ORIGINAL ARTICLE



A phase I dose escalation, dose expansion and pharmacokinetic trial of gemcitabine and alisertib in advanced solid tumors and pancreatic cancer

Justin A. Chen¹ · Jasmine C. Huynh¹ · Chun-Yi Wu² · Ai-Ming Yu³ · Karen Matsukuma⁴ · Thomas J. Semrad⁵ · David R. Gandara¹ · Tianhong Li¹ · Jonathan W. Riess¹ · Kit Tam¹ · Philip C. Mack⁶ · Anthony Martinez⁷ · Nichole Mahaffey⁷ · Karen L. Kelly¹ · Edward J. Kim¹

Received: 24 February 2022 / Accepted: 8 July 2022 / Published online: 30 July 2022 $\ensuremath{\mathbb{C}}$ The Author(s) 2022

Abstract

Purpose Aurora Kinase A (AKA) inhibition with gemcitabine represents a potentially synergistic cancer treatment strategy via mitotic catastrophe. The feasibility, safety, and preliminary efficacy of alisertib (MLN8237), an oral AKA inhibitor, with gemcitabine was evaluated in this open-label phase I trial with dose escalation and expansion.

Methods Key inclusion criteria included advanced solid tumor with any number of prior chemotherapy regimens in the dose escalation phase, and advanced pancreatic adenocarcinoma with up to two prior chemotherapy regimens. Four dose levels (DLs 1–4) of alisertib (20, 30, 40, or 50 mg) were evaluated in 3 + 3 design with gencitabine 1000 mg/m² on days 1, 8, and 15 in 28-day cycles.

Results In total, 21 subjects were treated in dose escalation and 5 subjects were treated in dose expansion at DL4. Doselimiting toxicities were observed in 1 of 6 subjects each in DL3 and DL4. All subjects experienced treatment-related adverse events. Grade ≥ 3 treatment-related adverse events were observed in 73% of subjects, with neutropenia observed in 54%. Out of 22 subjects evaluable for response, 2 subjects (9%) had partial response and 14 subjects (64%) had stable disease. Median PFS was 4.1 months (95% CI 2.1–4.5). No significant changes in pharmacokinetic parameters for gencitabine or its metabolite dFdU were observed with alisertib co-administration.

Conclusions This trial established the recommended phase 2 dose of alisertib 50 mg to be combined with gemcitabine. Gemcitabine and alisertib are a feasible strategy with potential for disease control in multiple heavily pre-treated tumors, though gastrointestinal and hematologic toxicity was apparent.

Keywords Alisertib · Gemcitabine · Pharmacokinetics · Aurora kinase a · Phase I

Edward J. Kim jhkim@ucdavis.edu

- Division of Hematology and Oncology, Davis Comprehensive Cancer Center, University of California, 4501 X Street, Suite 3016, Sacramento, CA 95817, USA
- ² Bioanalysis and Pharmacokinetics Core Facility, University of California, Sacramento, CA 95817, USA
- ³ Department of Biochemistry and Molecular Medicine, University of California, Sacramento, CA 95817, USA
- ⁴ Department of Pathology and Laboratory Medicine, University of California, Sacramento, CA 95817, USA

- ⁵ Gene Upshaw Memorial Tahoe Forest Cancer Center, Truckee, CA 96161, USA
- ⁶ Division of Hematology and Medical Oncology, Icahn School of Medicine at Mount Sinai, New York, NY 10029, USA
- ⁷ Office of Clinical Research, Davis Comprehensive Cancer Center, University of California, Sacramento, CA 95817, USA

Introduction

Taxanes stabilize microtubules to disrupt the dynamic polymerization and depolymerization necessary for mitosis. However, alteration of normal microtubule dynamics can cause side effects such as neuropathy which is a dose-limiting toxicity of taxanes. Newer generations of mitotic inhibitors are being developed to target proteins present only in cells undergoing active mitosis, thus limiting off-target effects noted with taxanes such as neuropathy.

Aurora Kinase A (AKA) is a member of the Aurora Kinase family of serine/threonine protein kinases. AKA is a serine/threonine kinase highly expressed during G2 transition to mitosis that supports assembly of spindle microtubules and facilitates centrosome maturation [1, 2]. Overexpression of AKA can lead to chromosomal instability [3–5] and has been observed in multiple solid cancers [4, 6–8]. Inhibition of AKA in pancreatic cancer cells causes increased mitotic arrest and apoptosis, leading to decreased proliferation and tumorigenicity [9], with similar apoptotic synergy observed when added to EGFR inhibition in resistant pre-clinical models of *EGFR*-driven NSCLC [8].

Alisertib (MLN-8237) is a potent, highly selective smallmolecule inhibitor of targeting the ATP-binding site of AKA with > 200-fold selectivity for AKA compared to Aurora Kinase B [10, 11]. Alisertib, which has yet to be approved for any indication and remains an investigational agent, has demonstrated modest single-agent activity in phase I trials for solid tumors [12, 13]. In vitro and in vivo solid tumor models have suggested enhanced anti-tumor activity with chemotherapy combinations [14–16], and alisertib plus paclitaxel has shown a trend for improved efficacy compared to paclitaxel alone in patients with small cell lung cancer [17]. A proposed mechanism of synergy is synthetic lethality leading to mitotic catastrophe [18].

Gemcitabine has been suggested to deplete ATP, which may augment alisertib binding to the ATP-site of AKA [19]. Additionally, AKA upregulates NF- κ B whose expression can be suppressed with aurora kinase inhibition [20, 21]. This is especially important in pancreatic cancer which usually demonstrates NF- κ B activation, and is consistent with downstream signaling activation from Kras mutations which are present in the majority of pancreatic cancers [22, 23]. To our knowledge, the combination of alisertib and gemcitabine has not yet been tested for treatment of pancreatic cancer.

Materials and methods

This clinical trial (NCT01924260) was conducted following all applicable regulatory requirements and was approved by the UC Davis Institutional Review Board. All participating subjects provided written informed consent prior to initiation of trial-associated procedures and treatment.

Study design and treatment

This was an open-label phase I clinical trial with two-phase design including a dose escalation and an expansion phase in pancreatic cancer. In the dose escalation phase, a standard 3+3 design was used to determine the maximum tolerated dose (MTD) of alisertib in combination with gemcitabine. Alisertib was administered orally twice daily (BID) on days 1-3, 8-10, and 15-17 of a 28-day treatment cycle. Gemcitabine was given concurrently at standard dosing of 1000 mg/ m² intravenously on days 1, 8, 15. A starting dose of alisertib 20 mg BID was used in the dose escalation phase and was escalated in cohorts of at least three evaluable subjects at 30 mg BID, 40 mg BID, and 50 mg BID until MTD or the highest feasible dose level (Table S1). The dose of 50 mg BID was the highest dose allowable and was previously reported as the recommended phase II dose (RP2D) for alisertib monotherapy [12, 24, 25]. In the expansion phase, subjects received the MTD or RP2D following the same cycle schedule. Dose adjustments were allowed for both drugs (Table S1B, 1C).

Treatment with alisertib and gemcitabine was repeated every 28 days. To proceed to the next cycle, lab parameters included ANC \geq 1500/mm³ and platelet count \geq 100,000/ mm³, and all other toxicity considered by the investigator to be related to therapy with alisertib or gemcitabine must have resolved to grade \leq 1 or to the subject's baseline values. If the subject failed to meet the above-cited criteria for initiating a cycle, then the next treatment was delayed for up to 1 week. Thereafter, the subject was re-evaluated to determine continuation eligibility. Dose modification (Table S1B, 1C) was required for cycle initiations delayed > 1 week due to incomplete recovery from treatment-related toxicity.

Objectives and statistical considerations

The primary objective of this phase I study was to determine the MTD and RP2D of alisertib in combination with gemcitabine. Upon determination of the MTD, an expansion cohort of subjects with pancreatic cancer was enrolled to further evaluate safety and evidence of clinical activity. Secondary objectives included preliminary efficacy as determined by objective response rate (ORR) and progression-free survival (PFS), and effects of alisertib drug–drug interactions on the pharmacokinetics of gemcitabine and its primary metabolite, 2', 2'-difluorodeoxyuridine (dFdU).

The MTD was defined as the highest dose tested in which fewer than 33% of subjects experienced DLT attributable to the study drugs, when at least six subjects were treated at that dose and evaluable for toxicity. The RP2D was to be selected based on the totality of safety and efficacy data, and did not necessarily equal the MTD. If the MTD was not reached by dose level 4, then that dose level would be the RP2D.

After identification of the MTD or RP2D, goal accrual was 16 subjects at that dose (6 patients in dose escalation and 10 patients in dose expansion) which allowed for an 81% chance of seeing at least 1 example of any toxicity that occurs in 10% or more of similar subjects, and a 93% chance of seeing at least one example of any toxicity that occurs in 15% or more of similar subjects.

Subject selection

Eligible subjects were required to be ≥ 18 years of age, have an Eastern Cooperative Oncology Group (ECOG) performance status of 0 to 2, and be able to swallow and retain oral medications. Female subjects of childbearing age were required to be willing to use effective birth control for the duration of the study. Male subjects were required to agree to use effective contraception during the entire study and for 4 months after the last dose of alisertib. Other eligibility criteria included: adequate bone marrow defined as absolute neutrophil count (ANC) > $1500/\text{mm}^3$ and platelet count \geq 100,000/mm³, adequate hepatic function defined as total bilirubin with institutional normal limits and ALT and AST < 2.5 times institutional upper limit of normal or ≤ 5 times institutional upper limit of normal in presence of liver metastases, and adequate renal function defined as creatinine ≤ 1.5 times institutional upper limit of normal or creatinine clearance $> 60 \text{ ml/min}/1.73\text{m}^2$ measured by 24-h urine collection. Prior treatment with chemotherapy, immunotherapy, targeted therapy, or radiation must have been completed at least 2 weeks prior to start of protocol treatment and side effects related to prior treatment (excluding alopecia, lymphopenia, and hyperglycemia) resolved to grade ≤ 1 . Prior gemcitabine-based regimens in the palliative setting were permitted if there was no evidence of progression on therapy or at least 6 months had elapsed after discontinuation of gemcitabine-based treatment. Prior gemcitabine in the adjuvant setting was permitted if the last treatment was greater than 6 months prior to registration.

In addition to the above criteria, criteria specific to subjects enrolled in the dose escalation phase of the trial included histologically or cytologically confirmed metastatic or unresectable solid tumor and any number of prior chemotherapies. Subjects enrolled in the dose expansion phase were required to have histologically or cytologically confirmed metastatic or unresectable pancreatic adenocarcinoma, up to 2 prior chemotherapy regimens in the palliative setting. Measurable disease was only required for the dose expansion phase.

Exclusion criteria for both cohorts included: prior treatment with AKA-targeted drugs, history of Gilbert's syndrome (due to metabolism of alisertib via glucuronidation), significant history of cardiac disease, symptomatic or uncontrolled brain metastases, prior radiation to greater than 25% of bone marrow or whole pelvis radiation, anticoagulation with warfarin, active clinical infection including active HIV, chronic hepatitis B, and pregnant or breast-feeding female subjects.

All subjects provided written consent. The study was approved by the UC Davis institutional review board (IRB) and was compliant with Good Clinical Practices guidelines and the Declaration of Helsinki.

Safety and efficacy assessments

Safety was monitored by performing physical examination and assessing vital signs, performance status, laboratory evaluations and an ECG as well as by collecting adverse events at every study visit. Toxicity was evaluated according to the National Cancer Institute Common Terminology Criteria for Adverse Events (NCI CTCAE, version 4.02). All subjects receiving any amount of study drug were evaluable for toxicity.

DLT was defined as any related (possibly, probably, or definitely) grade 3 non-hematologic toxicity or any attributable grade 4 toxicity. Grade 3 nausea or emesis was not considered dose-limiting unless it did not reverse to grade ≤ 2 within 96 h of appropriate management. Grade 3 fatigue was not considered dose-limiting unless it did not reverse to grade ≤ 2 in 7 days. Transient grade 4 neutropenia was not considered dose-limiting unless it did not resolve to grade 3 within 7 days or was associated with febrile neutropenia. DLT assessment was based on the first cycle of treatment. To be evaluable for DLT, a subject had to receive $\geq 80\%$ of the total intended dose of both gemcitabine and alisertib and be observed for at least 3 weeks after the start of the first cycle or have experienced a DLT. All subjects enrolled were followed for DLT, and any subjects who were not evaluable for toxicity were replaced.

Response was assessed using CT scans of the chest, abdomen, and pelvis (MRI could be substituted for abdomen and pelvis) at baseline and every 2 cycles on study per the Response Evaluation Criteria in Solid Tumors guideline (RECIST, version 1.1). PFS was evaluated using Kaplan–Meier survival analysis.

Adverse events were summarized according to organ system, laboratory category, and dose level in frequency tables graded according to CTCAE v4.02. Information regarding each subject's course including completion of therapy, dose delays, premature discontinuation, and major protocol violations were tabulated and summarized.

 Table 1 Baseline demographic and clinical information for all subjects

Characteristic	Total	Dose escalation	Dose expansion $(N=5)$	
	(N=26)	(N=21)		
Age-median (range)	57 (42-82)	57 (42–75)	63 (48-82)	
Sex				
Male	13	10	3	
Female	13	11	2	
ECOG performance s	tatus			
0	9	7	2	
1	16	13	3	
2	1	1		
Primary diagnosis				
NSCLC	7	7		
Colorectal	3	3		
Neuroendocrine (poorly differen- tiated)	3	3		
SCLC	2	2		
Head and neck	2	2		
Pancreas	6	1	5	
Gallbladder	1	1		
Small bowel	1	1		
Mesothelioma	1	1		
Prior lines of chemo				
0-1	6	4	2	
2–3	16	13	3	
4+	4	4		
Assigned dose level				
1	3	3		
2	3	3		
3	6	6		
4	14	9	5	

Pharmacokinetic (PK) assessments

Pharmacokinetic sampling was performed on the expanded cohort of subjects with pancreatic cancer treated at the RP2D. Alisertib was not administered on cycle 1, day 1 to enable gemcitabine pharmacokinetic sampling on day 1 and day 2 of cycle 1 in the absence of co-administered alisertib. This served as the reference baseline for comparison to gemcitabine pharmacokinetics when co-administered with alisertib on day 8 of cycle 1. On cycle 1 day 8, alisertib was administered within 10 min of the start of the gemcitabine infusion, and the second (evening) dose of alisertib was not administered to allow for gemcitabine measurement at the 24 h timepoint. Both the parent drug (gemcitabine, dFdC) and the metabolite (dFdU) were measured to evaluate changes in gemcitabine metabolism related to alisertib administration. Pharmacokinetic blood samples were collected with dipotassium ethylenediaminetetraacetic acid (K2EDTA) as the anticoagulant before treatment, immediately after gemcitabine infusion, and at additional prespecified post-infusion timepoints. Actual timepoints were recorded for PK parameter evaluation. Measurement of plasma gemcitabine, dFdU, and alisertib concentrations are described in the Supplementary Methods.

Gemcitabine, dFdU and alisertib PK parameters were estimated using non-compartmental analysis (NCA) with Certara Phoenix WinNonlin 8.0 (Princeton, NJ), including mean peak concentration (C_{max}) and exposure (Area Under the Curve, AUC). The AUC parameters were estimated using the linear-up-log-down trapezoidal rule. Tests of significance between day 1 and day 8 PK parameters were performed using paired t-tests with GraphPad Prism 6.07 (San Diego, CA).

AKA and pHH3 immunohistochemical staining assessments

When available, subject archival tumor specimens were collected for immunohistochemical analysis of AKA level and proliferative index assessment. Paraffin-embedded tumor sections were cut at 5 μ m and immunostained for AKA (mouse clone JLM28, Leica Biosystems, Buffalo Grove, IL; manual detection using a mouse HRP polymer) and phosphohistone H3 (pHH3, rabbit polyclonal, Millipore Sigma, Burlington, MA; manual detection using a rabbit HRP polymer). pHH3 is a marker of mitotic activity and a more specific marker than Ki-67, which is expressed during all active phases of the cell cycle.

AKA and pHH3 stained slides were scored using the following criteria: each tumor was scanned for "hot spots" of AKA expression, defined as areas of tumor with the highest density of immunopositive tumor nuclei according to visual scanning at low-power magnification. Nuclear staining (with or without cytoplasmic staining) of at least 2 + (intermediate) intensity (range 0–3, as per Allred scoring method [26]) was considered positive. Each hot spot was photocaptured at 200× magnification at the same level of illumination and individual tumor nuclei were counted. Tumor nuclei with 1+ or less intensity nuclear staining were counted as negative. A minimum of 500 cells were counted for each tumor (range 500–2082). pHH3 expression was assessed in the exact same area that the AKA count was performed using the same criteria. **Fig. 1** Proportion of planned dose delivered for alisertib (blue) and gemcitabine (red) in all treatment cycles for all subjects. For each cycle, the proportion of planned dose was calculated by the total dose delivered divided by total planned dose according to alisertib dose level and gemcitabine 1000 mg/m²



Results

Clinical characteristics

Between August 2013 and October 2016, a total of 26 subjects (median age 57 years, 13 men, 13 women) were enrolled at UC Davis Comprehensive Cancer Center. Subject characteristics are shown in Table 1. Twenty-one subjects were enrolled in the dose escalation phase with an additional 5 subjects enrolled and treated in the expansion phase. A total of 14 subjects were treated at dose level 4 (9 in dose escalation and 5 in dose expansion). Enrollment into dose expansion was discontinued due to poor accrual. Primary malignancies included non-small cell lung cancer (NSCLC), pancreatic cancer, and colorectal cancer, among others. A majority of subjects (77%) were treated with at least 2 prior lines of systemic therapy.

DLT and MTD

In the dose escalation phase, 21 subjects were evaluable for DLT. DLTs were observed in one out of 6 subjects treated at dose level 3 (grade 3 urinary tract infection; subject 7) and 1 out of 6 subjects treated at dose level 4 (grade 3 mucositis, lymphopenia, leukopenia, febrile neutropenia, hyponatremia, and dehydration; subject 19). Subject 7 was hospitalized on cycle 1 day 7 for septic shock from presumed urinary source requiring 3-day hospitalization for pressor support, fluid hydration, and antibiotics. Urinary tract infection was felt possibly related to both drugs. Subject 19 presented on cycle 1 day 8 with neutropenic fever, mucositis, hyponatremia, and dehydration. Despite antibiotics and resolution of neutropenia, fevers persisted and no infectious source was identified, although the subject was treated empirically for candida esophagitis. This subject was ultimately discharged on hospice. All grade 3 events were felt possibly or probably related to either study drugs. No DLTs were observed at dose level 1 or 2. The MTD and RP2D was determined to be dose level 4.

Treatment exposures, delays, and dose reductions

A total number of 94 cycles of alisertib and gemcitabine were delivered among the 26 subjects enrolled. Median number of cycles per subject was 3 (range 1–13), and median duration of treatment was 2.84 months (range 0.36–13.22). Similar median duration of treatment and cycles delivered were observed between the dose levels (Table S2). Proportion of planned dose received, defined as total dose received divided by total planned dose according to the dose level for alisertib and 1000 mg/m² for gemcitabine, was calculated according to each subject (Table S3) and each cycle (Fig. 1).

For alisertib, 18 subjects underwent dose delays/omissions and 9 subjects underwent dose reductions. A total of 27 cycles of alisertib were modified. For gemcitabine, 16 subjects required dose delays, 14 subjects required dose omission, and 5 subjects required dose reductions. A total

Table 2 Treatment-related adverse events occurring in > 10% of subjects or serious adverse events

Event, $N(\%)$	N(%) All subjects ($N=26$)		Dose level 4 ($N = 14$)			
	Any grade	Grade ≥ 3	Any grade	$Grade \ge 3$		
Any	26 (100)	24 (92)	14 (100)	14 (100)		
Hematologic						
Anemia	21 (81)	8 (31)	12 (86)	2 (14)		
Leukopenia	26 (100)	15 (58)	14 (100)	10 (71)		
Lymphopenia	20 (77)	12 (46)	10 (71)	6 (43)		
Lymphocytosis	2 (8)	2 (8)	2 (14)	2 (14)		
Neutropenia	23 (88)	17 (65)	14 (100)	11 (79)		
Febrile neutropenia	1 (4)	1 (4)	1 (7)	1 (7)		
Thrombocytopenia	23 (88)	4 (15)	13 (93)	2 (14)		
Constitutional						
Fatigue	18 (69)	1 (4)	11 (79)	1 (7)		
Myalgia	2 (8)	1 (4)	1 (7)	1 (7)		
Metabolic						
Total bilirubin elevated	3 (12)	0	1 (7)	0		
AST elevated	15 (58)	1 (4)	7 (50)	1 (7)		
ALT elevated	16 (62)	1 (4)	7 (50)	1 (7)		
ALP elevated	11 (42)	1 (4)	6 (43)	1 (7)		
Creatinine elevated	6 (23)	1 (4)	1 (7)	1 (7)		
Albumin decreased	11 (42)	1 (4)	5 (36)	0		
Hypoglycemia	4 (15)	0	2 (14)	0		
Potassium decreased	8 (31)	1 (4)	5 (36)	1(7)		
Magnesium decreased	3 (12)	0	1 (7)	0		
Sodium decreased	11 (42)	5 (19)	6 (43)	3 (21)		
Phosphate decreased	5 (19)	2 (8)	3 (21)	1 (7)		
Gastrointestinal						
Mucositis oral	16 (62)	6 (23)	11 (79)	3 (21)		
Oral pain	3 (12)	0	2 (14)	0		
Nausea	14 (54)	1 (4)	10 (71)	1 (7)		
Vomiting	10 (38)	1 (4)	5 (36)	0		
Diarrhea	10 (58)	3 (12)	8 (57)	3 (21)		
Constipation	6 (23)	0	0	0		
Anorexia	5 (19)	0	4 (29)	0		
Dry mouth	2 (8)	0	2 (14)	0		
Dehydration	2 (8)	2 (8)	2 (14)	2(14)		
Esophagitis	1 (4)	1 (4)	1 (7)	1 (7)		
Dermatologic						
Alopecia	3 (12)	0	2(14)	0		
Pruritus	3 (12)	0	2 (14)	0		
Neurologic			. ,			
Dizziness	3 (12)	0	2(14)	0		
Headache	3 (12)	0	2 (14)	0		
Lymphatic	- (-)		~ /			
Edema, limbs	3 (12)	0	0	0		
Cardiac	× /					
Pericardial effusion	1 (4)	1 (4)	0	0		

Table 2 (d	continued)
------------	------------

Event, <i>N</i> (%)	All subject	s(N=26)	Dose level 4 ($N = 14$)		
	Any grade	$Grade \ge 3$	Any grade	Grade ≥ 3	
Respiratory					
Dyspnea	3 (12)	1 (4)	1 (4)	0	
Pleural effusion	1 (4)	0	0	0	
Infection					
Urinary tract infec- tion	1 (4)	1 (4)	0	0	

of 17 cycles had dose omissions, 10 cycles were given at a reduced dose, and 43 cycles had delayed doses. Although dose delays were common for gemcitabine, subjects still received the majority of the planned doses (Table S3, Fig. 1).

Safety and toxicity

Treatment-related adverse events (Table 2) were observed in all subjects and included leukopenia (100%), neutropenia (88%), thrombocytopenia (88%), anemia (81%), lymphopenia (77%), fatigue (69%), mucositis (62%), and ALT elevation (62%). Grade \geq 3 adverse events were observed in 92% of all subjects and predominantly hematologic, including neutropenia (65%), leukopenia (58%), lymphopenia (46%), and anemia (31%). Similar adverse events were seen at dose level 4, and all 14 subjects experienced grade \geq 3 adverse events and neutropenia, of which 79% were grade \geq 3 with one patient having febrile neutropenia.

Serious adverse events

Serious treatment-related adverse events were observed in 6 subjects (23%). Subject 2 had pericardial effusion (grade 4), pleural effusion (grade 2), and dyspnea (grade 3). Pericardial effusion occured during cycle 14, was not malignant, and felt possibly related to study drugs; however, this resulted in cardiac tamponade requiring a pericardial window. Subject 7 had a urinary tract infection (grade 3), which was classified as a DLT. Subject 22 had hyponatremia (grade 3), mucositis (grade 3), and hematologic abnormalities (grade 4 leukopenia, neutropenia, lymphopenia; grade 3 anemia). Subject 14 had anemia (grade 3), and subject 19 had febrile neutropenia (grade 3) and mucositis (grade 3). Subject 23 had dehydration (grade 3), diarrhea (grade 3), acute kidney injury (grade 3), and vomiting (grade 1).

Response

Response was evaluable in 22 subjects. Best response of partial response, stable disease, and progressive disease was

Fig. 2 A Waterfall plot of best responses per evaluable subject. Dashed lines represent +20% and -30% change from baseline. 1–4: corresponding dose levels, PD: progressive disease, SD: stable disease, PR: partial response. **B** Kaplan–Meier plot for progression-free survival. Median PFS was 4.1 months (95% CI 2.1–4.5)



observed in 2 (9%), 14 (64%), and 6 (27%) subjects, respectively (Fig. 2A). Responses were observed in one subject with lung adenocarcinoma and one subject with pancreatic

adenocarcinoma. Median PFS was 4.1 months (95% CI 2.1–4.5, Fig. 2B).

Eighteen subjects had archival tumor specimens evaluable for immunohistochemical evaluation of AKA and pHH3 **Table 3** Estimated PKparameters of non-compartmentanalysis for gemcitabine.Day 1 (without alisertib) andDay 8 (with alisertib) werecompared using paired t testingand no significant differenceswere found (p > 0.05) for (a)gemcitabine and (b) dFdU

Parameters	Unit	Day 1			Day 8		
		N	Mean	SD	N	Mean	SD
Gemcitabine							
C _{max}	mcg/mL	5	7.42	2.13	4	6.24	2.62
AUC _{0->last}	min*mcg/mL	5	215.88	68.00	4	193.81	117.43
AUC _{0->inf}	min*mcg/mL	5	216.19	68.15	4	194.21	117.20
Percentage extrapolated AUC _{0->inf}	%	5	0.14	0.07	4	0.32	0.30
dFdU							
C_{\max}	mcg/mL	5	67.92	12.13	4	73.75	6.48
AUC _{0->last}	min*mcg/mL	5	$2.18\!\times\!10^4$	1.60×10^{3}	4	$2.06\!\times\!10^4$	2.92×10^{3}
AUC _{0->inf}	min*mcg/mL	5	2.47×10^{4}	4.93×10^{3}	4	2.18×10^4	1.60×10^{3}
Percentage extrapolated $AUC_{0->inf}$	%	5	18.13	7.49	4	15.73	6.17

and radiographic response (Table S4). Staining for AKA and pHH3 was generally low (AKA average 5.9% positive tumor nuclei, range 0–22.4%; pHH3 average 7.8% positive tumor nuclei, range 0–98%), although one subject was remarkable for pHH3 staining of 98.0% of evaluated tumor cells with best response of stable disease. Disease control and partial responses were observed regardless of AKA and pHH3 expression. PFS did not appear to correlate to AKA or pHH3 expression.

Pharmacokinetics

The PK parameters including C_{max} , AUC_{0->last}, AUC_{0->inf}, and the percentage of extrapolated AUC_{0->inf} of both gemcitabine and dFdU (Table 3) on day 1 (without alisertib) and day 8 (with alisertib) were evaluated by NCA with WinNonlin. One subject missed the PK day 8 timepoint and therefore was excluded from the gemcitabine NCA. The NCA PK parameters of alisertib were also evaluated (Table S5). Another subject had C_{max} at the last timepoint and therefore was excluded from the alisertib NCA since both the actual C_{max} and AUC_{0->inf} were unable to be estimated. All NCA PK parameters for gemcitabine and dFdU were not significantly affected by co-administered alisertib among individual subjects (paired t test, P > 0.05). The gemcitabine and dFdU concentration-time profiles on day 1 (no alisertib) and day 8 (with alisertib) within each subject were also visually overlapping (Fig. 3), suggesting the low possibility of DDI effect from alisertib on gemcitabine. Alisertib PK profiles on day 8 appeared similar between subjects (Fig. S1). Subject 22 (Fig. S1) had an unexpectedly high C_{max} (1750 ng/ mL), and Subject 24 (Fig. S1) had an unexpectedly long T_{max} (24 h) when compared to other subjects here (1.5-4.5 h) and in published literature (3-4 h) [30]. No offending concomitant medications were noted to affect the PK parameters of these two subjects, and no aberrancies were noted in drug administration dosage or timing.

Discussion

An alternative approach to chemotherapeutic, specifically taxane-induced mitotic inhibition, is to specifically target proteins critical to mitosis and only expressed in cells at the time of mitosis. Thus, there has been increasing interest in developing drugs to target proteins such as AKA that meet this description. The addition of gemcitabine may enhance alisertib activity by capitalizing on DNA instability. We therefore sought to evaluate the safety and preliminary efficacy of alisertib in combination with gemcitabine in this phase I dose escalation and expansion trial.

Alisertib with gemcitabine proved to be a feasible drug combination, and a majority of subjects were able to be treated at the highest dose level (DL4) and RP2D of gemcitabine (1000 mg/m² days 1, 8, 15) and alisertib 50 mg twice daily (days 1–3, 8–10, 15–17) in 28-day cycles. However, toxicity including hematologic abnormalities, fatigue, transaminitis, and mucositis resulted in frequent dose interruptions and reductions. Although subjects enrolled in this study were heavily pre-treated, disease control was noted in 16 evaluable subjects including two subjects with partial response. Although no clear relationship between efficacy and AKA or pHH3 expression was observed in these data, exploratory biomarkers in small cell lung cancer have demonstrated a potential benefit signal in certain genomic subsets, namely alterations in cell cycle regulation genes [17].

The metabolism of alisertib and its potential of perpetrating drug–drug interactions (DDIs) to other co-administered drugs has been extensively studied [27]. Briefly, the two main metabolites of alisertib are o-desmethyl alisertib and alisertib acyl glucuronide. In vitro phenotyping has demonstrated that CYP3A is largely involved in oxidative metabolism of alisertib. The acyl glucuronidation of alisertib is mainly due to uridine 5'-diphospho-glucuronosyltransferase Fig. 3 Gemcitabine (left panels)

and dFdU (right panels) concen-

tration-time profiles for subjects

(no alisertib, open markers) and

on day 8 (with alisertib, closed

markers in black)

(22-26) evaluable for gemcit-

abine and dFdU PK on day 1





(UGT) isoenzymes. In contrast, gemcitabine is primarily metabolized to dFdU in the liver and plasma by cytidine deaminase (CDA) for further clearance [28]. The CDA and transporter gene polymorphisms are responsible for the variance of metabolism and response [29]. In this study, we observed that gemcitabine PK parameters were not significantly affected by alisertib co-administration. In vitro, alisertib and its 2 main metabolites have not shown appreciable inhibition to selected CYPs [27], thus implying a low likelihood of perpetrating DDIs with co-administrated drugs through CYP and UGT inhibition/activation. Alisertib was orally administrated within 10 min after the starting of gemcitabine infusion. Although alisertib PK parameters were not compared between the presence and absence of gemcitabine intrasubject, similar $C_{\rm max}$ (700–900 ng/mL) and AUCs for alisertib were observed here as has been previously reported [30]. This is the first clinical DDI study for alisertib and gemcitabine. It is not yet known whether gemcitabine exerts DDI to alisertib. Further evaluation is required to confirm alisertib's effects on CDA-mediated gemcitabine metabolism in vitro. Our current finding suggests low possibility of DDI between the two drugs.

The RP2D of alisertib monotherapy has previously been reported as 50 mg twice daily for 7 days in a 21-day cycle [12, 24, 25]. In combination with paclitaxel, RP2D has been 40 mg twice daily at a schedule similar to the one reported here [31]. Tolerability issues have been raised with alisertib in either monotherapy or combination therapy. In a monotherapy trial of solid tumors, treatment-related adverse events included neutropenia (43%), leukopenia (21%), and anemia (10%), and a high rate of SAEs (43%) [32]. In combination with paclitaxel, tolerability concerns have included high rates of neutropenia, grade \geq 3 events, SAEs, and dose reductions or discontinuations [17, 31], although health-related quality of life appeared similar to paclitaxel alone [31]. Similarly, diarrhea, dehydration, and hematologic toxicities have been dose-limiting for alisertib plus irinotecan [33].

Alisertib monotherapy among multiple solid organ malignancies has demonstrated limited efficacy with response rates of approximately 5-20% depending on the primary tumor subtype [32]. Combination alisertib and paclitaxel has been evaluated in randomized phase 2 trials of subjects with small cell lung cancer and breast/ ovarian cancer. In small cell lung cancer, alisertib and paclitaxel has been compared to placebo and paclitaxel with 89 subjects in each arm. Although the primary endpoint of PFS was not met, the alisertib and paclitaxel arm trended toward improved PFS (mPFS 3.32 vs 2.17 months; HR 0.77, 95% CI 0.557–1.067, p = 0.113), and response rates were comparable (22 vs 18%) [17]. Similar findings have been noted in breast/ovarian subjects (mPFS 6.7 vs 4.7 months; HR 0.75, 80% CI 0.58–0.96, p = 0.14) with similar response rates in the two arms (60% vs 52%, p = 0.38 [31]. In acute myelogenous leukemia (AML), induction 7 + 3 chemotherapy plus alisertib 30 mg twice daily on days 8-15 has shown preliminary efficacy with complete remission rates of 64%. This is notable especially since subjects were required to have high-risk features [34].

Alisertib plus gemcitabine has a rational basis for combination in advanced solid organ malignancies. Alisertib coadministration did not affect gemcitabine PK characteristics, suggesting against any DDIs of alisertib on gemcitabine. Preliminary efficacy in terms of disease control among a heavily pre-treated group of subjects was observed, and toxicity, predominantly gastrointestinal and hematologic, was manageable. Further evaluation of this combinatorial strategy would benefit from the identification of a predictive biomarker(s). Supplementary Information The online version contains supplementary material available at https://doi.org/10.1007/s00280-022-04457-9.

Author contributions All authors were involved in writing of manuscript and performing research. In addition: JAC analyzed data and contributed analytical tools. JCH analyzed data. CW analyzed data and contributed analytical tools. AY analyzed data. KM analyzed data and contributed analytical tools. TJS designed research. PM analyzed data and contributed analytical tools. NM analyzed data. KK designed research. EJK designed research, analyzed data, and contributed analytical tools.

Funding Takeda Oncology.

Data availability The datasets generated during and/or analyzed during the current study are available from the corresponding author on reasonable request.

Declarations

Conflict of interests JAC received an institutional research grant from the AACR-AstraZeneca clinical immune-oncology research fellowship and is currently employed by BeiGene. Work was completed during fellowship employment at UC Davis. DRG receives institutional research Grants from Amgen, AstraZeneca, Genentech, and Merck, and is a consultant or on advisory boards for AstraZeneca (institutional), Roche-Genentech (institutional), Guardant Health (institutional), Inivata, IO Biotech (institutional), Lilly, Merck, Novartis, and Oncocyte (institutional). TL receives institutional research grants from Merck, Hengrui, LabyRx, OncoImmune (OncoC4), Tempus, AstraZeneca, and RasCal Therapeutics. JWR receives institutional research grants from Merck, Novartis, Spectrum, Revolution Medicine, AstraZeneca, and GlaxoSmithKline, is a consultant or on advisory boards for Blueprint, Beigene, Boehringer Ingelheim, Daiichi Sankyo, EMD Serano, Novartis, Turning Point, and Janssen. EJK receives institutional research grants from Celgene, OncoMed, Halozyme, Merck, Boston Biomedical, EpicentRx, Samumed, Bristol Myers Squibb, Astellas, Erytech, Fibrogen, NGM Biopharmaceuticals, Eureka Therapeutics, and Dynavax, and is a consultant or on advisory boards for Lilly, Eisai, and Taiho Oncology. JCH, CW, AY, KM, TJS, KT, PCM, AM, NM, and KK have no conflicts of interest to disclose.

Ethics approval All procedures performed in studies involving human participants were in accordance with the ethical standards of the institutional and/or national research committee and with the 1964 Helsinki Declaration and its later amendments or comparable ethical standards. This study was approved by the institutional review board of the University of California, Davis.

Open Access This article is licensed under a Creative Commons Attribution 4.0 International License, which permits use, sharing, adaptation, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons licence, and indicate if changes were made. The images or other third party material in this article are included in the article's Creative Commons licence, unless indicated otherwise in a credit line to the material. If material is not included in the article's Creative Commons licence and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a copy of this licence, visit http://creativecommons.org/licenses/by/4.0/.

References

- Barr AR, Gergely F (2007) Aurora-A: the maker and breaker of spindle poles. J Cell Sci 120(Pt 17):2987–2996. https://doi.org/ 10.1242/jcs.013136
- Cowley DO, Rivera-Perez JA, Schliekelman M, He YJ, Oliver TG, Lu L, O'Quinn R, Salmon ED, Magnuson T, Van Dyke T (2009) Aurora-A kinase is essential for bipolar spindle formation and early development. Mol Cell Biol 29(4):1059–1071. https://doi.org/10.1128/MCB.01062-08
- Zhou H, Kuang J, Zhong L, Kuo WL, Gray JW, Sahin A, Brinkley BR, Sen S (1998) Tumour amplified kinase STK15/BTAK induces centrosome amplification, aneuploidy and transformation. Nat Genet 20(2):189–193. https://doi.org/10.1038/2496
- Zhu J, Abbruzzese JL, Izzo J, Hittelman WN, Li D (2005) AURKA amplification, chromosome instability, and centrosome abnormality in human pancreatic carcinoma cells. Cancer Genet Cytogenet 159(1):10–17. https://doi.org/10.1016/j.cancergenc yto.2004.09.008
- Nikonova AS, Astsaturov I, Serebriiskii IG, Dunbrack RL Jr, Golemis EA (2013) Aurora A kinase (AURKA) in normal and pathological cell division. Cell Mol Life Sci 70(4):661–687. https://doi.org/10.1007/s00018-012-1073-7
- Li D, Zhu J, Firozi PF, Abbruzzese JL, Evans DB, Cleary K, Friess H, Sen S (2003) Overexpression of oncogenic STK15/ BTAK/Aurora A kinase in human pancreatic cancer. Clin Cancer Res 9(3):991–997
- Bavetsias V, Linardopoulos S (2015) Aurora kinase inhibitors: current status and outlook. Front Oncol 5:278. https://doi.org/ 10.3389/fonc.2015.00278
- Shah KN, Bhatt R, Rotow J, Rohrberg J, Olivas V, Wang VE, Hemmati G, Martins MM, Maynard A, Kuhn J, Galeas J, Donnella HJ, Kaushik S, Ku A, Dumont S, Krings G, Haringsma HJ, Robillard L, Simmons AD, Harding TC, McCormick F, Goga A, Blakely CM, Bivona TG, Bandyopadhyay S (2019) Aurora kinase A drives the evolution of resistance to third-generation EGFR inhibitors in lung cancer. Nat Med 25(1):111–118. https://doi.org/10.1038/s41591-018-0264-7
- Hata T, Furukawa T, Sunamura M, Egawa S, Motoi F, Ohmura N, Marumoto T, Saya H, Horii A (2005) RNA interference targeting aurora kinase a suppresses tumor growth and enhances the taxane chemosensitivity in human pancreatic cancer cells. Can Res 65(7):2899–2905. https://doi.org/10.1158/0008-5472. CAN-04-3981
- Gorgun G, Calabrese E, Hideshima T, Ecsedy J, Perrone G, Mani M, Ikeda H, Bianchi G, Hu Y, Cirstea D, Santo L, Tai YT, Nahar S, Zheng M, Bandi M, Carrasco RD, Raje N, Munshi N, Richardson P, Anderson KC (2010) A novel Aurora-A kinase inhibitor MLN8237 induces cytotoxicity and cell-cycle arrest in multiple myeloma. Blood 115(25):5202–5213. https://doi.org/ 10.1182/blood-2009-12-259523
- Green MR, Woolery JE, Mahadevan D (2011) Update on aurora kinase targeted therapeutics in oncology. Expert Opin Drug Discov 6(3):291–307. https://doi.org/10.1517/17460441.2011. 555395
- Cervantes A, Elez E, Roda D, Ecsedy J, Macarulla T, Venkatakrishnan K, Rosello S, Andreu J, Jung J, Sanchis-Garcia JM, Piera A, Blasco I, Manos L, Perez-Fidalgo JA, Fingert H, Baselga J, Tabernero J (2012) Phase I pharmacokinetic/pharmacodynamic study of MLN8237, an investigational, oral, selective aurora a kinase inhibitor, in patients with advanced solid tumors. Clin Cancer Res 18(17):4764–4774. https://doi.org/10.1158/1078-0432. CCR-12-0571
- Dees EC, Cohen RB, von Mehren M, Stinchcombe TE, Liu H, Venkatakrishnan K, Manfredi M, Fingert H, Burris HA 3rd,

Infante JR (2012) Phase I study of aurora A kinase inhibitor MLN8237 in advanced solid tumors: safety, pharmacokinetics, pharmacodynamics, and bioavailability of two oral formulations. Clin Cancer Res 18(17):4775–4784. https://doi.org/10.1158/1078-0432.CCR-12-0589

- Sehdev V, Katsha A, Ecsedy J, Zaika A, Belkhiri A, El-Rifai W (2013) The combination of alisertib, an investigational Aurora kinase A inhibitor, and docetaxel promotes cell death and reduces tumor growth in preclinical cell models of upper gastrointestinal adenocarcinomas. Cancer 119(4):904–914. https://doi.org/10. 1002/cncr.27801
- Katayama H, Sen S (2010) Aurora kinase inhibitors as anticancer molecules. Biochim Biophys Acta 1799(10–12):829–839. https:// doi.org/10.1016/j.bbagrm.2010.09.004
- 16. Zhou N, Singh K, Mir MC, Parker Y, Lindner D, Dreicer R, Ecsedy JA, Zhang Z, Teh BT, Almasan A, Hansel DE (2013) The investigational Aurora kinase A inhibitor MLN8237 induces defects in cell viability and cell-cycle progression in malignant bladder cancer cells in vitro and in vivo. Clin Cancer Res 19(7):1717–1728. https://doi.org/10.1158/1078-0432. CCR-12-2383
- 17. Owonikoko TK, Niu H, Nackaerts K, Csoszi T, Ostoros G, Mark Z, Baik C, Joy AA, Chouaid C, Jaime JC, Kolek V, Majem M, Roubec J, Santos ES, Chiang AC, Speranza G, Belani CP, Chiappori A, Patel MR, Czebe K, Byers L, Bahamon B, Li C, Sheldon-Waniga E, Kong EF, Williams M, Badola S, Shin H, Bedford L, Ecsedy JA, Bryant M, Jones S, Simmons J, Leonard EJ, Ullmann CD, Spigel DR, investigators Cs, (2020) Randomized phase II study of paclitaxel plus alisertib versus paclitaxel plus placebo as second-line therapy for SCLC: primary and correlative biomarker analyses. J Thorac Oncol 15(2):274–287. https://doi. org/10.1016/j.jtho.2019.10.013
- Mou PK, Yang EJ, Shi C, Ren G, Tao S, Shim JS (2021) Aurora kinase A, a synthetic lethal target for precision cancer medicine. Exp Mol Med 53(5):835–847. https://doi.org/10.1038/ s12276-021-00635-6
- VanderPorten EC, Taverna P, Hogan JN, Ballinger MD, Flanagan WM, Fucini RV (2009) The Aurora kinase inhibitor SNS-314 shows broad therapeutic potential with chemotherapeutics and synergy with microtubule-targeted agents in a colon carcinoma model. Mol Cancer Ther 8(4):930–939. https://doi.org/10.1158/ 1535-7163.Mct-08-0754
- Oh ET, Byun MS, Lee H, Park MT, Jue DM, Lee CW, Lim BU, Park HJ (2010) Aurora-a contributes to radioresistance by increasing NF-kappaB DNA binding. Radiat Res 174(3):265–273. https://doi.org/10.1667/RR2017.1
- Sun C, Chan F, Briassouli P, Linardopoulos S (2007) Aurora kinase inhibition downregulates NF-kappaB and sensitises tumour cells to chemotherapeutic agents. Biochem Biophys Res Commun 352(1):220–225. https://doi.org/10.1016/j.bbrc.2006.11.004
- 22. Ling JH, Kang YA, Zhao RY, Xia QH, Lee DF, Chang Z, Li J, Peng BL, Fleming JB, Wang HM, Liu JS, Lemischka IR, Hung MC, Chiao PJ (2012) Kras(G12D)-induced IKK2/beta/NF-kappa B activation by IL-1 alpha and p62 feedforward loops is required for development of pancreatic ductal adenocarcinoma. Cancer Cell 21(1):105–120. https://doi.org/10.1016/j.ccr.2011.12.006
- Wang WX, Abbruzzese JL, Evans DB, Larry L, Cleary KR, Chiao PJ (1999) The nuclear factor-kappa B RelA transcription factor is constitutively activated in human pancreatic adenocarcinoma cells. Clin Cancer Res 5(1):119–127
- Falchook G, Kurzrock R, Gouw L, Hong D, McGregor KA, Zhou X, Shi H, Fingert H, Sharma S (2014) Investigational Aurora A kinase inhibitor alisertib (MLN8237) as an enteric-coated tablet formulation in non-hematologic malignancies: phase 1 dose-escalation study. Invest New Drugs 32(6):1181–1187. https://doi.org/10.1007/s10637-014-0121-6

- Venkatakrishnan K, Zhou X, Ecsedy J, Mould DR, Liu H, Danaee H, Fingert H, Kleinfield R, Milton A (2015) Dose selection for the investigational anticancer agent alisertib (MLN8237): pharmacokinetics, pharmacodynamics, and exposure-safety relationships. J Clin Pharmacol 55(3):336–347. https://doi.org/10.1002/jcph.410
- Harvey JM, Clark GM, Osborne CK, Allred DC (1999) Estrogen receptor status by immunohistochemistry is superior to the ligand-binding assay for predicting response to adjuvant endocrine therapy in breast cancer. J Clin Oncol 17(5):1474–1481. https:// doi.org/10.1200/JCO.1999.17.5.1474
- 27. Pusalkar S, Zhou X, Li Y, Cohen L, Yang JJ, Balani SK, Xia C, Shyu WC, Lu C, Venkatakrishnan K, Chowdhury SK (2020) Biotransformation pathways and metabolite profiles of Oral [(14) C] Alisertib (MLN8237), an investigational aurora a kinase inhibitor, in patients with advanced solid tumors. Drug Metab Dispos 48(3):217–229. https://doi.org/10.1124/dmd.119.087338
- Ciccolini J, Serdjebi C, Peters GJ, Giovannetti E (2016) Pharmacokinetics and pharmacogenetics of Gemcitabine as a mainstay in adult and pediatric oncology: an EORTC-PAMM perspective. Cancer Chemother Pharmacol 78(1):1–12. https://doi.org/10. 1007/s00280-016-3003-0
- Tanaka M, Javle M, Dong X, Eng C, Abbruzzese JL, Li D (2010) Gemcitabine metabolic and transporter gene polymorphisms are associated with drug toxicity and efficacy in patients with locally advanced pancreatic cancer. Cancer 116(22):5325–5335. https:// doi.org/10.1002/cncr.25282
- 30. Zhou X, Pant S, Nemunaitis J, Craig Lockhart A, Falchook G, Bauer TM, Patel M, Sarantopoulos J, Bargfrede M, Muehler A, Rangachari L, Zhang B, Venkatakrishnan K (2018) Effects of rifampin, itraconazole and esomeprazole on the pharmacokinetics of alisertib, an investigational aurora a kinase inhibitor in patients with advanced malignancies. Invest New Drugs 36(2):248–258. https://doi.org/10.1007/s10637-017-0499-z
- Falchook G, Coleman RL, Roszak A, Behbakht K, Matulonis U, Ray-Coquard I, Sawrycki P, Duska LR, Tew W, Ghamande S, Lesoin A, Schwartz PE, Buscema J, Fabbro M, Lortholary A, Goff B, Kurzrock R, Martin LP, Gray HJ, Fu S, Sheldon-Waniga E, Lin HM, Venkatakrishnan K, Zhou X, Leonard EJ, Schilder RJ (2019)

Alisertib in combination with weekly paclitaxel in patients with advanced breast cancer or recurrent ovarian cancer: a randomized clinical trial. JAMA Oncol 5(1):e183773. https://doi.org/10.1001/jamaoncol.2018.3773

- 32. Melichar B, Adenis A, Lockhart AC, Bennouna J, Dees EC, Kayaleh O, Obermannova R, DeMichele A, Zatloukal P, Zhang B, Ullmann CD, Schusterbauer C (2015) Safety and activity of alisertib, an investigational aurora kinase A inhibitor, in patients with breast cancer, small-cell lung cancer, non-small-cell lung cancer, head and neck squamous-cell carcinoma, and gastrooesophageal adenocarcinoma: a five-arm phase 2 study. Lancet Oncol 16(4):395–405. https://doi.org/10.1016/S1470-2045(15) 70051-3
- 33. Semrad TJ, Kim EJ, Gong IY, Li T, Christensen S, Arora M, Riess JW, Gandara DR, Kelly K (2021) Phase 1 study of alisertib (MLN8237) and weekly irinotecan in adults with advanced solid tumors. Cancer Chemother Pharmacol 88(2):335–341. https://doi. org/10.1007/s00280-021-04293-3
- 34. Brunner AM, Blonquist TM, DeAngelo DJ, McMasters M, Fell G, Hermance NM, Winer ES, Lindsley RC, Hobbs GS, Amrein PC, Hock HR, Steensma DP, Garcia JS, Luskin MR, Stone RM, Ballen KK, Rosenblatt J, Avigan D, Nahas MR, Mendez LM, McAfee SL, Moran JA, Bergeron M, Foster J, Bertoli C, Manning AL, McGregor KL, Fishman KM, Kuo FC, Baltay MT, Macrae M, Burke M, Behnan T, Wey MC, Som TT, Ramos AY, Rae J, Lombardi Story J, Nelson N, Logan E, Connolly C, Neuberg DS, Chen YB, Graubert TA, Fathi AT (2020) Alisertib plus induction chemotherapy in previously untreated patients with high-risk, acute myeloid leukaemia: a single-arm, phase 2 trial. Lancet Haematol 7(2):e122–e133. https://doi.org/10.1016/S2352-3026(19) 30203-0

Publisher's Note Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.