



Regulatory B Cells and Its Role in Central Nervous System Inflammatory Demyelinating Diseases

Zhou Ran, Luo Yue-Bei, Zeng Qiu-Ming* and Yang Huan*

Department of Neurology, Xiangya Hospital, Central South University, Changsha, China

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*Correspondence:

Yang Huan zengqiuming1983@126.com Zeng Qiu-Ming 403850@csu.edu.cn

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Ran Z, Yue-Bei L, Qiu-Ming Z and Huan Y (2020) Regulatory B Cells and Its Role in Central Nervous System Inflammatory Demyelinating Diseases. Front. Immunol. 11:1884. doi: 10.3389/fimmu.2020.01884 Regulatory B (Breg) cells represent a population of suppressor B cells that participate in immunomodulatory processes and inhibition of excessive inflammation. The regulatory function of Breg cells have been demonstrated in mice and human with inflammatory diseases, cancer, after transplantation, and particularly in autoinflammatory disorders. In order to suppress inflammation, Breg cells produce anti-inflammatory mediators, induce death ligand-mediated apoptosis, and regulate many kinds of immune cells such as suppressing the proliferation and differentiation of effector T cell and increasing the number of regulatory T cells. Central nervous system Inflammatory demyelinating diseases (CNS IDDs) are a heterogeneous group of disorders, which occur against the background of an acute or chronic inflammatory process. With the advent of monoclonal antibodies directed against B cells, breakthroughs have been made in the treatment of CNS IDDs. Therefore, the number and function of B cells in IDDs have attracted attention. Meanwhile, increasing number of studies have confirmed that Breg cells play a role in alleviating autoimmune diseases, and treatment with Breg cells has also been proposed as a new therapeutic direction. In this review, we focus on the understanding of the development and function of Breg cells and on the diversification of Breg cells in CNS IDDs.

Keywords: regulatory B cells, central nervous system, inflammatory demyelinating diseases, multiple sclerosis, neuromyelitis optica

INTRODUCTION

The immune response feedback is an important mechanism that maintains the immune balance. Inflammatory diseases such as systemic lupus erythematosus (SLE), rheumatoid arthritis (RA) and multiple sclerosis (MS) are hallmarks of immunologic imbalances. As a major component of the immune system, B cells play both positive and negative roles in innate and adaptive immunity, through effector molecules such as antibodies and cytokines as well as through antigen-presention. On the one hand, B cells can mediate several negative processes such as amplifying immune responses.

Mechanistically, they differentiate into plasmablasts that secrete effector antibodies (1), may modulate effector T cell response through antigen presentation (2) and production of inflammatory cytokines (3). In addition, there is also a subset of B cells that regulates immune response to pathogens and autoantigens. These regulatory B cells are core targets in autoimmune and infectious diseases as well as cancer. These cells have a huge therapeutic potential against the aforementioned diseases. In one study performed in 1974 on delayed-type hypersensitivity, it was found that when B cells were removed, adoptively transferred splenocytes induced more intense reactions and lost their ability to suppress the delayed-type hypersensitivity reactions. This suggests that B cells or their products mediates inhibition of excessive inflammatory response (4). In another study conducted in 1996, it was found that mice with experimental autoimmune encephalomyelitis (EAE) but lacking B cells displayed greater differences in disease onset, severity, and recovery compared with the wild type group (5). Other studies on colitis and arthritis have demonstrated that B cells have antibody-independent immunoregulatory function (6, 7). Elsewhere, researchers have suggested that B cells inhibits excessive inflammation. B cells associated with inhibitory functions are referred to as Breg cells. IL-10 has been found to play a crucial role in the recovery of EAE (8). Other studies have further demonstrated that $IL-10^{-/-}$ mice display a non-remitting course of EAE, similar to the B cell-deficient mice (9). Combined, these findings suggest that B cells regulatory functions are mediated by IL-10. B cell-derived IL-10 has indeed been shown to play a key role in controlling autoimmunity (10). Accordingly, expression of IL-10 has been widely used to define suppressive B cell populations in mice and humans (11). B cells also regulate inflammation by a variety of IL-10-independent mechanisms (12).

Central nervous system Inflammatory demyelinating diseases (CNS IDDs) is a term referring to several CNS disorders, characterized by damaged myelin sheath of neurons, thus impairing transmission of signal by affected nerves. CNS IDDs can be differentiated based on disease severity and temporal courses, imaging, laboratory test and pathological characteristics. CNS IDDs mainly include MS, neuromyelitis spectrum disorders (NMOSD), and myelin oligodendrocyte glycoprotein antibodyassociated disease (MOG-Ab associated disease) (13). IDDs were considered to be primarily mediated by T lymphocytes. Given the success of therapeutic B cell depletion in MS (14) and NMOSD (15), there is growing concern on the role of B cells in the pathogenesis of IDDs. Studies on auto antibodies have improved our understanding of the role of B cells in the pathogenesis of immune-mediated diseases such as the appearance of oligoclonal IgG bands and deposition of IgG in the cerebrospinal fluid of MS, the presence of AQP4-IgG in NMOSD and antibodies against MOG in MOG-Ab associated disease. In addition, Breg cells also play a role in CNS IDDs. For instance, Breg cells deficiency is associated with severe symptoms of MS (16) and NMOSD (17), suggesting that Breg cells have the therapeutic potential to reduce immune-mediated inflammatory disorders. Subsequently, this review aimed at providing a summary of the current understanding on the development and function of Breg cells, and their role in the etiology of CNS IDDs.

DEVELOPMENT AND DIFFERENTIATION OF BREG CELLS

There are two distinct populations of B cells identified in mouse and human; the B1 and B2 subsets. Similar to other immune cells, B cells are derived from hematopoietic stem cells (HSCs), where they differentiate into progenitor B cells (Pro-B), precursor B cells (Pre-B) and immature B cells (Figure 1). Immature B cells undergo a "transitional" state, which is an early phase to the mature phenotype, after which they leave the bone marrow or fetal liver. B1 subset differentiates into mature B1a cells expressing CD5, and mature B1b cells. After stimulation with polysaccharides or lipids, mature B1 cells differentiate into antibody-secreting plasmablasts and short-lived plasma cells secreting antigen-specific antibodies. As for the B2 subset, they undergo three consecutive transitional B cells stages; transitional-1 (T1), transitional-2 (T2) and transitional-3 (T3). Transitional-B cells then migrates to the spleen and lymph node follicles, where they eventually differentiate into either follicular (FO) or marginal zone (MZ) B cells. The intermediate subset between T-2 B and MZ B cells are transitional-2 marginal-zone precursors (T2-MZP) B cells. Activated MZ B and FO B cells eventually differentiate into plasma cells, antibody producing B cells. Under special conditions, transitional B cells, MZ cells, T2-MZP cells, B1 cells, plasmablasts and plasma cells can all be activated to differentiate into Breg cells. Inflammatory microenvironment and intercellular interaction have been identified to activate this differentiation. Details of these processses will be discussed in the subsequent sections (Figure 2).

Inflammatory Microenvironment

The inflammatory microenvironment such as infiltration of cytokines and infection microenvironment can increase the expression as well as enhance the inhibitory property of Breg cells, implying that such factors play important roles in differentiation of Breg cells.

Numerous studies show that most inflammatory cytokines can indeed induce differentiation of Breg cells. As an immunosuppressive heterodimeric cytokine, IL-35 binds on its corresponding IL-35 receptor, activating signal transducer and activator of transcription (STAT) 1 and 3 to induce differentiation of resting B cells into IL-10 and IL-35-producing Breg cells. This suggests that IL-35 has the potential to induce autologous Breg cells as well as the treatment of autoimmune and inflammatory diseases (18, 19). More studies have demonstrated that IL-10-producing dendritic cells induced by IL-35 and phosphorylating STAT3 can induce immunosuppressive property of IL-10-producing B cells (20).

Similar to IL-35, IL-21 also induces production of IL-10 via phosphorylating STAT3. Accordingly, inhibition of phosphorylating STAT3 effectively blocks the production of IL-10 during the differentiation of the Breg cells. The effect of IL-21



differentiate into either mature B1a cells or mature B1b cells and eventually differentiate into plasmablast cells and plasma cells. Meanwhile, for the B2 subset, the transitional B cells differentiate either into follicular (FO) or marginal zone (MZ) B cells and eventually into plasmablast cells and plasma cells. Moreover, there is the transitional-2 marginal-zone precursors (T2-MZPs) B cells stage which is the precursor of MZ B cells. Studies have proved that the transitional B cells, MZ cells, T2-MZP cells, B1 cells, plasmablasts, and plasma cells can all differentiate into Breg cells with different phenotypes. Differentiation pathway of B10 cells in the middle of the photograph are listed separately because there are still not enough studies to classify them. In the dotted box, there are Breg cells with different phenotype and the source of them are still not clear.

on the differentiation of Breg cells strongly depends on additional signals including inhibitors of Toll-like receptors (TLR) and simulation of both B-cell receptor (BCR) stimulation and CD40 ligand. For instance, with the help of CD154 (CD40 ligand), IL-21 induces the differentiation of B cells into plasma cells or granzyme⁺ B lymphocytes (an important type of Breg cells) (21). In addition, the maturation of Breg cells into effector cells that secrete functional IL-10 requires homologous interactions with T cells mediated by IL-21 and CD40 (22).

Other inflammatory cytokines critical for differentiation of Breg cells include IL-1 β , IL-6 and granulocyte-macrophage colony-stimulating factor (GM-CSF). In mice with arthritis, deficiency of B cell specific IL-6 or IL-1 receptors is shown to exacerbate the disease compared with the controls (23). GM-CSF and IL-15 are strong immunosuppressive molecules that can

induce differentiation of naive CD19⁺ B cells into Breg cells, a process that can reverse the neuropathology of EAE (24).

Surprisingly, immune response to infectious diseases does not always worsen autoimmune diseases. In some instances, response to infectious diseases drives the development of IL-10producing Breg cells in both mice and humans. The helminth parasite *Schistosoma mansoni* contains TLR4 inhibitor, thus is able to induce secretion of IL-10 by B cells. This can then change the course of MS and reduce the severity of the disease (25). Similarly, *Mycobacterium tuberculosis* contains a TLR inhibitor, thus infection by this bacteria can aid in the recovery of EAE because it mediates the production of IL-10 by B cells. In a clinical trial, Bacillus Calmette-Guerin (BCG), a vaccine against tuberculosis disease, has been shown to alleviate clinically isolated syndrome (CIS) by reducing the number of lesions and



FIGURE 2 | Activation of Breg cells. Numerous mechanisms have been demonstrated to induce Breg cells, the most important of which are summarized as the inflammatory micro-environment and intercellular interactions. The inflammatory microenvironment is mainly produced by inflammatory factors, such as IL-35, IL-21, and other inflammatory cytokines. Cytokines often mediate activation of downstream signaling pathways by binding to cytokine receptors on the membrane. Microorganisms infection is another essential part of the inflammatory environment in the body, including invasive bacteria and commensal microbiota. Most microbial infections are activated by downstream molecular pathways mediated by membrane surface molecules, causing IL-10 transcription and translation and secretion. Intercellular interactions play a vital role in the induction of regulatory B cells, such as apoptotic cells or type3 innate lymphoid cells. Intercellular interactions mainly depend on the interaction of molecules on the surface of the membrane and the most classic surface molecules are Toll-like receptors (TLRs), CD40, B cell receptor (BCR). Many environmental and pharmaceutical factors can activate TLRs and cause downstream pathways activation, such as STATs and ERK orMyD88-NF-kB

FIGURE 2 | signaling pathway. CD40 can be activated by the CD40 ligand or binding immunoglobulin protein to mediate the activation of downstream signaling pathways STATs, and the downstream signaling pathways can be enhanced through synergy with other cell membrane surface molecules. BCR is thought to be involved in multiple functional processes of B cells, especially the induction of Breg cells. BCR combined with antigen can promote the release of calcium ions into the cell from the endoplasmic reticulum (ER) and also promote the activation of STIM on the endoplasmic reticulum, which open the calcium ion channels (SOC channel) on the cell membrane. A large amount of calcium influx increases the intracellular calcium ion concentration and promotes the phosphorylation of the nuclear factor of activated T cells (NFAT) in the downstream pathway, thereby increasing the transcription and translation secretion of IL-10. Other membrane surface molecules can promote Breg cell differentiation, such as Galectin-1and CD38. Research on regulatory B cell-specific transcription factors is still inconclusive, but the transcription factors such as Blimp1, IRF4, andHIF-1α have been shown to promote the transcription of IL-10.

improving long-term disease course (26). In MS, the severity of the disease significantly decreased after the reception with BCG vaccine (27). The underlying infection is not limited to invasive bacteria, but also includes the commensal microbiota in the intestines. These microorganisms have been shown to promote the differentiation of Breg cells in mesenteric lymph nodes and the spleen (23).

Intercellular Interaction

Intercellular interaction can also induce the differentiation of primary B cells into Breg cells, mainly through the activation of surface molecules on B cells (such as TLRs, CD40, BCR) and subsequent B cell downstream signaling pathway.

Gray M et al. found that apoptotic cells (ACs) affects the production of IL-10. This was demonstrated by injection of ACs into collagen-induced arthritis model, which induced the production of IL-10 by Breg cells, a process that alleviates inflammation (28). Gray M et al. also demonstrated the mechanism underlying secretion of IL-10 by B cells. Here, after recognizing the DNA containing complex on the surface of ACs, naturally occurring B cells (such as MZ B cells) bind and internalize the ACs surface chromatin complex, thereby activating TLR9 to regulate proliferation of B cells and secretion of IL-10 (29). Type 3 innate lymphoid cells (ILC3s) and innate B cells interact through IL-15 and B cell activating factors (BAFF), a process that promotes the development of ILC3s with CD40 ligand. CD40 positive ILC3s aid in the proliferation and differentiation of IL-10-secreting B cells. This mutually beneficial relationship between cells is important for maintaining immune tolerance, however, there are several deficiencies in this relationship in allergic asthmatic patients (30). By releasing IFN-a that interacts with CD40, dendritic cells can also drive the differentiation of immature B cells into IL-10-producing Breg cells. Conversely, Breg cells inhibits production of IFN-a by dendritic cells mediated by IL-10. In SLE, there are defects in this cross-talk, believed to be associated with abnormal activation of STAT1 and STAT3 (31).

TLRs are necessary for B cells to exert their inhibitory effects such as inhibition of inflammatory T cell responses and modulation of inflammation. TLRs-myeloid differentiation factor88 (MyD88) pathway is closely associated with the antiinflammatory immune mechanism. In mouse and human, the activation of TLR2, TLR4, and TLR9 transduction signal can induce production of IL-10 in B cells. For instance, trametes versicolor is a medicinal fungus that can promote differentiation of B cells into CD1d⁺ Breg cells in acute colitis, through the TLR2/4-mediated signaling pathway (32). Apart from chemical means, physical activation of B cells by factors such as ultraviolet radiation B has also been shown to induce differentiation of B cells into Breg cells. This process also suppresses the immune response through the TLR4-mediated signaling pathway (33). However, not all TLR stimulation can induce B cells to differentiate into Breg cells. For example, after activation through TLR7 and interferon- α , transitional B cells develop into pathogenic B cells, promoting the production of autoreactive antibodies (34). Studies on downstream mechanisms found that IFN-α can differentially regulate TLR7/8 and TLR9-activated STAT3 and ERK in B cells (35). More specifically, stimulation of B cells mediated by IFN-α and TLR7/8 inhibitors enhances phosphorylation of ERK1/2 and STAT3, which intern mediated production of IL-10 by B cells. Moreover, the activation of ERK and STAT3 is also important in TLR9- mediated IL-10 producing by B cells. However, IFN-a is not able to enhance the CpG-induced phosphorylation of ERK1/2 and STAT3 in B cells. MyD88 is a key downstream molecule in the inflammatory signaling pathway that also plays an important role in the regulation of cellular mediated immunity during infection (36). For instance, during Helicobacter felis infection, B cells activated by Helicobacter TLR-2 ligands can actuate IL-10-producing B cells in a MyD88 dependent manner (37). Endogenous TLR4 ligands are also found to be up-regulated, and activate B cells to produce IL-10 via TLR4-MyD88 signaling (38, 39). At transcription level, NF-kB plays an essential role in the inflammatory and immune response of cells, and the misregulation of NF-kB may cause autoimmune diseases, chronic inflammation and many types of cancer. Most importantly, with infectious diseases, activation of TLRs-MyD88-NF-kB can induce production of B cells specific IL-10 (40). On tumor research, one study found that activation of the TLRs-MyD88-NF-KB signaling pathway is necessary for Breg cells differentiation and the induced Breg cells with immunoregulatory functions can contribute to the suppression of anti-tumor immunity (41). This aside, IkBNS is a TLR-inducible nuclear IkB protein important in the TLRs-mediated IL-10 production in B cells. Mechanistically, IkBNS regulates inflammatory responses by inhibiting the induction of a subset of TLR-dependent genes through modulation of NF-κB activity (42). IκBNS-deficient B cells show reduced expression of Breg cells transcription factors including B lymphocyte induced maturation protein 1 (Blimp-1 protein) and interferon regulatory factor 4 (IRF4). They also fail to generate IL-10 producing CD138⁺ plasmablasts (a subset of Breg cells), suggesting that IkBNS is selectively required for IL-10 production in B cells, responding to TLR signals (43).

CD40, a membrane-associated protein, is a member of the tumor necrosis factor (TNF) receptor superfamily. The activation of CD40 on B cells not only induces the maturation of B cells into antibody-producing B cells, but is also crucial for the activation of Breg cells. In transgenic mice, ectopic expression of the CD154 is associated with increased CD40 signaling, which can in turn induce activation of the STAT3 pathway and an increase in the proportion of Breg cells (44). In experimental lupus, stimulation with CD40 inhibitor can induce IL-10-producing T2-like B cells to suppress Th1 responses and induce suppressive capacity to CD4⁺ T cells. This reduces the severity of the disease and delays IL-10 dependent progression of the disease (45). Furthermore, the synergistic effect of cell surface membrane molecules can activate B cells to perform their regulatory functions. For instance, co-stimulation of CD40 and TLRs has been shown to induce the highest proportion of IL-10 producing Breg cells, which plays a crucial role in recovery from MS relapse (46). Besides, an immunoglobulin protein (BIP), a member of the heat shock protein 70 family, can act in synergy with CD40 to induce the differentiation of Breg cells as well as suppress proliferation of T cells in a partially IL-10-dependent manner (47).

The BCR-STIM pathway is involved in most B cell function processes, such as activation, differentiation and antigen recognition, endocytosis, and presentation. BCR-stimulated B cells can maintain long-term tolerance and protect vulnerable mice from type 1 diabetes via an IL-10-dependent mechanism (48). At transcription level, many regulatory molecules are involved in IL-10 expression, including stromal interacting molecules (STIM) of endoplasmic reticulum (ER), nuclear factor of activated T cells (NFAT) family, IRF4 and a crucial cisregulatory element CNS-9 that is located 9kb upstream of the transcription start site of il10. After BCR activation, calcium sensory proteins STIM-1 and STIM-2 induce store-operated Ca²⁺ (SOC) influx and proliferation to increase the intracellular calcium concentration. This can activate the signaling pathway of NFAT family and mediate secretion of IL-10 (49). Without STIM1 or STIM2 proteins, B cells fail to produce IL-10 due to the defect in the activation of NFAT after BCR stimulation. Besides, the CNS-9 region contains clusters of NFAT and IRF binding motifs, which enhances the expression of IL-10 mRNA through the synergy effect of NFAT1 and IRF4. Therefore, deficiency of Irf4 specific to B cells impairs secretion of IL-10 and abnormal differentiation of Breg cell in dLNs. This intern predispose one to EAE (50).

Some other surface molecules such as CD38, Galectin-1 (Gal-1). may also play a role in the expression and function of Breg cells. For instance, CD38 is a transmembrane protein expressed in B lymphocytes. It can induce proliferation, differentiation or apoptosis of Breg cells. Currently, studies have found that the expression and effect of CD38 are inconsistent in different diseases. Some studies have demonstrated that CD38^{-/-} mice are more suitable for generating and expanding regulatory B10 cells than WT mice under appropriate stimulation (51). However, other studies have suggested that CD1d^{hi}CD5⁺ Breg cells highly expresses CD38, and in the presence of a CD38 stimulator, the percentage of Breg cells and their IL-10 production function increases (52). Galectin-1 (Gal-1) is a class of protein necessary for B cell development in the bone marrow. It plays a role in inducing B cell regulatory functions. Compared with wild-type B cells, Gal-1^{-/-} B cells have impaired IL-10 and Tim-1 expression, but with increased expression of TNF- α (53).

PHENOTYPES OF BREG CELLS

The activation process of immune cells is different in various disease states. Therefore, many surface markers used to identify Breg cells are either up- or down-regulated, which results in the non-uniform molecular expression of Breg cells. Studies in experimental animal models as well as in patients with autoimmune diseases have identified multiple subsets of Breg cells that exhibits diverse immune suppression mechanisms (see **Tables 1**, **2** for the function of various Breg cell subtypes in mice and humans). However, due to the intricate origins and activation pathways of Breg cells, there is an ever-increasing list of new phenotypic and functional markers associated with Breg cells.

Breg Cells in Mouse

Transitional-2 B Cells (CD1dhiCD21hiCD23hiCD24hi)

CD1d molecules are cell surface glycoproteins and the cytoplasmic tail of CD1d participates in signaling cascades associated with the transcription of IL-10 (82). In autoimmune diseases, the induced B-cell subpopulation is characterized by CD1d up-regulation, and the up-regulated CD1d can induce B-cell subpopulations to produce IL-10, promote antigenspecific regulatory T cell differentiation, and down-regulate inflammatory cascades associated with IL-1 upregulation and STAT3 activation (6). CD1d is expressed on a wide variety of cell types, of which three different B cell subsets express high levels of CD1d and have the potential to become Breg cells, including T2 B cells, MZ B cells and T2-MZP B cells. In the early stages of differentiation, B cells have already had the ability to differentiate into Breg cells. In SLE model, the adoptive transfer of T2 B cells can reverse autoimmunity and suppress the Th1 response (45). CD40 ligation halts the apoptosis of T2 B cells and prevents further differentiation into mature FO B cells (83), which can induce and expand the differentiation of IL-10⁺ T2 B cells. In allograft rejection model, T2 B cells isolated from tolerant mice show higher survival rates and inhibit cytokine production of T cells, thereby prolonging graft survival, suggesting that T2 B cells have the potential to treat allograft rejection (54).

MZ B Cells

(IgM^{hi}IgD^{lo}CD19⁺CD21^{hi}CD23⁻CD24^{hi}CD1d^{hi})

MZ B cells with CD1d high expression play a role in the prevention of autoimmunity through the production of regulatory cytokines and natural antibodies. Besides the polyreactive BCRs, MZ B cells also express high levels of TLRs, such as TLR9 which recognizes hypomethylated CpG motifs in bacterial DNA or chromatin complexes expressed on the surface of apoptotic cells (29). MZ B cells can differentiate into IL-10producing B cells and down-regulate the production of proinflammatory cytokines in response to stimulation of ligands or cytokines such as BAFF (84). Studies have demonstrated that through inflammatory stimuli, T-bet-expressing MZ B cells TABLE 1 | The phenotype and function of mouse regulatory B cell subsets.

| Name | Phenotype | Functions |
|-----------------------------|--|---|
| Transitional2 B cells | CD1d ^{hi} CD21 ^{hi} CD23 ^{hi} CD24 ^{hi} | 1.Supress the Th1 responses (45) |
| | | 2.Inhibit T cell cytokine production to prolong graft survival (54) |
| MZ B cells | lgM ^{hi} lgD ^{lo} CD19 ⁺ CD21 ^{hi} CD23 ⁻ CD24 ^{hi} CD1d ^{hi} | 1.Suppress CD8 and CD4 effector functions (55) |
| T2-MZP B cells* | CD1d ^{hi} CD21 ^{hi} CD23 ^{hi} lgM ^{hi} | Inhibit inflammatory cytokines production, suppress Ag-specific T cell activation and reduce in cells exhibiting Th1-type functional responses (56) |
| | | 2.Regulate both CD4 $^+$ CD25 $^-$ T cells and regulatory T cells (57) |
| | | 3.Promote tumor cell growth (58) |
| B10 cells | CD19 ⁺ CD1d ^{hi} CD5 ⁺ | 1.Suppress proliferation and differentiation of Th17 cells (59) |
| | | 2.Inhibit the production of Th1 cytokines, regulate the Th1/Th2 balance and maintain Treg cells (60) |
| | | 3.Inhibit microglial responses (61) |
| | | 4.Promote tumor cell proliferation (62) |
| B1a cells | CD19 ⁺ CD5 ⁺ | Inhibit macrophage proinflammatory responses and promote Treg cell responses (63) |
| | | Regulate neutrophil infiltration, CD4⁺ T cell activation and proinflammatory cytokine production (64) |
| | | Inhibit proinflammatory cytokine secretion, mediate CD4⁺ T cell apoptosis and prevent inflammation progressing (65) |
| | | 4.Negatively regulate anti-tumor immunity (66) |
| Plasmablast | CD138+CD44 ^{hi} | 1.Limit autoimmune inflammation and reduce disease severity (50) |
| Plasma B cells | IgM ⁺ CD138 ^{hi} TACI ⁺ CXCR4 ⁺ CD1d ^{int} Tim1 ^{int} | 1.Induce the Treg cell-mediated suppression (67) |
| | | 2.Activate macrophages to produce cytokines that reduce the anti-tumor immune response (68) |
| PD-L1 ^{hi} B cells | CD19 ⁺ PD-L1 ^{hi} | 1.Mediate the generation of regulatory T cells (69) |
| TIM-1 ⁺ B cells | TIM-1 ⁺ | 1.Promote Th2 responses and regulate immune tolerance (70) |
| | | 2.Promote the generation of regulatory T cells (71) |
| CD39+CD73+B cells | B220+CD39+CD73+ | 1.Suppress effector T cell functions (72) |
| | | |

*It is difficult to find a unified phenotype of T2 and T2-MZP B cells, so there are the most common and recognized cell phenotype of these two cells.

secrete IL-10, suggesting that T-bet might contribute to the remission of autoimmune diseases by activating the regulatory potential of MZ B cells (85). In collagen-induced arthritis model, MZ B cells produce most of IL-10 in response to TLR stimulation or apoptotic cells, and the adoptive transfer of MZ B cells could protect mice from infection (28). Study of Leishmania donovani infection have found that MZ B cells can interact with parasites to secrete IL-10 in a MyD88-dependent manner, and MZ B cells are involved in the suppression of CD8 and CD4 effector functions (55).

T2-MZP B Cells (CD1d^{hi}CD21^{hi}CD23^{hi}IgM^{hi})

Compared with MZ B cells, T2-MZP B cells also express highly CD1d but produce IL-10 in much larger quantities. It is difficult to find a unified phenotype of T2 and T2-MZP B cells, so the distinction between these two cells is worthy of further research and discussion. The immunomodulatory effects of T2-MZP B cells in a variety of immune-mediated pathologies including autoimmune diseases, allergy diseases and cancer. The regulatory function of T2-MZP B cells has been first demonstrated in experimental arthritis model, and the realization of its regulatory effect depends on IL-10 mediation, inhibition the production of inflammatory cytokines, suppression of Ag-specific T cell

activation, and reduction of Th1-type functional responses (56). Moreover, the regulatory effect of T2-MZP B cells can ameliorate the cellular infiltrates and the inflammatory damage by increasing Foxp3⁺ Treg cells and reducing the number of Th1 and Th17 cells (57). In Helicobacter felis infection model, T2-MZP B cells can induce the differentiation of T cells into a regulatory phenotype to ameliorate the inflammatory damage (37). In melanoma model, tumors initially signal via the lymphatic drainage to stimulate the preferential accumulation of T2-MZP Breg cells and this local response may be an early and critical step in generating an immunosuppressive environment to permit tumor growth and metastasis, suggesting T2-MZP B cells can promote tumor growth (58).

B10 (CD19+CD1dhiCD5+)

With the continuous expansion of the research scope, the exposure of B cells to different inflammatory environments had limited the use of CD1d markers to identify Breg cells. The co-expression of CD1d and CD5 on B10 cells has been therefore used to characterize the spleen B cell population. In mice, although B10 cells only account for about 1–2% of spleen B cells and 7–8% of peritoneal B cells, they are the main source of IL-10 production. Similar to B1a cells,

| Name | Phenotype | Functions |
|-------------------------------|---|---|
| Immature/Transitional B cells | CD19 ⁺ CD24 ^{hi} CD38 ^{hi} | 1.Suppress effector T cell but enhance Treg cell functions (44, 73) |
| Memory B cells/B10 cells | CD19 ⁺ CD24 ^{hi} CD27 ⁺ | 1.Induce Foxp3 expression on regulatory T cells (74) |
| | | 2.Have the value of evaluating the efficacy of biological drugs (75) |
| GrB ⁺ B cells | lgM ⁺ CD19 ⁺ CD38 ⁺ CD1d ⁺ CD147 ⁺ | 1. Inversely related to disease activity and clinical characteristics (76) |
| | | Negatively modulate Th1 and Th17 cells, induce T cell apoptosis and strongly suppress T cell proliferation (77) |
| | | 3.Suppress antitumor immune responses (78) |
| Br1 cells | CD25 ^{hi} CD71 ^{hi} CD73 ^{low} | 1.Suppress antigen-specific CD4 ⁺ T cell proliferation (79) |
| PD-L1 ^{hi} B cells | PD-L1 ⁺ | 1.Suppress pro-inflammatory cytokine production (80) |
| | | 2.Exhibit T cell suppressive capacity (69) |
| | | 3.Repress the proliferation and activation of CD8 ⁺ T cells (81) |

MZ B cells and T2-MZP B cells, B10 is able secrete a large amount of IL-10 and express similar surface markers such as CD19, CD1d, CD21, and CD24. However, each Breg cell type may have different stimulatory requirements for IL-10 production. In vitro, B10 cells stimulated via the TLR2 and TLR4 latter express cytoplasmic IL-10 at hour 5. Studies show that B10 cells have a regulatory function in suppressing immune responses such as IL-10-dependent regulation of T celldependent autoimmune responses (11). The adoptive transfer of B10 cells can suppress proliferation and differentiation of Th17 cells via the reduction of phosphorylating STAT3 and expression of retinoid-related orphan receptoryt (RORyt). This cascade of events delays the onset of inflammation and reduces clinical symptoms and inflammatory damage (59). In silicosis, a disease characterized by chronic lung inflammation and fibrosis, B10 can inhibit the production of Th1 cytokines, regulate the Th1/Th2 balance and maintain Treg cells (60). In viral infection, B10 cells can infiltrate the chronically infected brains and inhibit the microglial response (61). Generally, B10 cells are potent negative regulators of antigen-specific inflammation and T-celldependent autoimmune diseases. Therefore, the reinfusion of B10 cells to control disease progression may provide an effective treatment for both inflammatory and autoimmune conditions. B10 cells play a pro-tumorigenic role by promoting tumor cell proliferation. In pancreatic cancer, a recent research found that the bruton's tyrosine kinase signaling pathway can play a role in regulating differentiation of B10 cells, thereby controlling the cancer (62).

B1a Cells (CD19+CD5+)

B1a cells are another major source of IL-10, inhibiting the progression of both innate and adaptive immune responses, but at the cost of impeding pathogen clearance. The tissuespecific signals and unique pathogen-derived signals combine to determine whether the response of B1a cells is predominantly regulatory or proinflammatory. Gray M et al. found that in response to ACs, B1a cells can inhibit macrophage proinflammatory responses and promote Treg cell responses to self-antigens in an IL-10 dependent manner (63). In colitis model, IL-10 production by B1a significantly reduced disease severity by regulating neutrophil infiltration, $CD4^+$ T cell activation, and proinflammatory cytokine production during disease onset (64). In collagen-induced arthritis model, IL-10 produced by B1a cells inhibits proinflammatory cytokines secreted by activated macrophages and T cells in infectious lesions, and expressing Fas ligand (FasL) B1a cells can mediate $CD4^+$ T cell apoptosis and prevent inflammation progressing (65). In addition to secreting IL-10, in controlling immune homeostasis, B1a cells can also convert naive T cells into T cells with regulatory activity through cell-to-cell contact (86). In melanoma tumor immunity, B1a cells negatively regulate anti-tumor immunity by producing IL-10, suggesting they can be a target for immunotherapy of tumor (66).

Plasmablast (CD138⁺CD44^{hi}) and Plasma B Cells (IgM⁺CD138^{hi}TACI⁺CXCR4⁺CD1d^{int}Tim1^{int})

Studies have shown that in later stages of B-cell development such as plasmablasts and plasma cells can also produce IL-10 and have the inhibitory capacity. In autoimmune diseases, plasmablasts in the dLNs serve as IL-10 producers to limit autoimmune inflammation, while the absence of IL-10⁺ plasmablasts increases disease severity (50). In Salmonella Typhimurium infection, B cell-specific MyD88 signaling is essential for optimal development of IL-10-producing CD19+CD138+ B cells, especially in early stages of infection, and via MyD88 signaling, CD19⁺CD138⁺ B cells inhibit three key types of cells: neutrophils, natural killer cells, and inflammatory T cells (36). Similarly, plasma cells are also the major source of B-cell-derived IL-10 and IL-35 which can induce the Treg cell-mediated suppression (67). Plasma cells are found in the CNS of MS patients and the expression of IL-10 by plasma cells was necessary and sufficient to confer resistance toward inflammation, suggesting that plasma cells play an unexpected role in suppressing neuroinflammation (87). In hepatoma model, IgG-producing plasma cells activate macrophages to produce cytokines that reduce the anti-tumor immune response, while depletion of these plasma cells is able to prevent generation of activated macrophages, increase the anti-tumor T cell response, and reduce growth of tumor (68). As an essential regulator of plasma cell development, Prdm1 (encoding the Blimp-1 protein) is strongly correlated with IL-10 production, and during the formation of plasma cells, the prolonged elevation of Blimp-1 expression can elicit IL-10 production (88). Simon Fillatreau et al. identified a natural plasma cell subset characterized by the expression of the inhibitory receptor LAG-3, CD200, PD-L1, as well as PD-L2. Via a TLR-driven mechanism, natural regulatory plasma cells upregulate IL-10 expression within hours and without proliferating, suggesting that this group of plasma cells may be a potential disease treatment (89).

Other Breg Subsets

Programmed death ligand-1 (PD-L1) is important in controlling immune function, and promoting the proliferation of antigenspecific T cells. Besides this, programmed cell death receptor-1 (PD-1) binds to PD-L1 thereby transmitting inhibitory signals that reduce T cell proliferation. In RA, PD-L1^{hi} B cells can suppress disease development by elevating the expression of PD-L1. Presence of PD-L1 on B cells is positively correlated with Treg cells but negatively correlated with effector T cells, implying that PD-L1 mediates the generation of Tregs, an important molecule on B cells (69). T cell immunoglobulin mucin domain-1 (Tim)-1 is a membrane surface glycoprotein mainly expressed on cells, and is associated with regulation of immune responses. Apart from an inclusive marker for IL-10⁺ Breg cells derived from T2-MZP B cells, B10 cells and CD138⁺ B cells (90), Tim-1 is also critical for the induction and maintenance of Breg cells. Co-stimulation of IL-21, anti-Tim1 and CD40L can induce IL-10 activity in B10 cells and inhibit the progression of experimental periodontitis (91). TIM-1⁺ B cells strongly express IL-4 and IL-10, and promote Th2 responses, which can directly regulate immune tolerance (70). Conversely, B cells with Tim-1 defects are unable to produce IL-10 in response to ACs or by specific ligation with anti-TIM-1, or are unable to increase production of proinflammatory cytokine such as IL-1 and IL-6. This effect promotes Th1 and Th17 responses. In addition, B cells with defective Tim-1 can inhibit the generation of regulatory T cells and enhance the severity of autoimmune diseases (71). Collectively, these studies suggest that TIM-1 is critical in both the maintenance and induction of Breg cells under varied physiological conditions. CD39 and CD73 on their part are two ectoenzymes that together catalyze the dephosphorylation of adenine nucleotides to adenosine. Adenosine is known to suppress effector T cell function by binding on several adenosine receptors. Circulating B220+CD39+CD73+ B cells can drive a shift from an ATP-driven pro-inflammatory environment to an anti-inflammatory milieu induced by adenosine (72). A recent research found that decreased CD73 expression and the adoptive transfer of CD73⁺ B cells can impair production of adenosine, which can reduce the severity of colitis. This implies that CD39⁺CD73⁺ B cell adenosine can regulate autoimmune inflammation (92).

Breg Cells in Humans

In humans, Breg cells maintains immune homeostasis. Breg cells in both human and mice are predominantly identified based on their IL-10 producing property.

Immature/Transitional B Cells (CD19⁺CD24^{hi}CD38^{hi})

Previous studies have demonstrated that transitional B cells can exert IL-10 mediated inhibition of the expression of IFN-y and TNF- α in T cells (44). In healthy individuals, transitional B cells suppress proliferation of CD4⁺ T cell as well as release of pro-inflammatory cytokines, a function that partially mediated via the production of IL-10. However, in various autoimmune diseases such as primary Sjögren's syndrome (93) and diabetic nephropathy (94), there is an under production and defective functioning of transitional B cells, particularly during the active phase of the disease. In autoimmune diseases, defective transitional B cells have impaired IL-10 production upon activation via the CD40. Due to a defect in STAT3 phosphorylation (44), Breg cells are unable to suppress Th1 responses and fail to mediate differentiation of CD4⁺T cells into functionally suppressive Treg cells. This suggests that Breg cells may fail to prevent the development of autoreactive responses and inflammation. The higher number of transitional B cells in patients receiving rituximab is associated with long-term remissions, suggesting that the re-aggregation of Breg cells may be associated with better disease outcome (95). Apart from autoimmunity, transitional B cells have been shown to play a key role in establishing transplant tolerance (96). Breg cells can inhibit effector T cell function during the immune response in transplantation (73). In infectious diseases such as with viruses, enhanced production of IL-10-producing transitional B cells positively correlates with the viral load. The Breg cells were also shown to suppress virus-specific CD8⁺ T cell responses but enhances function of regulatory T-cells via the production of IL-10 and possibly expression of PD-L1. This suggests that transitional B cells may contribute to immune dysfunction in virus infection, which can hinder the elimination of the infection (97).

Memory B Cells/B10 Cells (CD19⁺CD24^{hi}CD27⁺)

Both memory B cells (also known as B10 cells in human) and transitional B cells are the major IL-10-producing B cells. They have similar functional characteristics such as suppressing proliferation of CD4⁺ T cells and inhibiting expression of proinflammatory cytokines. However, compared with transitional B cells, B10 cells have higher growth factor β (TGF- β) and shows stronger expression of granzyme B. B10 cells also express higher levels of surface integrins and CD39, suggesting the two Breg subsets have distinct functional characteristics (98). B10 cell subset additionally expresses high levels of TLR9, a receptor shown to be more sensitive to stimulation by CpG oligonucleotides. A proliferation-inducing ligand (APRIL) can stimulate signaling pathways activated by CpG (ERK1/2 and STAT3), which can induce an increase in the production of B10. This promotes production of IL-10 and induce expression of Foxp3 on regulatory T cells (74). Moreover, it is reported that miRNA-155 can regulate IL-10-Producing B10 cells in human by enhancing the expression of *il10* gene (99). Although B10 cells play a suppressive role, their function is altered differently in several autoimmune diseases such as Bullous pemphigoid (100). Moreover, B10 cells increases in RA patients treated with biopharmaceuticals, suggesting that B10 cells may represent a predictive biomarker for response to the treatment (75). In transplant-related diseases, the number of IL-10-producing $CD24^{hi}CD27^+$ B cells decreases. The function of the same cells is also impaired in graft-versus-host disease (cGVHD) (101). Similarly, in liver transplantation, patients that suffered from acute allograft rejection had significantly decreased proportions of B10 cells, but they dramatically increased after anti-rejection therapies (102).

Granzyme B⁺ B Cells (IgM⁺CD19⁺CD38⁺CD1d⁺CD147⁺)

Granzyme B(GrB) is a serine protease with several functions including antigen processing, matrix degradation, activation of inflammatory cytokines and immunoregulatory effects. GrB is dramatically elevated in chronic and inflammatory disorders. Secretion of GrB by B cells may play a significant role in early antiviral immune responses, regulation of autoimmune responses and in cancer immuno-surveillance. Studies show that co-stimulation of cytokines such as IL-21 and membrane surface molecules such as BCR, CD40 and TLRs induces B cells to differentiate into active forms that secrete cytotoxic serine protease (103). The immunoregulatory function of activated GrB⁺ B cells has been demonstrated in many human autoimmune diseases, suggesting that the impairment of this Breg cells subset is related to the pathogenesis of diseases (76). In RA, GrB-producing Breg cells are significantly decreased, and their proportion is negatively correlated with disease activity and clinical features. Moreover, the optimual level of these cells can be restored after effective therapy (104). Specifically, GrB⁺ B cells negatively modulate Th1 and Th17 cells, induce T cell apoptosis and strongly suppress T cell proliferation by downregulating the T cell receptor (TCR) zeta chain (77). Similar findings have been reported in tumor research. Other studies found that GrB⁺ B cells infiltrate tumor microenvironment and tumor-draining lymph nodes where they may participate in the suppression of antitumor immune responses (78). In transplantrelated diseases such as in renal transplantation, the affected patients showed a diminished level of GrB⁺ B cells compared to healthy controls (105). In general, GrB⁺ B cells may participate in early cell-mediated immune responses during inflammatory and neoplastic processes. Therefore, a better understanding of the role of GrB-secreting B cells in the immune system may help develop and improve new immunotherapy methods for infectious, autoimmune and malignant diseases.

Other Breg Subsets

Type 1 regulatory B (Br1) cells are characterized by CD25^{hi}CD71^{hi}CD73^{low}. They maintain peripheral blood tolerance by producing IgG4 antibodies (106). After receiving allergen-specific immunotherpay, the proportion of BR1 cells increases, secreting high levels of IL-10 (107). In autoimmune diseases, the function of Br1 is impaired while the inhibition of Th2 response islimited, implying that Br1 plays an important role in tolerance induction (79). Many Breg subtypes such as PD-L1^{hi} B cells are similar in both humans and mice. B cells can modulate T cell immune responses through the expression of regulatory molecules such as PD-L1. In autoimmune diseases,

CpG induces expression of PD-L1 on human B cells, which suppresses pro-inflammatory cytokines produced from antigenstimulated CD4⁺ T cells (80). PD-L1^{hi} B cells exhibiting T cell suppressive capacity are significantly decreased in untreated RA patients but normalize upon successful treatment (69). Besides, Tumor-infiltrating B cells that express high levels of PD-L1, IL-10 and TGF- β repress the proliferation and activation of CD8⁺ T cells (81).

BREG CELL EFFECTOR FUNCTIONS

Breg cells identified in both mice and human have been shown to downregulate inflammation associated with numerous pathological processes and the ability of each Breg cell subtype to negatively regulate immune responses as previously described. Generally, the functional mechanism of Breg cells is split into two parts; the immunomodulatory function by mediators produced by B cells and the immune effects mediated by surface molecules on B cells (see **Figure 3**).

The Immunomodulatory Function of Mediator Produced by B Cells

As the hallmark of Breg cells, IL-10 cytokine is commonly used as a marker for identification of Breg cells. As an anti-inflammatory cytokine, IL-10 plays a pivotal role in controlling excessive inflammation and downregulating the immune response. Plasmacytoid dendritic cells interact with Breg cells, where they drive the differentiation of immature B cells into IL-10-producing Breg cells. Here, plasmacytoid dendritic cells releases IFN- α that interacts with CD40 on B cells. Conversely, Breg cells inhibits the production of IFN-α from plasmacytoid dendritic cells by secreting IL-10. This cross-interaction is however compromised in autoimmune diseases (31). There is a regulatory feedback loop between macrophages or microglia and Breg cells. In viral infection, B10 cells are able to inhibit microglia secreted cytokines, in addition to modulating microglial cell responses within the infected lesion (61). Moreover, B cells can dampen the activation and influence the migration of macrophages by secreting IL-10 (108). Mutually, M2 polarized microglia can enhance the proportion of Breg cells to protects against hyperactive autoimmunity (109). As for the T cells, IL-10 inhibits secretion of cytokines by Th1, thus suppressing these cells. On the other hand, IL-10 enhances polarization of Th2, thus generating and maintaining the regulatory T cell pool (57).

In addition to producing IL-10, Breg cells produce other immune-regulatory cytokines such as TGF- β and IL-35. TGF- β is a pleiotropic cytokine involved in both suppressive and inflammatory immune responses that has been shown to suppress differentiation of Th1 by inhibiting the expression of STAT4, but promotes the development of Treg cells (110). IL-35 is the newest member of the IL-12 family. It is a potent anti-inflammatory cytokine secreted by Treg cells (111). In autoimmune diseases, IL-35 can induce production of IL-10 and IL-35 by Breg cells, which inhibits pathogenic Th1/Th17 cells (112). This prevents progression of the diseases and increases the proportion of Treg cells (18). with regard to immunoregulatory



intercellular contact. IL-10, as the main secreted inhibitory cytokine of regulatory B cells, has a variety of effects such as inhibiting the release of pro-inflammatory cytokines from immune cells, inhibiting the inflammatory differentiation of macrophage/microglia and promoting the conversion of T cells to regulatory T cells rather than effector T cells. Similarly, TGF- β , IL-35, IgG, and IgM can also regulate the differentiation of T cells into regulatory T cells. Membrane-bound molecules at the interface between regulatory B cells and T cells include CD80/86, CD40, MHC, FasL, PD-L1, and CD-1d, can regulate the differentiation of T cells into regulatory T cells into regulatory T cells and activate natural killer T (NKT) cells with suppressive function.

property of IL-35, this cytokin, as well as its derivatives have been shown to have a therapeutic potential against autoimmune and infectious diseases (113). Besides this function, B cells are best known for their ability to produce immunoglobulins, essential in the induction of protective immunity against many pathogens. IgM promotes the removal of apoptotic cells, phagocytosis by macrophages and modulates activation of pro-inflammatory signals through the FcyR (114). Moreover, in allograft rejection, Peter I Lobo et al. found that high levels of IgM minimizes the rejection rate of renal and cardiac allografts transplants in recipient individuals. This tolerance is partially mediated by inhibition of NF-kB translocation into the nucleus by blocking TLR4. This inhibition in effect impairs differentiation of activated T cells (115). Besides IgM, IgG can suppress overwhelming immune response, important in maintaining tolerance (116). In particular, IgG4, described as "anti-inflammatory IgG" has the ability to shorten compliment processes and reduce proinflammatory responses in natural immune cells (117).

The Immune Effects Mediated by Surface Molecules of B Cells

Immunity by cell dependent mechanism can mediate the inhibitory function of Breg cells, aided by several cell surface molecules on Breg cells. These molecules on the surface of Breg cells can induce the inhibition of immune cell function, promote differentiation of regulatory T cell and apoptosis of target cells.

CD40-activated B cells produce Foxp3⁺ Treg cells more efficiently compared with other antigen presenting cells. The longer contact time enables IL-10⁺ B cells to up-regulate expression of Foxp3 on CD4⁺ T cells, thus converts effector T cells into Treg cells. Breg cells can inhibit the proliferation of effector T cells via the CD40/CD40L interaction, suppressing autoimmune inflammation (118). Moreover, patients with the bare lymphocyte syndrome who do not express major histocompatibility complex class II (MHC II) molecules have impaired programmed cell death of autoreactive mature naive B cells, suggesting that tolerance to peripheral B cell is dependent on MHC II- TCR interaction (119). CD80 and CD86 molecules in the B7 family are expressed on antigen presenting cells and are important in establishing immune synapses and activating adaptive immune responses. Blocking antibodies against CD80 or CD86 partially reduces the regulatory effect of transitional B cells with regard to cytokine production, which implies that production of cytokines by B cells depends on interaction between B and T cells during antigen presentation (120). PD-L1 is another member of the B7 family with the ability to inhibit activation of T cell by binding to PD-1. PD-L1hi Breg cells negatively regulate differentiation of T cells. Adoptive transfer of PD-L1^{hi} B cells can inhibit EAE, demonstrating that B cells can activate Treg cells through PD-L1, thus suppressing immune responses (121). In addition, Breg cells can trigger pathogenic Th1 cells to undergo apoptosis through Fas-FasL interaction and effectively downregulate pathogenic immunity of these cells (122). CD1d expressed on MZB cells are recognized by TCR on NKT cells, thus can exert their regulatory functions by activating NKT cells (123). However, in autoimmune diseases, studies have found that the expression of CD1d on B cells in individuals with SLE patients is defective. This effect impairs the presentation and recognition of glycolipids presented by CD1d via the TCRs on NKT. Consequently, this inhibits the proliferation and activation of iNKT cells. In addition, some studies have found that the proportion of iNKT cells and the expression of CD1d in B cells in individuals with SLE who positively responded to rituximab treatment were significantly increased compared with patients who did not respond to the drug.

BREGS AND CENTRAL NERVOUS SYSTEM INFLAMMATORY DEMYELINATING DISEASES

Breg Cells and Multiple Sclerosis

MS is an autoimmune disease which affects \sim 2.5 million people worldwide. It is characterized by chronic inflammation of CNS and aberrant infiltration of inflammatory cells that eventually leads to demyelination and axonal damage. This disease is heterogeneous and complex, and thought to be caused by interactions of genetic and environmental factors (124). The disease was originally thought to be T cell-mediated, because activated T cells were abundantly present in MS lesions. In addition, EAE could also be induced by transfer of myelinreactive T cells. However, accumulating evidence on abnormal increase of immunoglobulin levels in the cerebrospinal fluid of affected patients, Antibody-deposition in brain lesions and the successful alleviation of the disease by B cell-depleting therapies shifted the focus to B cells as key players in immune-pathogenesis of MS. Breg cells also play a key role in demyelinating diseases of the nervous system. For instance, the occurrence of a radiologically isolated syndrome (RIS) or a CIS usually precedes MS. Further to this, individuals with RIS or CIS are more prone to develop MS within 6 months if they are deficient of IL-10producing B cells. This suggests that Breg cells have an inhibitory effect on the progression of MS (125, 126).

The B-cell-deficient mice failed to spontaneously recover from EAE. Interestingly, this was the first study to demonstrate that B cells modulate inflammation of the immune system (5). In addition, studies have shown that Breg cells are detected in CNS of EAE in a VLA-4 dependent manner, suggesting that Breg cells may contribute to regulation of CNS autoimmunity *in situ* (127). Recently, Simon Fillatreau et al. found that hypoxia-inducible factor-1 α (HIF-1 α) is a critical transcription factor for the production of IL-10 by B cells, and that HIF-1 α -dependent glycolysis facilitates increase in the proportion of CD1d^{hi}CD5⁺ B cells. Mice with B cells lacking Hif1 α have few IL-10-producing B cells, which exacerbates EAE (128).

When exploring the influence of external signals on the development and differentiation of immature B cells in the bone marrow, Simon Fillatreau et al. found that the bone marrow cells transiently stimulated by Toll-like receptor 9 can generate a new Breg cell subset CpG-proBs. CpG-proBs can slow down the development of EAE when transferred at the onset of clinical symptoms. Mechanistically, CpG-proBs can differentiate into mature Breg cells, trap T cells by releasing the CCR7 ligand and CCL19 and limit the immunopathogenesis of EAE through IL-10 production (129). Moreover, perforinexpressing regulatory B-cells (BRegs) are a new subset of Breg cells identified in patients with CIS and MS. BRegs exert their regulatory property on the disease by inhibiting proliferation of CD4⁺ T cells through the perforin/granzyme pathway (130). The proportion of BRegs in RRMS increased during relapse, suggesting that these new cells are associated with disease progression (131). Generally, the study on Breg cell subtypes and the mechanism with which they exert cell suppression have opened up interesting prospects on cell therapy during MS. Moreover, because some drugs used to treat MS patients were incidentally found to increase the production of IL-10 in human B cells, it provides promising prospect of regulating B cells for clinical treatment. As the first-line therapy against RRMS, IFN-β stimulates transformation of B lymphocytes into a subpopulation of regulatory transitional cells. Compared with untreated patients, the number of IL-10 producing transitional B lymphocytes in peripheral blood was significantly higher in IFN-B treated patients, demonstrating the role of IL-10-producing B cell populations in the disease therapy (132). Fingolimod is an immunosuppressant drug that modulates the sphingosine 1-phosphate receptor. It is the first oral, active disease-modifying drug approved for the treatment of MS. The proportion of IL-10 producing Breg cells is up-regulated while the migration capacity of the cells is enhanced after Fingolimod treatment (133). Alemtuzumab is another drug that inhibits monoclonal antibodies against CD52. It can restore the normal proportion of Breg cell subsets (CD19⁺CD24^{hi}CD38^{hi} cells and CD19⁺PD-L1^{hi} cells) in the peripheral blood of patients with relapsing MS patients. This suggests that CD19⁺CD24^{hi}CD38^{hi} and CD19⁺PD-L1^{hi} are promising candidate biomarkers for the efficacy of alemtuzumab therapy (134). Recently, siponimod, a selective sphingosine 1-phosphate receptor-1 and 5 modulator, was approved for active secondary progressive MS (SPMS). In one multi-centered randomized, double-blind, placebo-controlled clinical trial on SPMS patients, treatment with siponimod increased the number of transitional B cells and B1 cell subsets. In addition, the balance between Breg cells and memory B cells shifted in favor of Breg cells. Interestingly, it was a shift toward an anti-inflammatory and suppressive homeostatic immune state (135). However, in a phase 2 trial of atacicept (a recombinant fusion protein that suppresses B-cell function and proliferation), annualized relapse rates were higher in groups that received atacicept, compared to controls. One possible reason is that perhaps atacicept disrupt the B cell regulatory pathways but in turn stimulate T cell responses, a shift that sets in a proinflammatory environment and eventually relapses (136).

In conclusion, studies on the pathogenesis of MS have found that Breg cells regulates Th1/Th2 balance, induces apoptosis of effector T cells, neutralizes toxic substances, activates CD4⁺ T cells or natural killer cells (NK), inhibit the activation of dendritic cells and clear apoptotic bodies among many other functions. Further research on the Breg cell subtypes and underlying mechanism of Breg cells in MS can elucidate on the complex immune response in MS. This could provide a more comprehensive and systematic insight into its pathogenesis of the disease, and provide a basis for further exploration of new immunotherapy targets.

Breg Cells and Neuromyelitis Optica Spectrum Disorders

Neuromyletis optica sectrum disorder (NMOSD) is a rare autoimmune disease of the CNS that primarily attacks the optic neuritis and longitudinally extended transverse myelitis (137). Epidemiologically, the disease has a globally distribution, and is more common among young and middle-aged women and results in high disability rate. A major advancement that helped to distinguish NMOSD from MS was the discovery that 75% of patients with NMOSD have detectable serum IgG autoantibodies against the aquaporin-4 water channel (AQP4), an intergral water channel protein in astrocytes (138). After this discovery together with the pathogenic characteristics of the two diseases, it is believed that humoral mediated demyelination of astrocytes in the CNS is the major mechanism underlying the pathologenesis of the disease. The main features of the disease pathology can be reproduced using patient-derived monoclonal antibodies, thus reinforcing on the contribution of autoantibodies to the CNS injury associated with the disease (139). Moreover, studies on autoimmune diseases have found that impaired B cell tolerance potentially contributes to pathogenesis of the disease (140). In addition, accumulating evidence show that B cells play a vital role in NMOSD. This has been validated by B cell-targeted therapies such as rituximab, which have shown encouraging results for NMOSD (141).

The role of Breg cells in the pathogenesis of NMOSD has been extensively investigated. Some studies found that the proportion of Breg cells and expression of IL-10 are significantly lower in patients with NMOSD compared to those with MS, suggesting that the degree of impairment to B cell regulatory function can be considered as a distinctive marker between NMOSD and MS (142). CD19 $^+\mathrm{CD24^{hi}CD38^{hi}}$ Breg cells are less frequent in NMOSD patients positive for AQP4 antibodies than those without these antibodies. This phenomenon is also observed in CD19+CD5+CD1dhi Breg cells. In addition, NMOSD patients at acute relapse phase have lower IL-10 levels and significant impairment of CD19+CD24hiCD38hi Breg cells function (143). The advent of monoclonal antibodies has also provided a new direction in the treatment of NMOSD. For example, rituximab is an anti-CD20 chimeric monoclonal antibody shown to be well-tolerated, safe and efficient, with only minor risk of mild infusion reactions among NMOSD patients (141). After rituximab treatment, the functional balance between Breg cells and memory B cells inclines toward Breg cells as opposed to pro-inflammatory cytokines producing memory B cells (17). Tocilizumab on its part is a monoclonal antibody against IL-6 receptor shown to reduce relapse rate, neuropathic pain and fatigue in patients with NMOSD (144). Patients with autoimmune diseases treated with ocilizumab show an increased expression of TGF- β and CD25 molecule on the surface of B cells, reflective of activation of Breg cells (145). In summary, decrease in the proportion of Breg cells plays a role in the pathogenesis of neuroautoimmune diseases, and the number and function of Bregs can be restored after effective treatment.

Breg Cells and MOG-Ab Associated Demyelinating Disease

Myelin oligodendrocyte glycoprotein (MOG) is a glycoprotein located in the outer membrane of myelin, and is solely found within CNS, including in the brain, optic nerves and spinal cord. This implies that patients with encephalomyelitis attributed to MOG antibodies may develop bilateral optic nerve and lumbar spinal cord injury (146). Compared with NMOSD patients, patients with MOG antibody associated encephalomyelitis usually have a single course of the disease and show better recovery of neurological deficits after the attack. Based on clinical, immunological and histopathological evidence, encephalomyelitis associated with MOG antibodies has been regarded as a distinct disease entity different from MS and NMOSD (147).

In MOG antibody associated demyelinating disease, studies have demonstrated that the Breg cells such as CD19⁺CD24^{hi}CD38^{hi} and CD19⁺CD5⁺CD1d^{hi} B cells are numerically low and functionally impaired (148). Moreover, research found that IL-6 secreted by the dendritic cells promotes the differentiation of naïve CD4 T cells into TFH cells, whereas IL-21 secreted by TFH cells induce differentiation of B cells into memory B cells and plasma cells. The latter process results in the production of antibodies and disproportion of the memory B cells/Breg cells ratio. Imbalance between the memory B cells and Breg cells promotes pro-inflammatory cytokine responses that ultimately contributes to active demyelination (148). As the first-line therapy against NMOSD, methotrexate has also been found to extend the remission period as well as reduce the recurrence rate of the disease. It also results in minor side effects. Elsewhere, methotrexate has been shown to stimulate specific immune tolerance to auto-antigens mainly by enhancing secretion of B lymphocytes that produce effector IL-10 and TGF- β (149). Studies on the role of Breg cells in the pathogenesis of MOG antibody associated demyelinating diseases remain scanty, which creates the need to explore more potential therapeutic uses of Breg cells.

DISCUSSION AND PERSPECTIVE

Regulatory B cells are important modulators of the immune response and further promotes immune tolerance. From the early immature stage to the late plasma cell stage, Breg subpopulations have been found to evolve from different stages of B cell development. Due to the different microenvironment,

Breg cells have different phenotypes, but they all display immunomodulatory functions. Activation of Breg cells via BCR, TLR, or CD40 and cytokines has been shown to activate and expand the function of these cells, but mechanisms that can stabilize and maintain Breg cells remain elusive, thus there is need for further research on the plasticity and functional stability of Breg cells. In addition, numerous studies suggest that the number and function of Breg cells are involved in the pathogenesis of many diseases. Interestingly, the number and function of Breg cells among diseases and in different states are not exactly the same, which also suggests that the role of Breg cells in many disease pathologies is complicated. Therefore, it is necessary to expand our understanding on the mechanisms underlying activation, proliferation and precise functional mechanism of Breg cells in healthy individuals, as well as individuals with various immune, inflammatory or tumor diseases. Nonetheless, the important role of Breg cells in the pathogenesis of CNS IDDs has been revealed, and with the growing research on the function and contribution of Breg cells in the pathogenesis of autoimmune diseases, the therapeutic potential of Breg cells is gradually gaining acceptance. Although the therapy encompassing depletion of B-cells in treating autoimmune diseases achieved some success, this approach may exhausts Breg cells involved in the suppression of inflammation. Consequently, it would be advantageous to selectively increase Breg cells depending on the condition of the disease. In his study,

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Simon Fillatreau et al. suggested that reprogrammed quiescent of B cells is a novel tool for suppressing undesirable immune responses. This presents a noble research prospect for Breg cells (150). Substantially, treatment with Breg cells has certain theoretical feasibility and prospective clinical application, but its ultimate goal in the practice remains elusive.

CONSENT FOR PUBLICATION

Written informed consent for publication was obtained from all participants.

AUTHOR CONTRIBUTIONS

ZQ-M and YH conceived and planned the review. ZR wrote the manuscript. ZQ-M and LY-B critically revised the manuscript for important intellectual content. All authors contributed to the article and approved the submitted version.

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Conflict of Interest: The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

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