Characterizing skeletal muscle dysfunction in women with polycystic ovary syndrome

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Abstract: Polycystic ovary syndrome (PCOS) is the most common endocrine condition affecting women. It has traditionally been viewed as a primarily reproductive disorder; however, it is increasingly recognized as a lifelong metabolic disease. Women with PCOS are at increased risk of insulin resistance (IR), type 2 diabetes mellitus, non-alcoholic fatty liver disease and cardiovascular disease. Although not currently a diagnostic criterion, IR is a cardinal pathophysiological feature and highly prevalent in women with PCOS. Androgens play a bidirectional role in the pathogenesis of IR, and there is a complex interplay between IR and androgen excess in women with PCOS. Skeletal muscle has a key role in maintaining metabolic homeostasis and is also a metabolic target organ of androgen action. Skeletal muscle is the organ responsible for the majority of insulin-mediated glucose disposal. There is growing interest in the relationship between skeletal muscle, and rogen excess and mitochondrial dysfunction in the pathogenesis of metabolic disease in PCOS. Molecular mechanisms underpinning defects in skeletal muscle dysfunction in PCOS remain to be elucidated, but may represent promising targets for future therapeutic intervention. In this review, we aim to explore the role of skeletal muscle in metabolism, focusing particularly on perturbations in skeletal muscle specific to PCOS as observed in recent molecular and in vivo human studies. We review the possible role of androgens in the pathophysiology of skeletal muscle abnormalities in PCOS, and identify knowledge gaps, areas for future research and potential therapeutic implications. Despite increasing interest in the area of skeletal muscle dysfunction in women with PCOS, significant challenges and unanswered questions remain, and going forward, novel innovative approaches will be required to dissect the underlying mechanisms.

Keywords: androgen excess, insulin resistance, mitochondria, polycystic ovary syndrome, skeletal muscle

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Introduction

Polycystic ovary syndrome (PCOS) is a common and complex metabolic disorder of women, with a prevalence of 8–13%.¹ It is clinically defined by ovulatory dysfunction, androgen excess (clinical/ biochemical) and polycystic ovarian morphology on ultrasound. Androgen excess presents clinically in women with acne, hirsutism or androgenic alopecia, and biochemically is confirmed by the elevation of serum testosterone or other androgen precursors.² Insulin resistance (IR) and androgen excess are the cardinal pathophysiological features that underpin the clinical phenotype in PCOS, and are closely correlated clinically and biochemically.³ The precise direction of causality is unclear, however, and PCOS remains a poorly understood condition despite its high prevalence.

PCOS is increasingly recognized as a primarily metabolic disorder, and it is estimated that approximately 75% of women with PCOS have IR.⁴ It is hypothesized that skeletal muscle IR is a major pathophysiological player in driving the adverse metabolic phenotype in PCOS.^{5,6} Studies have reported reduced insulin-mediated glucose disposal (IMGD) in both lean^{7–9} and obese women with PCOS.^{10,11} In this review, we examine the role of skeletal muscle in metabolism, with

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a particular focus on perturbations in muscle function in PCOS. We also discuss a possible role for androgens in the pathophysiology of observed skeletal muscle abnormalities in PCOS and outline future potential research directions in this regard.

Skeletal muscle structure and function

Skeletal muscle mass compromises 40% of the total body mass and therefore is quantifiably the most abundant tissue mass in humans.¹² Skeletal muscle accounts for 30% of the metabolic rate in adults.^{12,13} Structurally, there are a number of fibre types that are defined as either slow or fast twitch based on their contractile properties. Classification of muscle fibre type also corresponds with histochemical staining for myofibrillar (myosin) ATPase as type I (slow-twitch) or type II (fast-twitch with highest ATPase activity). Oxidative metabolism is the highest in type 1 fibres, and they stain red because of the abundance of the oxygen transport protein myoglobin which is also related to the mitochondrial density.14 Similar to adipocytes, muscle fibres are heterogeneous but stable populations that can be regulated by hypertrophy and atrophy and even interconversion.¹⁵ Skeletal muscle is also a metabolic target of androgens. Androgens are known to increase muscle mass in both men and women, and are also associated with visceral adiposity in both sexes.16,17

Skeletal muscle energy metabolism

Seminal work by DeFronzo and others has highlighted the critical role of skeletal muscle in metabolic homeostasis and glucose metabolism.5,18,19 Skeletal muscle is the predominant site of glucose disposal under insulin control; termed insulinmediated glucose disposal. Skeletal muscle accounts for 80% of IMGD.20 Skeletal muscle is also the largest store of glycogen in the body and primary site for carbohydrate and lipid metabolism for energy production.²¹ Skeletal muscle is responsible for the bulk of fuel oxidation in the body, and has a striking capacity to rapidly modulate the rate of energy production, blood flow and substrate utilization adapting to numerous conditions, including the hormonal milieu and exercise, in both fed and fasted states.²² Notable enzymes include hexokinase II which phosphorylates glucose, glycogen synthase which controls glycogen synthesis, phosphofructokinase (PFK) which regulates glycolysis and pyruvate dehydrogenase (PDH) which regulates glucose oxidation.¹⁴ For this reason, there is an abundance of mitochondria, as muscle is heavily reliant on oxidative phosphorylation (OXPHOS) for energy production to maintain a continuous supply of ATP. Availability of ATP is modulated to accommodate increased need in times of increased demand, including exercise, in anaerobic conditions (phosphocreatine breakdown and glycogenolysis for substrate-level phosphorylation) and aerobic conditions (OXPHOS by utilizing reduced metabolites of carbohydrate and fat).^{14,23,24}

Regulation of metabolism occurs through interorgan crosstalk. Communication between adipose tissue, hepatocytes and skeletal muscle maintains metabolic homeostasis. Lipid metabolism and glucose homeostasis are highly interconnected as skeletal muscle utilizes both glucose and free fatty acids (FFAs) as fuel sources.^{25,26} The role of skeletal muscle metabolism in terms of glucose metabolism, fatty acid (FA) metabolism and the role of mitochondria will be further discussed below and graphically represented in Figure 1.

Key metabolic pathways

As shown in Figure 1, insulin binds to the tyrosine kinase transmembrane receptor which triggers auto-phosphorylation of the insulin receptor and insulin receptor substrate-1 (IRS-1). Phosphatidylinositol 3-kinase (PI3-K) activation follows, which is a critical insulin signalling node. One of the pathways for GLUT4 (glucose transporter 4) translocation is highlighted in this figure through Akt activation–induced activation of TBC1D4 (TBC1 domain family member 4), also known as Akt substrate 160 (AS160). Critical nodes in insulin signalling have been identified as the IRS, PI3-K and the Akt/protein kinase (PKB).²⁷

GLUT4 translocation to the plasma membrane allows for insulin-mediated glucose uptake into the myocyte. During insulin-stimulated states or fed states, most of the glucose entering the cells is stored as glycogen through the activation of glycogen synthase (GS). Insulin also stimulates protein synthesis effect through mTORC1 activations and inhibits protein breakdown through Forkhead box protein (FoxO) (not shown in Figure 1).



Figure 1. Key metabolic pathways in skeletal muscle. Insulin binding triggers auto-phosphorylation of the insulin receptor and subsequent auto-phosphorylation of IRS-1. Phosphatidylinositol 3-kinase (PI3-K) activation follows, which is a critical insulin signalling node. Akt activation-induced activation of TBC1D4, also known as AS160, initiates GLUT4 translocation to the plasma membrane. Insulin also stimulates the translocation of LCFA transporters to the plasma membrane, a downstream action of Akt, which promotes FA uptake by skeletal muscle while inhibiting FA release at adipocytes. Not shown here is that insulin stimulates protein synthesis effect through mTORC1 activation and inhibits protein breakdown mediated through FoxO. ACT, acyl-CoA transferase; ETC, electron-transport chain; FA, fatty acid; FATP, fatty acid transport protein; FABPP, fatty acid-binding protein; FoxO, Forkhead box protein O; GSV, Glut4 secretory vesicles; GLUT4, glucose transporter 4; IRS-1, insulin-responsive substrate-1; LCFA, long-chain fatty acid; MCT, monocarboxylate transporter; MT IM, mitochondria inner membrane; MT OM, mitochondria outer membrane; mTORC, mammalian target of rapamycin complex; P, phosphorylation; PDH, pyruvate dehydrogenase; PI(3)K, phosphatidylinositol-3-OH kinase; PIP3/2, phosphatidylinositol (3,4,5)-tris/diphosphate; TBC1D4, TBC1 domain family member 4, also known as Akt substrate 160 (AS160).

In contrast to the insulin-stimulated state, the fasted state relies on FFA for ATP synthesis. Long-chain fatty acids (LCFAs) are hydrophobic and can cross the plasma membrane, but there is now evidence of the role of membrane-associated FA-binding proteins. Insulin, through downstream activation of Akt, promotes FA update by skeletal muscle through the translocation of LCFA transporters to the plasma membrane and peripherally inhibits FA release at adipocytes. The metabolic status of the myocyte determines the fate of FFA. FFA can be stored once intracellular to monoglycerides, diacylglycerides (DAG) and then triglycerides (TAG) that are known as intramyocellular triglycerides (IMTGs).²⁸ Excessive lipid flux into skeletal muscle can influence



Figure 2. Proposed pathway disturbances in the pathogenesis of PCOS-related skeletal muscle dysfunction. Inter-organ crosstalk highlighting liver, adipose tissue, myokines, chronic inflammation and androgen excess with likely bidirectional implications for metabolic perturbations in PCOS. Intracellular androgen receptor binding with nuclear translocation of the androgen-bound AR receptor demonstrated. AMPK, adenosine monophosphate-activated protein kinase; CHO, carbohydrate; DAG, diacylglyceride; FFA, free fatty acid, IMTG, intramyocellular triglyceride; IR, insulin resistance; LCFA, long-chain fatty acid; PGC1a, peroxisome proliferator

coactivator 1a; PI(3)K, phosphatidylinositol-3-OH kinase; PKB/Akt, protein kinase B/Akt; TBC1D4, TBC1 domain family member 4, also known as Akt substrate 160 (AS160).

the accumulation of these lipid intermediates which correlates with reduced insulin sensitivity (IS).²⁹ Of these, diacylglycerol (DAGs) and ceramides are considered most metabolically active and implicated in potential muscle lipotoxicity.^{22,29–31}

The cellular powerhouse, the mitochondria, generates ATP through first β-oxidation of Acetyl-CoA which is an essential substrate for the common final pathway of fuel oxidation, OXPHOS, through the tricarboxylic acid (TCA) cycle.32 Mitochondrial dysfunction is of interest in metabolic disease, and there is much debate whether this is cause or consequence of IR, type 2 diabetes or obesity. Studies have shown a multitude of perturbations in mitochondrial function including increased reactive oxygen species leading to oxidative stress and mitochondrial damage and depletion, reduced mitochondrial size, downregulated OXPHOS pathways and a decrease in peroxisome proliferator coactivator 1a (PGC1a) activity, which is a master regulator of mitochondrial metabolism.33

In skeletal muscle, IR manifests as reduced insulin-mediated glucose uptake in the postprandial state and may be present decades before overt hyperglycaemia occurs.^{18,34} Skeletal muscle function is integrally linked with clinical and metabolic perturbations specific to PCOS, with inter-organ crosstalk graphically demonstrated in Figure 2.

Skeletal muscle dysfunction in PCOS

Poor metabolic health is recognized across the lifespan of women with PCOS.7,34,35 IR and PCOS are both independent risk factors for developing type 2 diabetes.³⁶⁻⁴¹ In PCOS, there is a well-established link between IR, impaired glucose tolerance, type 2 diabetes, non-alcoholic fatty liver disease (NAFLD) and cardiovascular disease (CVD).^{39,42-44} Circulating androgen burden is associated with an adverse metabolic phenotype, and correlates closely with indices of IR such as Homeostatic Model Assessment for Insulin Resistance (HOMA-IR) and insulin levels.45 A large-scale genome association study with participants from the UK Biobank has also recently demonstrated that the risk of type 2 diabetes is increased with increasing circulating testosterone concentrations.⁴⁶ Perturbations in

skeletal muscle energy metabolism appear to be a major player in mediating the adverse metabolic phenotype; understanding the molecular mechanisms underpinning PCOS-specific muscle dysfunction is therefore of critical importance.

IR and PCOS

Postprandial dysglycaemia is more common in women with PCOS than fasting dysglycaemia, and is reflective of a defect in insulin-mediated glucose uptake.47,48 Approximately, 75% of women with PCOS have IR, but this estimate varies across the literature.49 The variation in reported prevalence is likely a result of a number of factors: method utilized to assess IR, the population studied and also the changing diagnostic criteria for PCOS used in these studies.^{34,50} For example, different ethnicities may have a higher predisposition to IR.51 The changing definitions of PCOS over time also affect the interpretation of these phenotyping studies as the diagnostic criteria used introduce heterogeneity in the studied population of PCOS phenotype.52

Obesity has a strong association with IR, and every body mass index (BMI) increment raises the risk of T2DM by 10% for women with PCOS.^{53,54} While PCOS is associated with obesity in 50–60% of cases, IR is observed independent of BMI in this condition; IR and risk of diabetes are observed in both lean and obese women. Women with PCOS have significantly decreased *in vivo*–stimulated glucose disposal indicating IR.^{7,9} Fasting and dynamic measurements of IS indicate resistance in women with PCOS.^{9,55,56}

This decreased *in vivo* glucose disposal in PCOS women has been found to be independent of obesity or any alterations in fat-free mass.^{9,49,57} A systematic review of clamp studies in PCOS highlighted a 27% decrease in IS in PCOS patients, independent of BMI. This review included 28 studies with 38 study estimates, totalling 1224 participants with PCOS compared with 741 controls.⁴⁹ Cassar *et al.*⁴⁹ in this meta-analysis also found a relationship between total testosterone and IR, irrespective of the BMI.

Specific defects in insulin signalling in PCOS

Insulin signalling defects are recognized as proximal or distal in respect to Akt, with proximal defects understood as upstream of Akt. Earlier work on insulin signalling was carried out in cultured skin fibroblasts isolated from skin of women with PCOS.58 This research led to the hypothesis that proximal insulin signalling was impaired through increased phosphorylation of serine residues of the insulin receptor because of a serine kinase which limits progression of insulin signalling beyond the proximal protein of IRS1/2, although this abnormality was not universal in all samples.⁵⁸⁻⁶⁰ Dunaif et al.,⁶¹ investigating intact muscle in contrast to cultured myoblasts and adipose culture, demonstrated increased expression of IRS-2 but no change in expression IRS-1. In muscle, IRS-1 leads to GLUT4 translocation, whereas IRS-2 regulates downstream lipid metabolism.^{21,62,63} While there was no difference in the abundance of the insulin receptor or IRS-1 compared with muscle from controls in the Dunaif study, there was a significant decrease in the IRS-1-associated PI3-K activity in the same paper.⁶¹ Small reductions in the IRS-1 PKB response to insulin in intact muscle were also demonstrated by Hojlund et al.;64 interestingly, this signalling abnormality was reversed by treatment with insulin-sensitizing medications.

The IRS–PKB pathway is predominantly required for the metabolic effects of insulin such as glucose transport, gluconeogenesis and glycogen synthesis. The mitogenic effects of insulin are mitigated through the RAS-ERK (extracellular signal–regulated kinase) pathway. Corbould *et al.*⁶⁵ have highlighted increased basal phosphorylation of ERK in skeletal muscle of women with PCOS compared with controls, and this trend towards increased basal phosphorylation of ERK was also in agreement with a study by Rajkhowa.⁶⁶ Rajkhowa demonstrated that insulin reduced *in vivo* ERK activity.⁶⁶

Hansen *et al.*⁶⁷ have demonstrated perturbations in other glucose-regulating pathways. They focused on intrinsic PCOS factors and, in an attempt to avoid the confounding effect of obesity on IR, studied lean women with and without PCOS. Reduced expression and activity of AMPK (adenosine monophosphate–activated protein kinase) was found.⁶⁷ AMPK is a key regulator in the uptake of glucose into cells; AMPK is also linked to reduced circulating adiponectin levels.⁶⁷ This study also highlighted reduced metabolic flexibility in response to insulin, linked to reduced PDH.⁶⁷ In PCOS, many treatments have a focus on improving IS. Metformin, a biguanide, is commonly prescribed for women with PCOS and is a first line in the treatment of diabetes. Its exact mechanism of action has remained elusive, but interestingly it has been shown that it acts in an AMPK-dependent manner to modulate insulinmediated GLUT4 translocation. Metformin induces Rab4 expression via AMPK-AS160-PKC to increase insulin-mediated uptake, and AMPK knockdown inhibited this effect.68 Impaired insulin-stimulated phosphorylation of Akt and AS160 in skeletal muscle of women with PCOS has been shown to be partially but not completely reversible with peroxisome proliferator-activated receptor (PPAR) alpha therapy in the form of pioglitazone.64

Exercise is a core lifestyle recommendation for women with PCOS.⁶⁹ Exercise improves IS, insulin action and associated with increased skeletal muscle glucose uptake. Exercise improves IS in women with PCOS but does not normalize it.^{70,71} AMPK also plays a role in regulating glucose uptake during exercise; however, its role is not completely defined.⁷² Much of the available data about skeletal muscle metabolism during exercise are derived from studies in males. Females have a higher proportion of type 1 fibres. Females also have a higher reliance on IMTG during exercise which may be the reason women have higher IMTG content in type 1 fibres, with a lower capacity of glycolytic enzymes and a higher reliance on fat during exercise.73 This effect has been linked to oestrogen levels.73-75

In women with PCOS, Hansen *et al.* demonstrated that women with PCOS had an impaired ability to upregulate glucose handling proteins for insulin-mediated glucose uptake in muscle. Exercise improved IS in controls by increasing *GLUT4* and *HK2* gene and protein expressions, but this did not occur in PCOS women.⁷⁶

In a recent review, Stepto *et al.* have highlighted that advances in molecular mechanisms interrogating the perturbations in skeletal muscle in PCOS have to date created more questions than answers.³⁵ Dissecting mechanistic perturbations to date is hindered by the fact that most of the results are associative rather than causative.³⁵ Investigating intact muscle perturbations through *in vivo* studies is challenged by recruitment, expense and differing phenotypes of PCOS,

against a background of important confounders such as cardiorespiratory fitness, obesity and physical activity.^{49,35}

Metabolic inflexibility

The capacity of an organism to switch between oxidizing fat in the fasted state to carbohydrate in the fed state is termed metabolic flexibility.²² Metabolic inflexibility is well-documented in the setting of obesity, IR and type 2 diabetes. Its association with PCOS is also well-characterized.^{77–79}

While the molecular basis remains uncertain, it has been postulated that metabolic inflexibility implies the inability to appropriately activate glucose oxidation in response to glucose load; this may simply reflect a secondary consequence of impaired insulin action. In contrast to this theory, defects in mitochondrial fuel selection, independent of insulin signalling, could be a primary driver of IR, or it may occur as a result of oversupply of lipid fuel.⁸⁰ Metabolic inflexibility can lead to lipid accumulation in ectopic tissues such as skeletal muscle, driving lipotoxicity and exacerbating IR. In women with PCOS, ceramide has been increased threefold in one study with an overall 1.5-fold increase in IMTG compared with healthy controls.67

Metabolic inflexibility in PCOS is more pronounced with high BMI, worsening IR and hyperandrogenaemia.^{81,82} A recent systematic review has highlighted the association with hyperandrogenism, although the pathogenesis is unclear.⁸² There appears to be a common pathway in PCOS for inappropriate energy oxidation, metabolic inflexibility and impaired response to insulin in women with PCOS, but the role of androgen excess remains to be elucidated.^{82,83}

The role of androgen excess in skeletal muscle dysfunction

A direct causal relationship between androgen excess and IR in PCOS has been explored in a number of *in vivo* studies, with a focus on the effect of androgens on IS and whole-body glucose uptake through euglycaemic-hyperglycaemic clamp studies.⁴⁹ Abnormal insulin signalling is undoubtedly a feature of National Institutes of Health (NIH) and Rotterdam PCOS phenotypes with androgen excess, and may conceivably be milder in non-hyperandrogenic cohorts. In the

face of changing diagnostic criteria over time, abnormal insulin action is also still recognized in those with PCOS who have hyperandrogenism, polycystic ovaries and seemingly normal ovulatory function.⁸⁴ This IR may be milder than those with ovulatory dysfunction.⁸⁴ A recent population study has shown that women with PCOS exposed to combined oral contraceptive pills (COCPs) had a reduced risk of developing dysglycaemia across all BMI subgroups, raising the possibility of modulation of androgen action as a therapeutic tool to reduce metabolic dysfunction in PCOS.⁸⁵

Testosterone exposure can be amplified within target tissues. T is reduced by the intracellular enzyme 5-alpha reductase to dihydrotestosterone (DHT). DHT is considerably more potent at the androgen receptor than testosterone.86 Labrie et al.87 in 1988 coined the term intracrinology, which describes this mechanism of hormone precursor activation, action and inactivation within the peripheral target cells. At a cellular level, skeletal muscle cells exhibit intracellular 3-beta hydroxysteroid dehydrogenase (HSD), 17-beta-HSD and 5-alpha reductase activity resulting in the activation of dehydroepiandrosterone (DHEA) to T and DHT.86 AKR1 C3 (aldo-keto reductase type 1 C3), also known as 17 beta-HSD type 5, is an androgen activating enzyme, converting androstenedione (A4) to T, and has been proposed as an essential enzyme of androgen activation in skeletal muscle.88-91 Pre-receptor androgen metabolism has been extensively characterized in adipose tissue, but to date there is a relative paucity of data on androgen activation in skeletal muscle.86

There are a number of *in vivo* studies examining the role of androgen exposure on IS in women to understand a potential causative role for androgens in the pathogenesis of IR. Utilizing the hyperglycaemic-euglycaemic clamp, a significant reduction in whole-body IS has been demonstrated when short-term androgens were administered to women with normal menstrual cycles and BMI.92 Clamp studies were completed before and after at least 10 days of methyltestosterone.92 In one study, testosterone administration to patients undergoing female-to-male sex reassignment resulted in a significant decrease in IMGD measured by euglycaemic clamp studies before and after 4 months of treatment.93 Zang et al. studied testosterone administration to postmenopausal women before and after 3 months. They randomized women with average BMI 24.1 kg/m² to one of three groups: testosterone undecanoate, estradiol valerate or both. IS was significantly reduced in any of the testosterone groups without change in the oestradiol group.⁹⁴

Studies of anti-androgen treatment in hyperandrogenic women have also been shown to improve IS. Moghetti *et al.* implicated androgens in IR by highlighting the impact of androgen receptor blockade on IS before and after 3 months of antiandrogen treatment. They used three different anti-androgen treatments: spironolactone, flutamide and the gonadotropin-releasing hormone (GnRH) agonist buserelin to block androgen action. With all anti-androgen treatments, there was significant improvement in IS as measured by clamp in lean hyperandrogenic women before and after 3 months.⁹⁵

Intriguingly, 11-oxygenated androgens represent the majority of circulating androgens in women with PCOS, and these androgens do not decline with age in comparison with classic androgens and remain elevated after menopause.96 The question of whether these androgens contribute to metabolic risk and IR in PCOS is unresolved. The 11-oxygenated precursor androgens B11hydroxyandrostenedione (110HA4) and 11-ketoandrostenedione (11KA4) have been found to correlate with BMI, fasting insulin and HOMA-IR, but study results are conflicting and the same relationship with 11-ketotestosterone (11KT) and 11-hydroxytestosterone (11OHT) has not been found. Recently, Tosi et al.97 describe a positive relationship between 11-oxygenated androgens and IS, suggesting a relationship with IS that may diverge from that of classic androgens. This highlights the importance of future research to dissect these observations further.

Other mechanisms of PCOS-related muscle dysfunction

Myokines and PCOS

Skeletal muscle is increasingly recognized as an endocrine target organ.⁹⁸ Myokines are defined as cytokines and other peptides that are produced and released by muscle and exert autocrine, paracrine or endocrine effects.⁹⁹ Adiponectin, while a classical adipokine, is also now recognized as a myokine secreted from muscle with autocrine and paracrine effects.¹⁰⁰ Its effect on skeletal muscle includes metabolically favourable roles in the regulation of IS, metabolism, protein turnover, inflammatory signalling and myogenesis.¹⁰¹ Lower adiponectin levels have been associated with lower AMPK Thr172 phosphorylation in skeletal muscle.67 High-molecular-weight (HMW) adiponectin is inversely associated with intramyocellular lipid content detected by proton magnetic resonance spectroscopy in skeletal muscle.102 Adiponectin receptor 1 (AdipoR1) has a crucial role in skeletal muscle; adiponectin and AdipoR1 induce PGC-1a activation through Ca²⁺ signalling and by AMPK which leads to increase mitochondrial biogenesis.¹⁰³ After controlling for BMI-related effects, adiponectin levels are lower in women with PCOS compared with non-PCOS controls, likely related to IR.104 Androgen exposure can also cause a reduction in total circulating adiponectin by its inhibition of secretion from adipocytes.104-106

Irisin, a myokine discovered in 2012, is typically considered to have a positive association with energy metabolism; it is upregulated by exercise and downregulated in metabolic disease.¹⁰⁷ Its role in PCOS has recently attracted attention. Zhang et al.¹⁰⁸ demonstrated elevated irisin levels in women with hyperandrogenism and IR, while irisin levels were normal in women with PCOS without androgen excess. They postulated that elevated irisin levels may increase in a compensatory mechanism to metabolic stress at a molecular level in an attempt to counterbalance IR in women with hyperandrogenism and PCOS.¹⁰⁸ Li et al. also demonstrated irisin to be elevated in women with PCOS and androgen excess after adjusting for confounders such as BMI. Circulating irisin levels predicted hyperandrogenism, metabolic syndrome and IR in women with PCOS.¹⁰⁹ Overall, a meta-analysis in 2018 of eight studies did not observe any significant correlation between circulating irisin levels and IR in the PCOS population studied, and further studies are required to conclusively address this issue.¹¹⁰

Myostatin, a myokine belonging to the transforming growth factor beta (TGF- β) superfamily, negatively regulates muscle mass and growth. Myostatin also decreases insulin signalling and glucose uptake causing IR through nuclear factor kappa B (NF- κ B).¹¹¹ Inhibition of NF- κ B *in vitro* improves IR in L6-cultured muscle cell exposed to H₂O₂.¹¹² Chen *et al.*¹¹³ found that myostatin correlated with abdominal obesity and was increased in overweight women with PCOS. Further studies have not identified a difference in myostatin levels in women with or without PCOS.¹¹⁴ Obesity *per se* is associated with increased myostatin expression, and this has been demonstrated in muscle biopsies.¹¹⁵

Muscle mitochondrial dysfunction in PCOS

Abnormal mitochondrial function can affect whole-body metabolic homeostasis. Oxidative stress is considered to be a major contributor to IR in PCOS. Obesity is also a cause of oxidative stress; however, non-obese women with PCOS have increased oxidative stress, and therefore, this does not appear to be mediated by obesity alone.¹¹⁶ Reactive oxygen species–induced oxidative stress may also induce a pro-inflammatory state contributing to IR.^{117,118}

In women with IR and PCOS, genes involved in mitochondrial oxidative metabolism were down-regulated in skeletal muscle.¹¹⁹ Pioglitazone-induced improvements in IS were demonstrated in the same study cohort with increased expression of nuclear-encoded genes in skeletal muscle involved in mitochondrial phosphorylation pathways, OXPHOS. Pioglitazone increased the expression of PGC-1a in muscle of PCOS patients in this study cohort, and they postulated that this may occur *via* an increase in adiponectin which is an effect of thiazolidinedione (TZD) on muscle PPAR-y and an improved insulin action on mitochondrial biogenesis.^{120,121}

The PCOS muscle transcriptome and emerging role of microRNA

Women with PCOS have altered skeletal gene expression that is associated with specific changes in skeletal muscle DNA methylation patterns.¹²² Epigenetic changes are of increasing interest given genome-wide associated studies only explain a modest proportion of the heritability of PCOS, which was estimated at 70% from twin studies.^{123,124} Transcriptomic and epigenetic analysis show promise in delineating the perturbed molecular mechanisms in skeletal muscle IR in PCOS.

Nilsson *et al.*¹²² highlighted aberrant gene expression in a number of genes related to lipid, glucose metabolism and circadian rhythm that may be involved in the metabolic abnormalities of PCOS.

Decreased *COL1A1* expression correlated negatively with IR and interestingly *COL1A1* expression may be explained by hyperandrogenism in women with PCOS.¹²² Testosterone exposure *in vitro* decreased *COL1A1* expression in myotubes. This group also found that insulin regulated gene expression of Krüppel-like factor 10 control element (KLF10 or TGF- β -inducible early growth response protein 1). This is a DNA-binding protein which is already shown to be implicated in hepatic fibrosis *via* its binding with TGF- β control element.^{125,126}

There is increasing interest in skeletal muscle fibrosis and the aberrant gene expression in profibrotic pathways that have been demonstrated by Stepto et al.¹²⁷ The implication of these pathways in skeletal muscle IR is unknown but has been highlighted as a plausible contributor to IR; it is hypothesized that TGF-B ligand-mediated stromal deposition or fibrosis may be implicated in skeletal muscle in women with PCOS. This group demonstrated elevated gene expression in the TGF-β-regulated tissue fibrosis pathway. Specifically, changes have been noted in genes that encode extracellular matrix (ECM) components (COL1A2, COL3A1), enzymes in collagen deposition and assembly (LOX, DCN), ligands (TGFB2) and their receptor.127 Functional studies or proteomic analysis may advance this hypothesis in the future.128

Furthermore, in the study by Nilsen et al., COL1A1 was associated with increased DNA methylation of a CpG; DNA methylation at only two sites was highly correlated with differential gene expression in this particular study. The most overexpressed gene was noted to be DYRK1A that encodes an inhibitor of glycogen synthase kinase-3; this was not affected by in vitro androgen exposure, but 3 days of in vitro insulin exposure did reduce the cell response to insulin.¹²² NAMPT, a gene encoding visfatin, was upregulated in women with PCOS. Visfatin may have a role in mitochondrial biogenesis and has also been show to play a role in skeletal muscle uptake via the Ca2C-mediated AMPK-p38 mitogenactivated protein kinase (MAPK) pathway.129

Utilizing transcriptional profiling, it has been demonstrated that IR in skeletal muscle of women with PCOS is associated with altered gene expression. The authors reported reduced expression of genes involved in mitochondrial oxidative metabolism

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and postulated that reduced expression of peroxisome proliferator-activated receptor-y coactivator $(PGC)-1\alpha P$ could be implicated in this abnormality.¹¹⁹ PGC-1 α is a key regulator of mitochondrial biogenesis and also modulates remodelling of muscle to fibre-type composition, as well as participating in carbohydrate and lipid metabolism. Skov et al.¹¹⁹ indicated that these transcriptional abnormalities may contribute to the unique phenotype of IR in patients with PCOS. Exercise has positive effects on gene expression and epigenetic profiles in skeletal muscle of those with IR.130 Of interest, a study highlighted that electroacupuncture alters DNA methylation and could reverse PCOS-related transcriptomic disturbances in skeletal muscle, inducing metabolically advantageous alternations similar to those observed with exercise.131

There is also increasing interest in microRNA (miRNA) and PCOS development, and it is postulated that miRNA expression might be involved in the underlying pathophysiology of PCOS including glucose metabolism and IS.¹³² miRNA is short non-coding RNA molecules that are widely present in different tissue environments. MicroRNA plays critical role in gene regulation at post-transcription level and shows promise as potential biomarkers. Jing *et al.*¹³³ have identified three miRNAs (mrR-122, mrR-193b and miR-194) that are increased in women with PCOS and impaired glucose tolerance; there was also a modest correlation with HOMA-IR.

Conclusion

Although PCOS-specific mechanisms of skeletal muscle metabolic dysfunction have been identified, there is unlikely to be a single unifying molecular defect that underpins the adverse metabolic phenotype in this heterogeneous and complex disorder. Crosstalk between multiple contributory factors - including genetic, environmental, obesity and hormonal, coupled with intrinsic insulin signalling defects - is likely central to conferring the adverse phenotype observed. The role of androgen excess as a primary driver of some of these perturbations remains to be elucidated, and there is increasing interest in mitochondrial dysfunction as a potentially primary rather than secondary factor. Understanding the role played by androgen excess in PCOS-related skeletal muscle dysfunction, as well as the differential impact of classic and 11-oxygenated androgens in these processes, should be the focus of future research efforts. Novel and innovative approaches will be required to delineate these relationships.

Given the central role of skeletal muscle in IMGD, ameliorating IR at this site may effectively improve whole-body glucose homeostasis and may help abate the vicious cycle of hyperinsulinaemia and hyperandrogenaemia that is evident in PCOS. Future investigation into the role of skeletal muscle in PCOS and better understanding of the molecular basis may lead to future development of novel targeted therapies. The advent of 'omics'-based approaches such as genomics, transcriptomics, proteomics and metabolomics in large phenotyping studies may be helpful to further our understanding of skeletal muscle dysfunction and skeletal muscle IR in PCOS. It remains unclear whether modulation of defects in skeletal muscle energy metabolism, or indeed of local androgen exposure in muscle tissue, will live up to its potential as a novel target to reduce metabolic risk.

Declarations

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Consent for publication Not applicable.

Author contributions

Tara McDonnell: Data curation; Writing – original draft; Writing – review & editing.

Leanne Cussen: Writing – original draft; Writing – review & editing.

Marie McIlroy: Conceptualization; Supervision; Writing – review & editing.

Michael W. O'Reilly: Conceptualization; Funding acquisition; Supervision.

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References

- Azziz R, Woods KS, Reyna R, et al. The prevalence and features of the polycystic ovary syndrome in an unselected population. *J Clin* Endocrinol Metab 2004; 89: 2745–2749.
- Legro RS, Ehrmann DA, Hoeger KM, et al. Diagnosis and treatment of polycystic ovary syndrome: an Endocrine Society clinical practice guideline. *J Clin Endocrinol Metab* 2013; 98: 4565–4592.
- Franks S. Polycystic ovary syndrome. N Engl J Med 1995; 333: 853–861.
- Azziz R, Carmina E, Dewailly D, et al. Positions statement: criteria for defining polycystic ovary syndrome as a predominantly hyperandrogenic syndrome: an Androgen Excess Society guideline. *J Clin Endocrinol Metab* 2006; 91: 4237–4245.
- DeFronzo RA, Tobin JD and Andres R. Glucose clamp technique: a method for quantifying insulin secretion and resistance. *Am J Physiol* 1979; 237: E214–E223.
- 6. Azziz R, Carmina E, Chen Z, et al. Polycystic ovary syndrome. Nat Rev Dis Primers 2016; 2: 16057.
- Stepto NK, Cassar S, Joham AE, et al. Women with polycystic ovary syndrome have intrinsic insulin resistance on euglycaemic-hyperinsulaemic clamp. Hum Reprod 2013; 28: 777–784.
- Micic D, Sumarac-Dumanovic M, Kendereski A, et al. Total ghrelin levels during acute insulin infusion in patients with polycystic ovary syndrome. *J Endocrinol Invest* 2007; 30: 820–827.
- Dunaif A, Segal KR, Futterweit W, et al. Profound peripheral insulin resistance, independent of obesity, in polycystic ovary syndrome. *Diabetes* 1989; 38: 1165–1174.
- 10. Hutchison SK, Stepto NK, Harrison CL, *et al.* Effects of exercise on insulin resistance and body composition in overweight and obese women with

and without polycystic ovary syndrome. *J Clin Endocrinol Metab* 2011; 96: E48–E56.

- Eriksen M, Porneki AD, Skov V, et al. Insulin resistance is not conserved in myotubes established from women with PCOS. PLoS ONE 2010; 5: e14469.
- Zurlo F, Larson K, Bogardus C, *et al.* Skeletal muscle metabolism is a major determinant of resting energy expenditure. *J Clin Invest* 1990; 86: 1423–1427.
- Westerblad H, Bruton JD and Katz A. Skeletal muscle: energy metabolism, fiber types, fatigue and adaptability. *Exp Cell Res* 2010; 316: 3093– 3099.
- 14. Egan B and Zierath JR. Exercise metabolism and the molecular regulation of skeletal muscle adaptation. *Cell Metab* 2013; 17: 162–184.
- Schiaffino S and Reggiani C. Fiber types in mammalian skeletal muscles. *Physiol Rev* 2011; 91: 1447–1531.
- Lovejoy JC, Bray GA, Bourgeois MO, et al. Exogenous androgens influence body composition and regional body fat distribution in obese postmenopausal women–a clinical research center study. J Clin Endocrinol Metab 1996; 81: 2198–2203.
- Bagatell CJ and Bremner WJ. Androgens in men–uses and abuses. N Engl J Med 1996; 334: 707–714.
- DeFronzo RA and Tripathy D. Skeletal muscle insulin resistance is the primary defect in type 2 diabetes. *Diabetes Care* 2009; 32: S157–S163.
- DeFronzo RA, Jacot E, Jequier E, *et al.* The effect of insulin on the disposal of intravenous glucose results from indirect calorimetry and hepatic and femoral venous catheterization. *Diabetes* 1981; 30: 1000–1007.
- DeFronzo RA, Hendler R and Simonson D. Insulin resistance is a prominent feature of insulin-dependent diabetes. *Diabetes* 1982; 31: 795–801.
- 21. Sylow L, Tokarz VL, Richter EA, *et al.* The many actions of insulin in skeletal muscle, the paramount tissue determining glycemia. *Cell Metab* 2021; 33: 758–780.
- Goodpaster BH and Sparks LM. Metabolic flexibility in health and disease. *Cell Metab* 2017; 25: 1027–1036.
- Lundsgaard AM, Fritzen AM and Kiens B. Molecular regulation of fatty acid oxidation in skeletal muscle during aerobic exercise. *Trends Endocrinol Metab* 2018; 29: 18–30.

- 24. Hargreaves M and Spriet LL. Skeletal muscle energy metabolism during exercise. *Nat Metab* 2020; 2: 817–828.
- Groop LC, Bonadonna RC, Simonson DC, et al. Effect of insulin on oxidative and nonoxidative pathways of free fatty acid metabolism in human obesity. Am J Physiol 1992; 263: E79–E84.
- Abdul-Ghani MA and DeFronzo RA. Pathogenesis of insulin resistance in skeletal muscle. J Biomed Biotechnol 2010; 2010: 476279.
- Taniguchi CM, Emanuelli B and Kahn CR. Critical nodes in signalling pathways: insights into insulin action. *Nat Rev Mol Cell Biol* 2006; 7: 85–96.
- Morales PE, Bucarey JL and Espinosa A. Muscle lipid metabolism: role of lipid droplets and perilipins. J Diabetes Res 2017; 2017: 1789395.
- Krssak M, Falk Petersen K, Dresner A, et al. Intramyocellular lipid concentrations are correlated with insulin sensitivity in humans: a 1H NMR spectroscopy study. *Diabetologia* 1999; 42: 113–116.
- Brioche T, Pagano AF, Py G, et al. Muscle wasting and aging: experimental models, fatty infiltrations, and prevention. *Mol Aspects Med* 2016; 50: 56–87.
- Samuel VT, Kf P and Shulman GI. Lipidinduced insulin resistance: unravelling the mechanism. *Lancet* 2010; 375: 2267–2277.
- Watt MJ and Hoy AJ. Lipid metabolism in skeletal muscle: generation of adaptive and maladaptive intracellular signals for cellular function. *Am J Physiol Endocrinol Metab* 2012; 302: E1315–E1328.
- Montgomery MK and Turner N. Mitochondrial dysfunction and insulin resistance: an update. *Endocr Connect* 2015; 4: R1–R15.
- 34. Diamanti-Kandarakis E and Dunaif A. Insulin resistance and the polycystic ovary syndrome revisited: an update on mechanisms and implications. *Endocr Rev* 2012; 33: 981–1030.
- Stepto NK, Moreno-Asso A, McIlvenna LC, et al. Molecular mechanisms of insulin resistance in polycystic ovary syndrome: unraveling the conundrum in skeletal muscle? *f Clin Endocrinol Metab* 2019; 104: 5372–5381.
- 36. Persson S, Elenis E, Turkmen S, et al. Higher risk of type 2 diabetes in women with hyperandrogenic polycystic ovary syndrome. *Fertil Steril* 2021; 116: 862–871.
- 37. Lillioja S, Mott DM, Spraul M, *et al.* Insulin resistance and insulin secretory dysfunction as

precursors of non-insulin-dependent diabetes mellitus. Prospective studies of Pima Indians. *N Engl J Med* 1993; 329: 1988–1992.

- Randeva HS, Tan BK, Weickert MO, et al. Cardiometabolic aspects of the polycystic ovary syndrome. *Endocr Rev* 2012; 33: 812–841.
- Moran LJ, Misso ML, Wild RA, et al. Impaired glucose tolerance, type 2 diabetes and metabolic syndrome in polycystic ovary syndrome: a systematic review and meta-analysis. Hum Reprod Update 2010; 16: 347–363.
- Moran LJ, Strauss BJ and Teede HJ. Diabetes risk score in the diagnostic categories of polycystic ovary syndrome. *Fertil Steril* 2011; 95: 1742–1748.
- Rodgers RJ, Avery JC, Moore VM, et al. Complex diseases and co-morbidities: polycystic ovary syndrome and type 2 diabetes mellitus. Endocr Connect 2019; 8: R71–R75.
- 42. Bozdag G, Mumusoglu S and Zengin D. The prevalence and phenotypic features of polycystic ovary syndrome: a systematic review and metaanalysis. *Hum Reprod* 2016; 31: 2841–2855.
- Kempegowda P, Melson E, Manolopoulos KN, et al. Implicating androgen excess in propagating metabolic disease in polycystic ovary syndrome. *Ther Adv Endocrinol Metab* 2020; 11: 1–24.
- Legro RS, Kunselman AR, Dodson WC, et al. Prevalence and predictors of risk for type 2 diabetes mellitus and impaired glucose tolerance in polycystic ovary syndrome: a prospective, controlled study in 254 affected women. *J Clin Endocrinol Metab* 1999; 84: 165–169.
- O'Reilly MW, Taylor AE, Crabtree NJ, et al. Hyperandrogenemia predicts metabolic phenotype in polycystic ovary syndrome: the utility of serum androstenedione. *J Clin* Endocrinol Metab 2014; 99: 1027–1036.
- Ruth KS, Day FR, Tyrrell J, et al. Using human genetics to understand the disease impacts of testosterone in men and women. Nat Med 2020; 26: 252–258.
- Boudreaux MY, Talbott EO, Kip KE, et al. Risk of T2DM and impaired fasting glucose among PCOS subjects: results of an 8-year follow-up. *Curr Diab Rep* 2006; 6: 77–83.
- Legro RS, Kunselman AR and Dunaif A. Prevalence and predictors of dyslipidemia in women with polycystic ovary syndrome. *Am J Med* 2001; 111: 607–613.
- 49. Cassar S, Misso ML, Hopkins WG, *et al.* Insulin resistance in polycystic ovary syndrome:

a systematic review and meta-analysis of euglycaemic-hyperinsulinaemic clamp studies. *Hum Reprod* 2016; 31: 2619–2631.

- Vigil P, Contreras P, Alvarado JL, *et al.* Evidence of subpopulations with different levels of insulin resistance in women with polycystic ovary syndrome. *Hum Reprod* 2007; 22: 2974–2980.
- Goodarzi MO, Quinones MJ, Azziz R, et al. Polycystic ovary syndrome in Mexican-Americans: prevalence and association with the severity of insulin resistance. *Fertil Steril* 2005; 84: 766–769.
- 52. Moghetti P, Tosi F, Bonin C, *et al.* Divergences in insulin resistance between the different phenotypes of the polycystic ovary syndrome. *J Clin Endocrinol Metab* 2013; 98: 628–637.
- Joham AE, Ranasinha S, Zoungas S, et al. Gestational diabetes and type 2 diabetes in reproductive-aged women with polycystic ovary syndrome. *J Clin Endocrinol Metab* 2014; 99: E447–E452.
- Li W and Li Q. Dysregulation of glucose metabolism even in Chinese PCOS women with normal glucose tolerance. *Endocr J* 2012; 59: 765–770.
- 55. Legro RS, Finegood D and Dunaif A. A fasting glucose to insulin ratio is a useful measure of insulin sensitivity in women with polycystic ovary syndrome. *J Clin Endocrinol Metab* 1998; 83: 2694–2698.
- Dunaif A and Finegood DT. -Cell dysfunction independent of obesity and glucose intolerance in the polycystic ovary syndrome. *J Clin Endocrinol Metab* 1996; 81: 942–947.
- 57. Dunaif A and Graf M. Insulin administration alters gonadal steroid metabolism independent of changes in gonadotropin secretion in insulin resistant women with the polycystic ovary syndrome. *J Clin Invest* 1989; 83: 23–29.
- Dunaif A, Xia J, Book CB, *et al.* Excessive insulin receptor serine phosphorylation in cultured fibroblasts and in skeletal muscle. *J Clin Invest* 1995; 96: 801–810.
- Li M, Youngren JF, Dunaif A, et al. Decreased insulin receptor (IR) autophosphorylation in fibroblasts from patients with PCOS: effects of serine kinase inhibitors and IR activators. J Clin Endocrinol Metab 2002; 87: 4088–4093.
- Corbould A, Kim YB, Youngren JF, et al. Insulin resistance in the skeletal muscle of women with PCOS involves intrinsic and acquired defects in insulin signaling. Am J Physiol Endocrinol Metab 2005; 288: E1047–E1054.

- Dunaif A, Wu X, Lee A, et al. Defects in insulin receptor signaling in vivo in the polycystic ovary syndrome (PCOS). Am J Physiol Endocrinol Metab 2001; 281: E392–E399.
- 62. Bouzakri K, Zachrisson A, Al-Khalili L, *et al.* siRNA-based gene silencing reveals specialized roles of IRS-1/Akt2 and IRS-2/Akt1 in glucose and lipid metabolism in human skeletal muscle. *Cell Metab* 2006; 4: 89–96.
- 63. Huang C, Thirone AC, Huang X, et al. Differential contribution of insulin receptor substrates 1 versus 2 to insulin signaling and glucose uptake in 16 myotubes. *J Biol Chem* 2005; 280: 19426–19435.
- 64. Hojlund K, Glintborg D, Andersen NR, et al. Impaired insulin-stimulated phosphorylation of Akt and AS160 in skeletal muscle of women with polycystic ovary syndrome is reversed by pioglitazone treatment. *Diabetes* 2008; 57: 357–366.
- 65. Corbould A, Zhao H, Mirzoeva S, *et al.* Enhanced mitogenic signaling in skeletal muscle of women with polycystic ovary syndrome. *Diabetes* 2006; 55: 751–759.
- Rajkhowa M, Brett S, Cuthbertson DJ, et al. Insulin resistance in polycystic ovary syndrome is associated with defective regulation of ERK1/2 by insulin in skeletal muscle in vivo. *Biochem J* 2009; 418: 665–671.
- 67. Hansen SL, Svendsen PF and Jeppesen JF. Molecular mechanisms in skeletal muscle underlying insulin resistance in women who are lean with polycystic ovary syndrome. *J Clin Endocrinol Metab* 2019; 104: 1841–1854.
- Lee JO, Lee SK, Jung JH, et al. Metformin induces Rab4 through AMPK and modulates GLUT4 translocation in skeletal muscle cells. J Cell Physiol 2011; 226: 974–981.
- 69. Patten RK, Boyle RA, Moholdt T, *et al.* Exercise interventions in polycystic ovary syndrome: a systematic review and meta-analysis. *Front Physiol* 2020; 11: 606.
- 70. Frøsig C, Rose AJ, Treebak JT, et al. Effects of endurance exercise training on insulin signaling in human skeletal muscle: interactions at the level of phosphatidylinositol 3-kinase, Akt, and AS160. *Diabetes* 2007; 56: 2093–2102.
- 71. Stepto NK, Patten RK, Tassone EC, *et al.* Exercise recommendations for women with polycystic ovary syndrome: is the evidence enough. *Sports Med* 2019; 49: 1143–1157.
- O'Neill HM. AMPK and exercise: glucose uptake and insulin sensitivity. *Diabetes Metab J* 2013; 37: 1–21.

- Kiens B. Skeletal muscle lipid metabolism in exercise and insulin resistance. *Physiol Rev* 2006; 86: 205–243.
- Hortonc TJ, Pagliassotti MJ, Hobbs K, et al. Fuel metabolism in men and women during and after long-duration exercise. *Journal of Applied Physiology* 1998; 85: 1823–1832.
- 75. Friedlander AL, Casazza GA, Horning MA, et al. Training-induced alterations of carbohydrate metabolism in women: women respond differently from men. J Appl Physiol (1985) 1998; 85: 1175–1186.
- 76. Hansen SL, Bojsen-Møller KN, Lundsgaard AM, et al. Mechanisms underlying absent traininginduced improvement in insulin action in lean, hyperandrogenic women with polycystic ovary syndrome. *Diabetes* 2020; 69: 2267–2280.
- 77. Koves TR, Ussher JR, Noland RC, et al. Mitochondrial overload and incomplete fatty acid oxidation contribute to skeletal muscle insulin resistance. *Cell Metab* 2008; 7: 45–56.
- Kubota T, Kubota N, Kumagai H, et al. Impaired insulin signaling in endothelial cells reduces insulin-induced glucose uptake by skeletal muscle. *Cell Metab* 2011; 13: 294–307.
- Galgani JE, Moro C and Ravussin E. Metabolic flexibility and insulin resistance. Am J Physiol Endocrinol Metab 2008; 295: E1009–E1017.
- Thyfault JP, Rector RS and Noland RC. Metabolic inflexibility in skeletal muscle: a prelude to the cardiometabolic syndrome. J *Cardiometab Syndr* 2006; 1: 184–189.
- Broskey NT, Tam CS, Sutton EF, et al. Metabolic inflexibility in women with PCOS is similar to women with type 2 diabetes. Nutr Metab 2018; 15: 75.
- Rimmer M, Tan BK, Teede H, et al. Metabolic inflexibility in women with polycystic ovary syndrome: a systematic review. *Gynecol Endocrinol* 2020; 36: 501–507.
- Adamska A, Karczewska-Kupczewska M, Nikolajuk A, et al. Normal metabolic flexibility despite insulin resistance women with polycystic ovary syndrome. Endocr J 2013; 60: 1107–1113.
- Azziz R. Diagnostic criteria for polycystic ovary syndrome: a reappraisal. *Fertil Steril* 2005; 83: 1343–1346.
- 85. Kumarendran B, O'Reilly MW, Subramanian A, et al. Polycystic ovary syndrome, combined oral contraceptives, and the risk of dysglycemia: a population-based cohort study with a nested pharmacoepidemiological case-control study. Diabetes Care 2021; 44: 2758–2766.

- Schiffer LWA and Storbeck KH. Intracrine androgen biosynthesis, metabolism and action revisited. *Mol Cell Endocrinol* 2018; 465: 4–26.
- Labrie C, Belanger A and Labrie F. Androgenic activity of dehydroepiandrosterone and androstenedione in the rat ventral prostate. *Endocrinology* 1988; 123: 1412–1417.
- Lin H-K, Jez JM, Schlegel BP, et al. Expression and characterization of recombinant type 2 3 alpha hydroxysteroid dehydrogenase (HSD) from human prostate: demonstration of bifunctional 3 alpha/17 beta-HSD activity and cellular distribution. *Mol Endocrinol* 1997; 11: 1971– 1984.
- Luu-The V and Labrie F. Chapter 10: The intracrine sex steroid biosynthesis pathways. In: Martini L (ed.) *Progress in brain research*. Amsterdam: Elsevier, 2010, pp. 177–192.
- Longcope C and Fineberg SE. Production and metabolism of dihydrotestosterone in peripheral tissues. *J Steroid Biochem* 1985; 23: 415–419.
- Larionov AA, Vasyliev DA, Mason JI, et al. Aromatase in skeletal muscle. J Ster Biochem Mol Biol 2003; 84: 485–492.
- Diamond MP, Grainger D, Diamond MC, et al. Effects of methyltestosterone on insulin secretion and sensitivity in women. *J Clin Endocrinol Metab* 1998; 83: 4420–4425.
- Polderman KH, Gooren LJ, Asscheman H, et al. Induction of insulin resistance by androgens and estrogens. *J Clin Endocrinol Metab* 1994; 79: 265–271.
- Zang H, Carlström K, Arner P, et al. Effects of treatment with testosterone alone or in combination with estrogen on insulin sensitivity in postmenopausal women. *Fert Steril* 2006; 86: 136–144.
- 95. Moghetti P, Tosi F, Castello R, *et al.* The insulin resistance in women with hyperandrogenism is partially reversed by antiandrogen treatment: evidence that androgens impair insulin action in women. *J Clin Endocrinol Metab* 1996; 81: 952–960.
- 96. O'Reilly MW, Kempegowda P, Jenkinson C, et al. 11-oxygenated C19 steroids are the predominant androgens in polycystic ovary syndrome. *J Clin Endocrinol Metab* 2017; 102: 840–848.
- Tosi F, Villani M, Garofalo S, *et al.* Clinical value of serum levels of 11-oxygenated metabolites of testosterone in women with polycystic ovary syndrome. *J Clin Endocrinol Metab* 2022; 107: e2047–e2055.

- 98. Schnyder S. Skeletal muscle as endocrine organ and mediators. *Bone* 2015; 80: 115–125.
- Severinsen MCK and Pedersen BK. Muscleorgan crosstalk: the emerging roles of myokines. *Endocr Rev* 2020; 41: 594–609.
- 100. Liu Y, Chewchuk S, Lavigne C, et al. Functional significance of skeletal muscle adiponectin production, changes in animal models of obesity and diabetes, and regulation by rosiglitazone treatment. Am J Physiol Endocrinol Metab 2009; 297: E657–E664.
- Liu Y and Sweeney G. Adiponectin action in skeletal muscle. Best Pract Res Clin Endocrinol Metab 2014; 28: 33–41.
- 102. Perseghin G, Lattuada G, Danna M, et al. Insulin resistance, intramyocellular lipid content, and plasma adiponectin in patients with type 1 diabetes. Am J Physiol Endocrinol Metab 2003; 285: E1174–E1181.
- 103. Iwabu M, Yamauchi T, Okada-Iwabu M, et al. Adiponectin and AdipoR1 regulate PGC-1α and mitochondria by Ca2+ and AMPK/SIRT1. *Nature* 2010; 464: 1313–1319.
- 104. Toulis KA, Goulis DG, Farmakiotis D, et al. Adiponectin levels in women with polycystic ovary syndrome: a systematic review and a metaanalysis. Hum Reprod Update 2009; 15: 297–307.
- 105. Dankel SN, Degerud EM, Borkowski K, et al. Weight cycling promotes fat gain and altered clock gene expression in adipose tissue in C57BL/6J mice. Am J Physiol Endocrinol Metab 2014; 306: E210–E224.
- 106. Xu A, Chan KW, Hoo RL, et al. Testosterone selectively reduces the high molecular weight form of adiponectin by inhibiting its secretion from adipocytes. J Biol Chem 2005; 280: 18073– 18080.
- 107. Chen N, Li Q, Liu J, *et al.* Irisin, an exerciseinduced myokine as a metabolic regulator: an updated narrative review. *Diabetes Metab Res Rev* 2016; 32: 51–59.
- 108. Zhang L, Fang X, Li L, *et al.* The association between circulating irisin levels and different phenotypes of polycystic ovary syndrome. *J Endocrinol Invest* 2018; 41: 1401–1407.
- 109. Li H, Xu X, Wang X, et al. Free androgen index and Irisin in polycystic ovary syndrome. J Endocrinol Invest 2016; 39: 549–556.
- 110. Wang C, Zhang XY, Sun Y, et al. Higher circulating irisin levels in patients with polycystic ovary syndrome: a meta-analysis. *Gynecol* Endocrinol 2018; 34: 290–293.

- Sharma M, McFarlane C, Kambadur R, et al. Myostatin: expanding horizons. *IUBMB Life* 2015; 67: 589–600.
- 112. Xie F, Zhu J, Hou B, *et al.* Inhibition of NF-κB activation improves insulin resistance of L6 cells. *Endocr J* 2017; 64: 685–693.
- 113. Chen MJ, Han DS, Yang JH, *et al.* Myostatin and its association with abdominal obesity, androgen and follistatin levels in women with polycystic ovary syndrome. *Hum Reprod* 2012; 27: 2476–2483.
- 114. Arpaci H. Major determinants of circulating myostatin in polycystic ovary syndrome. *Exp Ther Med* 2019; 17: 1383–1389.
- 115. Hittel DS, Berggren JR, Shearer J, et al. Increased secretion and expression of myostatin in skeletal muscle from extremely obese women. *Diabetes* 2009; 58: 30–38.
- 116. Konopka AR, Asante A, Lanza IR, et al. Defects in mitochondrial efficiency and H2O2 emissions in obese women are restored to a lean phenotype with aerobic exercise training. *Diabetes* 2015; 64: 2104–2115.
- 117. González F, Sia CL, Bearson DM, *et al.* Hyperandrogenism induces a proinflammatory TNF α response to glucose ingestion in a receptor-dependent fashion. *J Clin Endocrinol Metab* 2014; 99: E848–E854.
- 118. González F, Rote NS, Minium J, et al. Reactive oxygen species-induced oxidative stress in the development of insulin resistance and hyperandrogenism in polycystic ovary syndrome. J Clin Endocrinol Metab 2006; 91: 336–340.
- Skov V, Glintborg D, Knudsen S, et al. Reduced expression of nuclear-encoded genes involved in mitochondrial oxidative metabolism in skeletal muscle of insulin-resistant women with polycystic ovary syndrome. *Diabetes* 2007; 56: 2349–2355.
- 120. Skov V, Glintborg D, Knudsen S, et al. Pioglitazone enhances mitochondrial biogenesis and ribosomal protein biosynthesis in skeletal muscle in polycystic ovary syndrome. PLoS ONE 2008; 3: e274.
- Ilie IR. Advances in PCOS pathogenesis and progression-mitochondrial mutations and dysfunction. Adv Clin Chem 2018; 86: 127–155.
- 122. Nilsson E, Benrick A and Kokosar M. Transcriptional and epigenetic changes

influencing skeletal muscle metabolism in women with polycystic ovary syndrome. *J Clin Endocrinol Metab* 2018; 103: 4465–4477.

- 123. Vink JM, Sadrzadeh S, Fau Lambalk CB, et al. Heritability of polycystic ovary syndrome in a Dutch twin-family study. J Clin Endocrinol Metab 2006; 91: 2100–2104.
- 124. Azziz R. PCOS in 2015: new insights into the genetics of polycystic ovary syndrome. *Nat Rev Endocrinol* 2016; 12: 74–75.
- 125. Holian J, Qi W, Kelly DJ, et al. Role of Krüppel-like factor 6 in transforming growth factor-beta1-induced epithelial-mesenchymal transition of proximal tubule cells. Am J Physiol Renal Physiol 2008; 295: F1388–F1396.
- 126. Raja-Khan N, Kunselman AR, Demers LM, et al. A variant in the fibrillin-3 gene is associated with TGF-β and inhibin B levels in women with polycystic ovary syndrome. *Fertil Steril* 2010; 94: 2916–2919.
- 127. Stepto NK, Hiam D, Gibson-Helm M, et al. Exercise and insulin resistance in PCOS: muscle insulin signalling and fibrosis. *Endocr Connect* 2020; 9: 346–359.
- 128. Manti M, Stener-Victorin E and Benrick A. Skeletal muscle immunometabolism in women with polycystic ovary syndrome: a meta-analysis. *Front Physiol* 2020; 11: 573505.
- 129. Lee JO, Kim N, Lee HJ, *et al.* Visfatin, a novel adipokine, stimulates glucose uptake through the Ca2⁺-dependent AMPK–p38 MAPK pathway in C2C12 skeletal muscle cells. *J Mol Endocrinol* 2015; 54: 251–262.
- 130. Barrès R, Yan J, Egan B, *et al.* Acute exercise remodels promoter methylation in human skeletal muscle. *Cell Metab* 2012; 15: 405–411.
- 131. Benrick A, Pillon NJ, Nilsson E, et al. Electroacupuncture mimics exercise-induced changes in skeletal muscle gene expression in women with polycystic ovary syndrome. J Clin Endocrinol Metab 2020; 105: 2027–2041.
- Chen B, Xu P, Wang J, et al. The role of MiRNA in polycystic ovary syndrome (PCOS). *Gene* 2019; 706: 91–96.
- 133. Jiang L, Huang J, Chen Y, et al. Identification of several circulating microRNAs from a genomewide circulating microRNA expression profile as potential biomarkers for impaired glucose metabolism in polycystic ovarian syndrome. Endocrine 2016; 53: 280–290.

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