



# Article Magnetometer-Guided Sentinel Lymph Node Dissection in Prostate Cancer: Rate of Lymph Node Involvement Compared with Radioisotope Marking

Svenja Engels<sup>†</sup>, Bianca Michalik<sup>†</sup>, Luca-Marie Meyer, Lena Nemitz, Friedhelm Wawroschek and Alexander Winter \*<sup>®</sup>

- University Hospital for Urology, Klinikum Oldenburg, Department of Human Medicine, School of Medicine and Health Sciences, Carl von Ossietzky University Oldenburg, 26122 Oldenburg, Germany; engels.svenja@klinikum-oldenburg.de (S.E.); michalik.bianca@klinikum-oldenburg.de (B.M.); luca-marie.meyer@uol.de (L.-M.M.); nemitz.lena@klinikum-oldenburg.de (L.N.); wawroschek.friedhelm@klinikum-oldenburg.de (F.W.)
- \* Correspondence: winter.alexander@klinikum-oldenburg.de

+ These authors contributed equally to this work.



Citation: Engels, S.; Michalik, B.; Meyer, L.-M.; Nemitz, L.; Wawroschek, F.; Winter, A. Magnetometer-Guided Sentinel Lymph Node Dissection in Prostate Cancer: Rate of Lymph Node Involvement Compared with Radioisotope Marking. *Cancers* 2021, 13, 5821. https://doi.org/ 10.3390/cancers13225821

Academic Editors: Moriaki Kusakabe and Akihiro Kuwahata

Received: 21 September 2021 Accepted: 17 November 2021 Published: 20 November 2021

**Publisher's Note:** MDPI stays neutral with regard to jurisdictional claims in published maps and institutional affiliations.



**Copyright:** © 2021 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (https:// creativecommons.org/licenses/by/ 4.0/). **Simple Summary:** Pelvic lymph node dissection is recommended in prostate cancer according to the patients' individual risk for nodal metastases. Targeted removal of sentinel lymph nodes increases the number of detected lymph node metastases in patients with prostate cancer. We previously established magnetometer-guided sentinel lymph node dissection in patients with prostate cancer to overcome logistical and technical disadvantages associated with the standard radioisotope-guided technique. This retrospective study compared the magnetometer-guided and standard techniques in terms of their ability to detect lymph node metastases. Using the magnetometer-guided technique, more sentinel lymph nodes were detected per patient. The detected rates of lymph node involvement matched the predictions in both techniques equally well. Our findings confirm the reliability of magnetometer-guided sentinel lymph node dissection and highlight the importance of the sentinel technique for detecting lymph node metastases in prostate cancer.

Abstract: Sentinel pelvic lymph node dissection (sPLND) enables the targeted removal of lymph nodes (LNs) bearing the highest metastasis risk. In prostate cancer (PCa), sPLND alone or combined with extended PLND (ePLND) reveals more LN metastases along with detecting sentinel LNs (SLNs) outside the conventional ePLND template. To overcome the disadvantages of radioisotope-guided sPLND in PCa treatment, magnetometer-guided sPLND applying superparamagnetic iron oxide nanoparticles as a tracer was established. This retrospective study compared the nodal staging ability between magnetometer- and radioisotope-guided sPLNDs. We analyzed data of PCa patients undergoing radical prostatectomy and magnetometer- (848 patients, 2015-2021) or radioisotopeguided (2092 patients, 2006–2015) sPLND. To reduce heterogeneity among cohorts, we performed propensity score matching and compared data considering sentinel nomogram-based probabilities for LN involvement (LNI). Magnetometer- and radioisotope-guided sPLNDs had SLN detection rates of 98.12% and 98.09%, respectively; the former detected more SLNs per patient. The LNI rates matched nomogram-based predictions in both techniques equally well. Approximately 7% of LN metastases were detected outside the conventional ePLND template. Thus, we confirmed the reliability of magnetometer-guided sPLND in nodal staging, with results comparable with or better than radioisotope-guided sPLND. Our findings highlight the importance of the sentinel technique for detecting LN metastases in PCa.

**Keywords:** prostate cancer; sentinel lymph node; lymphadenectomy; metastases; superparamagnetic iron oxide nanoparticles; radioisotopes

#### 1. Introduction

Pelvic lymph node dissection (PLND) is the most reliable technique for lymph node (LN) staging in clinically localized prostate cancer (PCa) [1]. LN status is a therapeutically crucial prognostic factor in PCa because the presence and extent of LN involvement (LNI) are related to an increased risk of systemic dissemination and progression of the disease [2–4]. Moreover, PLND or resection of LN metastases has been indicated to have therapeutic benefits, particularly in patients with minimal LNI [1,5–8].

The detection of LNI directly correlates with the number of dissected LNs as well as with the anatomical limits of PLND [9,10]. The European Association of Urology (EAU) guidelines, therefore, recommend an extended PLND (ePLND) for LN staging in patients with >5% risk of LNI as diagnosed by systematic random biopsy [11,12] or in those with >7% risk of LNI as diagnosed by multiparametric magnetic resonance imaging (MRI) and MRI-targeted biopsy [12,13].

The complication rate, however, also increases alongside the increase in the number of LNs removed [14–16]. Therefore, Wawroschek et al. [17] adopted the techniques and concepts of radioisotope-guided sentinel LN (SLN) identification from other tumor entities for use in PCa, and this has subsequently been independently confirmed in several studies [18,19]. Sentinel PLND (sPLND) enables the targeted removal of clinically negative LNs, which bear a high probability of containing metastases [20] because SLNs are the first lymphatic drainage stations of their primary organs or the respective tumor [21]. In PCa, sPLND alone or in combination with ePLND increases the number of detected LN metastases [19,22,23]. Moreover, during sPLND, SLNs occurring outside the conventional ePLND template can be removed [22–24]. Thus, sPLND provides additional diagnostic value by adjusting the degree and anatomical extent of PLND to the patient's individual lymphatic drainage situation [19,22,23,25].

Unfortunately, the use of radioisotope tracers for SLN marking is associated with several technical and logistical disadvantages [26,27]. For example, the practicality of the radioisotope-guided sPLND technique depends on the accessibility to radioisotope tracers and nuclear medicine facilities. Thus, this technique is used only in more developed countries or hospitals with access to such technology. Furthermore, this technique exposes patients and surgical staff to radiation, an aspect that is strongly controlled by legislation. In patients with breast cancer, superparamagnetic iron oxide nanoparticles (SPIONs) have been successfully developed as an equivalent, easy-to-use, and radiation-free alternative for SLN marking and intraoperative detection [26,28]. Our group subsequently adopted this technique of intraoperative magnetic detection of SLNs for use in patients with PCa; we use a system that comprises a magnetic tracer and a handheld magnetometer [23,24,27,29].

This retrospective study primarily aimed to compare magnetometer- and radioisotopeguided sPLND techniques in terms of their ability to detect LN metastases in patients with PCa. The secondary aim was to evaluate the anatomical distribution of dissected SLNs and detected LN metastases. We analyzed two large data sets from patients with PCa who underwent radical prostatectomy in combination with either of the two sPLND techniques at a tertiary referral hospital and performed a matched-pair analysis. The rates of LNI were compared between the two techniques while considering the patients' individual probabilities for LNI as inferred from our sentinel nomogram [30].

## 2. Materials and Methods

## 2.1. Patient Populations

This retrospective study considered two cohorts of patients with PCa consecutively documented in the database of the University Hospital for Urology Oldenburg. We initially enrolled 2186 patients with PCa who underwent open retropubic radical prostatectomy combined with radioisotope-guided sPLND between January 2006 and February 2015. Then, we excluded the data of 11 patients who received only one-sided sPLND and of one patient in whom the time between tracer injection and surgery extended the manufacturer-guaranteed tracer detectability period. Furthermore, 45 patients who underwent hormonal

treatment and 34 who underwent transurethral prostate surgeries before prostatectomy were also excluded. LN dissection data were incomplete in three additional patients. The final sample for the analysis of radioisotope-guided sPLND included 2092 patients.

Initially, we included 881 patients with PCa who underwent open retropubic radical prostatectomy combined with magnetometer-guided sPLND between February 2015 and May 2021. Then, we excluded the data of seven patients who received only onesided sPLND, three patients with limited tracer detectability because of metal implants, and two patients in whom the time between tracer injection and surgery extended the manufacturer-guaranteed detectability period. Furthermore, we also excluded 16 patients who underwent hormonal treatment and 4 who underwent transurethral prostate surgeries prior to prostatectomy. The LN dissection data were incomplete in one additional patient. The final sample for the analysis of magnetometer-guided sPLND included 848 patients.

All patients were informed verbally and in writing about the open retropubic radical prostatectomy and sPLND; all signed a consent form before surgery.

#### 2.2. sPLND Technique and Histopathological Examination

All patients were administered with transrectal tracer injection, either 99mTechnetium nanocolloid (160 MBq Nanocoll®, Nycomed Amersham Sorin, Milan, Italy) or superparamagnetic iron oxide nanoparticles (02/2015–01/2019 Sienna+®, Sysmex Europe GmbH, Norderstedt, Germany; 01/2019–05/2021 Magtrace<sup>®</sup>, Sysmex Europe GmbH, Norderstedt, Germany, into the prostate under ultrasonic guidance a day before surgery. SPLND was performed as described by Wawroschek et al. [31] and Winter et al. [32] for radioisotopeguided surgery and as described by Winter et al. [23] for magnetometer-guided surgery. All SLNs detected by a gamma probe (C-Trak System, Care Wise, Morgan Hill, CA, USA, or Crystal Probe SG04, Crystal Photonics GmbH, Berlin, Germany) or a magnetometer (Sentimag<sup>®</sup>, Sysmex Europe GmbH, Norderstedt, Germany) together with lymphatic fatty tissue directly adjoining or adhering to the identified SLNs were removed surgically. After sentinel-guided surgery, risk-adapted or if no SLN was detected at all, PLND was completed by ePLND using the anatomic template as described by Weingärtner et al. [33], which included all lymphatic fatty tissue along the external and internal iliac vessels and the obturator fossa as well as within the area dorsal to the obturator nerve, from the bifurcation of the common iliac artery (proximal limit) to the femoral canal (distal limit) and from the pelvic sidewall (lateral limit) to the perivesical fatty tissue (medial limit).

After surgery, LNs were cut into 3 mm transverse sections, routinely processed, and embedded into paraffin. Then, 4–5  $\mu$ m sections were stained with hematoxylineosin (HE; Figure 1). In rare cases of inconclusive conventional histology, samples were immunohistochemically stained with AE1/AE3 pancytokeratin antibodies to check for (micro-)metastases.

#### 2.3. Data Analyses

All data analyses were performed using R 4.1.0 [34]. For each sPLND technique, we calculated the median numbers of SLNs detected per patient, SLN detection rate (proportion of patients with detected SLNs), rate of LNI (proportion of patients with pathologic nodal stage 1; pN1), and the false-negative rate (proportion of LN-positive but SLN-negative cases). Numbers and proportions were compared statistically using the Wilcoxon rank-sum test and  $\chi^2$  proportions test, respectively. Tests were repeated after 1:1 optimal pair matching according to clinical information (i.e., age, prostate-specific antigen, clinical tumor stage, Gleason score, and percentage of positive biopsy cores) based on the results of a propensity score analysis. A summary of the propensity score-adjusted data is presented in Table A1 of Appendix A. We estimated the probability of LNI for each patient according to our nomogram [30]. This nomogram predicts a patient's individual probability for the presence of lymphogenic metastases based on clinical information such as prostate-specific antigen (PSA) value, clinical tumor stage, biopsy Gleason score, and percentage of positive biopsy Cores [30]. In our data set, clinical tumor stage and/or biopsy

data (biopsy Gleason score and/or the percentage of tumor-positive biopsy cores) were unavailable for 24 patients. To evaluate the diagnostic accuracy of each sPLND technique, we plotted the observed LNI rates against the nomogram-predicted probabilities of LNI in steps of approximately 3.33% (30 bins). Curves were smoothed, and 95% confidence intervals were calculated using the Loess method (local polynomial regression fitting). A  $\chi^2$  goodness-of-fit test was computed for each technique to estimate the deviation of the curves from the ideal curve wherein observed, and predicted rates of LNI match perfectly. Moreover, the numbers of dissected SLNs and LN metastases were counted for each anatomical region of sPLND. The region data were unavailable for 22 SLNs detected during magnetometer-guided sPLND.

### 3. Results

In total, we dissected 12,331 LNs in 848 patients who underwent magnetometer-guided sPLND and 22,565 LNs in 2092 patients who underwent radioisotope-guided sPLND. Among these dissected LNs, 6478 and 12,981 were SLNs. Table 1 summarizes clinical and histopathological tumor characteristics as well as LN dissection data of the two patient cohorts. A summary of these data after propensity score matching can be found in Table A1. The SLN detection rates did not differ between the magnetometer- and radioisotope-guided sPLND techniques (98.11%, n = 832 patients vs. 98.18%, n = 2054 patients, respectively; Table 2). However, we detected significantly more SLNs per patient using magnetometer-guided sPLND (median = 7, IQR: 4–10) than that using radioisotope-guided sPLND (median = 6, IQR: 4–8; Table 2).

Mathad	Magnetometer-Guided sPLND			Radioisotope-Guided sPLND		
Method	Overall	pN0	pN1	Overall	pN0	pN1
n (%)	848	655 (77.24)	193 (22.76)	2092	1696 (81.07)	396 (18.93)
Age (IQR)	67 (62–71)	67 (61–71)	68 (64–73)	67 (62–71)	67 (61–71)	68 (63–71)
Total PSA ng/mL (IQR)	8.7 (6.1–13.5)	8 (5.8–11.8)	12.8 (8.6–27.7)	7.8 (5.5–12.5)	7.2 (5.3–10.9)	12.0 (7.9–20.6)
Dissected LNs (IQR)	14 (10–18)	13 (10–17)	16 (12–21)	10 (7–14)	10 (7–13)	12 (9–15)
Positive LNs (IQR)	0 (-)	0 (-)	2 (1-4)	0 (-)	0 (-)	2 (1-3)
Dissected SLNs (IQR)	7 (4–10)	7 (5–10)	6 (4–10)	6 (4–8)	6 (4–8)	6 (3–8)
Positive SLNs (IQR)	0 (-)	0 (-)	1 (1-2)	0 (-)	0 (-)	1 (1-2)
Clinical tumor stage (%)	*		*	**	**	**
cT1	436 (51.42)	397 (60.61)	39 (20.21)	1129 (53.97)	1027 (60.55)	102 (25.76)
cT2	368 (43.40)	244 (37.25)	124 (64.25)	919 (43.93)	658 (38.80)	261 (65.91)
cT3	41 (4.83)	14 (2.14)	27 (13.99)	36 (1.72)	6 (0.35)	30 (7.58)
cT4	2 (0.24)	0	2 (1.04)	2 (0.10)	0	2 (0.51)
Biopsy Gleason sum (%)				***	***	
$\leq 6$	162 (19.10)	150 (22.90)	12 (6.22)	998 (47.71)	938 (55.31)	60 (15.15)
=7 (3 + 4)	402 (47.41)	354 (54.05)	48 (24.87)	724 (34.61)	570 (33.61)	154 (38.89)
=7 (4 + 3)	129 (15.21)	88 (13.44)	41 (21.24)	191 (9.13)	109 (6.43)	82 (20.71)
$\geq 8$	155 (18.28)	63 (9.62)	92 (47.67)	176 (8.41)	76 (4.48)	100 (25.25)
Postoperative Gleason						
sum (%)						
$\leq 6$	30 (3.54)	30 (4.58)	0	349 (16.68)	348 (20.52)	1 (0.25)
=7(3+4)	443 (52.24)	423 (64.58)	20 (10.36)	1122 (53.63)	1052 (62.03)	70 (17.68)
=7(4+3)	216 (25.47)	147 (22.44)	69 (35.75)	420 (20.08)	230 (13.56)	190 (47.98)
$\geq 8$	159 (18.75)	55 (8.40)	104 (53.89)	201 (9.61)	66 (3.89)	135 (34.09)

Table 1. Patient characteristics.

M. d 1	Magnetometer-Guided sPLND			Radioisotope-Guided sPLND		
Wiethod	Overall	pN0	pN1	Overall	pN0	pN1
Pathologic tumor stage (%)						
pT1c	2 (0.24)	2 (0.31)	0	1 (0.05)	1 (0.06)	0
pT2a	41 (4.83)	41 (6.26)	0	184 (8.80)	180 (10.61)	4 (1.01)
pT2b	21 (2.48)	19 (2.90)	2 (1.04)	40 (1.91)	39 (2.30)	1 (0.25)
pT2c	399 (47.05)	390 (59.54)	9 (4.66)	1086 (51.91)	1048 (61.79)	38 (9.60)
pT3a	178 (20.99)	141 (21.53)	37 (19.17)	407 (19.46)	300 (17.69)	107 (27.02)
pT3b	197 (23.23)	61 (9.31)	136 (70.47)	318 (15.20)	113 (6.66)	205 (51.77)
pT4	10 (1.18)	1 (0.15)	9 (4.66)	56 (2.68)	15 (0.88)	41 (10.35)

Table 1. Cont.

Data are presented as median (interquartile range) or frequency (percentage). sPLND: sentinel pelvic lymph node dissection; pN: pathologic nodal stage; IQR: interquartile range; (S)LN: (sentinel) lymph node; PSA: prostate-specific antigen; \* clinical T-category could not be assessed in one patient (pN1); \*\* clinical T-category could not be assessed in six patients (pN0: n = 5, pN1: n = 1); \*\*\* incomplete biopsy data in three patients (pN0).

**Table 2.** Comparison between magnetometer-guided sPLND and radioisotope-guided sPLND either with original data or with propensity score-adjusted data.

Comparison	<b>Original Results</b>	Test Statistic	Adjusted Results	<b>Test Statistic</b>
SLN detection rate	98.11% (832) vs. 98.18% (2054)	$\chi^2 < 0.001$ , df = 1, p = 1	98.11% (832) vs. 95.87% (812)	$\chi^2 = 6.55, df = 1,$ p = 0.011 *
Number of dissected SLNs	7 (4–10) vs. 6 (4–8)	W = 1,059,411, p < 0.001 ****	7 (4–10) vs. 5 (3–7)	W = 471,031, p < 0.001 ***
Rate of LNI	22.76% (193) vs. 18.93% (396)	$\chi^2 = 5.29, df = 1,$ p = 0.021 *	22.76% (192) vs. 25.97% (220)	$\chi^2 = 2.34, df = 1,$ p = 0.126
Rate of LN+ but SLN-	7.25% (14) vs. 9.85% (39)	$\chi^2 = 0.77, df = 1,$ p = 0.379	7.29% (14) vs. 14.55% (32)	$\chi^2 = 4.73, df = 1,$ p = 0.030 *
False-negative rate	3.63% (7) vs. 5.05% (20)	$\chi^2 = 0.32, df = 1,$ p = 0.572	3.65% (7) vs. 5.91% (13)	$\chi^2 = 0.700, df = 1,$ p = 0.403

Data are presented as percentage (n) or median (interquartile range). (S)LN: (sentinel) lymph node; LNI: lymph node involvement; LN+: lymph node positivity; SLN-: sentinel lymph node negativity. \* 5% significance level, \*\*\* 0.1% significance level.

We found metastases in 621 LNs of 193 patients who underwent magnetometerguided sPLND and in 1010 LNs of 396 patients who underwent radioisotope-guided sPLND (Table 1). Figure 1 shows a representative example of a HE staining of an SLN metastasis as revealed by the magnetic tracer. On the one hand, there were significantly more patients with LN positivity among those who underwent magnetometer-guided sPLND than among those who underwent radioisotope-guided sPLND (22.76% vs. 18.93%, respectively; Table 2), which leveled off after propensity score matching (Table 2). On the other hand, no difference was noted between the two techniques in the proportion of patients who had metastases only in non-SLNs (7.25%, n = 14 vs. 9.85%, n = 39, respectively; Table 2). Of these, in five patients who underwent magnetometer-guided sPLND and in 15 patients who underwent radioisotope-guided sPLND, respectively, no SLNs could be detected at all, and in two and four patients, respectively, macroscopically visible metastases were surgically removed without measuring tracer activity. Excluding these cases from the sample of only patients with non-SLN positivity, the resulting false-negative rates were 3.63% (*n* = 7) for magnetometer-guided sPLND and 5.05% (*n* = 20) for radioisotope-guided sPLND (Table 2).



**Figure 1.** Hematoxylin-eosin staining of a magnetically traced sentinel lymph node. In the center of the image, a 4 mm metastasis of a Gleason 8 adenocarcinoma of the prostate is present. Note the brownish discoloration (arrows) of macrophages containing the magnetic tracer (superparamagnetic iron oxide nanoparticles). Total magnification  $40 \times .$ 

The observed proportions of patients with LN positivity did not differ from the proportions predicted by the nomogram in both magnetometer- (goodness-of-fit test:  $\chi^2 = 0.73$ , df = 29, *p* = 1) and radioisotope-guided (goodness-of-fit test:  $\chi^2 = 0.75$ , df = 29, *p* = 1; Figure 2) sPLND techniques.



sPLND technique: - magnetometer-guided - radioisotope-guided

**Figure 2.** Observed proportion of patients with lymph node (LN) positivity in relation to the probability of LN involvement as predicted by the nomogram for magnetometer (blue)- and radioisotope (red)-guided sentinel pelvic lymph node dissection (sPLND). Gray shaded areas represent the 95% confidence intervals of the smoothed blue and red curves, respectively. The black dot-dashed line represents the ideal curve wherein the predicted probabilities and observed proportions match perfectly.

Anatomic regions outside the standard template for ePLND accounted for 4.5% of SLNs detected by a magnetometer and 2.9% of SLNs detected by a radioprobe, respectively (Figure 3a). Accordingly, approximately 7.6% and 5.7% of LN metastases detected by magnetometer-guided and radioisotope-guided sPLND, respectively, occurred in LNs outside the standard ePLND template (Figure 3b).



**Figure 3.** Anatomical distribution of (**a**) the dissected sentinel lymph nodes (SLNs) and (**b**) the lymph node (LN) metastases detected using magnetometer (light gray bars)- and radioisotope (dark gray bars)-guided sentinel pelvic lymph node dissection (sPLND) techniques, respectively. Data of the anatomic region were not available for 22 SLNs dissected using magnetometer-guided sPLND.

#### 4. Discussion

In this retrospective study of pelvic sentinel lymphadenectomy in patients with PCa, SLN detection rates were equally high for both magnetometer- and radioisotope-guided sPLND techniques; however, the former revealed a greater number of SLNs per patient. The accordance between the observed rates of LNI and nomogram-based predictions was equally high in both sPLND techniques. Both sPLND techniques revealed a considerable proportion of lymphogenic metastases outside the conventional ePLND template.

The SLN detection rates found in our study closely match those reported in other studies on sPLND in PCa [19,22,35]. Our observed rates of LNI were accordingly high [19,32,36] and even higher than expected from the ePLND data [37–40]. We resect a comparatively large number of SLN with both procedures. This can probably be explained by the fact that we do not perform dynamic imaging in clinical routine and remove all active LNs regardless of their activity level. Therefore, strictly speaking, our procedure could rather be called "lymphatic mapping".

To predict the individual probability of LNI in patients with PCa, Winter et al. [30] developed a nomogram, which is based on clinical information and radioisotope-guided sPLND data. This nomogram was subsequently validated externally [41]. Our analyses

revealed high accordance between nomogram-based predictions and LNI rates in both sPLND techniques. These results suggest that magnetometer-guided sPLND is a reliable and promising alternative that can be used in PCa to overcome the disadvantages of radioisotope marking [26,27], as observed in other tumor entities [26,28,42–45]. Furthermore, an improved flow of the magnetic tracer may lead to an increased number of detected SLNs in magnetometer-guided sPLND, as observed in the present study. The size of the magnetic tracer particles is approximately 60 nm, which is slightly smaller and more homogeneous than the radioisotope tracer [26]. The radioisotope tracer particles were 80 nm (95%), 80–100 nm (4%), and >100 nm (1%) [27]. Thus, the particle size might have influenced the drain of the two different tracers through the lymphatic pathways and/or trapping in the LNs, leading to more SLNs being detected using magnetometer-guided sPLND.

At our hospital, the magnetometer-guided sPLND has largely replaced radioisotopeguided sPLND, which is an obvious limitation of our retrospective study design as the two techniques were used in different study periods. Consequently, the change in surgeons, as well as their increasing experience over time, could be the reason why more SLNs per patient were detected when using the magnetometer technique. Routine histopathological examination of the dissected LNs and the distinct use of supplementary immunohistochemistry have also improved over the years at our hospital. Thus, the higher prevalence of LNI found in patients being treated with magnetometer-guided sPLND could have partly resulted from these developments. Furthermore, the advancements in PCa diagnostics as well as treatment options for low risk PCa (e.g., active surveillance) have led to updates in guidelines (e.g., EAU) for PCa over time with implications for clinical practice and a stage shift [46]. Therefore, it is unsurprising that the two patient cohorts in our retrospective study also differed in their clinical tumor properties. For example, the patient group treated with magnetometer-guided sPLND had more advanced tumors according to biopsy Gleason score (Table 1). As reflected in our results, the Gleason score may serve as a predictor for LNI [30,39], and this might partly explain the differences in the rates of LNI between the two sPLND techniques. In agreement with this, the observed differences in the rates of LNI are no longer present after propensity score analysis through optimal pair matching of the data according to clinical information. Nevertheless, the rates of LNI in both sPLND techniques accurately match the rates predicted by our sentinel-based nomogram [30]. Thus, our results show that both magnetometer- and radioisotope-guided sPLND techniques are reliable tools for LN staging in PCa. Since the performance of the magnetic method seems to be even slightly better and is also associated with a simpler clinical workflow, we will continue to use it to replace the radioactive sentinel procedure if there are no contraindications.

The magnetic SLN marking technique has some limitations [23,24,29]. The magnetic approach is not applicable in patients with pacemakers or other implanted electronic devices in the chest wall as well as those with hypersensitivity to iron or with iron overload disease. Furthermore, the detectability of the magnetic tracer is reduced in patients with metal implants such as hip prostheses or other metallic pelvic implants. In these cases, patients would still benefit from radioisotope-guided sPLND.

The sentinel approach bears one significant drawback. When LNs are fully metastasized, or lymphatic pathways are blocked, the afferent lymph could be directed to other LNs, which might not necessarily be SLNs [47,48]; this has been observed, for example, in inguinal SLNs in penile carcinoma [49,50]. Performing sPLND alone in these cases would yield false-negative results. The false-negative rates observed in our study were as low as those reported in other studies [19,22,35,48]. Thus, the diagnostic value of sPLND for LN staging in patients with PCa is similar to that when using a combination of sPLND and ePLND; this would have a higher prevalence of LNI than using ePLND alone [19,22,32,41].

In addition to the selective removal of LNs bearing the highest risk of containing metastases, the sentinel approach also enables surgical treatment, which is adjusted to the patient's individual lymphatic drainage situation [19,25,48]. Consequently, we identified a considerable proportion of LN metastases outside the conventional ePLND tem-

plate [33] using both sPLND techniques. These findings are well in line with those of other studies and highlight the diagnostic value of sPLND for LN staging in patients with PCa [19,22,25,36,51].

## 5. Conclusions

Our study confirms the diagnostic accuracy of magnetometer-guided sPLND in nodal staging in patients with PCa. We suggest magnetometer-guided sPLND as a reliable and promising alternative sPLND technique in PCa treatment to overcome the technical and logistic disadvantages of radioisotope marking. Furthermore, our results highlight the additional diagnostic value of the sentinel technique in PCa because it allows not only the selective removal of LNs bearing the highest risk of containing metastases but also surgical treatment that is adapted to the patient's individual lymphatic drainage situation.

**Author Contributions:** Conceptualization, A.W. and S.E.; methodology, S.E., B.M., and A.W.; validation, S.E. and B.M.; formal analysis, B.M.; resources, F.W.; data curation, S.E., L.-M.M., A.W., and L.N.; writing—original draft preparation, S.E. and B.M.; writing—review and editing, A.W., S.E., B.M., L.-M.M., L.N., and F.W.; visualization, B.M.; supervision, A.W. and F.W.; project administration, S.E. and A.W.; funding acquisition, A.W. and F.W. All authors have read and approved the final version of the manuscript.

Funding: This research received no external funding.

**Institutional Review Board Statement:** This retrospective study was conducted in accordance with the Declaration of Helsinki and received ethical approval from the Medical Ethics Committee of the University of Oldenburg (02/06/2021, reference: 2018-140). The study is registered in an international clinical trials register (Research Registry, researchregistry7004).

**Informed Consent Statement:** All patients provided their written informed consent for the documentation and scientific evaluation of their data.

**Data Availability Statement:** The data presented in this study are available upon reasonable request from the corresponding author.

**Conflicts of Interest:** The authors declare no conflict of interest. The funders had no role in the design of the study; collection, analyses, or interpretation of data; writing of the manuscript; or decision to publish the results.

#### Appendix A

Magnetometer-Guided sPLND			Radioisotope-Guided sPLND		
Overall	pN0	pN1	Overall	pN0	pN1
847 *	655 (77.33)	192 (22.67)	847	627 (74.03)	220 (25.97)
67 (62–71)	67 (61–71)	68 (64–73)	67 (62–71)	67 (62–71)	68 (63–72)
8.7 (6.1–13.4)	8.0 (5.8–11.8)	12.7 (8.6–27.3)	8.6 (6.0–14.3)	7.8 (5.6–12.4)	12.2 (8.1–20.2)
14 (10–18)	13 (10–17)	16 (12–20)	10 (8–14)	10 (7–13)	12 (9–15)
0 (-)	0 (-)	2 (1–4)	0 (0–1)	0 (-)	2 (1–3)
7 (4–10)	7 (5–10)	6 (4–10)	5 (3–7)	5 (3–7)	5 (3–8)
0 (-)	0 (-)	1 (1–2)	0 (-)	0 (-)	1 (1–2)
436 (51.48)	397 (60.61)	39 (20.31)	464 (54.78)	401 (63.96)	63 (28.64)
368 (43.45)	244 (37.25)	124 (64.58)	357 (42.15)	222 (35.41)	135 (61.36)
41 (4.84)	14 (2.14)	27 (14.06)	24 (2.83)	4 (0.64)	20 (9.09)
2 (0.24)	0	2 (1.04)	2 (0.24)	0	2 (0.91)
	Magne   Overall   847 *   67 (62–71)   8.7 (6.1–13.4)   14 (10–18)   0 (-)   7 (4–10)   0 (-)   436 (51.48)   368 (43.45)   41 (4.84)   2 (0.24)	Magnetometer-Guided   Overall pN0   847 * 655 (77.33)   67 (62–71) 67 (61–71)   8.7 (6.1–13.4) 8.0 (5.8–11.8)   14 (10–18) 13 (10–17)   0 (-) 0 (-)   7 (4–10) 7 (5–10)   0 (-) 0 (-)   436 (51.48) 397 (60.61)   368 (43.45) 244 (37.25)   41 (4.84) 14 (2.14)   2 (0.24) 0	$\begin{tabular}{ c c c } \hline Magnetometer-Guided $PLND$ \\ \hline Overall $pN0$ $pN1$ \\ \hline 847 * $655 (77.33)$ $192 (22.67)$ \\ \hline 67 (62-71)$ $67 (61-71)$ $68 (64-73)$ \\ \hline 67 (62-71)$ $67 (61-71)$ $68 (64-73)$ \\ \hline 8.7 (6.1-13.4)$ $8.0 (5.8-11.8)$ $12.7 (8.6-27.3)$ \\ \hline 14 (10-18)$ $13 (10-17)$ $16 (12-20)$ \\ \hline 0 (-)$ $0 (-)$ $2 (1-4)$ \\ \hline 7 (4-10)$ $7 (5-10)$ $6 (4-10)$ \\ \hline 0 (-)$ $0 (-)$ $1 (1-2)$ \\ \hline 436 (51.48)$ $397 (60.61)$ $39 (20.31)$ \\ \hline 368 (43.45)$ $244 (37.25)$ $124 (64.58)$ \\ \hline 41 (4.84)$ $14 (2.14)$ $27 (14.06)$ \\ \hline 2 (0.24)$ $0$ $2 (1.04)$ \\ \hline \end{tabular}$	$\begin{tabular}{ c c c c c } \hline Magnetometer-Guided $PLND & Radio \\ \hline \hline Overall & pN0 & pN1 & Overall \\ \hline 847 * & 655 (77.33) & 192 (22.67) & 847 \\ \hline 67 (62-71) & 67 (61-71) & 68 (64-73) & 67 (62-71) \\ \hline 8.7 (6.1-13.4) & 8.0 (5.8-11.8) & 12.7 (8.6-27.3) & 8.6 (6.0-14.3) \\ \hline 14 (10-18) & 13 (10-17) & 16 (12-20) & 10 (8-14) \\ \hline 0 (-) & 0 (-) & 2 (1-4) & 0 (0-1) \\ \hline 7 (4-10) & 7 (5-10) & 6 (4-10) & 5 (3-7) \\ \hline 0 (-) & 0 (-) & 1 (1-2) & 0 (-) \\ \hline 436 (51.48) & 397 (60.61) & 39 (20.31) & 464 (54.78) \\ \hline 368 (43.45) & 244 (37.25) & 124 (64.58) & 357 (42.15) \\ \hline 41 (4.84) & 14 (2.14) & 27 (14.06) & 24 (2.83) \\ \hline 2 (0.24) & 0 & 2 (1.04) & 2 (0.24) \\ \hline \end{tabular}$	$\begin{tabular}{ c c c c c c } \hline Magnetometer-Guided $$ $$ $$ $$ $$ $$ $$ $$ $$ $$ $$ $$ $$$

**Table A1.** Patient characteristics after 1:1 optimal pair matching according to clinical information (i.e., age, prostate-specific antigen, clinical tumor stage, Gleason score, and percentage of positive biopsy cores) and adjusted to propensity scores.

Mathad	Magnetometer-Guided sPLND			Radioisotope-Guided sPLND		
Wiethod -	Overall	pN0	pN1	Overall	pN0	pN1
Biopsy Gleason sum (%)						
$\leq 6$	162 (19.13)	150 (22.90)	12 (6.25)	162 (19.13)	153 (24.40)	9 (4.09)
=7 (3 + 4)	401 (47.34)	354 (54.05)	47 (24.48)	422 (49.82)	349 (55.66)	73 (33.18)
=7(4+3)	129 (15.23)	88 (13.44)	41 (21.35)	115 (13.58)	64 (10.21)	51 (23.18)
$\geq 8$	155 (18.30)	63 (9.62)	92 (47.92)	148 (17.47)	61 (9.73)	87 (39.55)
Postoperative Gleason						
sum (%)						
$\leq 6$	30 (3.54)	30 (4.58)	0	58 (6.85)	58 (9.25)	0
=7 (3 + 4)	443 (52.30)	423 (64.58)	20 (10.42)	438 (51.71)	406 (64.75)	32 (14.55)
=7(4+3)	215 (25.38)	147 (22.44)	68 (35.42)	204 (24.09)	111 (17.7)	93 (42.27)
$\geq 8$	159 (18.77)	55 (8.4)	104 (54.17)	147 (17.36)	52 (8.29)	95 (43.18)
Pathologic tumor stage (%)						
pT1c	2 (0.24)	2 (0.31)	0	1 (0.12)	1 (0.16)	0
pT2a	41 (4.84)	41 (6.26)	0	64 (7.56)	61 (9.73)	3 (1.36)
pT2b	21 (2.48)	19 (2.90)	2 (1.04)	8 (0.94)	8 (1.28)	0
pT2c	399 (47.11)	390 (59.54)	9 (4.69)	378 (44.63)	362 (57.74)	16 (7.27)
pT3a	177 (20.90)	141 (21.53)	36 (18.75)	180 (21.25)	127 (20.26)	53 (24.09)
pT3b	197 (23.26)	61 (9.31)	136 (70.83)	188 (22.20)	62 (9.89)	126 (57.27)
pT4	10 (1.18)	1 (0.15)	9 (4.69)	28 (3.31)	6 (0.96)	22 (10)

Data are presented as median (interquartile range) or frequency (percentage). sPLND: sentinel pelvic lymph node dissection; pN: pathologic nodal stage; IQR: interquartile range; (S)LN: (sentinel) lymph node; PSA: prostate-specific antigen; \* clinical T-category could not be assessed in one patient (pN1).

#### References

- 1. Withrow, D.R.; DeGroot, J.M.; Siemens, D.R.; Groome, P.A. Therapeutic value of lymph node dissection at radical prostatectomy: A population-based case-cohort study. *BJU Int.* **2011**, *108*, 209–216. [CrossRef] [PubMed]
- Cheng, L.; Bergstralh, E.J.; Cheville, J.C.; Slezak, J.; Corica, F.A.; Zincke, H.; Blute, M.L.; Bostwick, D.G. Cancer volume of lymph node metastasis predicts progression in prostate cancer. *Am. J. Surg. Pathol.* 1998, 22, 1491–1500. [CrossRef] [PubMed]
- 3. Cheng, L.; Zincke, H.; Blute, M.L.; Bergstralh, E.J.; Scherer, B.; Bostwick, D.G. Risk of prostate carcinoma death in patients with lymph node metastasis. *Cancer* 2001, *91*, 66–73. [CrossRef]
- Wawroschek, F.; Wagner, T.; Hamm, M.; Weckermann, D.; Vogt, H.; Märkl, B.; Gordijn, R.; Harzmann, R. The influence of serial sections, immunohistochemistry, and extension of pelvic lymph node dissection on the lymph node status in clinically localized prostate cancer. *Eur. Urol.* 2003, 43, 132–137. [CrossRef]
- Choo, M.S.; Kim, M.; Ku, J.H.; Kwak, C.; Kim, H.H.; Jeong, C.W. Extended versus standard pelvic lymph node dissection in radical prostatectomy on oncological and functional outcomes: A systematic review and meta-analysis. *Ann. Surg. Oncol.* 2017, 24, 2047–2054. [CrossRef]
- 6. Seiler, R.; Studer, U.E.; Tschan, K.; Bader, P.; Burkhard, F.C. Removal of limited nodal disease in patients undergoing radical prostatectomy: Long-term results confirm a chance for cure. *J. Urol.* **2014**, *191*, 1280–1285. [CrossRef]
- 7. Winter, A.; Henke, R.P.; Wawroschek, F. Targeted salvage lymphadenectomy in patients treated with radical prostatectomy with biochemical recurrence: Complete biochemical response without adjuvant therapy in patients with low volume lymph node recurrence over a long-term follow-up. *BMC Urol.* **2015**, *15*, 1–8. [CrossRef]
- 8. Preisser, F.; van den Bergh, R.C.N.; Gandaglia, G.; Ost, P.; Surcel, C.I.; Sooriakumaran, P.; Montorsi, F.; Graefen, M.; van der Poel, H.; de la Taille, A.; et al. Effect of extended pelvic lymph node dissection on oncologic outcomes in patients with D'Amico intermediate and high risk prostate cancer treated with radical prostatectomy: A multi-institutional study. *J. Urol.* **2020**, 203, 338–343. [CrossRef] [PubMed]
- 9. Bivalacqua, T.J.; Pierorazio, P.M.; Gorin, M.A.; Allaf, M.E.; Carter, H.B.; Walsh, P.C. Anatomic extent of pelvic lymph node dissection: Impact on long-term cancer-specific outcomes in men with positive lymph nodes at time of radical prostatectomy. *Urology* **2013**, *82*, 653–659. [CrossRef]
- 10. Briganti, A.; Blute, M.L.; Eastham, J.H.; Graefen, M.; Heidenreich, A.; Karnes, J.R.; Montorsi, F.; Studer, U.E. Pelvic lymph node dissection in prostate cancer. *Eur. Urol.* 2009, *55*, 1251–1265. [CrossRef]
- 11. Hinev, A.I.; Anakievski, D.; Kolev, N.H.; Hadjiev, V.I. Validation of nomograms predicting lymph node involvement in patients with prostate cancer undergoing extended pelvic lymph node dissection. *Urol. Int.* **2014**, *92*, 300–305. [CrossRef]

Table	A1.	Cont.
-------	-----	-------

- 12. Mottet, N.; Cornford, P.; van den Bergh, R.C.N.; Briers, E.; Expert Patient Advocate (European Prostate Cancer Coalition/Europa UOMO); De Santis, M.; Gillessen, S.; Grummet, J.; Henry, A.M.; van der Kwast, T.H.; et al. EAU-EANM-ESTRO-ESUR-ISUP-SIOG Guidelines on Prostate Cancer. Available online: https://uroweb.org/guideline/prostate-cancer/ (accessed on 6 September 2021).
- Gandaglia, G.; Ploussard, G.; Valerio, M.; Mattei, A.; Fiori, C.; Fossati, N.; Stabile, A.; Beauval, J.-B.; Malavaud, B.; Roumiguié, M.; et al. A novel nomogram to identify candidates for extended pelvic lymph node dissection among patients with clinically localized prostate cancer diagnosed with magnetic resonance imaging-targeted and systematic biopsies. *Eur. Urol.* 2019, 75, 506–514. [CrossRef]
- 14. Briganti, A.; Chun, F.K.H.; Salonia, A.; Suardi, N.; Gallina, A.; Da Pozzo, L.F.; Roscigno, M.; Zanni, G.; Valiquette, L.; Rigatti, P.; et al. Complications and other surgical outcomes associated with extended pelvic lymphadenectomy in men with localized prostate cancer. *Eur. Urol.* **2006**, *50*, 1006–1013. [CrossRef]
- 15. Musch, M.; Klevecka, V.; Roggenbuck, U.; Kroepfl, D. Complications of pelvic lymphadenectomy in 1,380 patients undergoing radical retropubic prostatectomy between 1993 and 2006. *J. Urol.* **2008**, *179*, 923–929. [CrossRef]
- Winter, A.; Vogt, C.; Weckermann, D.; Wawroschek, F. Complications of pelvic lymphadenectomy in clinically localised prostate cancer: Different techniques in comparison and dependency on the number of removed lymph nodes. *Aktuel. Urol.* 2011, 12, 179–183. [CrossRef] [PubMed]
- 17. Wawroschek, F.; Vogt, H.; Weckermann, D.; Wagner, T.; Harzmann, R. The sentinel lymph node concept in prostate cancer-first results of gamma probe-guided sentinel lymph node identification. *Eur. Urol.* **1999**, *36*, 595–600. [CrossRef] [PubMed]
- Fossati, N.; Willemse, P.-P.M.; Van den Broeck, T.; van den Bergh, R.C.N.; Yuan, C.Y.; Briers, E.; Bellmunt, J.; Bolla, M.; Cornford, P.; De Santis, M.; et al. The benefits and harms of different extents of lymph node dissection during radical prostatectomy for prostate cancer: A systematic review. *Eur. Urol.* 2017, *72*, 84–109. [CrossRef]
- 19. Wit, E.M.K.; Acar, C.; Grivas, N.; Yuan, C.; Horenblas, S.; Liedberg, F.; Valdes Olmos, R.A.; van Leeuwen, F.W.B.; van den Berg, N.S.; Winter, A.; et al. Sentinel node procedure in prostate cancer: A systematic review to assess diagnostic accuracy. *Eur. Urol.* **2017**, *71*, 596–605. [CrossRef]
- 20. Cabanas, R.M. An approach for the treatment of penile carcinoma. Cancer 1977, 39, 456–466. [CrossRef]
- 21. Gould, E.A.; Winship, T.; Philbin, P.H.; Kerr, H.H. Observations on a "sentinel node" in cancer of the parotid. *Cancer* **1960**, *13*, 77–78. [CrossRef]
- 22. Van der Poel, H.; Wit, E.; Acar, C.; van den Berg, N.S.; van Leeuwen, F.W.B.; Valdés Olmos, R.A.; Winter, A.; Wawroschek, F.; Liedberg, F.; Maclennan, S.; et al. Sentinel Node Prostate Cancer Consensus Panel Group members. Sentinel node biopsy for prostate cancer: Report from a consensus panel meeting. *BJU Int.* **2017**, *120*, 204–211. [CrossRef]
- Winter, A.; Engels, S.; Reinhardt, L.; Wasylow, C.; Gerullis, H.; Wawroschek, F. Magnetic marking and intraoperative detection of primary draining lymph nodes in high-risk prostate cancer using superparamagnetic iron oxide nanoparticles: Additional diagnostic value. *Molecules* 2017, 22, 2192. [CrossRef] [PubMed]
- Winter, A.; Kowald, T.; Paulo, T.S.; Goos, P.; Engels, S.; Gerullis, H.; Schiffmann, J.; Chavan, A.; Wawroschek, F. Magnetic resonance sentinel lymph node imaging and magnetometer-guided intraoperative detection in prostate cancer using superparamagnetic iron oxide nanoparticles. *Int. J. Nanomed.* 2018, 13, 6689–6698. [CrossRef]
- 25. Joniau, S.; Van den Bergh, L.; Lerut, E.; Deroose, C.M.; Haustermans, K.; Oyen, R.; Budiharto, T.; Ameye, F.; Bogaerts, K.; Van Poppel, H. Mapping of pelvic lymph node metastases in prostate cancer. *Eur. Urol.* **2013**, *63*, 450–458. [CrossRef]
- Douek, M.; Klaase, J.; Monypenny, I.; Kothari, A.; Zechmeister, K.; Brown, D.; Wyld, L.; Drew, P.; Garmo, H.; Agbaje, O.; et al. Sentinel node biopsy using a magnetic tracer versus standard technique: The SentiMAG multicentre trial. *Ann. Surg. Oncol.* 2014, 21, 1237–1245. [CrossRef]
- 27. Winter, A.; Woenkhaus, J.; Wawroschek, F. A novel method for intraoperative sentinel lymph node detection in prostate cancer patients using superparamagnetic iron oxide nanoparticles and a handheld magnetometer: The initial clinical experience. *Ann. Surg. Oncol.* **2014**, *21*, 4390–4396. [CrossRef] [PubMed]
- Thill, M.; Kurylcio, A.; Welter, R.; van Haasteren, V.; Grosse, B.; Berclaz, G.; Polkowski, W.; Hauser, N. The Central-European SentiMag study: Sentinel lymph node biopsy with superparamagnetic iron oxide (SPIO) vs. radioisotope. *Breast* 2014, 23, 175–179. [CrossRef]
- Geißen, W.; Engels, S.; Aust, P.; Schiffmann, J.; Gerullis, H.; Wawroschek, F.; Winter, A. Diagnostic accuracy of magnetometerguided sentinel lymphadenectomy after intraprostatic injection of superparamagnetic iron oxide nanoparticles in intermediateand high-risk prostate cancer using the magnetic activity of sentinel nodes. *Front. Pharmacol.* 2019, 10, 1123. [CrossRef] [PubMed]
- Winter, A.; Kneib, T.; Wasylow, C.; Reinhardt, L.; Henke, R.-P.; Engels, S.; Gerullis, H.; Wawroschek, F. Updated nomogram incorporating percentage of positive cores to predict probability of lymph node invasion in prostate cancer patients undergoing sentinel lymph node dissection. *J. Cancer* 2017, *8*, 2692–2698. [CrossRef]
- Wawroschek, F.; Vogt, H.; Weckermann, D.; Wagner, T.; Hamm, M.; Harzmann, R. Radioisotope guided pelvic lymph node dissection for prostate cancer. J. Urol. 2001, 166, 1715–1719. [CrossRef]
- 32. Winter, A.; Engels, S.; Süykers, M.-C.; Henke, R.-P.; Wawroschek, F. Radioisotope guided sentinel lymph node dissection in prostate cancer: Rate of lymph node involvment depending on preoperative tumor characteristics in more than 2100 patients. *SM J. Urol.* **2015**, *1*, 1002.

- 33. Weingärtner, K.; Ramaswamy, A.; Bittinger, A.; Gerharz, E.W.; Voge, D.; Riedmiller, H. Anatomical basis for pelvic lymphadenectomy in prostate cancer: Results of an autopsy study and implications for the clinic. *J. Urol.* **1996**, *156*, 1969–1971. [CrossRef]
- 34. R Core Team. *R: A Language and Environment for Statistical Computing;* R Foundation for Statistical Computing: Vienna, Austria, 2021. Available online: https://www.r-project.org/ (accessed on 4 June 2021).
- 35. Holl, G.; Dorn, R.; Wengenmair, H.; Weckermann, D.; Sciuk, J. Validation of sentinel lymph node dissection in prostate cancer: Experience in more than 2,000 patients. *Eur. J. Nucl. Med. Mol. Imaging* **2009**, *36*, 1377–1382. [CrossRef]
- Wawroschek, F.; Vogt, H.; Wengenmair, H.; Weckermann, D.; Hamm, M.; Keil, M.; Graf, G.; Heidenreich, P.; Harzmann, R. Prostate lymphoscintigraphy and radio-guided surgery for sentinel lymph node identification in prostate cancer. *Urol. Int.* 2003, 70, 303–310. [CrossRef] [PubMed]
- Abdollah, F.; Suardi, N.; Gallina, A.; Bianchi, M.; Tutolo, M.; Passoni, N.; Fossati, N.; Sun, M.; dell'Oglio, P.; Salonia, A.; et al. Extended pelvic lymph node dissection in prostate cancer: A 20-year audit in a single center. *Ann. Oncol.* 2013, 24, 1459–1466. [CrossRef]
- Briganti, A.; Chun, F.K.H.; Salonia, A.; Gallina, A.; Farina, E.; Da Pozzo, L.F.; Rigatti, P.; Montorsi, F.; Karakiewicz, P.I. Validation of a nomogram predicting the probability of lymph node invasion based on the extent of pelvic lymphadenectomy in patients with clinically localized prostate cancer. *BJU Int.* 2006, *98*, 788–793. [CrossRef] [PubMed]
- 39. Briganti, A.; Larcher, A.; Abdollah, F.; Capitanio, U.; Gallina, A.; Suardi, N.; Bianchi, M.; Sun, M.; Freschi, M.; Salonia, A.; et al. Updated nomogram predicting lymph node invasion in patients with prostate cancer undergoing extended pelvic lymph node dissection: The essential importance of percentage of positive cores. *Eur. Urol.* 2012, *61*, 480–487. [CrossRef] [PubMed]
- Godoy, G.; Chong, K.T.; Cronin, A.; Vickers, A.; Laudone, V.; Touijer, K.; Guillonneau, B.; Eastham, J.A.; Scardino, P.T.; Coleman, J.A. Extent of pelvic lymph node dissection and the impact of standard template dissection on nomogram prediction of lymph node involvement. *Eur. Urol.* 2011, 60, 195–201. [CrossRef]
- Grivas, N.; Wit, E.; Tillier, C.; van Muilekom, E.; Pos, F.; Winter, A.; van der Poel, H. Validation and head-to-head comparison of three nomograms predicting probability of lymph node invasion of prostate cancer in patients undergoing extended and/or sentinel lymph node dissection. *Eur. J. Nucl. Med. Mol. Imaging* 2017, 44, 2213–2226. [CrossRef]
- 42. Anninga, B.; White, S.H.; Moncrieff, M.; Dziewulski, P.; Geh, J.L.C.; Klaase, J.; Garmo, H.; Castro, F.; Pinder, S.; Pankhurst, Q.A.; et al. Magnetic technique for sentinel lymph node biopsy in melanoma: The MELAMAG trial. *Ann. Surg. Oncol.* **2016**, *23*, 2070–2078. [CrossRef]
- Baena Fustegueras, J.A.; González, F.H.; Calderó, S.G.; de la Fuente Juárez, M.C.; López, S.R.; Riu, F.R.; Petit, N.M.; Álvarez, P.M.; Torelló, A.L.; Matias-Guiu, X.; et al. Magnetic detection of sentinel lymph node in papillary thyroid carcinoma: The MAGIC-PAT study results. *Eur. J. Surg. Oncol.* 2019, 45, 1175–1181. [CrossRef]
- Hernando, J.; Aguirre, P.; Aguilar-Salvatierra, A.; Leizaola-Cardesa, I.O.; Bidaguren, A.; Gómez-Moreno, G. Magnetic detection of sentinel nodes in oral squamous cell carcinoma by means of superparamagnetic iron oxide contrast. *J. Surg. Oncol.* 2020, 121, 244–248. [CrossRef]
- Murakami, K.; Kotani, Y.; Suzuki, A.; Takaya, H.; Nakai, H.; Matsuki, M.; Sato, T.; Mandai, M.; Matsumura, N. Superparamagnetic iron oxide as a tracer for sentinel lymph node detection in uterine cancer: A pilot study. *Sci. Rep.* 2020, 10, 1–9. [CrossRef]
- Mottet, N.; Bellmunt, J.; Bolla, M.; Briers, E.; Cumberbatch, M.G.; De Santis, M.; Fossati, N.; Gross, T.; Henry, A.M.; Joniau, S.; et al. EAU-ESTRO-SIOG guidelines on prostate cancer. Part 1: Screening, diagnosis, and local treatment with curative intent. *Eur. Urol.* 2017, 71, 618–629. [CrossRef]
- 47. Morgan-Parkes, J.H. Metastases: Mechanisms, pathways, and cascades. *Am. J. Roentgenol.* **1995**, *164*, 1075–1082. [CrossRef] [PubMed]
- 48. Weckermann, D.; Dorn, R.; Holl, G.; Wagner, T.; Harzmann, R. Limitations of radioguided surgery in high-risk prostate cancer. *Eur. Urol.* 2007, *51*, 1549–1558. [CrossRef] [PubMed]
- 49. Kroon, B.K.; Horenblas, S.; Estourgie, S.H.; Lont, A.P.; Valdés Olmos, R.A.; Nieweg, O.E. How to avoid false-negative dynamic sentinel node procedures in penile carcinoma. *J. Urol.* 2004, *171*, 2191–2194. [CrossRef]
- 50. Leijte, J.A.P.; van der Ploeg, I.M.C.; Valdés Olmos, R.A.; Nieweg, O.E.; Horenblas, S. Visualization of tumor blockage and rerouting of lymphatic drainage in penile cancer patients by use of SPECT/CT. *J. Nucl. Med.* **2009**, *50*, 364. [CrossRef]
- 51. Mattei, A.; Fuechsel, F.G.; Bhatta Dhar, N.; Warncke, S.H.; Thalmann, G.N.; Krause, T.; Studer, U.E. The template of the primary lymphatic landing sites of the prostate should be revisited: Results of a multimodality mapping study. *Eur. Urol.* **2008**, *53*, 118–125. [CrossRef] [PubMed]