- Lawson WE, Crossno PF, Polosukhin VV, Roldan J, Cheng D-S, Lane KB, et al. Endoplasmic reticulum stress in alveolar epithelial cells is prominent in IPF: association with altered surfactant protein processing and herpesvirus infection. Am J Physiol Lung Cell Mol Physiol 2008;294:L1119–L1126.
- Korfei M, Ruppert C, Mahavadi P, Henneke I, Markart P, Koch M, et al. Epithelial endoplasmic reticulum stress and apoptosis in sporadic idiopathic pulmonary fibrosis. *Am J Respir Crit Care Med* 2008;178: 838–846.
- Takezaki A, Tsukumo S-I, Setoguchi Y, Ledford JG, Goto H, Hosomichi K, et al. A homozygous SFTPA1 mutation drives necroptosis of type II alveolar epithelial cells in patients with idiopathic pulmonary fibrosis. *J Exp Med* 2019;216:2724–2735.
- Torres-González E, Bueno M, Tanaka A, Krug LT, Cheng D-S, Polosukhin VV, *et al.* Role of endoplasmic reticulum stress in agerelated susceptibility to lung fibrosis. *Am J Respir Cell Mol Biol* 2012; 46:748–756.
- 10. Katzen J, Wagner BD, Venosa A, Kopp M, Tomer Y, Russo SJ, *et al.* An SFTPC BRICHOS mutant links epithelial ER stress and spontaneous lung fibrosis. *JCI Insight* 2019;4:126125.
- Ayaub EA, Kolb PS, Mohammed-Ali Z, Tat V, Murphy J, Bellaye P-S, et al. GRP78 and CHOP modulate macrophage apoptosis and the development of bleomycin-induced pulmonary fibrosis. J Pathol 2016;239:411–425.
- 12. Borok Z, Horie M, Flodby P, Wang H, Liu Y, Ganesh S, *et al. Grp78* loss in epithelial progenitors reveals an age-linked role for endoplasmic reticulum stress in pulmonary fibrosis. *Am J Respir Crit Care Med* 2020;201:198–211.
- 13. Hsu H-S, Liu C-C, Lin J-H, Hsu T-W, Hsu J-W, Su K, *et al.* Involvement of ER stress, PI3K/AKT activation, and lung fibroblast proliferation in bleomycin-induced pulmonary fibrosis. *Sci Rep* 2017;7:14272.
- Lawson WE, Polosukhin VV, Stathopoulos GT, Zoia O, Han W, Lane KB, et al. Increased and prolonged pulmonary fibrosis in surfactant protein C-deficient mice following intratracheal bleomycin. Am J Pathol 2005;167:1267–1277.

- Chen G, Ribeiro CMP, Sun L, Okuda K, Kato T, Gilmore RC, et al. XBP1S regulates MUC5B in a promoter variant-dependent pathway in idiopathic pulmonary fibrosis airway epithelia. Am J Respir Crit Care Med 2019;200:220–234.
- Hancock LA, Hennessy CE, Solomon GM, Dobrinskikh E, Estrella A, Hara N, et al. Muc5b overexpression causes mucociliary dysfunction and enhances lung fibrosis in mice. Nat Commun 2018;9:5363.
- Habermann AC, Gutierrez AJ, Bui LT, Yahn SL, Winters NI, Calvi CL, et al. Single-cell RNA-sequencing reveals profibrotic roles of distinct epithelial and mesenchymal lineages in pulmonary fibrosis [preprint]. *bioRxiv*; 2019 [accessed 2019 Oct 20]. Available from: https://www.biorxiv.org/content/10.1101/753806v1.
- Adams TS, Schupp JC, Poli S, Ayaub EA, Neumark N, Ahangari F, et al. Single cell RNA-seq reveals ectopic and aberrant lung resident cell populations in idiopathic pulmonary fibrosis [preprint]. *bioRxiv* 2019 [accessed 2019 Oct 20]. Available from: https://www.biorxiv.org/ content/10.1101/759902v3.
- Sisson TH, Mendez M, Choi K, Subbotina N, Courey A, Cunningham A, et al. Targeted injury of type II alveolar epithelial cells induces pulmonary fibrosis. Am J Respir Crit Care Med 2010;181:254–263.
- Alder JK, Barkauskas CE, Limjunyawong N, Stanley SE, Kembou F, Tuder RM, et al. Telomere dysfunction causes alveolar stem cell failure. Proc Natl Acad Sci USA 2015;112:5099–5104.
- Povedano JM, Martinez P, Flores JM, Mulero F, Blasco MA. Mice with pulmonary fibrosis driven by telomere dysfunction. *Cell Reports* 2015;12:286–299.
- Naikawadi RP, Disayabutr S, Mallavia B, Donne ML, Green G, La JL, et al. Telomere dysfunction in alveolar epithelial cells causes lung remodeling and fibrosis. JCI Insight 2016;1:e86704.
- 23. Nureki S-I, Tomer Y, Venosa A, Katzen J, Russo SJ, Jamil S, et al. Expression of mutant Sftpc in murine alveolar epithelia drives spontaneous lung fibrosis. *J Clin Invest* 2018;128:4008–4024.

Copyright © 2020 by the American Thoracic Society

Check for updates

a Hitting a HOMER: Epidemiology to the Bedside when Evaluating for Stereotactic Ablative Radiotherapy

In this issue of the *Journal*, Martinez-Zayas and colleagues (pp. 212–223) report and validate a novel prediction model (HOMER) to calculate the probability of patients with non-small cell lung cancer (NSCLC) having mediastinal lymph node involvement (1). Determining a patient's likelihood of lymph node metastasis is paramount in determining the stage of lung cancer and therefore appropriate treatment options. Clinical staging, including imaging modalities and biopsy techniques, remains a challenge and frequently falls short of surgical staging, depending on how aggressive the preoperative evaluation is (2). Accurate staging has been associated with improved survival and remains a huge

emphasis in the care of patients with lung cancer (3). The study by Martinez-Zayas and colleagues is the first to derive and validate a risk model aimed at discriminating between the most clinically useful forms of nodal disease in patients who were both surgical and nonsurgical candidates: N0, N1, and N2/3 disease.

The authors should be commended for the statistical rigor used to derive and validate their model. Covariates used to develop the model were pragmatic, clinically relevant, and appropriately limited by the last common outcome. By externally validating their prediction model at other medical centers, the authors offer a model with the possibility of geographic stability for patients with NSCLC without T4 tumors or distant metastasis, after adjusting for the local institution's population. The authors further supported their model with temporal validation to show stability over time (4). HOMER therefore has the potential to be generalizable in both the short term and the long term for patients with NSCLC seeking treatment at well-practiced thoracic oncology centers that use systematic endobronchial ultrasound-guided transbronchial needle aspiration (EBUS TBNA) lymph node staging. To carry out the systematic EBUS lymph node staging that the output of HOMER applies to, an examination of the intrathoracic nodes is required by EBUS,

This article is open access and distributed under the terms of the Creative Commons Attribution 4.0 International License (https://creativecommons. org/licenses/by/4.0/).

N.N. is supported by a UK Medical Research Council Clinical Academic Research Partnership. This work was in part undertaken at University College London, University College London Hospital, which received a proportion of funding from the UK Department of Health's NIHR Biomedical Research Centre's funding scheme (N.N.).

Originally Published in Press as DOI: 10.1164/rccm.201910-1933ED on October 28, 2019

EDITORIALS

beginning with contralateral N3 nodes, followed by N2 and then N1 lymph nodes. Any lymph node measuring \geq 5 mm in short axis is sampled, aiming for a minimum of three N2/3 lymph node stations sampled per procedure (5).

There are several clinically useful applications of HOMER. Assuming a patient is not a surgical candidate, the preferred treatment for N0 disease is definitive stereotactic ablative radiotherapy (SABR) and will not be confirmed surgically. Therefore, making an accurate clinical prediction of this disease state is crucial (6). Previous work was not able to discriminate between patients with N0 and N1 disease (7). HOMER presents two exciting ways to bring evidence-based decision making to these patients. First, the model can help predict the pretest probability that an EBUS TBNA will detect NSCLC, based on widely available clinical and radiographic data. As pointed out elegantly in the discussion, this can allow a more objective discussion about the risk and benefit of requiring an EBUS TBNA before SABR. As the low risk of complication is approached by the predicted probability of lymph node metastasis detected by EBUS TBNA, one can more confidently consider avoiding invasive mediastinal staging. This is especially relevant for patients at increased risk for complications during bronchoscopy. This may also be useful for patients with confirmed NSCLC from a transthoracic needle biopsy with radiographic N2/N3 disease who are at extremely high risk for bronchoscopy.

The other way HOMER can be used for these patients is to calculate a posttest probability of N1 disease in a patient being considered for SABR who has a negative EBUS TBNA. Current guidelines appropriately lean toward cytologic or pathologic confirmation for mediastinal staging. They suggest preoperative invasive mediastinal staging in patients with NSCLC unless the tumor is T1 (<3 cm) and peripheral, and the mediastinal lymph nodes are radiographically negative by computed tomography and positron emission tomography (7). This recommendation is based on a low false-negative rate (i.e., lymph node metastasis) in this patient population, as determined by older descriptive studies (8). Importantly, there are more recent data to support occult lymph node metastasis and a limited sensitivity for EBUS TBNA in similar patient populations (9). After making an assumption about the sensitivity of EBUS TBNA, a clinician can calculate a posttest probability, using HOMER, as is also demonstrated in the discussion. The ability to have an objective probability of N1 disease after a negative EBUS TBNA can assist the multidisciplinary lung cancer team when weighing the harm of SABR with occult N1 disease versus the harm of a larger radiation field and the addition of chemotherapy for presumed N1 disease.

As the authors warn in the discussion, HOMER should not be used to calculate the sensitivity of EBUS TBNA or the pretest probability of nodal disease, as there was no gold standard (i.e., surgical lymph node dissection) to compare with EBUS TBNA. Therefore, one must often put the model in the context of an assumed EBUS TBNA sensitivity, which is probably dependent on technique, lymph node size, necrosis, and tumor cellularity of each nodal metastasis. As mentioned here, EBUS TBNA may not be highly sensitive for NSCLC in radiographically normal lymph nodes (8). Clinicians may need to integrate HOMER with other observational studies associating standardized uptake value (SUV_{max}) of the primary tumor, adenocarcinoma histology, non-lower lobe tumors, and tumor size with occult lymph node metastasis after negative preoperative positron emission tomography/computed tomography (10–13).

HOMER affords the opportunity to integrate data-driven decision making into our NSCLC staging and treatment decisions, much in the way we use probability to guide the management of lung nodules (14). The model's performance could even be further refined as more data become available for patients with N1 disease. There are exciting ways to imagine an extended data set and similar methods being employed to predict other clinically meaningful outcomes in NSCLC. For nonsurgical patients, can we model the probability of long-term clinical response, using SABR, after a negative systematic EBUS? For surgical patients, can a model to predict occult lymph node metastasis after a negative systematic EBUS be similarly derived and validated? HOMER is an excellent example of using evidence collected from current practice to rigorously create a novel prediction tool to aid future clinical decisions. It is an important guide in practice and in principle, as we continue to strive for more evidence-based and data-driven care for patients with lung cancer.

Author disclosures are available with the text of this article at www.atsjournals.org.

David M. DiBardino, M.D. Division of Pulmonary, Allergy and Critical Care Medicine University of Pennsylvania Philadelphia, Pennsylvania

Neal Navani, M.A., M.B. B.S., M.Sc., Ph.D., F.R.C.P. University College London Respiratory and Department of Thoracic Medicine University College London Hospital London, England

References

- Martinez-Zayas G, Almeida FA, Simoff MJ, Yarmus L, Molina S, Young B, et al. A prediction model to help with oncologic mediastinal evaluation for radiation: HOMER. Am J Respir Crit Care Med 2020; 201:212–223.
- Navani N, Fisher DJ, Tierney JF, Stephens RJ, Burdett S; NSCLC Meta-Analysis Collaborative Group. The accuracy of clinical staging of stage I-IIIa non-small cell lung cancer: an analysis based on individual participant data. *Chest* 2019;155:502–509.
- Farjah F, Flum DR, Ramsey SD, Heagerty PJ, Symons RG, Wood DE. Multi-modality mediastinal staging for lung cancer among medicare beneficiaries. *J Thorac Oncol* 2009;4:355–363.
- Austin PC, van Klaveren D, Vergouwe Y, Nieboer D, Lee DS, Steyerberg EW. Validation of prediction models: examining temporal and geographic stability of baseline risk and estimated covariate effects. *Diagn Progn Res* 2017;1:12.
- Evison M, Crosbie P, Navani N, Callister M, Rintoul RC, Baldwin D, *et al.* How should performance in EBUS mediastinal staging in lung cancer be measured? *Br J Cancer* 2016;115:e9.
- Network NCC. Non-small cell lung cancer (Version 2.2019); 2018 [accessed 2019 Oct 1]. Available from: https://www.nccn.org/professionals/ physician_gls/pdf/nscl.pdf.
- O'Connell OJ, Almeida FA, Simoff MJ, Yarmus L, Lazarus R, Young B, et al. A prediction model to help with the assessment of adenopathy in lung cancer: HAL. Am J Respir Crit Care Med 2017;195: 1651–1660.
- Silvestri GA, Gonzalez AV, Jantz MA, Margolis ML, Gould MK, Tanoue LT, et al. Methods for staging non-small cell lung cancer: diagnosis and management of lung cancer, 3rd ed. American College of Chest

Physicians evidence-based clinical practice guidelines. *Chest* 2013; 143: e211S-e250S.

- El-Osta H, Jani P, Mansour A, Rascoe P, Jafri S. Endobronchial ultrasound for nodal staging of patients with non-small-cell lung cancer with radiologically normal mediastinum: a meta-analysis. *Ann Am Thorac Soc* 2018;15:864–874.
- Kaseda K, Asakura K, Kazama A, Ozawa Y. Risk factors for predicting occult lymph node metastasis in patients with clinical stage I nonsmall cell lung cancer staged by integrated fluorodeoxyglucose positron emission tomography/computed tomography. *World J Surg* 2016;40:2976–2983.
- Kanzaki R, Higashiyama M, Fujiwara A, Tokunaga T, Maeda J, Okami J, et al. Occult mediastinal lymph node metastasis in NSCLC patients diagnosed as clinical N0-1 by preoperative integrated FDG-PET/CT and CT: risk factors, pattern, and histopathological study. *Lung Cancer* 2011;71:333–337.
- Li L, Ren S, Zhang Y, Guan Y, Zhao J, Liu J, et al. Risk factors for predicting the occult nodal metastasis in T1-2N0M0 NSCLC patients staged by PET/CT: potential value in the clinic. *Lung Cancer* 2013; 81:213–217.
- Park HK, Jeon K, Koh WJ, Suh GY, Kim H, Kwon OJ, et al. Occult nodal metastasis in patients with non-small cell lung cancer at clinical stage IA by PET/CT. *Respirology* 2010;15:1179–1184.
- 14. Gould MK, Donington J, Lynch WR, Mazzone PJ, Midthun DE, Naidich DP, et al. Evaluation of individuals with pulmonary nodules: when is it lung cancer? Diagnosis and management of lung cancer, 3rd ed. American College of Chest Physicians evidence based clinical practice guidelines. Chest 2013;143:e93S–e120S.

Copyright © 2020 by the American Thoracic Society

Check for updates

a An Emotional Molecular Pathway in Pulmonary Hypertension–Alternative Complement System

The article by Frid and colleagues (pp. 224-239) in this issue of the Journal shows the significance of the alternative complement system in pulmonary hypertension (PH) (1). The discovery of this antimicrobial and immune-surveillance system has an emotional history (2). The classical pathway of complement activation by antibody-antigen complexes was discovered in the 1890s. Roughly half a century later, a brilliant biologist and biochemist, Dr. Pillemer, and his team discovered that the complement pathway could be activated by properdin in the absence of antibody-antigen complexes (3). Their discovery soon became very controversial; however, the Pillemer lab did not have the technical means to experimentally dispel all criticisms, and perhaps as a result of this controversy compounding existing mental issues, Dr. Pillemer died of suicide (2). His original observations were corroborated in the ensuing decades, and the pathway that he discovered was termed the alternative pathway (2). The alternative pathway is the ancient part of the complement system, with critical components present in insects and echinoderms (4). The lectin pathway was discovered later and represents yet another way to activate complement (5). Researchers in basic science are currently discovering further links between these pathways, such as those between the lectin and alternative pathways (6). What makes the complement system so important (5)? In a cascade of proteolytic activation steps, the complement factors C5b and C6-C9 form the lytic complex, which is an essential defense mechanism that can destroy microbes or faulty cells. In addition, many of the intermediary complement fragments (e.g., C3a and C5a) have major immune-regulatory effects.

Frid and colleagues investigated the role of complement cascade activation in regulating proinflammatory and

proproliferative processes during the initiation of experimental hypoxic PH and tested whether it can serve as a prognostic biomarker of outcome in human pulmonary arterial hypertension (PAH; Figure 1). The authors stained lung tissues from experimental PH models and patients with PAH, analyzed genetic murine models lacking specific complement components or circulating immunoglobulins, cultured human pulmonary adventitial fibroblasts, and performed a network medicine analysis of plasma from patients with PAH. Pulmonary perivascular-specific activation of the complement cascade was identified as a consistent critical determinant of PH/PAH in experimental animal models and humans. In experimental hypoxic PH, proinflammatory and proproliferative responses were complement (alternative pathway and C5) dependent, and immunoglobulins, particularly IgG, were critical for activation of the complement cascade in which Csf2/GM-CSF (granulocyte-macrophage colony-stimulating factor) was identified as a primary complement-dependent inflammatory mediator.

In their study, Frid and colleagues used omics to interrogate the association between the complement system and PAH. Omics studies are popular because of their potential for discovering new molecular disease mechanisms or new disease subgroups (for example, the pulmonary vascular omics network [7]). Here, the authors used omics to identify three classes of molecular deviations: 1) disease-causing genes (e.g., hemophilia caused by defective factor VIII or factor IX genes), 2) disease risk-increasing genes (e.g., increased risk of PAH in BMPR2 [bone morphogenic receptor 2] mutation carriers), and 3) molecular pathways that can be critical for disease pathogenesis (e.g., the complement pathway). The filters used to identify these three distinct molecular deviations have a decreasing stringency, with the third, molecular pathways that can cause disease, having the least-stringent discovery filter.

It is to be remembered that not all PH is caused by aberrant complement activation. C5-deficient mice can develop the PH phenotype. The FVB mouse strain is a popular transgenic

³This article is open access and distributed under the terms of the Creative Commons Attribution Non-Commercial No Derivatives License 4.0 (http://creativecommons.org/licenses/by-nc-nd/4.0/). For commercial usage and reprints, please contact Diane Gern (dgern@thoracic.org).

Originally Published in Press as DOI: 10.1164/rccm.201909-1790ED on October 10, 2019