

Preventive Effect of *Lactobacillus acidophilus* XY27 on DSS-Induced Ulcerative Colitis in Mice

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Introduction: *Lactobacillus acidophilus* is widely used as probiotic supplement in functional foods due to its beneficial regulatory effects on host, such as immune regulation, anti-inflammatory, and antioxidant activities.

Aim: This study aimed to determine the preventive effect of *Lactobacillus acidophilus* XY27 (*L. acidophilus* XY27) on colitis induced by dextran sodium.

Methods: The mice were randomly divided into five groups. Except for the control group, the other four groups were induced for ulcerative colitis (UC) with dextran sodium sulfate (DSS), and three groups in DSS-groups were treated with *L. acidophilus* XY27, *L. bulgaricus*, and salicylazosulfapyridine. The weight change, DAI score, colon length, and length to weight ratio were tested. The oxidation index and the levels of inflammatory cytokines in the serum were measured. Subsequently, the gene expression levels of inflammatory factors in the colon tissue were determined by the Real-Time quantitative polymerase chain reaction (qRT-PCR) method.

Results: The results showed that the mice in the *L. acidophilus* XY27 group performed better in terms of weight, DAI score, colon length, and length to weight ratio or colonic pathological sections compared with the DSS-induced group. Further, the levels of tumor necrosis factor α (TNF- α), Interleukin-6 (IL-6), Interleukin-12 (IL-12) and Interleukin-1 β (IL-1 β), malondialdehyde (MDA) content, and myeloperoxidase activity in the serum of UC mice treated with *L. acidophilus* XY27 significantly decreased, while the levels of Interferon- γ (IFN- γ), Interleukin-10 (IL-10), Catalase (CAT), and total superoxide dismutase (SOD) significantly increased. The gene expression levels of Ets-like transcription factor-1 (EIK-1), IL-12, IL-1 β , Cyclooxygenase 2 (COX-2), TNF- α , Escherichia coli, Lipopolysaccharide (LPS), and p100 in the colon significantly decreased while those of tight junction protein 1 (ZO-1), nuclear factor kappa B (NF- κ B), p53, and NF-kappa-B inhibitor alpha (I κ B- α) increased in the *L. acidophilus* XY27 group.

Conclusion: The results of the experiment suggested that *L. acidophilus* XY27 prevented colitis and alleviated symptoms in mice with DSS-induced UC, and also repaired the intestinal barrier function.

Keywords: dextran sodium sulfate, *Lactobacillus acidophilus* XY27, *Lactobacillus bulgaricus*, ulcerative colitis

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Introduction

Inflammatory bowel disease (IBD) is a chronic, recurrent, long-term inflammatory disease that eventually leads to colonic and rectal ulcers, including Crohn's disease (CD) and ulcerative colitis (UC). At present, no specific treatment is available for UC. Anti-inflammatory drugs and immunosuppressants are used mainly to relieve symptoms, which are effective only in the early stages of the disease with some side

effects.¹ Hence, more safe strategies for preventing and treating UC need to be explored.

The pathogenesis of UC is complex and related to both genetic and environmental factors. No matter what the exact mechanism of inducing and maintaining enteritis is, sufficient evidence shows that a strong local immune response is involved. Immune cells receive signals and are recruited and activated. They release soluble cytokines and inflammatory mediators and cause more serious inflammation and tissue damage, leading to the amplification and enhancement of local immune response.² Inflammatory mediators, especially cytokines, mediate the proliferation and differentiation of immune cells, regulate inflammatory cells, and balance the function of the intestinal mucosal barrier, thus playing a role in the immune response.³ The relative expression levels of TNF- α , IL-1 β , IFN- γ , IL-12, IL-6, EIK-1, COX-2, *E. coli*, LPS, and p100 genes increased in patients with colitis, the relative expression levels of ZO-1, NF- κ B, p53, and IKB- α genes decreased, and the contents of some of these factors were changed on the first day after DSS induction.⁴ The changes in the levels of inflammatory cytokines in colonic mucosa are the main causes of intestinal mucosal inflammatory injury and ulcers.

Many studies showed an important role of the intestinal flora in the mucosal immune response in intestinal diseases and inflammation.^{1,5,6} UC can be treated by regulating intestinal flora and increasing the number of probiotic.⁷⁻⁹ Therefore, increasing attention was paid to the potential and beneficial roles of probiotics in the control and regulation of IBD. Meta-analyses and systematic review reports showed that probiotics were beneficial to the treatment of IBD, especially the combined use of probiotics in treating UC.¹⁰ Millions of people worldwide consume probiotics, and the market size of probiotics is expected to reach nearly US \$70 billion by 2023.¹¹ Among these, *Lactobacillus*, as important commercial probiotics, is widely added to functional foods worldwide.

Lactobacillus acidophilus, which is a homofermentative species, is now one of the most typical *Lactobacillus* genera commonly used for various beneficial effects on human health.¹² *L. acidophilus* can not only decrease blood cholesterol, lactose, and the risk of constipation and diarrhea but also reduce the risk of mutation and carcinogenesis. *L. acidophilus* is recognized as a probiotic strain with anticancer and cholesterol-lowering properties, as well as an antagonist against intestinal and foodborne pathogens. It can survive in harsh environments and complex

gastrointestinal ecosystems.^{11,13} The beneficial effect of *L. acidophilus* lasts longer compared with that of microorganisms that cannot settle in the intestinal tract, and it is a potential strain.¹⁴ The anticancer and cholesterol-lowering characteristics, as well as the antagonism against intestinal and food pathogenic bacteria, are usually achieved by affecting the homeostasis of intestinal flora; its mechanism is under investigation.¹⁴ Intestinal flora and probiotics are vital in the occurrence, development, and treatment of UC. Probiotic can regulate inflammatory response by stimulating cells to produce cytokines. In particular, some *Lactobacillus* and *Bifidobacterium* genus can activate macrophages, natural killer (NK) cells, and T lymphocytes of the host to regulate the immune system. However, the evidence for the clinical effect of probiotics on these diseases is not sufficient probably because the individual differences and the health effect of probiotics are strain specific. Huge genomic diversity exists among different strains of *Lactobacillus*. The in vitro determination of many probiotics and animal model data showed that the efficacy was different among the potential probiotic strains tested.⁹ Different probiotics have different effects on different diseases because of strain and disease specificity. Therefore, it is very meaningful to find and verify strains with good efficacy.¹⁵

An effective probiotic to prevent and treat UC needs to be urgently developed. In this study, the preventive and therapeutic effects of *L. acidophilus* XY27 on DSS-induced UC were verified through detecting the colonic histological, blood and tissue-related inflammatory factors, oxidative stress factors of mice. This research aimed to obtain a *Lactobacillus* strain with the potential to ameliorate colitis.

Materials and Methods

Preparation of *L. acidophilus*

Source of *L. acidophilus*

L. acidophilus XY27 used in this study was isolated and identified from the traditional fermented yak yogurt samples of herdsmen in Xinyuan County, Northwest Xinjiang Uygur Autonomous region, China. The preservation number is CGMCC No. 18227 in China General Microbiological Culture Collection Center, Beijing, China (hereafter referred to as LA-XY27).

Activation of *L. acidophilus*

LA-XY27 frozen in glycerol was dissolved in the bath, inoculated in a 5 mL of MRS medium, and further cultured at 37°C for 1 day. Subsequently, it was activated

again in the liquid medium after growth. After LA-XY27 preparation, Gram staining was used and the microscopic examination was performed to observe whether the strains were contaminated. The number of live bacteria and the absorbance of the original solution were calculated by plate counting and absorbance measurement.

Preparation of the Suspension

The purebred colonies were inoculated on 5 mL of MRS medium and cultured at 37°C for 24 h. Then, 1 mL of the medium containing bacteria was centrifuged at 4000 rpm for 10 min. The supernatant was taken, and the bacteria were resuspended in sterile saline and finally stained.

Further, 200 µL of sample bacteria were poured into the aseptic medium and cultured in an incubator at 37°C for 16–40 h. The culture medium containing bacteria was centrifuged for 10 min at the speed of 4000 rpm. The supernatant was discarded, and the bacteria at the bottom were retained. The bacteria were resuscitated with 5 mL of aseptic saline. Finally, the OD value of the bacterial suspension was determined, which was about 1.9. The suspension could be used for gavage administration.

Animal Experiment

The experimental protocol is shown in Figure 1. Fifty C57BL/6 mice (SPF grade, male, aged 6 weeks) were

purchased from Chongqing Medical University and adaptively fed in an environment at constant temperature and humidity for a week. Fifty mice were randomly divided into five groups ($n = 10$): control group (C-G), DSS group (DSS-G), LA-XY27 group (LA-G), salicylazosulfapyridine group (SSZ-G), and *L. bulgaricus* group (BB-G). In the whole experiment, the mice in all groups were fed for 3 weeks. The mice in the five groups were separately randomized to receive normal saline (0.1 mL/10 g), LA-XY27 (1×10^9 CFU/mL, 0.1 mL/10 g), sulfasalazine (20 mg/kg), and *L. bulgaricus* (1.0×10^9 CFU/mL, 0.1 mL/10 g) by gastric lavage daily for 3 weeks. From the second week, the mice in the control group were fed normal saline, while the mice in the other groups were fed 3% DSS aqueous solution for 1 week. The body weight, vitality, fecal and anal bleeding of mice were recorded, and the condition of mice was observed after lavage administration on time every day. After 3 weeks of intervention, the mice were fed for 24 hours under the condition of fasting and water. The colon was preserved after the mice were euthanized, and the length and weight were recorded. This study was approved by the Ethics Committee of Chongqing Collaborative Innovation Center for Functional Food (201906019B, Chongqing, China) and followed the national standard of the people's Republic of China (GB/T 35892–2018) laboratory animal-guidelines for ethical review of animal welfare.

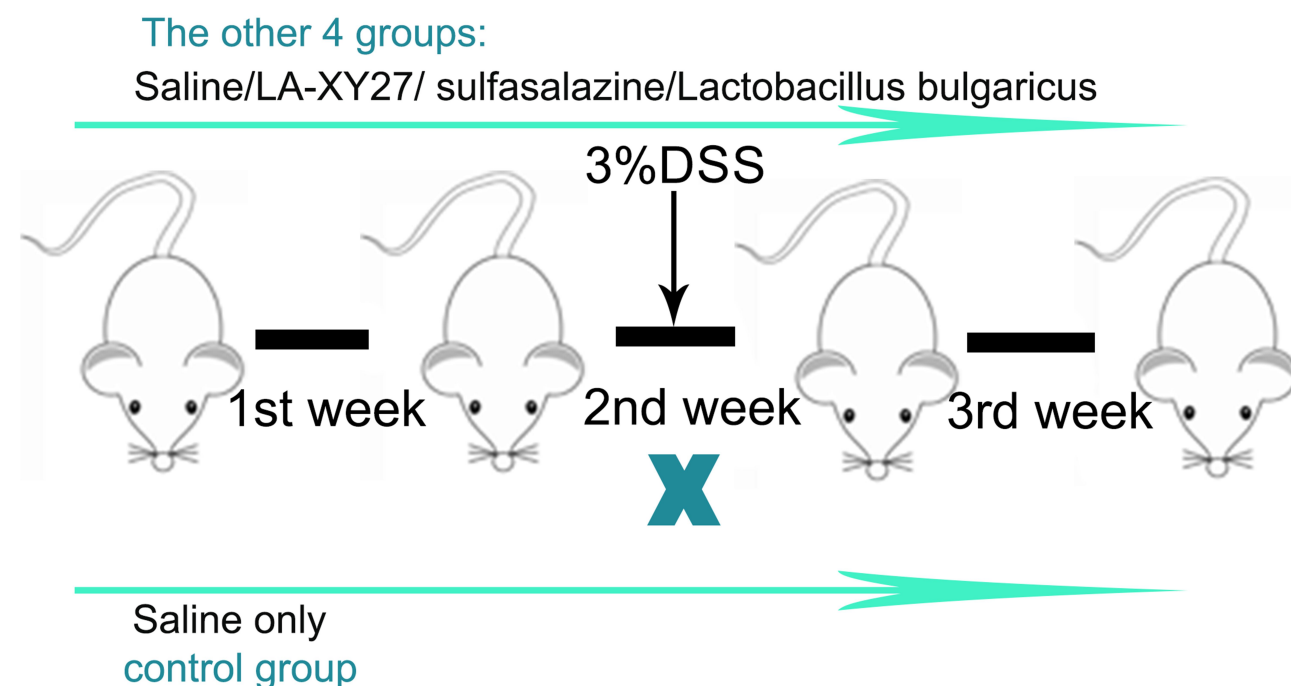


Figure 1 Illustrative diagram of the experimental process. In addition to the control group, the other four groups were induced by DSS in the second week and maintained for one week.

Disease Activity Index (DAI) Score of Mice

The disease activity was evaluated according to the evaluation method of human inflammatory bowel disease (IBD) activity.¹⁶ According to the blood stool condition and stool characteristics of mice every day, the score was as follows: 0 point for normal stool, 1 point for soft but formed stool, 2 points for unformed stool, and 3 points for loose stool. For bloody stool, 0 point for occult blood negative, 1 point for occult blood positive, 2 points for blood in stool, and 3 points for bloody stool. The disease activity was evaluated by adding the two scores.

Macroscopic Observation of Colon

The colon tissue with length no more than 0.5 cm was immersed in 10% formalin fixation solution for 48 h. The tissue blocks were then trimmed into smaller pieces and then followed by dehydration, embedding in paraffin, sectioning with a thickness of 5–8 μm , and staining with hematoxylin and eosin, and finally photographed under a microscope with different magnification (Olympus, Tokyo, Japan).

Detection of Oxidative Stress Factors and Inflammatory Factors in Serum

Blood samples were collected by enucleating eyeball from mice, then centrifuged at 4000 rpm for 10 min at 4°C to obtain serum. The serum was aspirate and saved standby at –80°C for future use.

The catalase (CAT), myeloperoxidase (MPO), and total superoxide dismutase (T-SOD), and the malondialdehyde (MDA) levels in the serum of mice were detected using

the related kits (Nanjing Jiancheng Biological Engineering Research Institute, Nanjing, Jiangsu, China) and enzyme-labeling instrument.

The inflammatory factors (TNF- α , IFN- γ , IL-1 β , IL-6, IL-10, and IL-12) in the serum were detected by using enzyme-linked immunosorbent assays (ELISA kits) (xyA049Rb, XY-R0012c, XY-R0019c, XYA399Mu, xyA058Gu, and XYA133Mu; BioLegend, San Diego, CA, USA) following the manufacturer's protocols.

Detection of Gene Expression in Colon

Total RNA was extracted from 50 to 100 mg colon tissue of each sample using TRIzol reagent (Invitrogen, Carlsbad, CA, USA) and then the concentration and purity of RNA were detected. RNA was reverse-transcribed into cDNA following the reverse transcription kit instructions (Tiangen Biotech Co., Ltd., Beijing, China). Finally, the target gene was amplified by real-time quantitative polymerase chain reaction using the gene primers listed in Table 1.

Statistical Analysis

All data were expressed as mean \pm standard deviation. One-way ANOVA analysis of variance was used for data analysis with SPSS 19.0 software. $P < 0.05$ indicated a statistically significant difference.

Results

Body Weight Change and Disease Activity Index (DAI) Score of Mice

The body weight change and DAI score of mice before and after the end of DSS induction in each group are

Table 1 Sequences of Gene Primers Used in RT-PCR

Gene Name	Forward	Reverse
β -actin	5'-ATGGAGCCGGACAGAAAAGC-3'	5'-TGGGAGGTGTCAACATCTTC-3'
EIK-1	5'-TTGTGTCTACCCAGAGGTTG-3'	5'-GCTATGGCCGAGGTTACAGA-3'
COX-2	5'-TTCCAATCCATGTCAAACCG-3'	5'-AGTCCGGGTACAGTCACACT-3'
LPS	5'-GAACAGTATGCAGTGTAAGAG-3'	5'-GCAGATGCCATAGTTGGTGT-3'
<i>Escherichia coli</i>	5'-GTTAATACCTTTGCTCATTGA-3'	5'-ACCAGGTACTTAATCCTGTT-3'
IL-12	5'-CCTCCACTGTGCTGGTTTTAT-3'	5'-TCAGCAACATGCTCCAGAAG-3'
NF- κ B	5'-TGCGATTCCGCTATAAATGCG-3'	5'-ACAAGTTCATGTGGATGAGG-3'
p100	5'-TGATGGAGGGAACCCAAACTC-3'	5'-GGATGACCCATTCGGGCTT-3'
p53	5'-TCACAGCGTCTGTTGACATT-3'	5'-ACCAAGCTCATTACCCTGAC-3'
IKB- α	5'-CGAGACTTTCGAGGAAATACC-3'	5'-GTCTGCGTCAAGACTGCTAC-3'
TNF- α	5'-CAGGCGGTGCCTATGTCTC-3'	5'-CGATCACCCCGAAGTTCAGT-3'
IL-1 β	5'-GAAATGCCACCTTTTGACAGT-3'	5'-TGGATGCTCTCATCAGGACA-3'

Note: The relative expression of target genes was normalized to the expression level of β -actin and the level in the DSS-G using the $2^{-\Delta\Delta CT}$ method.

shown in Figure 2 and Table 2. DSS induction lasted for 7 days from the first day of week 2. Except for the C-G, the weight of mice in the other four groups began to decline on day 3 and day 4. The weight loss in DSS-G, LA-G, and BB-G was the most obvious, followed by the SSZ-G. From the 11th day, that is, 3 days after the end of DSS induction, the weight of mice in the intervention groups

began to increase. As of the 14th day, the weight of mice in SSZ-G was the closest to the C-G, followed by the LA-G, followed by the DSS-G and BB-G.

On the last day of DSS induction, compared with the C-G (0.000 ± 0), the DAI score of the other four groups was significantly increased ($P < 0.01$), the highest in the DSS-G, followed by the other three intervention groups, but there was

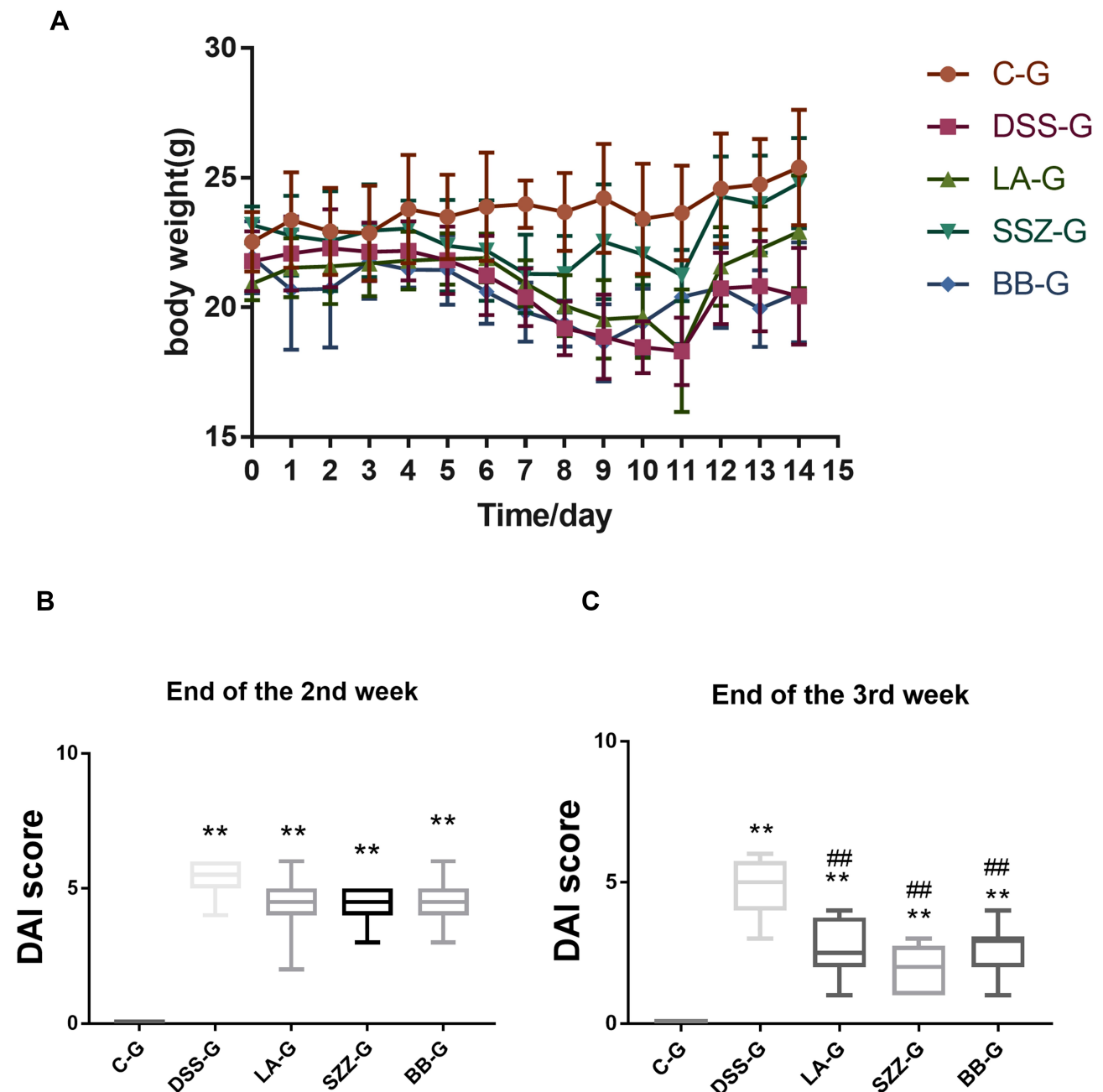


Figure 2 Body weight change curve and DAI score of mice. (A) Weight changes of mice in each group, DSS induction was started on day 1 and ended on day 7 (n=8). (B–C) DAI scores at the end of week 2 (end of DSS induction) (B) and at the end of week 3 (C). All the data were presented as mean \pm SD. **Means $p < 0.01$ compared with the C-G, ## means $p < 0.01$ compared with the DSS-G.

Abbreviations: C-G, control group; DSS-G, DSS-induced group; LA-G, LA-XY27 group; SSZ-G, salicylazosulfapyridine group; BB-G, *L. bulgaricus* group.

Table 2 DAI Score of Mice at the End of 2nd and 3rd Week

Group	End of 2nd Week	End of 3rd Week
C-G	0.000 ± 0	0.000 ± 0
DSS-G	5.375 ± 0.696**	4.750 ± 0.968**
LA-G	4.375 ± 1.111**	2.625 ± 0.992***
SSZ-G	4.375 ± 0.696**	1.875 ± 0.781***
BB-G	4.500 ± 0.866**	2.625 ± 0.857***

Notes: Values are presented as mean ± standard deviation (n = 8). **Means p < 0.01 compared with the C-G, *** means p < 0.01 compared with the DSS-G.

no statistical difference between the four groups (Figure 2B); after a week of recovery and intervention, compared with the DSS-G, the DAI score of LA-G, SSZ-G, and BB-G were significantly decreased ($P < 0.01$, Figure 2C), SSZ-G decreased the most, followed by LA-G.

Macroscopic Observation of Colon

As shown in Figure 3 and Table 3, the colon length was noticeably shorter in DSS-G (6.17 ± 0.87 cm) than in C-G (7.11 ± 0.35 cm) ($P < 0.05$). Although there was no statistical difference, it could be seen from the results that the colonic length in the SSZ-G (6.27 ± 0.79 cm) and BB-G (6.20 ± 0.90 cm) was longer than that in the DSS-G (6.17 ± 0.87 cm), but shorter than that in the C-G (7.11 ± 0.35 cm) and LA-G (7.11 ± 0.62 cm). The colonic length of mice treated with LA-XY27 was longer than that in the DSS-G, BB-G, and SSZ-G, indicating that LA-XY27 could prevent colon shortening. The ratio of colon weight to colon length was significantly increased in DSS-treated mice (0.05 ± 0.01) compared to that of control mice (0.03 ± 0.01) ($P < 0.01$). The ratio of LA-XY27-treated mice (0.04 ± 0.01) had a significant difference with DSS-treated mice (0.05 ± 0.01) ($P < 0.01$), but there was no significant difference compared with the other two treatment groups.

Morphological Observation of Colon Tissue

As shown in Figure 4, the colonic epithelium of mice in the C-G was intact, no ulcer or hyperplasia was found on the mucosal surface, and the crypt structure was regular and clear without obvious inflammatory infiltration; in the DSS-G, obvious inflammation with a large number of interstitial lymphocytes infiltrated and gathered between the basal layer and mucosal muscle (marked by asterisk), which caused thickening of the basal layer and villous defect (marked by black arrow), and the crypt showed structural changes such as shortening, branching, and curling (marked by yellow arrow).

Compared with the DSS-G, the inflammatory reaction of the three intervention groups was reduced, but the effect of LA-G was the best, which was the closest to that of C-G. The crypt structure of LA-G tended to be normal, with only slight lymphatic infiltration in the basement layer. In SSZ-G, there were a small number of lymphocyte infiltration and a few crypt abnormalities, while the effect of BB-G is not very good, with multiple lymphatic infiltration and villus defects.

Levels of Oxidative Stress Factors in Serum

Table 4 and Figure 5 shows that the CAT and T-SOD in the DSS-G (152.96 ± 53.9 and 64.79 ± 8.51) were significantly lower than that in the C-G (589.33 ± 61.9 and 91.88 ± 5.12) ($P < 0.01$), while MDA and MPO were higher in DSS-G (10.58 ± 2.95 and 830.04 ± 71.1) than that in C-G (5.50 ± 2.69 and 786.62 ± 60.5). The CAT and T-SOD levels increased both in LA-G (536.58 ± 68.1 and 89.86 ± 2.15) and SSZ-G (477.68 ± 103.1 and 89.96 ± 6.52) compared to DSS-G ($P < 0.05$). The MDA and MPO contents reduced significantly in LA-G (5.59 ± 2.59 and 794.64 ± 51.9) and SSZ-G (6.59 ± 3.69 and 795.22 ± 55.5) and BB-G (6.81 ± 2.67 and 797.34 ± 49.0) ($P < 0.01$). While there was no significant effect on CAT in BB-G. In conclusion, the data of LA-G were closer to that of C-G, which confirming that the effect of LA-XY27 was better.

Levels of Inflammatory Cytokines in Serum

Table 5 and Figure 6 shows that the levels of IFN- γ , IL-1 β , IL-6, IL-12, and TNF- α in the serum were the lowest in the C-G among all the groups, while the levels of IL-10 were the highest, in contrast, the aforementioned indicators followed the opposite trends in the DSS-G, there was significant difference between the two groups ($P < 0.01$). The concentrations of IFN- γ , IL-10, IL-12, IL-6, and TNF- α improved significantly in the three intervention groups ($P < 0.05$), especially there was no significant difference in IFN, TNF, and IL-12 between LA-G and C-G.

Gene Expression of the Colonic Tissue

As shown in Table 6 and Figure 7, the relative expression levels of EIK-1, IL-12, IL-1 β , COX-2, TNF- α , *E. coli*, LPS, and p100 genes were higher while the relative expression levels of ZO-1, NF- κ B, p53, and IKB- α genes were lower in the DSS-G compared with the C-G ($P < 0.01$), which were consistent with previously reported levels in serum. Although there were differences

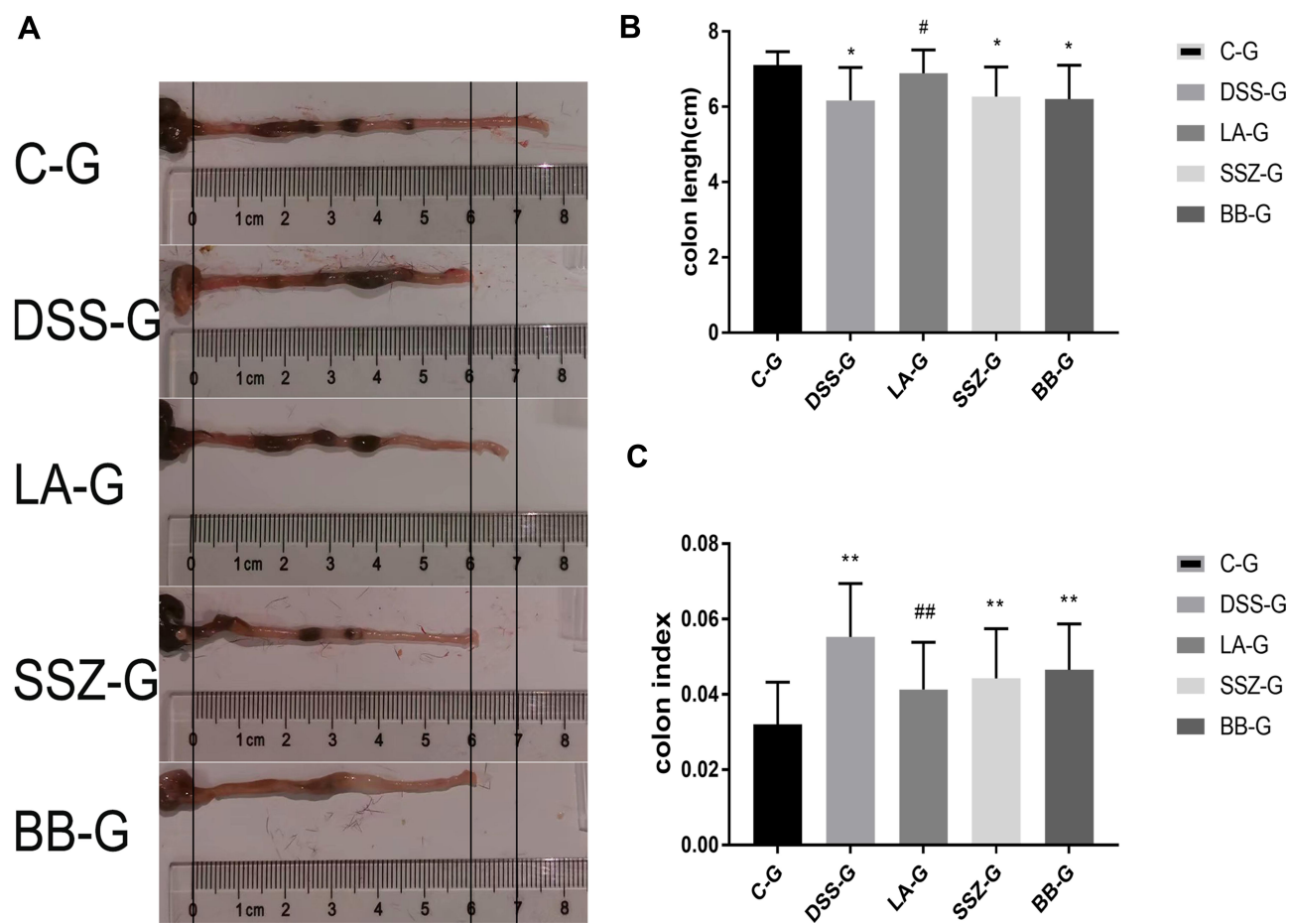


Figure 3 Colon length and colon index in each group. (A–B) Representative images and bar chart of colon length in each group (n=8). (C) colon index in each group (n=8). All the data were presented as mean \pm SD in B and C. * and ** means $p < 0.05$ and $p < 0.01$ respectively compared with the C-G; # and ### means $p < 0.05$ and $p < 0.01$ respectively compared with the DSS-G.

Abbreviations: C-G, control group; DSS-G, DSS-induced group; LA-G, LA-XY27 group; SSZ-G, salicylazosulfapyridine group; BB-G, *L. bulgaricus* group.

in LPS, EIK-1, p100, TNF- α between all the groups, the change of *E. coli*, IL-12 NF- κ B, ZO-1 and IL-1 β were more obvious. In general, the relative expression levels of EIK-1, IL-12, IL-1 β , TNF- α , *E. coli*, LPS, p100, and COX-2 genes were significantly lower in the LA-G than

in the DSS-G, while the relative expression levels of ZO-1, NF- κ B, p53, and IKB- α genes significantly increased and were closer to those in the C-G compared with the other two treatment groups (SSZ-G and BB-G), suggesting that LA-XY27 can adjust the levels of inflammatory factors more effectively.

Table 3 Colonic Length and the Ratio of Colonic Weight to Colonic Length

Group	Colonic Length (cm)	Colonic Weight/Colonic Length (g/cm)
C-G	7.11 \pm 0.35	0.03 \pm 0.01
DSS-G	6.17 \pm 0.87*	0.05 \pm 0.01**
LA-G	6.89 \pm 0.62#	0.04 \pm 0.01###
SSZ-G	6.27 \pm 0.79*	0.04 \pm 0.01**
BB-G	6.20 \pm 0.90*	0.04 \pm 0.01**

Notes: Values are presented as mean \pm standard deviation (n = 8). *Means $p < 0.05$ compared with the C-G, # means $p < 0.05$ compared with the DSS-G. **Means $p < 0.01$ compared with the C-G, ### means $p < 0.01$ compared with the DSS-G.

Discussion

In this study, the preventive and regulatory effects of LA-XY27 were verified by using the DSS-induced model. DSS induces superficial ulceration, mucosal damage, diarrhea, rectal bleeding, and loss of body weight, this model is elementary and reproducible and its clinical symptoms are similar to ulcerative colitis.^{17,18}

The results of DAI disease score scale showed that LA-XY27 had a good relieving effect on hematochezia and alleviate the weight loss caused by 3% DSS feeding. Due to colonic edema, the weight increased, the length shortened,

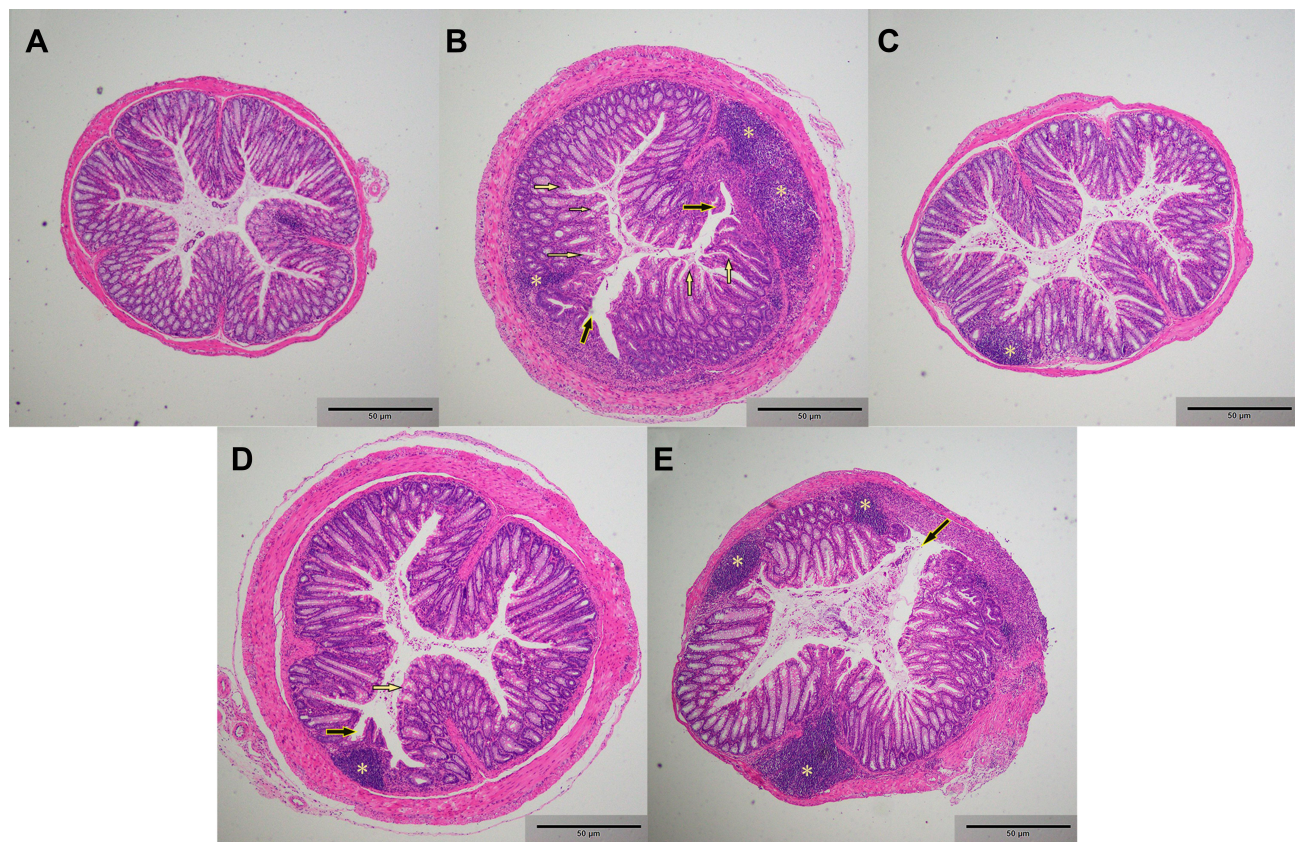


Figure 4 Morphological observation of colon. The asterisk, black arrow and yellow arrow in the picture respectively indicates interstitial lymphocyte infiltration, villous defect, and the crypt structural changes. (A): C-G, (B): DSS-G, (C): LA-G, (D): SSZ-G, (E): BB-G. Scale bar: 50 μ m.

and the ratio of colonic weight to colonic length was higher in the DSS model mice than that in normal mice.^{1,19,20} The colonic length was significantly longer in the LA-G than in the DSS-G, and the ratio of colonic weight/length was smaller in the LA-G than in the DSS-G. The pathological sections of the colon tissue further showed that LA-XY27 treatment significantly reduced the infiltration and proliferation of lymphocytes caused by inflammation, which preliminarily proved the anti-inflammatory effect of LA-XY27 on DSS-induced colitis. In this study, we observed treatment with LA-XY27 could not only relieve shortened colon and reduce the

ratio of colonic weight/length but also prevented mucosal damage, and remitted inflammatory cells infiltration. Therefore, we speculated that LA-XY27 may have a good effect on the intestinal mucosa.

Colitis leads to a rapid increase in the contents of free radicals such as reactive oxygen species (ROS) and reactive nitrogen species (RNS), further aggravating the damage and toxicity in the colon tissue, followed by a significant increase in the contents of peroxidation products MDA and MPO and a decrease in the levels of antioxidants SOD and CAT.^{21,22} Therefore, the antioxidant

Table 4 Determination of Oxidation Indexes in the Serum

Group	CAT (U/mL)	T-SOD (U/mL)	MDA (nmL/mL)	MPO(U/mL)
C-G	589.33 \pm 61.9 ^{##}	91.88 \pm 5.12 [#]	5.50 \pm 2.69 ^{###}	786.62 \pm 60.5 ^{###}
DSS-G	152.96 \pm 53.9 ^{**}	64.79 \pm 8.51 [*]	10.58 \pm 2.95 ^{**}	830.04 \pm 71.1 ^{**}
LA-G	536.58 \pm 68.1 ^{***}	89.86 \pm 2.15 ^{**}	5.59 \pm 2.59 ^{###}	794.64 \pm 51.9 ^{###}
SSZ-G	477.68 \pm 103.1 ^{***}	89.96 \pm 6.52 ^{**}	6.59 \pm 3.69 [#]	795.22 \pm 55.5 ^{###}
BB-G	219.69 \pm 86.6 ^{**}	87.99 \pm 3.34 ^{**}	6.81 \pm 2.67 [#]	797.34 \pm 49.0 ^{###}

Notes: Values are presented as mean \pm standard deviation (n = 8). *Means p < 0.05 compared with the C-G, # means p < 0.05 compared with the DSS-G. **Means p < 0.01 compared with the C-G, ### means p < 0.01 compared with the DSS-G.

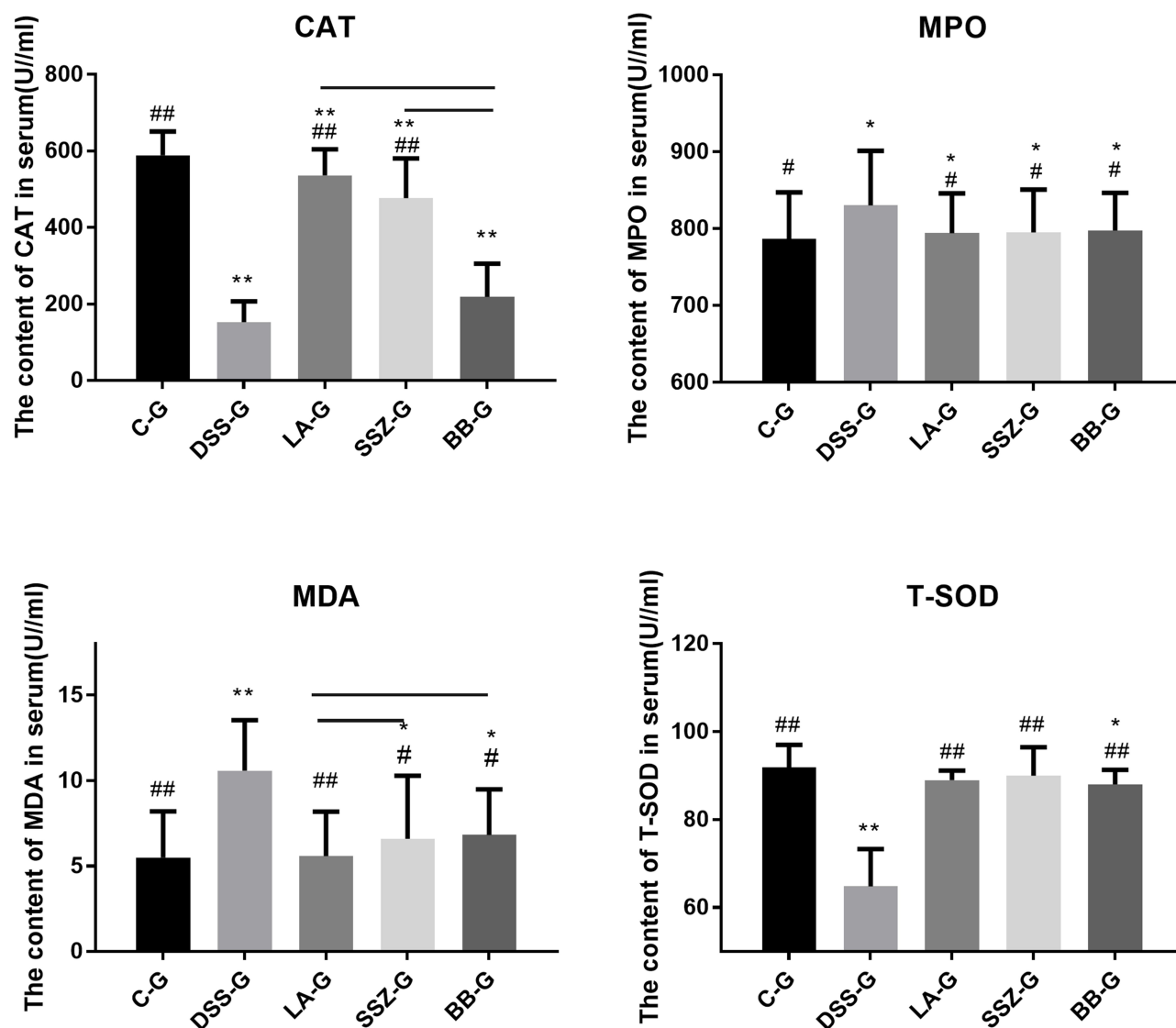


Figure 5 Oxidative stress factors in the serum. All the data were presented as mean \pm SD (n=8). *Represents a significant difference compared with the C-G, #represents a significant difference compared with the DSS-G, Horizontal lines were used to indicate whether there was a statistical difference between the three intervention groups; * or # means $p < 0.05$, ** or ### means $p < 0.01$.

Abbreviations: C-G, control group; DSS-G, DSS-induced group; LA-G, LA-XY27 group; SSZ-G, salicylazosulfapyridine group; BB-G, *L. bulgaricus* group.

and anti-inflammatory effects of LA-XY27 were verified by detecting the related oxidation indexes and inflammatory factors in serum and colon tissue. CAT is a marker

enzyme of peroxisomes, which exists in the peroxides of cells in various tissues of all known animals. CAT is one of the key enzymes in the biological defense system with

Table 5 Serum Inflammatory Cytokines

Group	IFN- γ (ng/L)	IL-1 β (ng/L)	IL-6 (pg/mL)	IL-10 (pg/mL)	TNF- α (ng/L)	IL-12 (ng/L)
C-G	5.5681 \pm 1.17 ^{###}	7.0394 \pm 1.16 ^{###}	5.3286 \pm 0.92 ^{###}	8.5216 \pm 1.21 ^{###}	1.4822 \pm 1.04 ^{###}	4.8204 \pm 1.79 ^{###}
DSS-G	7.6742 \pm 1.21 ^{**}	9.2506 \pm 1.02 ^{**}	8.5388 \pm 1.46 ^{**}	4.2114 \pm 1.39 ^{**}	3.8984 \pm 2.08 ^{**}	7.7288 \pm 1.65 ^{**}
LA-G	5.6702 \pm 1.63 ^{###}	8.0422 \pm 1.88 [#]	6.2290 \pm 0.132 ^{*#}	6.7188 \pm 1.10 ^{###}	1.7868 \pm 2.03 ^{###}	5.3218 \pm 1.97 ^{###}
SSZ-G	6.6724 \pm 1.18 ^{*#}	8.6497 \pm 1.35 ^{*#}	7.6342 \pm 1.11 [*]	5.8170 \pm 1.50 ^{**###}	1.8920 \pm 1.72 ^{###}	5.8230 \pm 1.57 ^{###}
BB-G	6.0730 \pm 1.78 [#]	8.8499 \pm 2.21 ^{*#}	7.3326 \pm 1.88 [*]	5.5178 \pm 1.19 ^{**###}	2.3884 \pm 1.14 ^{###}	6.4224 \pm 1.01 [#]

Notes: Values are presented as mean \pm standard deviation (n = 8). *Means $p < 0.05$ compared with the C-G, #means $p < 0.05$ compared with the DSS-G. **Means $p < 0.01$ compared with the C-G, ###means $p < 0.01$ compared with the DSS-G.

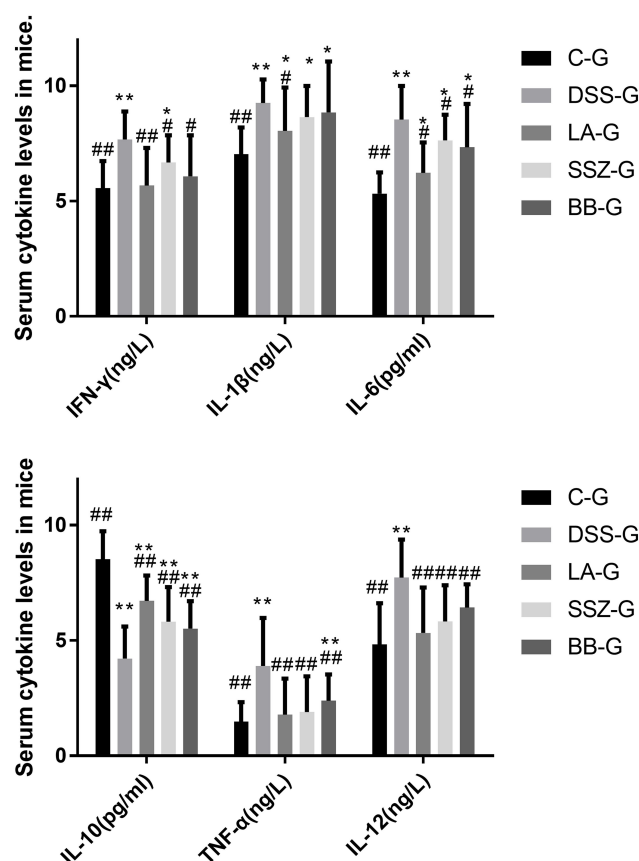


Figure 6 Inflammatory factors in serum. All the data were presented as mean \pm SD (n=8). *Represents a significant difference compared with the C-G, #represents a significant difference compared with the DSS-G, *or # means $p < 0.05$, ** or ### means $p < 0.01$.

Abbreviations: C-G, control group; DSS-G, DSS-induced group; LA-G, LA-XY27 group; SSZ-G, salicylazosulfapyridine group; BB-G, *L. bulgaricus* group.

strong antioxidant function. It can decompose hydrogen peroxide into water and oxygen out of the body, reduce the concentration and toxicity of hydrogen peroxide, and

protect cells and tissues from oxidative stress. MPO is a kind of heme protein, which is released by the accumulation and dispersion of neutrophils caused by inflammatory stimulation. The significant increase in MPO level indicates the inflammatory response.⁴ The decrease in MPO activity represents the decrease in neutrophil aggregation in inflamed tissues. In normal cells, SOD protects the body from tissue damage caused by free radicals, and inhibiting the decrease in the SOD level in the body can effectively alleviate the injury of the body.²³ The aforementioned indexes are biomarkers of oxidative stress. In this study, the levels of CAT and T-SOD in the serum increased and the levels of MDA and MPO effectively reduced in all intervention groups, while the effect of *L. acidophilus* XY27 was better than that of *L. bulgaricus* and salicylazosulfapyridine.

Inflammatory mediators, especially cytokines, mediate immune cells to regulate inflammatory response and balance intestinal mucosal barrier function. Many pieces of literature have reported that probiotics for intestinal flora can improve intestinal barrier function by regulating inflammatory factors.^{23–25} In the present study, LA-XY27 treatment could effectively reduce the levels of many pro-inflammatory cytokines, including IFN-γ, IL-1 β, IL-6, TNF-α, and IL-12, in the serum, and increase the levels of anti-inflammatory cytokine IL-10. This result was further verified by gene expression in the colon of mice with colitis. In addition, the relative expression levels of COX-2, EIK-1, *E. coli*, LPS, and p100 genes in mice with colitis treated with LA-XY27 were significantly lower than those in the model group, while the relative expression levels of protective factors ZO-1, NF-κB, p53, and

Table 6 Gene Expression in the Colon Tissue

Gene Name	C-G	DSS-G	LA-G	SSZ-G	BB-G
EIK-1	0.8561 \pm 0.086###	1 \pm 0**	0.9384 \pm 0.077###	0.9644 \pm 0.050****	0.9643 \pm 0.022###
COX-2	0.8073 \pm 0.014###	1 \pm 0**	0.9140 \pm 0.365****	0.9876 \pm 0.068**	0.9591 \pm 0.0494****
LPS	0.9020 \pm 0.074###	1 \pm 0**	0.9453 \pm 0.133****	0.9846 \pm 0.017****	0.9568 \pm 0.054**
<i>Escherichia coli</i>	0.1428 \pm 0.016###	1 \pm 0**	0.2339 \pm 0.023****	0.2798 \pm 0.012****	0.2943 \pm 0.198###
IL-12	0.4902 \pm 0.035###	1 \pm 0**	0.6046 \pm 0.099****	0.7491 \pm 0.055****	0.7926 \pm 0.042###
NF-κB	42.97 \pm 13.34###	1 \pm 0**	19.906 \pm 4.760****	2.7016 \pm 0.648****	1.1821 \pm 0.073**
p100	0.9703 \pm 0.065#	1 \pm 0**	0.9921 \pm 0.028**	0.9987 \pm 0.057*	0.9940 \pm 0.038*
p53	1.1807 \pm 0.052###	1 \pm 0**	1.0794 \pm 0.110****	1.0405 \pm 0.112****	1.0619 \pm 0.014###
IκB-α	10.2913 \pm 0.503###	1 \pm 0.92**	2.5118 \pm 0.044****	1.2174 \pm 0.110**	1.8500 \pm 0.06###
TNF-α	0.9241 \pm 0.164###	1 \pm 0**	0.9644 \pm 0.041****	0.9792 \pm 0.158****	0.9763 \pm 0.110###
IL-1β	0.3705 \pm 0.404###	1 \pm 0**	0.6221 \pm 0.172****	0.7873 \pm 0.182****	0.7835 \pm 0.020###
ZO-1	2.7308 \pm 0.346###	1 \pm 0**	1.9520 \pm 0.658****	1.2500 \pm 0.1637****	1.5747 \pm 0.060###

Notes: Values are presented as mean \pm standard deviation (n = 8). *Means $p < 0.05$ compared with the C-G, # means $p < 0.05$ compared with the DSS-G. **Means $p < 0.01$ compared with the C-G, ### means $p < 0.01$ compared with the DSS-G.

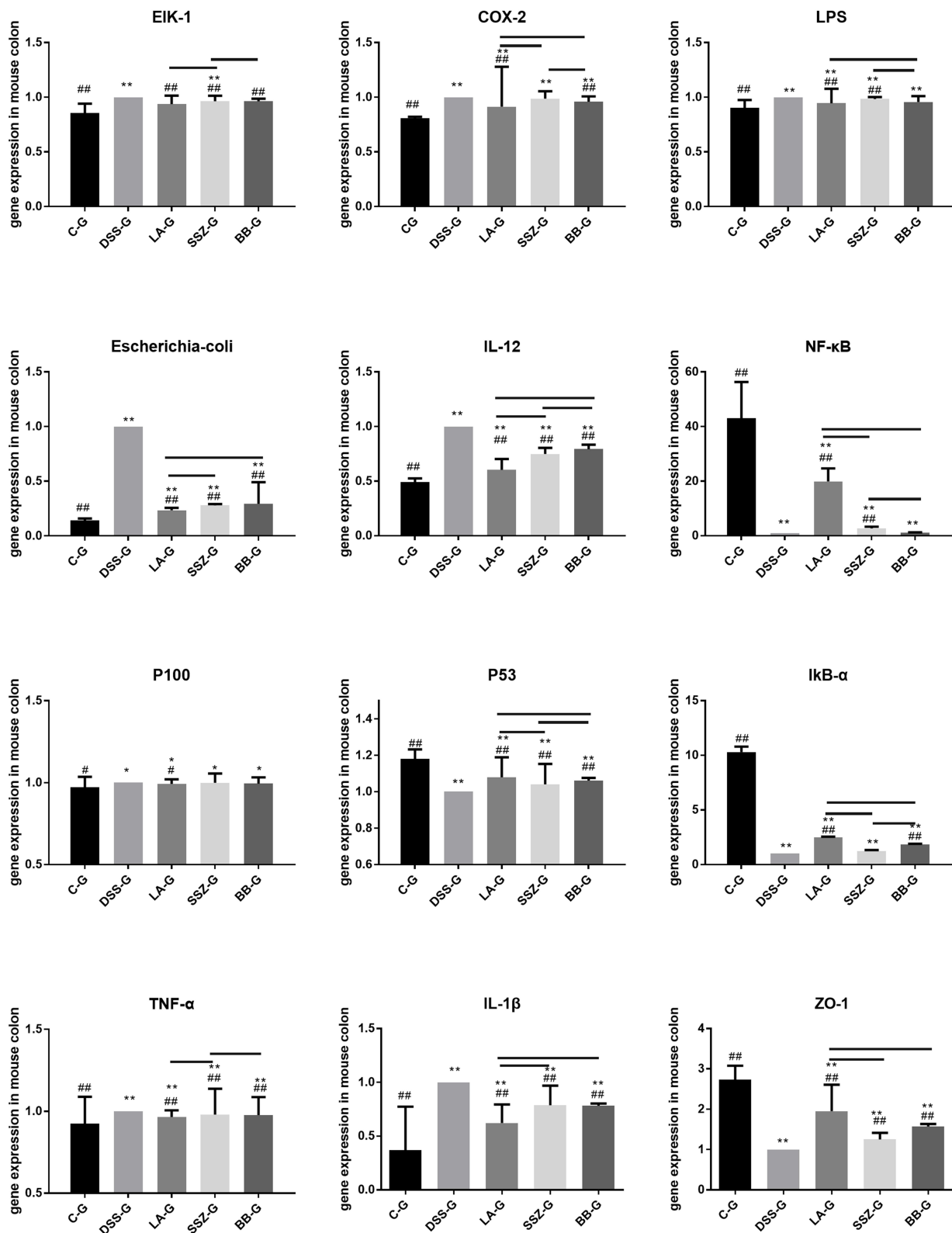


Figure 7 Gene expression in the colon. All the data were presented as mean \pm SD (n=8). *Represents a significant difference compared with the C-G, #represents a significant difference compared with the DSS-G, Horizontal lines were used to indicate whether there was a statistical difference between the three intervention groups; * or # means p < 0.05, ** or ## means p < 0.01.

Abbreviations: C-G, control group; DSS-G, DSS-induced group; LA-G, LA-XY27 group; SSZ-G, salicylazosulfapyridine group; BB-G, *L. bulgaricus* group.

IKB- α significantly increased. The decrease in the expression of the COX-2 gene also downregulated the expression of pro-inflammatory factors, inhibited inflammatory response, reduced intestinal inflammatory response and immune response, blocked inflammatory factors, and activated the NF- κ B pathway, which was important in inhibiting the local inflammatory response and treating UC. LPS is a potential candidate for endotoxin antagonists.²⁶ Recent studies revealed that LPS from some kind of bacteria is not only the inducer of pro-inflammatory cytokines, such as TNF- α , but also a competitive endotoxin antagonist with a universal type of action.²⁷

In this experiment, the effect of LA-XY27 seems to be better than that of *L. bulgaricus*, which may be due to the species specificity. Some authors conducted a meta-analysis on the effects of different kinds of probiotics on UC by analyzing 27 trials, demonstrating that probiotics were generally effective in treating CD and UC.^{23,28} *Lactobacillus* probiotics are more effective against UC.¹⁰ Although the effect is positive, the effective strains are still very limited, and the clinical efficacy and evidence are not sufficient.²⁹ The strains in this experiment were isolated and purified from herdsmen in Xinyuan County, Northwest Xinjiang Uygur Autonomous region, where the climate and geographical conditions were unique. For a long time, traditional yoghurt produced by herdsmen has a good health-care effect. Our preliminary experiments also proved that the *Lactobacillus fermentum* Suo isolated from yak yoghurt of Hongyuan grassland (Ngawa Tibetan and Qiang Autonomous Prefecture, Aba, Sichuan, China) house and attenuates HCl/Ethanol-induced gastric injury in mice through its antioxidant effects.³⁰ Another *Lactobacillus fermentum* HY01 (LF-HY01) isolated from traditional fermented yak yoghurt of Hongyuan has been proven to have prevented effects on dextran sulfate sodium-induced colitis in mice.³¹

The results suggested that LA-XY27 prevented and reduced the symptoms of colitis mainly by downregulating the levels of pro-inflammatory factors, enhancing anti-inflammatory factors and antioxidant capacity, and restoring intestinal barrier function. The effective mechanism of LA-XY27 is worth exploration. However, meta-analyses provided limited evidence for using *L. acidophilus* to treat acute gastroenteritis in children. Therefore, more large-scale, high-quality clinical trials should be conducted to verify the clinical application feasibility of the strains.^{32,33} The findings of this in vitro study might provide the basis and reference for future clinical research.

Conclusions

L. acidophilus XY27 had a better effect compared with *L. bulgaricus* and salicylazosulfapyridine in preventing DSS-induced colitis and relieving the symptoms of colitis in mice in this study. *L. acidophilus* XY27 exerted its effect mainly by enhancing the antioxidant capacity, repairing the intestinal barrier, and regulating the expression of inflammation-related genes and the secretion of inflammatory factors.

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Disclosure

The authors report no conflicts of interest in this work.

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