

Review Article

Traditional Uses, Chemical Constituents, and Biological Activities of *Bixa orellana* L.: A Review

Daniela de Araújo Vilar,¹ Marina Suênia de Araujo Vilar,¹
Túlio Flávio Accioly de Lima e Moura,² Fernanda Nervo Raffin,² Márcia Rosa de Oliveira,³
Camilo Flamarion de Oliveira Franco,⁴ Petrônio Filgueiras de Athayde-Filho,⁵
Margareth de Fátima Formiga Melo Diniz,¹ and José Maria Barbosa-Filho¹

¹ Laboratory of Pharmaceutical Technology, Federal University of Paraíba, 58051-900 João Pessoa, PB, Brazil

² Department of Pharmacy, Federal University of Rio Grande do Norte, 59010-180 Natal, RN, Brazil

³ Department of Molecular Biology, Federal University of Paraíba, 58051-900 João Pessoa, PB, Brazil

⁴ State Company of Agricultural Research of Paraíba, Rua Eurípedes Tavares 210, Tambiá, 58013-290 João Pessoa, PB, Brazil

⁵ Department of Chemistry, Federal University of Paraíba, 58051-900 João Pessoa, PB, Brazil

Correspondence should be addressed to José Maria Barbosa-Filho; jbarbosa@ltf.ufpb.br

Received 11 February 2014; Revised 17 May 2014; Accepted 31 May 2014; Published 23 June 2014

Academic Editor: Tsutomu Hatano

Copyright © 2014 Daniela de Araújo Vilar et al. This is an open access article distributed under the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Bixa orellana L., popularly known as “urucum,” has been used by indigenous communities in Brazil and other tropical countries for several biological applications, which indicates its potential use as an active ingredient in pharmaceutical products. The aim of this work was to report the main evidence found in the literature, concerning the ethnopharmacology, the biological activity, and the phytochemistry studies related to *Bixa orellana* L. Therefore, this work comprises a systematic review about the use of *Bixa orellana* in the American continent and analysis of the data collected. This study shows the well-characterized pharmacological actions that may be considered relevant for the future development of an innovative therapeutic agent.

1. Introduction

The use of natural compounds of mineral, animal, or plant origin in food products, cosmetics, and drugs began long ago. There are written records of ancient Egyptian and Chinese civilizations that have made use of these products. Nowadays, there has been a return to the search for products called “natural,” which in fact never ceased to exist. The analysis of the composition of many drugs shows that almost 50% of those in clinical use are derived from natural compounds. Furthermore, not only plants but also plant byproducts are widely used as preservatives and flavoring and coloring agents in various food and cosmetic preparations [1].

Bixa orellana is a plant native to Brazil but grows in other regions of South and Central America. It is grown in tropical countries such as Peru, Mexico, Ecuador, Indonesia, India, Kenya, and East Africa [2].

The seeds of this plant produce one of the dyes most frequently used worldwide, not only in food products but also in the textile, paint, and cosmetic industries. Its use has been stimulated due to the ban on the use of synthetic dyes in food and cosmetics, where it is one of the few accepted by the World Health Organization (WHO), since, in addition to being nontoxic, it does not seem to change the food value [3]. Another interesting fact is that 70% of all natural coloring agents consumed worldwide are derived from annatto [4].

Annatto first spread in the form of food coloring, also known as paprika, a condiment widely used in cooking to enhance the color of food. Today, however, its use has spread into many segments of industrial production. Thus, it is now applied on the skin—in the form of makeup and sunscreen—and there is research proving that its use brings health benefits, which makes producers thankful for cultivating it [5, 6]. Therefore, in the continuation of our research on

bioactive molecules from various species of different plant families [7–22], we offer this compilation of the traditional uses, chemical constituents, and biological activities of *Bixa orellana*.

The aim of this review is to highlight the biological and phytochemical studies that have been published about *Bixa orellana* in South and Central America and try to correlate these studies with the popular uses of this plant in those regions, as well as to evaluate whether its chemical composition can support the reported biomedical properties related to *Bixa orellana*.

2. Materials and Methods

In this work, the biological activities and compounds isolated from *Bixa orellana* were searched using the database of the Web of Science, Scielo, and the University of Illinois in Chicago NAPRALERT (acronym for “NATURALProducts ALERT”). The data were updated in April 2014, using “*Bixa orellana*, chemical, and bixin” as keywords for this review. The references found in the survey were later consulted for details about the models or mechanisms of bioassays used to test the extracts of *Bixa orellana*.

3. Botanical

The annatto tree belongs to the family Bixaceae and the genus *Bixa*. Despite the existence of several species, the most common in our country is *Bixa orellana* L., named after Francisco Orellana, who was the first European to navigate the Amazon [23].

According to Revilla [58], *B. orellana* is a small tree or shrub measuring from 3 to 5 meters in height, sometimes reaching a height of 10 meters. The trunk is short, measuring 20–30 cm in diameter, with dark gray bark with lenticels in vertical rows. The leaves are alternate, 10 to 20 cm long and 5 to 10 cm wide, sharp, green on both sides, and with extended petioles.

According to Oliveira et al. [59], seeds measure 0.3–0.5 cm in length and 0.2–0.3 cm in diameter, and their shape varies from pyramidal to almost conical. The number of seeds per capsule varies according to the author: Alonso [60] found that each bivalvar capsule may contain from 30 to 60 seeds, on average.

The seeds are considered the plant part of commercial importance, since the pericarp (layer that surrounds the seeds) contains the pigments that have wide industrial application. About 80% of this pigment is the carotenoid known as bixin, which has the dye property and can be extracted with vegetable oils or chemical bases. Depending on the cultivar and climatic conditions of the region, the bixin content can vary from 1 to 6% in the seed aril. The remainder is composed of other dyes and inert substances of minor importance [61].

4. Use in Traditional Medicine

Annatto is a native plant from South America, more specifically of the Amazon region. The popular name “urucum”

comes from the Tupi word “ru-ku,” which means “red.” In Brazil, this plant is commonly known as urucum, urucu, açafrao, açafroa, and açafroeira-da-terra. It is known by other popular names in other countries: atolé, achiote, and bija (Peru and Cuba); axiote (Mexico); achiote, achote, anatto, bija, and santo-domingo (Puerto Rico); bixa (Guyana); analto (Honduras); guajachote (El Salvador); onotto and onotillo (Venezuela); achiote and urucu (Bolivia); urucu (Argentina); roucou (Trinidad); roucou and koessewee (Suriname); and annatto (United States). The wide dissemination of its use in those regions is related to the growing demand for natural dyes by many pharmaceutical, cosmetic, textile, and especially food industries [57].

According to Córrea [27], seeds urucum supplies seeds that have been used as a condiment as well as laxative, cardiogenic, hypotensive, expectorant, and antibiotic. In addition, it has anti-inflammatory activity for bruises and wounds and has been used for the treatment of bronchitis and for wound healing purposes. Oil is also obtained from this plant. The infusion of the leaves has been shown to be effective against bronchitis, sore throat, and eye inflammation. The pulp, which includes the seed, is used for soft drinks and febrifuge. Moreover, it can provide valuable dyeing materials such as yellow (orellin) and red (bixin) substances, with the latter constituting a crystallized active ingredient.

In the food industry, it is used to color butter, margarine, mayonnaise, sauces, mustard, sausage, soup, juice, ice cream, bakery products, macaroni, and cheese, where it is commonly called “do reino” (of the kingdom), coming from Holland. It is also widely used in the printing industry and dye manufacturing. Many Aborigines use annatto for dyeing, where the dye is naturally obtained as a mixture and used to color ceramics and other vases for domestic use. In addition, most endogenous people use this dye on their skin to beautify themselves during religious rituals and mainly to protect themselves from ultraviolet radiation and from mosquitoes that infest forests [49]. The bast provides fibers for rough cordage, and the powder resulting from grinding the seeds has been used as an aphrodisiac. Finally, the infusion of cold buds serves to wash inflamed eyes, whereas the decoction of the leaves has been used for antiemetic therapy during pregnancy [27] (Table 1).

Thus, despite the different culture and traditions among the countries in South and Central America, several of the popular uses of *Bixa orellana* are the same, for example, antipyretic, aphrodisiac, antidiarrheal, antidiabetic, and insect repellent.

5. Chemical Compounds

Bixin, a red-colored carotenoid, is the pigment present in high concentration in the annatto seed aril. It is the main substance responsible for the dyeing characteristics of seeds, where its concentration can be as high as 5.0%. However, different seeds may have levels less than 2.0%, and because their commercial value is based on the bixin percentage, levels higher than 2.5% are usually required for export [61].

TABLE 1: Traditional uses of annatto in American countries.

Country/use	Plant part	References
Argentina		
Antipyretic/cardiogenic/antidiarrheal	Seeds	[24]
Antidiarrheal/dyes/condiment	Seeds	[25]
Brazil		
Body paint	Seeds	[26]
Insect repellent	Seeds	[27]
Condiment/food coloring	Seeds	[27]
Antipyretic	Seeds	[26]
Antipyretic/laxatives/burns	Seeds	[28]
Malaria	Seeds	[26]
Colombia		
Snakebite	Leaves	[29]
Aphrodisiac	Seeds	[30]
Cuba		
Aphrodisiac	Seeds	[31]
Guatemala		
Gonorrhea/dysentery	Leaves	[32]
Hepatitis	Leaves	[33]
Dysentery	Leaves	[34]
Blood diseases	Leaves	[35]
Gonorrhea	Roots	[32]
Diabetes	Roots	[36]
Honduras		
Aromatic/food coloring	Seeds	[37]
Pain/digestive/dysentery	Leaves	[38]
Jamaica		
Diabetes	Seeds	[39]
Nicaragua		
Respiratory and pulmonary disorders/diarrhea/diuretic/burns	Leaves + seeds	[40]
Labor pains	Seeds	[41]
Cough/cold/diuretic/diarrhea/burns/labor pains	Seeds	[40]
Paraguay		
Insecticide/repellent	Seeds	[42]
Diabetes	Seeds	[39]
Peru		
Aphrodisiac/aphrodisiac/diuretic/antidysentery/astringent	Fruits	[34]
Antipyretic/skin problems	Leaves	[33]
Alcoholic hepatitis/worms	Roots	[33]
Antipyretic/aphrodisiac/dysentery/astringent/stomach	Seeds	[34]
Trinidad and Tobago		
Diuretic	Leaves	[43]
Diabetes	Roots	[44]
Diabetes	Roots	[45]

Bixin was isolated for the first time from the seeds of *Bixa orellana* in 1875 and in 1961 its complete chemical structure and stereochemistry were determined by ^1H and ^{13}C -NMR. Bixin belongs to the small class of natural apocarotenoids, whose formation occurs by the oxidative degradation of C40 carotenoids (Table 2).

Bixin consists of a chain of 25 carbons and has the molecular formula $\text{C}_{25}\text{H}_{30}\text{O}_4$ (MW = 394.51). It has a carboxylic acid and methyl ester group at the ends of the chain. Bixin occurs in nature as 16-*Z* (*cis*), but during the extraction process it isomerizes resulting in the 16-*E* form (*trans*), which is called isobixin (Figure 1).

TABLE 2: The main carotenoids from the seeds of *Bixa orellana*.

Carotenoid	Country of isolation	Physical aspect	References
Apo- ψ -carotene, 9'Z-6'-ol	Brazil	Oil	[46]
Beta carotene	Brazil	183°C	[47]
	Suriname	*	[48]
	Brazil	198°C	[49–51]
	Peru	*	[52]
Bixin	Dominican Rep.	*	[53]
	Suriname	*	[48]
	USA	*	[54]
Z-Carotene	Brazil	Oil	[46]
Cryptoxanthin	Suriname	173°C	[48]
Dimethyl-(9Z)-6,6'-diapocarotene-6,6'-dioate	Brazil	Oil	[52]
Dimethyl-(9Z,9'Z)-6,6'-diapocarotene-6,6'-dioate	Brazil	Oil	[50]
Phytoene	Brazil	Oil	[46]
Phytofluene	Brazil	Oil	[46]
Geranylgeraniol	Brazil	Oil	[52]
Lutein	Suriname	196°C	[48]
Methyl-(9Z)-10'-oxo-6,10'-diapocarotene-6-oate	Brazil	Oil	[50]
Methyl-(9Z)-6'-oxo-6,5'-diapocarotene-6-oate	Brazil	Oil	[50]
Methyl-(9Z)-8'-oxo-6,8'-diapocarotene-6-oate	Brazil	Oil	[50]
Methyl-(9'Z)-apo-6'-lycopenoate	Brazil	Oil	[46]
Methyl-(7Z,9Z,9'Z)-apo-6'-lycopenoate	Brazil	Oil	[55]
Methyl-(9Z)-apo-8'-lycopenoate	Brazil	Oil	[46]
Methyl-(all-E)-apo-8'-lycopenoate	Brazil	Oil	[46]
Neurosporene	Brazil	Oil	[46]
Norbixin	Brazil	300°C	[52]
Trans-bixin	Jamaica	195°C	[56]
Zeaxanthin	Suriname	215°C	[48]

Source: [57].

Many other carotenoids (C_{19} , C_{22} , C_{24} , C_{25} , C_{30} , and C_{32}) occur in *Bixa orellana* but constitute a minor percentage of the pigments. The major oily constituent of annatto seeds is geranylgeraniol, representing 1% of dry seeds. Norbixin (Figure 1) is a demethylated derivative of bixin and although it is a naturally occurring compound, it is almost always referred to as a saponification product of bixin. This is the form used for commercial purposes [62].

Currently, more than two dozen substances have been isolated from the seeds of *Bixa orellana*. Besides bixin and norbixin, other compounds such as isobixin, beta-carotene, cryptoxanthin, lutein, zeaxanthin, orellin, bixein, bixol, crocetin, ishwarane, ellagic acid, salicylic acid, threonine, tomentosic acid, tryptophan, and phenylalanine have been found in the seeds of annatto. In addition, the following compounds, in their respective concentrations, are found in these seeds: 40 to 45% cellulose, 3.5 to 5.5% sugars, 0.3 to 0.9% essential oils, 3% fixed oils, 1.0 to 4.5% pigments, and 13 to 16 % proteins and *alpha* and *beta*-carotene, as well as tannins and saponins [63, 64].

Mercadante et al. [50, 65] isolated eight apocarotenoids from annatto seeds: methyl (7Z, 9Z, 9'Z)-apo-6'-lycopenoate, methyl (9Z)-apo-8'-lycopenoate, methyl 1(all-E)-apo-8'-lycopenoate, methyl (all-E)-8-apo-beta-carotene-8'-oate,

methyl (all-E)-apo-6'-lycopenoate, 6-geranylgeranyl-8'-methyl-6,8'-diapocaroten-6-8'-dioate, 6'-geranylgeranyl-6'-methyl-(9Z)-6,6'-diapocaroten-6-6'-dioate, and 6-geranylgeranyl-6'-methyl-6-6'-diapocaroten-6-6'-dioate.

More than 100 volatile compounds have been detected in aqueous and organic extracts, where 50 of these have already been identified (e.g., bornyl acetate, α -caryophyllene, copaene, α -cubebene, (+)-cyclosativene, geranyl phenylacetate, 1-heptanetriol, 3-methylpyridine, 4-methylpyridine γ -elemene, β -humulene, isolekene, β -pinene, seline-6-en-4-ol, δ -selinene, (-)-spathulenol, and (+)-ylangene) [66].

Because annatto is a rich source of carotenoids it is of great commercial importance. In fact, the therapeutic properties of annatto (e.g., antioxidant and hypoglycemic) have been attributed to its high levels of carotenoids [67–69]. Table 2 lists some of these compounds.

The pigments in annatto seeds can be extracted by mechanical processes through grinding the seeds and by physical-chemical methods using solvents or enzymes [46]. The solvent extraction can be performed using three basic methods: alkaline extraction (NaOH or KOH solutions), which results in the conversion of bixin to norbixin; extraction with oil (soybean, corn); and extraction using organic solvents (hexane, chloroform, ethanol, acetone, or

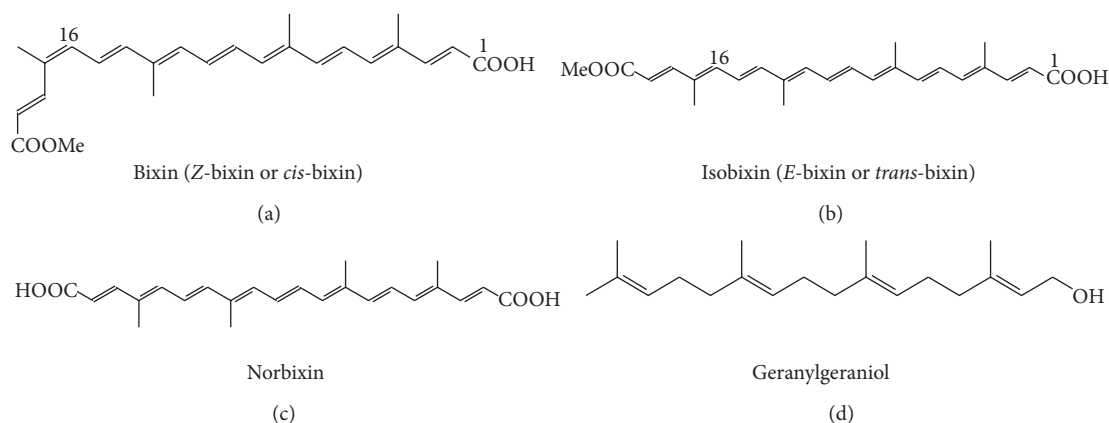


FIGURE 1: Chemical structure of some pigments of annatto.

propylene glycol), which results in the purest form of pigments.

Barbosa-Filho et al. [49] studied the seeds of four types of annatto cultivated in Paraíba State, Brazil, namely, “cascaverde” (“green peel”), “cascavermelha” (“red bark”), “bico de calango” (“lizard beak”), and “grão preto” (“black grain”), with respect to their oil (material extracted with hexane) and solid (material extracted with chloroform) contents, and also pure bixin, which was obtained by successive recrystallization from the chloroform fraction. Pure bixin appears as red-purple crystals with a melting point of 196–198°C. The different concentrations found for the oil fraction, chloroform extract, and bixin are as follows: red bark 5.8%, lizard beak 5.1%, green peel 4.9%, and black grain 4.6%. Red bark shows the highest yield for both solvent fractions, and the bixin amount is around 1%. This species has been reported as the most used in folk medicine. On the other hand, black grain shows negligible amounts of bixin.

6. Biological Activity

Table 3 shows data found in 38 studies performed with annatto in 15 different countries in the American countries. To obtain the extracts and fractions tested, several plant parts were used, such as leaf, root, seed, shoot, and even the whole plant. The data surveyed were classified according to the pharmacological activity tested.

Among the twenty-one activities tested, those with the largest number of studies performed were antifungal activity (12), antibacterial activity (12), antimalarial activity (6), and mutagenic activity (3). Cytotoxic activity and toxicity have been little studied, with three and two studies, respectively. Pharmacological activities have been evaluated in animal models (22 preclinical studies), human models (1 clinical study), cell cultures (2 studies), and *in vitro* tests (32 studies).

Antifungal activity has been investigated in one country in Central America (Guatemala) and in two countries in South America (Ecuador and Argentina) using eleven different fungal strains [70, 82, 84].

Freixa et al. [82] conducted a study in Ecuador to assess the antifungal activity of extracts from the dried leaves of

the annatto tree in response to 7 fungi species, obtaining satisfactory antifungal activity against *Trichophyton mentagrophytes* strains. In Guatemala, three different strains were used to evaluate antifungal activity, with no satisfactory activity being observed [84].

The extracts of annatto leaves have been evaluated for antibacterial activity against 8 different bacterial strains (*Bacillus subtilis*, *Escherichia coli*, *Micrococcus luteus*, *Pseudomonas aeruginosa*, *Staphylococcus aureus*, *Salmonella typhi*, *Shigella dysenteriae*, and *Staphylococcus epidermidis*), showing no activity.

Antimalarial activity has been determined against *Plasmodium gallinaceum*, *Plasmodium lophurae*, and *Plasmodium berghei*. Although the studies conducted previously in the United States did not show significant results [92], Valdés et al. (2011) reported a moderate activity of the seed extracts of *Bixa orellana* against *Plasmodium berghei* and *falciparum*.

7. Mutagenic and Cytotoxic Activities

No significant effect was observed when extracts of annatto seeds were tested for mutagenic activity in studies performed in the United States and Brazil [67, 93].

Extracts obtained from annatto seeds and leaves have been tested in cell cultures and the brine shrimp assay, respectively, and have been found to lack cytotoxicity in either model used. These experiments were carried out in Guatemala and the Dominican Republic [83, 89].

On the other hand, a study performed in Cuba with 10 medicinal plants that were active in inhibiting human lung carcinoma cell growth showed that the ethanolic extract of *Bixa orellana* presented cytotoxicity at concentrations below 100 µg/mL [81].

8. Toxicological Activities

Currently, concerns about the effect of synthetic dyes on human health are incontestable, making people increasingly choose those of natural origin, believing that they are devoid of toxic effects. This is not entirely true, because even a medication from a natural source can be a poison, depending

TABLE 3: Biological activities of extracts of annatto in American countries.

Country	biological activity	Part used	Type of extract	Organism tested	Model tested	Dose used	Activity	References
Argentina	Antibacterial	LE	EtOH	<i>Bacillus subtilis</i> <i>Escherichia coli</i> <i>Micrococcus luteus</i> <i>Pseudomonas aeruginosa</i> <i>Staphylococcus aureus</i>	<i>In vitro</i> <i>In vitro</i> <i>In vitro</i> <i>In vitro</i> <i>In vitro</i>	5 mg/mL 5 mg/mL 5 mg/mL 5 mg/mL 5 mg/mL	Inactive Inactive Inactive Inactive Inactive	[70] [70] [70] [70] [70]
	Antifungal	LE	EtOH	<i>Aspergillus niger</i> <i>Candida albicans</i>	<i>In vitro</i> <i>In vitro</i>	5 mg/mL 5 mg/mL	Inactive Inactive	[70] [70]
	Antiviral	SE	EtOH	Virus <i>Herpes simplex 1</i>	Cell culture	0.78 mg/mL	Inactive	[71]
	Insecticidal	AP	MeOH	Insect	<i>Sitophilus oryzae</i>	5%	Inactive	[71]
Brazil	Antimalarial	SE	CHCl ₃	Mouse	<i>Plasmodium berghei</i>	100 mg/kg	Inactive	[26]
	Antioxidant	SE	EtOH	<i>In vitro</i>	DPPH assay	0.1 g L ⁻¹	Active	[6]
	Insect repellent	SE	Petr. eth	Hamster	<i>Lutzomyia longipalpis</i>	1 g/L	Active	[72]
		SE	EtOH	Mosquito	<i>Aedes aegypti</i>	18.2 mg/mL	Active	[6]
	Molluscicidal	SE	EtOH	Conch	<i>Biomphalaria glabrata</i>	10,000 ppm	Inactive	[73]
	Mutagenic	SE	Powder	Mouse	Bone marrow cells	10,670 ppm	Inactive	[67]
	Toxicity	SE	Powder	Rat	<i>In vivo</i>	500 mg/kg	Inactive	[74]
	Antileishmanial	LE/RO	EtOH	<i>Leishmania amazonensis</i>	<i>In vitro</i>	0.12-2.5 mg/mL	Active	[75]
	Antileishmanial	SE	OE	<i>Leishmania amazonensis</i>	<i>In vitro</i>	10/50/100/500/1000 µg/mL	Active	[76]
	Hyperlipidemia	SE	H ₂ O	Mouse	<i>In vitro</i>	400 and 800 mg/kg	Active	[77]
Colombia	Snakebite	LE	EtOH	Mouse	<i>Bothrops atrox</i>	LD50 > 260 µg/animal	Active	[29]
Costa Rica	Anti-inflammatory	RO	EtOH	Rat	Paw edema/carrageenan	100 mg/kg	Inactive	[78]
Cuba	Positive inotropic effect	AP	H ₂ O	Guinea pig	Isolated atrium	320 µL	Inactive	[79]
	Antimalarial	SE	EtOH	<i>Plasmodium gallinaceum/falciparum</i>	<i>In vivo/in vitro</i>	500 mg/kg	Active	[80]
	Cytotoxic	SE	EtOH	Tumor cells	<i>In vitro</i>	3,9-250 mg/mL	Active	[81]
Ecuador	Antifungal	LE	MeOH	<i>Aspergillus niger</i> <i>Candida albicans</i> <i>Cryptococcus neoformans</i> <i>Fusarium oxysporum</i> <i>Neurospora crassa</i> <i>Penicillium purpurogenum</i> <i>Trichophyton mentagrophytes</i>	<i>In vitro</i> <i>In vitro</i> <i>In vitro</i> <i>In vitro</i> <i>In vitro</i> <i>In vitro</i> <i>In vitro</i>	10 mg/disk 10 mg/disk 10 mg/disk 10 mg/disk 10 mg/disk 10 mg/disk 10 mg/disk	Inactive Inactive Inactive Inactive Inactive Inactive Active	[82] [82] [82] [82] [82] [82] [82]

TABLE 3: Continued.

Country	biological activity	Part used	Type of extract	Organism tested	Model tested	Dose used	Activity	References
Guatemala								
	Antibacterial	LE	Various	<i>Escherichia coli</i> <i>Pseudomonas aeruginosa</i> <i>Salmonella typhi</i> <i>Shigella dysenteriae</i> <i>Staphylococcus aureus</i>	<i>In vitro</i> <i>In vitro</i> <i>In vitro</i> <i>In vitro</i> <i>In vitro</i>	50 µL/disk MIC > 10 mg/mL MIC > 10 mg/mL 50 µL MIC > 10 mg/mL	Inactive Inactive Inactive Inactive Inactive	[83] [83] [83] [83] [83]
	Antifungal	LE	H ₂ O	<i>Aspergillus flavus</i> <i>Candida albicans</i> <i>Microsporium gypseum</i>	<i>In vitro</i> <i>In vitro</i> <i>In vitro</i>	MIC > 10 mg/mL MIC > 10 mg/mL MIC > 10 mg/mL	Inactive Inactive Inactive	[84] [84] [84]
	Antigonorrheal	LE	EtOH	<i>Neisseria gonorrhoea</i>	<i>In vitro</i>	50 µL/disk	Active	[32]
	Antitrypanosomal	LE	EtOH	<i>Trypanosoma cruzi</i>	<i>In vitro</i>	MIC > 1 mg/mL	Inactive	[83]
	Cytotoxic	LE	H ₂ O	Crustacean	<i>Artemia salina</i>	LC50 > 1,000 ppm	Inactive	[83]
	Inhib. of platelet aggregation	SE	/	<i>In vitro</i>	Thrombin aggregation	IC50 0.795 mg/mL	Inactive	[35]
Hawaii								
	Contraceptive	RO	/	Mouse	/	0.2 mL/animal	Inactive	[85]
Jamaica								
	Hypoglycemic	SE	CHCl ₃	Dog		1 g	Active	[39]
		SE	H ₂ O	Dog		200 mL/animal	Active	[39]
	Antioxidant	SE	EtOH	<i>In vitro</i>	<i>In vitro</i>	0.25 and 2.5 µg/mL	Active	[86]
Mexico								
	Allergenic	SE	Oil	Human		25 µL/ person	Active	[87]
Paraguay								
	Insecticidal	SE	Petr. ether	Insect	<i>Rhodnius neglectus</i>	50 µg	Inactive	[42]
Puerto Rico								
	Molluscicidal	TP	H ₂ O	Conch	<i>Lymnaea cubensis</i>	LD100 > 1 M ppm	Inactive	[88]
Dominican Republic								
	Cytotoxic	SE	EtOH	Cell culture	Molt 4 cells	200 µg/mL	Inactive	[89]
Trinidad and Tobago								
	Antibacterial	SE	EtOAc	<i>Escherichia coli</i> <i>Pseudomonas aeruginosa</i>	<i>In vitro</i> <i>In vitro</i>	1,000 µg/mL 1,000 µg/mL	Inactive Inactive	[90] [90]
USA								
	Anticonvulsant	RA	EtOH	Mouse	Seizures/electroshock	100 mg/kg	Inactive	[91]
	Antimalarial	SE	CHCl ₃	Rat	Seizures/pentylentetrazole	400 mg/kg	Inactive	[91]
	Mutagenic	SE	MeOH	Chicken	<i>Plasmodium gallinaceum</i>	388 mg/kg	Inactive	[92]
		SE	H ₂ O	<i>Salmonella typhimurium</i>		100 mg/plaque	Inactive	[93]
	Hypotensive	RA	EtOH	<i>In vitro</i>	Placental trophoblasts	100 mg/plaque	Inactive	[93]
	Toxicity	RA	H ₂ O	Rat		50 mg/kg	Active	[93]
		RA		Mouse		LD50 > 700 mg/kg	Inactive	[92]

LE: leaf; AP: aerial part; RO: root; SE: seed; /: not given; LD50: 50% lethal dose; IC50: concentration that inhibits 50% of the effect; MIC: minimum inhibitory concentration.

on the dose that is administered. The failure to require in-depth data related to toxicological and chemical analyses for the registration of food additives derived from natural sources [67, 74] certainly makes the information about possible unwanted effects and/or pharmacological activities resulting from their use, much rarer than expected in view of the importance of the topic. In Brazil, the use of annatto is so widespread that its safety is not even questioned.

Paumgarten et al. [91] evaluated the toxicity of annatto extracts in rats. Doses up to 500 mg/kg body weight/day were introduced directly into the stomach of pregnant rats to evaluate the effect on the mother and fetus, and no adverse effects were found for either. The annatto extract did not induce an increase in the incidence of visible external, visceral, or skeletal anomalies in the fetuses. Therefore, the study suggested that the annatto extract was not toxic to rats nor was it embryotoxic. Studies performed in Brazil by Alves de Lima et al. [67], where extracts of annatto were mixed with the food of male rats, showed that the concentrations tested had no mutagenic or antimutagenic activity in their bone marrow cells. A parallel toxicity study conducted by Hagiwara et al. [74] showed that 0.1% annatto extract administered for thirteen weeks in the feed of male and female rats did not show any adverse effects. However, when higher doses were administered (0.3 and 0.9%), the authors noticed an increase in liver weight as well as changes in blood chemistries, including increase in alkaline phosphatase, phospholipids, and total protein, as well as albumin and albumin/globulin ratio.

Hagiwara et al. [74] also evaluated extracts of annatto for liver carcinogenicity in rats and found no evidence of liver tumors, even when given to animals at a high dose of 200 mg/kg body weight/day, compared to an acceptable dose of 0.0 to 0.065 mg/kg/day, thus indicating that the danger of a hepatocarcinogenic effect in humans may be absent or negligible.

A toxicity test was performed with extracts obtained from both plant seeds and shoots, and no significant effect was observed. The experiments were performed in the United States using mice as the animal model and it was found that the LD50 was greater than 700 mg/kg [92].

9. Correlation between the Biological Activities, Phytochemistry, and the Traditional Uses of *Bixa orellana*

Table 1 shows that many of the traditional uses of *Bixa orellana* are the same in several countries of South and Central America, which suggests its effectiveness as a therapeutic agent. Extracts of *Bixa orellana* showed biological activities such as antioxidant, hypotensive, molluscicide, and antimalarial against A549 cells for carcinoma of the lung, allergy, hypoglycemic, antifungal, antioxidant, insect repellent, antigonococcal, and antivenom serum and some of them are in accordance with the traditional use; for example, in Brazil it is used to extract the seeds with purpose repellent insecticide and antimalaria and scientific studies in the same country with the *Lutzomyia longipalpis* insect repellent action

and prove a study in Cuba proved the pharmacological action for antimalarial activity when tested against *Plasmodium berghei*. Some of them are in accordance with the traditional use; for example, seed extracts have been used in Brazil and Cuba as insect repellent and antimalarial. Antioxidant and insect repellent activities can be attributed to the carotenoids and the essential and fixed oils, respectively.

Despite the previous reports about the presence of components with anti-inflammatory properties, such as salicylic acid, lutein, polyphenols, and tannins, this activity has not yet been proven for *Bixa orellana* extracts. Similarly, the plant's essential and fixed oils have shown antibacterial properties, although this activity has not been proven too.

In general, the data obtained in this review do not allow correlations between the biologic activities tested *in vitro* or *in vivo* with the compounds identified in this species. However, taking into account the related activities such as the antiparasitic effect and the lack of mutagenic and cytotoxicity activity, it is possible to consider *Bixa orellana* as a potential source for the development of phytopharmaceutical products.

In conclusion, the studies discussed in this review represent a rich database around the *Bixa orellana* activities and its potential uses, which evokes the feasibility of phytopharmaceuticals to treat some diseases whenever an antioxidant, hypotensor, or hypoglycemic activity is necessary.

Although the commercial exploitation of this species is well established, there are very few studies on its pharmacological effects. Considering the need for developing a safe and effective product, more studies should be performed in order to confirm other biological activities supported by the popular uses of *Bixa orellana*.

Conflict of Interests

The authors declare that they have no conflict of interests.

Acknowledgments

The authors are grateful to CNPq/PRONEX/FAPESQ and CAPES for financial support and to the NAPRALERT Database of the University of Illinois, USA, for the literature on *Bixa orellana*. A. Leyva helped in editing the English language of the paper.

References

- [1] M. B. Perecin, O. A. Bovi, and N. B. Maia, "Pesquisa com plantas aromáticas, medicinais corantes: o papel do Instituto Agronômico," *O Agrônomo*, vol. 54, pp. 21–24, 2002.
- [2] M. E. A. Elias, G. Schroth, J. L. V. Macêdo, M. S. S. Mota, and S. A. D'Angelo, "Mineral nutrition, growth and yields of annatto trees (*Bixa orellana*) in agroforestry on an Amazonian Ferralsol," *Experimental Agriculture*, vol. 38, no. 3, pp. 277–289, 2002.
- [3] A. R. R. Bastos, J. G. Carvalho, R. P. Assis, and A. B. C. Filho, "Marcha de absorção de nutrientes em urucum (*Bixa orellana* L.) tipo cultivado piave vermelha em fase de viveiro," *Cerne*, vol. 5, pp. 76–85, 1999.

- [4] E. P. Thomas, M. Colvin, S. B. Rosen, C. Zuccarini, and S. Petzer, "HIV prevalence study and costing analysis undertaken for the development of an HIV/AIDS workplace strategy for buffalo city municipality," Tech. Rep., Medical Research Council, Buffalo, NY, USA, 2005.
- [5] C. F. Franco, "O Agronegócio do urucum na região Nordeste," 1 Reunião Nacional da Cadeia Produtiva de Urucum, CD-Rom, 2007.
- [6] A. Giorgi, P. De Marinis, G. Granelli, L. M. Chiesa, and S. Panseri, "Secondary metabolite profile, antioxidant capacity, and mosquito repellent activity of *Bixa orellana* from Brazilian Amazon region," *Journal of Chemistry*, vol. 2013, Article ID 409826, 10 pages, 2013.
- [7] J. S. Silva, M. D. Moura, R. A. G. Oliveira, M. F. F. Diniz, and J. M. Barbosa-Filho, "Natural product inhibitors of ovarian neoplasia," *Phytomedicine*, vol. 10, no. 2-3, pp. 221-232, 2003.
- [8] F. C. F. Sousa, C. T. V. Melo, C. O. M. Citó et al., "Plantas medicinais e seus constituintes bioativos: uma revisão da bioatividade e potenciais benefícios nos distúrbios da ansiedade em modelos animais," *Revista Brasileira de Farmacognosia*, vol. 18, pp. 642-654, 2008.
- [9] L. J. Quintans Jr., J. R. G. S. Almeida, J. T. Lima et al., "Plants with anticonvulsant properties—a review," *Revista Brasileira de Farmacognosia*, vol. 18, pp. 798-819, 2008.
- [10] H. D. S. Falcão, J. A. Leite, J. M. Barbosa-Filho et al., "Gastric and duodenal antiulcer activity of alkaloids: a review," *Molecules*, vol. 13, no. 12, pp. 3198-3223, 2008.
- [11] H. S. Falcão, I. R. Mariath, M. F. F. M. Diniz, L. M. Batista, and J. M. Barbosa-Filho, "Plants of the American continent with antiulcer activity," *Phytomedicine*, vol. 15, no. 1-2, pp. 132-146, 2008.
- [12] I. R. Mariath, H. D. S. Falcão, J. M. Barbosa-Filho et al., "Plants of the American continent with antimalarial activity," *Revista Brasileira de Farmacognosia*, vol. 19, no. 1, pp. 158-192, 2009.
- [13] J. E. R. Hon, P. M. Soares, C. L. Melo et al., "Atividade farmacológica da monocrotalina isolada de plantas do gênero *Crotalaria*," *Revista Brasileira de Farmacognosia*, vol. 20, pp. 453-458, 2010.
- [14] R. N. Almeida, D. S. Navarro, and J. M. Barbosa-Filho, "Plants with central analgesic activity," *Phytomedicine*, vol. 8, no. 4, pp. 310-322, 2001.
- [15] N. Z. T. Jesus, H. S. Falcão, I. F. Gomes et al., "Tannins, peptic ulcers and related mechanisms," *International Journal of Molecular Sciences*, vol. 13, no. 3, pp. 3203-3228, 2012.
- [16] L. G. Rocha, J. R. G. S. Almeida, R. O. Macêdo, and J. M. Barbosa-Filho, "A review of natural products with antileishmanial activity," *Phytomedicine*, vol. 12, no. 6-7, pp. 514-535, 2005.
- [17] G. R. M. Lima, C. A. Montenegro, C. L. F. Almeida, P. F. Athayde-Filho, J. M. Barbosa-Filho, and L. M. Batista, "Database survey of anti-inflammatory plants in South America: a review," *International Journal of Molecular Sciences*, vol. 12, no. 4, pp. 2692-2749, 2011.
- [18] K. S. De Lira Mota, G. E. N. Dias, M. E. F. Pinto et al., "Flavonoids with gastroprotective activity," *Molecules*, vol. 14, no. 3, pp. 979-1012, 2009.
- [19] A. L. Souto, J. F. Tavares, M. S. da Silva, M. F. F. M. de Diniz, P. F. de Athayde-Filho, and J. M. Barbosa-Filho, "Anti-inflammatory activity of alkaloids: an update from 2000 to 2010," *Molecules*, vol. 16, no. 10, pp. 8515-8534, 2011.
- [20] F. L. Silva, D. C. H. Fischer, J. F. Tavares, M. S. Silva, P. F. de Athayde-Filho, and J. M. Barbosa-Filho, "Compilation of secondary metabolites from *Bidens pilosa* L.," *Molecules*, vol. 16, no. 2, pp. 1070-1102, 2011.
- [21] J. R. Filho, H. de Sousa Falcão, L. M. Batista, J. M. B. Filho, and M. R. Piuvezam, "Effects of plant extracts on HIV-1 protease," *Current HIV Research*, vol. 8, no. 7, pp. 531-544, 2010.
- [22] J. M. Barbosa-Filho, A. A. Alencar, X. P. Nunes et al., "Sources of alpha-, beta-, gamma-, delta- and epsilon-carotenes: a twentieth century review," *Revista Brasileira de Farmacognosia*, vol. 18, no. 1, pp. 135-154, 2008.
- [23] S. N. S. Silva, C. L. F. Amaral, and T. N. H. Rebouças, "Adoption of conservation practices on farm and selection of varieties by producers of annatto in the city of Vitoria da Conquista-BA," *Revista Brasileira de Agroecologia*, vol. 5, pp. 106-113, 2010.
- [24] A. L. Bandoni, M. E. Mendiondo, R. V. D. Rondina, and J. D. Coussio, "Survey of Argentine medicinal plants. I. Folklore and phytochemical screening," *Lloydia*, vol. 35, no. 1-4, pp. 69-80, 1972.
- [25] G. H. Garcia, R. Campos, R. A. de Torres et al., "Antiherpetic activity of some Argentine medicinal plants," *Fitoterapia*, vol. 61, no. 6, pp. 542-546, 1990.
- [26] M. G. L. Brandao, T. S. M. Grandi, E. M. M. Rocha, D. R. Sawyer, and A. U. Krettli, "Survey of medicinal plants used as antimalarials in the Amazon," *Journal of Ethnopharmacology*, vol. 36, no. 2, pp. 175-182, 1992.
- [27] M. P. Corrêa, *Dicionário das Plantas Úteis do Brasil e das Exóticas Cultivadas*, vol. 4, Ministério da Agricultura/IBDF, Rio de Janeiro, Brasil, 1978.
- [28] G. S. Hirschmann and A. R. de Arias, "A survey of medicinal plants of minas gerais, Brazil," *Journal of Ethnopharmacology*, vol. 29, no. 2, pp. 159-172, 1990.
- [29] R. Otero, R. Fonnegra, S. L. Jiménez et al., "Snakebites and ethnobotany in the northwest region of Colombia. Part I: traditional use of plants," *Journal of Ethnopharmacology*, vol. 71, no. 3, pp. 493-504, 2000.
- [30] H. Garcia-Barriga, *Flora Medicinal de Colombia*, vol. 2, Universidad Nacional, Bogota, Colombia, 1975.
- [31] J. T. Roig y Mesa, *Plantas Medicinales, Aromaticas o Venenosas de Cuba*, Ministério de Agricultura, Havana, Cuba, 1945.
- [32] A. Cáceres, H. Menéndez, E. Méndez et al., "Antigonorrhoeal activity of plants used in Guatemala for the treatment of sexually transmitted diseases," *Journal of Ethnopharmacology*, vol. 48, no. 2, pp. 85-88, 1995.
- [33] J. A. Duke and R. V. Martinez, *Amazonian Ethnobotanical Dictionary*, CRC Press, Boca Raton, Fla, USA, 1994.
- [34] V. R. Ramirez, L. J. Mostacero, A. E. Garcia et al., *Vegetales Empleados en Medicina Tradicional Norperuana*, Banco Agrario del Peru, Universidad Nacional de Trujillo, Trujillo, Peru, 1988.
- [35] R. Villar, J. M. Calleja, C. Morales, and A. Cáceres, "Screening of 17 Guatemalan medicinal plants for platelet antiaggregant activity," *Phytotherapy Research*, vol. 11, pp. 441-445, 1997.
- [36] L. M. Giron, V. Freire, A. Alonzo, and A. Cáceres, "Ethnobotanical survey of the medicinal flora used by the Caribs of Guatemala," *Journal of Ethnopharmacology*, vol. 34, no. 2-3, pp. 173-187, 1991.
- [37] D. L. Lentz, "Medicinal and other economic plants of the Paya of Honduras," *Economic Botany*, vol. 47, no. 4, pp. 358-370, 1993.
- [38] D. L. Lentz, A. M. Clark, C. D. Hufford et al., "Antimicrobial properties of Honduran medicinal plants," *Journal of Ethnopharmacology*, vol. 63, no. 3, pp. 253-263, 1998.
- [39] E. Y. Morrison and M. E. West, "The effect of *Bixa orellana* (Annatto) on blood sugar levels in the anaesthetized dog," *West Indian Medical Journal*, vol. 34, no. 1, pp. 38-42, 1985.

- [40] F. G. Coe and G. J. Anderson, "Screening of medicinal plants used by the Garífuna of eastern Nicaragua for bioactive compounds," *Journal of Ethnopharmacology*, vol. 53, no. 1, pp. 29–50, 1996.
- [41] B. Barrett, "Medicinal plants of Nicaragua's Atlantic Coast," *Economic Botany*, vol. 48, no. 1, pp. 8–20, 1994.
- [42] G. Schmeda-Hirschmann and A. R. de Arias, "A screening method for natural products on triatomine bugs," *Phytotherapy Research*, vol. 6, no. 2, pp. 68–73, 1992.
- [43] G. E. Simpson, "Folk medicine in Trinidad," *The Journal of American Folklore*, vol. 75, no. 298, pp. 326–340, 1962.
- [44] D. Mahabir and M. C. Gulliford, "Use of medicinal plants for diabetes in Trinidad and Tobago," *Revista Panamericana de Salud Publica*, vol. 1, no. 3, pp. 174–179, 1997.
- [45] E. S. Ayensu, "Medicinal plants of the West Indies," Unpublished Manuscript, 1978.
- [46] A. Z. Mercadante, A. Steck, D. Rodriguez-Amaya, H. Pfander, and G. Britton, "Isolation of methyl 9'Z-APO-6'-lycopenoate from *Bixa Orellana*," *Phytochemistry*, vol. 41, no. 4, pp. 1201–1203, 1996.
- [47] E. Angelucci, H. K. Arima, E. A. Kumagai, and I. Annatto, "Preliminary data of the chemical composition," *Coletanea do Instituto de Tecnologia de Alimentos*, vol. 11, pp. 89–96, 1980.
- [48] A. S. L. Tirimanna, "Study of the carotenoid pigments of *Bixa orellana* L. Seeds by thin layer chromatography," *Mikrochimica Acta*, vol. 76, no. 1-2, pp. 11–16, 1981.
- [49] J. M. Barbosa-Filho, R. N. Silva-Filho, B. F. Lira et al., "Teor de bixina em quatro variedades de *Bixa orellana* L. cultivadas na Paraíba," *Revista Brasileira de Farmacognosia*, vol. 7-8, pp. 41–47, 1998.
- [50] A. Z. Mercadante, A. Steck, and H. Pfander, "Isolation and structure elucidation of minor carotenoids from annatto (*Bixa orellana* L.) seeds," *Phytochemistry*, vol. 46, no. 8, pp. 1379–1383, 1997.
- [51] C. Prentice-Hernandez and O. Rusig, "Annatto (*Bixa orellana* L.) extract obtained using ethyl alcohol as a solvent," *Arquivos de Biologia e Tecnologia*, vol. 35, pp. 63–74, 1992.
- [52] A. Z. Mercadante, "Composition of carotenoids from annatto," *ACS Symposium Series*, vol. 775, pp. 92–101, 2001.
- [53] M. J. Scotter, L. A. Wilson, G. P. Appleton, and L. Castle, "Analysis of Annatto (*Bixa orellana*) Food Coloring Formulations. 1. Determination of Coloring Components and Colored Thermal Degradation Products by High-Performance Liquid Chromatography with Photodiode Array Detection," *Journal of Agricultural and Food Chemistry*, vol. 46, no. 3, pp. 1031–1038, 1998.
- [54] A. J. Degnan, J. H. von Elbe, and R. W. Hartel, "Extraction of annatto seed pigment by supercritical carbon dioxide," *Journal of Food Science*, vol. 56, pp. 1655–1659, 1991.
- [55] A. Z. Mercadante, A. Steck, and H. Pfander, "Isolation and identification of new apocarotenoids from annatto (*Bixa orellana*) seeds," *Journal of Agricultural and Food Chemistry*, vol. 45, no. 4, pp. 1050–1054, 1997.
- [56] S. G. Anderson, M. G. Nair, A. Chandra, and E. Morrison, "Supercritical fluid carbon dioxide extraction of annatto seeds and quantification of *trans*-bixin by high pressure liquid chromatography," *Phytochemical Analysis*, vol. 8, pp. 247–249, 1997.
- [57] J. M. Barbosa-Filho, *Bixa orellana: Retrospectiva de usos populares, atividades biológicas, fitoquímica e emprego na fitocosmética, no continente americano*, Simpósio Brasileiro do Urucum—SIMBRAU, João Pessoa, Brazil, 2006.
- [58] J. Revilla, *Plantas da Amazônia: Oportunidades Econômicas e Sustentáveis*, Manaus: Programa de Desenvolvimento Empresarial e Tecnológico, 2nd edition, 2001.
- [59] F. Oliveira, G. Akisue, and M. K. Akisue, *Farmacognosia*, Atheneu, São Paulo, Brazil, 1996.
- [60] J. Alonso, *Tratado de Fitofármacos y Nutracêuticos*, Corpus, Rosário, Argentina, 2004.
- [61] C. O. Franco, E. G. Fabri, M. Barreiro Neto, M. H. Manfiolli, M. N. C. Harder, and N. C. Rucker, *Urucum: Sistema de Produção para o Brasil*, Emepa-Pb, Apta, João Pessoa, Brazil, 2008.
- [62] P. A. Venugopalan, G. A. Giridhar, and A. G. Ravishankar, "Food, Ethnobotanical and diversified applications of *Bixa orellana* L.: a scope for its improvement through biotechnological mediation," *Indian Journal of Fundamental and Applied Life Sciences*, vol. 1, pp. 9–31, 2011.
- [63] L. Taylor, *Herbal Secrets of the Rainforest*, Sage Press, Rocklin, Calif, USA, 2nd edition, 2002.
- [64] J. S. Oliveira, *Caracterização, Extração e Purificação por Cromatografia de Compostos de Urucum (Bixa orellana L.) [tese doctoral em Engenharia Química]*, Universidade Federal de Santa Catarina, Florianópolis, Brazil, 2005.
- [65] A. Z. Mercadante, A. Steck, and H. Pfander, "Three minor carotenoids from annatto (*Bixa orellana*) seeds," *Phytochemistry*, vol. 52, no. 1, pp. 135–139, 1999.
- [66] V. Galindo-Cuspinera, M. B. Lubran, and S. A. Rankin, "Comparison of volatile compounds in water- and oil-soluble annatto (*Bixa orellana* L.) extracts," *Journal of Agricultural and Food Chemistry*, vol. 50, no. 7, pp. 2010–2015, 2002.
- [67] R. O. Alves de Lima, L. Azevedo, L. R. Ribeiro, and D. M. F. Salvadori, "Study on the mutagenicity and antimutagenicity of a natural food colour (annatto) in mouse bone marrow cells," *Food and Chemical Toxicology*, vol. 41, no. 2, pp. 189–192, 2003.
- [68] L. R. P. Lima, T. T. Oliveira, T. J. Nagem et al., "Bixina, norbixina e quercetina e seus efeitos no metabolismo lipídico de coelhos," *Brazilian Journal of Veterinary Research and Animal Science*, vol. 38, pp. 196–200, 2001.
- [69] J. D. Fontana, S. V. Mendes, D. S. Persike, L. F. Peracetta, and M. Passos, "Carotenóides: cores atraentes e ação biológica," *Biotecnologia Ciência & Desenvolvimento*, vol. 2, p. 13, 2000.
- [70] C. A. Penna, M. Radice, G. O. Gutkind et al., "Antibacterial and antifungal activities of some Argentinean plants," *Fitoterapia*, vol. 65, no. 2, pp. 172–174, 1994.
- [71] A. M. Broussalis, G. E. Ferraro, V. S. Martino, R. Pinzón, J. D. Coussio, and J. C. Alvarez, "Argentine plants as potential source of insecticidal compounds," *Journal of Ethnopharmacology*, vol. 67, no. 2, pp. 219–223, 1999.
- [72] A. Rojas de Arias, G. Schmeda-Hirschmann, and A. Falcao, "Feeding deterrence and insecticidal effects of plant extracts on *Lutzomyia longipalpis*," *Phytotherapy Research*, vol. 6, no. 2, pp. 64–67, 1992.
- [73] M. S. Pinheiro and M. Z. Rouquayrol, "Molluscicidal activity of plants from Northeast Brazil," *Revista Brasileira de Pesquisas Médicas e Biológicas*, vol. 7, pp. 389–394, 1974.
- [74] A. Hagiwara, N. Imai, T. Ichihara et al., "A thirteen-week oral toxicity study of annatto extract (norbixin), a natural food color extracted from the seed coat of annatto (*Bixa orellana* L.), in Sprague-Dawley rats," *Food and Chemical Toxicology*, vol. 41, no. 8, pp. 1157–1164, 2003.
- [75] C. R. Almeida, R. B. Silva, M. J. Marques, and J. K. Chavasco, "Evaluation of antiparasitic activity of hydroethanolic extracts from root, stem and leaf of *Bixa orellana* L. on *Leishmania*

- amazonensis samples," *Revista da Universidade Vale do Rio Verde*, vol. 10, no. 2, pp. 384–391, 2012.
- [76] M. V. Lopes, V. C. Desoti, A. D. O. Caleare, T. Ueda-Nakamura, S. O. Silva, and C. V. Nakamura, "Mitochondria superoxide anion production contributes to geranylgeraniol-induced death in leishmania amazonensis," *Evidence-Based Complementary and Alternative Medicine*, vol. 2012, Article ID 298320, 9 pages, 2012.
- [77] J. M. Ferreira, D. F. Sousa, M. B. Dantas et al., "Effects of *Bixa orellana* L. seeds on hyperlipidemia," *Phytotherapy Research*, vol. 27, no. 1, pp. 144–147, 2013.
- [78] P. S. Benoit, H. H. S. Fong, G. H. Svoboda, and N. R. Farnsworth, "Biological and phytochemical evaluation of plants. XIV. Anti-inflammatory evaluation of 163 species of plants," *Lloydia*, vol. 39, no. 2-3, pp. 160–171, 1976.
- [79] D. Carabajal, A. Casaco, L. Arruzazabala, R. Gonzalez, and V. Fuentes, "Pharmacological screening of plant decoctions commonly used in Cuban folk medicine," *Journal of Ethnopharmacology*, vol. 33, no. 1-2, pp. 21–24, 1991.
- [80] A. F. C. Valdés, J. M. Martínez, D. A. Rodríguez, R. S. Lizama, and Y. G. Gaitén, "Actividad antimalárica de un extracto hidroalcohólico de *Bixa orellana* L.," *Revista Cubana de Medicina Tropical*, vol. 63, pp. 181–185, 2011.
- [81] A. D. García, H. R. Sánchez, and R. S. Lizama, "Citotoxicidad de extractos de plantas medicinales sobre la línea celular de carcinoma de pulmón humano A549," *Revista Cubana de Farmacia*, vol. 45, pp. 101–108, 2011.
- [82] B. Freixa, R. Vila, L. Vargas, N. Lozano, T. Adzet, and S. Canigueral, "Screening for antifungal activity of nineteen Latin American plants," *Phytotherapy Research*, vol. 12, no. 6, pp. 427–430, 1998.
- [83] A. Cáceres, B. López, S. González, I. Berger, I. Tada, and J. Maki, "Plants used in Guatemala for the treatment of protozoal infections. I. Screening of activity to bacteria, fungi and American trypanosomes of 13 native plants," *Journal of Ethnopharmacology*, vol. 62, no. 3, pp. 195–202, 1998.
- [84] A. Caceres, O. Cano, B. Samayoa, and L. Aguilar, "Plants used in Guatemala for the treatment of gastrointestinal disorders. I. Screening of 84 plants against enterobacteria," *Journal of Ethnopharmacology*, vol. 30, no. 1, pp. 55–73, 1990.
- [85] A. S. Matsui, S. Hoskin, M. Kashiwagi et al., "A survey of natural products from Hawaii and other areas of the Pacific for an antifertility effect in mice," *Internationale Zeitschrift fur Klinische Pharmakologie, Therapie, und Toxikologie*, vol. 5, no. 1, pp. 65–69, 1971.
- [86] M. Abayomi, A. S. Adebayo, D. Bennett, R. Porter, and J. S. Campbell, "In vitro antioxidant activity of *Bixa orellana* (Annatto) seed extract," *Journal of Applied Pharmaceutical Science*, vol. 4, no. 2, pp. 101–106, 2014.
- [87] N. W. Dunham and K. R. Allard, "A preliminary pharmacologic investigation of the roots of *Bixa orellana*," *Journal of the American Pharmacist Association*, vol. 49, pp. 218–219, 1960.
- [88] F. R. Medina and R. Woodbury, "Terrestrial plants molluscicidal to lymnaeid hosts of *Fascioliasis hepatica* in Puerto Rico," *Journal of Agricultura of the University of Puerto Rico*, vol. 63, pp. 366–376, 1979.
- [89] B. Weniger, Y. Jiang, A. Oulad-Ali, L. Italiano, J. P. Beck, and R. Anton, "Biological effects of bixin and *Bixa orellana* extracts on lymphoid cells in culture," *Planta Medica*, vol. 59, no. 7, article A680, 1993.
- [90] C. M. Chariandy, C. E. Seaforth, R. H. Phelps, G. V. Pollard, and B. P. S. Khambay, "Screening of medicinal plants from Trinidad and Tobago for antimicrobial and insecticidal properties," *Journal of Ethnopharmacology*, vol. 64, no. 3, pp. 265–270, 1999.
- [91] F. J. R. Paumgartten, R. R. Carvalho, I. B. Araújo et al., "Evaluation of the developmental toxicity of annatto in the rat," *Food and Chemical Toxicology*, vol. 40, no. 11, pp. 1595–1601, 2002.
- [92] C. F. Spencer, F. R. Koniuszy, E. F. Rogers et al., "Survey of plants for antimalarial activity," *Lloydia*, vol. 10, no. 3, pp. 145–174, 1947.
- [93] H. Mikkelsen, J. C. Larsen, and F. Tarding, "Hypersensitivity reactions to food colours with special reference to the natural colour annatto extract (butter colour)," *Toxicological Aspects of Food Safety*, vol. 1, pp. 141–143, 1978.