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# Research Paper

# Safety, Tolerability and Pharmacokinetics of Single Doses of Oxytocin Administered via an Inhaled Route in Healthy Females: Randomized, Single-blind, Phase 1 Study



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#### ABSTRACT

Background: The utility of intramuscular (IM) oxytocin for the prevention of postpartum hemorrhage in resource-poor settings is limited by the requirement for temperature-controlled storage and skilled staff to administer the injection. We evaluated the safety, tolerability and pharmacokinetics (PK) of a heat-stable, inhaled (IH) oxytocin formulation.

Methods: This phase 1, randomized, single-center, single-blind, dose-escalation, fixed-sequence study (NCT02542813) was conducted in healthy, premenopausal, non-pregnant, non-lactating women aged 18–45 years. Subjects initially received IM oxytocin 10 international units (IU) on day 1, IH placebo on day 2, and IH oxytocin 50  $\mu$ g on day 3. Subjects were then randomized 4:1 using validated GSK internal software to IH placebo or ascending doses of IH oxytocin (200, 400, 600  $\mu$ g). PK was assessed by comparing systemic exposure (maximum observed plasma concentration, area under the concentration-time curve, and plasma concentrations at 10 and 30 min post dose) for IH versus IM oxytocin. Adverse events (AEs), spirometry, laboratory tests, vital signs, electrocardiograms, physical examinations, and cardiac telemetry were assessed.

Findings: Subjects were recruited between September 14, 2015 and October 12, 2015. Of the 16 subjects randomized following initial dosing, 15 (IH placebo n=3; IH oxytocin n=12) completed the study. IH (all doses) and IM oxytocin PK profiles were comparable in shape. However, systemic exposure with IH oxytocin 400  $\mu$ g most closely matched IM oxytocin 10 IU. Systemic exposure was approximately dose proportional for IH oxytocin. No serious AEs were reported. No clinically significant findings were observed for any safety parameters. Interpretation: These data suggest that similar oxytocin systemic exposure can be achieved with IM and IH admiration.

Interpretation: These data suggest that similar oxytocin systemic exposure can be achieved with IM and IH administration routes, and no safety concerns were identified with either route. The inhalation route may offer the opportunity to increase access to oxytocin for women giving birth in resource-poor settings.

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# 1. Introduction

Oxytocin is a neuropeptide involved in the onset and progression of labor (Arrowsmith and Wray, 2014; Blanks and Thornton, 2003).

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Oxytocin acts as a potent endogenous uterotonic (Arrowsmith and Wray, 2014), causing an increase in uterine contraction intensity and frequency, which facilitates labor and delivery (Blanks and Thornton, 2003). Following delivery, the mother is, however, at risk of postpartum hemorrhage (PPH), defined as blood loss of ≥500 mL within 24 h after birth (World Health Organization, 2012), most commonly due to uterine atony (Weeks, 2015). In the 25 years between 1990 and 2015, an estimated 10.7 million women worldwide died in pregnancy and childbirth (World Health Organization, 2015a). Of the estimated 303,000 women who died during and following pregnancy and childbirth in 2015, 99% were from low-income countries, with

approximately two-thirds of deaths occurring in sub-Saharan Africa and one-third in Asia (World Health Organization, 2015a). PPH is estimated as the single largest contributor to maternal mortality, responsible for 19.7% of maternal deaths worldwide (Say et al., 2014). Many of these deaths could be prevented by effective access to treatments to reduce PPH (World Health Organization, 2012). Active management of the third stage of labor, including prophylactic administration of injectable oxytocin immediately after delivery, has been shown to significantly reduce primary blood loss ≥ 500 mL compared with expectant management (Begley et al., 2015).

Intramuscular (IM) oxytocin (10 international units [IU]) was identified as one of 13 life-saving commodities for women and children in a 2012 United Nations (UN) Commission Report (United Nations, 2012), and was included in the World Health Organization (WHO) model list of essential medicines (World Health Organization, 2015b). Oxytocin is supplied as an aqueous solution in an ampoule and requires coldchain storage, sterile needles with sharps disposal, and trained healthcare professionals for administration. This significantly limits access in resource-poor settings, where many women cannot attend medical facilities, their birth attendants are not allowed to administer injections, or the oxytocin cannot be refrigerated, leading to use of material of degraded quality. A review of studies assessing the quality of oxytocin ampoules for injection in low- and middle-income countries found that 58% and 22% of ampoules collected in Africa and Asia, respectively, contained less than the specified content of oxytocin, according to pharmacopoeial limits (Torloni et al., 2016).

Inhaled (IH) delivery of heat-stable oxytocin could offer a practical means to deliver this medicine without the need for cold-chain storage and provide access in those resource-poor settings of greatest unmet need. The Innovation Countdown 2030 Initiative has estimated that the introduction of non-injectable, heat-stable formulations of oxytocin could prevent 146,000 maternal deaths over a period of 8 years (Innovation Countdown 2030, 2015).

We have developed a heat-stable dry powder formulation of oxytocin for inhalation and sought to evaluate the safety, tolerability and pharmacokinetics (PK) in this study. Subjects also received IM oxytocin 10 IU for PK comparison.

# 2. Materials and Methods

# 2.1. Study Design

This phase 1, randomized, single-center, single-blind, ascending dose-escalation, fixed-sequence study with IH oxytocin (NCT02542813) was conducted at the GSK Clinical Unit Cambridge, Addenbrooke's Centre for Clinical Investigation (Addenbrooke's Hospital, UK). Ethical approval for the study was obtained from the Office for Research Ethics Committees Northern Ireland. The study was conducted in accordance with International Conference on Harmonisation Good Clinical Practice, all applicable subject privacy requirements, and the ethical principles outlined in the Declaration of Helsinki 2013. The full protocol can be accessed at https://gsk-clinicalstudyregister.com/search/ps/1/?study\_ids=201558.

# 2.2. Subjects

Subjects eligible for inclusion in the study were healthy, premeno-pausal, non-pregnant, non-lactating women aged 18–45 years inclusive, with a body mass index of 18–30 kg/m², who were taking an estrogen-containing oral contraceptive pill both prior to and for the duration of the study. Key exclusion criteria included a history of chronic lung conditions, respiratory tract infection within 4 weeks of screening, and history of smoking within 1 year of screening. A full list of exclusion criteria are included in Supplementary Appendix A. Potential subjects were identified from the volunteer panel of the GSK Unit and by advertising.

Written informed consent was obtained prior to the performance of any study-specific procedures.

#### 2.3. Randomization and Masking

Suitable subjects were enrolled for the study by the principal investigator or designee based on the inclusion and exclusion criteria. Subjects were randomized 1:4 to receive either IH placebo or ascending doses of IH oxytocin (50, 200, 400, 600 µg). Subjects were assigned to study treatment in accordance with the randomization schedule generated by GSK prior to the start of the study using validated internal software. Dosing with IH placebo and IH oxytocin was single blind (blinded to subjects only) for all subjects throughout the study. The excipients in the placebo formulation had not, to our knowledge, previously been included in an inhaled product in this combination. Although the respiratory risk was deemed to be low, the investigator remained unblinded to allow differentiation of adverse events related to inhaled placebo from inhaled oxytocin in order to monitor the tolerability of the study medication and the safety of the subjects. Blinding for subjects was maintained by the use of a matched inhaler device and capsule presentation for both IH placebo and IH oxytocin. The timings of all procedures after IH placebo and IH oxytocin were identical.

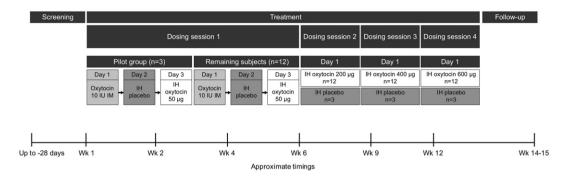
#### 2.4. Procedures

# 2.4.1. Interventions

In dosing session one, all subjects received oxytocin 10 IU (17 µg) (EVER Neuro Pharma GmbH, Unterach, Austria) by IM injection in the thigh on day 1. The excipients in the dry powder IH formulation capsules have been used previously in other IH products, but not in the combination used in this study. Therefore, the safety and tolerability of the excipients alone (IH placebo) were tested in all subjects on day 2. IH oxytocin was then administered to all subjects at a dose of 50 µg on day 3. All IH treatment capsules were administered by oral inhalation using a Modified Air Inlet ROTAHALER™ Dry Powder Inhaler. Study investigators were trained to use the inhaler, and trained subjects prior to use. The first dosing session employed a sentinel cohort approach, in which a pilot group of three subjects was evaluated first, before the remaining 12 subjects were dosed in a staggered fashion, with no more than four subjects dosed per day.

The doses of IH oxytocin selected for investigation were expected to encompass the likely IH therapeutic dose (ie, with a similar systemic exposure to IM oxytocin 10 IU). Dose selection considerations encompassed the predicted amount of drug delivered to the lungs (% nominal) for the current dry powder formulation delivered via the ROTAHALER™ inhaler device, and assumptions around the likely range of relative bioavailability values of IH oxytocin compared with recently reported exposure values following IM administration of oxytocin 10 IU to healthy subjects. Lung bioavailability was assumed to be between 10% and 150% of IM availability, with the upper bound of the range reflecting lung bioavailability reported for other low molecular weight proteins, such as leuprolide (Adjei and Garren, 1990) and insulin (Patton et al., 2004).

In dosing sessions two, three, and four, subjects randomized to placebo received IH placebo (excipients only), while subjects randomized to IH oxytocin received escalating doses of IH oxytocin: 200  $\mu g$ , 400  $\mu g$ , and 600  $\mu g$  (administered as 200  $\mu g$  and 400  $\mu g$  capsules using one ROTAHALER inhaler device per capsule). The decision to proceed to the next increased dose level was made by the dose-escalation committee based on the available safety and tolerability data from not less than nine subjects receiving oxytocin at the prior dose level, as well as any available PK data from the previous dose levels. A follow-up visit was conducted 7–21 days after the last IH oxytocin or IH placebo administration. An overview of the study design is shown in Fig. 1.



**Fig. 1.** Study design. IH = inhaled; IM = intramuscular; IU = international units.

## 2.4.2. Assessments and Analyses

2.4.2.1. Safety Assessments. Adverse events (AEs) and serious AEs (SAEs) were monitored throughout the study. Spirometry, clinical laboratory tests, vital signs, electrocardiograms (ECGs) (PR, QRS, QT, corrected QT [QTc] intervals), physical examinations, and cardiac telemetry were assessed at specified times after dosing.

2.4.2.2. PK Assessments. Blood samples were drawn pre-dose and 3, 5, 10, 20, 30, and 45 min, 1, 1.5, 2, 2.5, 3, 4, and 8 h after dosing. An additional 24-h post-dose blood sample was drawn in dosing session one for the pilot group and for the first three subjects at each new dose level in dosing sessions two, three, and four. Plasma samples were taken via an indwelling cannula (or by direct venepuncture), collected into tripotassium ethylenediaminetetraacetic acid (K3 EDTA) tubes and placed immediately on ice prior to centrifugation. Supernatant plasma was then prepared and analyzed for oxytocin by inVentiv Health Clinique Inc. (Québec, Canada) using a validated analytical method based on automated solid phase extraction, followed by liquid chromatography/mass spectrometry/mass spectrometry analysis. The range of quantification for oxytocin was 2–500 pg/mL using a 350 μL aliquot of human plasma.

The PK parameters determined for both IM oxytocin and IH oxytocin were maximum observed plasma concentration ( $C_{max}$ ), time to  $C_{max}$  ( $t_{max}$ ), apparent terminal phase heat stable, inhaled (IH) oxytocin formulation area under the concentration-time curve from time zero (pre-dose) to last time of quantifiable concentration ( $AUC_{[0-t]}$ ), area under the concentration-time curve from zero (pre-dose) to 3 h post dose ( $AUC_{[0-3h]}$ ), observed plasma concentration at 10 min (nominal time) post-dose (Cp10), and 30 min (nominal time) post-dose (Cp30).

#### 2.5. Outcomes

The objectives and endpoints of the study were to evaluate the safety and tolerability of IH oxytocin by monitoring AEs and changes over time in hematology, clinical chemistry, urinalysis, vital signs, spirometry, cardiac telemetry, and 12-lead ECG parameters from pre-dose values; evaluate the safety and tolerability of the five excipients in the placebo by monitoring for respiratory AEs by spirometry, including forced expiratory volume in 1 s (FEV<sub>1</sub>), and pulse oximetry; and to establish the PK characteristics of oxytocin by assessing the plasma concentration profile ( $C_{max}$ ,  $t_{max}$ , AUC, and  $t_{1/2}$ ) for IH and IM oxytocin at the specified doses.

IM oxytocin is reported to have a rapid onset (3-5 min) and limited duration (2-3h) of uterine activity (Pitocin prescribing information). To assess whether therapeutic concentrations of oxytocin were achieved rapidly and maintained for a sufficient duration post-inhalation compared with the IM route of administration, Cp10 and Cp30 [representing the median and upper bound of the range of  $t_{max}$  following IM administration in a previous study with IM oxytocin (Wong et al., 2016)] were estimated, representing the onset of uterine activity, along

with systemic exposure over the 0–3-h post-dose period (AUC $_{[0-3h]}$ ), representing the duration of uterine activity.

#### 2.6. Statistical Analysis

The sample size was based on feasibility. The study populations assessed included the all subjects/safety population, which included all subjects who received at least one dose of study medication; and the PK population, which included all subjects who received at least one dose of study medication and had at least one PK sample analyzed.

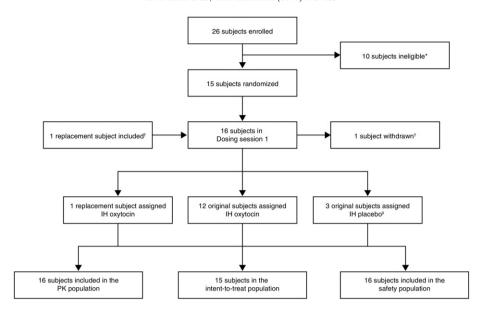
For PK comparisons (IH vs IM), point estimates and two-sided 90% confidence intervals (CI) were computed using a mixed-effect model. Loge-transformed AUC $_{[0-t]}$ , AUC $_{[0-3h]}$ , Cp10, Cp30, and C $_{\rm max}$  were analyzed with a fixed-effect term for treatment and a random-effect term for subject. Point estimates and CIs were exponentially back-transformed to obtain adjusted (least squares) geometric means, point estimates and associated 90% CI for the ratios. Summary statistics were carried out using SAS v9.3 (Cary, CA, USA). Plasma oxytocin concentration-time data were analyzed by non-compartmental methods with WinNonlin v6.3 (Pharsight Corporation, St. Louis, MO, USA). Safety data were summarized descriptively.

## 3. Results

The study was conducted between September 14, 2015 (first subject first visit) and December 16, 2015 (last subject last visit). Subjects were recruited between September 14, 2015 and October 12, 2015. A total of 26 subjects were enrolled, of whom 16 received study treatment and 15 completed the study (trial profile Fig. 2). In the pilot group, two subjects were randomized to receive IH oxytocin and one subject to IH placebo. Among the other subjects, ten subjects were randomized to receive IH oxytocin and two subjects to IH placebo. One subject was withdrawn prior to dosing, at the investigator's discretion, due to inability to cannulate. This subject was replaced and assigned the same randomization sequence. All 16 subjects received IM oxytocin 10 IU and were included in the safety and PK populations.

Baseline demographics and characteristics are shown in Table 1. The mean (range) age of the study population was 28.5 (23–35) years, and all subjects were of White/Caucasian/European heritage.

A total of 13 (81%) subjects experienced at least one AE during the study. AEs occurred in four (25%) subjects who received oxytocin IM 10 IU; five (33%) subjects who received IH oxytocin 50 µg; three (25%) subjects who received IH oxytocin 200 µg; five (42%) subjects who received IH oxytocin 400 µg; six (50%) subjects who received IH oxytocin 600 µg; and five (33%) subjects who received IH placebo. Overall, 45 AEs were reported, of which seven were moderate in intensity, and none were severe. The most common AE was headache, reported in six (38%) subjects. Nine AEs reported in five subjects were determined by the investigator to be drug-related. Of these drug-related AEs, headache and cough were the most common drug-related AEs (three subjects each, 19%). Drug-related AEs occurred in one (6%) subject who received



**Fig. 2.** Trial profile. \*Ineligible subjects: did not meet inclusion/met exclusion criteria (n = 5); subject withdrew consent (n = 2); reserve, not used (n = 2); investigator discretion (n = 1). †One subject was withdrawn at the investigator's discretion due to inability to cannulate. She received IM oxytocin only. This subject was replaced and assigned the same randomization sequence, thus, a total of 16 subjects were randomized; ‡subjects had received IM oxytocin and IH oxytocin  $(50 \, \mu g)$  in the initial dosing session. IH = inhaled; PK = pharmacokinetics.

oxytocin IM 10 IU (headache); three (25%) subjects who received IH oxytocin 400 μg (cough and headache in two subjects [17%] each); three (25%) subjects who received IH oxytocin 600 μg (cough, headache and flushing in one [6%] subject each); and one (6%) subject who received IH placebo (cough). No trends or clinically important differences in AEs were noted between groups or with escalating doses. There were no discontinuations or withdrawals from the study due to AEs, and no deaths or SAEs were reported (Table 2). No trends or clinically significant abnormal spirometry, laboratory, vital signs, ECG, or telemetry findings were reported or noted during review of the data.

The shape of the observed PK concentration-time profile following IH administration was consistent with the IM PK profile (Fig. 3). Plasma oxytocin was detectable in all subjects at the first sampling time (3 min post-dose) and remained quantifiable in 5/16 subjects up to 4 h following IM administration and in at least half of subjects for 1 h with the 50  $\mu g$  IH dose and 2.5 to 4 h with the higher IH doses (200  $\mu g$ , 400  $\mu g$ , and 600  $\mu g$ ). Following each route of administration (IH or IM) there was rapid absorption of oxytocin into plasma ( $t_{\rm max} \sim 10$  min following IM administration and  $t_{\rm max} \sim 8{-}20$  min following IH oxytocin; Table 3). Thereafter, plasma oxytocin concentrations declined rapidly, irrespective of administration route or dose, with the apparent terminal elimination half-life comparable for IH versus IM oxytocin (Table 3). Following IH oxytocin,  $C_{\rm max}$  and AUC(0-t) increased in an approximately linear manner between 200  $\mu g$  and 600  $\mu g$  (Table 4).

PK exposure (as defined by Cp10 and Cp30, AUC<sub>(0-3 h)</sub> and C<sub>max</sub>) was higher with IH oxytocin 400  $\mu g$  and 600  $\mu g$  and much lower with 50  $\mu g$  and 200  $\mu g$  IH compared with IM oxytocin. Oxytocin exposure with IH oxytocin 400  $\mu g$  most closely matched that with IM oxytocin 10 IU, based on the geometric mean ratio for IH versus IM for the PK parameters Cp10, Cp30, and AUC<sub>(0-3 h)</sub> (Table 4).

**Table 1**Baseline demographics and characteristics.

Characteristic	Total population ( $N = 16$ )
Age (years), mean (range)	28.5 (25-35)
Sex, female, n (%)	16 (100)
BMI (kg/m <sup>2</sup> ), mean (range)	22.8 (18.3-28)
Height (cm), mean (SD)	166.5 (5.7)
Weight (kg), mean (SD)	63.3 (8.7)
Race, White/Caucasian/European Heritage, n (%)	16 (100)

BMI = body mass index; SD = standard deviation.

#### 4. Discussion

PPH is a major cause of maternal death with 99% of these deaths occurring in resource-poor settings (World Health Organization, 2015a). Oxytocin is considered the gold standard preventative treatment for PPH, but the requirement for refrigerated storage and needle-based administration for the currently available IM formulations substantially limits its utility in these settings. Other treatments, such as ergometrine or misoprostol, are used but are not considered to be as effective as oxytocin and have contraindications to their use (World Health Organization, 2012). The WHO states that recommendations concerning alternative uterotonics should not detract from the objective of increasing access to oxytocin (World Health Organization, 2012) and in response to this, several initiatives have been launched. The Uniject single-use prefilled syringe, which does not require additional sterile equipment, eliminates the need for dosage estimation and can be used by unskilled personnel (Diop et al., 2016; Tsu et al., 2003; Tsu et al., 2009; Althabe et al., 2011; Strand et al., 2005; Stanton et al., 2013). However, the Uniject devices still require maintenance of cold-chain supply, and a study in rural Ghana highlighted that the single-use prefilled syringe was highly sensitive to small changes in ambient temperature and could expire in as few as 6 days (Mullany et al., 2014). The preferred approach would be the development of a heat-stable, non-parenteral oxytocin formulation as identified by the UN Commission on Life-saving Commodities for Women and Children (United Nations, 2012). A heat-stable oxytocin analog, carbetocin is in development, but it will still require parenteral administration (Boucher et al., 2004). Non-parenteral routes of administration include sublingual, IH, and intranasal administration. A PK study carried out in male volunteers showed that sublingual oxytocin had poor bioavailability, a long lag time and long absorption half-life and was therefore not a reliable formulation for accurate dosing for immediate prevention of PPH (De Groot et al., 1995). We selected the IH route, as this has proven a reliable route for other potent drugs and offers the potential for rapid absorption (Cazzola et al., 2013). Additionally, the formulation of peptides and proteins in a spray-dried matrix for IH delivery is known to confer enhanced thermostability compared with aqueous solutions (Vehring, 2008). This approach has been applied to the formulations used in this study: robust heat stability was demonstrated at 30 °C and at 40 °C during product development (Supplementary Fig. 1) and recent data support a minimum 30 month shelf-life at 25 °C (approved by the UK

**Table 2** Adverse event summary (safety population).

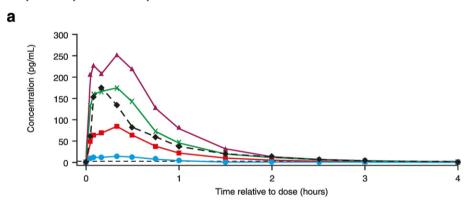
	IM oxytocin 10 IU (17 μg)		IH oxytocin 50 μg		IH oxytocin 200 μg		IH oxytocin 400 μg		IH oxytocin 600 μg		IH placebo		Total	
	Number of subjects $N = 16$	Number of AEs	Number of subjects $N = 15$	Number of AEs	Number of subjects $N = 12$	Number of AEs	Number of subjects $N = 12$	Number of AEs	Number of subjects $N = 12$	Number of AEs	Number of subjects $N = 15$	Number of AEs	Number of subjects $N = 16$	Number of AEs
All AEs														
Any	4 (25%)		5 (33%)		3 (25%)		5 (42%)		6 (50%)		5 (33%)		13 (81%)	45
Drug-related	1 (6%)		0		0		3 (25%)		3 (25%)		1 (7%)		5 (31%)	9
Cough	0		0		0		2 (17%)	2	1 (8%)	1	1 (7%)	1	3 (19%)	3
Headache	1 (6%)	1	0		0		2 (17%)	2	1 (8%)	1	0		3 (19%)	3
Flushing	0		0		0		0		1 (8%)	1	0		1 (6%)	1
Led to discontinuation of study drug or withdrawal from the study	0	0	0	0	0	0	0	0	0	0	0	0	0	0
SAEs														
Non-fatal	0	0	0	0	0	0	0	0	0	0	0	0	0	0
Deaths	0	0	0	0	0	0	0	0	0	0	0	0	0	0

AE = adverse event; IH = inhaled; IM = intramuscular; IU = international units; SAE = serious adverse event.

Medicines & Healthcare products Regulatory Agency for clinical supply). An in vitro preclinical study further demonstrated that dry powder formulations of oxytocin suitable for oral inhalation can be prepared and are stable after extended storage under tropical and subtropical conditions (Fabio et al., 2015). The potential for an IH powder form of oxytocin was also demonstrated in an in vivo preclinical study, which showed that administration of an ultra-fine oxytocin powder to the airways of postpartum sheep led to rapid absorption and comparable uterine

electromyographic activity to that observed with IM administration (Prankerd et al., 2013).

This study reports successful administration of oxytocin by the IH route in humans. The five non-pharmacologically active components contained in the IH oxytocin formulation are approved individually for use in other IH products. However, they had never been used previously in this particular combination. Administration of the excipients alone was not associated with features of irritancy (eg, coughing and



LLQ = 2 pg/mL

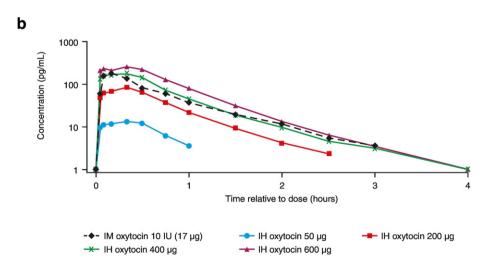


Fig. 3. Median oxytocin plasma concentration-time profile on A) non-logarithmic and B) logarithmic scales (PK population). IH = inhaled; IM = intramuscular; IU = international units; LLQ = lower limit of quantification; PK = pharmacokinetic.

**Table 3** Summary of plasma oxytocin  $t_{max}$  and  $t_{1/2}$  (PK population).

Parameter	Number of subjects (n/N)	Oxytocin dose and formulation	
T <sub>max</sub> (h), median (min, max)	16/16	10 IU IM	0.17 (0.09, 0.34)
	15/15	50 μg IH	0.32 (0.06, 0.34)
	12/12	200 μg IH	0.33 (0.05, 0.50)
	12/12	400 μg IH	0.33 (0.05, 0.34)
	12/12	600 μg IH	0.14 (0.05, 0.34)
T <sub>1/2</sub> (h), geometric mean (95% CI)	13/16	10 IU IM	0.64 (0.55, 0.74)
	8/15	50 μg IH	0.36 (0.30, 0.44)
	12/12	200 μg IH	0.45 (0.38, 0.54)
	9/12	400 μg IH	0.55 (0.43, 0.70)
	12/12	600 µg IH	0.60 (0.49, 0.74)

CI = confidence interval; IH = inhaled; IM = intramuscular; IU = international units; PK = pharmacokinetics; t<sub>1/2</sub> = terminal phase half-life; t<sub>max</sub> = time to maximum observed plasma concentration

bronchospasm) in this study. Indeed, the IH route of delivery was well tolerated in this small-scale study.

Since IM oxytocin reaches its site of action, the uterus, via the plasma it is proposed that demonstrating a comparable plasma drug profile following IH and IM routes of administration allows for extrapolation from the extensive clinical safety and efficacy data for the approved IM oxytocin product for the prevention of PPH to the IH product. Overall, the shape of the plasma PK profile following IH administration was similar to that following IM administration. Both administration routes demonstrated rapid absorption of oxytocin into plasma (median t<sub>max</sub> 8-20 min), followed by rapid elimination, with an apparent  $t_{1/2}$  consistent across administration routes. Oxytocin exposure with IH oxytocin was approximately linear over the 200-600 µg dose range, while dose normalized exposure at 50 µg was lower. Lower exposure (AUC) following 50 µg may in part reflect censoring of the PK profile; since concentrations of oxytocin fell below the lower limit of quantification in at least half the subjects by 1 h post-dose. The dose range of IH oxytocin assessed in this study was predicted to cover the therapeutic range

**Table 4**Statistical analysis of oxytocin PK parameters (PK population).

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Parameter	Comparison (IH oxytocin vs IM	Adjusted geometric	mean	Adjusted geometric mean ratio for IH vs IM
	oxytocin 10 IU)	IH oxytocin	IM oxytocin	oxytocin (90% CI)
$C_{max}$	50 μg IH	15.3	189.96	0.08 (0.06, 0.10)
(pg/mL) <sup>a</sup>	200 μg IH	103.03		0.54 (0.41, 0.71)
	400 μg IH	255.95		1.35 (1.03, 1.77)
	600 μg IH	365.42		1.92 (1.47, 2.52)
$AUC_{(0-3 h)}$	50 μg IH	14.75	127.61	0.12 (0.10, 0.14)
$(pg \cdot h/mL)^a$	200 μg IH	69.78		0.55 (0.46, 0.65)
	400 μg IH	161.67		1.27 (1.07, 1.50)
	600 μg IH	234.91		1.84 (1.56, 2.18)
$AUC_{(0-t)}$	50 μg IH	9.16	119.83	0.08 (0.06, 0.10)
$(pg \cdot h/mL)^a$	200 μg IH	65.02		0.54 (0.42, 0.70)
	400 μg IH	153.83		1.28 (1.00, 1.66)
	600 μg IH	224.34		1.87 (1.45, 2.41)
Cp10	50 μg IH	12.01	171.24	0.07 (0.05, 0.09)
(pg/mL) <sup>a</sup>	200 μg IH	72.71		0.42 (0.33, 0.55)
	400 μg IH	188.13		1.10 (0.85, 1.43)
	600 μg IH	258.31		1.51 (1.16, 1.96)
Cp30	50 μg IH	9.58	82.16	0.12 (0.09, 0.15)
(pg/mL)a	200 μg IH	58.63		0.71 (0.56, 0.91)
	400 μg IH	135.57		1.65 (1.30, 2.09)
	600 μg IH	190.96		2.32 (1.83, 2.95)

%CVb = inter-subject variability;  $AUC_{(0-3\ h)} = area$  under the concentration-time curve from zero (pre-dose) to 3 h;  $AUC_{(0-1)} = area$  under the concentration-time curve from time zero (pre-dose) to last time of quantifiable concentration; CI = confidence interval;  $C_{max} = maximum$  observed plasma concentration; Cp10 = quantified concentration at 10 min (nominal time) post-dose; Cp30 = quantified concentration at 30 min (nominal time) post-dose; Cp30 = quantified concentration at 30 min (nominal time) post-dose; Cp30 = quantified concentration at 30 min (nominal time) post-dose; Cp30 = quantified concentration at 30 min (nominal time) post-dose; Cp30 = quantified concentration at 30 min (nominal time) post-dose; Cp30 = quantified concentration at 30 min (nominal time) post-dose; Cp30 = quantified concentration at 30 min (nominal time) post-dose; Cp30 = quantified concentration at 30 min (nominal time) post-dose; Cp30 = quantified concentration at 30 min (nominal time) post-dose; Cp30 = quantified concentration at 30 min (nominal time) post-dose; Cp30 = quantified concentration at 30 min (nominal time) post-dose; Cp30 = quantified concentration at 30 min (nominal time) post-dose; Cp30 = quantified concentration at 30 min (nominal time) post-dose; Cp30 = quantified concentration at 30 min (nominal time) post-dose; Cp30 = quantified concentration at 30 min (nominal time) post-dose; Cp30 = quantified concentration at 30 min (nominal time) post-dose; Cp30 = quantified concentration at 30 min (nominal time) post-dose; Cp30 = quantified concentration at 30 min (nominal time) post-dose; Cp30 = quantified concentration at 30 min (nominal time) post-dose; Cp30 = quantified concentration at 30 min (nominal time) post-dose; Cp30 = quantified concentration at 30 min (nominal time) post-dose; Cp30 = quantified concentration at 30 min (nominal time) post-dose; Cp30 = quantified concentration at 30 min (nominal time) post-dose; Cp30 = quantified concentration at 3

and the  $400 \,\mu g$  dose of IH oxytocin was found to result in systemic exposure most similar to that of IM oxytocin 10 IU; with the rate and extent of absorption of oxytocin into plasma most consistent with that observed following IM oxytocin 10 IU (as reflected by the geometric mean ratio for IH versus IM for the PK parameters Cp10, Cp30, and  $AUC_{[0-3h]}$ ). However, between subject variability tended to be higher following IH compared with IM dosing.

This study provides important preliminary evidence in humans that a heat-stable, dry powder IH formulation has the potential to deliver a systemic profile of oxytocin comparable with parenteral (IM) administration, although important questions remain. This study was conducted in healthy non-pregnant women. The physiological changes associated with pregnancy, including higher levels of circulating oxytocin, increased renal/hepatic blood flow, increased blood/plasma volume and cardiac output, increased total body water, and decreased plasma proteins have the potential to alter the PK of oxytocin (Sheffield et al., 2014; Prevost et al., 2014). However, unless affecting lung absorption, these factors would be expected to affect IH and IM administration equally. To determine whether there are differences in PK between pregnant and non-pregnant women, investigations are ongoing to study women in third stage of labor. We conducted an enabling study in women in the third stage of labor and observed that their inspiratory profile using the ROTAHALER™ inhaler device was not materially different from non-pregnant volunteers (Wong et al., 2016). However, any effect of altered inspiratory profile during labor on the PK of IH oxytocin will need to be determined in future investigations in women in the third stage of labor. In this study IH oxytocin was not associated with evidence of respiratory tract irritancy, but the information is preliminary; an evaluation of tolerability in smokers and people with asthma would form part of future investigations.

The device for delivery was selected on the basis of its simplicity. The capsule is pushed into the device, the device twisted once to break open the capsule and the powder IH. There has been over 20 years of experience with this type of device, which has optimized and simplified the design to support practical use. Effective delivery in the field will depend on appropriate educational materials and training; in use studies are planned as part of the development program.

This study provides proof-of-concept that an IH oxytocin formulation resistant to thermo-degradation can achieve systemic exposure comparable with IM delivery. If confirmed in women in the third stage of labor with an optimized formulation and delivery device, this approach has the potential to address one of the most important unmet health needs in developing countries.

# **Role of the Funding Source**

GSK, the funder of this study (NCT02542813), was involved in the study design, data collection, data analysis, data interpretation, and writing of the study report. The corresponding author had full access to all the data in the study and had final responsibility for the decision to submit. The McCall MacBain Foundation and Grand Challenges

 $<sup>^</sup>a$  %CVb for IM oxytocin and IH oxytocin, respectively:  $C_{\rm max}$ : 47% and 61-98%; AUC $_{(0-3~\rm h)}$ : 27% and 41-61%; AUC $_{(0-t)}$ : 41% and 43-72%; Cp10: 45% and 64-92%; Cp30: 45% and 42-58%.

Canada (funded by the Government of Canada) provided funding support to the project team at Monash University and had no role in the design, conduct, interpretation, or reporting of this study.

# **Conflicts of Interest**

Disala Fernando, Sarah Siederer, Sunita Singh, Ian Schneider, Ashutosh Gupta, Marcy Powell and Duncan Richards are employees of GSK. Sarah Siederer, Sunita Singh, Ian Schneider, Ashutosh Gupta, Marcy Powell are shareholders of GSK. Susan Fowles was an employee and shareholder of GSK during the conduct of the study and during the initial stages of manuscript development. Peter Lambert has no conflicts of interest to declare. Michelle McIntosh has a patent application for a method and formulation for inhalation (WO2013/016754 A1 [PCT/AU2011001430]) pending.

#### **Author Contributions**

Disala Fernando, Sarah Siederer, Sunita Singh, and Marcy Powell contributed to the study concept and design, data acquisition, and analysis and interpretation. Ian Schneider contributed to study concept and design, and data acquisition. Ashutosh Gupta and Duncan Richards contributed to analysis and interpretation. Michelle McIntosh and Peter Lambert contributed to the study concept and design and interpretation. Susan Fowles contributed to study concept and design, and analysis and interpretation. All authors, apart from Susan Fowles, were involved in the preparation and review of the manuscript and approved the final version to be submitted. Posthumous authorship was granted for Susan Fowles as she was involved in the initial stages of manuscript development prior to her death in September 2016.

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## Appendix A. Supplementary data

Supplementary data to this article can be found online at http://dx.doi.org/10.1016/j.ebiom.2017.07.020.

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