Orientation-Cue Invariant Population Responses to Contrast-Modulated and Phase-Reversed Contour Stimuli in Macaque V1 and V2



Xu An^{1,2}, Hongliang Gong¹, Jiapeng Yin¹, Xiaochun Wang¹, Yanxia Pan¹, Xian Zhang^{1,2}, Yiliang Lu¹, Yupeng Yang², Zoltan Toth³, Ingo Schiessl³, Niall McLoughlin³, Wei Wang^{1*}

1 Institute of Neuroscience, State Key Laboratory of Neuroscience and Key Laboratory of Primate Neurobiology, Shanghai Institutes for Biological Sciences, Chinese Academy of Sciences, Shanghai, P. R. China, **2** Key Laboratory of Brain Function and Diseases, School of Life Sciences, University of Science and Technology of China, Hefei, P. R. China, **3** Faculty of Life Science, University of Manchester, Manchester, United Kingdom

Abstract

Visual scenes can be readily decomposed into a variety of oriented components, the processing of which is vital for object segregation and recognition. In primate V1 and V2, most neurons have small spatio-temporal receptive fields responding selectively to oriented luminance contours (first order), while only a subgroup of neurons signal non-luminance defined contours (second order). So how is the orientation of second-order contours represented at the population level in macaque V1 and V2? Here we compared the population responses in macaque V1 and V2 to two types of second-order contour stimuli generated either by modulation of contrast or phase reversal with those to first-order contour stimuli. Using intrinsic signal optical imaging, we found that the orientation of second-order contour stimuli was represented invariantly in the orientation columns of both macaque V1 and V2. A physiologically constrained spatio-temporal energy model of V1 and V2 neuronal populations could reproduce all the recorded population responses. These findings suggest that, at the population level, the primate early visual system processes the orientation of second-order contours initially through a linear spatio-temporal filter mechanism. Our results of population responses to different second-order contour stimuli support the idea that the orientation maps in primate V1 and V2 can be described as a spatial-temporal energy map.

Citation: An X, Gong H, Yin J, Wang X, Pan Y, et al. (2014) Orientation-Cue Invariant Population Responses to Contrast-Modulated and Phase-Reversed Contour Stimuli in Macaque V1 and V2. PLoS ONE 9(9): e106753. doi:10.1371/journal.pone.0106753

Editor: Nicholas Seow Chiang Price, Monash University, Australia

Received April 7, 2014; Accepted August 1, 2014; Published September 4, 2014

This is an open-access article, free of all copyright, and may be freely reproduced, distributed, transmitted, modified, built upon, or otherwise used by anyone for any lawful purpose. The work is made available under the Creative Commons CC0 public domain dedication.

Data Availability: The authors confirm that all data underlying the findings are fully available without restriction. All relevant data are within the paper and its Supporting Information files.

Funding: This work was supported by National '973' Programs 2011CBA00400 and 2009CB941303. The funders had no role in study design, data collection and analysis, decision to publish, or preparation of the manuscript.

Competing Interests: The authors have declared that no competing interests exist.

* Email: w.wang@ion.ac.cn

Introduction

Visual perception arises from the transformation of neural signals along the visual hierarchy with neurons having different sizes of receptive fields (RFs) in each of its processing stages [1-2]. Humans and non-human primates can effortlessly see oriented contours or boundaries of objects, regardless of whether they are defined solely by a change in luminance (first order) or by contrast, texture, or other visual cues (second order). In contrast to a luminance defined first-order stimulus, all regions of a secondorder stimulus contain the same average luminance (Fig. 1). Second-order stimuli were initially manifested by second-order motion as globally drift-balanced stimuli [3-4] and so it has been suggested that there exists separate visual channels specifically to process such stimuli [5-10]. General speaking, second-order stimuli reveal the dissociation between retinal inputs (Fourier components, first order) and visual percepts (non-Fourier features, second order).

It has been known for more than a half century that most neurons in early visual cortices have small spatio-temporal oriented RFs with precise retinotopic coordinates, exhibiting orientation selectivity to luminance-defined contours [11-19]. Therefore, our hypothesis is that the population responses in early visual cortices might be directly activated by the local luminance cues that define the global second-order contours. Specifically, we ask how is the orientation of second-order contours processed at the population level in macaque V1 and V2. This is an important question not only pertaining to the processing of orientation regardless of its defining cues (known as orientation-cue invariance) but also to the subsequent invariant representation of shapes and forms observed in the middle temporal (MT) area and V4 [20-26]. Population responses to contrast-modulated contour stimuli were previously found to be orientation-cue invariant in cat area 18 and a non-linear "filter-rectify-filter" model was subsequently proposed to account for this observation [9]. Recently, it was reported that neurons responding to contrast-defined contours in cat area 18 [8,27] also encoded motion-defined second-order contours [28]. This is not the case in macaques as only a small number of cells in V1 and V2 were selective to the orientation of motion-defined contours [29-30]. A recent population study in macaque found that the preferences of population responses within V1 and V2 activated by illusory contour stimuli, that were



Figure 1. Synthetic first- and second-order contour stimuli. LG, sine-wave luminance gratings. CM, contrast modulated contours. PR, phase-reversal defined contours. Each column depicts one type of contour stimuli with an orientation of 90°. Arrows superimposed on each stimulus type in the top row represent the bidirectional motion of the global contours. The contours move leftward for 2 seconds and then rightward for another 2 seconds, as depicted below by the traces in the space-time plots. The square brackets and black arrows point to the second-order contours. doi:10.1371/journal.pone.0106753.q001

defined by abutting lines, depended critically on the spatial frequency of the local carriers [25]. These results are compatible with a recent single-cell electrophysiological study, which demonstrated that most neurons in macaque V1 and V2 signal the orientation of first-order carriers within texture-defined herringbone patterns [31]. It appears that only a small number of neurons in the early visual cortices of non-human primate exhibit clear responses to second-order stimuli [23,29,32–37]. Thus, in this study we specifically investigated whether and how the orientation of second-order contours defined by contrast modulation and phase reversal is encoded by population responses in macaque V1 and V2.

We measured the cortical population responses to sine-wave luminance gratings (LG, first order), to contrast-modulated (CM) second-order contours, as well as to another set of second-order contours defined by phase reversal (PR) (Fig. 1), using intrinsic signal optical imaging of macaque V1 and V2. We found that contrast- and phase-defined contours both activated orientation domains with preferences in close register with those activated by first-order luminance-defined contours in V1 and V2, exhibiting orientation-cue invariance. In addition a physiologically constrained spatio-temporal energy model of V1 and V2 neuronal population responses was able to account for all the population responses recorded. Our experimental findings and energy model simulations suggest that the population responses in primate V1 and V2 reflect the spatio-temporal filter properties of orientationselective neurons, and hence the orientation maps in the primate can also be described as a spatio-temporal energy map [38–41].

Materials and Methods

Ethics Statement

The experimental subjects were rhesus macaques (Macaca mulatta) and came from our institutional non-human primate breeding colony under licenses issued by National Forestry Ministry - Shanghai Bureau and Shanghai Animal Management Office. The animals were housed without cage in strict secure rooms with various environmental enrichment items, secure windows, and a balcony access to daylight. Specifically, the environmental enrichment items mainly included a large mirror on the wall, a swing, a rope for climbing, ladders, platforms, and an outdoor balcony for the animals to sunbathe and have outdoor activities (Fig. S1). The housing, husbandry, and breeding standards comply with National Laboratory Animal - Requirements of environment and housing facilities (GB: 14925-2010). All experimental procedures for primate research including animal euthanasia were approved by the Institute of Neuroscience Institutional Animal Care and Use Committee and by the local ethical review committee of the Shanghai Institutes for Biological Sciences.

Animal surgical preparation and maintenance for optical imaging

A total of six adult male rhesus macaques each weighing $3.0 \sim 4.5$ kg were prepared and maintained for acute *in vivo* intrinsic signal optical imaging as described elsewhere [42–44]. In brief in each experiment, anesthetic induction was achieved by intra-muscular injection of ketamine hydrochloride (15 mg·kg⁻¹, i.m.). After a tracheotomy, all surgical procedures were carried out under gaseous anesthesia (Isoflurane $0.5 \sim 2\%$ in 2:1 N₂O:O₂).

General anesthesia was maintained by bolus injections of Pentothal (sodium thiopental $\sim 2 \text{ mg/kg/h IV}$) administered by IV catheter. Depth of anesthesia was verified with respect to an electrocardiogram (ECG), pulse oximeter (SpO₂), and end-tidal carbon dioxide (CO_2) , all of which were monitored continuously. A maintenance solution, of 5% glucose in saline, was administered by constant infusion through an intravenous catheter $(3 \sim 5 \text{ cm}^3/$ kg/h), whilst a thermister controlled electric blanket maintained the animal's core temperature at around 38°C. The animal's eyes were first washed with dH₂O before the application of Atropine drops (1% solution) and the insertion of Plano hard gas-permeable contact lenses. Refractive errors were estimated using a slit ophthalmoscope and when necessary corrected using external lenses such that the animal focused on a screen placed 57 cm from the centre of its head. Craniotomy and durotomy were performed on both sides of the skull over V1 and V2 for dual optical imaging using two stainless steel chambers of 25 mm diameter secured to skull using dental cement. The lunate sulcus (LS) and superior temporal sulcus (STS) were used as cortical landmarks for surgeries. At the end of each experiment, euthanasia was achieved by the administration of a lethal IV injection of sodium pentobarbitone (50 mg). After death, the animal was immediately perfused transcardially with a saline rinse followed by ice-cold 1% paraformaldehyde in 0.1 M potassium phosphate buffer. The brain was then removed and the imaged visual cortices were flattened and sectioned for subsequent histological processing.

Visual stimuli

A CRT monitor (Sony Trinitron Multiscan G520, 1280×960 pixels, 100 Hz) was placed 57 cm in front of the animal eyes extending 40×30 degrees as described elsewhere [25,42]. The gamma of the monitor was carefully corrected by using the Color Calibration device (ColorCAL) from Cambridge VS system. Visual stimuli were computer-generated using custom software based on Psychtoolbox-3. For the contrast-modulated noise stimuli, the individual noise size of the noise carrier used in previous psychophysics and fMRI studies was between 1.81-13 arcmin [10,45-54]. In this study, full screen noise textures were composed of noise elements randomly positioned with each element spanning approximately 5.4 arcmin. In some cases, we tested the noise size of 1.8 arcmin that corresponded to one pixel size on the CRT monitor, but we did not observe clear responses. We enlarged the noise size to 5.4 arcmin which activated reliable population responses across animals; therefore, we used this size in all cases. The luminance of the elements ranged from $0.2 \sim 82 \text{ cd} \cdot \text{m}^{-2}$ following a uniform distribution. All stimuli employed had a mean luminance of approximately 45 cd \cdot m⁻². As illustrated in Figure 1, standard sine-wave luminance gratings (LG) were defined by the sequence of peaks and troughs where the contrast was defined as (Lmax-Lmin)/(Lmax+Lmin). Lmax and Lmin are the maximum and minimum luminance level of the stimuli, respectively. LG is regarded as a first-order stimulus (equation 1 and Movie S1). By comparison, the contrastmodulated grating was composed of noise texture whose pixels were modulated in contrast by a traveling sinusoid (equation 2 and Movie S2) and is called a second-order contrast-modulated (CM) stimulus [3,8-9,53,55-57]. The appearance of CM stimulus is comparable to a traveling wave on the surface of water. In the texture phase reversed (PR) stimuli (equation 3 and Movie S3), second-order contours were generated from spatial locations where noise pixels had their polarity flipped simultaneously between frames. There is no change in either luminance or contrast in this stimulus type. Only the leading and trailing edges of the second-order contours are visible in the space-time plot (Fig. 1). The mean luminance in both first- and second-order stimuli was kept the same. The first- and second-order contours described above drifted back and forth perpendicular to their orientations for a total of 4 seconds, with 2 seconds for each moving direction, respectively (Fig. 1). The phase-reversal contours are dynamic second-order contours and disappear as soon as the stimuli stop moving. The top row in Figure 1 displays a single spatial frame (x, y) from each stimulus type with a space-time plot (x, t) below to aid the visualization of the movement.

For simplicity, we first define a two-dimensional moving sine wave as: $S(x,y,t) = \sin(2\pi f(\sin(\theta)x + \cos(\theta)y) + 2\pi vt))$, where f is the spatial frequency, θ is the orientation, and v is the temporal frequency. Then the first- and second-order stimuli can be expressed as:

$$LG(x,y,t) = l_0 + l_0 c_{LG} S_{LG}(x,y,t)$$
(1)

$$CM(x,y,t) = l_0 + l_0 \frac{mS_{CM}(x,y,t) + 1}{2} N(x,y)$$
(2)

$$PR(x,y,t) = l_0 + l_0 \operatorname{sgn}(S_{PR}(x,y,t))N(x,y)$$
(3)

Where N(x,y) is the uniform distribution $[-1 \ 1]$ for two dimensional noise texture, l_0 is the mean luminance (45 cd·m⁻² in the experiment), contrast c_{LG} is set to 1, modulation depth *m* is set to 1, for LG, PR, and CM, the spatial frequency *f* and temporal frequency v were in the range of 1.0~1.5 cycles per degree (cpd) and 4~6 Hz.

Optical imaging

Details of all the equipment and recording procedures of our custom built optical imaging system were as described elsewhere [25,42,58]. In brief, visual responses were recorded at sixteen frames per second for a period of 8 s, including 1 s prior to the stimulus onset under 630 ± 10 nm red light illumination. The inter-stimulus interval was 13 s. Data were collected in an interleaved fashion for first- and second-order stimuli with different orientations. For first-order stimuli, data were typically averaged over 32 or 64 trials, while for second-order stimuli, the data were often averaged over 256 trials. The boundary of V1 and V2 was classically defined using either retinotopic space mapping or ocular dominance mapping [59].

Optical image analysis

For each trial, frames taken between 3 and 7 s after the stimulus onset were averaged, and then subtracted and divided by a blank frame (the average response from the 1 s interval prior to the stimulus onset) to generate a map of reflectance change ($\Delta R/R$ map). Differential orientation maps were then created via pixel-bypixel subtraction of reflectance maps generated by a pair of stimuli with orthogonal orientations (e.g. $0^{\circ}-90^{\circ}$). Orientation preference maps were constructed using a vector summation algorithm [25,42,60]. We adopted a published method for removing pixels with large variability (e.g. those from blood vessels) and a mask was generated based on an objectively chosen threshold [9,25,42,58]. Pixels covered by the mask were interpolated from surrounding un-occluded values just for display purposes but were never used in quantitative analysis. The interpolated images were then highpass filtered (1.1~1.2 mm in diameter) and smoothed $(85 \sim 323 \ \mu m$ in diameter) by circular averaging filters when necessary to suppress low and high frequency noise while avoiding signal distortion. Fractures of orientation preference map for LG were derived following Bonhoeffer and Grinvald (1993) [61], based upon a map of the magnitude of orientation gradient:

$$\begin{aligned} |\nabla I(x,y)| &= \\ \sqrt{\left[I(x+1,y) - I(x-1,y)\right]^2 + \left[I(x,y+1) - I(x,y-1)\right]^2} \end{aligned} \tag{4}$$

where I is the orientation angle value and (x,y) the pixel coordinates. Pinwheel centers of orientation preference map for LG were identified using a method similar to that of Crair et al. (1997) [62]: a pixel is considered to be a pinwheel center if the sum of orientation differences (wrapped into $-90 \sim 90^{\circ}$) between its four counterclockwise neighbors is $\pm 180^{\circ}$. Previous and our current studies all found that pixels in pinwheel centers and fractures with rapid change of orientation preference were noisesensitive and contributed disproportionately to the measurement variance in orientation preference maps [9,63-65]. The pinwheel center pattern of an orientation preference map was dilated with a 5 pixel radius disk; its union with the fracture pattern (gradient of orientation change $>15^{\circ}$ /pixel) was used to form a binary mask to remove these noise-sensitive pixels from the comparison of different orientation preference maps [9]. Note that keeping these pixels in the calculation did not change the overall signature of the results. Using orientation preference maps obtained in an experiment with CM and LG stimuli as an example, the angular differences of orientation preference maps between first- and second-order contour stimuli that were less than 30° were 81% in both V1 and V2 if these pixels were kept in the calculation, while only slightly increased to 82% and 84%, respectively, in V1 and V2 after removing these pixels. For the quantification of the response amplitude, the max $\Delta R/R$ values in each responsive patch of a differential map were averaged, and this average intensity was taken as the response amplitude. A response profile analysis was performed in order to extract the orientation best represented by differential orientation maps [9,25,40,42,58,66]. In brief, the signs of the $\Delta R/R$ values in the differential map were reversed, so that pixels responsive to the first condition of a trial had positive values while pixels responsive to the second condition exhibited negative values. The mean of all pixels was subtracted from each pixel thereafter. This $\Delta R/R$ map was then divided into 12 iso-orientation domains $(0^{\circ}-180^{\circ})$ based on the orientation preference map produced by LG stimuli, and the mean value of each domain was plotted as a function of its corresponding orientation. In addition, we adopted the spatial correlation coefficient (SCC) metric as an index of similarity between two differential maps that were evoked by first- and second-order stimuli (see Ramsden et al., 2001 for details). The value of SCC ranged between -1 and 1, with 1 indicating two identical maps while -1 indicating two identical but inverted maps. As in previous studies [67-68], coefficients within the range between -0.2 and 0.2 were deemed neither significantly overlapped nor segregated.

Visual stimulus analysis

We analyzed the spectral power distributions of the first- and second-order stimuli in the frequency domain as follows:

$$P(\omega_x, \omega_y, \tau) = |F(S(x, y, t))|^2$$
(5)

where S(x,y,t) is the stimulus sequence, F represents the Fourier transform (matlab function ff(tn), ω_x and ω_y are the spatial frequency coordinates corresponding to the x and y directions

respectively, τ is the temporal frequency coordinate. To reveal the power distribution difference between the 90° and 0° oriented stimuli in the two-dimensional spatial-frequency space, the power distribution difference was integrated over all temporal frequencies:

$$D(\omega_x, \omega_y) = \int P_{90}(\omega_x, \omega_y, \tau) - P_0(\omega_x, \omega_y, \tau) d\tau$$
(6)

the coordinate system of which can be converted to $\tilde{D}(\rho,\theta)$, where ρ and θ represent spatial frequency and orientation respectively. To show the differential power distribution in the individual spatial frequency, orientation, or temporal frequency dimension, the differential power was further integrated as follows:

$$S(\rho) = \int |\tilde{D}(\rho,\theta)| d\theta \tag{7}$$

$$O(\theta) = \int \tilde{D}(\rho, \theta) d\rho \tag{8}$$

$$T(\tau) = \iint |P_{90}(\omega_x, \omega_y, \tau) - P_0(\omega_x, \omega_y, \tau)| d\omega_x d\omega_y \tag{9}$$

Model simulations

We used the same model structure as that in our previous studies [42,58], which was originally described by Mante and Carandini [41,69]. Essentially, neuronal populations of both V1 and V2 were modeled as a bank of spatio-temporal filters and the average response of a set of neurons having the same tuning properties (orientation, spatial, and temporal preference) was given by integrating the energy of the stimulus falling into the receptive field (RF) of the population. To calculate the population responses of V1 and V2, here we further assumed that V1 and V2 are composed of neurons having a mixture of different tuning properties based on experimental observations [12–13,70]. The spatial filtering property of neurons was modeled as a twodimensional Gaussian function in frequency space:

$$S(p_{x},p_{y}) = \exp\left(-\frac{[p_{x}-S_{p}\sin(\phi)]^{2}+[p_{y}-S_{p}\cos(\phi)]^{2}}{2\sigma_{S}^{2}}\right)/(2\pi\sigma_{S}^{2})$$
(10)

where p_x and p_y are the spatial frequency coordinates. S_p and σ_S are the preferred spatial frequency and spatial bandwidth. ϕ is the preferred orientation. Similarly, the temporal filtering property was modeled as a Gaussian function as follows:

$$T(\rho) = \exp\left(-\frac{(\rho - T_{\rho})^2}{2\sigma_T^2}\right) / (\sqrt{2\pi}\sigma_T)$$
(11)

in which ρ is the temporal frequency coordinate. T_{ρ} and σ_T are the preferred temporal frequency and temporal bandwidth. Response of neuronal population with the same spatial frequency, temporal frequency, and orientation preferences was modeled as the integral over all spatio-temporal frequencies of the stimulus, scaled by the spatial and temporal filter functions:

$$R = \int_0^\infty \int_0^\infty \int_0^\infty \mu_S \mu_T A(p_x, p_y, \rho) S(p_x, p_y) T(\rho) dp_x dp_y d\rho \qquad (12)$$

where R and A are the response and Fourier transformed stimulus, respectively. μ_S and μ_T are two weights, corresponding to the proportion of neurons preferring the specific spatial and temporal frequencies respectively. In the simulations, preferred orientations (24 values) were uniformly distributed over 180°. Spatial frequencies (14 values) were logarithmically spaced between 0.13 and 11 cpd with a mean of 2.2 cpd for V1 [13] and 1.4 cpd for V2 [12], following Gaussian distributions which decided the weights $\mu_{\rm S}$. Temporal frequencies (16 values) were again logarithmically spaced between 0.25 to 45 Hz with a mean of 3.7 Hz for V1 and 3.5 Hz for V2 [13], following Gaussian distributions which decided the weights μ_{T} . The spatial frequency and temporal frequency bandwidths were scaled to be 1/3 of the preferred spatial and temporal frequencies, as in Mante and Carandini [41]. For the simulation results, we averaged 256 trials as displayed on the monitor during experiments. We only analyzed a 15 by 15 degree sub-region of the full screen to reduce computation time, but the results were well approximated by this reduction. As in the optical imaging data, responses to two stimuli with orthogonal contour orientations were subtracted.

Results

We initially used second-order contours defined by carriers with small noise size corresponding to one pixel on the CRT monitor (about 1.8 arcmin). Perceptually the second-order contours can be clearly seen, however, we did not observe reliable and clear population responses to these stimuli in our optical imaging experiments of both V1 and V2. This may be because the SF components of these stimuli largely exceed the responsive range of the recorded areas. However all our second-order contour stimuli defined by noise size up to 5.4 arcmin elicited weak, but clear population responses in the recorded cortical regions of both V1 and V2 (Fig. 2).

Orientation-invariant population responses activated by second-order contour stimuli

We generated differential maps of orientation preference within a region of interest (ROI) (Fig. 2A, B), by subtracting the intrinsic optical signals evoked by alternating full-field contour stimuli that had orthogonal orientations (0° and 90°). Dark regions prefer the first stimulus condition (0°) and bright regions prefer the second (90°). Within the same ROI of both V1 and V2, all contour stimuli activated orientation domains (Fig. 2B). However, responses to first-order stimuli were much stronger than those to second-order stimuli (Fig. 2C). Interestingly, all the stimuli evoked stronger responses in V2 than in V1. The response to luminance grating (LG) stimuli $(1.40\pm0.04\times10^{-4} \text{ in V1}; 2.32\pm0.16\times10^{-4} \text{ in V2})$ was nearly six times greater than that evoked by contrastmodulated (CM) stimuli $(2.53\pm0.09\times10^{-5})$ in V1: $4.00\pm0.27\times10^{-5}$ in V2) and twice as large as that to phasereversal (PR) stimuli $(6.38 \pm 0.30 \times 10^{-5} \text{ in V1}; 7.97 \pm 0.52 \times 10^{-5} \text{ in}$ V2) (Fig. 2C). The standard error (SEM) was computed for the response from each black and white patch in the differential maps. For a direct comparison of the population responses activated by different stimuli, we presented the maps based on the gray-scale range of the population responses activated by CM (Fig. 2D). All comparisons were made between signals recorded from the same locations in V1 and V2. We performed response profile analysis [9,25,40,42,58] to determine the orientation preference of the functional domains activated by each of the oriented contour stimuli (Fig. 2E). As expected, we found that the response profiles produced by first-order contours closely matched the preferences of the underlying orientation columns in both V1 and V2. Interestingly, the functional domains activated by CM and PR stimuli were in close register with those evoked by the first-order contour stimuli of the same orientations. Data obtained with the same CM and PR stimuli yielded similar results in all animals studied (See Figs. 3 and 4 for further examples). By using a spatial correlation coefficient (SCC) index [25,68], we also calculated the similarity between the differential orientation maps generated by LG and by CM and PR stimuli in V1 and V2 across all animals studied (Fig. 5A). The mean and median SCC values for CM and PR stimuli in both V1 and V2 were above or very close to 0.2, indicating significant overlap. The SCC value was just below 0.2 for the PR stimuli in V1. In addition, the SCC values for these two types of second-order stimuli in V2 were significantly higher than those in V1 (F(1, 16) = 13.22, p < 0.01, two-way ANOVA),suggesting that V2 exhibits a higher degree of cue invariance than V1. Together, these results demonstrate that different types of second-order contour stimuli elicit clear population responses within orientation columns of both V1 and V2 in macaque. The orientation preferences of the population responses evoked by contrast-modulated and phase-reversed contour stimuli are closely matched to those activated by luminance gratings.

Differences in response strength between first- and second-order contour stimuli

Figure 5B summarizes our findings on the average strength of responses induced by first and second-order contour stimuli across all six macaques studied. The relative change in the amount of light reflected $(\Delta R/R)$ from the ROIs in both V1 and V2 was measured and averaged for each stimulus type. We found that all the population responses to second-order contour stimuli (CM and PR) were significantly weaker than those to first-order LG stimuli in both V1 and V2 (F(2, 73) = 40.52, p < 0.01, two-way ANOVA). This is most likely due to the fact that in contrast to sine-wave gratings the noise texture has a broadband power spectrum and as such contains energy at many different orientations. The contrastmodulated stimuli elicited the weakest responses, but were not significantly different from the phase-reversed stimuli (p > 0.05, two-way ANOVA, post hoc Bonferroni test). In addition, the response to the first-order stimulus was significantly stronger in V2 than in V1 (p < 0.05, two-way ANOVA, post hoc Bonferroni test). While the response strength to the second-order stimuli was also slightly higher in V2 than that in V1, this was not significant (p >0.05, two-way ANOVA, post hoc Bonferroni test). Altogether, these results imply that a similar mechanism may underlie the responses to these first- and second-order contour stimuli in both V1 and V2.

Spatio-temporal energy model can account for the population responses recorded

Visual perception to global visual features or texture patterns is often modeled using a combination of linear and non-linear mechanisms [71–74]. The "filter-rectify-filter" (FRF) model [75], that was used to account for second-order motion perception, was also proposed to be responsible for neural responses to the orientation of contrast modulated second-order contours in cat area 18 [8–9]. However, the neural substrates corresponding to the different stages of the filter-rectify-filter model are still unclear [76].



Figure 2. Orientation preference domains were activated by different first- and second-order contours in macaque V1 and V2. (A), Picture of the cortical surface taken from the left hemisphere of macaque 766 with region of interest (ROI) indicated by red box. This image was obtained under 550 nm green-light illumination. The broken white line indicates the border between V1 and V2. LS, lunate sulcus. L, lateral. A, anterior. (B), Differential orientation maps of 0° minus 90° in V1 and V2. Blood vessels were masked gray on all the maps. White boxes represent regions of V1 and V2 that were further analyzed and compared. (C), Response strength comparison for first- and second-order stimuli from ROIs of V1 and V2 in B. (D), Representative areas of V1 and V2 as boxed in B. Pixels covered by blood-vessel masks as shown in B were interpolated for clarity. Iso-orientation preferences as indicated by the color code below figure. (E), Normalized orientation response profiles for first- and second-order contours indicate from V1 and V2 areas in D. Error bars represent S.E.M. Scale bar: 1 mm. doi:10.1371/journal.pone.0106753.g002

In fact, the majority, if not all, of second-order stimuli contain both, global second-order features and local first-order luminance changes [9,25,31,41,55–56,77–78]. For example, the PR and CM stimuli actually contain local luminance changes, as the luminance of pixels that define the boundary in the PR stimulus varies locally, so does the luminance of pixels that define the CM boundary (from black or white to mean grey). We thus examined whether a linear filter model could simulate our experimental results rather than the more complex "filter-rectify-filter" model suggested by an early study [9]. To do this we chose to implement a well-known spatio-temporal energy based model [79]. This spatio-temporal energy model has previously been employed to capture many response properties of early visual cortex at the single-cell level [80–83]. It has also been successfully applied to model the population responses of ferret visual area 17 elicited by different combinations of local visual features [38,41]. More recently it has reproduced the population activities induced by motion axes of moving noise and dots in macaque V1 and V2 [42,84] and in cat visual areas 17 and 18 [58]. The model of population responses is composed of spatial frequency, temporal frequency, and orientation filters with all parameters acquired from single-unit studies in macaque V1 and V2 [12–13,70] (Fig. 6A).



Figure 3. Orientation preference domains in V1 and V2 activated by phase-reversed contours. (A), Picture of the surface vasculature with the region of interest (ROI) in V1 and V2 as outlined by the red box. LS: lunate sulcus. A, anterior. L, lateral. (B), Differential orientation maps of V1 and V2 derived from LG and PR stimuli with orthogonal orientation pairs of 45° and 135°. Blood vessels were masked gray on all the maps. (C), Differential orientation maps from the representative areas of V1 and V2 (white boxes in B). Both pairs of the grayscale images were displayed based on the intensity range of the PR map and were superimposed with iso-orientation outlines derived from the orientation map generated by LG stimuli. (D), Normalized response profiles for LG and PR stimuli. The two pairs of curves were closely matched in the orientation preference for both V1 and V2. Scale bar: 1 mm. doi:10.1371/journal.pone.0106753.q003

We first simulated response strength to two orthogonal orientations (horizontal and vertical contours). Responses were derived from simulated population neurons of V1 and V2. The model produced weak responses to second-order contour stimuli (Fig. 6B) and the relative response strengths for all three types of stimuli were preserved (compare Figs. 5B and 6B). The ratios of response strength between LG and the rest of the stimuli were not perfectly matched between the model simulations and the optical imaging experiments. Two possible reasons may account for this

discrepancy. First, the energy model did not take into consideration of a potential non-linear gain control mechanism within the circuits of orientation columns in V1 and V2 for the processing of second-order contours with low saliency. Secondly, a subgroup of specialized neurons within V1 and V2, which was previously reported to respond to second-order stimuli, contributed to the recorded population responses. Obviously this part of population response cannot be captured by a simple linear model. Nevertheless, the energy model predicted almost identical orientation



Figure 4. Orientation differential and preference maps in V1 and V2 for contrast modulated contour stimuli. (A), Picture of the surface vasculature of the left hemisphere of macaque 740 with the region of interest (ROI) in V1 and V2 outlined in red. Diagrams of the 0° and 90° oriented LG and CM stimuli were shown on top. (B), Differential orientation maps of V1 and V2 derived from LG and CM stimuli with orthogonal orientation pairs of 0°–90° and 45°–135°. Blood vessels were masked gray on all the maps. (C), Color coded orientation preference maps generated by LG and CM stimuli with blood vessels masked gray. (D), Orientation preference maps from representative areas of V1 and V2 as boxed in C. (E), Histograms produced by pixel-wise subtraction of the two pairs of orientation preferences less than 30° were 82% and 84% in V1 and V2, respectively. Note that orientation ranges from 0° to 180°, so the difference between two orientation values will be in the range of -180° to 180° by direct subtraction. Scale bar: 1 mm.

doi:10.1371/journal.pone.0106753.g004

response profiles for both first- and second-order (CM and PR) contour stimuli in V1 and V2 (Fig. 6C). It is known that dynamic and high-pass filtered noise textures can reduce the interference of first-order information in the processing of second-order features [6]. Besides static noise carrier, some psychophysical and fMRI studies also employed dynamic and filtered noise carriers [10,47,49,52–54,85–86]. Therefore, we also simulated the response profiles to CM and PR stimuli with dynamic and high-pass filtered noise carriers. The high-pass filtering of noise carriers eliminated components with SFs below 9 cpd that exceeded the

optimal SF range of V1 and V2 neurons reported previously [13]. CM and PR stimuli formed by both dynamic and high-pass filtered noise carriers also evoked orientation cue-invariant population response in both V1 and V2 (Fig. 6D). This is consistent with the results obtained using static noise carriers.

A previous computational study has demonstrated that the analysis of the spectral power distributions in the frequency domain can reveal essential properties of second-order stimuli [78]. Our model results also can be intuitively understood through analyzing the first- and second-order stimuli in the frequency



Figure 5. Spatial correlation coefficients (SCC) and response strengths across all animals studied. (A), Spatial correlation coefficients for both V1 and V2 between the first-order stimulus and the two second-order stimuli. SCC represents spatial similarity between the orientation differential maps activated by LG stimuli and those by CM and PR stimuli. The numbers of orientation differential maps computed were 6 and 4 for CM and PR stimuli respectively in both V1 and V2. The small solid black squares represent mean values while the red lines represent median values. (B), Comparison of population response strengths (Δ R/R) in V1 and V2. Error bars represent S.E.M. Data from 6 monkeys. doi:10.1371/journal.pone.0106753.g005

space (Fig. 7; see Materials and methods for details of the analysis). We firstly computed the power distribution difference between a pair of contour stimuli with orthogonal orientations (here using 90° and 0° orientations as examples) (Fig. 7A, D, and G). Then the distribution of the differential power in orientation, spatial frequency, and temporal frequency dimensions was analyzed. For the stimulus pair of luminance gratings, as expected, the differential power had a sharp peak and trough at 90° and 0°, respectively, in the orientation dimension (Fig. 7B, C). There were single peaks at ~1.5 cpd and ~5 Hz in the spatial and temporal frequency dimensions, respectively, corresponding to the spatial and temporal frequencies of the sine-wave gratings (Fig. 7C).

We further examined how neuronal populations with different preferences to orientation, spatial, and temporal frequencies respond to the pair of 90° and 0° luminance gratings, and the results for V1 and V2 were presented in Figures 8A and 9A. The maximal differential population responses $(0^{\circ}-90^{\circ})$ occurred in the spatio-temporal frequency range corresponding to the stimulus parameters used for the pair of luminance gratings (Figs. 8A and 9A), demonstrating that model neurons are capable of reproducing the population responses activated by luminance gratings in both V1 and V2. In the case of the CM stimuli as the sinusoidal contrast modulation of a noise texture, this modulation causes a collinear distortion or inhomogeneous distribution in the local luminance of the broadband noise texture along the orientation of the contours (i.e. serrated edges). Thus the initial isotropic-noise structures within the noise texture tend to appear after modulation as oval shapes parallel to the contour orientation. This induced orientation signal can be clearly seen in the frequency space, as the differential power of the CM stimulus pair had a peak and trough at 90° and 0° respectively in the orientation dimension (Fig. 7E, F). Due to the broadband noise-texture carrier used, the differential power also existed in other orientations and distributed in a broad range of spatial frequencies (Fig. 7F). There was a sharp peak at \sim 5 Hz in the temporal frequency dimension, corresponding to the temporal frequency of the stimuli (Fig. 7F). Thereby, the slight induced orientation signal is detected by the corresponding simulated neuronal populations, resulting in weak but consistent differential population responses selective to the orientation of the CM stimuli (Figs. 8B and 9B). Analogously, the response to the PR stimulus can be similarly explained by the fact that the PR stimulus is related to the CM stimulus, because it is in principle a noise texture modulated by a square wave (compare equations 2 and 3). Thus, also in this case, the modulation will introduce an anisotropic oriented component that was revealed by the analysis in the frequency domain (Fig. 7H, I). Noting that in comparison with the CM stimulus pair, the differential power of the PR stimulus pair spread to higher spatial and temporal frequencies (Fig. 7I) and this is caused by the square wave modulation. Similar to the CM stimuli, the power of the PR stimuli can be detected by the corresponding neuronal populations, leading to the orientation-cue invariant model responses (Figs. 8C and 9C). These results are comparable with recent studies on contrast-modulated second-order motion [46,87]. In these studies, it was found that the texture contribution to second-order motion perception was mediated solely by the residual distortion products through the first-order mechanism and the non-existence of a separate secondorder motion channel was consequently suggested. We further simulated the population responses to CM and PR stimuli with small noise size of 1.8 arcmin (corresponding to one pixel size on the CRT monitor) for the carrier. However, this condition generated the weakest response strength and no reliable and robust response profiles could be obtained, consistent with our experimental observations using optical imaging.

Discussion

First- and second-order cues may occur spatially and temporally next to each other in natural scenes (often seen as texture) [8,73]. The presence of local luminance fluctuations has been previously recognized within the synthetic contrast-modulated second-order stimulus [6,55–56,77]. This so-called luminance artifact cannot be completely avoided but only reduced. However, the second-order motion stimulus as a whole is drift-balanced without mean luminance changes [3] and the contrast modulated noise stimulus remains an important tool to study the processing of non-Fourier features [53,55,87–88]. Thus, noise texture has been widely used in human psychophysics and fMRI studies including those for making second-order motion stimuli. Similar to luminance cues in natural scenes or images, local contrast information has been



Figure 6. Energy model simulations of population responses to all stimulus types across V1 and V2. (A), Illustration of the receptive field of the modeled neuronal populations in the spatio-temporal frequency space. RFs with different tuning properties are assumed to be Gabor wavelets of different shapes, resulting in Gaussians in the frequency space, whose elliptic outlines are displayed. RFs illustrated in the same color correspond to a same orientation preference lying on radial sections in the frequency space (e.g. red: horizontal orientation; light blue vertical orientation). (B), Population response magnitudes produced by the energy model to different types of stimuli. (C), Normalized response profiles from V1 and V2 simulated by the energy model to the first- and second-order contour stimuli $(0^{\circ}-90^{\circ})$. As for the optical imaging experiment, the noise-texture

carriers used for the simulation were static and were not filtered. (D), Normalized response profiles simulated by using dynamic and high-pass filtered (eliminate components with SFs below 9 cpd) noise textures as the carrier for CM and PR stimuli ($0^{\circ}-90^{\circ}$). The error bars indicate S.E.M over 256 trials and are smaller than the data marker. doi:10.1371/journal.pone.0106753.q006

suggested as an independent variable encoded by the early visual system [89–95]. Our findings suggest that it is these local firstorder visual cues and related variations, which are collinearly distributed along the second-order contours, that drive the orientation cue-invariant population responses in macaque V1 and V2. This is also reflected in the observation that the population responses, activated by these second-order contour stimuli, are quite consistent between V1 and V2. This is because that macaque V2 not only receives its major feed-forward projections from V1 [96–97], but also responds to a similar range of spatial and temporal frequencies [12–13].

The spatio-temporal oriented filter mechanism underlying population responses

In a natural setting, second-order contours are defined by a broad range of physical cues or components. In the laboratory, synthetic second-order contours can be defined by all sorts of first-order carriers regardless of their spatial components. To the second-order contour stimuli used in this study, a spatio-temporal energy model predicted all the orientation-selective responses (Figs. 6 and 8–9). These model results strongly suggest that it is the

linear filter property of the small oriented spatio-temporal RFs of most V1 and V2 neurons that underlies the population responses. These population responses are consistent with an earlier observation in cat area 18, in which orientation-cue invariant population responses to contrast-modulated contour stimuli were also found [9]. By contrast, the population responses in cat area 18 were regarded as the responses to global second-order contours and a non-linear "filter-rectify-filter" model was subsequently proposed [9]. The main argument in this previous study was based on the hypothesis that the SFs of the carriers were above the cutoff SF of area 18; therefore, the population responses in the recorded region of area 18 should represent the non-linear neural responses to second-order contours. However, a recent study using electrophysiological single-unit recording of cat area 18 and monkey V1 and V2 found very few neurons responding to pure second-order visual features [31]. In fact, the responses of most neurons greatly decreased when the spatial frequency of the carriers was set beyond the sensitive range of the neurons. In the current study of macaque V1 and V2, we also did not observe reliable population responses when the individual noise element of



Figure 7. Distribution of differential power between stimuli with orthogonal orientations in the frequency space. (A, D, G), Diagrams of pairs of stimuli with orthogonal orientations of 90° and 0°. White arrows indicate the bidirectional motion of the stimuli. The stimuli moved 0.51 second in each direction to reduce the computational time. We then used a Fourier transform (matlab function 'fftn') to compute the power of each stimulus. (B, E, H), Power distribution difference $(90^\circ - 0^\circ)$ in the two dimensional spatial frequency space corresponding to each stimulus pair. Note that to reduce dimensions, the power was integrated over all temporal frequencies. 32 pairs of the CM and PR stimuli were used for the computation. (C, F, I), Differential power distributions in the orientation, spatial frequency, and temporal frequency dimensions for each stimulus pair, respectively. The gray shades in the left panels of F and I represent SEM over 32 pairs of stimuli.



Figure 8. Simulated differential population responses with different preferences in V1. (A–C), Details of differential responses of neuronal populations preferring different orientations, spatial, and temporal frequencies to each stimulus pair (0°–90°). N = 10 trials. doi:10.1371/journal.pone.0106753.g008

the noise carrier was set to 1.8 arcmin corresponding to high carrier spatial frequency.

It is also important to note that the stimuli were not presented exclusively within a restricted visual field corresponding to the recorded cortical region, but instead presented across the full screen that activated a large section of the retina including central vision. Thus, another interesting question arises as whether neurons having RFs at different eccentricities within different regions of V1 and V2 employ different mechanisms for the processing of the same second-order contour stimulus. Specifically, do neurons in the region corresponding to the central visual field of high spatial resolution utilize linear mechanisms, while neurons representing peripheral space, with low spatial resolution utilize



Figure 9. Simulated differential population responses with different preferences in V2. (A–C), Details of differential responses of neuronal populations preferring different orientations, spatial, and temporal frequencies to each stimulus pair (0°–90°). N = 10 trials. doi:10.1371/journal.pone.0106753.q009

non-linear mechanisms? Together, it is conceivable that for the processing of contrast-modulated contours defined by noise carriers with different SFs, cue-invariant orientation responses may be produced by different neuronal mechanisms in early visual cortices. However, this requires further investigations, particularly when considering the fact that orientation is represented in both V1 and V2 with spatial frequency invariance.

Using the spatio-temporal energy model, but with the receptivefield parameters of neurons for V1 and V2, we were able to reproduce the previous observation of orientation-cue invariant population responses to different texture-defined or contrastmodulated contour stimuli. This result suggests that the "filterrectify-filter" model is not essential to account for the population responses recorded in macaque V1 and V2. Interestingly, it was previously found that the orientation preferences of population responses evoked by illusory contours were always orthogonal to those activated by luminance gratings not only in cat areas 17 and 18 but also in macaque V1 and V2 [25,68,98]. Furthermore, the linear energy model could reproduce population responses with an orthogonal orientation preference evoked by illusory-contour stimuli in both species, but not the partially shifted orientation preference observed in monkey V1 and V2 [25]. This suggests that the processing of second-order contours created by modulating noise carriers are different from that of other type illusory or filling-in figures, which are more complicated and clearly need to employ not only separate non-linear integrative mechanisms but also cortical feedback [25,34,99–101].

The processing of local first-order visual cues has been previously observed and predicted for the neuronal firings to other stimuli of texture-defined patterns or forms in macaque V2 [31,102-103]. Psychophysical studies have also suggested that spatial and temporal pooling of orientation-selective filters is a fundamental aspect of the low-level visual processing that underlies orientation discrimination [104-106]. Similar first-stage filters for orientation and spatial frequency have been proposed to sub-serve a common mechanism for the detection of contrast- and texturedefined boundaries [31,46,73,87-88]. Furthermore, results from numerous psychophysical studies suggest that arrays of V1 neurons may integrate their responses to local components into global contours when the local components are collinearly aligned [107-112]. This idea is reinforced by anatomical studies investigating lateral and horizontal connections and population responses of V1 [113-118]. However, at the levels of columns and circuits particularly for generating the visual responses of a subgroup of specialized neurons previously reported to signal second-order stimuli [27,29,32-37], the actual neuronal mechanisms could be much more complicated. So is the case for the processing of the direction signal of second-order motion, which was proposed to invoke a separate non-linear visual mechanism [3,6-7,53,56,78,119-120].

Non-linear integrative processing for second-order contours

The spatio-temporal energy model can only capture the linear aspect of neural population responses in V1 and V2 to our secondorder contour stimuli. However, visual non-linearities start with phototransduction within the retina. Indeed, nonlinear spatial integration and fine-scale heterogeneities in spatial sampling for ganglion cells of mouse retina have been revealed recently [121]. Similarly, responses to some second-order cues have been demonstrated as early as in the guinea pig retina and thalamic Y-type cells in cats [122–124]. Thus, it may not be surprising that a small percentage of high-order neurons within V1 and V2 of non-human primates respond to non-luminance defined boundaries [23,29,32,37,125]. It is impossible to extract the contribution of this small group of neurons from our recordings as our responses are derived from pooled neuronal activities. It is also worth noting that the weak intrinsic signals generated by secondorder stimuli in this study are not consistent with the subjective experience when viewing the stimuli. We readily perceive all these second-order contours without any extra effort and largely independently of the carriers. This suggests that neuronal mechanisms, particularly those underlying response gain control or normalization and retinotopic spatial pooling, involve nonlinearities in visual cortices [25-26,73-74,83,126]. The contribution of surround suppression to encode second-order features at multiple stages may also need to be considered [86,127]. Previous studies have found that the feedback from higher visual cortices can enhance neuronal responses in V1 to higher-order visual stimuli of low saliency [128–129]. This interaction between lowand high-level visual areas also needs to be considered [130–135]. Thus, it appears that the perception of second-order stimuli engages the hierarchical processing from early visual cortices to high brain stages with different focuses in each processing stage. Regardless of the mechanisms involved, the spatio-temporal filtering mechanism revealed here by energy model at the population level in macaque V1 and V2, constitutes the foundation for processing the orientation of second-order contours defined specifically by modulating noise-texture carriers.

Conclusions

It is well-known that the complexity of encoded visual features increases greatly from V1 to V4 and IT, consistent with the great increase of RF size along the visual hierarchy [1-2,136-137]. Therefore, it is likely that at the population level the higher processing stages may capture different features from second-order contour stimuli [26,138-140]. In contrast, the small spatiotemporal RFs of the V1 and V2 neurons are more suitable to the detection of the local luminance variations in these stimuli such as the serrated edges produced by the white and black pixels. It is also known that foveal and parafoveal RFs in primate V1 and V2 are quite small and have super-high spatial resolution. The grating acuity is >26 cpd in monkey [141] and that is higher than the SFs of stimuli used in the current and most if not all previous studies on second-order visual processing. Thus, population activities in early visual cortices of V1 and V2 mainly reflect the processing of local luminance changes while those in higher visual areas represent global second-order features. Our experimental results demonstrate that the orientation of contrast-modulated and phasereversed contours is invariantly represented in the population responses of macaque V1 and V2. Simulations based on a physiologically constrained energy model further suggest that the spatio-temporal filter mechanism of the majority of V1 and V2 neurons underlies the recorded population responses. Our findings also suggest that the orientation maps in macaque V1 and V2 can be described as a spatio-temporal energy map [40].

Supporting Information

Figure S1 The inside and outside view of our nonhuman primate housing facilities. (TIF)

Movie S1 An illustrative movie shows sine-wave luminance gratings of 0° and 90° orientations. $\rm (AVI)$

Movie S2 An illustrative movie shows contrast-modulated noise stimuli of 0° and 90° orientations. $\rm (AVI)$

Movie S3 An illustrative movie shows second-order contour stimuli of 0° and 90° orientations defined by the phase reversal of noise texture. (AVI)

Acknowledgments

We thank Drs. M-m. Poo, M. Stryker, C. Baker and L. Spillmann for comments and suggestions on the manuscript. We thank Dr. M.J. Rasch for his invaluable input toward the model work. Conceived and designed the experiments: WW. Performed the experiments: XA HLG JPY YXP XZ XCW NM WW. Analyzed the data: XA

References

- Rousselet GA, Thorpe SJ, Fabre-Thorpe M (2004) How parallel is visual processing in the ventral pathway? Trends Cogn Sci 8: 363–370.
- Kravitz DJ, Saleem KS, Baker CI, Ungerleider LG, Mishkin M (2013) The ventral visual pathway: an expanded neural framework for the processing of object quality. Trends Cogn Sci 17: 26–49.
- Chubb C, Sperling G (1988) Drift-balanced random stimuli: A general basis for studying non-Fourier motion perception. J Opt Soc of Amer A: 1986–2007.
- Cavanagh P, Mather G (1989) Motion: the long and short of it. Spat Vision, 4 2: 103–129.
- Mather G, West S (1993) Evidence for second-order motion detectors. Vis Res 33: 1109–1112.
- Smith AT, Ledgeway T (1997) Separate detection of moving luminance and contrast modulations: fact or artifact? Vis Res 37: 45–62.
- Vaina LM, Cowey A, Kennedy D (1999) Perception of first- and second-order motion: separable neurological mechanisms? Hum Brain Mapp 7: 67–77.
- Baker C, Mareschal I (2001) Processing of second-order stimuli in the visual cortex. Prog Brain Res 134: 171–191.
- Zhan CA, Baker CL, Jr. (2006) Boundary cue invariance in cortical orientation maps. Cereb Cortex 16: 896–906.
- 10. Glasser DM, Tadin D (2011) Increasing stimulus size impairs first- but not second-order motion perception. J Vis 11.
- Hubel DH, Wiesel TN (1968) Receptive fields and functional architecture of monkey striate cortex. J Physiol 195: 215–243.
 Levitt JB, Kiper DC, Movshon JA (1994) Receptive fields and functional
- Levitt JB, Kiper DC, Movshon JA (1994) Receptive fields and functional architecture of macaque V2. J Neurophysiol 71: 2517–2542.
- Foster KH, Gaska JP, Nagler M, Pollen DA (1985) Spatial and temporal frequency selectivity of neurones in visual cortical areas V1 and V2 of the macaque monkey. J Physiol 365: 331–363.
- Hubel DH, Wiesel TN (1962) Receptive fields, binocular interaction and functional architecture in the cat's visual cortex. J Physiol 160: 106–154.
- Rust NC, Schwartz O, Movshon JA, Simoncelli EP (2005) Spatiotemporal elements of macaque V1 receptive fields. Neuron 46: 945–956.
- Skottun BC, De Valois RL, Grosof DH, Movshon JA, Albrecht DG, et al. (1991) Classifying simple and complex cells on the basis of response modulation. Vis Res 31: 1079–1086.
- Movshon JA, Thompson ID, Tolhurst DJ (1978) Spatial and temporal contrast sensitivity of neurones in areas 17 and 18 of the cat's visual cortex. J Physiol 283: 101–120.
- De Valois RL, Albrecht DG, Thorell LG (1982) Spatial frequency selectivity of cells in macaque visual cortex. Vision Res 22: 545–559.
- De Valois RL, Cottaris NP, Mahon LE, Elfar SD, Wilson JA (2000) Spatial and temporal receptive fields of geniculate and cortical cells and directional selectivity. Vis Res 40: 3685–3702.
- Albright TD (1992) Form-cue invariant motion processing in primate visual cortex. Science 255: 1141–1143.
- Sary G, Vogels R, Kovacs G, Orban GA (1995) Responses of Monkey Inferior Temporal Neurons to Luminance-Defined, Motion-Defined, and Texture-Defined Gratings. J Neurophysiol 73: 1341–1354.
- Sary G, Vogels R, Orban GA (1993) Cue-invariant shape selectivity of macaque inferior temporal neurons. Science 260: 995–997.
- O'keele L, Movshon J (1998) Processing of first-and second-order motion signals by neurons in area MT of the macaque monkey. Vis Neurosci 15: 305– 317.
- Mysore SG, Vogels R, Raiguel SE, Orban GA (2008) Shape selectivity for camouflage-breaking dynamic stimuli in dorsal V4 neurons. Cereb Cortex 18: 1429–1443.
- Pan Y, Chen M, Yin J, An X, Zhang X, et al. (2012) Equivalent representation of real and illusory contours in macaque V4. J Neurosci 32: 6760–6770.
- Kastner S, De Weerd P, Ungerleider LG (2000) Texture segregation in the human visual cortex: A functional MRI study. J Neurophysiol 83: 2453–2457.
- Mareschal I, Baker CL Jr. (1998) A cortical locus for the processing of contrastdefined contours. Nat Neurosci 1: 150–154.
- Gharat A, Baker CL, Jr. (2012) Motion-defined contour processing in the early visual cortex. J Neurophysiol 108: 1228–1243.
- Marcar V, Raiguel S, Xiao D, Orban G (2000) Processing of kinetically defined boundaries in areas V1 and V2 of the macaque monkey. J Neurophysiol 84: 2786–2798.
- Mysore SG, Vogels R, Raiguel SE, Orban GA (2006) Processing of kinetic boundaries in macaque V4. J Neurophysiol 95: 1864–1880.
- El-Shamayleh Y, Movshon JA (2011) Neuronal responses to texture-defined form in macaque visual area V2. J Neurosci 31: 8543–8555.
- Lui LL, Bourne JA, Rosa MG (2005) Single-unit responses to kinetic stimuli in New World monkey area V2: physiological characteristics of cue-invariant neurones. Exp Brain Res 162: 100–108.
- Lamme VA, van Dijk BW, Spekreijse H (1993) Contour from motion processing occurs in primary visual cortex. Nature 363: 541–543.

YLL YPY WW. Contributed reagents/materials/analysis tools: ZT IS. Contributed to the writing of the manuscript: XA WW.

- Lee TS, Nguyen M (2001) Dynamics of subjective contour formation in the early visual cortex. Proc Natl Acad Sci USA 98: 1907–1911.
- Grosof DH, Shapley RM, Hawken MJ (1993) Macaque V1 neurons can signal 'illusory' contours. Nature 365: 550–552.
- Chaudhuri A, Albright TD (1997) Neuronal responses to edges defined by luminance vs. temporal texture in macaque area V1. Vis Neurosci 14: 949– 962.
- von der Heydt R, Peterhans E (1989) Mechanisms of contour perception in monkey visual cortex. I. Lines of pattern discontinuity. J Neurosci 9: 1731– 1748.
- Baker TI, Issa NP (2005) Cortical maps of separable tuning properties predict population responses to complex visual stimuli. J Neurophysiol 94: 775–787.
- Basole A, Kreft-Kerekes V, White LE, Fitzpatrick D (2006) Cortical cartography revisited: A frequency perspective on the functional architecture of visual cortex. Prog Brain Res 154: 121–134.
- Basole A, White LE, Fitzpatrick D (2003) Mapping multiple features in the population response of visual cortex. Nature 423: 986–990.
- Mante V, Carandini M (2005) Mapping of stimulus energy in primary visual cortex. J Neurophysiol 94: 788–798.
- An X, Gong H, Qian L, Wang X, Pan Y, et al. (2012) Distinct functional organizations for processing different motion signals in V1, V2, and V4 of macaque. J Neurosci 32: 13363–13379.
- Schiessl I, Wang W, McLoughlin N (2008) Independent components of the haemodynamic response in intrinsic optical imaging. Neuroimage 39: 634–646.
- Mc Loughlin NP, Blasdel GG (1998) Wavelength-dependent differences between optically determined functional maps from macaque striate cortex. Neuroimage 7: 326–336.
- Aaen-Stockdale C, Bowns L (2006) Motion-detection thresholds for first- and second-order gratings and plaids. Vision Research 46: 925–931.
- Allard R, Faubert J (2013) No dedicated second-order motion system. J Vis 13.
 Allard R, Faubert J (2013) No second-order motion system sensitive to high
- temporal frequencies. J Vis 13.
 Bertone A, Faubert J (2003) How is complex second-order motion processed?
- Vision Research 43: 2591–2601.
- Dakin SC, Williams CB, Hess RF (1999) The interaction of first- and secondorder cues to orientation. Vis Res 39: 2867–2884.
- Ellemberg D, Allen HA, Hess RF (2004) Investigating local network interactions underlying first- and second-order processing. Vision Research 44: 1787–1797.
- Nishida S, Ledgeway T, Edwards M (1997) Dual multiple-scale processing for motion in the human visual system. Vision Research 37: 2685–2698.
- Seiffert AE, Somers DC, Dale AM, Tootell RB (2003) Functional MRI studies of human visual motion perception: texture, luminance, attention and aftereffects. Cereb Cortex 13: 340–349.
- Smith A, Greenlee M, Singh K, Kraemer F, Hennig J (1998) The processing of first-and second-order motion in human visual cortex assessed by functional magnetic resonance imaging (fMRI). J Neurosci 18: 3816–3830.
- Pavan A, Campana G, Guerreschi M, Manassi M, Casco C (2009) Separate motion-detecting mechanisms for first- and second-order patterns revealed by rapid forms of visual motion priming and motion aftereffect. J Vis 9: 2721–16.
- Benton CP, Johnston A (1997) First-order motion from contrast modulated noise? Vis Res 37: 3073–3078.
- Ledgeway T, Hutchinson CV (2006) Is the direction of second-order, contrastdefined motion patterns visible to standard motion-energy detectors: a model answer? Vis Res 46: 556–567.
- Landy MS, Bergen JR (1991) Texture segregation and orientation gradient. Vis Res 31: 679–691.
- An X, Gong H, McLoughlin NP, Yang Y, Wang W (2014) The Mechanism for Processing Random-Dot Motion at Various Speeds in Early Visual Cortices. PloS ONE 9(3): e93115.
- Blasdel G, Campbell D (2001) Functional retinotopy of monkey visual cortex. J Neurosci 21: 8286–8301.
- Bonhoeffer T, Grinvald A (1996) Optical Imaging based on intrinsic signals: the methodology. Brain mapping: the methods: 55–97.
- Bonhoeffer T, Grinvald A (1993) The layout of iso-orientation domains in area 18 of cat visual cortex: optical imaging reveals a pinwheel-like organization. J Neurosci 13: 4157–4180.
- Crair MC, Ruthazer ES, Gillespie DC, Stryker MP (1997) Ocular dominance peaks at pinwheel center singularities of the orientation map in cat visual cortex. J Neurophysiol 77: 3381–3385.
- Maldonado PE, Godecke I, Gray CM, Bonhoeffer T (1997) Orientation selectivity in pinwheel centers in cat striate cortex. Science 276: 1551–1555.
- Ohki K, Chung S, Kara P, Hubener M, Bonhoeffer T, et al. (2006) Highly ordered arrangement of single neurons in orientation pinwheels. Nature 442: 925–928.

- Nauhaus I, Benucci A, Carandini M, Ringach DL (2008) Neuronal selectivity and local map structure in visual cortex. Neuron 57: 673–679.
- Zhan CA, Baker CL, Jr. (2008) Critical spatial frequencies for illusory contour processing in early visual cortex. Cereb Cortex 18: 1029–1041.
- 67. Cole LC (1949) The measurement of inter-specific association. JSTOR: Ecology 30: 411–424.
- Ramsden BM, Hung CP, Roe AW (2001) Real and illusory contour processing in area V1 of the primate: a cortical balancing act. Cereb Cortex 11: 648–665.
- Mante V, Carandini M (2003) Visual cortex: seeing motion. Curr Biol 13: R906–908.
- De Valois RL, Yund EW, Hepler N (1982) The orientation and direction selectivity of cells in macaque visual cortex. Vis Res 22: 531–544.
- Hallum LE, Landy MS, Heeger DJ (2011) Human primary visual cortex (V1) is selective for second-order spatial frequency. J Neurophysiol 105: 2121–2131.
 Bust NC, Mante V, Simoncelli EP, Moyshon IA (2006) How MT cells analyze
- Rust NC, Mante V, Simoncelli EP, Movshon JA (2006) How MT cells analyze the motion of visual patterns. Nat Neurosci 9: 1421–1431.
 Landy MS, Graham N (2004) Visual perception of texture, in The Visual
- Landy MS, Graham N (2004) Visual perception of texture. in The Visual Neurosciences, eds Chalupa LM, Werner JS (MIT, Massachusetts), pp. 1106– 1118.
- 74. Carandini M, Demb JB, Mante V, Tolhurst DJ, Dan Y, et al. (2005) Do we know what the early visual system does? J Neurosci 25: 10577–10597.
- Wilson HR, Ferrera VP, Yo C (1992) A psychophysically motivated model for two-dimensional motion perception. Vis Neurosci 9: 79–97.
- Shapley R (1998) Visual cortex: pushing the envelope. Nat Neurosci 1: 95–96.
 Benton CP, Johnston A, McOwan PW, Victor JD (2001) Computational
- model of , Joinston A, McOwal TW, Victo JD (2001) Completational modeling of non-Fourier motion: further evidence for a single luminance-based mechanism. J Opt Soc Am A Opt Image Sci Vis 18: 2204–2208.
- Fleet DJ, Langley K (1994) Computational Analysis of Non-Fourier Motion. Vis Res 34: 3057–3079.
- Adelson EH, Bergen JR (1985) Spatiotemporal energy models for the perception of motion. J Opt Soc Am A 2: 284–299.
- DeAngelis GC, Ohzawa I, Freeman RD (1993) Spatiotemporal organization of simple-cell receptive fields in the cat's striate cortex. II. Linearity of temporal and spatial summation. J Neurophysiol 69: 1118–1135.
- Movshon JA, Thompson ID, Tolhurst DJ (1978) Spatial summation in the receptive fields of simple cells in the cat's striate cortex. J Physiol 283: 53–77.
 Reid RC, Soodak RE, Shapley RM (1991) Directional selectivity and
- Reid RC, Soodak RE, Shapley RM (1991) Directional selectivity and spatiotemporal structure of receptive fields of simple cells in cat striate cortex. J Neurophysiol 66: 505–529.
- 83. Rust NC, Movshon JA (2005) In praise of artifice. Nat Neurosci 8: 1647-1650.
- Rasch MJ, Chen M, Wu S, Lu HD, Roe AW (2012) Quantitative inference of population response properties across eccentricity from motion induced maps in macaque V1. J Neurophysiol.
- Larsson J, Landy MS, Heeger DJ (2006) Orientation-selective adaptation to first- and second-order patterns in human visual cortex. J Neurophysiol 95: 862–881.
- Wang HX, Heeger DJ, Landy MS (2012) Responses to second-order texture modulations undergo surround suppression. Vis Res 62: 192–200.
- Manahilov V, Simpson WA, Calvert J (2005) Why is second-order vision less efficient than first-order vision? Vis Res 45: 2759–2772.
- Dakin SC, Mareschal I (2000) Sensitivity to contrast modulation depends on carrier spatial frequency and orientation. Vis Res 40: 311–329.
- Mante V, Frazor RA, Bonin V, Geisler WS, Carandini M (2005) Independence of luminance and contrast in natural scenes and in the early visual system. Nat Neurosci 8: 1690–1697.
- Shapley R (1986) The importance of contrast for the activity of single neurons, the VEP and perception. Vis Res 26: 45–61.
- Frazor RA, Geisler WS (2006) Local luminance and contrast in natural images. Vis Res 46: 1585–1598.
- Weliky M, Fiser J, Hunt RH, Wagner DN (2003) Coding of natural scenes in primary visual cortex. Neuron 37: 703–718.
- Geisler WS, Albrecht DG, Crane AM (2007) Responses of neurons in primary visual cortex to transient changes in local contrast and luminance. J Neurosci 27: 5063–5067.
- Boynton GM, Demb JB, Glover GH, Heeger DJ (1999) Neuronal basis of contrast discrimination. Vis Res 39: 257–269.
- Ayzenshtat I, Gilad A, Zurawel G, Slovin H (2012) Population response to natural images in the primary visual cortex encodes local stimulus attributes and perceptual processing. J Neurosci 32: 13971–13986.
- Nassi JJ, Callaway EM (2009) Parallel processing strategies of the primate visual system. Nat Rev Neurosci 10: 360–372.
- Sincich LC, Horton JC (2005) The circuitry of V1 and V2: integration of color, form, and motion. Annu Rev Neurosci 28: 303–326.
- Sheth BR, Sharma J, Rao SC, Sur M (1996) Orientation maps of subjective contours in visual cortex. Science 274: 2110–2115.
- Sary G, Koteles K, Kaposvari P, Lenti L, Csifcsak G, et al. (2008) The representation of Kanizsa illusory contours in the monkey inferior temporal cortex. Eur J Neurosci 28: 2137–2146.
- Stanley DA, Rubin N (2003) fMRI activation in response to illusory contours and salient regions in the human Lateral Occipital Complex. Neuron 37: 323– 331.
- 101. Cox MA, Schmid MC, Peters AJ, Saunders RC, Leopold DA, et al. (2013) Receptive field focus of visual area V4 neurons determines responses to illusory surfaces. Proc Natl Acad Sci U S A 110: 17095–17100.

- Anzai A, Peng X, Van Essen DC (2007) Neurons in monkey visual area V2 encode combinations of orientations. Nat Neurosci 10: 1313–1321.
- Ito M, Komatsu H (2004) Representation of angles embedded within contour stimuli in area V2 of macaque monkeys. J Neurosci 24: 3313–3324.
- Westheimer G, Ley EJ (1997) Spatial and temporal integration of signals in foveal line orientation. J Neurophysiol 77: 2677–2684.
- Parkes L, Lund J, Angelucci A, Solomon JA, Morgan M (2001) Compulsory averaging of crowded orientation signals in human vision. Nat Neurosci 4: 739–744.
- Jones DG, Anderson ND, Murphy KM (2003) Orientation discrimination in visual noise using global and local stimuli. Vis Res 43: 1223–1233.
- 107. Field DJ, Hayes A, Hess RF (1993) Contour integration by the human visual system: evidence for a local "association field". Vis Res 33: 173–193.
- Polat U, Mizobe K, Pettet MW, Kasamatsu T, Norcia AM (1998) Collinear stimuli regulate visual responses depending on cell's contrast threshold. Nature 391: 580–584.
- Li W, Gilbert CD (2002) Global contour saliency and local colinear interactions. J Neurophysiol 88: 2846–2856.
- Kapadia MK, Ito M, Gilbert CD, Westheimer G (1995) Improvement in visual sensitivity by changes in local context: parallel studies in human observers and in V1 of alert monkeys. Neuron 15: 843–856.
- Nelson JI, Frost BJ (1985) Intracortical facilitation among co-oriented, coaxially aligned simple cells in cat striate cortex. Exp Brain Res 61: 54–61.
- Dresp B, Bonnet C (1995) Subthreshold summation with illusory contours. Vision Research 35: 1071–1078.
- 113. Rockland KS, Lund JS (1982) Widespread periodic intrinsic connections in the tree shrew visual cortex. Science 215: 1532–1534.
- Gilbert CD, Wiesel TN (1989) Columnar specificity of intrinsic horizontal and corticocortical connections in cat visual cortex. J Neurosci 9: 2432–2442.
- 115. Malach R, Amir Y, Harel M, Grinvald A (1993) Relationship between intrinsic connections and functional architecture revealed by optical imaging and in vivo targeted biocytin injections in primate striate cortex. Proc Natl Acad Sci U S A 90: 10469–10473.
- Angelucci A, Levitt JB, Walton EJ, Hupe JM, Bullier J, et al. (2002) Circuits for local and global signal integration in primary visual cortex. J Neurosci 22: 8633–8646.
- 117. Bosking WH, Zhang Y, Schofield B, Fitzpatrick D (1997) Orientation selectivity and the arrangement of horizontal connections in tree shrew striate cortex. J Neurosci 17: 2112–2127.
- 118. Das A, Gilbert CD (1999) Topography of contextual modulations mediated by short-range interactions in primary visual cortex. Nature 399: 655–661.
- Baker CL, Jr. (1999) Central neural mechanisms for detecting second-order motion. Curr Opin Neurobiol 9: 461–466.
- Nishida S, Sasaki Y, Murakami I, Watanabe T, Tootell R (2003) Neuroimaging of direction-selective mechanisms for second-order motion. J Neurophysiol 90: 3242.
- 121. Schwartz GW, Okawa H, Dunn FA, Morgan JL, Kerschensteiner D, et al. (2012) The spatial structure of a nonlinear receptive field. Nat Neurosci 15: 1572–1580.
- Demb JB, Zaghloul K, Sterling P (2001) Cellular basis for the response to second-order motion cues in Y retinal ganglion cells. Neuron 32: 711–721.
- Rosenberg A, Husson TR, Issa NP (2010) Subcortical representation of non-Fourier image features. J Neurosci 30: 1985–1993.
- Rosenberg A, Issa NP (2011) The y cell visual pathway implements a demodulating nonlinearity. Neuron 71: 348–361.
- 125. Super H, Spekreijse H, Lamme VA (2001) Two distinct modes of sensory processing observed in monkey primary visual cortex (V1). Nat Neurosci 4: 304–310.
- Carandini M, Heeger DJ (2012) Normalization as a canonical neural computation. Nat Rev Neurosci 13: 51–62.
- Tanaka H, Ohzawa I (2009) Surround suppression of V1 neurons mediates orientation-based representation of high-order visual features. J Neurophysiol 101: 1444–1462.
- Hupe JM, James AC, Payne BR, Lomber SG, Girard P, et al. (1998) Cortical feedback improves discrimination between figure and background by V1, V2 and V3 neurons. Nature 394: 784–787.
- Lee TS, Mumford D, Romero R, Lamme VA (1998) The role of the primary visual cortex in higher level vision. Vision Res 38: 2429–2454.
- Koivisto M, Railo H, Revonsuo A, Vanni S, Salminen-Vaparanta N (2011) Recurrent processing in V1/V2 contributes to categorization of natural scenes. J Neurosci 31: 2488–2492.
- Wokke ME, Vandenbroucke AR, Scholte HS, Lamme VA (2013) Confuse your illusion: feedback to early visual cortex contributes to perceptual completion. Psychol Sci 24: 63–71.
- Bullier J (2001) Integrated model of visual processing. Brain Research Reviews 36: 96–107.
- Lee TS, Yang CF, Romero RD, Mumford D (2002) Neural activity in early visual cortex reflects behavioral experience and higher-order perceptual saliency. Nat Neurosci 5: 589–597.
- Gilbert CD, Li W (2013) Top-down influences on visual processing. Nat Rev Neurosci 14: 350–363.
- Ramalingam N, McManus JN, Li W, Gilbert CD (2013) Top-down modulation of lateral interactions in visual cortex. J Neurosci 33: 1773–1789.

- 136. Lennie P (1998) Single units and visual cortical organization. Perception 27: $889{-}935.$
- 137. Orban GA (2008) Higher order visual processing in macaque extrastriate cortex. Physiol Rev 88: 59–89.
- Kourtzi Z, Tolias AS, Altmann CF, Augath M, Logothetis NK (2003) Integration of local features into global shapes: monkey and human FMRI studies. Neuron 37: 333–346.
- 139. Mendola JD, Dale AM, Fischl B, Liu AK, Tootell RB (1999) The representation of illusory and real contours in human cortical visual areas

revealed by functional magnetic resonance imaging. J Neurosci 19: 8560–8572.

- Hedges JH, Gartshteyn Y, Kohn A, Rust NC, Shadlen MN, et al. (2011) Dissociation of neuronal and psychophysical responses to local and global motion. Curr Biol 21: 2023–2028.
- Kiorpes L (1992) Development of vernier acuity and grating acuity in normally reared monkeys. Vis Neurosci 9: 243–251.