Effect of Some Clinically Used Proteolytic Enzymes on Inflammation in Rats

A. H. M. VISWANATHA SWAMY* AND P A. PATIL¹

¹Department of Pharmacology and Pharmacotherapeutics, J. N. Medical College, Nehru Nagar, Belgaum - 580 010, India

Swamy, et al.: Effects of Proteolytic Enzymes on Inflammation in Rats

The study was designed to investigate the role of three proteolytic enzymes *viz.*, chymotrypsin, trypsin and serratiopeptidase on hind paw edema and cotton pellet induced granuloma and their possible interactions with aspirin in albino rats. Animals were treated with proteolytic enzymes alone in three different doses or aspirin or in combination with subantiinflammatory dose of aspirin or saline, 30 min before injection of 0.1ml 1% carrageenan. Paw volume was measured before and 3 h after the injection of carrageenan. Chymotrypsin, (5, 18 and 36 mg/kg), trypsin (1.44, 2.88 and 5.76 mg/kg) and serratiopeptidase (0.45, 0.9 and 2.70 mg/kg) were showed dose dependent antiinflammatory activity in acute model of inflammation. Serratiopeptidase showed better antiinflammatory activity on carrageenan induced inflammation than other two proteolytic enzymes and aspirin. However, chymotrypsin, trypsin and serratiopeptidase were found to be more effective than aspirin in subacute model of inflammation. Chymotrypsin, trypsin and serratiopeptidase possess antiinflammatory activity and exhibit synergistic effect with aspirin in both acute and subacute models of inflammation in rats.

Key words: Inflammation, proteolytic enzymes, aspirin

Inflammation is a normal response to protect the tissues from various noxious stimuli and is one of the most normal clinical conditions. A wide variety of enzymes and enzyme mixtures have been used as adjunctive therapeutic agents in a number of clinical conditions particularly in trauma and orthopedic clinics. Proteolytic enzymes are co-administered with non-steroidal antiinflammatory agents. Based on the earlier reports, it is suggested that the presence of proteolytic enzymes like chymotrypsin, cathepsin D¹ and other proteases² in inflammatory exudates indicate their role in the process of inflammation³⁻⁵. On the other hand, proteolytic activities of these enzymes have been proposed to be vital for the control of inflammation by clearing inflammatory debris^{6,7}. There are several reports advocating the use of these proteolytic enzymes for the treatment of inflammatory disorders^{8,9}. While other reports indicate that despite extensive clinical experience their antiinflammatory activity is unproved⁵ and controversial⁸. The literature survey indicates that there is paucity of information regarding serratiopeptidase and interaction with NSAID's. As controversies about

*For correspondence E-mail: vmhiremath2004@yahoo.com Department of Pharmacology, KLES College of Pharmacy, Vidyanagar, Hubli - 580 031, India. the role of proteolytic enzymes like chymotrypsin, trypsin and serratiopeptidase on inflammation still exists, the present study was undertaken to probe antiinflammatory activity of these enzymes and their possible interactions with aspirin.

Wistar rats of either sex weighing between 120 and 150 g were used for the study. Animals were housed in a room temperature maintained at $22 \pm 1^{\circ}$ with an alternating 12 h light-dark cycle. They were subjected to standard diet and water *ad libitum*. Trypsin, purchased from S. D. Fine Chemicals, Mumbai and aspirin was procured form Swastik Pharmaceuticals, Mumbai. Chymotrypsin and serratiopeptidase were purchased from the local market as alfapsin and bidanzen respectively. All the animal experimental protocol has been approved by the institutional animal ethics committee.

The clinical doses of all the three proteolytic enzymes used were converted to rat equivalents with the help of conversion Table¹⁰. All the drugs were dissolved in saline and were administered orally half an hour prior to the carrageenan injection and thereafter repeated once daily for 10 days in subacute model of inflammation.

Acute inflammation was produced by subplantar injection of 0.1 ml of 1% suspension of carrageenan in normal saline in the right hind paw of the rats¹¹. Measurement of paw volume was done using

plethysmograph at '0' and '3' h after injection of carrageenan as per the procedure given in the literature¹². Oral administration of proteolytic enzymes (treatment/dose as shown in table) and aspirin is done 30 min before inducing inflammation and control group animals received normal saline. Mean increase in paw volume was measured and percentage inhibition was calculated by using the formula

% inhibition =
$$100(1-V_r/V_c)$$

where $V_{\rm t}$ is mean paw volume of drug treated, $V_{\rm c}$ mean paw volume of control.

Sub-acute inflammation was produced by cotton pellet induced granuloma in rats^{13,14}. Sterile cotton $(10 \pm 1 \text{ mg})$ soaked in 0.2 ml of distilled water containing penicillin (0.1 mg) and streptomycin (0.13 mg) was implanted subcutaneously bilaterally in axilla under ether anesthesia. The animals were treated with chymotrypsin (18 mg/kg), trypsin (2.88 mg/kg) and Serratiopeptidase (0.9 mg/kg) for consecutive 10 days. Saline treated animals served as control and aspirin (200 mg/kg) was administered as standard drug. The animals were sacrificed on the 11th day and the cotton pellet granuloma was dissected out. After removal of fat and extraneous tissue, the cotton pellets were dried overnight at 55° and their dry weight was recorded. The weight of the cotton pellet before implantation was subtracted from the dried pellet to obtain the net granuloma weight.

Along with subacute antiinflammatory study, antiulcer activity was also performed by removing stomach and adrenal glands from animals treated with proteolytic enzymes, aspirin and control for studying gastric mucosa and weight changes in adrenal glands. The stomach thus removed were cut opened along the greater curvature and washed in saline. The severity of haemorrhagic erosions in the acid secreting glandular mucosa was assessed on a scale. The ulcer area was calculated under a dissecting microscope with a square grid. Gastric mucosal lesions were seen in the form of hemorrhages or linear breaks. Ulcer index was calculated using the following method¹⁵. Ulcer index = 10/X, where X = Total mucosal area/total area of mucosal lesions. The results were analyzed by Student's t test and *P* values <0.05 were considered to be significant.

In acute inflammation model, the carrageenan induced paw edema was significantly reduced by the proteolytic enzymes when compared to control (Table 1). The percentage protection (inhibition) of edema for chymotrypsin (36 mg/kg), trypsin (5.76 mg/kg) and serratiopeptidase (2.70 mg/kg) and aspirin (200 mg/kg) was found to be 62.03, 49.07, 62.81 and 56.09, respectively. The lowest dose of all the three enzymes and aspirin (54 mg/kg) individually has failed to show antiinflammatory activity, but in combination have shown antiinflammatory activity. In the model of subacute inflammation, % weight of the granulation tissue was significantly reduced in animals treated with chymotrypsin (42%), trypsin (36%) and serratiopeptidase (45%) and its combination with subantiinflammatory dose of aspirin showed antiinflammatory effect (Table 2). The chymotrypsin, trypsin and serratiopeptidase treated animals showed significant reduction in ulcer index as compared to control and its combination with aspirin as compared to aspirin treated animals (Table 3).

The proteolytic enzymes treated animals showed significant increase in adrenal gland weights as compared to control, i.e., chymotrypsin showed

TARIE 1.	FEFECT OF DROT		DRUG ON CARR		
IADLE I.	EFFECT OF PhOT	S AND STANDARL		AGEENAN INDUCED	

Treatment	Dose (mg/kg)	Edema at 3 rd h	% Inhibition
Control	Normal saline	1.08 ± 0.03	
Aspirin	200.00	0.18 ± 0.03	56.09*
Chymotrypsin	9.00	$\textbf{0.98} \pm \textbf{0.24}$	9.25
Chymotrypsin	18.00	0.73 ± 0.12	32.40*
Chymotrypsin	36.00	0.41 ± 0.06	62.03*
Trypsin	1.44	0.83 ± 0.11	23.14
Trypsin	2.88	0.70 ± 0.10	35.18*
Trypsin	5.76	0.55 ± 0.05	49.07*
Serratiopeptidase	0.45	1.03 ± 0.04	4.62
Serratiopeptidase	0.90	$\textbf{0.47} \pm \textbf{0.18}$	56.48*
Serratiopeptidase	2.70	0.38 ± 0.05	62.81*
Chymotrypsin + Aspirin	9.00 + 54.00	$\textbf{0.43} \pm \textbf{0.04}$	60.18*
Trypsin + Aspirin	1.44 + 54.00	0.61 ± 0.04	43.51*
Serratiopeptidase + Aspirin	0.45 + 54.00	$\textbf{0.16} \pm \textbf{0.02}$	60.97*

Values are mean \pm SEM; n = 6 animals in each group; *p < 0.05 when compared to control

TABLE 2: EFFECT OF PROTEOLYTIC ENZYMES AND STANDARD DRUG ON COTTON PELLET INDUCED GRANULOMA FORMATION

Treatment	Dose (mg/kg)	Mean granuloma wt (mg/100 g)	% Inhibition
Control	Normal Saline	45.54 ± 1.19	
Aspirin	200.00	27.66 ± 2.60	39.24*
Chymotrypsin	18.00	$\textbf{26.46} \pm \textbf{2.88}$	41.88*
Trypsin	2.88	29.22 ± 2.43	35.85*
Serratiopeptidase	0.90	25.15 ± 1.13	44.77*
Chymotrypsin + Aspirin	9.00 + 54.00	28.88 ± 1.82	36.58*
Trypsin + Aspirin	1.44 + 54.00	$\textbf{30.14} \pm \textbf{0.58}$	33.00*
Serratiopeptidase + Aspirin	0.45 + 54.00	$\textbf{28.10} \pm \textbf{4.23}$	38.28*

Values are mean \pm SEM; n = 6 animals in each group; *p < 0.05 when compared to control

TABLE 3: EFFECT OF PROTEOLYTIC ENZYMES AND STANDARD DRUG ON GASTRIC MUCOSA (ULCER INDEX)

Treatment	Dose (mg/kg)	Mean ulcer index \pm SEM	% Inhibition
Control	Normal Saline	10.00 ± 6.32	
Aspirin	200.00	40.00 ± 0.00	
Chymotrypsin	18.00	$\textbf{5.00} \pm \textbf{0.60}$	50.00*
Trypsin	2.88	$\textbf{5.60} \pm \textbf{0.66}$	44.00*
Serratiopeptidase	0.90	4.21 ± 0.26	57.90*
Chymotrypsin + Aspirin	9.00 + 54.00	$\textbf{26.42} \pm \textbf{5.83}$	33.95#
Trypsin + Aspirin	1.44 + 54.00	30.00 ± 4.21	25.00#
Serratiopeptidase + Aspirin	0.45 + 54.00	$\textbf{23.33} \pm \textbf{7.602}$	41.67#

Values are mean \pm SEM; n = 6 animals in each group; *p < 0.05 when compared to control and #p < 0.05 when compared to aspirin

TABLE 4: EFFECT OF PROTEOLYTIC ENZYMES AND STANDARD DRUG ON WEIGHT OF ADRENAL GLANDS

Treatment	Dose (mg/kg)	Mean weight (mg%BW)	% Increases
Control	Normal saline	9.58 ± 0.767	
Aspirin	200.00	$\textbf{5.38} \pm \textbf{0.23}$	43.84**
Chymotrypsin	18.00	16.45 ± 1.81	71.71*
Trypsin	2.88	13.41 ± 1.24	39.97*
Serratiopeptidase	0.90	18.12 ± 1.93	89.94
Chymotrypsin + Aspirin	9.00 + 54.00	13.25 ± 1.02	38.30*
Trypsin + Aspirin	1.44 + 54.00	15.54 ± 2.01	62.21*
Serratiopeptidase + Aspirin	0.45 + 54.00	17.83 ± 3.07	86.11*

Values are mean \pm SEM; *n* = 6 animals in each group; **p* < 0.05 when compared to control. [†]: % decrease

71.71% increase in adrenal gland weight while trypsin and serratiopeptidase showed 39.97% and 89.94%, respectively. But in aspirin treated group the adrenal gland weight was decreased significantly as compared to control (Table 4).

Findings of the present study clearly indicate that chymotrypsin, trypsin and serratiopeptidase have suppressed inflammation significantly both in carrageenan as well as cotton pellet induced granuloma and appear to be dose dependent. The lowest dose of all the three enzymes and aspirin individually has failed to show any significant antiinflammatory activity on carrageenan induced inflammation, but in combination of all the above have pontentiated antiinflammatory activity of aspirin. The potentiated antiinflammatory activity of aspirin was comparable to that of aspirin 200 mg/kg, not only in carrageenan induced but also in cotton pellet induced granuloma study. The antiinflammatory activity of these enzymes in both models of inflammation may be attributed due to stimulation of neutrophil apoptosis¹⁶, inhibition of neutrophil migration at the inflammatory site¹⁷, inhibition of bradykinin synthesis, decreased vascular permeability and by clearing inflammatory debris¹⁸⁻²⁰.

Aspirin has expected to produce significant increase in the ulcer index as compared to control. While chymotrypsin, trypsin and serratiopeptidase alone and in combination with aspirin reduced ulcer index significantly as compared to aspirin treated animals. The reduction in ulcer index observed in this study may be due to boost of the defensive factors.

Adrenal gland weights of the animals treated with all the three proteolytic enzymes were significantly increased as compared to control but in contrast to aspirin treated animals in which it decreased significantly as compared to control. The reduction in the weight of adrenal glands in aspirin treated group may be because of production of corticosteroids which may in turn explain the antiinflammatory activity of aspirin²¹. Increase in adrenal glands weight in proteolytic enzymes treated groups may indicate the release of catecholamines that are responsible for antiinflammatory activity^{22,23}. Further investigations are needed to establish these facts by biochemical estimations of corticosteroids and catecholamines in plasma.

The observation of the present study clearly indicates that antiinflammatory activity of all the three enzymes were dose dependent. The combination of low doses of these enzymes with sub antiinflammatory dose of aspirin resulted in synergistic antiinflammatory activity without ulcerogenic potential appears to be clinically a beneficial interaction. If the present findings could be extrapolated to human beings, such combination therapies may reduce the adverse effects of NSAID's like aspirin. However clinical trials are further needed to be studied to confirm the same.

ACKNOWLEDGEMENTS

The authors are thankful to Dr. F. V. Manvi, Principal, K. L. E. S. College of Pharmacy, Belgaum, for his constant support during the present research.

REFERENCES

- Whithouse MW. Chemistry and pharmacology. London: Academic Press; 1974. p. 7.
- Roxvall L, Sennerby L, Heideman M. Anti-inflammatory agents inhibit leukocyte accumulation and vascular leakage induced by trypsin and trypsin-digested serum in hamster cheek pouch. J Surg Res 1993;54:207-11.
- Ferreira SH, Rocha M, Silva E. Liberation of a bradykinin-like substance in the circulating blood of dogs by trypsin, chymotrypsin and nagarse. Br J Pharmacol 1969;36:611-22.
- Roxvall L, Sennerby L, Johansson BR, Heideman M. Trypsin-induced vascular permeability and leukocyte accumulation in hamster cheek pouch: The role of complement activation. J Surg Res 1990;49:504-13.
- 5. Ervin G. Advance in Pharmacology, London: Academic Press; 1966 p. 50.
- 6. Weiner M, Piliero SJ. Nonsteroidal anti-inflammatory agents. Ann Rev

Pharmacol 1990;10:171-98.

- Fisher JD. Antiinflammatory Agents: Chemistry and Pharmacology, New York: Academic Press; 1974. p. 369.
- David AC, Richard JR. Advance in Pharmacology and Chemotherapy, New York: Academic Press; 1975. p. 266.
- 9. Ernest BH. AMA Council on Drugs, Chicago: American Medical Association, Publishing Sciences Group; 1965. p. 399.
- Paget GE, Barnes JM. Evaluation of Drug Activities of Pharmacometrics, London: Academic Press; 1964. p. 134.
- Winter CA, Risley EA, Nuss GW. Carrageenan-induced oedema in hind paw of the rats an assay for anti-inflammatory drugs. Proc Soc Exp Biol Med 1962;11:544-51.
- Chattopadhyay RN, Chattopadhyay R, Roy S, Moitra SK. A simple method for plethysmometric measurement of paw volume of small laboratory animals in evaluation of anti-inflammatory effects. Bull Calcutta School Trop Med 1986;34:5-8.
- Winter CA, Porter CC. Effect of alteration in side chain upon antiinflammatory and liver glycogen activities of hydrocortisone ester. J Am Pharm Ass Sci 1957;46:515-9.
- 14. Tuner RA. Screening Methods in Pharmacology, New York: Academic Press; 1965. p. 158.
- Parmar NS, Jagruti KD. A review of the current methodology for the evaluation of gastric and duodenal anti-ulcer agents. Indian J Pharmacol 1993;25:120-35
- Trevani AS, Andonegui G, Giordano M, Nociari M, Fontan P, Dran G, et al. Neutrophil apoptosis induced by proteolytic enzymes. Lab Invest 1996;74:711-21.
- Jutila MA, Kishimoto TK, Finken M. Low-dose chymotrypsin treatment inhibits neutrophil migration into sites of inflammation in vivo: Effects on Mac-1 and MEL-14 adhesion protein expression and function. Cell Immunol 1991;132:201-14.
- Esch PM, Gerngross H, Fabian A. Reduction of postoperative swelling: Objective measurement of swelling of the upper ankle joint in treatment with serrapeptase a prospective study. Fortschr Med 1989;107:67-72.
- Mazzone A, Catalani M, Costanzo M, Drusian A, Mandoli A, Russo S, *et al.* Evaluation of Serratia peptidase in acute or chronic inflammation of otorhinolaryngology pathology: A multicentre, doubleblind, randomized trial versus placebo. J Int Med Res 1990;18:379-88.
- Merten HA, Muller K, Drubel F, Halling F. Volumetric verification of edema protection with Serrapeptase after third molar osteotomy. Dtsch Z Mund Kiefer Gesichtschir 1991;15:302-5.
- Rainford KD. Aspirin and Salicylates, London: Butterworth and Co; 1984. p. 77.
- 22. Green KL. The anti-inflammatory effect of catecholamines in the peritoneal cavity and hind paw of the mouse. Br J Pharmacol 1972;45:322-32.
- 23. Green KL. Role of endogenous catecholamines in the anti-inflammatory activity of α -adrenoceptor blockin g agents Br J Pharmacol 1974;51: 45-53.

Accepted 6 February 2008 Revised 20 August 2007 Received 22. July 2006 Indian J. Pharm. Sci., 2008, 70 (1): 114-117