

# Untangling the association of amyloid- $\beta$ and tau with synaptic and axonal loss in Alzheimer's disease

Joana B. Pereira,<sup>1,2</sup> Shorena Janelidze,<sup>1</sup> DRik Ossenkoppele,<sup>1,3</sup> Hlin Kvartsberg,<sup>4,5</sup> Ann Brinkmalm,<sup>4,5</sup> Niklas Mattsson-Carlgren,<sup>6,7,8</sup> Erik Stomrud,<sup>1,9</sup> Ruben Smith,<sup>1,7</sup> Henrik Zetterberg,<sup>4,5,10,11</sup> Kaj Blennow<sup>4,5</sup> and Oskar Hansson<sup>1,9</sup>

See Jagust (doi:10.1093/brain/awaa402) for a scientific commentary on this article.

It is currently unclear how amyloid- $\beta$  and tau deposition are linked to changes in synaptic function and axonal structure over the course of Alzheimer's disease. Here, we assessed these relationships by measuring presynaptic (synaptosomal-associated protein 25, SNAP25; growth-associated protein 43, GAP43), postsynaptic (neurogranin, NRGN) and axonal (neurofilament light chain) markers in the CSF of individuals with varying levels of amyloid- $\beta$  and tau pathology based on <sup>18</sup>F-flutemetamol PET and <sup>18</sup>F-flortaucipir PET. In addition, we explored the relationships between synaptic and axonal markers with cognition as well as functional and anatomical brain connectivity markers derived from resting-state functional MRI and diffusion tensor imaging. We found that the presynaptic and postsynaptic markers SNAP25, GAP43 and NRGN are elevated in early Alzheimer's disease i.e. in amyloid- $\beta$ -positive individuals without evidence of tau pathology. These markers were associated with greater amyloid- $\beta$  pathology, worse memory and functional changes in the default mode network. In contrast, neurofilament light chain was abnormal in later disease stages, i.e. in individuals with both amyloid- $\beta$  and tau pathology, and correlated with more tau and worse global cognition. Altogether, these findings support the hypothesis that amyloid- $\beta$  and tau might have differential downstream effects on synaptic and axonal function in a stage-dependent manner, with amyloid-related synaptic changes occurring first, followed by tau-related axonal degeneration.

- 1 Clinical Memory Research Unit, Department of Clinical Sciences, Lund University, Malmö, Sweden
- 2 Division of Clinical Geriatrics, Department of Neurobiology, Care Sciences and Society, Karolinska Institute, Stockholm, Sweden
- 3 Department of Neurology and Alzheimer Center, Neuroscience Campus Amsterdam, VU University Medical Center, Amsterdam, The Netherlands
- 4 Institute of Neuroscience and Physiology, the Sahlgrenska Academy at University of Gothenburg, Mölndal, Sweden
- 5 Clinical Neurochemistry Laboratory, Sahlgrenska University Hospital, Mölndal, Sweden
- 6 Department of Clinical Sciences, Malmö, Lund University, Lund, Sweden
- 7 Department of Neurology, Skåne University Hospital, Lund University, Lund, Sweden
- 8 Wallenberg Center for Molecular Medicine, Lund University, Lund, Sweden
- 9 Memory Clinic, Skåne University Hospital, Malmö, Sweden
- 10 Department of Neurodegenerative Disease, UCL Institute of Neurology, London, UK
- 11 UK Dementia Research Institute at UCL, London, UK

Correspondence to: Joana B. Pereira Memory Clinic, Skåne University Hospital, SE-20502 Malmö, Sweden E-mail: joana.pereira@ki.se

Correspondence may also be addressed to: Oskar Hansson E-mail: Oskar.Hansson@med.lu.se

Received May 19, 2020. Revised September 4, 2020. Accepted September 21, 2020. Advance access publication December 5, 2020 © The Author(s) (2020). Published by Oxford University Press on behalf of the Guarantors of Brain.

This is an Open Access article distributed under the terms of the Creative Commons Attribution Non-Commercial License (http://creativecommons.org/licenses/by-nc/4.0/), which permits non-commercial re-use, distribution, and reproduction in any medium, provided the original work is properly cited. For commercial re-use, please contact journals.permissions@oup.com

**Keywords:** neurogranin; neurofilament; amyloid-β PET; tau PET; MRI **Abbreviation:** NfL = neurofilament light chain

### Introduction

Alzheimer's disease is a slowly progressing disorder in which pathophysiological changes precede clinical symptoms by many years (Jack *et al.*, 2010). There is growing evidence suggesting that increases in amyloid- $\beta$  may be one of the triggers that initiates Alzheimer's disease by setting off a chain of events that include the accumulation of toxic forms of tau, which eventually cause downstream neurodegeneration and dementia (Jack *et al.*, 2010, 2013*a*, 2014). However, it is currently unclear how rising amyloid- $\beta$  levels lead to tau deposition and why there is such a long delay between them.

Recently, a hypothesis has been proposed that could explain how these two events are temporally related (Edwards, 2019). This hypothesis suggests that, in early stages of Alzheimer's disease, as amyloid- $\beta$  levels start to rise, plaques begin to be deposited and progressively increase in size, reducing glutamatergic transmission and damaging nearby synapses (Wu et al., 2010; Burgold et al., 2011). This initial damage to synapses close to amyloid-ß plaques produces a local network dysfunction that evokes a response from microglia, which remove damaged synapses to prevent further damage or major network disruption (Cummings et al., 2017). However, as more amyloid- $\beta$  plaques continue to accumulate, particularly across multiple locations, the synaptic damage becomes more pronounced and spreads, resulting in phosphorylation of tau (Howlett et al., 2008; Hong et al., 2016), dissociation of tau from microtubules and the formation of tau tangles, which in turn triggers axon loss and neurodegeneration (Jack et al., 2018).

Although this proposed sequence of toxic effects is supported by several animal studies of Alzheimer's disease (Zempel et al., 2010, 2013; Rozkalne et al., 2011; Cohen et al., 2013; Spires-Jones and Hyman, 2014; Koffie et al., 2019), to our knowledge it has yet to be extensively tested in human subjects. This is particularly timely because of the recent development of tau tracers with PET and the novel assays for measuring synaptic and axonal proteins in the CSF. Thus, the aim of this study was to understand the differential impact of amyloid- $\beta$  and tau on synaptic loss and axonal integrity during earlier Alzheimer's disease stages, i.e. in amyloid-positive and amyloid-negative individuals who do not yet have tau pathology, as well as later Alzheimer's disease stages, i.e. in tau-positive and tau-negative individuals with established amyloid- $\beta$  pathology. To this end, we measured the concentrations of presynaptic (synaptosomalassociated protein 25, SNAP25; growth-associated protein 43, GAP43), postsynaptic (neurogranin, NRGN) and axonal (neurofilament light chain, NfL) markers in the CSF. We then tested which of these CSF markers was associated with

regional amyloid-B and tau deposition measured with amyloid-PET and tau-PET, and we assessed their relationship with memory and global cognition. Finally, using restingstate functional MRI and diffusion tensor imaging, we identified which of the available CSF and PET markers was more closely associated with changes in the default-mode network and degeneration of the parahippocampal tract, which are well-established connectivity measures of Alzheimer's disease (Pievani et al., 2011). Our underlying hypothesis was that global amyloid-ß deposition would be associated with changes in synaptic markers, memory dysfunction and default-mode network activity, whereas elevated tau PET would be accompanied by more pronounced axonal degeneration, global cognitive decline and parahippocampal white matter damage. The unique combination of imaging and CSF measures places this study in an ideal position to test the multimodal biomarker relationships in the course of Alzheimer's disease.

### Materials and methods

### **Participants**

The present study included 128 individuals from the Swedish BioFINDER cohort (http://biofinder.se/), an ongoing longitudinal study designed to identify and develop new markers for neurodegenerative disorders, particularly Alzheimer's disease. For this study, only subjects with <sup>18</sup>F-flutemetamol PET, <sup>18</sup>F-flortaucipir PET and CSF measures were included. Cognitively normal subjects were required to have a Clinical Dementia Rating score of 0, 27-30 points on the Mini-Mental State Examination, not fulfil criteria for mild cognitive impairment or dementia, have no history of cognitive change over time, and be fluent in Swedish. Cognitively impaired subjects were required to fulfil the DSM-5 criteria for mild neurocognitive disorder or major neurocognitive disorder due to Alzheimer's disease (American Psychiatric Association, 2013), and exhibit abnormal amyloid-B accumulation based on <sup>18</sup>F-flutemetamol PET. All subjects underwent the Mini-Mental State Examination (Folstein et al., 1975) and delayed word list recall test from the ADAS-Cog (Alzheimer's Disease Assessment Scale - Cognitive Subscale) (Rosen et al., 1984) to assess global cognition and episodic memory, respectively. There were not global ADAS-Cog scores available for this sample.

The Regional Ethical Review Board of Lund University, the Swedish Medicines and Products Agency, and the Radiation Safety Committee of Skåne University Hospital in Sweden approved the study and written, informed consent was obtained from all participants according to the Declaration of Helsinki.

### Image acquisition

All subjects underwent structural MRI on a Siemens Tim Trio 3 T scanner, <sup>18</sup>F-flutemetamol PET on a Philips Gemini TF 16 scanner, and <sup>18</sup>F-flortaucipir PET on a General Electrics Discovery 690 scanner. <sup>18</sup>F-flutemetamol PET images were acquired 90 to 110 min after injection of 185 MBq <sup>18</sup>F-flutemetamol and reconstructed into  $4 \times 5$  frames using the line-of-response row-action maximum-likelihood algorithm (Palmqvist et al., 2016). <sup>18</sup>F-flortaucipir PET images were acquired 80 to 100 min after injection of 370 MBg <sup>18</sup>F-flortaucipir, reconstructed into  $4 \times 5$  frames using an iterative Vue Point HD algorithm with six subsets, 18 iterations with 3 mm filter and no time-of-flight correction (Hahn et al., 2016). The structural T<sub>1</sub>weighted images were acquired using a magnetization-prepared rapid gradient echo sequence using the following parameters: 176 slices, repetition time: 1950 ms, echo time: 3.4 ms, inversion time: 900 ms, flip angle: 9°, 1 mm isotropic voxels.

In addition, a subsample also underwent resting-state functional MRI and diffusion tensor imaging on a Siemens Tim Trio 3 T scanner. Resting-state functional MRI images were acquired using a gradient-echo planar imaging pulse sequence with the following parameters: 180 volumes, 33 slices, repetition time: 2000 ms, echo time: 30 ms, 3 mm isotropic voxels. Diffusion tensor imaging scans were acquired with 64 diffusion-weighted directions at a b-value of 1000 s/mm<sup>2</sup> using an echo-planar imaging sequence with the following parameters: 65 axial slices; repetition time: 8200 ms; echo time: 86 ms, no inter-slice gap, 2 mm isotropic voxels.

### Image preprocessing

All PET images were motion-corrected, time-averaged and coregistered to their skull stripped  $T_1$ -weighted scans. Amyloid- $\beta$ positivity was established using a composite cortical region normalized by the whole cerebellum, brainstem and eroded subcortical white matter (Landau et al., 2015) on <sup>18</sup>F-flutemetamol data with a cut-off of > 0.693. This cut-off was obtained from linear mixed models in a large group of subjects of BioFINDER (n = 406) using a composite cortical region of interest in addition to the whole cerebellum, the pons/brainstem and eroded cortical white matter as a reference region (Palmqvist et al., 2016). We excluded borderline cases and used cut-offs that were  $\pm 5\%$  from the original cut-offs to avoid drawing conclusions based on cases that could be easily misclassified because of variability in the measurements or subthreshold amyloid-ß positivity. Tau positivity was calculated using a composite set of brain regions corresponding to a temporal meta-region of interest normalized by the inferior cerebellum grey matter on<sup>18</sup>F-flortaucipir data with a previously established cut-off of >1.34 (Ossenkoppele et al., 2018).

The resting-state functional MRI scans were analysed using multivariate exploratory linear decomposition into independent components (MELODIC), as implemented in FSL (FMRIB's Software Library, www.fmrib.ox.ac.uk/fsl). This is a data-driven method that extracts temporally related signals, which represent functionally relevant networks. Using different modules of FSL, the following preprocessing was applied to the functional MRI scans: removal of the first five volumes, motion correction, slicetiming correction, brain extraction, spatial smoothing using a Gaussian kernel of 8 mm and high-pass temporal filtering of 100 s. After preprocessing, all images were registered to the MNI (Montreal Neurological Institute) space using a mean echo-planar image (EPI) generated from all subjects. The subject's time series were then temporally concatenated into a single 4D time series and separated in 20 independent components. Of these 20 components, two quantitatively overlapped with the anterior and posterior default mode networks provided by Biswal *et al.* (2010). We binarized these network maps and extracted the mean temporal time series of each map from all subjects.

Diffusion tensor images were corrected for distortions caused by eddy currents and head motion, and skull-striped using the eddy\_correct tool of FSL (https://fsl.fmrib.ox.ac.uk/fsl/fslwiki). A diffusion tensor model was then fitted at each voxel to calculate the fractional anisotropy maps for each subject using the Diffusion Toolbox (Behrens et al., 2007). All images were visually inspected to ensure whole brain coverage, absence of artefacts (venetian blind, checkers and stripe blurring artefacts) or spatial distortions in the temporal poles caused by field inhomogeneities. To extract structural connectivity values from a relevant Alzheimer's disease connectivity marker we used the hippocampal cingulum mask from the John Hopkins University (JHU) white matter tractography atlas (Hua et al., 2008), corresponding to the parahippocampal tract. We binarized this mask, coregistered it to the native fractional anisotropy images of all subjects and extracted their mean fractional anisotropy values.

### **CSF** biomarker measurements

NRGN and NfL were measured at the Clinical Neurochemistry Laboratory at University of Gothenburg, Mölndal, Sweden, using an in-house immunoassay for NRGN (Kvartsberg *et al.* 2019) and GAP43 (Sandelius *et al.* 2019) and a commercial ELISA for NfL (NF-light<sup>®</sup> ELISA, Uman Diagnostics) (Zetterberg *et al.*, 2016). For SNAP25 measurements, we used an assay that consists of enrichment with immunoprecipitation (KingFisher<sup>TM</sup> Flex System) followed by digestion, addition of heavy isotope-labelled standards, and quantitation with liquid chromatography/selected reaction monitoring mass spectrometry (LC-SRM/MS) (Agilent 6490 QQQ MS).

### Statistical analyses

Statistical analyses were carried out using SPSS 25.0 (IBM Corp., Armonk, NY, USA) and R (version 3.5.1). Square root, logarithmic or reciprocal transformations were applied to CSF biomarkers that were not normally distributed. Then, a set of pairwise *t*-tests was used to compare these biomarkers between amyloid-positive and amyloid-negative individuals without evidence of tau pathology (earlier disease stages) and between taupositive and tau-negative individuals with amyloid- $\beta$  pathology (later disease stages). These analyses were adjusted for age in addition to sex for the NfL comparisons since this marker was significantly higher in males compared to females (P < 0.05). Sixteen subjects had missing SNAP25 levels due to technical problems. For all the analyses, we report the degrees of freedom or number of subjects.

To assess the ability of synaptic and axonal markers in predicting amyloid- $\beta$  and tau pathology, we built linear regression models with the markers as dependent variables and amyloid- $\beta$ and tau as the outcomes, using the forward selection method and adjusting for age and sex. For these analyses, we verified that the residuals were normally distributed, there was no heteroscedasticity, and no multicollinearity between the variables, which was determined using a variance inflation factor below five.

Spearman partial correlation analyses were then used to assess the relationship between memory or global cognition with synaptic and axonal markers, while adjusting for the previous covariates in addition to education. These analyses were carried out in earlier and later Alzheimer's disease stages.

We then used linear regression models to assess which of the PET (amyloid- $\beta$ , tau), synaptic (SNAP25, GAP43, NRGN) and axonal (NfL) markers was the best predictor of functional activity in the anterior and posterior default-mode networks as well as fractional anisotropy in the parahippocampal tract. Again, here we verified that all linear regression model assumptions were met.

Finally, we conducted mediation analyses to test different models based on the expression 'X  $\rightarrow$  Y, mediated by M', while controlling for covariates. The significance of the mediation was assessed by calculating bias-corrected 95% confidence intervals (CIs) using bootstrapping (500 resamples).

All analyses were adjusted for multiple comparisons using false discovery rate (FDR) corrections (q < 0.05, two-tailed).

### **Data availability**

Anonymized data will be shared by request from a qualified academic investigator for the sole purpose of replicating procedures and results presented in the article and as long as data transfer is in agreement with EU legislation on the general data protection regulation and decisions by the Ethical Review Board of Sweden and Region Skåne, which should be regulated in a material transfer agreement.

### Results

### **Study participants**

In this cross-sectional and multimodal study of 128 participants, 62 were cognitively normal and 66 were cognitively impaired, the latter being diagnosed with mild cognitive impairment or Alzheimer's disease dementia. Since previous evidence shows that amyloid and tau accumulation have different non-linear trajectories over the course of Alzheimer's disease, with amyloid initially accelerating and then later decelerating or even decreasing, whereas tau is initially low and then later accelerates (Kadir et al., 2012; Jack et al., 2010, 2013a, b, 2014, 2018), we divided the participants into three groups using a global amyloid-β composite cortical region and a temporal tau-PET meta-region of interest. Thus, using PET measures, in the current study 37 subjects were amyloid-negative and tau-negative (amyloid-β-Tau-), 35 were amyloid-positive and tau-negative (amyloid- $\beta$  + Tau-) and 53 were both amyloid-positive and tau-positive  $(amyloid-\beta + Tau +)$  (Table 1). Only three subjects were amyloid- $\beta$ -Tau+, who were excluded from the analyses because of the small number of cases in the group. To motivate our approach of dividing the sample into groups, we explored the non-linear relationships between the

biomarkers using spline models across the entire cohort (Supplementary material). These models showed that the relationships between amyloid PET and synaptic markers are quadratic: they are positively correlated in initial stages, then reach a stable point, and afterwards they become slightly negatively correlated. This suggests that amyloid seems to have a detrimental effect on synaptic damage only in early stages and that this effect cannot be captured by assessing these relationships across the entire sample. In contrast, the spline models also show that tau PET only has an effect on NfL in later disease stages, with Tau– subjects being concentrated on one side of the plot, whereas Tau + subjects spread out on the other side, driving the correlations.

A subsample of our cohort also underwent resting state functional MRI (66%, 43 cognitively unimpaired, 41 cognitively impaired) and diffusion tensor imaging (49%, 25 cognitively unimpaired, 38 cognitively impaired) to assess the relationship between synaptic, axonal and PET maskers with brain connectivity measures of Alzheimer's disease.

All the analyses performed in this study were repeated using a less stringent cut-off of 1.22 to define tau positivity and divide subjects into Tau + and Tau- groups. This cutoff was calculated following the recommendations by Jack *et al.* (2017) to calculate a sensitivity-based tau PET cut-off. The results of these analyses can be found in the Supplementary material.

### Synaptic and axonal biomarkers change in earlier and later Alzheimer's disease stages

Our findings showed significant increases in the synaptic markers SNAP25 [F(2,63) = 9.08, P = 0.004], GAP43 [F(2,70) = 6.12, P = 0.016] and NRGN [F(2,70) = 7.75, P = 0.007] but not in the axonal marker NfL [F(2,70) = 0.73, P = 0.395] (Fig. 1A) in earlier Alzheimer's disease stages, i.e. in amyloid- $\beta$  + Tau- compared to amyloid- $\beta$ -Tau- cases. Similar results were found after excluding subjects who did not have values for all CSF markers [SNAP25: F(2,61) = 10.63, P = 0.002; GAP43: F(2,63) = 4.35, P = 0.041; NRGN: F(2,61) = 10.30, P = 0.002; NfL: F(2,61) = 2.75, P = 0.103].

In contrast, significant increases in NfL [F(2,86) = 18.70, P < 0.001] but not SNAP25 [F(2,75) = 1.83, P = 0.180], GAP43 [F(2,86) = 1.99, P = 0.162] or NRGN [F(2,86) = 0.66, P = 0.421] were found in later Alzheimer's disease stages, i.e. in amyloid- $\beta$  + Tau + versus amyloid- $\beta$  + Tau-cases (Fig. 1B). Again, these results were similar after excluding subjects without all markers [SNAP25: F(2,73) = 0.76, P = 0.385; GAP43: F(2,75) = 3.78, P = 0.100; NRGN: F(2,73) = 0.944, P = 0.334; NfL: F(2,73) = 17.79, P < 0.001]. To examine whether the later results were influenced by the presence of cognitive impairment, we performed an additional analysis that compared the same panel of markers between amyloid- $\beta$  + Tau + and amyloid- $\beta$  + Tau - groups but only in subjects who were cognitively



**Figure 1 Synaptic and axonal markers change in earlier and later Alzheimer's disease stages.** (**A**) Representation of the neuronal aspects measured by the postsynaptic [NRGN (Ng)], axonal (NfL) and presynaptic markers (SNAP25; GAP43). (**B**) In early Alzheimer's disease stages, SNAP25, GAP43 and NRGN show significant increases in amyloid- $\beta$  + ( $A\beta$  +) compared to amyloid- $\beta$  – ( $A\beta$ -) individuals without evidence of tau PET retention (Tau-). (**C**) In later Alzheimer's disease stages, NfL shows significant increases in Tau + compared to Tau- individuals who are all amyloid- $\beta$  + . (**D**) The increases in NfL in the previous groups can still be observed when only cognitively impaired (Cl) individuals are considered. These analyses were performed using t-tests, while controlling for covariates. The values presented in the plots are the residuals of SNAP25, GAP43, NRGN and NfL after regressing out the effects of age (all CSF markers) in addition to sex (NfL).

impaired. This analysis showed that NfL was still significantly increased in cognitively impaired tau-positive compared to tau-negative individuals [F(2,60) = 8.23, P = 0.006], whereas synaptic proteins did not show changes between these groups [SNAP25: F(2,49) = 1.21, P = 0.277; GAP43: F(2,60) = 0.17, P = 0.678; NRGN: F(2,60) = 0.61, P = 0.437] (Fig. 1C). Altogether, these results are in line with the hypothesis that amyloid-related synaptic changes occur early in Alzheimer's disease, even when widespread tau pathology is not yet present, whereas axonal degeneration is more closely associated with the development of tau pathology in cases that are amyloid-positive. We note that these results do not indicate there is no synaptic damage in later stages of Alzheimer's disease. In fact, if we compare amyl $oid-\beta + Tau + subjects$  to the amyloid- $\beta$ -Tau- group, there are significant differences in all synaptic markers [NRGN: F(2,88) = 16.79, P < 0.001; GAP43: F(2,88) = 17.043,P < 0.001; SNAP25: F(2,74) = 19.76, P < 0.001]. Our results suggest that synaptic damage is not significantly different between amyloid- $\beta$  + Tau- and amyloid- $\beta$  + Tau + subjects, in contrast to axonal damage, which is significantly different, indicating that synaptic changes emerge in earlier disease stages, whereas axonal damage emerges in later disease stages.

### Synaptic degeneration and axonal damage correlate with high amyloid- $\beta$ and tau

To assess whether synaptic and axonal proteins are associated with the severity of amyloid- $\beta$  and tau deposition in earlier and later Alzheimer's disease stages, we performed linear regression models using global amyloid-PET and temporal tau-PET standard uptake value ratio (SUVR) values as the outcome and SNAP25, GAP43, NRGN and NfL as the predictors. The goal of these analyses is to select the CSF marker that is the best predictor of amyloid and tau deposition, while excluding the other markers that were also associated with amyloid and tau but did not significantly improve the fit of the model when added to it. We found that increasing NRGN levels were associated with elevated global amyloid-PET SUVR  $(r^2 = 0.11, t = 2.51, P = 0.015)$  in cases with normal tau-PET (i.e. in amyloid- $\beta$ -Tau- and amyloid- $\beta$  + Tau- cases) (Fig. 2A), explaining 11% of the variance in amyloid- $\beta$ deposition, in contrast to SNAP25 (t = 0.933, P = 0.355), GAP43 (t = -0.573, P = 0.569) or NfL (t = 0.858, P = 0.394), which did not reach significance. These findings indicate that postsynaptic changes are associated with the amount of amyloid-B pathology even before development of evident tau pathology.

On the other hand, increasing NfL levels correlated best with tau-PET SUVR values ( $r^2 = 0.39$ , t = 5.42, P < 0.001), explaining 39% of the variance in tau deposition in cases with amyloid- $\beta$  pathology (i.e. in amyloid- $\beta$ +Tau- and amyloid- $\beta$ +Tau+) (Fig. 2B), in contrast to SNAP25

(t = 0.35, P = 0.728), GAP43 (t = 1.66, P = 0.102) and NRGN (t = 1.48, P = 0.145), indicating that axonal degeneration is more closely related to the levels of tau pathology in Alzheimer's disease.

### Memory and global cognition correlate with synaptic and axonal markers in a stage-dependent manner

The temporal course of cognitive decline in Alzheimer's disease is characterized by early changes in memory, which are then followed by other deficits and global cognitive decline. To test the hypothesis that synaptic and axonal proteins would be differentially associated with cognitive functions that are affected in earlier or later disease stages, we conducted correlation analyses between delayed memory (word list recall from ADAS-Cog), global cognition (Mini-Mental State Examination), SNAP25, GAP43, NRGN and NfL.

When the analyses were restricted to individuals without evident tau pathology (i.e. in amyloid-B-Tau- and amyloid- $\beta$  + Tau- cases), we found that the only correlations that remained significant were between memory and both presynaptic (GAP43;  $\rho = 0.30$ , P = 0.011) (Fig. 3A) and postsynaptic (NRGN:  $\rho = 0.28$ , P = 0.017) (Fig. 3B) markers (there were also a trend for SNAP25:  $\rho = 0.26$ , P = 0.039, uncorrected for multiple comparisons). In contrast, when the analyses were restricted to individuals with evident amyloid- $\beta$  pathology (i.e. amyloid- $\beta$  + Tau- and amyloid- $\beta$  + Tau + cases), the only remaining significant correlation was between NfL and global cognition ( $\rho = -0.25$ , P = 0.020) (Fig. 3C). This correlation was no longer significant when it was tested in amyloid- $\beta$  + Tau- and amyloid- $\beta$  + Tau + cases separately, possibly due to a lower number of subjects in the separate groups.

### Functional and structural connectivity changes are associated with synaptic loss and tau pathology

For individuals who underwent resting state functional MRI and diffusion tensor imaging, we extracted connectivity measures that are known to be sensitive to Alzheimer's disease (Pievani *et al.*, 2011) and tested their relationship with our panel of CSF and PET markers. These connectivity measures consisted of brain activation signals from the anterior and posterior default mode networks as well as the fractional anisotropy values of the parahippocampal tract.

Our models showed that NRGN was associated with both reduced connectivity in the posterior default-mode network ( $r^2 = 0.33$ , t = -3.27, P = 0.002) (Fig. 4A) and increased connectivity in the anterior default-mode network ( $r^2 = 0.37$ , t = 2.278, P = 0.020) (Fig. 4B) in early Alzheimer's disease. When the signals of the anterior and posterior default-mode networks were combined, no significant associations with any marker were



**Figure 2 Synaptic degeneration and axonal damage correlate with high amyloid-\beta and tau. (A)** Among the synaptic and axonal markers, increasing NRGN (Ng) correlates best with greater amyloid- $\beta$  PET retention, i.e. in individuals who are amyloid- but do not show evidence of tau pathology. (B) In contrast, higher NfL correlates best with greater tau PET retention in later Alzheimer's disease stages, i.e. in individuals who are Tau- and Tau + with amyloid- $\beta$  pathology. The analyses were carried out using linear regression models. The values presented in the plots are the residuals of NRGN and NfL after regressing out the effects of age (all markers) in addition to sex (NfL). AD = Alzheimer's disease.



Figure 3 Memory and global cognition are associated with synaptic and axonal markers in a stage-dependent manner. (A) Memory measured by the word list recall test (ADAS-Cog) correlates with GAP43 and NRGN levels in early Alzheimer's disease stages (amyloid- $\beta$ +Tau- and amyloid- $\beta$ -Tau-), whereas (B) global cognition measured by the Mini-Mental State Examination correlates with NfL in later Alzheimer's disease stages (amyloid- $\beta$ +Tau+ and amyloid- $\beta$ +Tau-). The correlation analyses were performed using Spearman's  $\rho$ . These results are in line with findings showing memory deficits in early Alzheimer's disease and global cognitive impairment in later disease stages. The values presented in the plots are the residuals of GAP43, NRGN and NfL after regressing out the effects of age, sex and education. AD = Alzheimer's disease.

found. On the other hand, contrary to what we expected, tau PET SUVR correlated with reduced parahippocampal white matter integrity in later disease stages ( $r^2 = 0.16$ , t = -2.91, P = 0.006) (Fig. 5), instead of NfL (t = 0.90, P = 0.374).

## Synaptic loss mediates the effect of amyloid- $\beta$ on tau, whereas tau mediates the effect of NfL on cognition

To test the hypothesis that amyloid- $\beta$  deposition is followed by synaptic damage, tau aggregation, axonal degeneration and cognitive impairment, we conducted three mediation analyses. The first analysis tested whether the relationship between amyloid- $\beta$  and tau was mediated by synaptic loss in earlier Alzheimer's disease stages. Since NRGN was the synaptic marker that was most strongly associated with amyloid- $\beta$  in our previous analyses, we selected it as the synaptic mediator. The results of this analysis showed that NRGN mediates the effect between amyloid- $\beta$  and tau pathologies in early Alzheimer's disease (0.118, 95% CI: 0.03 to 0.24, P = 0.008).

The second analysis assessed whether, in later Alzheimer's disease stages, the effects of tau on global cognition were mediated by axonal damage measured with NfL. The results



**Figure 4 Synaptic loss predicts functional connectivity changes in the default-mode networks.** In earlier Alzheimer's disease stages (amyloid- $\beta$  + Tau– and amyloid- $\beta$ -Tau–), our linear regression models show that NRGN (Ng) is the marker that correlates best with connectivity increases in the anterior default-mode network (**A**) and connectivity decreases in the posterior default-mode network (**B**). The values presented in the plots are the residuals of functional MRI (fMRI) signals and NRGN after regressing out the effects of age and sex. AD = Alzheimer's disease.

of this analysis showed that NfL was not a mediator of this relationship (0.654, 95% CI: -0.351 to 1.700, P = 0.190).

Finally, the third analysis tested whether the relationship between NfL and global cognition was mediated by tau instead and this proved to be significant (4056.8, 95% CI: 2274.4 to 6099.5, P < 0.001).

Altogether, these results suggest that the relationship between amyloid- $\beta$  and tau is mediated by synaptic changes, in line with our initial hypothesis. In contrast, contrary to what we had predicted, the relationship between NfL and global cognition is mediated by tau pathology.

### Discussion

The aggregation of amyloid- $\beta$  and tau in brain regions serving memory and cognition are thought to be responsible for a

sequence of events that lead to clinical Alzheimer's disease (Hardy and Selkoe, 2002). However, a better understanding of the *in vivo* downstream effects of amyloid- $\beta$  and tau in human subjects is needed. Our findings agree with the hypothesis that amyloid- $\beta$  and tau are associated with synaptic dysfunction and axonal degeneration, respectively. These changes correlate with memory, global cognition and brain connectivity in a stage-dependent manner, suggesting they may be useful to track disease progression. Although the associations revealed by these data are cross-sectional, the most likely interpretation based on previous animal studies (Zempel et al., 2010, 2013; Rozkalne et al., 2011; Cohen et al., 2013; Spires-Jones and Hyman, 2014; Koffie et al., 2019), is that amyloid-β deposition is followed by synaptic damage, tau aggregation and neurofilament changes (Fig. 6). These findings might open possibilities for new therapeutic targets for Alzheimer's disease based on synaptic and axonal function.



Figure 5 Tau pathology correlates with structural connectivity changes in the parahippocampal tract. In later Alzheimer's disease stages (amyloid- $\beta$  + Tau + and amyloid- $\beta$  + Tau-), our linear regression models show that tau PET retention is the measure that correlates best with reduced fractional anisotropy (FA) in the parahippocampal tract (*left*). The values presented in the plots are the residuals of fractional anisotropy values and tau PET levels after regressing out the effects of age and sex.

There is growing evidence that amyloid- $\beta$  may be part of a mechanism controlling synaptic activity (Fagiani et al., 2019). In particular, it has been shown that amyloid- $\beta$  production is enhanced by action potential-dependent synaptic activity, leading to increased amyloid- $\beta$  at synapses and alterations of synaptic excitatory transmission (Palop and Mucke, 2010). These changes can affect both presynaptic and postsynaptic mechanisms of neuronal communication and impair overall brain activity (Gulisano et al., 2019). However, several other studies have also proposed that, compared to amyloid- $\beta$ , tau might have a more critical role in synaptic dysfunction. This is due to the presence of tau at the synapses in Alzheimer's disease brains (Tai *et al.*, 2012), the fact that tau pathology is more strongly associated with cognitive decline (Nelson et al., 2012) and its ability to spread trans-synaptically between neurons (Pooler et al., 2013).

Thus, motivated by these two different lines of evidence, in this study we sought to untangle the effects of amyloid- $\beta$ and tau on presynaptic and postsynaptic markers in earlier and later Alzheimer's disease stages, during which amyloid-β and tau could have a differential impact on synaptic loss and axonal integrity. We used SNAP25 and GAP43 as presynaptic markers because of their role in initiating fusion of synaptic vesicles for synaptic communication (Brinkmalm et al., 2014) or synaptic plasticity (Allegra Mascaro et al., 2013), respectively. In addition, we used NRGN as a postsynaptic marker since it is involved in long-term potentiation (Zhong and Gerges, 2010; Zetterberg and Blennow, 2015). We found that increases in SNAP25, GAP43 and NRGN occur already in early Alzheimer's disease stages in amyloid- $\beta$  + Tau- individuals. The increase in these synaptic proteins was associated with memory impairment and NRGN was

specifically associated with functional connectivity changes in the anterior and posterior default-mode networks. It is well known that memory loss is one of the first clinical symptoms of Alzheimer's disease (Carlesimo and Oscar-Berman, 1992). In addition, changes in network hyperactivity and hypoactivity are common in early Alzheimer's disease (Sheline and Raichle, 2013; Huijbers et al., 2015; Jones et al., 2016; Sepulcre et al., 2017), being part of a potentially compensatory mechanism that the brain enables to cope with initial pathology and maintain normal function (Elman et al., 2014). Although reduced connectivity is more expected in the context of a neurodegenerative disorder like Alzheimer's disease, increased connectivity can also occur and may be due to the overactivation of N-methyl-D-aspartic acid receptors by glutamate at the synaptic level, which are mediated by amyloid-ß (Gouras et al., 1997; Palop and Mucke, 2010). Altogether, these findings would fit a model in which amyloid- $\beta$  and synaptic loss are the key players in initial memory impairment and early network dysfunction. The assumption that amyloid- $\beta$  and synaptic damage are linked in Alzheimer's disease is supported by several animal studies showing that neurons lacking the amyloid precursor protein show greater excitatory synaptic transmission (Priller et al., 2006), neuritic outgrowth (Koo et al., 1993) and an increased number of synapses (Steinbach et al., 1998). They also agree with human studies showing that amyloid plaques are associated with both presynaptic and postsynaptic changes in non-demented individuals (Potter et al., 2011), a mechanism that might be potentially regulated by mitochondria (Reddy and Beal, 2008) or cholinergic receptors (Jürgensen and Ferreira, 2010). Thus, our findings confirm previous reports showing the detrimental role of amyloid pathology on synapses. The fact that the



Figure 6 Schematic hypothetical representation of the downstream effects of amyloid- $\beta$  and tau on synaptic and axonal function in earlier and later Alzheimer's disease stages. In earlier disease stages (amyloid- $\beta$  + Tau– and amyloid- $\beta$ -Tau–), the accumulation of amyloid- $\beta$  into plaques reduces the number of postsynaptic spines and the number of presynaptic vesicles, altering local synaptic communication. In later disease stages (amyloid- $\beta$  + Tau + and amyloid- $\beta$  + Tau–), as synaptic changes become more pronounced, tau becomes phosphorylated and aggregates into insoluble paired helical filaments (PHF), which form neuropil threads and neurofibrillary tangles. These changes are followed by the dissociation of neurofilaments from microtubules, axonal degeneration and ultimately global cognitive impairment. A $\beta$  = amyloid- $\beta$ ; AD = Alzheimer's disease.

postsynaptic marker NRGN was the best predictor of amyloid pathology compared to the presynaptic markers SNAP25 and GAP43 could be related to the fact that postsynaptic glutamate receptor trafficking has been shown to be a prime initial target for amyloid- $\beta$  by several previous studies (Almeida *et al.*, 2005; Roselli *et al.*, 2005; Snyder *et al.*, 2005; Shemer *et al.*, 2006), suggesting that postsynaptic terminals might be more affected by amyloid pathology in early stages of Alzheimer's disease. Thus, altogether, these findings agree with our hypothesis that synaptic markers would be associated with functional connectivity in earlier disease stages and are in line with previous evidence showing that functional connectivity is an early marker that changes in response to amyloid deposition (Sheline and Raichle, 2013; Huijbers *et al.*, 2015; Jones *et al.*, 2016; Sepulcre *et al.*, 2017).

As previously proposed by animal models (Howlett *et al.*, 2008; Hong *et al.*, 2016; Edwards, 2019), in later

#### Table | Cohort characteristics

|                              | Amyloid-β–Tau–<br>(n = 37) | Amyloid-β+<br>Tau–(n = 35) | Amyloid-β+<br>Tau+(n = 53) | Amyloid-β–<br>Tau– versus<br>amyloid-β+<br>Tau–(P-value) | Amyloid-<br>β+Tau-<br>versus amyl-<br>oid-β+Tau +<br>(P-value) |
|------------------------------|----------------------------|----------------------------|----------------------------|--|--|
| Age                          | 74 (52-87)                 | 77 (56-88)                 | 71 (40-88)                 | 0.124  | 0.003  |
| Sex, male/female             | 21/16                      | 19/16                      | 28/25                      | 0.833  | 0.893  |
| Education                    | 12 (7–18)                  | 10 (6–23)                  | 12 (5–20)                  | 0.235  | 0.264  |
| MMSE                         | 29.0 (27–30)               | 29.0 (21–30)               | 23.5 (7–29)                | 0.070  | < 0.00 l   |
| Delayed memory               | I.0 (0–5)                  | 3.0 (0–9)                  | 8.0 (0–10)                 | 0.005  | < 0.00 l   |
| Cognitively impaired, %      | 2.7                        | 37.1                       | 92.5                       | < 0.00 l   | < 0.00 l   |
| Global amyloid- $\beta$ SUVR | 0.65 (0.56–0.74)           | 0.95 (0.76–1.18)           | 1.02 (0.81–1.31)           | < 0.00 l   | < 0.00 l   |
| Temporal meta-ROI tau SUVR   | 1.15 (1.02–1.23)           | 1.21 (1.05–1.30)           | 1.80 (1.34–2.84)           | 0.002  | < 0.00 l   |
| SNAP25, pmol/l               | 16.83 (6.99–25.51)         | 21.92 (7.27–36.17)         | 24.82 (5.82–54.66)         | 0.004  | 0.180  |
| GAP43, pg/ml                 | 3576.16 (1788–6475)        | 4272.97 (2009–7926)        | 5003.62 (1193–9086)        | 0.016  | 0.162  |
| NGRN, pg/ml                  | 222.92 (90.98–527.49)      | 263.45 (90.04–494.03)      | 300.36 (112.34–547.84)     | 0.007  | 0.421  |
| NfL, pg/ml                   | 792.51 (388.44–11 632.0)   | 1065.10 (470.95–11 759.0)  | 1429.60 (517.78–13 322.0)  | 0.395  | < 0.00 l   |
| Subsample with rs-fMRI (%)   | 70.3                       | 82.9                       | 83.0                       | -  | -  |
| Subsample with DTI (%)       | 56.8                       | 62.9                       | 79.2                       | -  | _  |

Data are presented as median (range). P-values were derived from ANOVA for continuous normally distributed measures, Mann-Whitney tests for continuous non-normally distributed measures and chi-squared tests for categorical measures. DTI = diffusion tensor imaging; MMSE = Mini-Mental State Examination; ROI = region of interest; SUVR = standard uptake value ratio; rs-fMRI = resting-state functional MRI.

Alzheimer's disease stages the spread of amyloid- $\beta$ -induced synaptic damage along the length of the axon, results in phosphorylation, subsequent dissociation of tau from microtubules and the formation of tangles. Since the microtubules are the structures that support the axon, the dissociation of tau from these structures gives place to axonal degeneration (Spires-Jones et al., 2009). Axonal degeneration is thus an important aspect of Alzheimer's disease that can be measured indirectly using NfL, a scaffolding protein involved in the growth of axons (Gaetani et al., 2019), or with fractional anisotropy on diffusion tensor imaging. In this study, we found that NfL was significantly increased in later Alzheimer's disease stages, i.e. in tau-positive individuals with amyloid- $\beta$  pathology. These increases were associated with global cognition, which normally becomes impaired in more advanced Alzheimer's disease stages. However, contrary to what we expected, tau pathology was the best predictor of reduced parahippocampal white matter integrity, instead of NfL. This finding may be due to the size of the damaged microstructures associated with tau pathology or high NfL levels. Tau proteins can be found in microtubules, which have a diameter of ~25 nm (Ledbetter and Porter, 1963), whereas neurofilaments are much smaller, being  $\sim$ 1.42 nm (Gaetani *et al.*, 2019). Although the contribution of cellular components to diffusion anisotropy is not completely understood (DaSilva et al., 2003), there is evidence supporting that certain axonal structures such as neurofilaments and microtubules might play an important role in this process (Beaulieu and Allen, 1994). In addition, other metabolic mechanisms such as axonal transport, which is primarily carried out through the microtubules, has also been suggested to be associated with anisotropy (Beaulieu, 2002). Thus, it is plausible that, compared to neurofilament injury,

damage to the microtubules, which are larger in size, would have a greater impact on the integrity of white matter fibres by increasing the number of larger obstacles to directional water diffusion, which is captured by fractional anisotropy on diffusion brain images. Altogether, these findings suggest that, in the presence of amyloid- $\beta$  pathology, tau may be the main contributor to both increasing NfL levels and structural connectivity loss. In addition, we also found that tau mediates the effects of NfL on global cognition, suggesting that the effects of NfL on cognitive function might not be independent of tau. We note that, in the current study, we did not hypothesize that functional connectivity would be associated with NfL or axonal damage because functional connectivity often occurs between brain areas that are not anatomically connected or linked by axons, as reported in previous studies (Honey et al., 2009; Hermundstad et al., 2013; Park and Friston, 2013; Palmqvist et al., 2017).

Regarding previous literature, our results are in line with a study showing early increases in SNAP25 in presymptomatic stages of familial Alzheimer's disease (Schindler et al., 2019). They also agree with findings in sporadic Alzheimer's disease showing elevated NRGN in amyloid-\u03b3+ non-demented individuals compared to amyloid- $\beta$ - subjects (Portelius et al., 2015; Mattsson et al., 2016), an association between GAP43 and amyloid pathology measured using CSF amyloid- $\beta_{1-42}$  (Sandelius *et al.*, 2019), and finally an association between NRGN with memory (Casaletto et al., 2017) or between NfL and global cognitive decline (Zetterberg et al., 2016; Mattsson et al., 2019). Regarding NfL, although familial Alzheimer's studies suggest that it is an early disease marker (Preische et al., 2019), they have not compared it to synaptic markers within the same sample to establish which changes occur earlier or later over the course of the disease. In fact, previous studies by our group comparing different CSF and plasma markers across the same subjects suggest that the changes in NfL in early disease stages are rather subtle, arguing against NfL being a reliable early marker of Alzheimer's disease (Palmqvist et al., 2019, 2020). To our knowledge, there are no previous studies assessing the relationship between changes in synaptic and axonal CSF markers with measures of tau aggregation into insoluble paired helical filaments captured by tau PET. The studies published so far have mainly used CSF measures of tau (Mattsson et al., 2016; Sutphen et al., 2018; Sandelius et al., 2019), which mostly reflect soluble tau forms that change in earlier stages of Alzheimer's disease at least partly in response to amyloid pathology (Sato et al., 2018; Mattsson-Carlgren et al. 2020). Thus, our study has the advantage of measuring a constellation of synaptic and axonal markers in individuals who also underwent amyloid PET and tau PET, which are thought to reflect the formation of neuritic plaques (Ikonomovic et al., 2008) and tangles (Sato et al., 2018; Smith et al., 2019) that characterize earlier and later stages of Alzheimer's disease (Braak and Braak, 1991).

It should be noted that our cross-sectional design relies on the assumption that amyloid- $\beta$  deposition is followed by tau pathology, which is in line with current models of Alzheimer's disease (Jack et al. 2010, 2013a, 2014, 2018). Future studies assessing longitudinal changes in SNAP25, GAP43, NRGN, NfL in relation to longitudinal amyloid-B PET and tau-PET imaging would be needed to understand their dynamic trajectories over the course of Alzheimer's disease, as described in the previous models (Jack et al., 2010, 2013a, 2014, 2018). In addition, studies assessing the relationship between cognitive resilience and synaptic integrity (Arnold et al., 2013; Boros et al., 2017; Latimer et al., 2019) or synaptic changes in primary age-related tauopathy (PART) (Crary et al., 2014) would also be useful to further understand the implications of synaptic and axonal loss. Finally, note also that we used tau PET imaging instead of CSF tau to assess later disease stages in the current study. This choice was driven by recent evidence showing that CSF tau reflects mainly soluble tau forms, which change in earlier stages of Alzheimer's disease at least partly in response to amyloid pathology, in contrast to tau PET, which most likely reflects the formation of tau into paired helical filaments (Sato et al., 2018; Mattsson-Carlgren et al., 2020), correlating strongly with neurofibrillary tangles (Smith et al., 2019) and being therefore a better late disease marker. To ensure that our results were not driven by a specific threshold to define tau-positivity, we replicated all of our findings using a less stringent cut-off of 1.22, which can be found in Supplementary material.

In summary, the results of our *in vivo* study agree with the hypothesis that amyloid- $\beta$  and tau have toxic effects on synaptic function and axonal integrity, respectively, linking these events over the course of Alzheimer's disease. By combining several established and innovative methods, we show that early disease stages are characterized by amyloidinduced synaptic damage, memory impairment and functional connectivity changes, whereas later disease stages are characterized by tau-associated axonal damage, global cognitive decline and reduced anatomical connectivity. While we recognize that the current techniques do not have the resolution to assess the molecular mechanisms of amyloid- $\beta$  and tau at the single neuron level, these results are consistent with several animal studies and current disease models that place synaptic dysfunction and axonal degeneration as central events in the pathogenesis of Alzheimer's disease.

### Funding

Work at the authors' research centre was supported by the Swedish Research Council, the Knut and Alice Wallenberg foundation, the Marianne and Marcus Wallenberg foundathe Strategic Research Area MultiPark tion. (Multidisciplinary Research in Parkinson's disease) at Lund University, the Swedish Alzheimer Foundation, the Swedish Brain Foundation, Demensfonden, Eivind och Elsa K: son Sylvans stiftelse, Märtha och Gustaf Ågrens stiftelse, Gun och Bertil Stohnes stiftelse, Stiftelsen Gamla Tjänarinnor, The Parkinson foundation of Sweden, The Parkinson Research Foundation, the Skåne University Hospital Foundation, and the Swedish federal government under the ALF agreement. Doses of <sup>18</sup>F-flutemetamol injection were sponsored by GE Healthcare. The precursor of <sup>18</sup>F-flortaucipir was provided by AVID radiopharmaceuticals. J.B.P. is supported by grants from the Swedish Research Council (#2018-02201),Hjärnfonden (#FO2019-0289), Alzheimerfonden (#AF-930827), the Strategic Research Programme in Neuroscience at Karolinska Institutet Startup Grant), The Center for Medical (Stratneuro Innovation (#20200695), Gamla Tjänarinnor (#2019-00803) and Stohnes. K.B. is supported by the Swedish Research Council (#2017-00915), the Alzheimer Drug Discovery Foundation (ADDF), USA (#RDAPB-201809-2016615), the Swedish Alzheimer Foundation (#AF-742881), Hjärnfonden, Sweden (#FO2017-0243), the Swedish state under the agreement between the Swedish government and the County Councils, the ALF-agreement (#ALFGBG-715986), and European Union Joint Program for Neurodegenerative Disorders (JPND2019-466-236). H.Z. is a Wallenberg Scholar supported by grants from the Swedish Research Council (#2018-02532), the European Research Council (#681712), Swedish State Support for Clinical Research (#ALFGBG-720931) and the UK Dementia Research Institute at UCL.

### **Competing interests**

K.B. has served as a consultant or at advisory boards for Abcam, Axon, Biogen, Lilly, MagQu, Novartis and Roche Diagnostics, and is a co-founder of Brain Biomarker Solutions in Gothenburg AB, a GU Venture-based platform company at the University of Gothenburg. H.Z. has served at scientific advisory boards for Denali, Roche Diagnostics, Wave, Samumed and CogRx, has given lectures in symposia sponsored by Fujirebio, Alzecure and Biogen, and is a cofounder of Brain Biomarker Solutions in Gothenburg AB, a GU Ventures-based platform company at the University of Gothenburg. O.H. has acquired research support (for the institution) from Roche, Pfizer, GE Healthcare, Biogen, AVID Radiopharmaceuticals and Euroimmun. In the past 2 years, he has received consultancy/speaker fees (paid to the institution) from Biogen and Roche. All other authors report no competing interests.

### Supplementary material

Supplementary material is available at Brain online.

### References

- Allegra Mascaro AL, Cesare P, Sacconi L, Grasselli G, Mandolesi G, Maco B, et al. In vivo single branch axotomy induces GAP-43-dependent sprouting and synaptic remodeling in cerebellar cortex. Proc Natl Acad Sci USA 2013; 110: 10824–9.
- Almeida CG, Tampellini D, Takahashi RH, Greengard P, Lin MT, Snyder EM, et al. Beta-amyloid accumulation in APP mutant neurons reduces PSD-95 and GluR1 in synapses. Neurobiol Dis 2005; 20: 187–98.
- American Psychiatric Association. Diagnostic and statistical manual of mental disorders: diagnostic and statistical manual of mental disorders. 5th edn. Arlington, VA: American Psychiatric Association; 2013.
- Arnold SE, Louneva N, Cao K, Wang LS, Han LY, Wolk DA, et al. Cellular, synaptic, and biochemical features of resilient cognition in Alzheimer's disease. Neurobiol Aging 2013; 34: 157–68.
- Beaulieu C. The basis of anisotropic water diffusion in the nervous system—a technical review. NMR Biomed 2002; 15: 435–55.
- Beaulieu C, Allen PS. Determinants of anisotropic water diffusion in nerves. Magn Reson Med 1994; 31: 394–400.
- Behrens TE, Berg HJ, Jbabdi S, Rushworth MF, Woolrich MW. Probabilistic diffusion tractography with multiple fibre orientations: what can we gain? Neuroimage 2007; 34: 144–55.
- Biswal BB, Mennes M, Zuo XN, Gohel S, Kelly C, Smith SM, et al. Toward discovery science of human brain function. Proc Natl Acad Sci USA 2010; 107: 4734–9.
- Boros BD, Greathouse KM, Gentry EG, Curtis KA, Birchall EL, Gearing M, et al. Dendritic spines provide cognitive resilience against Alzheimer's disease. Ann Neurol 2017; 82: 602–14.
- Braak H, Braak E. Neuropathological stageing of Alzheimer-related changes. Acta Neuropathol 1991; 82: 239–59.
- Brinkmalm A, Brinkmalm G, Honer WG, Frölich L, Hausner L, Minthon L, et al. SNAP-25 is a promising novel cerebrospinal fluid biomarker for synapse degeneration in Alzheimer's disease. Mol Neurodegeneration 2014; 9: 53.
- Burgold S, Bittner T, Dorostkar MM, Kieser D, Fuhrmann M, Mitteregger G, et al. In vivo multiphoton imaging reveals gradual growth of newborn amyloid plaques over weeks. Acta Neuropathol 2011; 121: 327–35.
- Carlesimo GA, Oscar-Berman M. Memory deficits in Alzheimer's patients: a comprehensive review. Neuropsychol Rev 1992; 3: 119–69.
- Casaletto KB, Elahi FM, Bettcher BM, Neuhaus J, Bendlin BB, Asthana S, et al. Neurogranin, a synaptic protein, is associated with

memory independent of Alzheimer biomarkers. Neurology 2017; 89: 1782-8.

- Cohen RM, Rezai-Zadeh K, Weitz TM, Rentsendorj A, Gate D, Spivak I, et al. A transgenic Alzheimer rat with plaques, tau pathology, behavioral impairment, oligomeric abeta, and frank neuronal loss. J Neurosci 2013; 33: 6245–56.
- Crary JF, Trojanowski JQ, Schneider JA, Abisambra JF, Abner EL, Alafuzoff I, et al. Primary age-related tauopathy (PART): a common pathology associated with human aging. Acta Neuropathol 2014; 128: 755–66.
- Cummings DM, Benway TA, Ho H, Tedoldi A, Fernandes Freitas MM, Shahab L, et al. Neuronal and peripheral pentraxins modify glutamate release and may interact in blood–brain barrier failure. Cereb Cortex 2017; 27: 3437–48.
- DaSilva AF, Tuch DS, Wiegell MR, Hadjikhani N. A primer on diffusion tensor imaging of anatomical substructures. Neurosurg Focus 2003; 15: E4.
- Edwards FA. A unifying hypothesis for Alzheimer's disease: from plaques to neurodegeneration. Trends Neurosci 2019; 42: 310–22.
- Elman JA, Oh H, Madison CM, Baker SL, Vogel JW, Marks SM, et al. Neural compensation in older people with brain amyloid-β deposition. Nat Neurosci 2014; 17: 1316–8.
- Fagiani F, Lanni C, Racchi M, Pascale A, Govoni S. Amyloid-β and synaptic vesicle dynamics: a cacophonic orchestra. J Alzheimers Dis 2019; 72: 1–14.
- Folstein MF, Folstein SE, McHugh PR. "Mini-mental state". A practical method for grading the cognitive state of patients for the clinician. J Psych Res 1975; 12: 189–98.
- Gaetani L, Blennow K, Calabresi P, Di Filippo M, Parnetti L, Zetterberg H. Neurofilament light chain as a biomarker in neurological disorders. J Neurol Neurosurg Psychiatry 2019; 90: 870–81.
- Gouras GK, Relkin NR, Sweeney D, Munoz DG, Mackenzie IR, Gandy S. Increased apolipoprotein E epsilon 4 in epilepsy with senile plaques. Ann Neurol 1997; 41: 402–4.
- Gulisano W, Melone M, Ripoli C, Tropea MR, Puma DD, Giunta S, et al. Neuromodulatory action of picomolar extracellular Amyloidβ42 oligomers on presynaptic and postsynaptic mechanisms underlying synaptic function and memory. J Neurosci 2019; 39: 5986–6000.
- Hahn A, Schain M, Erlandsson M, Sjölin P, James GM, Strandberg OT, et al. Modeling strategies for quantification of in vivo 18F-AV1451 binding in patients with tau pathology. J Nucl Med 2016; 58: 623–31.
- Hardy J, Selkoe DJ. The amyloid hypothesis of Alzheimer's disease: progress and problems on the road to therapeutics. Science 2002; 297: 353–6.
- Hermundstad AM, Bassett DS, Brown KS, Aminoff EM, Clewett D, Freeman S, et al. Structural foundations of resting-state and taskbased functional connectivity in the human brain. Proc Nat Acad Sci USA 2013; 110: 6169–74.
- Honey CJ, Sporns O, Cammoun L, Gigandet X, Thiran JP, Meuli R, et al. Predicting human resting-state functional connectivity from structural connectivity. Proc Nat Acad Sci USA 2009; 106: 2035–40.
- Hong S, Beja-Glasser VF, Nfonoyim BM, Frouin A, Li S, Ramakrishnan S, et al. Complement and microglia mediate early synapse loss in Alzheimer mouse models. Science 2016; 352: 712–6.
- Howlett DR, Bowler K, Soden PE, Riddell D, Davis JB, Richardson JC, et al. Abeta deposition and related pathology in an APP x PS1 transgenic mouse model of Alzheimer's disease. Histol Histopathol 2008; 23: 67–76.
- Hua K, Zhang J, Wakana S, Jiang H, Li X, Reich DS, et al. Tract probability maps in stereotaxic spaces: analyses of white matter anatomy and tract-specific quantification. Neuroimage 2008; 39: 336–47.
- Huijbers W, Mormino EC, Schultz AP, Wigman S, Ward AM, Larvie M, et al. Amyloid-b deposition in mild cognitive impairment is

associated with increased hippocampal activity, atrophy and clinical progression. Brain 2015; 138: 1023–35.

- Ikonomovic MD, Klunk WE, Abrahamson EE, Mathis CA, Price JC, Tsopelas ND, et al. Post-mortem correlates of in vivo PiB-PET amyloid imaging in a typical case of Alzheimer's disease. Brain 2008; 131: 1630–45.
- Jack CR Jr, Bennett DA, Blennow K, Carrillo MC, Dunn B, Haeberlein SB, et al. NIA-AA research framework: toward a biological definition of Alzheimer's disease. Alzh & Dem 2018; 14: 535–62.
- Jack CR Jr, Knopman DS, Jagust WJ, Petersen RC, Weiner MW, Aisen PS. Tracking pathophysiological processes in Alzheimer's disease: an updated hypothetical model of dynamic biomarkers. Lancet Neurol 2013a; 12: 207–16.
- Jack CR Jr, Knopman DS, Jagust WJ, Shaw LM, Aisen PS, Weiner MW, et al. Hypothetical model of dynamic biomarkers of the Alzheimer's pathological cascade. Lancet Neurol 2010; 9: 119–28.
- Jack CR, Wiste HJ, Lesnick TG, Weigand SD, Knopman DS, Vemuri P, et al. Brain β-amyloid load approaches a plateau. Neurology 2013b; 80: 890–6.
- Jack CR Jr, Wiste HJ, Weigand SD, Rocca WA, Knopman DS, Mielke MM, et al. Age-specific population frequencies of cerebral β-amyloidosis and neurodegeneration among people with normal cognitive function aged 50–89 years: a cross-sectional study. Lancet Neurol 2014; 13: 997–1005.
- Jack CR Jr, Wiste HJ, Weigand SD, Therneau TM, Lowe VJ, Knopman DS, et al. Defining imaging biomarker cut points for brain aging and Alzheimer's disease. Alzh & Dem 2017; 13: 205–16.
- Jones DT, Knopman DS, Gunter JL, Graff-Radford J, Vemuri P, Boeve BF, et al. Cascading network failure across the Alzheimer's disease spectrum. Brain 2016; 139: 547–62.
- Jürgensen S, Ferreira ST. Nicotinic receptors, amyloid-β, and synaptic failure in Alzheimer's disease. J Mol Neurosci 2010; 40: 221–9.
- Kadir A, Almkvist O, Forsberg A, Wall A, Engler H, Långström B, et al. Dynamic changes in PET amyloid and FDG imaging at different stages of Alzheimer's disease. Neurobiol Aging 2012; 33: 198–e1.
- Koffie RM, Meyer-Luehmann M, Hashimoto T, Adams KW, Mielke ML, Garcia-Alloza M, et al. Oligomeric amyloid beta associates with postsynaptic densities and correlates with excitatory synapse loss near senile plaques. Proc Natl Acad Sci USA 2009; 106: 4012–7.
- Koo E H, Park L, Selkoe D J.Amyloid beta-protein as a substrate interacts with extracellular matrix to promote neurite outgrowth. Proceedings of the National Academy of Sciences 1993; 90: 4748–52.
- Kvartsberg H, Lashley T, Murray CE, Brinkmalm G, Cullen NC, Höglund K, et al. The intact postsynaptic protein neurogranin is reduced in brain tissue from patients with familial and sporadic Alzheimer's disease. Acta Neuropathol 2019; 137: 89–102.
- Landau SM, Fero A, Baker SL, Koeppe R, Mintun M, Chen K, et al. Measurement of longitudinal β-amyloid change with 18F-florbetapir PET and standardized uptake value ratios. J Nucl Med 2015; 56: 567.
- Latimer CS, Burke BT, Liachko NF, Currey HN, Kilgore MD, Gibbons LE, et al. Resistance and resilience to Alzheimer's disease pathology are associated with reduced cortical pTau and absence of limbic-predominant age-related TDP-43 encephalopathy in a community-based cohort. Acta Neuropathol Comm 2019; 7: 91.
- Ledbetter MC, Porter KR. A microtubule in plant cell fine structure. J Cell Biol 1963; 19: 239–50.
- Mattsson-Carlgren N, Andersson E, Janelidze S, Ossenkoppele R, Insel P, Strandberg O, et al. Aβ deposition is associated with increases in soluble and phosphorylated tau that precede a positive Tau PET in Alzheimer's disease. Sci Adv 2020; 6: eaaz2387.
- Mattsson N, Cullen NC, Andreasson U, Zetterberg H, Blennow K. Association between longitudinal plasma neurofilament light and neurodegeneration in patients with Alzheimer disease. JAMA Neurol 2019; 76: 791–9.

- Mattsson N, Insel PS, Palmqvist S, Portelius E, Zetterberg H, Weiner M, et al. Alzheimer's disease neuroimaging initiative. Cerebrospinal fluid tau, neurogranin, and neurofilament light in Alzheimer's disease. EMBO Mol Med 2016; 8: 1184–96.
- Nelson PT, Alafuzoff I, Bigio EH, Bouras C, Braak H, Cairns NJ, et al. Correlation of Alzheimer disease neuropathologic changes with cognitive status: a review of the literature. J Neuropathol Exp Neurol 2012; 71: 362–81.
- Ossenkoppele R, Rabinovici GD, Smith R, Cho H, Schöll M, Strandberg O, et al. Discriminative accuracy of [18F] flortaucipir positron emission tomography for Alzheimer disease vs other neurodegenerative disorders. JAMA 2018; 320: 1151–62.
- Palmqvist S, Insel P S, Stomrud E, Janelidze S, Zetterberg H, Brix B, et al. Cerebrospinal fluid and plasma biomarker trajectories with increasing amyloid deposition in Alzheimer's disease. EMBO Mol Med 2019; 11: e11170.
- Palmqvist S, Janelidze S, Quiroz YT, Zetterberg H, Lopera F, Stomrub E, et al. Discriminative accuracy of plasma phospho-tau217 for alzheimer disease vs other neurodegenerative disorders. JAMA 2020; 324: 772–81.
- Palmqvist S, Mattsson N, Hansson O. Alzheimer's disease neuroimaging initiative. Cerebrospinal fluid analysis detects cerebral amyloidbeta accumulation earlier than positron emission tomography. Brain 2016; 139: 1226–36.
- Palmqvist S, Schöll M, Strandberg O, Mattsson N, Stomrud E, Zetterberg H, et al. Earliest accumulation of β-amyloid occurs within the default-mode network and concurrently affects brain connectivity. Nat Comm 2017; 8: 1–3.
- Palop JJ, Mucke L. Amyloid-β-induced neuronal dysfunction in Alzheimer's disease: from synapses toward neural networks. Nat Neurosci 2010; 13: 812–8.
- Park HJ, Friston K. Structural and functional brain networks: from connections to cognition. Science 2013; 342: 1238411.
- Pievani M, de Haan W, Wu T, Seeley WW, Frisoni GB. Functional network disruption in the degenerative dementias. Lancet Neurol 2011; 10: 829–43.
- Pooler AM, Polydoro M, Wegmann S, Nicholls SB, Spires-Jones TL, Hyman BT. Propagation of tau pathology in Alzheimer's disease: identification of novel therapeutic targets. Alzh Res Ther 2013; 5: 1–8.
- Portelius E, Zetterberg H, Skillbäck T, Törnqvist U, Andreasson U, Trojanowski JQ, et al. Cerebrospinal fluid neurogranin: relation to cognition and neurodegeneration in Alzheimer's disease. Brain 2015; 138: 3373–85.
- Potter PE, Rauschkolb PK, Pandya Y, Sue LI, Sabbagh MN, Walker DG, et al. Pre-and post-synaptic cortical cholinergic deficits are proportional to amyloid plaque presence and density at preclinical stages of Alzheimer's disease. Acta Neuropathol 2011; 122: 49–60.
- Preische O, Schultz SA, Apel A, Kuhle J, Kaeser SA, Barro C, et al. Serum neurofilament dynamics predicts neurodegeneration and clinical progression in presymptomatic Alzheimer's disease. Nat Med 2019; 25: 277–83.
- Priller C, Bauer T, Mitteregger G, Krebs B, Kretzschmar HA, Herms J. Synapse formation and function is modulated by the amyloid precursor protein. J Neurosci 2006; 26: 7212–21.
- Reddy PH, Beal MF. Amyloid beta, mitochondrial dysfunction and synaptic damage: implications for cognitive decline in aging and Alzheimer's disease. Trends Mol Med 2008; 14: 45–53.
- Rosen WG, Mohs RC, Davis KL. A new rating scale for Alzheimer's disease. Am J Psychiatry 1984; 141: 1356–64.
- Roselli F, Tirard M, Lu J, Hutzler P, Lamberti P, Livrea P, et al. Soluble  $\beta$ -amyloid1-40 induces NMDA-dependent degradation of postsynaptic density-95 at glutamatergic synapses. J Neurosci 2005; 25: 11061–70.
- Rozkalne A, Hyman BT, Spires-Jones TL. Calcineurin inhibition with FK506 ameliorates dendritic spine density deficits in plaque-bearing Alzheimer model mice. Neurobiol Dis 2011; 41: 650–4.
- Sandelius Å, Portelius E, Källén Å, Zetterberg H, Rot U, Olsson B, et al. Elevated CSF GAP-43 is Alzheimer's disease specific and

associated with tau and amyloid pathology. Alzheimers Dement 2019; 15: 55-64.

- Sato C, Barthélemy NR, Mawuenyega KG, Patterson BW, Gordon BA, Jockel-Balsarotti J, et al. Tau kinetics in neurons and the human central nervous system. Neuron 2018; 97: 1284–98.
- Schindler SE, Li Y, Todd KW, Herries EM, Henson RL, Gray JD, et al. Emerging cerebrospinal fluid biomarkers in autosomal dominant Alzheimer's disease. Alzheimers Dement 2019; 15: 655–65.
- Sepulcre J, Sabuncu MR, Li Q, El Fakhri G, Sperling R, Johnson KA. Tau and amyloid  $\beta$  proteins distinctively associate to functional network changes in the aging brain. Alzheimers Dement 2017; 13: 1261–9.
- Sheline YI, Raichle ME. Resting state functional connectivity in preclinical Alzheimer's disease. Biol Psychiatry 2013; 74: 340–7.
- Shemer I, Holmgren C, Min R, Fülöp L, Zilberter M, Sousa KM, et al. Non-fibrillar β-amyloid abates spike-timing-dependent synaptic potentiation at excitatory synapses in layer 2/3 of the neocortex by targeting postsynaptic AMPA receptors. Eur J Neurosci 2006; 23: 2035–47.
- Smith R, Wibom M, Pawlik D, Englund E, Hansson O. Correlation of in vivo [18F] Flortaucipir with postmortem Alzheimer disease tau pathology. JAMA Neurol 2019; 76: 310–7.
- Snyder EM, Nong Y, Almeida CG, Paul S, Moran T, Choi EY, et al. Regulation of NMDA receptor trafficking by amyloid-β. Nat Neurosci 2005; 8: 1051–8.
- Spires-Jones TL, Hyman BT. The intersection of amyloid beta and tau at synapses in Alzheimer's disease. Neuron 2014; 82: 756–71.
- Spires-Jones TL, Stoothoff WH, de Calignon A, Jones PB, Hyman BT. Tau pathophysiology in neurodegeneration: a tangled issue. Trends Neurosci 2009; 32: 150–9.
- Steinbach JP, Müller U, Leist M, Li ZW, Nicotera P, Aguzzi A. Hypersensitivity to seizures in β-amyloid precursor protein deficient mice. Cell Death Differ 1998; 5: 858–66.

- Sutphen CL, McCue L, Herries EM, Xiong C, Ladenson JH, Holtzman DM, et al. Longitudinal decreases in multiple cerebrospinal fluid biomarkers of neuronal injury in symptomatic late onset Alzheimer's disease. Alzheimers Dement 2018; 14: 869–79.
- Tai HC, Serrano-Pozo A, Hashimoto T, Frosch MP, Spires-Jones TL, Hyman BT. The synaptic accumulation of hyperphosphorylated tau oligomers in Alzheimer disease is associated with dysfunction of the ubiquitin- proteasome system. Am. J Pathol 2012; 181: 1426–35.
- Wu HY, Hudry E, Hashimoto T, Kuchibhotla K, Rozkalne A, Fan Z, et al. Amyloid beta induces the morphological neurodegenerative triad of spine loss, dendritic simplification, and neuritic dystrophies through calcineurin activation. J Neurosci 2010; 30: 2636–49.
- Zempel H, Luedtke J, Kumar Y, Biernat J, Dawson H, Mandelkow E, et al. Amyloid-b oligomers induce synaptic damage via Tau-dependent microtubule severing by TTLL6 and spastin. EMBO J 2013; 32: 2920–37.
- Zempel H, Thies E, Mandelkow E, Mandelkow E-M. Abeta oligomers cause localized Ca(2+) elevation, missorting of endogenous Tau into dendrites, Tau phosphorylation, and destruction of microtubules and spines. J Neurosci 2010; 30: 11938–50.
- Zetterberg H, Blennow K. Neurogranin levels in cerebrospinal fluid a new addition to the Alzheimer disease diagnostic toolbox. JAMA Neurol 2015; 72: 1237–8.
- Zetterberg H, Skillbäck T, Mattsson N, Trojanowski JQ, Portelius E, Shaw LM, et al. Association of cerebrospinal fluid neurofilament light concentration with Alzheimer disease progression. JAMA Neurol 2016; 73: 60–7.
- Zhong L, Gerges NZ. Neurogranin and synaptic plasticity balance. Commun Integr Biol 2010; 3: 340–2.