Van Immerseel F, 2018. Vaccines as alternatives to antibiotics for food producing animals. Part 2: new approaches and potential solutions. Veterinary Research, 49.
- Hosseini S, Curilovs A and Cutting SM, 2018. Biological containment of genetically modified *Bacillus subtilis*. Applied and Environmental Microbiology, 84.
- Huang Y, Xiao X, Huang H, Jing J, Zhao H, Wang L and Long X-E, 2018. Contrasting beneficial and pathogenic microbial communities across consecutive cropping fields of greenhouse strawberry. Applied Microbiology and Biotechnology, 102, 5717–5729.
- Imarhiagbe EE and Ikhajiagbe B, 2018. Antibiotic susceptibility of bacterial isolates and water quality index of water sourced from closed ground water and open hand dug well in Koko Community, Delta State, Nigeria. Studia Universitatis Babes-Bolyai Biologia, 63, 47–57.
- Iseppi R, e Niederhausern S, Bondi M, Messi P and Sabia C, 2018. Extended-Spectrum -Lactamase, AmpC, and MBL-Producing Gram-Negative Bacteria on Fresh Vegetables and Ready-to-Eat Salads Sold in Local Markets. Microbial Drug Resistance, 24, 1156–1164.
- Ivanko OG and Solianyk OV, 2018. Antibiotic-associated disorders of prothrombin synthesis and their probiotic correction with *B. clausii* in breastfeed infants. Zaporozhye Medical Journal, 384–387.
- Javed MT, Akram MS, Abid A, Shahid M, Ali Q and Iqbal N, 2018. *Bacillus pumilus* enhances the safflower (*Carthamus tinctorius* L.) growth under chromium stress by an antioxidative potential and nutrient acquisition. Phytopathology, 108, 43–43.
- Jebakumar RM and Selvarajan R, 2018. Biopriming of micropropagated banana plants at pre- or post-BBTV inoculation stage with rhizosphere and endophytic bacteria determines their ability to induce systemic resistance against BBTV in cultivar Grand Naine. Biocontrol Science and Technology, 28, 1074–1090.
- Jeong D-E, So Y, Lim H, Park S-H and Choi S-K, 2018. Scarless genomic point mutation to construct a *Bacillus subtilis* strain displaying increased antibiotic plipastatin production. Journal of Microbiology and Biotechnology, 28, 1030–1036.
- Jeong J-J, Moon H-J, Pathiraja D, Park B, Choi I-G and Kim KD, 2018. Draft genome sequences of *Bacillus megaterium* KU143, *Microbacterium testaceum* KU313, and *Pseudomonas protegens* AS15, Isolated from Stored Rice Grains. Microbiology Resource Announcements 6.
- Ji H, Wu L, Pu F, Ren J and Qu X, 2018. Point-of-care identification of bacteria using proteinencapsulated gold nanoclusters. Advanced Healthcare Materials, 7.
- Jiang Y, Zhang Z, Wang Y, Jing Y, Liao M, Rong X, Li B, Chen G and Zhang H, 2018. Effects of probiotic on microfloral structure of live feed used in larval breeding of turbot *Scophthalmus maximus*. Journal of Oceanology and Limnology, 36, 1002–1012.
- Jimenez-Tototzintle M, Ferreira IJ, Duque SdS, Guimaraes Barrocas PR and Saggioro EM, 2018. Removal of contaminants of emerging concern (CECs) and antibiotic resistant bacteria in urban wastewater using UVA/TiO2/H2O2 photocatalysis. Chemosphere, 210, 449–457.

- Jujjavarapu SE and Dhagat S, 2018. In silico discovery of novel ligands for antimicrobial lipopeptides for computer-aided drug design. Probiotics and Antimicrobial Proteins, 10, 129–141.
- Jujjavarapu SE, Dhagat S and Kurrey V, 2018. Identification of novel ligands for therapeutic lipopeptides: daptomycin, surfactin and polymyxin. Current Drug Targets, 19, 1589–1598.
- Ka-ot AL, Banerjee S, Haldar G and Joshi SR, 2018. Acid and heavy metal tolerant *Bacillus* sp from rathole coal mines of Meghalaya, India. Proceedings of the Indian National Science Academy Part B Biological Sciences, 88, 1187–1198.

Kapoor Y, Sharma R and Kumar A, 2018. Repurposing of existing drugs for the bacterial infections: An In silico and In vitro study. Infectious disorders drug targets.

Karunakaran T, Ismail IS, Ee GCL, Nor SMM, Palachandran K and Santhanam RK, 2018. Nitric oxide inhibitory and anti-*Bacillus* activity of phenolic compounds and plant extracts from *Mesua* species. Revista Brasileira De Farmacognosia-Brazilian Journal of Pharmacognosy, 28, 231–234.

Kavitha M, Raja M and Perumal P, 2018. Evaluation of probiotic potential of *Bacillus* spp. isolated from the digestive tract of freshwater fish Labeo calbasu (Hamilton, 1822). Aquaculture Reports, 11, 59–69.

Khadieva GF, Lutfullin MT, Mochalova NK, Lenina OA, Sharipova MR and Mardanova AM, 2018. New *Bacillus subtilis* strains as promising probiotics. Microbiology, 87, 463–471.

- Khan T, Abbasi BH, Iqrar I, Khan MA and Shinwari ZK, 2018. Molecular identification and control of endophytic contamination during in vitro plantlet development of *Fagonia indica*. Acta Physiologiae Plantarum, 40.
- Knight CA, Bowman MJ, Frederick L, Day A, Lee C and Dunlap CA, 2018. The first report of antifungal lipopeptide production by a *Bacillus subtilis* for subsp inaquosorum strain. Microbiological Research, 216, 40–46.
- Koelbl T, Bernhard S, Egger B, Meuli-Simmen C and Ly T, 2018. Fulminant wound infection with *Clostridium perfringens* and *Bacillus cereus* in a healthy five year old boy. Journal of Pediatric Surgery Case Reports, 38, 64–65.
- Kulengowski B, Rutter WC, Campion JJ, Lee GC, Feola DJ and Burgess DS, 2018. Effect of increasing meropenem MIC on the killing activity of meropenem in combination with amikacin or polymyxin B against MBL- and KPC-producing *Enterobacter cloacae*. Diagnostic Microbiology and Infectious Disease, 92, 262–266.
- Kuno Y, Sriyam S, Nakagawa R and Kimura K, 2018. A Survey of phage contamination in nattoproducing factories and development of phage-resistant *Bacillus subtilis* (natto) strains. Food Science and Technology Research, 24, 485–492.
- Kvan OV, Gavrish IA, Lebedev SV, Korotkova AM, Miroshnikova EP, Serdaeva VA, Bykov AV and Davydova NO, 2018. Effect of probiotics on the basis of *Bacillus subtilis* and *Bifidobacterium longum* on the biochemical parameters of the animal organism. Environmental Science and Pollution Research, 25, 2175–2183.
- Lam AK, Hill MA, Moen EL, Pusavat J, Wouters CL and Rice CV, 2018. Cationic Branched Polyethylenimine (BPEI) disables antibiotic resistance in methicillin-resistant *Staphylococcus epidermidis* (MRSE). Chemmedchem, 13, 2240–2248.
- Lang J-F, Tian X-L, Shi M-W and Ran L-X, 2018. Identification of endophytes with biocontrol potential from *Ziziphus jujuba* and its inhibition effects on *Alternaria alternata*, the pathogen of jujube shrunken-fruit disease. Plos One, 13.
- Lapointe JF, McCarthy CD, Dunphy GB and Mandato CA, 2018. Physiological evidence of integrinantibody reactive proteins influencing the innate cellular immune responses of larval *Galleria mellonella* hemocytes. Insect Science.
- Laue M, Han H-M, Dittmann C and Setlow P, 2018. Intracellular membranes of bacterial endospores are reservoirs for spore core membrane expansion during spore germination. Scientific Reports, 8.
- Ledwoch K, Dancer SJ, Otter JA, Kerr K, Roposte D, Rushton L, Weiser R, Mahenthiralingam E, Muir DD and Maillard JY, 2018. Beware biofilm! Dry biofilms containing bacterial pathogens on multiple healthcare surfaces; a multi-centre study. Journal of Hospital Infection, 100, E47–E56.
- Li A, Jiang X, Wang Y, Zhang L, Zhang H, Mehmood K, Li Z, Waqas M and Li J, 2018. The impact of *Bacillus subtilis* 18 isolated from Tibetan yaks on growth performance and gut microbial community in mice. Microbial Pathogenesis, 128, 153-161.
- Li B, Li H, Liu N, Liu Y, Liu L, Liu J, Zhang X and Xing X, 2018. Analysis of pathogenic bacteria of the infected chicks. Chinese Veterinary Science/Zhongguo Shouyi Kexue, 48, 971–978.
- Li L, Ge H, Gu D, Meng H, Li Y, Jia M, Zheng C and Zhou X, 2018. The role of two-component regulatory system in beta-lactam antibiotics resistance. Microbiological Research, 215, 126–129.

- Li X, Wu S, Li X, Yan T, Duan Y, Yang X, Duan Y, Sun Q and Yang X, 2018. Simultaneous Supplementation of *Bacillus subtilis* and antibiotic growth promoters by stages improved intestinal function of pullets by altering gut microbiota. Frontiers in Microbiology, 9.
- Lin L, Liang H, Zhuang H, Lian K, Zhong Q, Cai Y, Luo C and Tang J, 2018. Effect of Zaozhu Yinchen Recipe on intestinal flora in treatment of nonalcoholic steatohepatitis. Chinese Journal of Integrated Traditional and Western Medicine, 38, 673–676.
- Liu R, Wang W, Liu X, Lu Y, Xiang T, Zhou W and Wan Y, 2018. Characterization of a lipase from the silkworm intestinal bacterium *Bacillus pumilus* with antiviral activity against *Bombyx mori* (Lepidoptera: Bombycidae) nucleopolyhedrovirus in vitro. Journal of Insect Science, 18.
- Livingstone PG, Morphew RM, Cookson AR and Whitworth DE, 2018. Genome analysis, metabolic potential, and predatory capabilities of *Herpetosiphon ilansteffanense* sp nov. Applied and Environmental Microbiology, 84.
- Lopes R, Tsui S, Goncalves PJRO and e Queiroz MV, 2018. A look into a multifunctional toolbox: endophytic *Bacillus* species provide broad and underexploited benefits for plants. World Journal of Microbiology & Biotechnology, 34.
- Mahmmod YS, Nonnemann B, Svennesen L, Pedersen K and Klaas IC, 2018. Typeability of MALDI-TOF assay for identification of non-aureus staphylococci associated with bovine intramammary infections and teat apex colonization. Journal of Dairy Science, 101, 9430–9438.
- Marag PS and Suman A, 2018. Growth stage and tissue specific colonization of endophytic bacteria having plant growth promoting traits in hybrid and composite maize (*Zea mays* L.). Microbiological Research, 214, 101–113.
- Martins KS, e Assis Magalhaes LT, e Almeida JG and Pieri FA, 2018. Antagonism of bacteria from dog dental plaque against human cariogenic bacteria. Biomed Research International.
- Matteoli FP, Passarelli-Araujo H, Reis RJA, Rocha LO, e Souza EM, Aravind L, Olivares FL and Venancio TM, 2018. Genome sequencing and assessment of plant growth-promoting properties of a *Serratia marcescens* strain isolated from vermicompost. BMC Genomics, 19.
- Mazhar S, Hill C and McAuliffe O, 2018. The genus macrococcus: an insight into its biology, evolution, and relationship with *Staphylococcus*. Advances in Applied Microbiology, 105, 1–50.
- McClure J-AM, Lakhundi S, Kashif A, Conly JM and Zhang K, 2018. Genomic comparison of highly virulent, moderately virulent, and avirulent strains from a genetically closely-related MRSA ST239 sub-lineage provides insights into pathogenesis. Frontiers in Microbiology, 9.
- McGovern E, Waters SM, Blackshields G and McCabe MS, 2018. Evaluating established methods for rumen 16S rRNA amplicon sequencing with mock microbial populations. Frontiers in Microbiology, 9.
- McHugh AJ, Feehily C, Tobin JT, Fenelon MA, Hill C and Cotter PD, 2018. Mesophilic sporeformers identified in whey powder by using shotgun metagenomic sequencing. Applied and Environmental Microbiology, 84.
- Menz J, Mueller J, Olsson O and Kuemmerer K, 2018. Bioavailability of antibiotics at soil-water interfaces: a comparison of measured activities and equilibrium partitioning estimates. Environmental Science & Technology, 52, 6555–6564.
- Midhun SJ, Neethu S, Vysakh A, Radhakrishnan EK and Jyothis M, 2018. Antagonism against fish pathogens by cellular components/preparations of *Bacillus coagulans* (MTCC-9872) and it's in vitro probiotic characterisation. Current Microbiology, 75, 1174–1181.
- Milivojevic D, Sumonja N, Medic S, Pavic A, Moric I, Vasiljevic B, Senerovic L and Nikodinovic-Runic J, 2018. Biofilm-forming ability and infection potential of *Pseudomonas aeruginosa* strains isolated from animals and humans. Pathogens and Disease, 76.
- Moharam MHA, Stephan D and Koch E, 2018. Evaluation of plant-derived preparations and microorganisms as seed treatments for control of covered kernel smut of sorghum (*Sporisorium sorghi*). Journal of Plant Diseases and Protection, 125, 159–166.
- Mohr T, Aliyu H, Kuechlin R, Zwick M, Cowan D, Neumann A and e Maayer P, 2018. Comparative genomic analysis of *Parageobacillus thermoglucosidasius* strains with distinct hydrogenogenic capacities. BMC Genomics, 19.
- Molloy-Simard V, Lemyre J-L, Martel K and Catalone BJ, 2018. Elevating the standard of endoscope processing: Terminal sterilization of duodenoscopes using a hydrogen peroxide-ozone sterilizer. American Journal of Infection Control.
- Moore JE, McCaughan J, Stirling J, Bell J and Millar BC, 2018. Snow angels the microbiology of freshly fallen snow: implications for immunocompromised patients. Journal of Water and Health, 16, 1029–1032.

- Moradi M, Nejad FJ, Bonjar GHS, Fani SR, Mimand BM, Probst C and Madani M, 2018. Efficacy of *Bacillus subtilis* native strains for biocontrol of *Phytophthora* crown and root rot of pistachio in Iran. Tropical Plant Pathology, 43, 306–313.
- Mukhtar T, Afridi MS, McArthur R, Van Hamme JD, Rineau F, Mahmood T, Zahid M, Salam A, Khan MN, Ali F, Mehmood S, Bangash N and Chaudhary HJ, 2018. Draft genome sequence of *Bacillus pumilus* SCAL1, an endophytic heat-tolerant plant growth-promoting bacterium. Microbiology Resource Announcements, 6.
- Mulyani Y., Aryantha INP, Suhandono S and Pancoro A, 2018. Intestinal Bacteria of Common Carp (Cyprinus carpio L.) as a Biological Control Agent for *Aeromonas*. Journal of Pure and Applied Microbiology, 12, 601–610.
- Munoz-Moreno CY, De La Cruz-Rodriguez Y, Vega-Arreguin J, Alvarado-Rodriguez M, Manuel Gomez-Soto J, Alvarado-Gutierrez A and Fraire-Velazquez S, 2018. Draft genome sequence of *Bacillus subtilis* 2C-9B, a strain with biocontrol potential against chili pepper root pathogens and tolerance to Pb and Zn. Microbiology Resource Announcements, 6.
- Murina V, Kasari M, Takada H, Hinnu M, Saha CK, Grimshaw JW, Seki T, Reith M, Putrins M, Tenson T, Strahl H, Hauryliuk V and Atkinson GC, 2018. ABCF ATPases involved in protein synthesis, ribosome assembly and antibiotic resistance: structural and functional diversification across the tree of life. Journal of Molecular Biology.
- Nandi A, Banerjee G, Dan SK, Ghosh K and Ray AK, 2018. Evaluation of in vivo probiotic efficiency of *Bacillus amyloliquefaciens* in labeo rohita challenged by pathogenic strain of *Aeromonas hydrophila* MTCC 1739. Probiotics and Antimicrobial Proteins, 10, 391–398.
- Nannan C, Gillis A, Caulier S and Mahillon J, 2018. Complete genome sequence of *Bacillus velezensis* CN026 exhibiting antagonistic activity against gram-negative foodborne pathogens. Microbiology Resource Announcements, 6.
- Ngo DQ, Dao HH, Hanaoka S, Hasegawa Y and Le TT, 2018. Methods for treatment of skin infectious diseases using microorganisms.
- Nigris S, Baldan E, Tondello A, Zanella F, Vitulo N, Favaro G, Guidolin V, Bordin N, Telatin A, Barizza E, Marcato S, Zottini M, Squartini A, Valle G and Baldan B, 2018. Biocontrol traits of Bacillus licheniformis GL174, a culturable endophyte of *Vitis vinifera* cv. Glera. BMC Microbiology, 18.
- Noormohammadi H, Abolmaali S and Astaneh SDA, 2018. Identification and characterization of an endolysin Like from *Bacillus subtilis*. Microbial Pathogenesis, 119, 221–224.
- Obszanska K, Kern-Zdanowicz I and Sitkiewicz I, 2018. Efficient construction of *Streptococcus anginosus* mutants in strains of clinical origin. Journal of Applied Genetics, 59, 515–523.
- Osman KM, Kappell AD, Orabi A, Al-Maary KS, Mubarak AS, Dawoud TM, Hemeg HA, Moussa IMI, Hessain AM, Yousef HMY and Hristova KR, 2018. Poultry and beef meat as potential seedbeds for antimicrobial resistant enterotoxigenic *Bacillus* species: a materializing epidemiological and potential severe health hazard. Scientific Reports, 8.
- Oztopuz O, Pekin G, Park RD and Eltem R, 2018. Isolation and evaluation of new antagonist *Bacillus* Strains for the control of pathogenic and mycotoxigenic fungi of fig orchards. Applied Biochemistry and Biotechnology, 186, 692–711.
- Pan Z, Qi G, Andriamanohiarisoamanana FJ, Yamashiro T, Iwasaki M, Nishida T, Tangtaweewipat S and Umetsu K, 2018. Potential of anaerobic digestate of dairy manure in suppressing soil-borne plant disease. Animal Science Journal, 89, 1512–1518.
- Panda SK, Padhi L and Sahoo G, 2018. Oral bacterial flora of Indian cobra (*Naja naja*) and their antibiotic susceptibilities. Heliyon, 4.
- Passari AK, Leo VV, Chandra P, Kumar B, Nayak C, Hashem A, Abd Allah EF, Alqarawi AA and Singh BP, 2018. Bioprospection of actinobacteria derived from freshwater sediments for their potential to produce antimicrobial compounds. Microbial Cell Factories, 17.
- Passari AK, Yadav MK and Singh BP, 2018. In vitro evaluation of antimicrobial activities and antibiotic susceptibility profiling of culturable actinobacteria from fresh water streams. Indian Journal of Experimental Biology, 56, 665–673.
- Pawar RR and Borkar SG, 2018. Isolation and characterization of thermophilic bacteria from different habitats and their assessment for antagonism against soil-borne fungal plant pathogens. African Journal of Microbiology Research, 12, 556–566.
- Pisheh SM and Madani M, 2018. Evaluation of effect the cyclic lipopeptides from *Bacillus atrophaeus* HNSQJYH170 on *Candida* species. Qom University of Medical Sciences Journal, 12, Pe20-En19.

- Poulsen A-SR, e Jonge N, Nielsen JL, Hojberg O, Lauridsen C, Cutting SM and Canibe N, 2018. Impact of *Bacillus* spp. spores and gentamicin on the gastrointestinal microbiota of suckling and newly weaned piglets. Plos One, 13.
- Reda RM, El-Hady MA, Selim KM and El-Sayed HM, 2018. Comparative study of three predominant gut *Bacillus* strains and a commercial *B. amyloliquefaciens* as probiotics on the performance of *Clarias gariepinus*. Fish & Shellfish Immunology, 80, 416–425.
- Reda RM, Selim KM, El-Sayed HM and El-Hady MA, 2018. In vitro selection and identification of potential probiotics isolated from the gastrointestinal tract of Nile Tilapia, *Oreochromis niloticus*. Probiotics and Antimicrobial Proteins, 10, 692–703.
- Sabate DC, Perez Brandan C, Petroselli G, Erra-Balsells R and Carina Audisio M, 2018. Biocontrol of *Sclerotinia sclerotiorum* (Lib.) de Bary on common bean by native lipopeptide-producer *Bacillus* strains. Microbiological Research, 211, 21–30.
- Sadek ZI, Abdel-Rahman MA, Azab MS, Darwesh OM and Hassan MS, 2018. Microbiological evaluation of infant foods quality and molecular detection of *Bacillus cereus* toxins relating genes. Toxicology Reports, 5, 871–877.
- Saini S, Battan B, Maan S and Sharma J, 2018. Decolourization of dyes by *Alcaligenes faecalis* and *Bacillus flexus* isolated from textile effluent. Indian Journal of Experimental Biology, 56, 820–826.
- Sakthi Thesai A, Rajakumar S and Ayyasamy PM, 2018. Removal of fluoride in aqueous medium under the optimum conditions through intracellular accumulation in *Bacillus flexus* (PN4). Environmental Technology, 1–14.
- Sanchez-Bautista A, De Leon-Garcia de Alba C, Aranda-Ocampo S, Zavaleta-Mejia E, Nava-Diaz C, Goodwin PH and Leyva-Mir SG, 2018. Root endophyte bacteria in drought-tolerant and drought-susceptible maize lines. Revista Mexicana de Fitopatologia, 36, 35–55.
- Santana JO, Gramacho KP, e Souza Eduvirgens Ferreira KT, Rezende RP, Oliveira Mangabeira PA, Moreira Dias RP, Couto FM and Pirovani CP, 2018. Witches' broom resistant genotype CCN51 shows greater diversity of symbiont bacteria in its phylloplane than susceptible genotype catongo. BMC Microbiology, 18.
- Santesmases MJ and Santesmases MJ, 2018. Circulation of Penicillin in Spain: Health, Wealth and Authority. Circulation of Penicillin in Spain: Health, Wealth and Authority.
- Sarwar A, Brader G, Corretto E, Aleti G, Abaidullah M, Sessitsch A and Hafeez FY, 2018. Qualitative analysis of biosurfactants from *Bacillus* species exhibiting antifungal activity. Plos One, 13.
- Schlievert PM, Kilgore SH, Kaus GM, Ho TD and Ellermeier CD, 2018. Glycerol Monolaurate (GML) and a Nonaqueous Five-Percent GML Gel Kill *Bacillus* and *Clostridium* Spores. Msphere, 3.
- Selvakumar S, Chandra AP and Balakrishnan G, 2018. Anti-bacterial effects of citrus medica on some clinically important human pathogens. Research Journal of Pharmaceutical Biological and Chemical Sciences, 9, 1442–1448.
- Sharaf A, Obornik M, Hammad A, El-Afifi S and Marei E, 2018. Characterization and comparative genomic analysis of virulent and temperate *Bacillus megaterium* bacteriophages. Peerj, 6.
- Shukla S, Park JH, Chung SH and Kim M, 2018. Ochratoxin A reduction ability of biocontrol agent Bacillus subtilis isolated from Korean traditional fermented food Kimchi. Scientific Reports, 8.
- Silva TR, Duarte AWF, Passarini MRZ, Ruiz ALTG, Franco CH, Moraes CB, e Melo IS, Rodrigues RA, Fantinatti-Garboggini F and Oliveira VM, 2018. Bacteria from Antarctic environments: diversity and detection of antimicrobial, antiproliferative, and antiparasitic activities. Polar Biology, 41, 1505–1519.

Smith BR and Unckless RL, 2018. Draft genome sequence of lysinibacillus fusiformis strain juneja, a laboratory-derived pathogen of *Drosophila melanogaster*. Microbiology Resource Announcements, 6.

- Sorokan AV, Benkovskaya GV, Blagova DK, Maksimova TI and Maksimov IV, 2018. Defense responses and changes in symbiotic gut microflora in the colorado potato beetle *Leptinotarsa decemlineata* under the effect of endophytic bacteria from the genus *Bacillus*. Journal of Evolutionary Biochemistry and Physiology, 54, 300–307.
- Stanley CN and Arueyingho VO, 2018. Microbiological evaluation and antibiotic susceptibility patterns of organisms isolated from frozen chicken products sold in Port Harcourt, Nigeria. Journal of Pharmaceutical Research International, 23.
- Sudha MR, Jayanthi N, Aasin M, Dhanashri RD and Anirudh T, 2018. Efficacy of *Bacillus coagulans* Unique IS2 in treatment of irritable bowel syndrome in children: a double blind, randomised placebo controlled study. Beneficial Microbes, 9, 563–572.

- Sun Q, Jiang L, Guo H, Xia F, Wang B, Wang Y, Xia Q and Zhao P, 2018. Increased antiviral capacity of transgenic silkworm via knockdown of multiple genes on *Bombyx mori* bidensovirus. Developmental and Comparative Immunology, 87, 188–192.
- Sun Q, Li W, Xu B, Wei F, Wang L, Lin P and Li S, 2018. Effects of curcumin and Bacillus licheniformis on growth performance, serum antioxidant function, intestinal microbe counts and immune organ indexes of broilers. Chinese Journal of Animal Nutrition, 30, 3176–3183.
- Sun W, Shi C, Zhang M, Zhao D, Chen X, Feng L, Luo Y, Guo W and Xing D, 2018. Isolation and identification of *Bacillus* spp. controlling *Neocosmospora* pod rot of peanut. Weishengwu Xuebao, 58, 1573–1581.
- Susilowati A, Puspita AA and Yunus A, 2018. Drought resistant of bacteria producing exopolysaccharide and IAA in rhizosphere of soybean plant (Glycine max) in Wonogiri Regency Central Java Indonesia. 4th International Conference on Sustainable Agriculture and Environment 142.
- Suyamud B, Inthorn D, Panyapinyopol B and Thiravetyan P, 2018. Biodegradation of bisphenol A by a newly isolated *Bacillus megaterium* Strain ISO-2 from a polycarbonate industrial wastewater. Water Air and Soil Pollution, 229.
- Swarge BN, Roseboom W, Zheng L, Abhyankar WR, Brul S, e Koster CG and e Koning LJ, 2018. "One-Pot" sample processing method for proteome-wide analysis of microbial cells and spores. Proteomics Clinical Applications, 12.
- Syromyatnikov MY, Kokina AV, Savinkova OV, Panevina AV, Solodskikh SA, Orlova MV, Grabovich MY, Starkov AA and Popov VN, 2018. Study of the microbiological composition of dairy products and mayonnaise using DNA barcoding and metabarcoding. Foods and Raw Materials, 6, 144–153.
- Takimoto T, Hatanaka M, Hoshino T, Takara T, Tanaka K, Shimizu A, Morita H and Nakamura T, 2018. Effect of *Bacillus subtilis* C-3102 on bone mineral density in healthy postmenopausal Japanese women: a randomized, placebo-controlled, double-blind clinical trial. Bioscience of Microbiota Food and Health, 37, 87–96.
- Terra L, Dyson PJ, Hitchings MD, Thomas L, Abdelhameed A, Banat IM, Gazze SA, Vujaklija D, Facey PD, Francis LW and Quinn GA, 2018. A Novel Alkaliphilic *Streptomyces* Inhibits ESKAPE Pathogens. Frontiers in Microbiology, 9.
- Thamthaweechok N, Tiengrim S and Thamlikitkul V, 2018. Heat stability of antibiotics commonly used in food animals and agriculture in Thailand. Journal of the Medical Association of Thailand, 101, 863–867.
- Townsend JR, Bender D, Vantrease WC, Sapp PA, Toy AM, Woods CA and Johnson KD, 2018. Effects of probiotic (*Bacillus subtilis* DE111) supplementation on immune function, hormonal status, and physical performance in division I baseball players. Sports, 6.
- Tozlu E, Tekiner N, Kotan R and Ortucu S, 2018. Investigation on the biological control of Alternaria alternata. Indian Journal of Agricultural Sciences, 88, 1241–1247.
- Tsonis I, Karamani L, Xaplanteri P, Kolonitsiou F, Zampakis P, Gatzounis G, Marangos M and Assimakopoulos SF, 2018. Spontaneous cerebral abscess due to *Bacillus subtilis* in an immunocompetent male patient: A case report and review of literature. World Journal of Clinical Cases, 6, 1169–1174.
- Turck D, Bresson JL, Burlingame B, Dean T, Fairweather-Tait S, Heinonen M, Hirsch-Ernst KI, Mangelsdorf I, McArdle H, Naska A, Neuhauser-Berthold M, Nowicka G, Pentieva K, Sanz Y, Siani A, Sjodin A, Stern M, Tome D, Vinceti M, Willatts P, Engel KH, Marchelli R, Poting A, Poulsen M, Schlatter JR, Germini A and N. EFSA Panel on Dietetic Products, 2018. Safety of p-ribose as a novel food pursuant to Regulation (EU) 2015/2283. EFSA Journal, 16, e05265–e05265.
- Val-Calvo J, Luque-Ortega JR, Crespo I, Miguel-Arribas A, Abia D, Sanchez-Hevia DL, Serrano E, Gago-Cordoba C, Ares S, Alfonso C, Rojo F, Wu LJ, Boer DR and Meijer WJJ, 2018. Novel regulatory mechanism of establishment genes of conjugative plasmids. Nucleic Acids Research, 46, 11910–11926.
- Vidal S, Brandt BW, Dettwiler M, Abril C, Bressan J, Greub G, Frey CF, Perreten V and Rodriguez-Campos S, 2018. Limited added value of fungal ITS amplicon sequencing in the study of bovine abortion. Heliyon, 4.
- Vignesh V, Sathiyanarayanan G, Parthiban K, Kumar KS and Thirumurugan R, 2018. Functional assessment of subtilosin A against *Aeromonas* spp. causing gastroenteritis and hemorrhagic septicaemia. Indian Journal of Biotechnology, 17, 27–32.
- Viichez JI, Tang Q, Kaushal R, Wang W, Lv S, He D, Chu Z, Zhang H, Liu R and Zhang H, 2018. Complete Genome Sequence of *Bacillus megaterium* Strain TG1-E1, a Plant Drought Tolerance-Enhancing Bacterium. Microbiology Resource Announcements, 7.

- Vilchez JI, Tang Q, Kaushal R, Wang W, Lv S, He D, Chu Z, Zhang H, Liu R and Zhang H, 2018. Genome Sequence of *Bacillus megaterium* Strain YC4-R4, a plant growth-promoting rhizobacterium isolated from a high-salinity environment. Microbiology Resource Announcements, 6.
- Villegas-Escobar V, Maria Gonzalez-Jaramillo L, Ramirez M, Natalia Moncada R, Sierra-Zapata L, Orduz S and Romero-Tabarez M, 2018. Lipopeptides from *Bacillus* sp EA-CB0959: Active metabolites responsible for in vitro and in vivo control of Ralstonia solanacearum. Biological Control, 125, 20–28.
- Villeneuve C-A, Marchand G, Gardette M, Lavoie J, Neesham-Grenon E, Begin D and Debia M, 2018. Assessment of workers' exposure to microorganisms when using biological degreasing stations. Food and Chemical Toxicology, 116, 53–59.
- Vinodkumar S, Nakkeeran S, Renukadevi P and Mohankumar S, 2018. Diversity and antiviral potential of rhizospheric and endophytic *Bacillus* species and phyto-antiviral principles against tobacco streak virus in cotton. Agriculture Ecosystems & Environment, 267, 42–51.
- Vorobets NM, Kryvtsova MV, Rivis OY, Spivak MY, Yavorska HV and Semenova HM, 2018. Antimicrobial activity of phytoextracts on opportunistic oral bacteria, yeast and bacteria from probiotics. Regulatory Mechanisms in Biosystems, 9, 374–378.
- Wang J, Song L, Jiao Q, Yang S, Gao R, Lu X and Zhou G, 2018. Comparative genome analysis of jujube witches'-broom Phytoplasma, an obligate pathogen that causes jujube witches'-broom disease. BMC Genomics, 19.
- Wang X, Huang N, Shao J, Hu M, Zhao Y and Huo M, 2018. Coupling heavy metal resistance and oxygen flexibility for bioremoval of copper ions by newly isolated *Citrobacter freundii* JPG1. Journal of Environmental Management, 226, 194–200.
- Wang X, Kiess AS, Peebles ED, Wamsley KGS and Zhai W, 2018. Effects of Bacillus subtilis and zinc on the growth performance, internal organ development, and intestinal morphology of male broilers with or without subclinical coccidia challenge. Poultry Science, 97, 3947–3956.
- Wang Y, Zhang H, Yan H, Yin C, Liu Y, Xu Q, Liu X and Zhang Z, 2018. Effective Biodegradation of Aflatoxin B1 Using the *Bacillus licheniformis* (BL010) Strain. Toxins, 10.
- Wang Z, Li P, Luo L, Simpson DJ and Ganzle MG, 2018. Daqu fermentation selects for heat-resistant enterobacteriaceae and bacilli. Applied and Environmental Microbiology, 84.
- Wolf IR, Paschoal AR, Quiroga C, Domingues DS, e Souza RF, Pretto-Giordano LG and Vilas-Boas LA, 2018. Functional annotation and distribution overview of RNA families in 27 *Streptococcus agalactiae* genomes. BMC Genomics, 19.
- Wong SF, Lim PKC, Mak JW, Ooi SS and Chen DKF, 2018. Molecular characterization of culturable bacteria in raw and commercial edible bird nests (EBNs). International Food Research Journal, 25, 966–974.
- Xu B-H, Lu Y-Q, Ye Z-W, Zheng Q-W, Wei T, Lin J-F and Guo L-Q, 2018. Genomics-guided discovery and structure identification of cyclic lipopeptides from the *Bacillus siamensis* JFL15. Plos One, 13.
- Xu X, Luo D, Bao Y, Liao X and Wu J, 2018. Characterization of diversity and probiotic efficiency of the autochthonous lactic acid bacteria in the fermentation of selected raw fruit and vegetable juices. Frontiers in Microbiology, 9.
- Yang BW, Yeo I-C, Choi JH, Sumi CD and Hahm YT, 2018. RNA-Seq Analysis of Antibiotic-Producing *Bacillus subtilis* SC-8 Reveals a Role for Small Peptides in Controlling PapR Signaling. Applied Biochemistry and Biotechnology, 185, 359–369.
- Yendyo S, Ramesh GC and Pandey BR, 2018. Revised evaluation of *Trichoderma* spp., *Pseudomonas fluorescens* and *Bacillus subtilis* for biological control of Ralstonia wilt of tomato version 3; referees: 2 approved. F1000Research 6(2028): (22 March 2018)-(2022 March 2018).
- Yin Y, Zhang P, Yue X, Du X, Li W, Yin Y, Yi C and Li Y, 2018. Effect of sub-chronic exposure to lead (Pb) and *Bacillus subtilis* on *Carassius auratus* gibelio: Bioaccumulation, antioxidant responses and immune responses. Ecotoxicology and Environmental Safety, 161, 755–762.
- Yu L, Muralidharan S, Lee NA, Lo R, Stokes JR, Fitzgerald MA and Turner MS, 2018. The impact of variable high pressure treatments and/or cooking of rice on bacterial populations after storage using culture-independent analysis. Food Control, 92, 232–239.
- Yu WQ, Zheng GP, Qiu DW, Yan FC, Liu WZ and Liu WX, 2018. Draft genome sequence, diseaseresistance genes, and phenotype of a *Paenibacillus terrae* strain (NK3-4) with the potential to control plant diseases. Genome, 61, 725–734.
- Zafar H and Saier MH, 2018. Comparative genomics of transport proteins in seven Bacteroides species. Plos One, 13.

- Zarza E, Alcaraz LD, Aguilar-Salinas B, Islas A and Olmedo-Alvarez G, 2018. Complete Genome Sequences of Two *Bacillus pumilus* Strains from Cuatrocienegas, Coahuila, Mexico (vol 6, e00364-18, 2018). Microbiology Resource Announcements, 7.
- Zhang Y, Zhang Y, Li P, Wang Y, Wang J, Shao Z and Zhao G, 2018. GlnR positive transcriptional regulation of the phosphate-specific transport system pstSCAB in *Amycolatopsis mediterranei* U32. Acta Biochimica Et Biophysica Sinica, 50, 757–765.
- Zhao K, Li J, Shen M, Chen Q, Liu M, Ao X, Liao D, Gu Y, Xu K, Ma M, Yu X, Xiang Q, Chen J, Zhang X and Penttinen P, 2018. Actinobacteria associated with Chinaberry tree are diverse and show antimicrobial activity. Scientific Reports, 8.
- Zheng J, Zhang S, He Y, Xue X and Guo X, 2018. Optimization of spore formation conditions of an aflatoxin-degrading *Bacillus subtilis* strain used in silage preparation. Animal Husbandry and Feed Science (Inner Mongolia), 39, 19–22.
- Zhou S-Y, Hu Y-J, Meng F-C, Qu S-Y, Wang R, Andersen RJ, Liao Z-H and Chen M, 2018. Bacillamidins A-G from a Marine-Derived *Bacillus pumilus*. Marine Drugs, 16.
- Zommiti M, Bouffartigues E, Maillot O, Barreau M, Szunerits S, Sebei K, Feuilloley M, Connil N and Ferchichi M, 2018. In vitro Assessment of the Probiotic Properties and Bacteriocinogenic Potential of *Pediococcus pentosaceus* MZF16 Isolated From Artisanal Tunisian Meat Dried Ossban. Frontiers in Microbiology, 9.

Geobacillus stearothermophilus

None

Gram-negative bacteria

Gluconobacter oxydans

None

Xanthomonas campestris

None

Yeasts

- (2018). Fungal Genomics: Methods and Protocols, Second Edition. Fungal Genomics: Methods and Protocols, Second Edition 1775.
- Abarike ED, Cai J, Lu Y, Yu H, Chen L, Jian J, Tang J, Jun L and Kuebutornye FKA, 2018. Effects of a commercial probiotic BS containing *Bacillus subtilis* and *Bacillus licheniformis* on growth, immune response and disease resistance in Nile tilapia, Oreochromis niloticus. Fish & Shellfish Immunology 82, 229–238.
- Abu-Freha N, Badarna W, Sigal-Batikoff I, Abu Tailakh M, Etzion O, Elkrinawi J, Segal A, Mushkalo A and Fich A, 2018. ASCA and ANCA among Bedouin Arabs with inflammatory bowel disease, the frequency and phenotype correlation. BMC Gastroenterology, 18.
- Acs-Szabo L, Papp LA, Antunovics Z, Sipiczki M and Miklos I, 2018. Assembly of *Schizosaccharomyces cryophilus* chromosomes and their comparative genomic analyses revealed principles of genome evolution of the haploid fission yeasts. Scientific Reports 8.
- Agamennone V, Krul CAM, Rijkers G and Kort R, 2018. A practical guide for probiotics applied to the case of antibiotic-associated diarrhea in The Netherlands. BMC Gastroenterology, 18.
- Aguiar T, Luiz C, Rocha Neto AC and Di Piero RM, 2018. Residual polysaccharides from fungi reduce the bacterial spot in tomato plants. Bragantia, 77, 299–313.
- Ahmadsah LSF, Kim E, Jung Y-S and Kim H-Y, 2018. Identification of LAB and Fungi in Laru, a Fermentation Starter, by PCR-DGGE, SDS-PAGE, and MALDI-TOF MS. Journal of Microbiology and Biotechnology, 28, 32–39.
- Alassane-Kpembi I, Pinton P, Hupe J-F, Neves M, Lippi Y, Combes S, Castex M and Oswald IP, 2018. *Saccharomyces cerevisiae* boulardii Reduces the Deoxynivalenol-Induced Alteration of the Intestinal Transcriptome. Toxins, 10(5).
- Albertin W, Chernova M, Durrens P, Guichoux E, Sherman DJ, Masneuf-Pomarede I and Marullo P, 2018. Many interspecific chromosomal introgressions are highly prevalent in Holarctic *Saccharomyces uvarum* strains found in human-related fermentations. Yeast, 35, 141–156.



- Alcazar-Fabra M, Trevisson E and Brea-Calvo G, 2018. Clinical syndromes associated with Coenzyme Q (10) deficiency. Mitochondrial Diseases, 62, 377–398.
- Alderees F, Mereddy R, Webber D, Nirmal N and Sultanbawa Y, 2018. Mechanism of Action against Food Spoilage Yeasts and Bioactivity of *Tasmannia lanceolata*, *Backhousia citriodora* and *Syzygium anisatum* Plant Solvent Extracts. Foods, 7.
- Al-Khalaifah HS, 2018. Benefits of probiotics and/or prebiotics for antibiotic-reduced poultry. Poultry Science, 97, 3807–3815.
- Alobaid K and Khan Z, 2018. Epidemiologic characteristics of adult candidemic patients in a secondary hospital in Kuwait: A retrospective study. Journal de Mycologie Medicale.
- Amer M, Nadeem M, Nazir SUR, Fakhar M, Abid F, Ain Q-U and Asif E, 2018. Probiotics and Their Use in Inflammatory Bowel Disease. Alternative Therapies In Health And Medicine, 24, 16–23.
- Amorim Ventura LL, e Oliveira DR, Gomes MA and Fantoni Torres MR, 2018. Effect of probiotics on giardiasis. Where are we? Brazilian Journal of Pharmaceutical Sciences, 54.
- Araujo R and Sampaio-Maia B, 2018. Fungal Genomes and Genotyping. Advances in Applied Microbiology, Vol 102: 37-81.
- Arendrup MC, Chowdhary A, Astvad KMT and Jorgensen KM, 2018. APX001A in vitro activity against contemporary blood isolates and *Candida auris* determined by the EUCAST reference method. Antimicrobial Agents and Chemotherapy, 62.
- Argy N, Le Gal S, Coppee R, Song Z, Vindrios W, Massias L, Kao W-C, Hunte C, Yazdanpanah Y, Lucet J-C, Houze S, Clain J and Nevez G, 2018. Pneumocystis Cytochrome b Mutants Associated With Atovaquone Prophylaxis Failure as the Cause of Pneumocystis Infection Outbreak Among Heart Transplant Recipients. Clinical Infectious Diseases, 67, 913–919.
- Awad L, Tamim H, Abdallah D, Salameh M, Mugharbil A, Jisr T, Zahran K, Droubi N, Ibrahim A and Moghnieh R, 2018. Correlation between antifungal consumption and the distribution of *Candida* species in different hospital departments of a Lebanese medical Centre. BMC Infectious Diseases, 18.
- Awais MM, Jamal MA, Akhtar M, Hameed MR, Anwar MI and Ullah MI, 2018. Immunomodulatory and ameliorative effects of *Lactobacillus* and *Saccharomyces* based probiotics on pathological effects of eimeriasis in broilers. Microbial pathogenesis, 126, 101–108.
- Baeza M, Fernandez-Lobato M, Alcaino J and Cifuentes V, 2018. Isolation and Characterization of Extrachromosomal Double-Stranded RNA Elements from Carotenogenic Yeasts. Microbial Carotenoids: Methods and Protocols, 1852, 327–339.
- Bagheri B, Zambelli P, Vigentini I, Bauer FF and Setati ME, 2018. Investigating the Effect of Selected Non-*Saccharomyces* Species on Wine Ecosystem Function and Major Volatiles. Frontiers in Bioengineering and Biotechnology, 6.

Beauvais A and Latge J-P, 2018. Special Issue: Fungal Cell Wall. Journal of Fungi, 4.

- Belinato de Souza JR, Kupper KC and Augusto F, 2018. In vivo investigation of the volatile metabolome of antiphytopathogenic yeast strains active against *Penicillium digitatum* using comprehensive two-dimensional gas chromatography and multivariate data analysis. Microchemical Journal, 141, 204–209.
- Beyer R, Jandric Z, Zutz C, Gregori C, Willinger B, Jacobsen ID, Kovarik P, Strauss J and Schuller C, 2018. Competition of *Candida glabrata* against Lactobacillus is Hog1 dependent. Cellular Microbiology, 20.
- Bhakt P, Shivarathri R, Choudhary DK, Borah S and Kaur R, 2018. Fluconazole-induced actin cytoskeleton remodeling requires phosphatidylinositol 3-phosphate 5-kinase in the pathogenic yeast *Candida glabrata*. Molecular Microbiology, 110, 425–443.
- Bharathi R, 2018. Comparison of Chromogenic Media with the Corn Meal Agar for Speciation of *Candida*. Journal of Pure and Applied Microbiology, 12, 1617–1622.
- Bhatt RS, Sahoo A, Karim SA and Gadekar YP, 2018. Effects of *Saccharomyces cerevisiae* and rumen bypass-fat supplementation on growth, nutrient utilisation, rumen fermentation and carcass traits of lambs. Animal Production Science, 58, 530–538.
- Biagiotti C, Ciani M, Canonico L and Comitini F, 2018. Occurrence and involvement of yeast biota in ripening of Italian Fossa cheese. European Food Research and Technology, 244, 1921–1931.
- Biasucci G, 2018. Gut perturbation and probiotics in neonatology. Journal of Pediatric and Neonatal Individualized Medicine, 7.
- Bijelic B, Matic IZ, Besu I, Jankovic L, Juranic Z, Marusic S and Andrejevic S, 2018. Celiac diseasespecific and inflammatory bowel disease-related antibodies in patients with recurrent aphthous stomatitis. Immunobiology.

- Bonciani T, De Vero L, Giannuzzi E, Verspohl A and Giudici P, 2018. Qualitative and quantitative screening of the -glucosidase activity in *Saccharomyces cerevisiae* and *Saccharomyces uvarum* strains isolated from refrigerated must. Letters in Applied Microbiology, 67, 72–78.
- Buckova M, Puskarova A, Zenisova K, Krakova L, Piknova L, Kuchta T and Pangallo D, 2018. Novel insights into microbial community dynamics during the fermentation of Central European ice wine. International Journal of Food Microbiology, 266, 42–51.
- Burr R and Espenshade PJ, 2018. Oxygen-responsive transcriptional regulation of lipid homeostasis in fungi: Implications for anti-fungal drug development. Seminars in Cell & Developmental Biology, 81, 110–120.
- Cabot A, Roquefeuil S and Soula JB, 2018. Yeast probiotics in broilers: benefits for the whole production chain. International Poultry Production, 26, 55–53, 55.
- Calahorra M, Sanchez NS and Pena A, 2018. Influence of phenothiazines, phenazines and phenoxazine on cation transport in *Candida albicans*. Journal of Applied Microbiology, 125, 1728–1738.
- Caldara M and Marmiroli N, 2018. Tricyclic antidepressants inhibit *Candida albicans* growth and biofilm formation. International Journal of Antimicrobial Agents, 52, 500–505.
- Cassilly CD and Reynolds TB, 2018. PS, It's Complicated: The Roles of Phosphatidylserine and Phosphatidylethanolamine in the Pathogenesis of *Candida albicans* and Other Microbial Pathogens. Journal of Fungi, 4.
- Castilho DG, Alencar Chaves AF, Navarro MV, Conceicao PM, Ferreira KS, Silva LS, Xander P and Batista WL, 2018. Secreted aspartyl proteinase (PbSap) contributes to the virulence of *Paracoccidioides brasiliensis* infection. Plos Neglected Tropical Diseases, 12.
- Cen Q-W, Chen T, Zheng W, Zhang Y, Ying R-F, Tang Z-X and Shi L-E, 2018. Isolation, Identification and Partial Characterization of Film-forming Microorganisms from Chinese Homemade Pickle, a Traditional Fermented Vegetable Product. Chiang Mai Journal of Science, 45, 2283–2293.
- Chen J, Wan C-M, Gong S-T, Fang F, Sun M, Qian Y, Huang Y, Wang B-X, Xu C-D, Ye L-Y, Dong M, Jin Y, Huang Z-H, Wu Q-B, Zhu C-M, Fang Y-H, Zhu Q-R and Dong Y-S, 2018. Chinese clinical practice guidelines for acute infectious diarrhea in children. World Journal of Pediatrics, 14, 429–436.
- Cioch-Skoneczny M, Satora P, Skotniczny M and Skoneczny S, 2018. Quantitative and qualitative composition of yeast microbiota in spontaneously fermented grape musts obtained from cool climate grape varieties 'Rondo' and 'Regent'. FEMS Yeast Research, 18.
- Cohen PA, 2018. Probiotic Safety-No Guarantees. Jama Internal Medicine, 178, 1577–1578.
- Corbaci C and Ucar FB, 2018. Purification, characterization and in vivo biocontrol efficiency of killer toxins from *Debaryomyces hansenii* strains. International Journal of Biological Macromolecules, 119, 1077–1082.
- Cristina dos Reis K, Arrizon J, Amaya-Delgado L, Gschaedler A, Freitas Schwan R and Ferreira Silva C, 2018. Volatile compounds flavoring obtained from Brazilian and Mexican spirit wastes by yeasts. World Journal of Microbiology & Biotechnology, 34.
- Csoma H, Acs-Szabo L, Papp LA and Sipiczki M, 2018. Application of different markers and dataanalysis tools to the examination of biodiversity can lead to different results: a case study with *Starmerella bacillaris* (synonym *Candida zemplinina*) strains. Fems Yeast Research, 18.
- Cully M, 2018. ANTIFUNGAL DRUGS Small molecules targeting a tertiary RNA structure fight fungi. Nature Reviews Drug Discovery, 17.
- Cummings D, Cruise M, Lopez R, Roggenbuck D, Jairath V, Wang Y, Shen B and Rieder F, 2018. Loss of tolerance to glycoprotein 2 isoforms 1 and 4 is associated with Crohn's disease of the pouch. Alimentary Pharmacology & Therapeutics, 48, 1251–1259.
- Czech A, Smolczyk A, Ognik K, Wlazlo L, Nowakowicz-Debek B and Kiesz M, 2018. Effect of dietary supplementation with *Yarrowia lipolytica* or *Saccharomyces cerevisiae* yeast and probiotic additives on haematological parameters and the gut microbiota in piglets. Research in Veterinary Science, 119, 221–227.
- Dalhoff A, 2018. Does the use of antifungal agents in agriculture and food foster polyene resistance development? A reason for concern. Journal of Global Antimicrobial Resistance, 13, 40–48.
- Daliri EB-M, Tango CN, Lee BH and Oh D-H, 2018. Human microbiome restoration and safety. International Journal of Medical Microbiology, 308, 487–497.
- Das S, Rai G, Tigga RA, Srivastava S, Singh PK, Sharma R, Datt S, Singh NP and Dar SA, 2018. *Candida auris* in critically ill patients: Emerging threat in intensive care unit of hospitals. Journal De Mycologie Medicale, 28, 514–518.

- Davis SE, Tams RN, Solis NV, Wagner AS, Chen T, Jackson JW, Hasim S, Montedonico AE, Dinsmore J, Sparer TE, Filler SG and Reynolds TB, 2018. *Candida albicans* Cannot Acquire Sufficient Ethanolamine from the Host To Support Virulence in the Absence of De Novo Phosphatidylethanolamine Synthesis. Infection and Immunity, 86.
- De Marco S, Sichetti M, Muradyan D, Piccioni M, Traina G, Pagiotti R and Pietrella D, 2018. Probiotic Cell-Free Supernatants Exhibited Anti-Inflammatory and Antioxidant Activity on Human Gut Epithelial Cells and Macrophages Stimulated with LPS. Evidence-Based Complementary and Alternative Medicine.
- Demirhan B, Guragac FT, Er-Demirhan B, Tastan H and Kupeli-Akkol E, 2018. A PRECLINICAL STUDY: EFFECT OF NEW PROBIOTIC PRODUCTS, "*BACILLUS SUBTILIS* AND *BACILLUS AMYLOLIQUEFACIENS*", ON DIARRHEA. Fresenius Environmental Bulletin, 27, 4139–4148.
- Demuyser L, Van Genechten W, Mizuno H, Colombo S and Van Dijck P, 2018. Introducing fluorescence resonance energy transfer-based biosensors for the analysis of cAMP-PKA signalling in the fungal pathogen Candida glabrata. Cellular Microbiology, 20.
- Devi G, Harikrishnan R, Paray BA, Al-Sadoon MK, Hoseinifar SH and Balasundaram C, 2018. Comparative immunostimulatory effect of probiotics and prebiotics in *Channa punctatus* against Aphanomyces invadans. Fish & Shellfish Immunology, 86, 965–973.
- Diazid TG, Branco AF, Jacovaci FA, Jobim CC, Pratti Daniel JL, Iank Bueno AV and Ribeiro MG, 2018. Use of live yeast and mannan-oligosaccharides in grain-based diets for cattle: Ruminal parameters, nutrient digestibility, and inflammatory response. Plos One, 13.
- Diba K, Makhdoomi K, Nasri E, Vaezi A, Javidnia J, Gharabagh DJ, Jazani NH, Chavshin AR, Badiee P, Badali H and Fakhim H, 2018. Emerging *Candida* species isolated from renal transplant recipients: Species distribution and susceptibility profiles. Microbial Pathogenesis, 125, 240–245.
- Dudzicz S, Kujawa-Szewieczek A, Kwiecien K, Wiecek A and Adamczak M, 2018. *Lactobacillus plantarum* 299v Reduces the Incidence of Clostridium difficile Infection in Nephrology and Transplantation Ward-Results of One Year Extended Study. Nutrients, 10.
- Duo Saito RA, Connell L, Rodriguez R, Redman R, Libkind D and e Garcia V, 2018. Metabarcoding analysis of the fungal biodiversity associated with Castano Overa Glacier Mount Tronador, Patagonia, Argentina. Fungal Ecology, 36, 8–16.
- De Paula Menezes R, Silva FF, Melo SGO, Alves PGV, Brito MO, e Souza Bessa MA, Amante Penatti MP, Pedroso RS, Abdallah VOS and Roder DvDB, 2018. Characterization of *Candida* species isolated from the hands of the healthcare workers in the neonatal intensive care unit. Medical Mycology.
- El Khoury S, Rousseau A, Lecoeur A, Cheaib B, Bouslama S, Mercier P-L, Demey V, Castex M, Giovenazzo P and Derome N, 2018. Deleterious Interaction Between Honeybees (*Apis mellifera*) and its Microsporidian Intracellular Parasite *Nosema ceranae* Was Mitigated by Administrating Either Endogenous or Allochthonous Gut Microbiota Strains. Frontiers in Ecology and Evolution, 6.
- El Kurdi B, Khatua B, Snozek C and Singh VP, 2018. ADMISSION SERUM UNBOUND FATTY ACIDS CORRELATE WITH LENGTH OF HOSPITAL STAY IN PATIENTS WITH ACUTE PANCREATITIS (AP). Gastroenterology, 154, S294–S295.
- El Mouzan MI, Korolev KS, Al Mofarreh MA, Menon R, Winter HS, Al Sarkhy AA, Dowd SE, Al Barrag AM and Assiri AA, 2018. Fungal dysbiosis predicts the diagnosis of pediatric Crohn's disease. World Journal of Gastroenterology, 24, 4510–4516.
- Eldarov MA, Beletsky AV, Tanashchuk TN, Kishkovskaya SA, Ravin NV and Mardanov AV, 2018. Whole-Genome Analysis of Three Yeast Strains Used for Production of Sherry-Like Wines Revealed Genetic Traits Specific to Flor Yeasts. Frontiers in Microbiology, 9.
- Ene LV, Farrer RA, Hirakawa MP, Agwamba K, Cuomo CA and Bennett RJ, 2018. Global analysis of mutations driving microevolution of a heterozygous diploid fungal pathogen. Proceedings of the National Academy of Sciences of the United States of America, 115, E8688–E8697.
- Fan G, Xu D, Fu Z, Xu C, Teng C, Sun B and Li X, 2018. Screen of aroma-producing yeast strains from Gujinggong Daqu and analysis of volatile flavor compounds produced by them. Journal of Chinese Institute of Food Science and Technology, 18, 220–229.
- Ferraz Dellias, MdT, Borges CD, Lopes ML, Cruz SH, e Amorim HV and Tsai SM, 2018. Biofilm formation and antimicrobial sensitivity of lactobacilli contaminants from sugarcane-based fuel ethanol fermentation. Antonie Van Leeuwenhoek International Journal of General and Molecular Microbiology, 111, 1631–1644.
- Ferreira RM, Aguiar F, Rocha TM, Ganhao S, Agueda A, Guerra M, Pimenta S, Bernardes M and Costa L, 2018. ANTI-*SACCHAROMYCES CEREVISIAE* ANTIBODIES IN SPONDYLARTHROPATHIES: PREVALENCE AND ASSOCIATIONS WITH DISEASE PHENOTYPE. Annals of the Rheumatic Diseases, 77, 856–856.

- Florencia Perez M, Sofia Isas A, Aladdin A, El Enshasy HA and Rafael Dib J, 2018. Killer Yeasts as Biocontrol Agents of Postharvest Fungal Diseases in Lemons. Sustainable Technologies for the Management of Agricultural Wastes, 87–98.
- Florez ID, Veroniki A-A, Al Khalifah R, Yepes-Nunez JJ, Sierra JM, Vernooij RWM, Acosta-Reyes J, Granados CM, Perez-Gaxiola G, Cuello-Garcia C, Zea AM, Zhang Y, Foroutan N, Guyatt GH and Thabane L, 2018. Comparative effectiveness and safety of interventions for acute diarrhea and gastroenteritis in children: A systematic review and network meta-analysis. Plos One, 13.
- Forche A, Cromie G, Gerstein AC, Solis NV, Pisithkul T, Srifa W, Jeffery E, Abbey D, Filler SG, Dudley AM and Berman J, 2018. Rapid Phenotypic and Genotypic Diversification After Exposure to the Oral Host Niche in Candida albicans. Genetics, 209, 725–741.
- Foster AJ, 2018. Identification of Fungicide Targets in Pathogenic Fungi. Physiology and Genetics: Selected Basic and Applied Aspects, 2nd Edition, 277–296.
- Freedman SB, Williamson-Urquhart S, Farion KJ, Gouin S, Willan AR, Poonai N, Hurley K, Sherman PM, Finkelstein Y, Lee BE, Pang X-L, Chui L, Schnadower D, Xie J, Gorelick M and Schuh S, 2018. Multicenter Trial of a Combination Probiotic for Children with Gastroenteritis. New England Journal of Medicine, 379, 2015–2026.
- Frohlich-Wyder M-T, Arias-Roth E and Jakob E, 2018. Cheese Yeasts. Yeast (Chichester, England).
- Gabaldon T and Fairhead C, 2018. Genomes shed light on the secret life of *Candida glabrata*: not so asexual, not so commensal. Current Genetics.
- Gallone B, Mertens S, Gordon JL, Maere S, Verstrepen KJ and Steensels J, 2018. Origins, evolution, domestication and diversity of *Saccharomyces* beer yeasts. Current Opinion in Biotechnology, 49, 148–155.
- Gao J, Wang H, Li Z, Wong AH-H, Wang Y-Z, Guo Y, Lin X, Zeng G, Wang Y and Wang J, 2018. Candida albicans gains azole resistance by altering sphingolipid composition. Nature Communications 9.
- Garcia MD, Chua SMH, Low Y-S, Lee Y-T, Agnew-Francis K, Wang J-G, Nouwens A, Lonhienne T, Williams CM, Fraser JA and Guddat LW, 2018. Commercial AHAS-inhibiting herbicides are promising drug leads for the treatment of human fungal pathogenic infections. Proceedings of the National Academy of Sciences of the United States of America, 115, E9649–E9658.
- Garcia-Agudo L, Rodriguez-Iglesias M and Carranza-Gonzalez R, 2018. Nosocomial Candiduria in the Elderly: Microbiological Diagnosis. Mycopathologia, 183, 591–596.
- Ghasemian A, Eslami M, Shafiei M, Najafipour S and Rajabi A, 2018. Probiotics and their increasing importance in human health and infection control. Reviews in Medical Microbiology, 29, 153–158.
- Ghodhbane H, Ben Hamed K, Alesendria V, Abdelly C, Cocolin L and Regaya I, 2018. ANTIBACTERIAL AND ANTIFUNGAL ACTIVITY OF LAB BACTERIOCINS DERIVED FROM THE VALORIZATION OF DAIRY BYPRODUCTS USING AWDA ASSAY. Toxicon, 149, 96–96.
- Glinka P, Nowacka K, Kulesza K, Ejdys E and Dynowska M, 2018. Share of the Saccharomyces genus in the mycobiota of the gastrointestinal tract of oncology patients potential effects of a fruit-based diet. Annals of Parasitology, 64, 199–202.
- Gobesso, A. A. O., G. V. Pombo, R. L. Costa, Y. S. Pereira and K. Feltre (2018). Effect of yeast supplementation on digestibility, fecal microbiota and serum endotoxin levels in non-exercising and exercising horses. Livestock Science 215: 21-24.
- Gonzalez T, Malagon C, Guarnizo P, Mosquera AC, Chila-Moreno L and Romero-Sanchez C, 2018. Autoantibodies and Gastrointestinal Symptoms in Colombian Children with Juvenile Idiopathic Arthritis. Current Rheumatology Reviews, 14, 163–171.
- Gordun E, Puig A, Pinol L and Carbo R, 2018. Identification of yeast isolated from laboratory sourdoughs prepared with grape, apple, and yogurt. Journal of Microbiology, Biotechnology and Food Sciences, 7, 399–403.
- Goren I, Godny L, Reshef L, Yanai H, Gophna U, Tulchinsky H and Dotan I, 2018. Starch Consumption May Modify Antiglycan Antibodies and Fecal Fungal Composition in Patients With Ileo-Anal Pouch. Inflammatory bowel diseases.
- Gorgel A, Cankaya C and Tecellioglu M, 2018. Anti-*Saccharomyces cerevisiae* as Unusual Antibody in Autoimmune Polyglandular Syndrome Type III: A Case Report. Endocrine, metabolic & immune disorders drug targets.
- Gress A, Serratorre N and Briggs SD, 2018. Determining the role of the epigenetic factor SET4 in antifungal drug resistance in budding yeast. Faseb Journal, 32.

- Gupta S, Bhathena ZP, Kumar S, Srivastava PP and Jadhao SB, 2018. Quantification and Characterization of Mannan Oligosaccharide Producing Yeasts isolated from Various Food Products. Proceedings of the Indian National Science Academy Part B Biological Sciences, 88, 1237–1247.
- Gurina OP, Stepanova AA, Dementieva EA, Blinov AE, Varlamova ON and Blinov GA, 2018. TITLE. Klinicheskaia laboratornaia diagnostika, 63, 44–50.
- Gut AM, Vasiljevic T, Yeager T and Donkor ON, 2018. *Salmonella* infection prevention and treatment by antibiotics and probiotic yeasts: a review. Microbiology-Sgm, 164, 1327–1344.
- Gutierrez A, Boekhout T, Gojkovic Z and Katz M, 2018. Evaluation of non-*Saccharomyces* yeasts in the fermentation of wine, beer and cider for the development of new beverages. Journal of the Institute of Brewing, 124, 389–402.
- Haastrup MK, Johansen P, Malskaer AH, Castro-Mejia JL, Kot W, Krych L, Arneborg N and Jespersen L, 2018. Cheese brines from Danish dairies reveal a complex microbiota comprising several halotolerant bacteria and yeasts. International Journal of Food Microbiology, 285, 173–187.
- Hardwick JM, 2018. Do Fungi Undergo Apoptosis-Like Programmed Cell Death? Mbio, 9.
- Hasan KAM and Yassein SN, 2018. Prevalence and type of fungi in milk from goats with sub clinical mastitis. Online Journal of Veterinary Research, 22, 669–674.
- Hayashi K, Yamaguchi Y, Ogita A, Tanaka T, Kubo I and Fujita K-I, 2018. Effect of nagilactone E on cell morphology and glucan biosynthesis in budding yeast *Saccharomyces cerevisiae*. Fitoterapia, 128, 112–117.
- Healey KR, Kordalewska M, Ortigosa CJ, Singh A, Berrio I, Chowdhary A and Perlin DS, 2018. Limited ERG11 Mutations Identified in Isolates of Candida au ris Directly Contribute to Reduced Azole Susceptibility. Antimicrobial Agents and Chemotherapy, 62.
- Hernandez A, Perez-Nevado F, Ruiz-Moyano S, Serradilla MJ, Villalobos MC, Martin A and Cordoba MG, 2018. Spoilage yeasts: What are the sources of contamination of foods and beverages? International Journal of Food Microbiology, 286, 98–110.
- Hernandez-Chavez MJ, Franco B, Clavijo-Giraldo DM, Hernandez NV, Estrada-Mata E and Manuel Mora-Montes H, 2018. Role of protein phosphomannosylation in the *Candida tropicalis*-macrophage interaction. Fems Yeast Research, 18.
- Hernanz N, Martin-de Argila C and Albillos A, 2018. ARE PROBIOTICS USEFUL IN GASTROINTESTINAL DISEASES? Revista Castellana De Gastroenterologia, 34, 48–54.
- Horn MP, Peter AM, Grunder FR, Leichtle AB, Spalinger J, Schibli S and Sokollik C, 2018. PR3-ANCA and panel diagnostics in pediatric inflammatory bowel disease to distinguish ulcerative colitis from Crohn's disease. Plos One, 13.
- Hu L, Wang J, Ji X, Liu R, Chen F and Zhang X, 2018. Selection of non-*Saccharomyces* yeasts for orange wine fermentation based on their enological traits and volatile compounds formation. Journal of Food Science and Technology-Mysore, 55, 4001–4012.
- Hu XQ, Liu Q, Hu JP, Zhou JJ, Zhang X, Peng SY, Peng LJ and Wang XD, 2018. Identification and characterization of probiotic yeast isolated from digestive tract of ducks. Poultry Science, 97, 2902–2908.
- Ienascu IMC, Balaes T, Petre CV, Pop RO, Cata A, Stefanut MN, Albu P and Poenaru M, 2018. Novel N-(2-bromo-phenyl)-2-hydroxy-benzamide Derivatives with Antifungal Activity. Revista De Chimie, 69, 1876–1880.
- Imarhiagbe EE and Ikhajiagbe B, 2018. Antibiotic susceptibility of bacterial isolates and water quality index of water sourced from closed ground water and open hand dug well in Koko Community, Delta State, Nigeria. Studia Universitatis Babes-Bolyai Biologia, 63, 47–57.
- Jakopovic Z, Cica KH, Mrvcic J, Pucic I, Canak I, Frece J, Pleadin J, Stanzer D, Zjalic S and Markov K, 2018. Properties and Fermentation Activity of Industrial Yeasts *Saccharomyces cerevisiae*, *S. uvarum, Candida utilis* and *Kiuyveromyces marxianus* Exposed to AFB(1), OTA and ZEA. Food Technology and Biotechnology, 56, 208–217.
- Jiang L and Yang Y, 2018. The putative transient receptor potential channel protein encoded by the orf19.4805 gene is involved in cation sensitivity, antifungal tolerance, and filamentation in Candida albicans. Canadian Journal of Microbiology, 64, 727–731.
- Johnston BC, Lytvyn L, Lo CK-F, Allen SJ, Wang D, Szajewska H, Miller M, Ehrhardt S, Sampalis J, Duman DG, Pozzoni P, Colli A, Lonnermark E, Selinger CP, Wong S, Plummer S, Hickson M, Pancheva R, Hirsch S, Klarin B, Goldenberg JZ, Wang L, Mbuagbaw L, Foster G, Maw A, Sadeghirad B, Thabane L and Mertz D, 2018. Microbial Preparations (Probiotics) for the Prevention of *Clostridium difficile* Infection in Adults and Children: An Individual Patient Data Meta-analysis of 6,851 Participants. Infection Control and Hospital Epidemiology, 39, 771–781.

- Jung J, Moon YS, Yoo JA, Lim J-H, Jeong J and Jun J-B, 2018. Investigation of a nosocomial outbreak of fungemia caused by *Candida pelliculosa* (*Pichia anomala*) in a Korean tertiary care center. Journal of Microbiology Immunology and Infection, 51, 794–801.
- Kara I, Yildirim F, Ozgen O, Erganis S, Aydogdu M, Dizbay M, Gursel G and Kalkanci A, 2018. *Saccharomyces cerevisiae* fungemia after probiotic treatment in an intensive care unit patient. Journal De Mycologie Medicale, 28, 218–221.
- Kaur R, Mehra B, Dhakad MS, Goyal R, Bhalla P and Dewan R, 2018. Clinico-mycological analysis and antifungal resistance pattern in human immunodeficiency virus-associated candidiasis: An Indian perspective. Indian journal of sexually transmitted diseases and AIDS, 39, 111–119.
- Kawamoto S, Moranova Z, Virtudazo E, Ohusu M and Raclavsky V, 2018. Cell Cycle Regulation and Hypoxic Adaptation in the Pathogenic Yeast *Cryptococcus neoformans*. Faseb Journal, 32.
- Kayser WC, Carstens GE, Washbun KE, Lawhon SD, Reddy SM, Skidmore AL, Chevaux E and Pinchak WE, 2018. Effects of Live Yeast Supplementation on Complete Blood Cell Count and Febrile Responses in Heifers after Viral-Bacterial Respiratory Challenge. Journal of Animal Science, 96, 57–57.
- Keniya MV, Ruma YN, Tyndall JDA and Monka BC, 2018. Heterologous Expression of Full-Length Lanosterol 14 alpha-Demethylases of Prominent Fungal Pathogens *Candida albicans* and *Candida glabrata* Provides Tools for Antifungal Discovery. Antimicrobial Agents and Chemotherapy, 62.
- Keniya MV, Sabherwal M, Wilson RK, Woods MA, Sagatova AA, Tyndall JDA and Monk BC, 2018. Crystal Structures of Full-Length Lanosterol 14 alpha-Demethylases of Prominent Fungal Pathogens *Candida albicans* and *Candida glabrata* Provide Tools for Antifungal Discovery. Antimicrobial Agents and Chemotherapy, 62.
- Kesmen Z, Buyukkiraz ME, Ozbekar E, Celik M, Ozkok FO, Kilic O, Cetin B and Yetim H, 2018. Assessment of Multi Fragment Melting Analysis System (MFMAS) for the Identification of Food-Borne Yeasts. Current Microbiology, 75, 716–725.
- Kesmen Z, Ozbekar E and Buyukkiraz ME, 2018. Multifragment melting analysis of yeast species isolated from spoiled fruits. Journal of Applied Microbiology, 124, 522–534.
- Khedri S, Santos ALS, Roudbary M, Hadighi R, Falahati M, Farahyar S, Khoshmirsafa M and Kalantari S, 2018. Iranian HIV/AIDS patients with oropharyngeal candidiasis: identification, prevalence and antifungal susceptibility of *Candida* species. Letters in Applied Microbiology, 67, 392–399.
- Kirkland TN, 2018. Special Issue: Genomic Data in Pathogenic Fungi. Journal of Fungi, 4.
- Kobayashi Y, Maeda T, Yamaguchi K, Kameoka H, Tanaka S, Ezawa T, Shigenobu S and Kawaguchi M, 2018. The genome of Rhizophagus clarus HR1 reveals a common genetic basis for auxotrophy among arbuscular mycorrhizal fungi. BMC Genomics, 19.
- Kodedova M, Valachovic M, Csaky Z and Sychrova H, 2018. Effectiveness of three families of antifungal peptides is influenced by lipid composition of the yeast plasma membrane. Febs Open Bio, 8, 364–364.
- Kong HS and Sani AN, 2018. Antimicrobial properties of the acetone leaves and stems extracts of Clinacanthus nutans from three different samples/areas against pathogenic microorganisms. International Food Research Journal, 25, 1698–1702.
- Kregiel D, James SA, Rygala A, Berlowska J, Antolak H and Pawlikowska E, 2018. Consortia formed by yeasts and acetic acid bacteria *Asaia* spp. in soft drinks. Antonie Van Leeuwenhoek International Journal of General and Molecular Microbiology, 111, 373–383.
- Krogerus K, Preiss R and Gibson B, 2018. A Unique *Saccharomyces cerevisiae* x *Saccharomyces uvarum* Hybrid Isolated From Norwegian Farmhouse Beer: Characterization and Reconstruction. Frontiers in Microbiology, 9.
- Kulesza K, Biedunkiewicz A, Nowacka K and Glinka P, 2018. Potentially pathogenic fungi of the *Candida* genus isolated from the Lyna River a 20-year study. Annals of Parasitiology, 64, 217–223.
- Kumari S, Kumar M, Khandelwal NK, Kumari P, Varma M, Vishwakarma P, Shahi G, Sharma S, Lynn AM, Prasad R and Gaur NA, 2018. ABC transportome inventory of human pathogenic yeast *Candida glabrata*: Phylogenetic and expression analysis. Plos One, 13.
- Laghi L, Zhu C, Campagna G, Rossi G, Bazzano M and Laus F, 2018. Probiotic supplementation in trained trotter horses: effect on blood clinical pathology data and urine metabolomic assessed in field. Journal of Applied Physiology, 125, 654–660.
- Lauterbach A, Wilde C, Bertrand D, Behr J and Vogel RF, 2018. Rating of the industrial application potential of yeast strains by molecular characterization. European Food Research and Technology, 244, 1759–1772.



- Lazo-Velez MA, Serna-Saldivar SO, Rosales-Medina MF, Tinoco-Alvear M and Briones-Garcia M, 2018. Application of *Saccharomyces cerevisiae* var. *boulardii* in food processing: a review. Journal of Applied Microbiology, 125, 943–951.
- Lee KK, Kubo K, Abdelaziz JA, Cunningham I, e Silva Dantas A, Chen X, Okada H, Ohya Y and Gow NAR, 2018. Yeast species-specific, differential inhibition of beta-1,3-glucan synthesis by poacic acid and caspofungin. Cell surface (Amsterdam), 3, 12–25.
- Leiter E, Csernoch L and Pocsi I, 2018. Programmed cell death in human pathogenic fungi a possible therapeutic target. Expert Opinion on Therapeutic Targets, 22, 1039–1048.
- Leiva-Pelaez O, Gutierrez-Escobedo G, Lopez-Fuentes E, Cruz-Mora J, De Las Penas A and Castano I, 2018. Molecular characterization of the silencing complex SIR in *Candida glabrata* hyperadherent clinical isolates. Fungal Genetics and Biology, 118, 21–31.
- Li N, Zheng B, Cai HF, Chen YH, Qiu MQ and Liu MB, 2018. Cost-effectiveness analysis of oral probiotics for the prevention of *Clostridium* difficile-associated diarrhoea in children and adolescents. Journal of Hospital Infection, 99, 469–474.
- Li R-Y, Zheng X-W, Zhang X, Yan Z, Wang X-Y and Han B-Z, 2018. Characterization of bacteria and yeasts isolated from traditional fermentation starter (Fen-Daqu) through a H-1 NMR-based metabolomics approach. Food Microbiology, 76, 11–20.
- Liao E, Xu Y, Jiang Q and Xia W, 2018. Characterisation of dominant autochthonous strains for nitrite degradation of Chinese traditional fermented fish. International Journal of Food Science and Technology, 53, 2633–2641.
- Libkind D, Moline M and Colabella F, 2018. Isolation and Selection of New Astaxanthin-Producing Strains of *Phaffia rhodozyma*. Microbial Carotenoids: Methods and Protocols, 1852, 297–310.
- Lin X, Cai Z and Lin G, 2018. Observation on the clinical effects of *Saccharomyces boulardii* sachets in preventing antibiotic-associated diarrhea in children. Zhongguo Weishengtaxixue Zazhi/Chinese Journal of Microecology, 30, 327–337.
- Liu Y, Huang X, Wan Y, Ma W, Li Y and Qin Q, 2018. Pathogen distribution and drug resistance analysis of clinical fungal bloodstream infection. Journal of Modern Laboratory Medicine, 33, 81–86.
- Liu Y, Liu Q, Hesketh J, Huang D, Gan F, Hao S, Tang S, Guo Y and Huang K, 2018. Protective effects of selenium-glutathione-enriched probiotics on CCl4-induced liver fibrosis. Journal of Nutritional Biochemistry, 58, 138–149.
- Liu Y, Tran DQ and Rhoads JM, 2018. Probiotics in Disease Prevention and Treatment. Journal of Clinical Pharmacology, 58, S164–S179.
- Lu X, Zhang M, Zhao L, Ge K, Wang Z, Jun L and Ren F, 2018. Growth Performance and Post-Weaning Diarrhea in Piglets Fed a Diet Supplemented with Probiotic Complexes. Journal of Microbiology and Biotechnology, 28, 1791–1799.
- Luraschi A, Richard S and Hauser PM, 2018. Site-Directed Mutagenesis of the 1,3-beta-Glucan Synthase Catalytic Subunit of *Pneumocystis jirovecii* and Susceptibility Assays Suggest Its Sensitivity to Caspofungin. Antimicrobial Agents and Chemotherapy, 62.
- Lv C, Ming D and Lei T-T, 2018. A performance evaluation of an immuno-latex chromatography card for the rapid detection of Candida spp. Journal of microbiological methods, 157, 4–8.
- Mahmood NN and Raham DH, 2018. PURIFICATION AND ANTIMICROBIAL ACTIVITY OF BIOSURFACTANTS FROM SACCHAROMYCES CEREVISIAE AGAINST PLANKTONIC CELL AND BIOFILM OF ACINETOBACTER BAUMANNII. Biochemical and Cellular Archives, 18, 17–23.
- Maldonado I, Cataldi S, Garbasz C, Relloso S, Striebeck P, Guelfand L, Lopez Moral L and B. Red Micologia Ciudad Autonoma, 2018. Identification of *Candida* yeasts: Conventional methods and MALDI-TOF MS. Revista Iberoamericana De Micologia, 35, 151–154.
- Maneerattanaporn M, Preechakawin N, Suttikulsombat M and Kiratisin P, 2018. Role of *Saccharomyces boulardii* as adjunctive treatment in non-*Clostridium difficile* infection, nosocomial diarrhea in a tertiary care hospital. Neurogastroenterology and Motility, 30.
- Manohar K, Peroumal D and Acharya N, 2018. TLS dependent and independent functions of DNA polymerase eta (Pol eta/Rad30) from Pathogenic Yeast *Candida albicans*. Molecular Microbiology, 110, 707–727.
- Mantegazza C, Molinari P, D'Auria E, Sonnino M, Morelli L and Zuccotti GV, 2018. Probiotics and antibiotic-associated diarrhea in children: A review and new evidence on *Lactobacillus rhamnosus* GG during and after antibiotic treatment. Pharmacological Research, 128, 63–72.

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- Manuel Galaz-Galaz V, Francisco Moreno-Salazar S, Luis Davila-Ramirez J, Sosa-Castaneda J, Celaya-Michel H, Cesar Morales-Munguia J, Marlene Barrales-Herendia S and Angel Barrera-Silva M, 2018. EFFECTS OF LIVE YEAST (*Saccharomyces cerevisiae*) SUPPLEMENTATION AND DIFFERENT NUTRIENT DENSITY DIETS IN GROWING-FINISHING PIGS UNDER HEAT STRESS. Interciencia, 43, 574–579.
- Mares M, Minea B, Nastasa V, Rosca I, Bostanaru A-C, Marincu I, Toma V, Cristea VC, Murariu C and Pinteala M, 2018. In vitro activity of echinocandins against 562 clinical yeast isolates from a Romanian multicentre study. Medical Mycology, 56, 442–451.
- Martin E, Boccazzi IV, De Marco L, Bongiorno G, Montagna M, Sacchi L, Mensah P, Ricci I, Gradoni L, Bandi C and Epis S, 2018. The mycobiota of the sand fly *Phlebotomus perniciosus*: Involvement of yeast symbionts in uric acid metabolism. Environmental Microbiology, 20, 1064–1077.
- Martorana A, Giuffre AM, Capocasale M, Zappia C and Sidari R, 2018. Sourdoughs as a source of lactic acid bacteria and yeasts with technological characteristics useful for improved bakery products. European Food Research and Technology, 244, 1873-1885.
- Mayerhofer CCK, Awoyemi AO, Moscavitch SD, Lappegard KT, Hov JR, Aukrust P, Hovland A, Lorenzo A, Halvorsen S, Seljeflot I, Gullestad L, Troseid M and Broch K, 2018. Design of the GutHearttargeting gut microbiota to treat heart failuretrial: a Phase II, randomized clinical trial. Esc Heart Failure, 5, 978–985.
- McColl AI, Bleackley MR, Anderson MA and Lowe RGT, 2018. Resistance to the Plant Defensin NaD1 Features Modifications to the Cell Wall and Osmo-Regulation Pathways of Yeast. Frontiers in Microbiology, 9.
- McFarland LV and Goh S, 2018. Are probiotics and prebiotics effective in the prevention of travellers' diarrhea: A systematic review and meta-analysis. Travel Medicine And Infectious Disease.
- McNair LKF, Siedler S, Vinther JMO, Hansen AM, Neves AR, Garrigues C, Jager AK, Franzyk H and Staerk D, 2018. Identification and characterization of a new antifungal peptide in fermented milk product containing bioprotective *Lactobacillus* cultures. FEMS Yeast Research, 18.
- Mendez-Palacios N, Mendez-Mendoza M, Vazquez-Flores F, Castro-Colombres JG and Efren Ramirez-Bribiesca J, 2018. Productive and economic parameters of pigs supplemented from weaning to finishing with prebiotic and probiotic feed additives. Animal Science Journal, 89, 994–1001.
- Mendoza LM, Vega-Lopez GA, Fernandez de Ullivarri M and Raya RR, 2018. Population and oenological characteristics of non-*Saccharomyces* yeasts associated with grapes of Northwestern Argentina. Archives of Microbiology.
- Metz KA, Teng X, Coppens I, Lamb HM, Wagner BE, Rosenfeld JA, Chen X, Zhang Y, Kim HJ, Meadow ME, Wang TS, Haberlandt ED, Anderson GW, Leshinsky-Silver E, Bi W, Markello TC, Pratt M, Makhseed N, Garnica A, Danylchuk NR, Burrow TA, Jayakar P, McKnight D, Agadi S, Gbedawo H, Stanley C, Alber M, Prehl I, Peariso K, Ong MT, Mordekar SR, Parker MJ, Crooks D, Agrawal PB, Berry GT, Loddenkemper T, Yang Y, Maegawa GHB, Aouacheria A, Markle JG, Wohlschlegel JA, Hartman AL and Hardwick JM, 2018. KCTD7 deficiency defines a distinct neurodegenerative disorder with a conserved autophagy-lysosome defect. Annals of Neurology, 84, 766–780.
- Michel M, Meier-Dornberg T, Hutzler M and Jacob F, 2018. Alternative brewers' yeasts what's ahead of us? Brauwelt International, 36, 267–269.
- Miranda-Yuquilema JE, Marin-Cardenas A, Lazo-Perez L and Sanchez-Macias D, 2018. Repercussion of lactic bacteria and yeasts on the productive performance and health of piglets. Revista De Investigaciones Veterinarias Del Peru, 29, 1203–1212.
- Mitrofanova D, Dakik P, McAuley M, Medkour Y, Mohammad K and Titorenko VI, 2018. Lipid metabolism and transport define longevity of the yeast *Saccharomyces cerevisiae*. Frontiers in Bioscience-Landmark, 23, 1166–1194.
- Miyoshi J, Sofia MA and Pierre JF, 2018. The evidence for fungus in Crohn's disease pathogenesis. Clinical Journal of Gastroenterology, 11, 449–456.
- Mohy El-Din SM and Mohyeldin MM, 2018. Component analysis and antifungal activity of the compounds extracted from four brown seaweeds with different solvents at different seasons. Journal of Ocean University of China, 17, 1178–1188.

Morillon A, 2018. Antisense (cryptic) IncRNA from yeast to human. Febs Open Bio, 8, 14–14.

- Munir MB, Marsh TL, Blaud A, Hashim R, Joshua WJA and Nor SAM, 2018. Analysing the effect of dietary prebiotics and probiotics on gut bacterial richness and diversity of Asian snakehead fingerlings using T-RFLP method. Aquaculture Research, 49, 3350–3361.
- Nagy F, Bozo A, Toth Z, Daroczi L, Majoros L and Kovacs R, 2018. In vitro antifungal susceptibility patterns of planktonic and sessile *Candida* kefyr clinical isolates. Medical Mycology, 56, 493–500.

- Nally MC, Ponsone ML, Pesce VM, Toro ME, Vazquez F and Chulze S, 2018. Evaluation of behaviour of *Lachancea thermotolerans* biocontrol agents on grape fermentations. Letters in Applied Microbiology, 67, 89–96.
- Naparlo K, Zyracka E, Grzesik M, Bartosz G and Sadowska-Bartosz I, 2018. Influence of catechins on *Saccharomyces cerevisiae* subjected to thermal stress. Free Radical Biology and Medicine, 120, S129–S129.
- Naquin D, Panozzo C, Dujardin G, van Dijk E, D'Aubenton-Carafa Y and Thermes C, 2018. Complete Sequence of the Intronless Mitochondrial Genome of the *Saccharomyces cerevisiae* Strain CW252. Microbiology Resource Announcements, 6.
- Nasiri AH, Towhidi A, Shakeri M, Zhandi M, Dehghan-Banadaky M and Colazo MG, 2018. Effects of live yeast dietary supplementation on hormonal profile, ovarian follicular dynamics, and reproductive performance in dairy cows exposed to high ambient temperature. Theriogenology, 122, 41–46.
- Naumov GI, Shalamitskiy MY, Naumova ES and Lee CF, 2018. Phylogenetics, Biogeography, and Ecology of Methylotrophic Yeasts of the Heterogeneous Genus *Ogataea*: Achievements and Prospects. Microbiology, 87, 443–452.
- Carmo MS, os Santos CI, Araujo MC, Alberto Giron J, Fernandes ES and Monteiro-Neto V, 2018. Probiotics, mechanisms of action, and clinical perspectives for diarrhea management in children. Food & Function, 9, 5074–5095.
- Obiakor CV, Tun HM, Bridgman SL, Arrieta M-C and Kozyrskyj AL, 2018. The association between early life antibiotic use and allergic disease in young children: recent insights and their implications. Expert Review of Clinical Immunology, 14, 841–855.
- Ochiai A, Ogawa K, Fukuda M, Ohori M, Kanaoka T, Tanaka T, Taniguchi M and Sagehashi Y, 2018. Rice Defensin OsAFP1 is a New Drug Candidate against Human Pathogenic Fungi. Scientific Reports, 8.
- Okmen F, Ekici H and Ari SA, 2018. Case Report of a Tubo-ovarian Abscess Caused by *Candida* kefyr. Journal of Obstetrics and Gynaecology Canada, 40, 1466–1467.
- Omran AN and Mansori AG, 2018. Pathogenic Yeasts Recovered From Acne Vulgaris: Molecular Characterization and Antifungal Susceptibility Pattern. Indian Journal of Dermatology, 63, 386–390.
- Padayachee M, Visser J, Viljoen E, Musekiwa A and Blaauw R, 2018. Efficacy and safety of *Saccharomyces boulardii* in the treatment of acute gastroenteritis in the pediatric population: systematic review. Clinical Nutrition, 37(Suppl. 1), S154-S154.
- Padilla B, Gil JV and Manzanares P, 2018. Challenges of the Non-Conventional Yeast *Wickerhamomyces anomalus* in Winemaking. Fermentation-Basel, 4, 68-Article No.: 68.
- Paliy I and Zaika S, 2018. *Saccharomyces boulardii* normalize the colon microflora during eradication of *Helicobacter pylori* in patients with gastroesophageal reflux disease. Helicobacter, 23.
- Panthee S, Hamamoto H, Ishijima SA, Paudel A and Sekimizu K, 2018. Utilization of Hybrid Assembly Approach to Determine the Genome of an Opportunistic Pathogenic Fungus, Candida *albicans* TIMM 1768. Genome Biology and Evolution, 10, 2017–2022.
- Papp C, Kocsis K, Toth R, Bodai L, Willis JR, Ksiezopolska E, Lozoya-Perez NE, Vagvolgyi C, Montes HM, Gabaldon T, Nosanchuk JD and Gacser A, 2018. Echinocandin-Induced Microevolution of *Candida parapsilosis* Influences Virulence and Abiotic Stress Tolerance. Msphere, 3.
- Paramithiotis S and Drosinos EH, 2018. Probiotic Dairy Products: Inventions Toward Ultramodern Production. Innovations in Technologies for Fermented Food and Beverage Industries, 143–157.
- Patino B, Vazquez C, Manning JM, Roncero MIG, Cordoba-Canero D, Di Pietro A and Martinez-del-Pozo A, 2018. Characterization of a novel cysteine-rich antifungal protein from *Fusarium graminearum* with activity against maize fungal pathogens. International Journal of Food Microbiology, 283, 45–51.
- Perpetuini G, Tittarelli F, Mattarelli P, Modesto M, Cilli E, Suzzi G and Tofalo R, 2018. Intraspecies polymorphisms of *Kluyveromyces marxianus* strains from Yaghnob valley. Fems Microbiology Letters, 365.
- Pfaller MA, Rhomberg PR, Wiederhold NP, Gibas C, Sanders C, Fan H, Mele J, Kovanda LL and Castanheira M, 2018. In Vitro Activity of Isavuconazole against Opportunistic Fungal Pathogens from Two Mycology Reference Laboratories. Antimicrobial Agents and Chemotherapy, 62.
- Pinoargote G, Flores G, Cooper K and Ravishankar S, 2018. Effects on survival and bacterial community composition of the aquaculture water and gastrointestinal tract of shrimp (*Litopenaeus vannamei*) exposed to probiotic treatments after an induced infection of acute hepatopancreatic necrosis disease. Aquaculture Research, 49, 3270–3288.
- Preiss R, Tyrawa C, Krogerus K, Garshol LM and van der Merwe G, 2018. Traditional Norwegian Kveik Are a Genetically Distinct Group of Domesticated *Saccharomyces cerevisiae* Brewing Yeasts. Frontiers in Microbiology, 9.

- Prendes LP, Merin MG, Fontana AR, Bottini RA, Ramirez ML and Morata de Ambrosini VI, 2018. Isolation, identification and selection of antagonistic yeast against *Alternaria alternata* infection and tenuazonic acid production in wine grapes from Argentina. International Journal of Food Microbiology, 266, 14–20.
- Pukrop JR, Brennan KM, Funne BJ and Schoonmaker JP, 2018. Effect of a hydrolyzed mannan- and glucan-rich yeast fraction on performance and health status of newly received feedlot cattle. Journal of Animal Science, 96, 3955–3966.
- Quan AS and Eisen MB, 2018. The ecology of the *Drosophila*-yeast mutualism in wineries. Plos One, 13.
- Quintilla R, Kolecka A, Casaregola S, Daniel HM, Houbraken J, Kostrzewa M, Boelchout T and Groenewald M, 2018. MALDI-TOF MS as a tool to identify foodborne yeasts and yeast-like fungi. International Journal of Food Microbiology, 266, 109–118.
- Raghavan V, Bui DT, Al-Sweel N, Friedrich A, Schacherer J, Aquadro CF and Alani E, 2018. Incompatibilities in Mismatch Repair Genes MLH1-PMS1 Contribute to a Wide Range of Mutation Rates in Human Isolates of Baker's Yeast. Genetics, 210, 1253–1266.
- dos Reis LF, Sato Cabral de Araujo CA, Sousa RS, Hamad Minervino AH, Costa Oliveira FL, Mazzocca Lopes Rodrigues FA, Meira-Junior EB, Barreto-Junior RA, Mori CS and Ortolan EL, 2018. Prevention of acute ruminal lactic acidosis in sheep by probiotic or monensin supplementation: clinical aspects. Semina-Ciencias Agrarias, 39, 1575–1584.
- Rex F and Scharfenberger-Schmeer M, 2018. Rapid analysis of contaminants. Deutsche Weinmagazin (No. 19), 11–13.
- Ribeiro MRS, Oliveira DR, Oliveira FMS, Caliari MV, Martins FS, Nicoli JR, Torres MF, Andrade MER, Cardoso VN and Gomes MA, 2018. Effect of probiotic *Saccharomyces boulardii* in experimental giardiasis. Beneficial Microbes, 9, 789–797.
- Ringot-Destrez B, D'Alessandro Z, Lacroix J-M, Mercier-Bonin M, Leonard R and Robbe-Masselot C, 2018. A sensitive and rapid method to determine the adhesion capacity of probiotics and pathogenic microorganisms to human gastrointestinal mucins. Microorganisms, 6.
- Roa ML, Guzman YE and Navarro CA, 2018. Effect of using probiotics in the intestinal morphometry of broiler. Archivos de Zootecnia, 67, 486–492.
- Robinson JL, 2018. Probiotics for Modification of the Incidence or Severity of Respiratory Tract Infections. Pediatric Infectious Disease Journal, 37, 722–724.
- Rocha Gauch LM, Pedrosa SS, F. Silveira-Gomes, R. A. Esteues and S. H. Marques-da-Silva (2018). Isolation of Candida spp. from denture-related stomatitis in Para, Brazil. Brazilian Journal of Microbiology 49(1): 148-151.
- Rodrigues CF and Henriques M, 2018. Portrait of Matrix Gene Expression in *Candida glabrata* Biofilms with Stress Induced by Different Drugs. Genes 9.
- Rodriguez-Nogales A, Algieri F, Garrido-Mesa J, Vezza T, Pilar Utrilla M, Chueca N, Garcia F, M. Elena Rodriguez-Cabezas F and Galvez J, 2018. Intestinal anti-inflammatory effect of the probiotic *Saccharomyces boulardii* in DSS-induced colitis in mice: Impact on microRNAs expression and gut microbiota composition. Journal of Nutritional Biochemistry, 61, 129–139.
- Rokas A, Wisecaver JH and Lind AL, 2018. The birth, evolution and death of metabolic gene clusters in fungi. Nature Reviews Microbiology, 16, 731–744.
- Ruelle L, Deyi VYM, Konopnicki D and Dauby N, 2018. *Saccharomyces cerevisiae* fungemia: risk factors, outcome and link with Enterol (R) administration. Acta Clinica Belgica, 73, 8–8.
- Rui-Yao L, Xiao-Wei Z, Xin Z, Zheng Y, Xiao-Yong W and Bei-Zhong H, 2018. Characterization of bacteria and yeasts isolated from traditional fermentation starter (Fen-Daqu) through a 1H NMR-based metabolomics approach. Food Microbiology, 76, 11–20.
- Sadrossadati SZ, Ghahri M, Imani Fooladi AA, Sayyahfar S, Beyraghi S and Baseri Z, 2018. Phenotypic and genotypic characterization of *Candida* species isolated from candideamia in Iran. Current Medical Mycology, 4, 14–20.
- Sagatova AA, Keniya MV, Tyndall JDA and Monk BC, 2018. Impact of Homologous Resistance Mutations from Pathogenic Yeast on *Saccharomyces cerevisiae* Lanosterol 14 alpha-Demethylase. Antimicrobial Agents and Chemotherapy, 62.
- Sakai K, Hirose T, Iwatsuki M, Chinen T, Kimura T, Suga T, Nonaka K, Nakashima T, Sunazuka T, Usui T, Asami Y, Omura S and Shiomi K, 2018. Pestynol, an Antifungal Compound Discovered Using a *Saccharomyces cerevisiae* 12gene Delta 0HSR-iERG6-Based Assay. Journal of Natural Products, 81, 1604–1609.

- Salas B, Conway HE, Kunta M, Vacek D and Vitek C, 2018. Pathogenicity of *Zygosaccharomyces bailii* and other yeast species to mexican fruit Fly (Diptera:Tephritidae) and mass rearing implications. Journal of Economic Entomology, 111, 2081–2088.
- Sam QH, Yew WS, Seneviratne CJ, Chang MW and Chai LYA, 2018. Immunomodulation as Therapy for Fungal Infection: Are We Closer? Frontiers in Microbiology, 9.
- Sampaio JP, 2018. Microbe Profile: *Saccharomyces eubayanus*, the missing link to lager beer yeasts. Microbiology-Sgm, 164, 1069–1071.
- Sanders ME, Merenstein D, Merrifield CA and Hutkins R, 2018. Probiotics for human use. Nutrition Bulletin, 43, 212–225.
- Sarde SJ, Kempken F and Kumar A, 2018. Spliceosomal proteins encoded by fungal genomes. Current Science, 114, 1677–1686.
- Schlafer S, Kamp A and Garcia JE, 2018. A confocal microscopy based method to monitor extracellular pH in fungal biofilms. Fems Yeast Research, 18.
- Schnabl B, 2018. GUT DYSBIOSIS AND ALCOHOLIC LIVER DISEASE (ALD). Alcoholism-Clinical and Experimental Research, 42, 135A–135A.
- Sendid B, Jawhara S, Sarter H, Maboudou P, Thierny C, Gower-Rousseau C, Colombel JF and Poulain D, 2018. Uric acid levels are independent of anti-*Saccharomyces cerevisiae* antibodies (ASCA) in Crohn's disease: A reappraisal of the role of *S. cerevisiae* in this setting. Virulence, 9, 1224–1229.
- Sgouros G, Chalvantzi I, Mallouchos A, Paraskevopoulos Y, Banilas G and Nisiotou A, 2018. Biodiversity and Enological Potential of Non-*Saccharomyces* Yeasts from Nemean Vineyards. Fermentation-Basel, 4, 32-Article No.: 32.
- Sha SP, Suryavanshi MV, Jani K, Sharma A, Shouche Y and Tamang JP, 2018. Diversity of Yeasts and Molds by Culture-Dependent and Culture-Independent Methods for Mycobiome Surveillance of Traditionally Prepared Dried Starters for the Production of Indian Alcoholic Beverages. Frontiers in Microbiology, 9.
- Shibayama J, Kuda T, Shikano A, Fukunaga M, Takahashi H, Kimura B and Ishizaki S, 2018. Effects of rice bran and fermented rice bran suspensions on caecal microbiota in dextran sodium sulphate-induced inflammatory bowel disease model mice. Food Bioscience, 25, 8–14.
- Shibo W, Pinglan L and Jinlan Z, 2018. Screening and identification of yeasts with dipeptidyl peptidase-IV inhibitory activity. China Brewing, 37, 48–51.
- Shokohi T, Aslani N, Ahangarkani F, Meyabadi MF, Hagen F, Meis JF, Boekhout T, Kolecka A and Badali H, 2018. *Candida infanticola* and *Candida spencermartinsiae* yeasts: Possible emerging species in cancer patients. Microbial Pathogenesis, 115, 353–357.
- Shokohi T, Moradi N, Badram L, Badali H, Ataollahi MR and Afsarian MH, 2018. Molecular Identification of Clinically Common and Uncommon Yeast Species. Jundishapur Journal of Microbiology, 11.
- Simi WB, Leite-Jr DP, Paula CR, Hoffmann-Santos HD, Takahara DT and Hahn RC, 20198. Yeasts and filamentous fungi in psittacidae and birds of prey droppings in midwest region of Brazil: a potential hazard to human health. Brazilian journal of biology = Revista brasleira de biologia.
- Sims JC, 2018. Effects of the cell cycle on vacuole size in *S. cerevisiae* yeast. Faseb Journal, 32.
- Siriyappagouder P, Galindo-Villegas J, Lokesh J, Mulero V, Fernandes JMO and Kiron V, 2018. Exposure to yeast shapes the intestinal bacterial community assembly in Zebrafish Larvae. Frontiers in Microbiology, 9.
- Sniffen JC, McFarland LV, Evans CT and Goldstein EJC, 2018. Choosing an appropriate probiotic product for your patient: An evidence-based practical guide. Plos One, 13.
- Sokol I, Gawel A and Bobrek K, 2018. The prevalence of yeast and characteristics of the isolates from the digestive tract of clinically healthy turkeys. Avian Diseases, 62, 286–290.
- Soman RX, Gupta N, Chaudhari P, Sunavala A, Shetty A and Rodrigues C, 2018. Implications for diagnosis and treatment of infective endocarditis: eight year experience of an infectious disease team in a private tertiary care centre. The Journal of the Association of Physicians of India, 66, 22–25.
- Sony P, Kalyani M, Jeyakumari D, Kannan I and Sukumar RG, 2018. In vitro antifungal activity of cassia fistula extracts against fluconazole resistant strains of *Candida* species from HIV patients. Journal De Mycologie Medicale, 28, 193–200.
- Sovran B, Planchais J, Jegou S, Straube M, Lamas B, Natividad JM, Agus A, Dupraz L, Glodt J, Da Costa G, Michel M-L, Langeila P, Richard ML and Sokol H, 2018. Enterobacteriaceae are essential for the modulation of colitis severity by fungi. Microbiome, 6.



- Sozio E, Pieralli F, Azzini AM, Tintori G, Demma F, Furneri G, Sbrana F, Bertolino G, Fortunato S, Meini S, Bragantini D, Morettini A, Nozzoli C, Menichetti F, Concia E and Tascini C, 2018. A prediction rule for early recognition of patients with candidemia in Internal Medicine: results from an Italian, multicentric, case-control study. Infection, 46, 625–633.
- Su H, Han L and Huang X, 2018. Potential targets for the development of new antifungal drugs. Journal of Antibiotics, 71, 978–991.
- Sulik-Tyszka B, Snarski E, Niedzwiedzka M, Augustyniak M, Myhre TN, Kacprzyk A, Swoboda-Kopec E, Roszkowska M, Dwilewicz-Trojaczek J, Jedrzejczak WW and Wroblewska M, 2018. Experience with *Saccharomyces boulardii* Probiotic in Oncohaematological Patients. Probiotics and Antimicrobial Proteins, 10, 350–355.
- Sun Z and Xiao D, 2018. Review in metabolic modulation of higher alcohols in top-fermenting yeast. Advances in Applied Biotechnology, 444, 767–773.
- Sutthiwong N, Fouillaud M and Dufosse L, 2018. The Influence of pH, NaCl, and the Deacidifying Yeasts *Debaryomyces hansenii* and *Kluyveromyces marxianus* on the Production of Pigments by the Cheese-Ripening Bacteria Arthrobacter arilaitensis. Foods, 7.
- Takahashi H, Sakagawa E, Sakakibara I, Machida C, Miyaki S, Takahashi A, Onai S, Fukuyoshi S, Ohta A, Satou K and Kanamasa S, 2018. Draft Genome Sequence of *Saccharomyces cerevisiae* Strain Hm-1, Isolated from Cotton Rosemallow. Microbiology Resource Announcements, 7.
- Tao F, Xuzeng W, Yifei W, Min S, Lingyun Y and Zhimin X, 2018. Isolation and identification of yeast strains from vineyard soils and aroma compounds of wines fermented by them. Food Science, China, 39, 213–220.
- Teblick A, Jansens H, Dams K, Somville FJ and Jorens PG, 2018. Boerhaave's syndrome complicated by a *Saccharomyces cerevisiae* pleural empyema. Case report and review of the literature. Acta Clinica Belgica, 73, 377-381.
- Thakre A, Zore G, Kodgire S, Kazi R, Mulange S, Patil R, Shelar A, Santhakumari B, Kulkarni M, Kharat K and Karuppayil SM, 2018. Limonene inhibits *Candida albicans* growth by inducing apoptosis. Medical Mycology, 56, 565–578.
- Till P, Mach RL and Mach-Aigner AR, 2018. A current view on long noncoding RNAs in yeast and filamentous fungi. Applied Microbiology and Biotechnology, 102, 7319–7331.
- Treguier P, David M, Gargala G, Camus V, Stamatoullas A, Menard A-L, Lenain P, Contentin N, Lemasle E, Lanic H, Tilly H, Jardin F and Lepretre S, 2018. *Cyberlindnera jadinii* (teleomorph *Candida utilis*) candidaemia in a patient with aplastic anaemia: a case report. JMM Case Reports, 5, e005160-e005160.
- Trivic I and Hojsak I, 2018. Use of probiotics in the prevention of nosocomial infections. Journal of Clinical Gastroenterology, 52, S62–S65.
- Trunk K, Peltier J, Liu Y-C, Dill BD, Walker L, Gow NAR, Stark MJR, Quinn J, Strahl H, Trost M and Coulthurst SJ, 2018. The type VI secretion system deploys antifungal effectors against microbial competitors. Nature Microbiology, 3, 920–931.
- Tuao Gava CA, Carvalho de Castro AP, Pereira CA, Goncalves JS, Cajuhi Araujo LF and Paz CD, 2018. Photoprotector adjuvants to enhance UV tolerance of yeast strains for controlling mango decay using pre-harvest spraying. Biocontrol Science and Technology, 28, 811–822.
- Ulerio-Nunez F, Gjestvang M, O'Handley SF and Gehret AU, 2018. Exploration of a pho13 knockout growth phenotype in non-engineered strains of *Saccharomyces cerevisiae*. Faseb Journal, 32.
- Vazquez J, Grillitsch K, Daum G, Mas A, Torija M-J and Beltran G, 2018. Melatonin Minimizes the Impact of Oxidative Stress Induced by Hydrogen Peroxide in *Saccharomyces* and Non-conventional Yeast. Frontiers in Microbiology, 9.
- Vermeersch SJ, Vandenplas Y, Tanghe A, Elseviers M and Annemans L, 2018. Economic evaluation of *S. boulardii* CNCM 1-745 for prevention of antibiotic-associated diarrhoea in hospitalized patients. Acta Gastro-Enterologica Belgica, 81, 269–276.
- Viitasalo L, Kurppa K, Ashorn M, Saavalainen P, Huhtala H, Ashorn S, Maki M, Ilus T, Kaukinen K and Iltanen S, 2018. Microbial biomarkers in patients with nonresponsive celiac disease. Digestive Diseases and Sciences, 63, 3434–3441.
- Vu K, Thompson GR, Iii R, Chandler CS, Jane ED, Elizabeth ML, Shawn RM, Wieland E and David MG, 2018. Flucytosine resistance in *Cryptococcus gattii* is indirectly mediated by the FCY2-FCY1-FUR1 pathway. Medical Mycology, 56, 857–867.
- Wachholz PA, Nunes VdS, o Valle AP, Jacinto AF and Fortes Villas-Boas PJ, 2018. Effectiveness of probiotics on the occurrence of infections in older people: systematic review and meta-analysis. Age and Ageing, 47, 527–536.



- Wang JM, Bennett RJ and Anderson MZ, 2018. The Genome of the Human Pathogen Candida albicans Is Shaped by Mutation and Cryptic Sexual Recombination. Mbio, 9.
- Whaley SG, Zhang Q, Caudle KE and Rogers PD, 2018. Relative Contribution of the ABC Transporters Cdr1, Pdh1, and Snq2 to Azole Resistance in *Candida glabrata*. Antimicrobial Agents and Chemotherapy, 62.

Williams B, Addepalli B and Limbach P, 2018. Detection of N4-acetylcytidine modification in *Saccharomyces* cerevisiae mRNA. Abstracts of Papers of the American Chemical Society, 255.

- Woo S-Y, Win NN, Wong CP, Ito T, Hoshino S, Ngwe H, Aye AA, Han NM, Zhang H, Hayashi F, Abe I and Morita H, 2018. Two new pyrrolo-2-aminoimidazoles from a Myanmarese marine sponge, *Clathria prolifera*. Journal of Natural Medicines, 72, 803–807.
- Xiang X, 2018. Nuclear movement in fungi. Seminars in Cell & Developmental Biology, 82, 3–16.
- Xiao J, Peng Z, Liao Y, Sun H, Chen W, Chen X, Wei Z, Yang C, Nuessler AK, Liu J and Yang W, 2018. Organ transplantation and gut microbiota: current reviews and future challenges. American Journal of Translational Research, 10, 3330–3344.
- Xiao-yu L, Ke L and Wei-guang Z, 2018. Effects of pure and mixed fermentation on the quality of Stauntonia chinensis wine. Food and Fermentation Industries(No. 10), 134–140.
- Xu J, Li Y, Yang Z, Li C, Liang H, Wu Z and Pu W, 2018. Yeast probiotics shape the gut microbiome and improve the health of early-weaned piglets. Frontiers in Microbiology, 9.
- Xue F, Nan X, Sun F, Pan X, Guo Y, Jiang L and Xiong B, 2018. Metagenome sequencing to analyze the impacts of thiamine supplementation on ruminal fungi in dairy cows fed high-concentrate diets. Amb Express, 8.
- Yang Y, 2018. Regulatory effect of *Saccharomyces boulardii* combined with Lactulose on serum braingut peptides in children with functional constipation. Zhongguo Weishengtaxixue Zazhi/Chinese Journal of Microecology, 30, 441–447.
- Yang Z, Wang Q, Ma K, Shi P, Liu W and Huang Z, 2018. Fluconazole inhibits cellular ergosterol synthesis to confer synergism with berberine against yeast cells. Journal of Global Antimicrobial Resistance, 13, 125–130.
- Yanni AE, Stamataki NS, Konstantopoulos P, Stoupaki M, Abeliatis A, Nikolakea I, Perrea D, Karathanos VT and Tentolouris N, 2018. Controlling type-2 diabetes by inclusion of Cr-enriched yeast bread in the daily dietary pattern: a randomized clinical trial. European Journal of Nutrition, 57, 259–267.
- Yis R and Doluca M, 2018. Identification of Candida species by restriction enzyme analysis. Turkish Journal of Medical Sciences, 48, 1058–1067.
- Yurkov AM, 2018. Yeasts of the soil obscure but precious. Yeast, 35, 369–378.
- Zalewski Z, McManus PS and Page R, 2018. The role of yeasts in the cranberry fruit rot complex. Phytopathology, 108.
- Zhang F, Tang Y, Ren Y, Yao K, He Q, Wan Y and Chi Y, 2018. Microbial composition of spoiled industrial-scale Sichuan paocai and characteristics of the microorganisms responsible for paocai spoilage. International Journal of Food Microbiology, 275, 32–38.

Viruses used for plant protection

Alphaflexiviridae

Potyviridae

Baculoviridae

None.



Appendix E – The 2016 updated list of QPS Status recommended biological agents in support of EFSA risk assessments

The list of QPS status recommended biological agents (EFSA BIOHAZ Panel, 2016) is being maintained in accordance with the self-task mandate of the BIOHAZ Panel (2017–2019). Possible additions to this list are included around every 6 months, with the first Panel Statement adopted in June 2017 and the last Panel Statement planned for adoption in December 2019. These additions are published as updates to the Scientific Opinion (EFSA BIOHAZ Panel, 2016); the latest update is available at https://doi.org/10.2903/j.efsa.2018.5315 and, as of January 2018, also as supporting information linked to every Panel Statement available on the Knowledge Junction at https://doi.org/10.5281/zenodo.1146566.



Appendix F – Microbial species as notified to EFSA, received between October 2018 and March 2019 (reply to ToR 1)

EFSA risk assessment area	Microorganism species/strain	Intended use	EFSA Question number ^(a) and EFSA webpage link ^(b)	Additional information provided by the EFSA Scientific Unit	Previous QPS status of the respective TU? ^(C)	To be evaluated? yes or no ^(d)
Bacteria						
Feed additives	<i>Bacillus amyloliquefaciens</i> CBS143954	Zootechnical additives/Digestibility enhancer endo-1,4-beta-xylanase (EC 3.2.1.8) produced by GMM <i>Trichoderma reesei</i> , subtilisin protease (EC 3.4.21.62) produced by GMM <i>Bacillus subtilis</i> and alpha- amylase (EC 3.2.1.1) produced by GMM <i>Bacillus amyloliquefaciens</i>	EFSA-Q-2018-01040		Yes	No
Plant protection products	<i>Bacillus amyloliquefaciens</i> FZB42	Fungicide	EFSA-Q-2019-00096		Yes	No
Feed additives	<i>Bacillus licheniformis</i> DSM 19670	Zootechnical additives/Digestibility enhancers RONOZYME [®] ProAct (serine protease produced by a GMM strain of <i>Bacillus licheniformis</i> DSM 19670)	EFSA-Q-2019-00156		Yes	No
Feed additives	Bacillus licheniformis ENV01/DSM 32457	Technological additives/Silage additive	EFSA-Q-2018-00690		Yes	No
Feed additives	<i>Bacillus subtilis</i> CBS143946	Zootechnical additives/Digestibility enhancer? endo-1,4-beta-xylanase (EC 3.2.1.8) produced by GMM <i>Trichoderma reesei</i> , subtilisin protease (EC 3.4.21.62) produced by GMM <i>Bacillus subtilis</i> and alpha- amylase (EC 3.2.1.1) produced by GMM <i>Bacillus amyloliquefaciens</i>	EFSA-Q-2018-01040		Yes	No
Feed additives	Bacillus subtilis DSM32324, Bacillus subtilis DSM32325 and Bacillus amyloliquefaciens DSM25840	Zootechnical feed additive/Flora stabiliser	EFSA-Q-2019-00117	Bacillus subtilis DSM32324, Bacillus subtilis DSM32325 and Bacillus amyloliquefaciens DSM25840 are non-GMM, and are all three of natural origin	Yes	No



EFSA risk assessment area	Microorganism species/strain	Intended use	EFSA Question number ^(a) and EFSA webpage link ^(b)	Additional information provided by the EFSA Scientific Unit	Previous QPS status of the respective TU? ^(C)	To be evaluated? yes or no ^(d)
Food enzymes, food additives and flavourings	Burkholderia ubonensis	Food enzyme triacylglycerol lipase produced by <i>Burkholderia</i> <i>ubonensis</i>	EFSA-Q-2019-00056	<i>Burkholderia ubonensis</i> has been used for over 20 years for the production of the food enzyme	No	Yes
Feed additives	Corynebacterium ammoniagenes KCCM 80161	Sensory additives/ Flavouring compounds IMP (disodium 5'-inosinate) produced by fermentation with <i>Corynebacterium ammoniagenes</i> KCCM 80161	EFSA-Q-2019-00040	Disodium 5'-inosinate feed grade is a highly purified product and does not contain any microorganisms. After the fermentation, the cells of the production strain <i>Corynebacterium ammoniagenes</i> KCCM80161 are eliminated by filtration and centrifugation from the fermentation broth	No	Yes
Feed additives	Corynebacterium casei KCCM80190	Nutritional Additives/Amino Acids, their salts and analogues. L-lysine produced by fermentation with a GMM <i>Corynebacterium casei</i> KCCM80190	EFSA-Q-2019-00195		No	Yes
Feed additives	<i>Corynebacterium glutamicum</i> KCCM 80184	Nutritional additives/Amino acids/ Production of L-methionine	EFSA-Q-2018-01017		Yes	No
Feed additives	<i>Corynebacterium glutamicum</i> KCCM80188	Sensory additives/ Flavouring compounds MSG (monosodium L-glutamate) produced by fermentation with <i>Corynebacterium glutamicum</i> KCCM80188	EFSA-Q-2019-00037		No	No
Feed additives	<i>Corynebacterium glutamicum</i> KFCC11043	Nutritional Additives/Amino Acids, their salts and analogues L-lysine sulphate feed grade produced by fermentation with <i>Corynebacterium</i> <i>glutamicum</i> KFCC11043 (GMM)	EFSA-Q-2019-00194		Yes	No



EFSA risk assessment area	Microorganism species/strain	Intended use	EFSA Question number ^(a) and EFSA webpage link ^(b)	Additional information provided by the EFSA Scientific Unit	Previous QPS status of the respective TU? ^(C)	To be evaluated? yes or no ^(d)
Novel foods	<i>Escherichia coli</i> BL21 (DE3) strain	Production of a recombinant protein (novel food) for food supplements	EFSA-Q-2018-00316	GMM <i>E. coli</i> Summary of the application under "APOAEQUORIN" at https://ec.europa.eu/food/safety/novel_ food/authorisations/summary-ongoing- applications-and-notifications_en	No	No
Novel foods	<i>Escherichia coli</i> DSM 32833	Production of novel food 6-SL (6'-sialyllactose sodium salt)	EFSA-2019-0169	GMM E. coli (K12-DH1 derivative)	No	No
Novel foods	<i>Escherichia coli</i> DSM 32834	Production of novel food 3-SL (3'-sialyllactose sodium salt)	EFSA-2019-0204	GMM E. coli (K12-DH1 derivative)	No	No
Feed additives	<i>Escherichia coli</i> K12 KCCM 80096	Nutritional additives/Amino acids/ Production of L-methionine	EFSA-Q-2018-01017		No	No
Feed additives	<i>Escherichia coli</i> <i>KCCM</i> 80109 <i>Escherichia coli</i> <i>KCCM</i> 80197	Sensory additives/ Flavouring compounds/ L-cysteine hydrochloride monohydrate by fermentation with <i>Escherichia coli</i> KCCM 80109 and <i>Escherichia coli</i> KCCM 80197	EFSA-Q-2019-00041	L-cysteine hydrochloride monohydrate feed grade is a highly purified product and does not contain any microorganisms. After the fermentation, the cells of the production strain <i>E. coli</i> K12 KCCM 80109 and KCCM 80197 are removed by filtration and centrifugation from the fermentation broth	No	No
Novel foods	<i>Escherichia coli</i> strain K12 DH1 (DSM 32774)	Production of 2'-fucosyllactose/ difucosyllactose (novel food)	EFSA-Q-2018-00374 http://www.efsa. europa.eu/en/efsa journal/pub/5717	GMM <i>E. coli</i> (K12-DH1 derivative) Summary of application: https://ec.europa.eu/food/sites/ food/files/safety/docs/novel-food_sum_ ongoing-app_2fl-dfl.pdf	No	No
Novel foods	<i>Escherichia coli</i> K12 derivative	For the production of allulose (novel food); a recombinant derivative of <i>Escherichia coli</i> strain K-12 is used in the fermentation process to produce D-psicose 3- epimerase	EFSA-Q-2018-00756	Summary of application: https://ec.europa.eu/food/sites/food/file s/safety/docs/novel-food_sum_ongoing- app_allulose_sum.pdf	No	No
Feed additives	<i>Escherichia coli</i> K12 KCCM 80159 (C 001)	Nutritional additives/Amino acids/ Production of L-valine	EFSA-Q-2018-00712		No	No



EFSA risk assessment area	Microorganism species/strain	Intended use	EFSA Question number ^(a) and EFSA webpage link ^(b)	Additional information provided by the EFSA Scientific Unit	Previous QPS status of the respective TU? ^(C)	To be evaluated? yes or no ^(d)
Feed additives	<i>Escherichia coli NITE</i> BP-02526	Nutritional additives/Amino acids/ Flavouring compounds/Sensory additives/Production of L-histidine monohydrochloride monohydrate	EFSA-Q-2018-00782		No	No
Feed additives	<i>Gluconobacter frateurii</i> NBRC 103465	Zootechnical additives/ <i>Gluconobacter frateurii</i> NBRC 103465 is a glyceric acid producing strain. Used for the production of the active ingredient: 2,3-dihydroxy propanoic acid/glyceric acid (GA), p-enantiomer (p-GA)	EFSA-Q-2018-00999	Other Zootechnical additives/Draft Genome Sequence of <i>Gluconobacter</i> <i>frateurii</i> NBRC 103465, has been published online by Sato et al. in 2013 Jul 25. https://doi.org/10.1128/ genomea.00369-13	No	Yes
Feed additives	<i>Lactobacillus buchneri</i> DSM 29026	Technological Additives/Silage Additives	EFSA-Q-2019-00180	This is a non-holder specific additive. The strain has also been designated MFLBU1 or LBU1	Yes	No
Feed additives	<i>Microbacterium</i> <i>foliorum</i> SYG27B-MF	Production of the novel food D-allulose	EFSA-Q-2018-00797	Non-GMM strain of <i>Microbacterium</i> foliorum	No	Yes
Feed additives	<i>Sphingomonas</i> <i>elodea</i> PS-60 ATCC 31461	Technological additives/Gelling agents, Stabilisers, Thickeners/ Production of Gellan Gum	EFSA-Q-2018-00815		No	Yes
Filamentous	fungi					
Food enzymes, food additives and flavourings	Aspergillus niger (DSM 25770)	Zootechnical additives Digestibility enhancers 6-phytase produced by a GMM strain of <i>Aspergillus niger</i> (DSM 25770)	EFSA-Q-2019-00042	Natuphos [®] E is a preparation of 6- phytase produced by <i>Aspergillus niger</i> , LU17257 (DSM 25770)		
Food enzymes, food additives and flavourings	<i>Aspergillus niger</i> DSM 32805	Food enzyme chymosin IUBMB name and number of the enzyme protein: 3.4.23.4; Chymosin from GMM strain of <i>Aspergillus niger</i> DSM 32805. This strain was previously known under the name <i>A. niger</i> var. awamori	EFSA-Q-2019-00125		No	No



EFSA risk assessment area	Microorganism species/strain	Intended use	EFSA Question number ^(a) and EFSA webpage link ^(b)	Additional information provided by the EFSA Scientific Unit	Previous QPS status of the respective TU? ^(C)	To be evaluated? yes or no ^(d)
Feed additives	Aspergillus niger strain ATCC 26550 and NRC A-1-233	Zootechnical functional group/ Performance enhancer AviPlus [®] is a preparation based on a mixture of sorbic acid, citric acid, thymol and vanillin	EFSA-Q-2019-00154	The production of citric acid process is via fermentation of a stable non-GMM <i>Aspergillus niger</i> strain	No	No
Feed additives	Aspergillus niger strain ATCC 26550/ NRC A-1-233	Zootechnical additive/Performance enhancer AviPlus [®] is a preparation based on a mixture of sorbic acid, citric acid, thymol and vanillin	EFSA-Q-2019-00157	The production of citric acid process is via fermentation of a stable non-GMM <i>Aspergillus niger</i> strain	No	No
Plant protection products	Trichoderma atroviride AGR2	Plant protection product	EFSA-Q-2018-00476	Pesticide risk assessment and peer review of <i>Trichoderma atroviride</i> AGR2 in accordance with Article 12 of Regulation (EC) No 1107/2009 (application for approval)	No	No
Feed additives	Trichoderma longibrachiatum MUCL 49754 /Trichoderma longibrachiatum MUCL 49755	Zootechnical additives/Production of endo-1,4-beta-xylanase and endo- 1,3(4)-beta-glucanase	EFSA-Q-2018-00762		No	No
Feed additives	<i>Trichoderma reesei</i> CBS143953	Zootechnical additives/endo-1,4- beta-xylanase (EC 3.2.1.8) produced by GMM <i>Trichoderma</i> <i>reesei</i> , subtilisin protease (EC 3.4.21.62) produced by GMM <i>Bacillus subtilis</i> and alpha-amylase (EC 3.2.1.1) produced by GMM <i>Bacillus amyloliquefaciens</i>	EFSA-Q-2018-01040		No	No
Feed additives	<i>Trichoderma reesei</i> DSM 32338	Zootechnical additives/Production of muramidase	EFSA-Q-2018-00952		No	No

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EFSA risk assessment area	Microorganism species/strain	Intended use	EFSA Question number ^(a) and EFSA webpage link ^(b)	Additional information provided by the EFSA Scientific Unit	Previous QPS status of the respective TU? ^(C)	To be evaluated? yes or no ^(d)
Feed additives	<i>Trichoderma reesei</i> MUCL 49755	Zootechnical additives/Digestibility enhancers Used for the production of endo- 1,4-beta-xylanase (Bergazym [®] P 100)	EFSA-Q-2019-00097	The strain used for the production of the xylanase is <i>Trichoderma reesei</i> MUCL 49755		
Feed additives	<i>Trichoderma reesei</i> RF5427 strain	Zootechnical additives/Digestibility enhancers/Production of endo-1,4- beta-xylanase	EFSA-Q-2018-00824		No	No
Feed additives	<i>Trichoderma reesei</i> strain Morph-Y5#2 (CBS143953)	Zootechnical additives/Digestibility enhancers/Production of Xylanase by GMM <i>Trichoderma reesei</i>	EFSA-Q-2018-01039		No	No
Yeasts						
Feed additives	Komagataella pastoris appaT75 (CGMCC 12056) previously known as Pichia pastoris	Zootechnical feed additive/ Digestibility enhancer APSA PHYTAFEEDR 20,000 is a preparation of 6-phytase produced in two formulations. APSA PHYTAFEEDR 20,000 is produced by fermentation of a GMM yeast	EFSA-Q-2019-00188		Yes	No
Feed additives	Komagataella pastoris appaT75 (CGMCC 12056) previously known as Pichia pastoris	Zootechnical feed additive/ Digestibility enhancer APSA PHYTAFEEDR 20,000 is a preparation of 6-phytase produced in two formulations. APSA PHYTAFEEDR 20,000 is produced by fermentation of a GMM yeast	EFSA-Q-2019-00192		Yes	No
Feed additives	Pichia pastoris	Nutrase P (6-phytase) belongs to the category 4 'zootechnical additives' in its functional group 'digestibility enhancers'	EFSA-Q-2019-00155	The strain used for the production is a GMM <i>Pichia pastoris</i>	Yes	No



EFSA risk assessment area	Microorganism species/strain	Intended use	EFSA Question number ^(a) and EFSA webpage link ^(b)	Additional information provided by the EFSA Scientific Unit	Previous QPS status of the respective TU? ^(C)	To be evaluated? yes or no ^(d)
Feed additives	Saccharomyces cerevisiae	Nutritional additives/vitamins, pro- vitamins and chemically well- defined substances having a similar effect 25-hydroxycholecalciferol (25-OH-D3)	EFSA-Q-2019-00155	25-OH-D3 is obtained from the raw material 5,7,24-cholestatrienol ('trienol'). Trienol is produced by a fermentation process using a GMM <i>Saccharomyces</i> <i>cerevisiae</i> strain number SC0639		
Feed additives	<i>Saccharomyces</i> <i>cerevisiae</i> CBS 493.94	Zootechnical additives/Digestibility enhancers	EFSA-Q-2018-00893		Yes	No
Feed additives	<i>Saccharomyces</i> <i>cerevisiae</i> CNCM I-3399	Nutritional additives/Selenium- enriched yeast	EFSA-Q-2018-00908	Trace elements	Yes	No
Novel foods	<i>Yarrowia lipolytica</i> A-101	As a novel food production of yeast biomass/Chromium enriched	EFSA-Q-2018-00769	Application from same applicant as <i>Yarrowia lipolytica</i> biomass (see above)/Summary of application: NA Qualification: 'for production purpose only' implies the absence of viable cells of the production organism in the final product and can also be applied for food and feed products based on microbial biomass	Yes	No
Novel foods	<i>Yarrowia lipolytica</i> A-101	As a novel food production of yeast biomass/Selenium enriched	EFSA-Q-2018-00796	Application from same applicant as <i>Yarrowia lipolytica</i> biomass (See above)/Summary of application: NA Qualification: 'for production purpose only' implies the absence of viable cells of the production organism in the final product and can also be applied for food and feed products based on microbial biomass	Yes	No



EFSA risk assessment area	Microorganism species/strain	Intended use	EFSA Question number ^(a) and EFSA webpage link ^(b)	Additional information provided by the EFSA Scientific Unit	Previous QPS status of the respective TU? ^(C)	To be evaluated? yes or no ^(d)
Viruses						
Plant protection products	Spodoptera exigua multicapsid nucleopolyhedrovirus (SeMNPV)	Plant protection product	EFSA-Q-2018-00726	Pesticide risk assessment and peer review of <i>Spodoptera exigua multicapsid</i> <i>nucleopolyhedrovirus</i> (SeMNPV) in accordance with Article 12 of Regulation (EC) No 1107/2009 (application for approval)	Yes	No
Algae						
Novel foods	Euglena gracilis	As a novel food consisting on dried whole cells of <i>Euglena gracilis</i>	EFSA-Q-2019-00043	A minimally processed dried biomass of <i>E. gracilis</i> preparation, containing a minimum of 50% algae beta-glucan	No	Yes
Novel foods	Phaeodactylum tricornutum	PhaeoSOL is a novel food consisting on an extract of the microalgae <i>Phaeodactylum tricornutum</i> to be uses as a source of the naturally occurring carotenoid, fucoxanthin	EFSA-Q-2019-00091	PhaeoSOL consists of an ethanolic extraction of microalgae dried biomass	No	Yes

(a): To find more details on specific applications please access the EFSA website - Register of Questions: http://registerofquestions.efsa.europa.eu/roqFrontend/ListOfQuestionsNoLogin?0&panel= ALL

(b): Where no link is given this means that the risk assessment has not yet been published at the time of the publication of this Panel Statement.

(c): Included in the QPS list as adopted in December 2016 (EFSA BIOHAZ Panel, 2017) and respective updates which include new additions (latest: EFSA BIOHAZ Panel, 2018).

(d): In the current Panel Statement.