



ELSEVIER

Contents lists available at ScienceDirect

## Data in Brief

journal homepage: [www.elsevier.com/locate/dib](http://www.elsevier.com/locate/dib)



### Data Article

# Data set on the bioprecipitation of sulfate and trivalent arsenic by acidophilic non-traditional sulfur reducing bacteria



Letícia Paiva de Matos, Patrícia Freitas Costa, Mariana Moreira, Paula Cristine Silva Gomes, Silvana de Queiroz Silva, Leandro Vinícius Alves Gurgel, Mônica Cristina Teixeira \*

Federal University of Ouro Preto, Campus Universitário Morro do Cruzeiro, s/nº, Bauxita, 35400-000 Ouro Preto, Minas Gerais, Brazil

#### ARTICLE INFO

##### Article history:

Received 8 November 2017

Received in revised form

18 December 2017

Accepted 28 December 2017

Available online 2 January 2018

##### Keywords:

Arsenite

Sulfate reduction

Bioremediation

Immobilized cells

Acid pH

#### ABSTRACT

Data presented here are related to the original paper “Simultaneous removal of sulfate and arsenic using immobilized non-traditional sulfate reducing bacteria (SRB) mixed culture and alternative low-cost carbon sources” published by same authors (Matos et al., 2018) [1]. The data set here presented aims to facilitate this paper comprehension by giving readers some additional information. Data set includes a brief description of experimental conditions and the results obtained during both batch and semi-continuous reactors experiments. Data confirmed arsenic and sulfate were simultaneously removed under acidic pH by using a biological treatment based on the activity of a non-traditional sulfur reducing bacteria consortium. This microbial consortium was able to utilize glycerol, powdered chicken feathers as carbon donors, and proved to be resistant to arsenite up to 8.0 mg L<sup>-1</sup>. Data related to sulfate and arsenic removal efficiencies, residual arsenite and sulfate contents, pH and Eh measurements obtained under different experimental conditions were depicted in graphical format.

Refers to <https://doi.org/10.1016/j.cej.2017.11.035>

© 2018 The Authors. Published by Elsevier Inc. This is an open access article under the CC BY license (<http://creativecommons.org/licenses/by/4.0/>).

DOI of original article: <https://doi.org/10.1016/j.dib.2017.12.064>

\* Corresponding author. Fax: +55 31 3559 1069.

E-mail address: [teixeira@ef.ufop.br](mailto:teixeira@ef.ufop.br) (M.C. Teixeira).

<https://doi.org/10.1016/j.dib.2017.12.064>

2352-3409/© 2018 The Authors. Published by Elsevier Inc. This is an open access article under the CC BY license (<http://creativecommons.org/licenses/by/4.0/>).

## Specifications Table

Subject area	Chemistry, Biology, Engineering
More specific subject area	Biotechnology processes, Bioremediation.
Type of data	Table, image, graph, figure
How data was acquired	pH and Eh measurements: digital potentiostat with a combined platinum electrode (Digimed, DM-22). Residual sulfate concentration: turbidimetric method [2]. Total arsenic content: determined by inductively coupled plasma optical emission spectroscopy (ICP-OES) (Varian, 725-ES).
Data format	Analyzed. Averaged data
Experimental factors	Brief description of any pretreatment of samples
Experimental features	Culture media was prepared as previously described [1,3] and incubated in an oven (Fanem, model A-LT). Samples were centrifuged (10,000 rpm, 15 min, Fiberlite F155-8×50cy, Thermo, Multifuge X1R) and filtered (0.45 μm acetate cellulose membrane - Sartorius-Stedium) and acidified with nitric acid (50 μL) before residual As(III) measurements.
Data source location	Ouro Preto, Brazil
Data accessibility	All data are included in this document.
Related research article	L. P. Matos, P. F. Costa, M. Moreira, P. C. S. Gomes, S. Q. Silva, L. V. A. Gurgel, M. C. Teixeira, Simultaneous removal of sulfate and arsenic using immobilized non-traditional SRB mixed culture and alternative low-cost carbon sources, Chemical Engineering Journal, 334 (2018), 1630–1641.

## Value of the Data

- Different experimental conditions were compared. Free and immobilized bacterial cells were used. Different organic electron donors were tested including some low cost waste material.
- Data compare results obtained under batch and semi-continuous experimental conditions.
- Semi-continuous experiments were carried out for a long time. Data were collected for more than 150 days.
- Arsenite (bio)precipitation by sulfate reducing microorganisms was achieved under acidic pH.

## 1. Data

Data described simultaneous  $\text{SO}_4^{2-}$  and As(III) removal obtained by using a non-traditional SRB microbial consortium previously adapted to the growth under acidic pH using Glycerol and PCF as electron donors. The main bacterial species identified are: *Pantoea agglomerans*, *Enterobacter* sp., *Citrobacter* sp., *Cupriavidus metallidurans*, *Ralstonia* sp. and *Burkholderia cepacia*. Arsenic and sulfate are commonly found as contaminants in industrial effluents from mining and metallurgical industries.

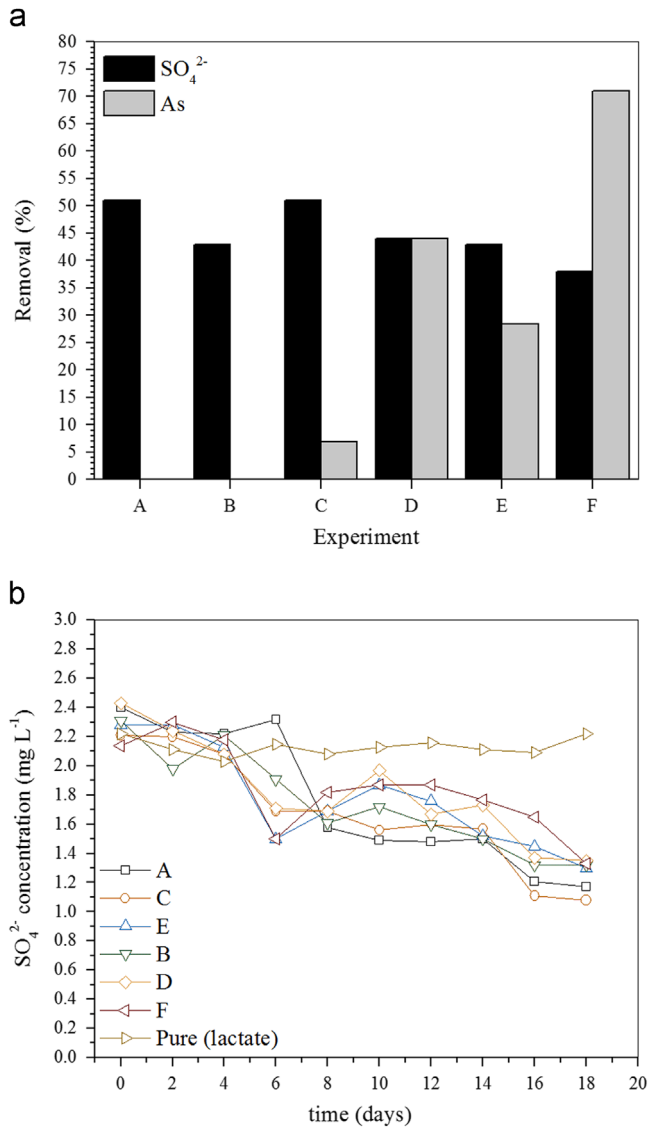
Arsenic and sulfate removal were obtained under batch and semi-continuous culture conditions. Semi-continuous up-flow reactors were constructed and operated for more than 150 days to prove their efficiency. At the final, effluent pH was neutralized and, depending on the operational conditions,  $\text{SO}_4^{2-}$  and As(III) ions were removed with 74.8% and 80% efficiency, respectively.

### 1.1. Batch experiments

Data presented in Figs. 1–4 are referred to experimental conditions summarized in Table 1.

### 1.2. Semi-continuous experiments

Figures 5 and 6 depict some results obtained during semi-continuous experiments. Experiments were carried out in bioreactors operated according conditions described in section 2.2.4.



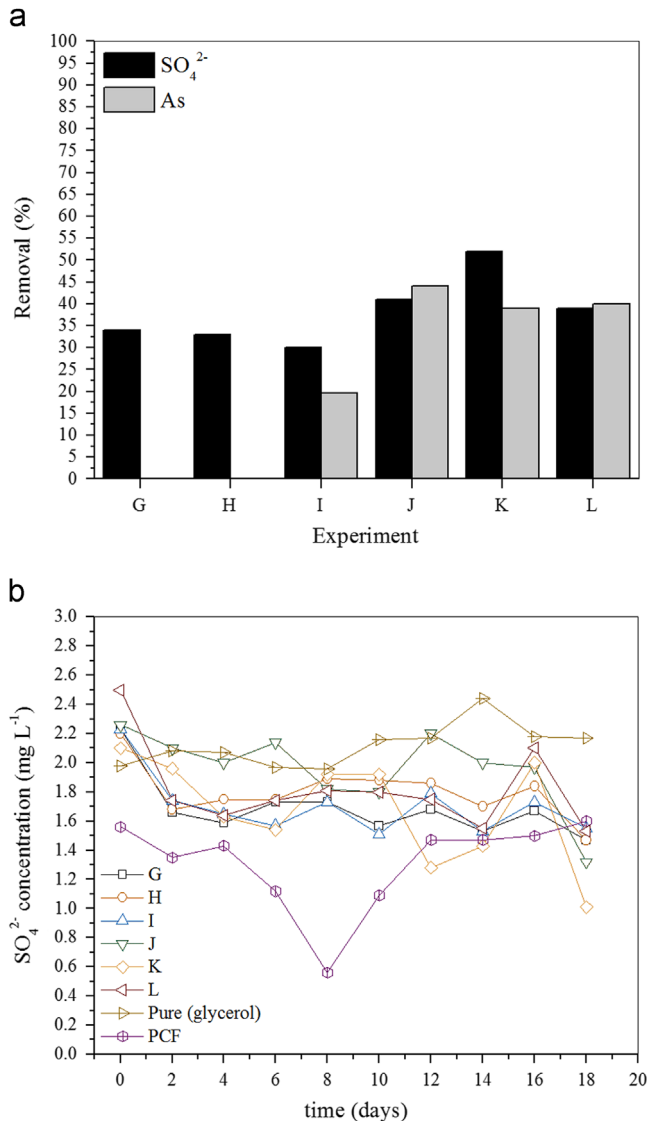
**Fig. 1.** a) Sulfate and As(III) removal (batch reactors) in acidic medium (pH 5.5) under different experimental conditions. Carbon source, sodium lactate; PCF (B, C, F); As(III), 0 (A, B), 4.0 (C, D) or 8.0 (E, F) mg L<sup>-1</sup>. b) Monitoring the sulfate removal (batch reactors) in acidic medium (pH 5.5) under different experimental conditions. Carbon source, sodium lactate; PCF (B, C, F); As(III), 0 (A, B), 4.0 (C, D) or 8.0 (E, F) mg L<sup>-1</sup>.

## 2. Experimental design, materials, and methods

Experimental design is detailed in Matos et al. [1]. All experiments and analyses were replicated twice and data were averaged.

### 2.1. Materials

Analytical grade reagents – Calcium alginate,  $\text{HNO}_3$ ,  $\text{HCl}$ ,  $\text{NaOH}$ ,  $\text{KH}_2\text{PO}_4$ ,  $\text{NH}_4\text{Cl}$ ,  $\text{Na}_2\text{SO}_4$ ,  $\text{MgSO}_4 \cdot 7\text{H}_2\text{O}$ ,  $\text{HCl}$ ,  $\text{FeSO}_4$ , sodium thioglycolate, ascorbic acid, ethylenediaminetetraacetic acid (EDTA), glycerol, sodium lactate - were purchased from different suppliers. A Brazilian poultry plant provided the powdered chicken feathers (PCF) used as solid support material and chemical substrate for



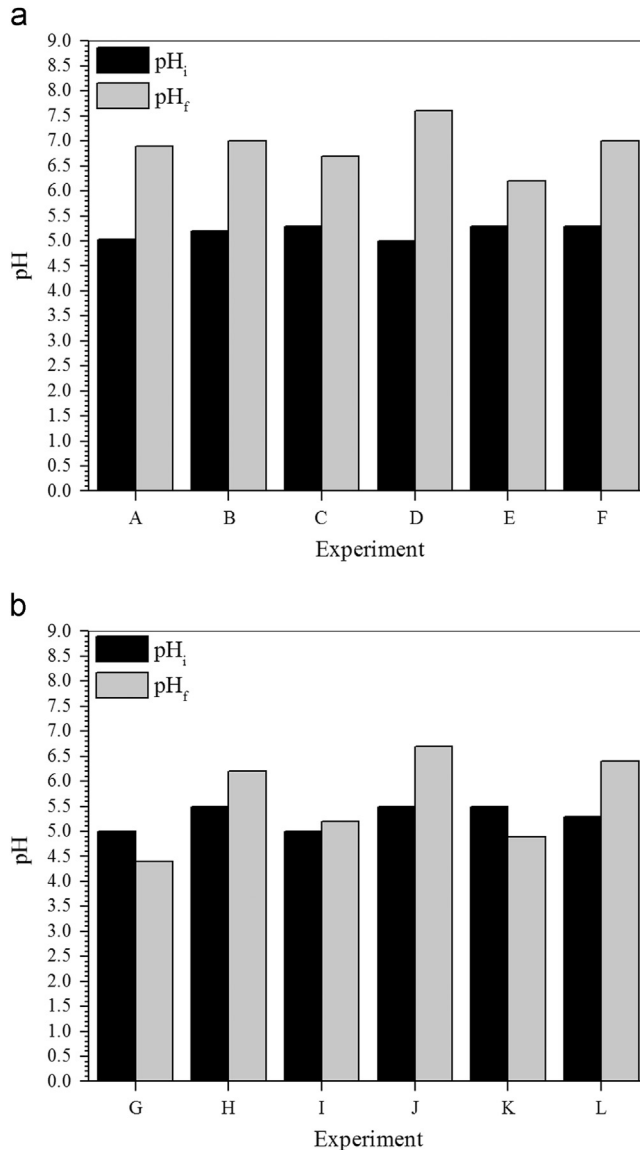
**Fig. 2.** a Sulfate and As(III) removal (batch reactors) in acidic medium (pH 5.5) under different experimental conditions. Carbon source, glycerol; PCF (H, J, L); As(III), 0 (G, H), 4.0 (I, J) or 8.0 (K, L)  $\text{mg L}^{-1}$ . b Monitoring the sulfate removal (batch reactors) in acidic medium (pH 5.5) under different experimental conditions. Carbon source, glycerol; PCF (H, J, L); As(III), 0 (G, H), 4.0 (I, J) or 8.0 (K, L)  $\text{mg L}^{-1}$ .

microbial growth. Distilled or tap (when mentioned) waters were used for preparing solutions and culture medium.

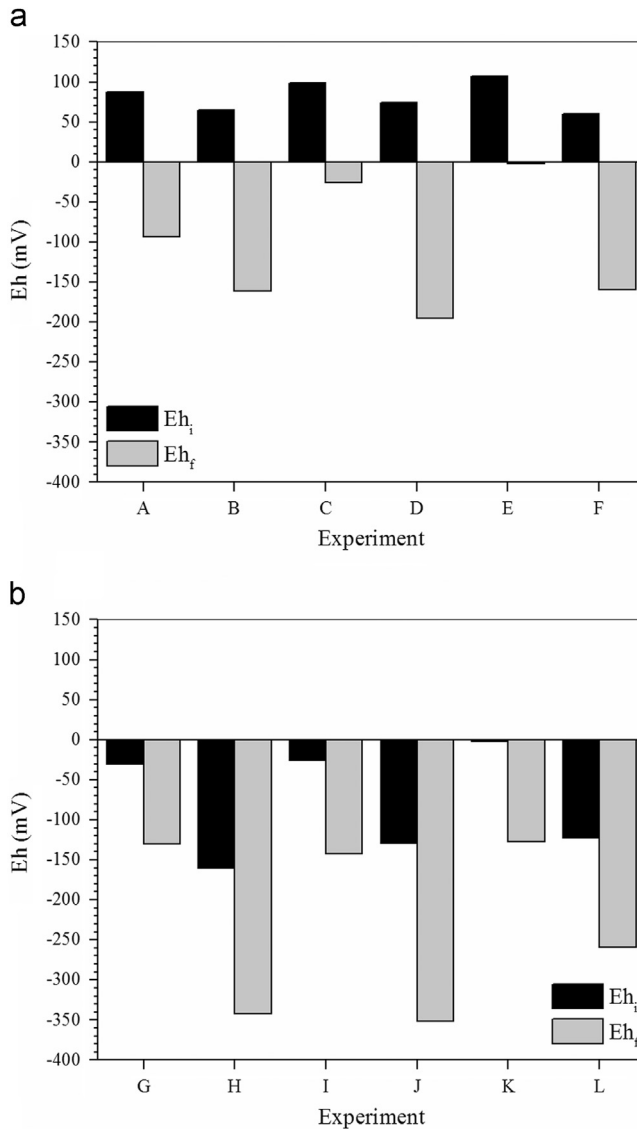
## 2.2. Methods

### 2.2.1. Microbial consortium

A microbial culture was obtained after enrichment of some sediment samples, collected from an urban pond, using modified Postgate C liquid medium [1,2]. Microbial culture was adapted to acidic



**Fig. 3.** a Changes in pH observed during the microbial growth (batch reactors) under different experimental conditions. Carbon source, sodium lactate; PCF (B, C, F); As(III), 0 (A, B), 4.0 (C, D) or 8.0 (E, F) mg L<sup>-1</sup>. b Changes in pH observed during the microbial growth (batch reactors) under different experimental conditions. Carbon source, glycerol; PCF (H, J, L); As(III), 0 (G, H), 4.0 (I, J) or 8.0 (K, L) mg L<sup>-1</sup>.



**Fig. 4.** a Changes in Eh (oxidation/reduction potential) observed during microbial growth (pH 5.5) under different experimental conditions. Carbon source, sodium lactate; PCF (B, C, F); As(III), 0 (A, B), 4.0 (C, D) or 8.0 (E, F) mg L<sup>-1</sup>. b Changes in Eh (oxidation/reduction potential) observed during microbial growth (pH 5.5) under different experimental conditions. Carbon source, glycerol; PCF (H, J, L); As(III), 0 (G, H), 4.0 (I, J) or 8.0 (K, L) mg L<sup>-1</sup>.

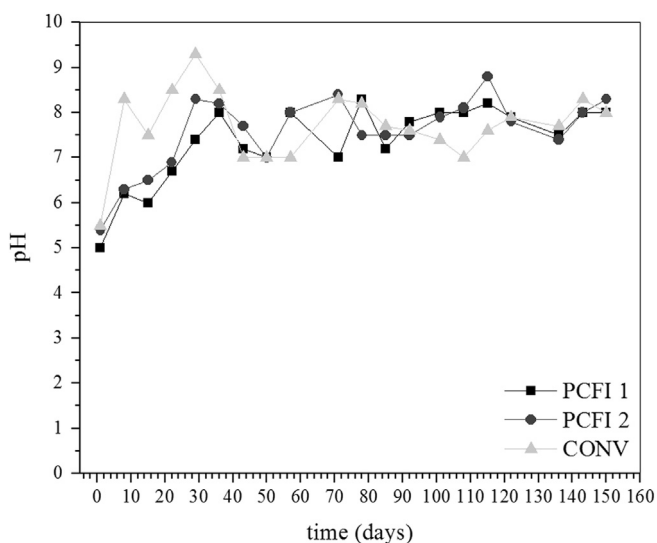
pH (5–5.5) and to the use of Glycerol as electron donor. Main identified microbial species were *Pantoea agglomerans*, *Enterobacter sp.*, *Citrobacter sp.*; *Cupriavidus metallidurans*, *Ralstonia sp.* and *Burkholderia cepacia* [1]. Microbial mixed culture was considered as a non-traditional sulfate reducing bacteria (SRB) consortium.

### 2.2.2. Culture parameters

Microbial growth and process efficiency were indirectly estimated by measuring arsenic and sulfate contents and pH and Eh (mV) changes as well.

**Table 1**  
Batch experiments – visual summary chart.

Experimental Condition	Medium Postgate B	Electron donor		PCF	As(III) (mg L <sup>-1</sup> )
		Sodium lactate	Glycerol		
A	Medium Postgate B	Sodium lactate	Glycerol	PCF	4.0
B					
C					
D					
E					
F					
G		Glycerol	PCF	8.0	
H					
I					
J					
K					
L					

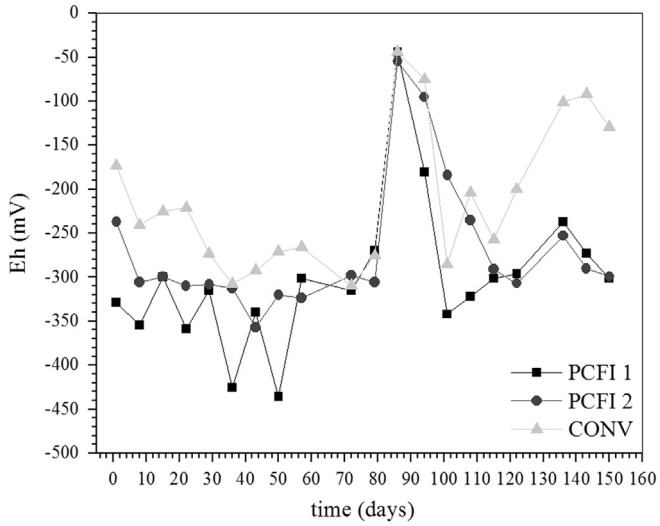


**Fig. 5.** Changes in pH during sulfate and arsenic removal in semi-continuous reactors. PCFI 1: sulfate, immobilized PCF and tap water, PCFI 2: sulfate, immobilized PCF and distilled water and CONV: modified Postgate B liquid medium and distilled water. Initial As(III) concentration of 8.0 mg L<sup>-1</sup>, pH 5.0 and 34 °C.

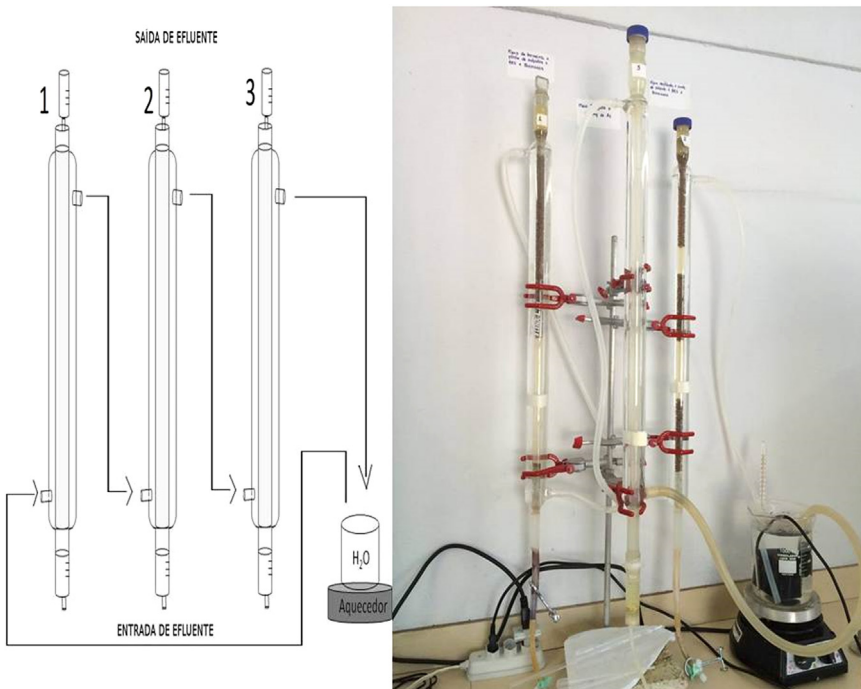
Both, batch and semi-continuous experiments were conducted using free and calcium alginate encapsulated microbial cells [1]. Postgate C liquid medium, at pH 5.5 was enriched with sodium lactate, glycerol and PCF as carbon sources. Culture medium pH was acidified to pH 5.5. Microbial tolerance to arsenic (4.0 and 8.0 mg of As(III) L<sup>-1</sup>) was accessed.

### 2.2.3. Batch experiments

Chemical oxygen demand (COD)-to-sulfate ratio used were 2.5 or 3.0, using sodium lactate or glycerol as main soluble carbon sources, respectively. Cultures were incubated in sterilized glass bottles (Postgate B medium, 473 mL, pH 5.0) containing a 5% (w v<sup>-1</sup>) inoculum, PCF (2%) and 4.0 or 8.0 mg L<sup>-1</sup> of As(III). An abiotic control flask was also compared. Flasks were sealed to minimize



**Fig. 6.** Changes in Eh during sulfate and arsenic removal in semi-continuous reactors. PCFI 1: sulfate, immobilized PCF and tap water, PCFI 2: sulfate, immobilized PCF and distilled water and CONV: modified Postgate B liquid medium and distilled water. Initial As(III) concentration of  $8.0 \text{ mg L}^{-1}$ , pH 5.0 and  $34^\circ\text{C}$ .



**Fig. 7.** Bioreactors PCFI1, CONV and PCFI2 - Visual aspect.



Oxygen dissolution and incubated at 35 °C for 360 h. 2 mL aliquots were withdrawn for residual sulfate concentration, pH and Eh measurements. Initial and final As(III) concentration were determined.

#### 2.2.4. Semi-continuous experiments

Four glass bioreactors (Fig. 7) were constructed as described by Matos et al. [1]. Bioreactors were supplied with Postgate C medium and inoculated with microbial consortium adapted to different electron donors and pH values. Sodium lactate and PCF (PCFF, CONV), glycerol and immobilized PCF (PCFI 1, PCFI 2, CONV) and free PCF (PCFF) were tested as carbon sources. Free (PCFF, CONV) or encapsulated (PCFI 1, PCFI 2) microbial cells were inoculated into the systems. Additionally, the suitability of using tap water (PCFI 1) instead of purified water (PCFF, PCFI 2, CONV) was tested. Operating temperature was  $34 \pm 2$  °C and bioreactors were supplied by an up-flow flux. Monitored parameters were pH, oxidation/reduction potential (Eh), residual sulfate and As(III) concentration. Bioreactors were operated during more than 150 days.

### Acknowledgments

This study was supported by Federal University of Ouro Preto - UFOP and National Council for the Improvement of Higher Education CAPES (undergraduate and graduate scholarships) and Brazilian National Research Council – CNPq (CNPq grant number 308861/2013-2) and Minas Gerais State Research Funding Agency -FAPEMIG (FAPEMIG grant numbers CRA APQ-01807-09 and APQ-02366-14) funding agencies.

### Transparency document. Supporting information

Supplementary data associated with this article can be found in the online version at <http://dx.doi.org/10.1016/j.dib.2017.12.064>.

### References

- [1] L.P. Matos, P.F. Costa, M. Moreira, P.C.S. Gomes, S.Q. Silva, L.V.A. Gurgel, M.C. Teixeira, Simultaneous removal of sulfate and arsenic using immobilized non-traditional SRB mixed culture and alternative low costs carbon sources, *Chem. Eng. J.* 334 (2018) 1630–1641.
- [2] A. Kolmert, P. Wikström, K.B. Hallberg, A fast and simple turbidimetric method for the determination of sulfate in sulfate-reducing bacterial cultures, *J. Microbiol. Methods* 41 (2000) 179–184.
- [3] P.F. Costa, L.P. Matos, V.A. Leão, M.C. Teixeira, Bioremoval of arsenite and sulfate by a mixed culture with sulfate-reducing capacity growing on powdered chicken feathers, *J. Environ. Chem. Eng.* 2 (2014) 70–75.