

# Autoinflammatory and autoimmune contributions to complex regional pain syndrome

Molecular Pain  
Volume 14: 1–13  
© The Author(s) 2018  
Article reuse guidelines:  
sagepub.com/journals-permissions  
DOI: 10.1177/1744806918799127  
journals.sagepub.com/home/mpi



J David Clark<sup>1,2</sup>, Vivianne L Tawfik<sup>2</sup>, Maral Tajerian<sup>2</sup>, and Wade S Kingery<sup>3</sup>

## Abstract

Complex regional pain syndrome (CRPS) is a highly enigmatic syndrome typically developing after injury or surgery to a limb. Severe pain and disability are common among those with chronic forms of this condition. Accumulating evidence suggests that CRPS may involve both autoinflammatory and autoimmune components. In this review article, evidence for dysfunction of both the innate and adaptive immune systems in CRPS is presented. Findings from human studies in which cytokines and other inflammatory mediators were measured in the skin of affected limbs are discussed. Additional results from studies of mediator levels in animal models are evaluated in this context. Similarly, the evidence from human, animal, and translational studies of the production of autoantibodies and the potential targets of those antibodies is reviewed. Compelling evidence of autoinflammation in skin and muscle of the affected limb has been collected from CRPS patients and laboratory animals. Cytokines including IL-1 $\beta$ , IL-6, TNF $\alpha$ , and others are reliably identified during the acute phases of the syndrome. More recently, autoimmune contributions have been suggested by the discovery of self-directed pain-promoting IgG and IgM antibodies in CRPS patients and model animals. Both the autoimmune and the autoinflammatory components of CRPS appear to be regulated by neuropeptide-containing peripheral nerve fibers and the sympathetic nervous system. While CRPS displays a complex neuroimmunological pathogenesis, therapeutic interventions could be designed targeting autoinflammation, autoimmunity, or the neural support for these phenomena.

## Keywords

Autoinflammation, autoimmunity, cytokine, nerve growth factor, pain, autonomic, peripheral nervous system

Date Received: 12 July 2018; revised: 6 August 2018; accepted: 7 August 2018

## Introduction

Complex regional pain syndrome (CRPS) usually develops after limb injury followed by immobilization.<sup>1</sup> It presents with a disparate array of nociceptive, vascular, and autonomic changes that exceed the expected clinical course of the inciting injury in both magnitude and duration, frequently resulting in significant functional impairment and disability. Additionally, trophic changes consisting of acutely hypertrophic and later atrophic skin may be seen along with changes in nail texture and hair growth. Osteopenia is sometimes observed on radiographic studies. The population incidence of CRPS is approximately 26 per 100,000 person-years, though the incidence is much higher in specific settings.<sup>2</sup> For example, prospective clinical studies report a 31%

incidence of CRPS after distal tibia fracture and an average 18.8% (1047 patients, range: 1%–37%) incidence of CRPS after distal radius fracture.<sup>3</sup> Additional sources of limb injury linked to CRPS include hand and foot surgery, sports-related trauma, and injuries suffered on the job. Traditionally, CRPS is divided into Type I and

<sup>1</sup>Anesthesiology Service, Veterans Affairs Palo Alto Health Care System, Palo Alto, CA, USA

<sup>2</sup>Department of Anesthesiology, Perioperative & Pain Medicine, Stanford University School of Medicine, Stanford, CA, USA

<sup>3</sup>Palo Alto Veterans Institute for Research, Palo Alto, CA, USA

### Corresponding Author:

J David Clark, Anesthesia Service, Veterans Affairs Palo Alto Health Care System, 3801 Miranda Ave., Palo Alto, CA 94304, USA.

Email: djclark@stanford.edu



Type II with the latter less common variety involving a verifiable nerve lesion.

Patients with CRPS typically progress from an acute stage in which the affected limb is painful, warm, and edematous to a chronic stage in which warmth and edema resolve but the pain remains.<sup>4</sup> Quantitative sensory testing of both CRPS I and II patients has revealed high rates of thermal and mechanical hyperalgesia and slightly lower rates of cold and mechanical allodynia.<sup>5</sup> CRPS symptoms gradually improve over the first year after injury in some patients, but persistent CRPS is a serious problem resulting in severe pain, weakness, contractures, and bone loss.<sup>6</sup> Even with this improvement, however, the majority (~75%) continue to meet the commonly used Budapest criteria for diagnosis.<sup>7</sup> Over 80% of chronic CRPS patients are severely disabled.<sup>8</sup> Treatment strategies are most often symptomatic and focus on rehabilitation; no currently available treatment is disease modifying. Moreover, there is considerable uncertainty as to whether any treatment for CRPS is reliably effective.<sup>9</sup>

The fundamental obstacle to the rational design of therapies to prevent or treat CRPS is our lack of understanding of the syndrome's underlying mechanisms. Indeed, the diverse and temporally evolving range of manifestations of the syndrome makes studies in humans and animal models difficult to execute and interpret. To this point, theories based on the dysfunction of a specific type of tissue, alterations in a single signaling pathway, or change in abundance of a single specific biochemical mediator have failed to explain the complexity of aptly named "Complex" regional pain syndrome. It is notable in this regard that a growing number of enigmatic multisystem syndromes and diseases are now believed to be the result of deranged immune system function. We hypothesize that CRPS may have a similar basis. Furthermore, dysfunctional neuroimmune interactions are likely responsible for initiating and perpetuating CRPS.

## Review

### *Autoinflammatory and autoimmune processes*

Two types of immune system dysfunction are increasingly recognized for their contributions to a broad variety of disease states, autoinflammation and autoimmunity. As the "auto" prefix implies, both these processes involve immune activity against self though through distinct mechanisms. In autoinflammation, the innate immune system is directly responsible for tissue inflammation; in autoimmunity, the innate immune system activates the adaptive immune system against self.<sup>10</sup> Both sets of processes potentially affect the physiology of multiple systems including the neurological,

musculoskeletal, vascular, and integumentary systems, all systems being involved in CRPS. The disease states currently recognized to be supported by autoinflammation and autoimmunity are highly diverse.<sup>10,11</sup> Autoinflammation and autoimmunity were once thought of as mutually exclusive conditions, though it is now appreciated that diseases such as recurrent pericarditis, psoriasis, and ankylosing spondylitis have mixed autoimmunologic etiologies.<sup>12–14</sup> Both autoinflammation and autoimmunity can support inflammation in the absence of infection, a sign of CRPS recognized from the time of Paul Sudeck's descriptions of the syndrome more than 100 years ago.<sup>15</sup> While autoinflammation and autoimmunity are known components of several conditions and diseases involving painful symptoms, the specific roles of these phenomena as pain mechanisms are less well recognized.

### *Autoinflammation*

Autoinflammatory conditions are currently recognized as "clinical disorders marked by abnormally increased inflammation, mediated predominantly by the cells and molecules of the innate immune system, with a significant host predisposition."<sup>16</sup> These conditions are characterized by episodic or continuous inflammation commonly manifest as joint swelling, myalgias, and rashes. Pain is frequently experienced in the setting of autoinflammation and has been reported to occur in skin, joints, muscle, and viscera (Table 1). Both monogenic (Mendelian) and highly complex polygenic conditions may be considered autoinflammatory.<sup>10,14,16,17</sup> Although many of the currently recognized autoinflammatory diseases are rare, several common pain-related conditions involve autoinflammatory processes including osteoarthritis and gout.<sup>18</sup>

The basis for these conditions is dysfunction of the innate immune system, though specific conditions may involve different pathways. The innate immune system is that branch of immunity involving both humoral and cellular response elements nonselective for specific threats or antigens. This type of immunity provides an immediate defense against invading organisms, and the cellular components include dendritic cells such as the skin's Langerhans cells, mast cells, and others. Pattern recognition receptors expressed on these cell types and involved in the innate immune response include Toll-like receptors, NOD and NALP receptors, RIG-I receptors, and several others.<sup>19,20</sup> When these receptors and cells are activated, the involved cells produce cytokines, complement fragments, prostaglandins, bradykinin, and other pro-inflammatory and pronociceptive mediators. The activation of inflammasomes, multiprotein complexes that facilitate the maturation and subsequent secretion of inflammatory cytokines,<sup>21</sup> is common to

**Table 1.** Diseases associated with autoinflammation.

Disease	Gene	Protein	Pain
Familial Mediterranean fever	<i>MEFV</i>	Pyrin	Chest, abdomen, and joints
Cryopyrin-associated autoinflammatory syndrome	<i>NLRP3</i>	Cryopyrin	Joints and muscles
Mevalonate-kinase deficiency	<i>MVK</i>	MVK	Abdomen, joints, and headache
Early-onset sarcoidosis	<i>NOD2</i>	NOD2	Joints and tendons
Familial cold autoinflammatory syndrome type 2	<i>NLRP12</i>	NLRP12	Joints
Pyogenic arthritis, pyoderma gangrenosum, and acne	<i>PSTPIP1</i>	PSTPIP1	Joints
Deficiency of IL-1 receptor antagonist	<i>IL1RN</i>	IL-1Ra	Bones and joints
TNF receptor associated periodic fever	<i>TNFRSF1A</i>	TNFR1	Headache, muscles, and joints
Chronic recurrent multifocal osteomyelitis	<i>LPIN2</i> and <i>PSTPIP2</i>	LIPIN2 and PSTPIP2	Bone
Idiopathic recurrent acute pericarditis	NA	NA	Chest
Schnitzler syndrome	NA	NA	Bone and muscles

Genes and corresponding proteins, where known, are included in this table. TNF: tumor necrosis factor; NA: not applicable.

many innate immune pathways resulting in the production of IL-1 $\beta$  and IL-18. The IL-1 $\beta$  produced by inflammasome activation in autoinflammatory disorders is thought to be a key mediator underlying painful symptoms.<sup>22–25</sup>

The complement system can be activated by multiple pathways including ones that are a part of the innate system of immunity generating complement split products.<sup>26,27</sup> Anaphylatoxin split products including C3a and C5a both intensify inflammation and can contribute to pain directly through interactions with their respective receptors on neurons or indirectly by stimulating the production of cytokines and neurotrophins in the surrounding tissue.<sup>28–30</sup> In addition, the membrane attack complex C5b-9 (MAC) formed by complement system activation can lead to nerve damage and Wallerian degeneration.<sup>31</sup> Recently, it was suggested that sublytic concentrations of MAC help to regulate acute and chronic inflammation through the activation of ERK1 and other intracellular signaling pathways in neurons and additional cell types.<sup>32,33</sup>

### Autoinflammation and CRPS (cytokine production)

Observations made in CRPS patients, as well as the results of experiments using animal models, support the notion that autoinflammation contributes to multiple manifestations of CRPS. Early stage CRPS patients often exhibit the classical signs of acute inflammation including *rubor* (erythema), *tumor* (swelling and edema), *calor* (warmth), *dolor* (pain), and loss of function. Indeed, acute CRPS can be mistaken for infection, compartment syndrome, and other conditions involving acute inflammation. As reviewed above, autoinflammatory conditions have as their hallmark the generation of inflammation-related immune molecules including cytokines and complement fragments, the accumulation of innate immune cells such as mast cells, and the

activation of dendritic cells in the absence of a foreign pathogen. All of these occur in CRPS.

Elevations in skin cytokine levels including TNF $\alpha$ , IL-1 $\beta$ , IL-6, and others have been demonstrated in human volunteers and patients after minor mechanical trauma,<sup>34</sup> fracture,<sup>35</sup> burns,<sup>36</sup> and surgery.<sup>37</sup> Using immunohistochemical analysis and immunoassays of skin suction blister fluid, elevated skin cytokine levels have been documented in CRPS patients at various stages of the syndrome as well. For example, suction blister fluid from CRPS patients was found to contain elevated levels of IL-6, TNF $\alpha$ , and ET-1,<sup>38,39</sup> though these levels did not correlate strongly with the stage of the syndrome.<sup>40</sup> Immunohistochemical studies demonstrated that keratinocytes in the skin ipsilateral to CRPS symptoms express higher levels of IL-6 and TNF $\alpha$  than the skin of the contralateral limb.<sup>41</sup> Similar studies have been performed on serum from CRPS and control patients showing higher levels of cytokines such as IL-6 and TNF $\alpha$  along with lower levels of anti-inflammatory cytokines such as IL-10.<sup>42,43</sup> Local TNF $\alpha$  activity may correlate with mechanical allodynia in CRPS patients.<sup>44</sup> Imaging studies indicate that TNF $\alpha$  accumulates in the joints and other tissues of CRPS limbs during the acute phase of the syndrome,<sup>45</sup> and biologic anti-TNF $\alpha$  agents have shown some promise in the treatment of CRPS.<sup>46–49</sup>

The mechanisms by which cytokine levels increase and support the varied manifestations of CRPS have been studied extensively in animal models. Using the well-validated rodent tibia fracture/cast immobilization model, it has been shown that skin and, to a lesser extent, muscle levels of IL-1 $\beta$ , IL-6, and TNF $\alpha$  are elevated.<sup>50–54</sup> Although better investigated in animal models than in humans, the pain-related neurotrophin nerve growth factor (NGF) also appears to be strongly elevated in the skin of these model animals.<sup>55</sup> Furthermore, both small molecule and biologic therapies

targeting NGF and cytokine signaling reduced the allodynia, hindpaw unweighting, and some of the vascular and autonomic CRPS-like features demonstrated in the model animals in these studies. Similar to the findings in humans, the keratinocyte layer was found to be responsible for generating most of the involved mediators,<sup>41,56</sup> though again deeper tissues such as muscle do seem to generate pain-related cytokines.<sup>50</sup> Mitogen-activated protein kinase (MAPK) activation within keratinocytes was functionally linked to the enhanced cytokine production.<sup>56</sup> Additional studies on CRPS model animals demonstrated the activation of inflammasomes in keratinocytes as a required step in the production of IL-1 $\beta$  in the tibial fracture/cast model consistent with established autoinflammatory pathways.<sup>52</sup>

A striking feature of these observations was the requirement for intact neuropeptide and sympathetic nervous system signaling. Blockade of the substance P (SP) NK1 receptor using the selective antagonist LY303870 was observed to block nociceptive and vascular CRPS-like changes in models of both CRPS Type I (fracture/cast) and CRPS Type II (sciatic transection).<sup>57,58</sup> Subsequent studies using neuropeptide signaling-deficient animals showed that SP and calcitonin gene-related peptide (CGRP) signaling through efferent peptidergic neurons is required for enhanced levels of IL-1 $\beta$ , TNF $\alpha$ , and NGF.<sup>59</sup> The production of these mediators by keratinocytes is reliant upon neuropeptide-mediated activation of ERK1/2 and JNK MAPK signaling pathways.<sup>56</sup> Clinical evidence exists for exaggerated SP signaling in CRPS; the application of exogenous SP in CRPS patient limbs shows increased SP-induced plasma protein extravasation in both affected and unaffected limbs of patients.<sup>60</sup>

The mechanism for upregulation of SP and CGRP signaling after fracture has not been fully elucidated, but oxidative stress may be involved. Several lines of evidence from human studies suggest oxidative stress in the limbs of patients with CRPS.<sup>61-63</sup> Nociceptive sensitization and elevated markers of oxidative stress have been noted in the limbs of laboratory animals used with either the ischemia-reperfusion or the tibial fracture model of CRPS, changes that can be reduced by the administration of antioxidant drugs like N-acetyl cysteine and vitamin C.<sup>50,64</sup> Using the tibial fracture model, it was observed that oxidative stress caused the upregulation of SP and CGRP that in turn were critical for the production of inflammatory cytokines (IL-6) and the pain-related neurotrophin NGF.<sup>50</sup>

While the most extensive data regarding inflammation and cytokine production in CRPS pertain to skin, muscle, and joints, elevations in cytokine production in spinal cord tissue have been noted as well after fracture and cast immobilization. Specifically, elevations in IL-6 and CCL2, cytokines often associated with activated

glia, were identified in spinal cord tissue ipsilateral to tibial fracture in rats one and four weeks after injury.<sup>65</sup> Similar to the results in skin, c-fiber activity was shown to underlie microglial activation and cytokine production, and the NK1 receptor antagonist LY303870 blocked the glial and cytokine changes.

The autonomic nervous system may also control the production of inflammatory mediators in CRPS. For example, studies using chemical sympathectomy suggested that the sympathetic nervous system is responsible primarily for IL-6 upregulation after fracture and cast immobilization, and that this sympathetically supported IL-6 production is required for the full manifestation of CRPS-like symptoms.<sup>66</sup> Further investigation demonstrated that  $\beta$ 2 adrenergic receptors are particularly important to IL-6 upregulation. The parasympathetic nervous system, on the other hand, has been much less well explored in relation of CRPS. The well-demonstrated roles of cholinergic stimulation, particularly through the vagus nerve, suggest that inadequate parasympathetic tone might contribute to the excessive production of inflammatory mediators as may be the case in rheumatoid arthritis.<sup>67</sup>

### *Autoinflammation and CRPS (immune cell contributions)*

Additional cellular immune system components including mast cells and Langerhans dendritic cells, which contribute to both innate and adaptive immunity, may contribute to CRPS. Evidence for mast cell involvement comes from skin blister fluid studies in which the mast-cell marker protein tryptase was found to be elevated in the CRPS affected versus contralateral limbs of patients.<sup>68</sup> Complementary data using skin biopsies from the ipsilateral versus contralateral skin of CRPS patients showed an accumulation of mast cells ipsilateral to the CRPS symptoms, particularly in subjects with relatively acute disease.<sup>41</sup> Patients with very chronic CRPS failed to show such differences in mast cell abundance.<sup>69</sup> Mast cell migration and degranulation is influenced by SP signaling in humans, and such signaling could be involved in the accumulation of mast cells in the limbs of CRPS patients.<sup>70,71</sup> In humans, the contribution of mast cells to CRPS pain has not been established directly, though pain is a prominent feature of mastocytosis, a disease in which abnormally large numbers of mast cells accumulate.<sup>72</sup> Additionally, mast cells are contributors to neurogenic inflammation, a well-described component of CRPS.<sup>73</sup>

Extensive investigation of mast cell function has been undertaken using animal models. Mast cell accumulation and degranulation in the dermis has been observed after tibial fracture and cast immobilization in rats.<sup>74</sup> The accumulation and degranulation of these cells



controls nociceptive sensitization in this model of CRPS. Further investigation showed that the degranulation of the dermal mast cells was dependent upon SP signaling through the NK1 receptor consistent with established human physiology governing mast cell degranulation. In fact, SP-containing nerve fibers were found to be in close proximity to the infiltrating mast cells. Mast cells release a wide range of mediators capable of supporting nociception, though the specific nociceptive pathways have not been defined in clinical CRPS or animal models.

Likewise, abundant Langerhans cells were identified in skin samples from some CRPS patients using immunohistochemical techniques,<sup>75,76</sup> although a more definitive study involving primarily later stage patients found diminished Langerhans cell numbers in CRPS skin.<sup>69</sup> Interestingly, the migration of Langerhans cells in skin is regulated in part through  $\alpha 1$  adrenergic receptor signaling,<sup>77</sup> and signaling through  $\beta$  adrenergic receptors controls cytokine release from these cells.<sup>78</sup> Aberrant adrenergic signaling is a key component of CRPS pathophysiology.<sup>4</sup> However, recent studies using the well-validated rodent tibial fracture/cast immobilization model of CRPS failed to demonstrate any effects of Langerhans cell depletion casting some doubt on this hypothesis or perhaps highlighting immunological differences between humans and mice.<sup>76</sup>

### Autoimmunity

Autoimmune conditions are ones in which the body produces an immune response against its own tissues. Autoimmunity involves the adaptive immune system, a system of cells and processes targeting specific antigens. Both B and T lymphocytes participate in adaptive immunity, and in many instances, this form of immunity involves the formation of antibodies released into the blood and other tissues to find their targets. An additional component of the complex biology governing adaptive immunity are dendritic or Langerhans cells. Langerhans cells are found in especially high numbers in the gut and skin where they are positioned to detect

early the presence of foreign proteins on invading microorganisms.<sup>79</sup> Mentioned previously as participants in the innate system of immunity due to their ability to phagocytose invading organisms and to produce inflammatory mediators such as cytokines, these are also one type of antigen-presenting cell. Under certain conditions, Langerhans cells activate CD4+ helper T cells which in turn stimulate B cells to produce antibodies against the presented antigen.<sup>80</sup> Langerhans cells are capable of presenting self-antigens thus stimulating an adaptive immune response against intrinsic tissues. However, antigen presentation capability is not limited to Langerhans cells.<sup>81</sup>

A wide variety of tissues can be the target of autoimmunity, and the list of diseases with autoimmune pathology is growing rapidly. In addition to classic autoimmune diseases such as rheumatoid arthritis, type-1 diabetes, Hashimoto's thyroiditis, and myasthenia gravis, autoimmunity is now believed to play a role in several heart and lung diseases, narcolepsy, and certain forms of encephalitis.<sup>82-84</sup> Many autoimmune conditions involve antibodies directed against components of the central or peripheral nervous systems, and some of these conditions involve pain as a primary symptom (Table 2). For example, Guillain-Barre syndrome (GBS) involves a polyneuropathy affecting both motor and sensory components of peripheral nerves.<sup>85,86</sup> Autoantibodies have been identified in variable percentages of GBS patients against neurofascin, gliomedin, contactin, GM1 ganglioside, and several additional proteins.<sup>87,88</sup> Recent evidence suggests that many cases of idiopathic small fiber neuropathy may be due to autoantibodies against these fibers.<sup>89</sup> Although rare, a pain syndrome caused by anti-voltage-gated potassium channel complex antibodies has been described.<sup>90</sup> Nearly 50% of patients with such antibodies had pain, and nearly one-third had pain as the only presenting symptom. In the case of paraneoplastic neuropathies, autoimmunity involving anti-Hu and anti-CV2/CRIMP5 antibodies has been linked to painful sensory changes.<sup>91</sup>

Autonomic changes tend to be less of a therapeutic focus when confronted with a patient suffering from

**Table 2.** Autoimmune diseases characterized by painful symptoms.

Autoimmune peripheral neuropathy	Target proteins
Guillain-Barré syndrome	Gangliosides (several)
Chronic inflammatory demyelinating polyneuropathy	Glycolipids and P0
Multifocal motor neuropathy	Neurofascin-186 and gliomedin
Anti-myelin-associated glycoprotein antibody-mediated neuropathy	Myelin-associated glycoprotein and gangliosides
Paraneoplastic peripheral neuropathies	Hu, CV2, and Ma
Voltage-gated potassium channel autoimmunity	Voltage-gated potassium channel complexes
Autoimmune autonomic neuropathy	P/Q Ca <sup>2+</sup> channels and nicotinic ACh receptor

Several of the most commonly recognized autoimmune diseases in which patients frequently report pain are listed along with one or more of the target proteins for disease-related autoantibodies.

CRPS, yet autonomic dysfunction is a component of this syndrome.<sup>43,92,93</sup> It is notable then that several types of autoimmune autonomic neuropathies have been described,<sup>94</sup> and some degree of autonomic dysfunction, for example, cardiovascular reflex tests, heart rate variability, and sympathetic skin tests, has been found in rheumatoid arthritis, the archetypical painful autoimmune disease.<sup>95</sup>

### *Autoimmunity and CRPS*

Emerging evidence suggests that autoimmunity may contribute to CRPS.<sup>96</sup> A collection of observations made over the past decade suggest an autoimmune etiology for CRPS, thus helping to explain the seemingly unrelated nature of the syndrome's signs and symptoms as well as difficulties in achieving adequate symptom control, remission, or cure using standard therapies. Exploration of CRPS-related autoimmunity began with the serendipitous observation of symptom improvement in CRPS patients treated with intravenous immunoglobulin for unrelated conditions. Later, a small randomized clinical trial using low-dose intravenous immunoglobulin provided positive results,<sup>97</sup> although a subsequent larger scale trial failed to reveal significant effects using this drug.<sup>98</sup> Plasma exchange therapy has also been shown effective in reducing pain in CRPS patients,<sup>99</sup> but larger trials are required to confirm these results.

The autoimmune hypothesis is bolstered by several additional sets of observations. First, it was demonstrated that a disproportionate number of patients had IgM and IgG profiles consistent with antecedent infections by chlamydia, parvovirus, and campylobacter.<sup>100,101</sup> Cross-reactivity of antichlamydia and campylobacter antibodies with self-antigens explains some cases of autoimmune neuropathy.<sup>102,103</sup> Second, experiments using immunohistochemical techniques and cytometric analysis identified sympathetic nervous system neurons as targets for autoantibodies from some CRPS patients with little evidence of such autoimmunity from patients with other types of peripheral neuropathy.<sup>101,104</sup> Subsequent experiments using an *in vitro* beating cardiomyocyte preparation suggested that a majority of CRPS patients but not healthy controls have autoantibodies binding to and activating the M-2 muscarinic,  $\beta$ -2 adrenergic ( $\beta$ 2-AR), or  $\alpha$ -1 adrenergic receptors.<sup>105,106</sup> Interestingly, other patients expressing anti- $\beta$ 2-AR autoantibodies display orthostatic hypotension and additional nonpain symptoms suggesting adrenergic receptor autoantibody expression alone may not be sufficient to cause CRPS.<sup>107</sup> Third, CD14+ monocytes are elevated in the blood of CRPS sufferers, and those cell counts correlate with allodynia severity.<sup>108</sup> Fourth, specific human leukocyte antigen (HLA) immune alleles are associated with CRPS.<sup>109–111</sup> HLA-B62 and HLA-DQ8 have been

associated with CRPS with fixed dystonia, while HLA-DQ8 alone was associated with CRPS without dystonia. Last, CRPS shows a 3–4:1 female:male predominance similar to the strong female predominance characterizing most autoimmune conditions.<sup>2,112</sup>

Animal models have been employed to pursue the hypothesis that autoimmunity contributes to the signs and symptoms of CRPS. Paradigms of passive transfer of immunoglobulins have been used to address the question of whether antibodies themselves are involved. For example, Goebel et al. injected purified IgG from CRPS patients intraperitoneally in control mice. Although nociceptive behaviors were not changed in these mice, the animals displayed both changes in rearing behavior and performance in a task involving balance and coordination perhaps consistent with the motor changes in CRPS patients.<sup>113</sup> A follow-up study involved the administration of CRPS or control patient IgG to mice in which hindpaw incision had been made mimicking the trauma normally preceding the development of CRPS. In this model, both mechanical hyperalgesia and edema were enhanced by the CRPS patient IgG. In addition, the CRPS IgG increased wound area SP levels.<sup>114</sup> Separate experiments employed the well-characterized tibial fracture/cast immobilization model of CRPS in which to study autoimmune contributions. In CRPS model mice in which CD20+ B cells had been depleted using a biologic agent, the manifestation of allodynia, postural unweighting, and vascular changes were all attenuated. The results were nearly identical when the same measurements were made in the muMT mice that do not produce mature B cells.<sup>115</sup> More recent autoantigen discovery experiments identified several potential autoantigenic proteins, and one protein, keratin 16, seemed to be reactive with both murine CRPS model IgM and IgG antibodies from CRPS patients.<sup>116</sup> The passive transfer of IgM but not IgG antibodies purified from CRPS model wild-type mice reconstituted nociceptive sensitization in CRPS model muMT mice.<sup>117</sup> It is not clear whether CRPS-related IgM autoantibodies lead to pain via a direct interaction with their targets, or whether the deposition of antibodies promotes subsequent responses such as the activation of complement.

Additional experiments showed that knockdown of B cells did not affect postfracture increases in skin cytokine levels, though the deposition of complement fragments in peripheral tissues was diminished.<sup>115</sup> The complement cascade is activated through the classical pathway by antibodies bound to tissue antigens. With regard to complement activation, IgM class antibodies are particularly efficient.<sup>118</sup> This activation produces C5a, a nociceptive mediator we have shown to support allodynia through the activation of C5a receptors on sensory neurons.<sup>28,30</sup> C5a also causes the migration and degranulation of mast cells in skin that we and others have linked to pain in

CRPS patients and in the fracture/cast model.<sup>41,74</sup> Separately, complement activation leads to the formation of MAC complexes thus damaging peripheral nerves.<sup>119,120</sup> In fact, cutaneous neurite loss has been reported in CRPS patients,<sup>121</sup> and we have shown skin and sciatic nerve MAC levels to be increased in B cell-dependent fashion in the mouse tibial fracture CRPS model.<sup>115</sup>

### *The vexing question of regional symptoms*

One of the principal characteristics of CRPS is that in the majority of cases a single limb is involved, although spread, most often to the contralateral limb, is sometimes seen.<sup>122</sup> How are symptoms limited to a single limb if contributing autoimmune antibodies are present in serum? We are only beginning to understand how regional autoimmunity might function, but it is notable that in the mouse fracture/cast immobilization model we found deposits of immunoglobulins in the skin and sciatic nerves ipsilateral but not contralateral to the fractures.<sup>115</sup> Furthermore, in both humans as well as animal models, Langerhans cell accumulation was greater in the skin of the affected compared to the contralateral limbs<sup>75,76</sup> although, as mentioned above, the analysis of skin from patients with longstanding CRPS (years) did not show such changes.<sup>69</sup> In the setting of CRPS, new antigens (neoantigens) could be regionally expressed, posttranslational modifications of existing proteins might render them immunogenic such as by citrullination or carbamylation, the compartmentalization of proteins might be altered or the target antigens might lose their “immune privileged” status in the setting of CRPS by virtue of a change in a tissue barrier. Each has been posited as an explanation for autoimmunity in other disease states, though none of these mechanisms have been conclusively demonstrated to occur in CRPS.

### *Neural control of autoimmune mechanisms in CRPS*

An intriguing possibility is that regional changes in peptidergic and sympathetic function in the affected limbs could be supporting autoimmune-related changes similar to how these systems support changes in innate immune function as previously discussed. Neural control of adaptive immunity is a novel concept, but one for which evidence is beginning to accumulate. Using the fracture/cast mouse model of CRPS, it was shown that the accumulation of immunoglobulins in the skin ipsilateral to injury was dependent on NK1 receptor signaling. Likewise, the sera of CRPS model mice in which the SP coding *tac1* gene had been deleted was not capable of reconstituting nociceptive sensitization in muMT fracture/cast model mice, and the *tac1*<sup>-/-</sup> mice did not display IgM accumulation in skin, peripheral nerves, or

spinal cord tissue after fracture as was observed in the wild-type mice. The results were similar for mice in which CGRP signaling was disrupted.<sup>76</sup> Interestingly, SP signaling has been implicated in the disruption of immune privilege in autoimmune diseases of the CNS and in regionally localized conditions like alopecia areata.<sup>123,124</sup> Furthermore, it was observed that the enhanced postfracture expression of one confirmed autoantigen, Krt16, was dependent on intact SP and CGRP signaling.<sup>76</sup>

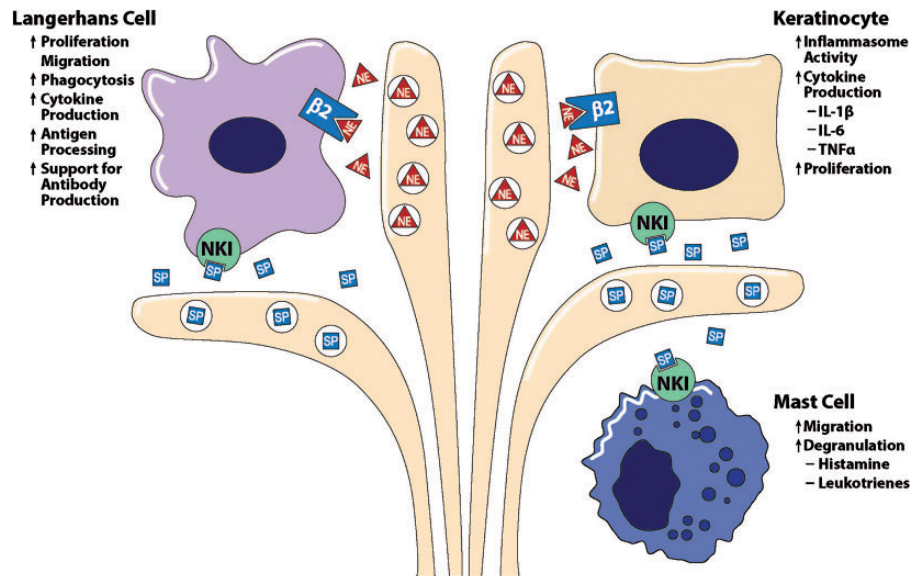
Loss of immune privilege of the CNS including the spinal cord and higher centers is particularly intriguing. Most studies to this point have examined autoimmune responses against peripheral nerves and other tissues, but IgM antibodies are found in spinal cord tissue in the rodent fracture model.<sup>76</sup> Pain and other consequences might result from interactions of autoantibodies with specific CNS targets, for example, ion channels, cell surface receptors, or might modulate nociception through the fixation of complement. Elevated spinal levels of C5a in nerve injury models have been linked to nociceptive sensitization.<sup>125</sup>

Dysfunction of the sympathetic nervous system has been observed in some CRPS patients, and the older name for the syndrome “Reflex Sympathetic Dystrophy” highlighted this association. Furthermore, it has been reported that sympathectomy can reduce the inflammation and pain associated with the classic autoimmune condition rheumatoid arthritis.<sup>126,127</sup> In animal models of rheumatoid arthritis, sympathetic signaling through  $\beta$ 2-AR on B cells is responsible for stimulating autoantibody production particularly in the acute phases of the condition.<sup>128,129</sup> The precise mechanisms for these effects are unclear, though both B cell and Langerhans cell activity can be regulated by norepinephrine through the activation of  $\alpha$  and  $\beta$  adrenergic receptors.<sup>78,130</sup> Thus, regionally activated neuroimmune networks involving peptidergic, sympathetic, or both systems may play a role in the autoimmune manifestations of CRPS.

## **Conclusions**

CRPS is an enigmatic syndrome for which no convincingly effective treatments exist. One of the most confounding aspects of the syndrome has been the disparate mix of manifestations involving multiple tissues thereby defying the identification of a clear unifying etiology. Recent evidence suggests that the interrelated phenomena of autoinflammation and autoimmunity may be the basis for this syndrome. Dysfunction of neuropeptide-containing afferent neurons and sympathetic fibers may support the inflammation and autoimmune phenomena, at least in CRPS Type I where no clear nerve lesion is present. Much less information is





**Figure 1.** A neuroinflammatory model of Complex Regional Pain Syndrome. Data from patients and laboratory models suggest both neuropeptides and autonomic nervous system activity contribute to the pathogenesis of CRPS. Antigen-presenting cells, epithelial cells and mast cells may all be contributing components. NE: norepinephrine; SP: Substance P;  $\beta 2$ :  $\beta 2$  adrenergic receptor.

available regarding mechanistic overlap with CRPS Type II, although neuropeptide dysregulation seems to be involved.<sup>131</sup> In Figure 1, a summary of the interactions of neuropeptidergic and sympathetic transmitters with key cells participating in CRPS-related autoimmunity and autoinflammation is presented. An attractive aspect of this hypothesis is that a number of new therapeutic approaches could be considered to combat autoinflammation and autoimmunity. For example, biologic anti-cytokine agents (TNF $\alpha$ , IL-1 $\beta$ , IL-6, etc.) are available and could be tested in clinical populations. Likewise, agents targeting components of the adaptive immune system such as rituximab (anti-CD20) or calcineurin inhibitors might be tested. Peptidergic and sympathetic signaling blockers may reduce both autoinflammatory and autoimmune responses. One challenge, however, will be to weigh the potential benefits of these powerful immunological agents against susceptibility to infection and certain forms of cancer, problems currently faced by patients taking these drugs to relieve rheumatological conditions. Still, for patients not improving with more conservative therapies or time, immunomodulatory agents might at some point become viable options for reducing the pain, disability, and other consequences of CRPS.

#### Author Contributions

JDC conceived, outlined, and wrote substantial portions of the manuscript. VLT and MT provided discussion and feedback on the manuscript's goals, read the manuscript, and provided text, references, and comments. WSK provided extensive comments, text, and revisions of the manuscript.

#### Declaration of Conflicting Interests

The author(s) declared no potential conflicts of interest with respect to the research, authorship, and/or publication of this article.

#### Funding

The author(s) disclosed receipt of the following financial support for the research, authorship, and/or publication of this article: This study was supported by grants from the Department of Veteran Affairs I01RX001475 and NIH NS072143 and NS094438.

#### References

- Schwartzman RJ, Erwin KL and Alexander GM. The natural history of complex regional pain syndrome. *Clin J Pain* 2009; 25: 273–280.
- de Mos M, de Bruijn AG, Huygen FJ, Dieleman JP, Stricker BH and Sturkenboom MC. The incidence of complex regional pain syndrome: a population-based study. *Pain* 2007; 129: 12–20.
- Sarangi PP, Ward AJ, Smith EJ, Staddon GE and Atkins RM. Algodystrophy and osteoporosis after tibial fractures. *J Bone Joint Surg Br* 1993; 75-B: 450–452.
- Bruehl S. An update on the pathophysiology of complex regional pain syndrome. *Anesthesiology* 2010; 113: 1–725.
- Gierthmuhlen J, Maier C, Baron R, Tolle T, Treede RD, Birbaumer N, Hugel V, Koroschetz J, Krumova EK, Lauchart M, Maihofner C, Richter H and Westermann A; German Research Network on Neuropathic Pain (DFNS) Study Group. Sensory signs in complex regional pain syndrome and peripheral nerve injury. *Pain* 2012; 153: 765–774.



6. Bean DJ, Johnson MH and Kydd RR. The outcome of complex regional pain syndrome type 1: a systematic review. *J Pain* 2014; 15: 677–690.
7. Bean DJ, Johnson MH, Heiss-Dunlop W and Kydd RR. Extent of recovery in the first 12 months of complex regional pain syndrome type-1: a prospective study. *Eur J Pain* 2016; 20: 884–894.
8. Subbarao J and Stillwell GK. Reflex sympathetic dystrophy syndrome of the upper extremity: analysis of total outcome of management of 125 cases. *Arch Phys Med Rehabil* 1982; 14: 198–554.
9. O’Connell NE, Wand BM, McAuley J, Marston L and Moseley GL. Interventions for treating pain and disability in adults with complex regional pain syndrome. *Cochrane Database Syst Rev* 2013; 4: CD009416.
10. Doria A, Zen M, Bettio S, Gatto M, Bassi N, Nalotto L, Ghirardello A, Iaccarino L and Punzi L. Autoinflammation and autoimmunity: bridging the divide. *Autoimmun Rev* 2012; 12: 22–30.
11. Borella E, Palma L, Zen M, Bettio S, Nalotto L, Gatto M, Domeneghetti M, Laccarino L, Punzi L and Doria A. The body against self: autoinflammation and autoimmunity. *Isr Med Assoc J* 2014; 16: 608–610.
12. Cantarini L, Lopalco G, Selmi C, Napodano S, De Rosa G, Caso F, Costa L, Iannone F and Rigante D. Autoimmunity and autoinflammation as the yin and yang of idiopathic recurrent acute pericarditis. *Autoimmun Rev* 2015; 14: 90–97.
13. Hedrich CM. Shaping the spectrum – from autoinflammation to autoimmunity. *Clin Immunol* 2016; 165: 21–28.
14. McGonagle D and McDermott MF. A proposed classification of the immunological diseases. *PLoS Med* 2006; 3: e297.
15. Iolascon G, de Sire A, Moretti A and Gimigliano F. Complex regional pain syndrome (CRPS) type I: historical perspective and critical issues. *Clin Cases Miner Bone Metab* 2015; 12: 4–10.
16. Kastner DL, Aksentijevich I and Goldbach-Mansky R. Autoinflammatory disease reloaded: a clinical perspective. *Cell* 2010; 140: 784–790.
17. Russo RA and Brogan PA. Monogenic autoinflammatory diseases. *Rheumatology* 2014; 53: 1927–1939.
18. Goldbach-Mansky R. Immunology in clinic review series; focus on autoinflammatory diseases: update on monogenic autoinflammatory diseases: the role of interleukin (IL)-1 and an emerging role for cytokines beyond IL-1. *Clin Exp Immunol* 2012; 167: 391–404.
19. Kigerl KA, de Rivero Vaccari JP, Dietrich WD, Popovich PG and Keane RW. Pattern recognition receptors and central nervous system repair. *Exp Neurol* 2014; 258: 5–16.
20. Plato A, Hardison SE and Brown GD. Pattern recognition receptors in antifungal immunity. *Semin Immunopathol* 2014; 37: 97–106.
21. Zhang H, Li F, Li WW, Stary C, Clark JD, Xu S and Xiong X. The inflammasome as a target for pain therapy. *Br J Anaesth* 2016; 117: 693–707.
22. Alvarez-Errico D, Vento-Tormo R and Ballestar E. Genetic and epigenetic determinants in autoinflammatory diseases. *Front Immunol* 2017; 8: 318.
23. de Torre-Minguela C, Mesa del Castillo P and Pelegrín P. The NLRP3 and pyrin inflammasomes: implications in the pathophysiology of autoinflammatory diseases. *Front Immunol* 2017; 8: 43.
24. Hoffman HM and Broderick L. The role of the inflammasome in patients with autoinflammatory diseases. *J Allergy Clin Immunol* 2016; 138: 3–14.
25. Peckham D, Scambler T, Savic S and McDermott MF. The burgeoning field of innate immune-mediated disease and autoinflammation. *J Pathol* 2017; 241: 123–139.
26. Chimenti MS, Ballanti E, Triggianese P and Perricone R. Vasculitides and the complement system: a comprehensive review. *Clin Rev Allergy Immunol* 2015; 49: 333–346.
27. Orsini F, De Blasio D, Zangari R, Zanier ER and De Simoni MG. Versatility of the complement system in neuroinflammation, neurodegeneration and brain homeostasis. *Front Cell Neurosci* 2014; 8: 380.
28. Clark JD, Qiao Y, Li X, Shi X, Angst MS and Yeomans DC. Blockade of the complement C5a receptor reduces incisional allodynia, edema, and cytokine expression. *Anesthesiology* 2006; 104: 1274–1282.
29. Jang JH, Liang D, Kido K, Sun Y, Clark DJ and Brennan TJ. Increased local concentration of complement C5a contributes to incisional pain in mice. *J Neuroinflammation* 2011; 8: 80.
30. Liang DY, Li X, Shi X, Sun Y, Sahbaie P, Li WW and Clark JD. The complement component C5a receptor mediates pain and inflammation in a postsurgical pain model. *Pain* 2012; 153: 366–372.
31. Ramaglia V, King RH, Nourallah M, Wolterman R, de Jonge R, Ramkema M, Vigar MA, van der Wetering S, Morgan BP, Troost D and Baas F. The membrane attack complex of the complement system is essential for rapid Wallerian degeneration. *J Neurosci* 2007; 27: 7663–7672.
32. Tegla CA, Cudrici C, Patel S, Trippe R, Rus V, Niculescu F and Rus H. Membrane attack by complement: the assembly and biology of terminal complement complexes. *Immunol Res* 2011; 51: 45–60.
33. Tran GT, Hodgkinson SJ, Carter NM, Killingsworth M, Nomura M, Verma ND, Plain KM, Boyd R and Hall BM. Membrane attack complex of complement is not essential for immune mediated demyelination in experimental autoimmune neuritis. *J Neuroimmunol* 2010; 229: 98–106.
34. Eberle T, Doganci B, Kramer H, Fehrer M, Wagner I, Sommer C and Birklein F. Mechanical but not painful electrical stimuli trigger TNF alpha release in human skin. *Exp Neurol* 2010; 221: 246–250.
35. Kramer HH, Eberle T, Uceyler N, Wagner I, Klonschinsky T, Muller LP, Sommer C and Birklein F. TNF-alpha in CRPS and ‘normal’ trauma—significant differences between tissue and serum. *Pain* 2011; 152: 285–290.
36. Angst MS, Clark JD, Carvalho B, Tingle M, Schmelz M and Yeomans DC. Cytokine profile in human skin in response to experimental inflammation, noxious stimulation, and administration of a COX-inhibitor: a microdialysis study. *Pain* 2008; 139: 15–27.

37. Pepper A, Li W, Kingery WS, Angst MS, Curtin CM and Clark JD. Changes resembling complex regional pain syndrome following surgery and immobilization. *J Pain* 2013; 14: 516–524.
38. Groeneweg JG, Huygen FJ, Heijmans-Antonissen C, Niehof S and Zijlstra FJ. Increased endothelin-1 and diminished nitric oxide levels in blister fluids of patients with intermediate cold type complex regional pain syndrome type 1. *BMC Musculoskelet Disord* 2006; 7: 91.
39. Huygen FJ, De Bruijn AG, De Bruin MT, Groeneweg JG, Klein J and Zijlstra FJ. Evidence for local inflammation in complex regional pain syndrome type 1. *Mediators Inflamm* 2002; 11: 47–51.
40. Munnikes RJ, Muis C, Boersma M, Heijmans-Antonissen C, Zijlstra FJ and Huygen FJ. Intermediate stage complex regional pain syndrome type 1 is unrelated to proinflammatory cytokines. *Mediators Inflamm* 2005; 2005: 366–372.
41. Birklein F, Drummond PD, Li W, Schlereth T, Albrecht N, Finch PM, Dawson LF, Clark JD and Kingery WS. Activation of cutaneous immune responses in complex regional pain syndrome. *J Pain* 2014; 15: 485–495.
42. Lenz M, Uceyler N, Frettlow J, Hoffken O, Krumova EK, Lissek S, Reinersmann A, Sommer C, Stude P, Waaga-Gasser AM, Tegenthoff M and Maier C. Local cytokine changes in complex regional pain syndrome type I (CRPS I) resolve after 6 months. *Pain* 2013; 154: 2142–2149.
43. Schlereth T, Drummond PD and Birklein F. Inflammation in CRPS: role of the sympathetic supply. *Auton Neurosci* 2014; 182: 102–107.
44. Maihofner C, Handwerker HO, Neundorfer B and Birklein F. Mechanical hyperalgesia in complex regional pain syndrome: a role for TNF-alpha? *Neurology* 2005; 65: 311–313.
45. Bernateck M, Karst M, Gratz KF, Meyer GJ, Fischer MJ, Knapp WH, Koppert W and Brunkhorst T. The first scintigraphic detection of tumor necrosis factor-alpha in patients with complex regional pain syndrome type 1. *Anesth Analg* 2010; 110: 211–215.
46. Dirckx M, Groeneweg G, Wesseldijk F, Stronks DL and Huygen FJ. Report of a preliminary discontinued double-blind, randomized, placebo-controlled trial of the anti-TNF-alpha chimeric monoclonal antibody infliximab in complex regional pain syndrome. *Pain Pract* 2013; 13: 633–640.
47. Dirckx M, Stronks DL, Groeneweg G and Huygen FJ. Effect of immunomodulating medications in complex regional pain syndrome: a systematic review. *Clin J Pain* 2012; 28: 355–363.
48. Eisenberg E, Sandler I, Treister R, Suzan E and Haddad M. Anti tumor necrosis factor-alpha adalimumab for complex regional pain syndrome type 1 (CRPS-I): a case series. *Pain Pract* 2013; 13: 649–656.
49. Miculescu AA, Nordquist L, Hysing EB, Butler S, Basu S, Lind AL and Gordh T. Targeting oxidative injury and cytokines' activity in the treatment with anti-tumor necrosis factor-alpha antibody for complex regional pain syndrome 1. *Pain Pract* 2013; 13: 641–648.
50. Guo TZ, Wei T, Huang TT, Kingery WS and Clark JD. Oxidative stress contributes to fracture/cast-induced inflammation and pain in a rat model of complex regional pain syndrome. *J Pain*. Epub ahead of print 30 April 2018. DOI: 10.1016/j.jpain.2018.04.006.
51. Li WW, Guo TZ, Li XQ, Kingery WS and Clark JD. Fracture induces keratinocyte activation, proliferation, and expression of pronociceptive inflammatory mediators. *Pain* 2010; 151: 843–852.
52. Li WW, Guo TZ, Liang D, Shi X, Wei T, Kingery WS and Clark JD. The NALP1 inflammasome controls cytokine production and nociception in a rat fracture model of complex regional pain syndrome. *Pain* 2009; 147: 277–286.
53. Sabsovich I, Guo TZ, Wei T, Zhao R, Li X, Clark DJ, Geis C, Sommer C and Kingery WS. TNF signaling contributes to the development of nociceptive sensitization in a tibia fracture model of complex regional pain syndrome type I. *Pain* 2008; 137: 507–519.
54. Wei T, Sabsovich I, Guo TZ, Shi X, Zhao R, Li W, Geis C, Sommer C, Kingery WS and Clark DJ. Pentoxifylline attenuates nociceptive sensitization and cytokine expression in a tibia fracture rat model of complex regional pain syndrome. *Eur J Pain* 2009; 13: 253–262.
55. Sabsovich I, Wei T, Guo TZ, Zhao R, Shi X, Li X, Yeomans DC, Klyukin M, Kingery WS and Clark JD. Effect of anti-NGF antibodies in a rat tibia fracture model of complex regional pain syndrome type I. *Pain* 2008; 138: 47–60.
56. Shi X, Wang L, Clark JD and Kingery WS. Keratinocytes express cytokines and nerve growth factor in response to neuropeptide activation of the ERK1/2 and JNK MAPK transcription pathways. *Regul Pept* 2013; 186: 92–103.
57. Guo TZ, Offley SC, Boyd EA, Jacobs CR and Kingery WS. Substance P signaling contributes to the vascular and nociceptive abnormalities observed in a tibial fracture rat model of complex regional pain syndrome type I. *Pain* 2004; 108: 95–107.
58. Kingery WS, Davies MF and Clark JD. A substance P receptor (NK1) antagonist can reverse vascular and nociceptive abnormalities in a rat model of complex regional pain syndrome type II. *Pain* 2003; 104: 75–84.
59. Guo TZ, Wei T, Shi X, Li WW, Hou S, Wang L, Tsujikawa K, Rice KC, Cheng K, Clark DJ and Kingery WS. Neuropeptide deficient mice have attenuated nociceptive, vascular, and inflammatory changes in a tibia fracture model of complex regional pain syndrome. *Mol Pain* 2012; 8: 85.
60. Leis S, Weber M, Isselmann A, Schmelz M and Birklein F. Substance-P-induced protein extravasation is bilaterally increased in complex regional pain syndrome. *Exp Neurol* 2003; 183: 197–204.
61. Baykal T, Seferoglu B, Karsan O, Kiziltunc A and Senel K. Antioxidant profile in patients with complex regional pain syndrome type I. *Int J Rheum Dis* 2014; 17: 156–158.
62. Eisenberg E, Shtahl S, Geller R, Reznick AZ, Sharf O, Ravbinovich M, Erenreich A and Nagler RM. Serum and salivary oxidative analysis in Complex Regional Pain Syndrome. *Pain* 2008; 138: 226–232.

63. Taha R and Blaise GA. Update on the pathogenesis of complex regional pain syndrome: role of oxidative stress. *Can J Anesth/J Can Anesth* 2012; 59: 875–881.
64. Coderre TJ, Xanthos DN, Francis L and Bennett GJ. Chronic post-ischemia pain (CPIP): a novel animal model of complex regional pain syndrome-type I (CRPS-I; reflex sympathetic dystrophy) produced by prolonged hindpaw ischemia and reperfusion in the rat. *Pain* 2004; 112: 94–105.
65. Li WW, Guo TZ, Shi X, Sun Y, Wei T, Clark DJ and Kingery WS. Substance P spinal signaling induces glial activation and nociceptive sensitization after fracture. *Neuroscience* 2015; 310: 73–90.
66. Li W, Shi X, Wang L, Guo T, Wei T, Cheng K, Rice KC, Kingery WS and Clark JD. Epidermal adrenergic signaling contributes to inflammation and pain sensitization in a rat model of complex regional pain syndrome. *Pain* 2013; 154: 1224–1236.
67. Provan SA, Olstad DS, Solberg EE, Smedslund G and Dagfinrud H. Evidence of reduced parasympathetic autonomic regulation in inflammatory joint disease: a meta-analyses study. *Semin Arthritis Rheum* 2018; 48: 134–140.
68. Huygen FJ, Ramdhani N, van Toorenenbergen A, Klein J and Zijlstra FJ. Mast cells are involved in inflammatory reactions during complex regional pain syndrome type 1. *Immunol Lett* 2004; 91: 147–154.
69. Osborne S, Farrell J, Dearman RJ, MacIver K, Naisbitt DJ, Moots RJ, Edwards SW and Goebel A. Cutaneous immunopathology of long-standing complex regional pain syndrome. *Eur J Pain* 2015; 19: 1516–1526.
70. Kulka M, Sheen CH, Tancowny BP, Grammer LC and Schleimer RP. Neuropeptides activate human mast cell degranulation and chemokine production. *Immunology* 2008; 123: 398–410.
71. Wang GD, Wang XY, Liu S, Qu M, Xia Y, Needleman BJ, Mikami DJ and Wood JD. Innervation of enteric mast cells by primary spinal afferents in guinea pig and human small intestine. *Am J Physiol Gastrointest Liver Physiol* 2014; 307: G719–G731.
72. Carter MC, Metcalfe DD and Komarow HD. Mastocytosis. *Immunol Allergy Clin North America* 2014; 34: 181–196.
73. Rosa AC and Fantozzi R. The role of histamine in neurogenic inflammation. *Br J Pharmacol* 2013; 170: 38–45.
74. Li WW, Guo TZ, Liang DY, Sun Y, Kingery WS and Clark JD. Substance P signaling controls mast cell activation, degranulation, and nociceptive sensitization in a rat fracture model of complex regional pain syndrome. *Anesthesiology* 2012; 116: 882–895.
75. Calder JS, Holten I and McAllister RM. Evidence for immune system involvement in reflex sympathetic dystrophy. *J Hand Surg Br* 1998; 23: 147–150.
76. Li WW, Guo TZ, Shi X, Birklein F, Schlereth T, Kingery WS and Clark JD. Neuropeptide regulation of adaptive immunity in the tibia fracture model of complex regional pain syndrome. *J Neuroinflammation* 2018; 15: 105.
77. Maestroni GJ. Dendritic cell migration controlled by alpha 1b-adrenergic receptors. *J Immunol* 2000; 165: 6743–6747.
78. Goyarts E, Matsui M, Mammone T, Bender AM, Wagner JA, Maes D and Granstein RD. Norepinephrine modulates human dendritic cell activation by altering cytokine release. *Exp Dermatol* 2008; 17: 188–196.
79. Sparber F. Langerhans cells: an update. *J Dtsch Dermatol Ges* 2014; 12: 1107–1111.
80. Romani N, Ratzinger G, Pfaller K, Salvenmoser W, Stossel H, Koch F and Stoitzner P. Migration of dendritic cells into lymphatics—the Langerhans cell example: routes, regulation, and relevance. *Int Rev Cytol* 2001; 207: 237–270.
81. Kashem SW, Haniffa M and Kaplan DH. Antigen-presenting cells in the skin. *Annu Rev Immunol* 2017; 35: 469–499.
82. Agmon-Levin N and Selmi C. The autoimmune side of heart and lung diseases. *Clinic Rev Allerg Immunol* 2013; 44: 1–5.
83. Fontana A, Gast H, Reith W, Recher M, Birchler T and Bassetti CL. Narcolepsy: autoimmunity, effector T cell activation due to infection, or T cell independent, major histocompatibility complex class II induced neuronal loss? *Brain* 2010; 133: 1300–1311.
84. Kayser MS and Dalmau J. Anti-NMDA receptor encephalitis, autoimmunity, and psychosis. *Schizophr Res* 2016; 176: 36–40.
85. Joseph SA and Tsao CY. Guillain-Barre syndrome. *Adolesc Med* 2002; 13: 487–494.
86. Ruts L, Drenthen J, Jongen JL, Hop WC, Visser GH, Jacobs BC, van Doorn PA and Dutch GBSSG. Pain in Guillain-Barre syndrome: a long-term follow-up study. *Neurology* 2010; 75: 1439–1447.
87. Devaux JJ, Odaka M and Yuki N. Nodal proteins are target antigens in Guillain-Barre syndrome. *J Peripher Nerv Syst* 2012; 17: 62–71.
88. Yuki N. Guillain-Barre syndrome and anti-ganglioside antibodies: a clinician-scientist's journey. *Proc Jpn Acad Ser B Phys Biol Sci* 2012; 88: 299–326.
89. Liu X, Treister R, Lang M and Oaklander AL. IVIg for apparently autoimmune small-fiber polyneuropathy: first analysis of efficacy and safety. *Ther Adv Neurol Disord* 2018; 11: 1756285617744484.
90. Klein CJ, Lennon VA, Aston PA, McKeon A and Pittock SJ. Chronic pain as a manifestation of potassium channel-complex autoimmunity. *Neurology* 2012; 79: 1136–1144.
91. Koike H, Tanaka F and Sobue G. Paraneoplastic neuropathy: wide-ranging clinicopathological manifestations. *Curr Opin Neurol* 2011; 24: 504–510.
92. Terkelsen AJ, Gierthmuhlen J, Petersen LJ, Knudsen L, Christensen NJ, Kehr J, Yoshitake T, Madsen CS, Wasner G, Baron R and Jensen TS. Cutaneous noradrenaline measured by microdialysis in complex regional pain syndrome during whole-body cooling and heating. *Exp Neurol* 2013; 247: 456–465.
93. Terkelsen AJ, Molgaard H, Hansen J, Finnerup NB, Kroner K and Jensen TS. Heart rate variability in complex regional pain syndrome during rest and mental and orthostatic stress. *Anesthesiology* 2012; 116: 133–146.



94. Mazzeo A, Stancanelli C, Di Leo R and Vita G. Autonomic involvement in subacute and chronic immune-mediated neuropathies. *Autoimmun Dis* 2013; 2013: 549465.
95. Adlan AM, Lip GY, Paton JF, Kitas GD and Fisher JP. Autonomic function and rheumatoid arthritis-A systematic review. *Semin Arthritis Rheum* 2014; 44: 283–304.
96. Goebel A and Blaes F. Complex regional pain syndrome, prototype of a novel kind of autoimmune disease. *Autoimmun Rev* 2013; 12: 682–686.
97. Goebel A, Baranowski A, Maurer K, Ghiai A, McCabe C and Ambler G. Intravenous immunoglobulin treatment of the complex regional pain syndrome: a randomized trial. *Ann Intern Med* 2010; 152: 152–158.
98. Goebel A, Bisla J, Carganillo R, Frank B, Gupta R, Kelly J, McCabe C, Murphy C, Padfield N, Phillips C, Sanders M, Serpell M, Shenker N, Shoukrey K, Wyatt L and Ambler G. Low-dose intravenous immunoglobulin treatment for long-standing complex regional pain syndrome. *Ann Intern Med* 2017; 167: 476–483.
99. Aradillas E, Schwartzman RJ, Grothusen JR, Goebel A and Alexander GM. Plasma exchange therapy in patients with complex regional pain syndrome. *Pain Phys* 2015; 18: 383–394.
100. Goebel A. Screening of patients with complex regional pain syndrome for antecedent infections. *Clin J Pain* 2001; 17: 378–379.
101. Goebel A, Vogel H, Caneris O, Bajwa Z, Clover L, Roewer N, Schedel R, Karch H, Sprotte G and Vincent A. Immune responses to *Campylobacter* and serum auto-antibodies in patients with complex regional pain syndrome. *J Neuroimmunol* 2005; 162: 184–189.
102. Caudie C, Quittard Pinon A, Taravel D, Sivadon-Tardy V, Orlikowski D, Rozenberg F, Sharshar T, Raphaël J C and Gaillard J L. Preceding infections and anti-ganglioside antibody profiles assessed by a dot immunoassay in 306 French Guillain-Barre syndrome patients. *J Neurol* 2011; 258: 1958–1964.
103. Orlikowski D, Porcher R, Sivadon-Tardy V, Quincampoix JC, Raphael JC, Durand MC, Sharshar T, Roussi J, Caudie C, Annane D, Rozenberg F, Leruez-Ville M, Gaillard JL and Gault E. Guillain-Barre syndrome following primary cytomegalovirus infection: a prospective cohort study. *Clin Infect Dis* 2011; 52: 837–844.
104. Kohr D, Tschernatsch M, Schmitz K, Singh P, Kaps M, Schafer KH, Diener M, Mathies J, Matz O, Kummer W, Maihofner C, Fritz T, Birklein F and Blaes F. Autoantibodies in complex regional pain syndrome bind to a differentiation-dependent neuronal surface autoantigen. *Pain* 2009; 143: 246–251.
105. Dubuis E, Thompson V, Leite MI, Blaes F, Maihofner C, Greensmith D, Vincent A, Shenker N, Kuttikat A, Leuwer M and Goebel A. Longstanding complex regional pain syndrome is associated with activating autoantibodies against alpha-1a adrenoceptors. *Pain* 2014; 155: 2408–2417.
106. Kohr D, Singh P, Tschernatsch M, Kaps M, Pouokam E, Diener M, Kummer W, Birklein F, Vincent A, Goebel A, Wallukat G and Blaes F. Autoimmunity against the beta2 adrenergic receptor and muscarinic-2 receptor in complex regional pain syndrome. *Pain* 2011; 152: 2690–2700.
107. Yu X, Stavrakis S, Hill MA, Huang S, Reim S, Li H, Khan M, Hamlett S, Cunningham MW and Kem DC. Autoantibody activation of beta-adrenergic and muscarinic receptors contributes to an “autoimmune” orthostatic hypotension. *J Am Soc Hypertens* 2012; 6: 40–47.
108. Ritz BW, Alexander GM, Nogusa S, Perreault MJ, Peterlin BL, Grothusen JR and Schwartzman RJ. Elevated blood levels of inflammatory monocytes (CD14+ CD16+) in patients with complex regional pain syndrome. *Clin Exp Immunol* 2011; 164: 108–117.
109. de Rooij AM, Florencia Gosso M, Haasnoot GW, Marinus J, Verduijn W, Claas FH, van den Maagdenberg AM and van Hilten JJ. HLA-B62 and HLA-DQ8 are associated with complex regional pain syndrome with fixed dystonia. *Pain* 2009; 145: 82–85.
110. Jin EH, Zhang E, Ko Y, Sim WS, Moon DE, Yoon KJ, Hong JH and Lee WH. Genome-wide expression profiling of complex regional pain syndrome. *PLoS One* 2013; 8: e79435.
111. van Rooijen DE, Roelen DL, Verduijn W, Haasnoot GW, Huygen FJ, Perez RS, Claas FH, Marinus J, van Hilten JJ and van den Maagdenberg AM. Genetic HLA associations in complex regional pain syndrome with and without dystonia. *J Pain* 2012; 13: 784–789.
112. Sandroni P, Benrud-Larson LM, McClelland RL and Low PA. Complex regional pain syndrome type I: incidence and prevalence in Olmsted county, a population-based study. *Pain* 2003; 103: 199–207.
113. Goebel A, Leite MI, Yang L, Deacon R, Cendan CM, Fox-Lewis A and Vincent A. The passive transfer of immunoglobulin G serum antibodies from patients with longstanding complex regional pain syndrome. *Eur J Pain* 2011; 15: 504.e1–506.
114. Tekus V, Hajna Z, Borbely E, Markovics A, Bagoly T, Szolcsanyi J, Thompson V, Kemeny A, Helyes Z and Goebel A. A CRPS-IgG-transfer-trauma model reproducing inflammatory and positive sensory signs associated with complex regional pain syndrome. *Pain* 2014; 155: 299–308.
115. Li WW, Guo TZ, Shi X, Czirr E, Stan T, Sahbaie P, Wyss-Coray T, Kingery WS and Clark JD. Autoimmunity contributes to nociceptive sensitization in a mouse model of complex regional pain syndrome. *Pain* 2014; 155: 2377–2389.
116. Tajerian M, Hung V, Khan H, Lahey LJ, Sun Y, Birklein F, Kramer HH, Robinson WH, Kingery WS and Clark JD. Identification of KRT16 as a target of an autoantibody response in complex regional pain syndrome. *Exp Neurol* 2017; 287: 14–20.
117. Guo TZ, Shi X, Li WW, Wei T, Clark JD and Kingery WS. Passive transfer autoimmunity in a mouse model of complex regional pain syndrome. *Pain* 2017; 158: 2410–2421.
118. Daha NA, Banda NK, Roos A, Beurskens FJ, Bakker JM, Daha MR and Trouw LA. Complement activation by (auto-) antibodies. *Mol Immunol* 2011; 48: 1656–1665.



119. Putzu GA, Figarella-Branger D, Bouvier-Labit C, Liprandi A, Bianco N and Pellissier JF. Immunohistochemical localization of cytokines, C5b-9 and ICAM-1 in peripheral nerve of Guillain-Barre syndrome. *J Neurol Sci* 2000; 174: 16–21.
120. Rosoklija GB, Dwork AJ, Younger DS, Karlikaya G, Latov N and Hays AP. Local activation of the complement system in endoneurial microvessels of diabetic neuropathy. *Acta Neuropathol* 2000; 99: 55–62.
121. Oaklander AL, Rissmiller JG, Gelman LB, Zheng L, Chang Y and Gott R. Evidence of focal small-fiber axonal degeneration in complex regional pain syndrome-I (reflex sympathetic dystrophy). *Pain* 2006; 120: 235–243.
122. van Rijn MA, Marinus J, Putter H, Bosselaar SR, Moseley GL and van Hilten JJ. Spreading of complex regional pain syndrome: not a random process. *J Neural Transm* 2011; 118: 1301–1309.
123. Reinke E and Fabry Z. Breaking or making immunological privilege in the central nervous system: the regulation of immunity by neuropeptides. *Immunol Lett* 2006; 104: 102–109.
124. Siebenhaar F, Sharov AA, Peters EM, Sharova TY, Syska W, Mardaryev AN, Freyschmidt-Paul P, Sundberg JP, Maurer M and Botchkarev VA. Substance P as an immunomodulatory neuropeptide in a mouse model for autoimmune hair loss (alopecia areata). *J Invest Dermatol* 2007; 127: 1489–1497.
125. Griffin RS, Costigan M, Brenner GJ, Ma CH, Scholz J, Moss A, Allchorne AJ, Stahl GL and Woolf CJ. Complement induction in spinal cord microglia results in anaphylatoxin C5a-mediated pain hypersensitivity. *J Neurosci* 2007; 27: 8699–8708.
126. Kidd BL, Cruwys S, Mapp PI and Blake DR. Role of the sympathetic nervous system in chronic joint pain and inflammation. *Ann Rheum Dis* 1992; 51: 1188–1191.
127. Levine JD, Fye K, Heller P, Basbaum AI and Whiting-O'Keefe Q. Clinical response to regional intravenous guanethidine in patients with rheumatoid arthritis. *J Rheumatol* 1986; 13: 1040–1043.
128. Pongratz G and Straub RH. B-cell involvement in the pathogenesis of RA-is there a contribution of the sympathetic nervous system? *Immunol Res* 2008; 40: 148–163.
129. Pongratz G and Straub RH. Role of peripheral nerve fibres in acute and chronic inflammation in arthritis. *Nat Rev Rheumatol* 2013; 9: 117–126.
130. Sanders VM. The beta2-adrenergic receptor on T and B lymphocytes: do we understand it yet? *Brain Behav Immun* 2012; 26: 195–200.
131. Kingery WS, Offley SC, Guo TZ, Davies MF, Clark JD and Jacobs CR. A substance P receptor (NK1) antagonist enhances the widespread osteoporotic effects of sciatic nerve section. *Bone* 2003; 33: 927–936.