microbial biotechnology



Corrigendum

Methanol-based cadaverine production by genetically engineered *Bacillus methanolicus* strains

Specification of the mistake identified

The cadaverine concentrations reported were miscalculated and overestimated by 41.6% because the HPLC standard solution was made of cadaverine dihydrochloride (175.1 g/mol) not cadaverine (102.18 g/mol).

Implications of the corrections

The identified mistake does not destroy the novelty of the paper and all main hypotheses and conclusions remain valid.

Minor adjustments due to the correction (changes in colour red)

1 The published sentence 'Notably, combined formation of cadaverine and L-lysine by the cadA and ldcC expressing strains was above 3-fold higher than L-lysine formation by the parent strain (Table 1), which might indicate feedback deregulation by L-lysine as consequence of a metabolic pull by lysine decarboxylase.'

should be adjusted to

- 'Notably, combined formation of cadaverine and L-lysine by the cadA expressing strain was nearly 2-fold higher than L-lysine formation by the parent strain (Table 1), which might indicate feedback deregulation by L-lysine as consequence of a metabolic pull by lysine decarboxylase'.
- 2 The statement below concerning three different lysine decarboxylase expressing strains is no longer true for MGA3 (pTH1mp-ldcC-lysA) because it in fact produces less cadaverine than strain MGA3(pTH1mp-lysA) produces L-lysine.

Table 1. Specific L-lysine decarboxylase activities, cadaverine and L-lysine production levels in recombinant B. methanolicus M168-20 strains.

Plasmid	L-lysine decarboxylase specific activity nmol/min/mg protein	Cadaverine mg/L	L-lysine mg/L	Cadaverine + L-lysine mg/L
pHP13	< 1 ± 0.2	0.0	140 \pm 10	140
pTH1mp-ldcC	7.0 ± 1.0	75 ± 6	40 ± 5	115
pTH1mp-cadA	88.0 ± 11.0	245 ± 15	10 ± 2	255

The results shown are from triplicate (cadaverine and L-lysine) and duplicate (lysine decarboxylase activity) shake flask cultures. Activity was measured using crude extracts from exponentially growing cells, whereas the production levels were found from late stationary cultures, approximately 20 h after inoculation.

Table 2. Production of cadaverine and L-lysine by recombinant B. methanolicus M168-20 strains cultivated at different medium pH.

	M168-20(pHP13)	M168-20(pHP13)		M168-20(pTH1mp-ldcC)		M168-20(pTH1mp- <i>cadA</i>)	
рН	Cadaverine	L-lysine	Cadaverine	L-lysine	Cadaverine	L-lysine	
6.5	0	50 ± 10	30 ± 3	< 15	26 ± 3	< 15	
7.2	0	130 ± 10	78 ± 6	40 ± 5	250 ± 12	< 30	
7.6	0	140 ± 10	185 \pm 12	< 30	263 ± 12	< 30	
8.0	0	140 ± 10	178 ± 18	< 15	292 ± 18	< 30	
8.5	0	140 ± 10	178 ± 18	< 15	303 ± 18	< 30	

The mean values (mg/L) and standard deviation of triplicate shake flask cultures are presented. The production levels were found from late stationary cultures, from 20-30 h after inoculation.

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Table 3. Cadaverine and L-lysine production by recombinant B. methanolicus MGA3 strains.

Plasmid	Cadaverine mg/L	L-lysine mg/L
pHP13	0	7 ± 1 ^a
pTH1mp-lysC	0	55 ± 5^{a}
pTH1mp-lysA	0	150 ± 10^{a}
pTH1mp-ldcC	12 ± 2	7 ± 1
pTH1mp-ldcC-lysC	82 ± 6	< 10
pTH1mp-ldcC-lysA	111 ± 6	< 10
pTH1mp-cadA	262 ± 18	< 10
pTH1mp- <i>cadA-lysA</i>	280 ± 18	< 10

The production levels were found from late stationary shake flask cultures, approximately 20 h after inoculation.

Table 4. Fed-batch methanol fermentation production data of strains MGA3(pTH1mp-cadA) and MGA3(pHP13).

Strain	CDW g/L	$\begin{array}{c} \mu^a \\ h^{-1} \end{array}$	Asp ^b g/L	Glu ^b g/L	Ala ^b g/L	Lys ^b g/L	Cad ^b g/L
MGA3(pTH1mp- <i>cadA</i>)	65.5	0.45	1.5	71.8	10.2	0.0	6.5
MGA3(pHP13)	45.0	0.49	1.1	59.0	12.0	0.4	0.0

μ, specific growth rate; Ala, L-alanine; Asp, L-aspartate; Cad, Cadaverine; CDW, cell dry weight; Glu, L-glutamate; Lys, L-lysine. The maximum mean values from early (CDW) or late stationary growth phase is presented for the MGA3(pTH1mp-cadA) duplicate cultures and the deviation never exceed ten per cent. The MGA3(pHP13) data were imported from (Brautaset, T., et al., 2010).

'Interestingly, L-lysine production was in each case lower (7, 55 and 150 mg/L, respectively; Table 3) for the three isogenic strains that do not express IdcC, i.e. MGA3(pHP13), MGA3(pTH1mp-lysC) and MGA3(pTH1mp-lysA), respectively, indicating that LdcC exerts a metabolic pull deregulating flux through the L-lysine biosynthesis pathway'.

SUMMARY

Methanol is regarded as an attractive substrate for biotechnological production of value-added bulk products, such as amino acids and polyamines. In the present study, the methylotrophic and thermophilic bacterium Bacillus methanolicus was engineered into a microbial cell factory for the production of the platform chemical 1,5-diaminopentane (cadaverine) from methanol. This was achieved by the heterologous expression of the Escherichia coli genes cadA and ldcC encoding two different lysine decarboxylase enzymes and by increasing the overall L-lysine production levels in this host. Both CadA and LdcC were functional in B. methanolicus cultivated at 50°C and expression of cadA resulted in cadaverine production levels up to 300 mg l⁻¹ during shake flask conditions. A volume-corrected concentration of 6.5 g l⁻¹ of cadaverine was obtained by high-cell-density fed-batch methanol fermentation. Our results demonstrated that efficient conversion of L-lysine into cadaverine presumably has severe effects on feedback regulation of the L-lysine biosynthetic pathway in B. methanolicus. By also investigating the cadaverine tolerance level, B. methanolicus proved to be an exciting alternative host and comparable to the well-known bacterial hosts E. coli and Corynebacterium glutamicum. This study represents the first demonstration of microbial production of cadaverine from methanol.

The authors apologize to all readers for the inconvenience caused to the identified mistake.

Reference

Nærdal, I., Pfeifenschneider, J., Brautaset, T. and Wendisch, V.F. (2015) Methanol-based cadaverine production by genetically engineered Bacillus methanolicus strains. Microb Biotechnol 8: 342-350.

a. Data imported from (Nærdal, I., et al., 2011).

a. Specific growth rates are maximum values calculated from the exponential growth period.

b. CDW, cadaverine and amino acid concentrations are maximum values and volume corrected (see "Experimental Procedures" section).