

J Neurogastroenterol Motil, Vol. 22 No. 2 April, 2016 pISSN: 2093-0879 eISSN: 2093-0887 http://dx.doi.org/10.5056/jnm15138 Journal of Neurogastroenterology and Motility



Association of the Serotonin Receptor 3E Gene as a Functional Variant in the MicroRNA-510 Target Site with Diarrhea Predominant Irritable Bowel Syndrome in Chinese Women

Yu Zhang,^{1,2} Yaoyao Li,^{1,3} Zhenfeng Hao,¹ Xiangming Li,¹ Ping Bo,^{1*} and Weijuan Gong^{2,3,4*}

¹Department of Chinese and Western Integrative Medicine, Medical College of Yangzhou University, Yangzhou, China; ²Jiangsu Key Laboratory of Integrated Traditional Chinese and Western Medicine for Prevention and Treatment of Senile Diseases, Yangzhou, China; ³Jiangsu Coinnovation Center for Prevention and Control of Important Animal Infectious Diseases and Zoonoses, Yangzhou, China; and ⁴Department of Immunology, School of Medicine, Yangzhou University, Yangzhou, China

Background/Aims

The functional variant (rs56109847) in the 3'-untranslated regions (3'-UTR) of the serotonin receptor 3E (*HTR3E*) gene is associated with female diarrhea predominant irritable bowel syndrome (IBS-D) in British populations. However, the relationship of the polymorphism both to *HTR3E* expression in the intestine and to the occurrence of Chinese functional gastrointestinal disorders has yet to be examined.

Methods

Polymerase chain reaction amplification and restriction fragment length polymorphism analyses were employed to detect polymorphisms among Chinese Han women, particularly 107 patients with IBS-D, 99 patients with functional dyspepsia (FD), 115 patients with mixed IBS and 69 patients with IBS-D + FD. We also assessed microRNA-510 (miR-510) and *HTR3E* expression in human colonic mucosal tissues with immunohistochemistry and other methods. Dual-luciferase reporter assays were conducted to examine the binding ability of miR-510 and *HTR3E* 3'-UTR.

Results

Genotyping data showed the variant rs56109847 was significantly associated with IBS-D, but not with FD, mixed-IBS, or FD + IBS-D. *HTR3E* was abundantly expressed around the colonic mucosal glands but less expressed in the stroma. miR-510 expression decreased, whereas *HTR3E* expression increased in the colonic mucosal tissue of patients with IBS-D compared with those in controls. *HTR3E* expression was significantly higher in patients with the GA genotype than that in patients with the GG genotype. The single-nucleotide polymorphisms disrupted the binding site of miR-510 and significantly upregulated luciferase expression in HEK293 and HT-29 cells.

Conclusions

The single-nucleotide polymorphisms rs56109847 led to reduced microRNA binding and overexpression of the target gene in intestinal cells, thereby increasing IBS-D risk in the Chinese Han population. The decreased expression of miR-510 might contribute to IBS-D. (J Neurogastroenterol Motil 2016;22:272-281)

Key Words

Female; Functional gastrointestinal disorders; MicroRNA-510; Receptors, Serotonin, 5-HT₃

Received: August 27, 2015 Revised: December 22, 2015 Accepted: December 31, 2015 This is an Open Access article distributed under the terms of the Creative Commons Attribution Non-Commercial License (http://creativecommons. org/licenses/by-nc/4.0) which permits unrestricted non-commercial use, distribution, and reproduction in any medium, provided the original work is properly cited.

*Correspondence: Ping Bo and Weijuan Gong are equally responsible for this study.

Ping Bo, PhD

Department of Chinese and Western Integrative Medicine, Medical College of Yangzhou University, Yangzhou, China Tel: +86-158-52553020, Fax: +86-514-87978842, E-mail: 13783655@qq.com Weijuan Gong, PhD

- Department of Immunology, School of Medicine, Yangzhou University, Yangzhou, China
- Tel: +86-139-52790811, E-mail address: wjgong@yzu.edu.cn.
- Yu Zhang and Yaoyao Li contributed to this work equally.

Introduction

Functional dyspepsia (FD) and irritable bowel syndrome (IBS) are common functional gastrointestinal disorders (FGID) in the human population, particularly in females.^{1,2} The etiology of FGID is assumed to be associated with infection,³ alterations in immune system or visceral sensitivity,⁴ imbalance of the intestinal flora,⁵ and psychiatric factors.^{6,7} Recent studies on the familial aggregation of patients with FGID have indicated that hereditary factors may contribute to susceptibility to FGID.^{68,9}

MicroRNAs (miRNAs) are a class of small noncoding RNAs that regulate gene expression by binding to the 3'-untranslated region (3'-UTR) of the messenger RNA (mRNA) of the target gene.¹⁰ MicroRNAs are involved in the pathogenesis of FGID.¹¹ Moreover, genomic variation within miRNA target sites may be important genetic sources that influence FGID risk in humans.^{12,13}

Mechanistic studies of FGID showed that serotonin receptor 3 (5-HT₃) plays an important role in the motor-sensory function of the gut.¹⁴ In all the subtypes of the 5-HT₃ receptor, only the 5-HT_{3E} subunit is exclusively expressed in gastrointestinal (GI) tissues, including colon and small intestine.^{15,16} This finding indicates that the 5-HT_{3E} subunit may play a distinct and specific role in the formation and function of 5-HT₃ receptors in the human GI tract. Previous studies in England indicated that the GA genotype of *HTR3E* may serve as a predisposing factor in patients with IBS.¹² The high prevalence of overlap between FD and IBS has been consistently reported both locally and internationally¹⁷; this phenomenon demonstrated that both disorders share common pathophysiological disturbances.¹⁸ Female gender was also associated with FD overlapping IBS.¹⁹

In this study, we examined the association of the 5-HT_{3E} polymorphism with susceptibility to FD, diarrhea predominant IBS (IBS-D), mixed IBS (IBS-M), and FD + IBS-D in a Chinese female Han population. We also assessed the mRNA and protein expression of miR-510 and 5-HT_{3E} in the colonic mucosa of patients with IBS-D and controls to determine the association between the expression and the 5-HT_{3E} polymorphism. We further examined the binding ability of miR-510 to the *HTR3E* 3'-UTR containing G or A alleles.

Materials and Methods

Study Population

In this study, 107 patients with IBS-D, 99 patients with FD, 115 patients with IBS-M, and 69 patients with IBS-D + FD were diagnosed by using the Rome III criteria. All patients were of the southern Chinese female Han population. The control group (n = 294) was age matched, and the subjects were healthy based on checkup examinations and had no history of GI diseases.

All the participants provided informed consent. Briefly, 10 mL of peripheral blood was collected. Each subject was asked to complete a questionnaire with data, including age, profession, and other demographic information. Due to reasons such as the patients' wishes, age, and the high correlation between IBS-D and the single-nucleotide polymorphisms (SNP) rs56109847, we extracted a random 53 patients with IBS-D for biopsy.

Basic characteristics, such as age, profession, and place of residence of the patients and control subjects are presented in Table 1. Most of the patients were younger than 50 years, and were classified as people who lived in Yangzhou city. Over 50% of the patients underwent mental labor in their profession. Age, profession, place of residence, marital status, alcohol or smoking habits, and family history of bowel symptoms were not significantly different between the patients and control subjects (P > 0.05).

Ethics

This study was reviewed and approved by the Medical Ethics Committee of Yangzhou University prior to participant recruitment.

Genotyping

Genomic DNA was extracted from whole blood in buffy coat preparations through the method of Miller with minor modifications by using the TIANamp Blood DNA Midi Kit (TIANGEN, Beijing, China).²⁰ Genotyping was performed using polymerase chain reaction (PCR) amplification and restriction fragment length polymorphism (RFLP) analyses. The reaction volume contained 100 ng of DNA, 10 μ L of 2× Taq PCR Master Mix (Boerdi, Nanjing, China), and 5 pmol oligonucleotide primers flanking the polymorphic region (forward: 5'-CGTCATATGCCTCTGGAA-CA-3' and reverse: 5'-ATAGGCGTGAACCACTGCAC-3') in double-distilled water (ddH₂O) to a final volume of 20 μ L. PCR reactions were performed using a PTC-200 (MJ Research, Minnesota, USA) or a Mastercycler gradient thermal cycler (MJ

Table 1. Basic Characteristics of the Patients and Control Subjects

	IBS-D (n = 107)	FD (n = 99)	IBS-D + FD $(n = 69)$	Controls $(n = 294)$	χ^2	<i>P</i> -value
Age (yr)						
18-30	40 (37.4)	32 (32.3)	25 (36.2)	108 (36.7)	5.932	0.748
31-40	29 (27.1)	25 (25.3)	19 (27.5)	88 (29.9)		
41-50	20 (18.7)	17 (17.2)	10 (14.5)	52 (17.7)		
51-71	18 (16.8)	25 (25.3)	15 (21.7)	46 (15.6)		
Profession						
Mental labor	74 (69.2)	72 (72.7)	53 (76.8)	220 (74.8)	1.721	0.632
Manual labor	33 (30.8)	27 (27.3)	16 (23.2)	74 (25.2)		
Place of residence						
Yangzhou city	104 (97.2)	94 (94.9)	66 (95.7)	289 (98.3)	4.185	0.221
Other cities	3 (2.8)	5 (5.1)	3 (4.3)	5 (1.7)		
Marital status						
Yes	61 (57.0)	66 (66.7)	38 (55.1)	185 (62.9)	3.484	0.323
No	46 (43.0)	33 (33.3)	31 (44.9)	109 (37.1)		
Alcohol or smoking habits						
Yes	10 (9.3)	15 (15.2)	11 (15.9)	28 (9.5)	4.297	0.231
No	97 (90.7)	84 (84.8)	58 (84.1)	266 (90.5)		
Family history of bowel symptoms						
Yes	14 (13.1)	10 (10.1)	10 (14.4)	27 (9.2)	2.416	0.491
No	93 (86.9)	89 (89.9)	59 (85.6)	267 (90.8)		

IBS-D, diarrhea predominant irritable bowel syndrome; FD, functional dyspepsia. Data are presented as n(%).

Research) with the following protocols: incubation at 94°C for 2 minutes; 5 cycles of 94°C for 30 seconds, 68°C for 30 seconds, and 72°C for 30 seconds; 5 cycles of 94°C for 30 seconds, 64°C for 30 seconds, and 72°C for 30 seconds; 25 cycles of 94°C for 30 seconds, 60°C for 30 seconds, and 72°C for 30 seconds; and a final extension step of 72°C for 5 minutes. A 3 μ L aliquot of the PCR product was run on 1.5% agarose gel to visualize a 397-bp fragment. SNP were identified by restriction enzyme digestion. Briefly, 3 μ L of the PCR product of *HTR3E* was digested with 0.5 μ L of the restriction enzyme Hpy188III (New England Biolabs, Ipswich, MA, USA), according to the manufacturer's recommendations. Two types of genotypes were determined through electrophoresis of the enzyme-digested product of *HTR3E* 3'UTR, GG (298 bp), and GA (344 and 298 bp). The enzyme-digested products were confirmed using nucleotide sequencing.

MicroRNA-510 and Serotonin Receptor 3E Expression in Colonic Mucosa

Preparation of tissue sections

Colonic mucosal tissues were obtained from 45 female patients

with IBS-D and the GG genotype and 8 female patients with IBS-D and the GA genotype (21-63 years old). We collected colonic mucosal tissue samples from 53 healthy females (matched to the patient sample with respect to age and ethnicity) as controls.

Immunohistochemistry studies

Immunohistochemical staining was performed as below. Colonic mucosal tissues were immediately washed in cold phosphatebuffered saline (PBS, pH 7.4) and then fixed in 4% buffered paraformaldehyde at 4°C for 24 hours. The tissues were dehydrated, embedded in paraffin, and then sectioned sagittally at a thickness of 4 μ m. Slides were incubated in boiled 0.01 mol/L citrate buffer (pH 7.2) for 10 minutes, cooled at room temperature, immersed in 3% H₂O₂ for 10 minutes, and washed in PBS (0.01 mol/L, pH 7.2) 3 times. Subsequently, 10% normal goat serum was added for 20 minutes. The sections were incubated with primary anti-5-HT_{3E} (1:1000; Sigma, Saint Louis, USA) at 4°C for 14-16 hours. PBS was used as a substitute for antibodies and as a negative control. The slides were then washed in PBS (0.01 mol/L, pH 7. 2) 3 times and then incubated with the anti-rabbit IgG-horseradish peroxidase (HRP) for 20 minutes at room temperature. The slides were stained with 3'3-diaminbezidine tetrahydrochloride. The presence of dark brown granules was considered as a positive result. Negative controls were prepared through incubation with equivalent concentrations of non-immune rabbit antiserum. Integrated optical density measurements of images were performed under the same illuminating setting for each sample. The images were captured using the JEDA801D capture software. Optical density was determined in the microscopic fields of the malformed portion of colonic mucosal tissues. Immunohistochemically stained slides were reviewed independently by 2 researchers.

Quantitative reverse-transcription polymerase chain reaction detection

Total RNA was isolated from colonic mucosal tissues by using Trizol reagent (Invitrogen, Carlsbad, CA, USA) and subjected to reverse transcription with Takara PrimeScriptTM First-strand cDNA Synthesis kit (Takara Bio, Kusatsu, Japan) according to the manufacturer's instructions. Quantitative reverse-transcription polymerase chain reaction (qRT-PCR) was performed using an All-in-One miRNA gRT-PCR Detection Kit (GeneCopoeia, Rockville, MD, USA) on the 7500 Real-Time PCR system (Applied Biosystems, South San Francisco, CA, USA). The U6 small RNA and glyceraldehyde-3-phosphate dehydrogenase (GAPDH) mRNA were used as internal controls. All reactions were performed in triplicate. The primers for miR-510 and U6 were 5'-CTCAACTGGT-GTCGTGGAGTCGGCAATTCAGTTGAGGTGATTG-3' and TACAT ACTGGTGTCGTGGAGTC; and 5' -CGCTTC-GGCAGCACATATAC-3' and 5'-TTCACGAATTTGCGT-GTCAT-3', respectively. The primers for 5-HT_{3E} and GAPDH mRNAs included 5'-GTCCCCACCCAAGTCAACAT-3' and 5'- AAATGTCTGGGAGCCACAGG-3'; and 5'-AAGGT-GAAGGTCGGAGTCAAC-3', and 5'-GGGGTCATTGATG-GCAACAATA-3', respectively.

Western blot analysis

Approximately 100 mg of tissues were minced into small pieces by using surgical blades and then sonicated in protein lysis buffer. Protein concentration was measured by using the Bradford method. The specimens were adjusted to the same protein concentration, aliquoted, and then stored at -80° C until further assay. Equal amounts of total proteins from tissues were separated on SDS-polyacrylamide gels and then electro-transferred onto polyvinylidene fluoride (PVDF) membranes (Millipore, Bedford, MA, USA). The blots were incubated with anti-5-HT_{3E}, an antibody produced in rabbit (1:1000; Sigma), overnight at 4°C. The samples were washed and incubated with goat anti-rabbit IgG-HRP secondary antibodies (1:10 000) for 1 hour at room temperature and detected using an enhanced chemiluminescence kit (BeyoECL Plus; Beyotime, ShangHai, China).

Role of MicroRNA-510 on the 3' Untranslated Regions of the Serotonin Receptor 3E Wild Type and Mutant Messenger RNA

Based on the bioinformatics prediction and data in the literature, the SNP rs56109847 of the *HTR3E 3'*-UTR is located in the region of miRNA-510 recognition for binding. Variations in this region could affect miRNA-510 binding 5-HT_{3E} mRNA and 5-HT_{3E}. To test our hypothesis, we determined whether SNP could affect 5-HT_{3E} expression by using the dual luciferase reporter assay.

Construction of pRL-TK-HTR3E 3'-UTR-c.*76G/A

The following procedures were performed to construct the pRL-TK-HTR3E 3'-UTR-c.*76G/A. The full-length HTR3E 3'-UTR fragment (Supplementary Figure A) was amplified from genomic DNA by using the forward primer 5'-ATTATCTAGAGCAGGTGCTCACCTGCCAACTT-3'(P1) and the reverse primer 5'-CATGTCTGCTCGAAGCGG CCGCCTCTGCAGAATTATTTATTGGG-3'(P2); the former contains a restriction site for Xba I (in bold), whereas the latter contains the reverse complementary sequence of the SV40 polyA 5'-downstream of the pRL-TK Renilla luciferase (Rluc) to vield HTR3E 3'-UTR. Second, the SV40 poly A between Xba I and BamH I of pRL-TK vector (Supplementary Figure B) was amplified using the forward primer 5'-CCCAATAAATAATTCTG CAGAGGCGGCCGCTTCGAGCAGACATG-3'(P3), with 22 bases of the downstream HTR3E 3'-UTR (in bold), and the reverse primer 5'-CGCGGATCCTTATCGATTTTACCACAT-3'(P4), with the restriction site for BamH I in bold, to generate the SV40 poly A fragment. The recombinant DNA fragment (Supplementary Figure C) was generated after PCR by using P1 and P4 primers from the mixture template of HTR3E 3'-UTR and SV40 poly A fragment. The recombinant PCR product was also digested with Xba I and BamH I and inserted with the corresponding restriction sites of the pRL-TK Rluc vector (Promega, Madison, WI, USA) to construct pRL-TK-HTR3E 3'-UTR-c.*76G/A vector.

Cell Culture, Transfection, and Dual Luciferase Reporter Assays

We obtained HEK293 and HT29 cell lines from the Shanghai Cell Bank, Chinese Academy of Sciences and which were then cultured in DMEM (HyClone, Logan, Utah, USA) supplemented with 10% fetal bovine serum (HvCloneUSA). For the luciferase assays, the cells were split into 24-well plates at approximately 5 $\times 10^{5}$ cells per well prior to transfection. Subsequently, 2.5 μ L of Lipofectamine 2000 (Invitrogen) per well was added with 400 ng (per well) of the Rluc recombinant plasmid pRL-TK-HTR3E 3'-UTR-c.*76A/G (constructed above) and 35 ng of the pGL3control (firefly luciferase; Promega). The solution was then cotransfected into cultured cells with 500 ng (per well) of miR-NA-510 plasmid or negative control miRNA plasmid (synthesized by Genechem, Shanghai, China), according to the manufacturer's instructions. After transfection for 48 hours, the dual-luciferase reporter assay system (Promega) was used and cell lysates were prepared according to the manufacturer's instructions. The pRL-TK-HTR3E 3'-UTR-c.*76A/G and pGL3-control were transfected into cultured cells to investigate whether the SNP without miR-510 could affect HTR3E expression. Three procedures were performed for each transfection, and luciferase activity was measured at 3-fold.

Statistical Methods

The Statistical Program for Social Sciences, version 19.0 (SPSS, Chicago, IL, USA) was used for statistical analysis. Pearson's χ^2 test was performed to assess deviation from the Hardy-Weinberg equilibrium. Chi-square test was used to evaluate the basic characteristics of patients and control subjects, as well as differences in the frequency distributions of each allele and genotype of the miRSNPs between cases and controls. Group means were compared using the *t* test. A *P*-value < 0.05 was considered statistically significant, and all the statistical tests were two-sided.

Results

Significant Association of the Single-nucleotide Polymorphisms rs56109847 with Diarrhea Predominant Irritable Bowel Syndrome but Not Functional Dyspepsia or Functional Dyspepsia + Diarrhea Predominant Irritable Bowel Syndrome

For the *HTR3E* c.*76G>A variant, no deviation from Hardy-Weinberg equilibrium was detected in FD, IBS-D, and FD + IBS-D or healthy controls (P > 0.05). Genotyping data showed a significant association between female patients with IBS-D and the SNP rs56109847 in the *HTR3E* 3'-UTR ($\chi^2 = 7.731$, P = 0.005; Table 2). The frequency of the GA genotype relative to the GG genotype for the presence of FD alone, as well as the coexistence of FD and IBS-D, did not significantly differ among the groups.

Positive Expression Rate of Serotonin Receptor 3E in Patients with Diarrhea Predominant Irritable Bowel Syndrome and the GA Genotype Was Significantly Higher Compared with Patients with the GG Genotype, as Determined Using Immunohistochemistry Studies

5-HT_{3E} was abundantly expressed around the colonic mucosal glands with minimal expression in the stroma (Fig. 1A and 1B). 5-HT_{3E} expression was significantly higher in patients with the GA genotype as compared with patients with the GG genotype (P < 0.05; Fig. 1C).

Group -	Genotype				Allele frequency			
	G/G	G/A	χ^2	Р	G	А	χ^2	P
IBS-D $(n_{female} = 107)$	90 (84)	17 (16)	7.731	0.005	197 (92)	17 (8)	7.357	0.007
$FD(n_{female} = 99)$	90 (91)	9 (9)	0.567	0.451	189 (96)	9 (4)	0.546	0.460
$IBS-D + FD (n_{female} = 69)$	61 (88)	8 (12)	1.802	0.179	130 (94)	8 (6)	1.730	0.188
IBS-M ($n_{female} = 115$)	102 (89)	13 (11)	2.258	0.133	217 (94)	13 (6)	2.163	0.141
Controls $(n_{female} = 294)$	274 (93)	20(7)			568 (97)	20 (3)		

Table 2. Genotype and Allele Frequency of the Serotonin Receptor 3E 3'-Untranslated Regions Variant c._76G>A (rs56109847) by Female Diarrhea Predominant Irritable Bowel Syndrome and Healthy Control Groups

IBS-D, diarrhea predominant irritable bowel syndrome; FD, functional dyspepsia; IBS-M, mixed irritable bowel syndrome. Data are presented as n (%).

Serotonin Receptor 3E Protein Expression in Colonic Mucosal Tissues in Patients with Diarrhea Predominant Irritable Bowel Syndrome and the GA Genotype Was Significantly Higher than that in Serotonin Receptor 3E GG Genotype Patients

We detected 5-HT_{3E} protein expression in colonic mucosal tissues in 53 IBS-D patients and 53 normal controls by using Western blot analysis. 5- HT_{3E} protein expression in the colonic mucosal tissues of patients with IBS-D patients was higher compared with that in normal colonic mucosal tissues (P < 0.01; Fig. 2A). These results suggest that 5-HT_{3E} may play an important role in the occurrence of IBS-D, particularly in female IBS-D. We detected 5-HT_{3E} protein expression in colonic mucosal tissues in 45 patients with IBS-D, and the GG genotype and in 8 patients with IBS-D and the GA genotype by using the same method. 5-HT_{3E} protein expression in the colonic mucosal tissues of patients with the GA genotype was significantly higher (P < 0.01; Fig. 2B).

Results of Ouantitative Reverse-transcription Polymerase Chain Reaction Detection

We detected miR-510 expression in the colonic mucosal tissues of 53 patients with IBS-D and 53 normal controls by using qRT-PCR. MicroRNA-510 expression was lower in the colonic mucosal tissues of patients with IBS-D as compared with normal controls (Fig. 3A). However, miR-510 expression in colonic mucosal tissues was not significantly different between patients with IBS-D and the GA genotype and patients with the GG genotype (Fig. 3B). The expression of 5-HT3E mRNA in tissue samples was also not significantly different between patients with IBS-D and normal controls (Fig. 3C) or between patients with IBS-D with the GA and GG genotypes (Fig. 3D) (P > 0.05).







С 0.30 0.25 IOD/area of 5-HT_{3E} 0.20 0.15 0.10 0.05 0.00 GG GΑ

Figure 1. Expression of serotonin receptor $3E (5-HT_{3E})$ in colonic mucosa of female diarrhea predominant irritable bowel syndrome (IBS-D) patients with GA and GG genotypes detected by immunohistochemistry. (A) Expression of $5-HT_{3E}$ in colonic mucosa of female IBS-D patients with GA genotype as detected using immunohistochemistry (magnification, $\times 100$). (B) Expression of 5-HT_{3E} in colonic mucosa of female IBS-D patients with GG genotype as detected using immunohistochemistry (magnification, ×100). (C) Positive expression rate of 5-HT_{3E} in patients with GA genotype was significantly higher than that in patients with GG genotype as revealed by immunohistochemistry studies (*P < 0.05). IOD, integrated optical density.



Figure 2. Expression of the serotonin receptor $3E (5-HT_{3E})$ protein in colonic mucosal tissues of female diarrhea predominant irritable bowel syndrome (IBS-D) patients and normal controls. (A) The expression of 5-HT_{3E} protein in colonic mucosal tissue of female IBS-D patients and normal controls, as detected using Western blot analysis, I (GA) and I (GG) refer to IBS-D patients with the GA genotype and patients with the GG genotype, respectively; C refers to normal control. (B) The 5-HT_{3E} protein expression in colonic mucosal tissues in patients with GA genotype were significantly higher than in patients with GG genotype and normal controls (**P < 0.01).

Serotonin Receptor 3E rs56109847 Disrupts the Binding Site for MicroRNA-510 and Significantly Upregulates Luciferase Expression in HEK293 and HT-29 Cells

The *HTR3E* 3'-UTR variant c.*76G>A affected the binding of miR-510 to the *HTR3E* 3'-UTR, and the c.*76A variant constructs co-transfected with miR-510 showed significantly higher luciferase expression than the c.*76G allele constructs in HEK293 (Fig. 4A) and HT-29 (Fig. 4B) cells (P < 0.001). In both cell lines, luciferase activity was not significantly different when cotransfected with the negative controls (P > 0.05). When transfected with constructs containing the SNP (G/A) without miR-510 or negative control, no difference in luciferase activity was detected in both HEK293 and HT-29 cells (P > 0.05; Fig. 4C and Fig. 4D).

Discussion

In this study the SNP rs56109847 was identified to be linked with the diarrhea phenotype of IBS (IBS-D) in the South Chinese Han female population, and the functional variant of *HTR3E 3'*-UTR could inhibit the binding of miR-510 to *HTR3E 3'*-UTR. The 5-HT_{3E} protein expression level in human colonic mucosal tissues was shown to be enhanced in colonic tissues of IBS-D patients. The 5-HT_{3E} protein expression level was significantly higher in IBS-D patients with the GA genotype than patients with the GG genotype. The polymorphic allele resulted in high expression of 5-HT_{3E} in colonic cell lines. This indicated that microRNAs which target on *HTR3E 3'*-UTR could be potentially used for IBS-D therapy.

High prevalence of overlap between FD and IBS-D has been consistently displayed both locally and internationally.¹⁷ The SNP rs56109847 has been also shown in significant association with female IBS-D in the United Kingdum population.¹² However, we failed to demonstrate an association between *HTR3E* polymorphism and occurrence of FD that overlapped with IBS-D or IBS-M. Furthermore, there was no significant association of the SNP rs56109847 with FD alone or 2 subtypes of FD, which is called epigastric pain syndrome and postprandial distress syndrome. This indicates that pathogenic mechanisms of IBS-D may be different from other functional gastrointestinal disorders.

The 5-HT₃ receptor consists of 5 subunits, including 5-HT_{3A}, 5-HT_{3B}, 5-HT_{3C}, 5-HT_{3D}, and 5-HT_{3E}.^{15,21,22} Only 5-HT_{3E} is exclusively expressed in GI tissues, both colons and small intestines.¹⁵ Although the occurrence of IBS-D is associated with multiple genetic polymorphisms, 5-HT_{3E} may play an important and specific role in the GI tract. SNPs residing in miRNA-binding sites affect expression of target mRNA and contribute to susceptibility of a lot of human diseases, such as cancer, kidney disease, and cognitive disorders.²³⁻²⁶ The enhanced level of gastrointestinal miR-29a expression increased intestinal permeability via down-regulation of target genes in patients with IBS-D.^{27,28} Here the 5-HT_{3E} rs56109847 could directly inhibit the binding of miR-510 to HTR3E 3'-UTR in HEK293 and HT-29 cells confirmed by luciferase assays. This confirmed that the SNP (rs56109847) of the non-coding region of HTR3E affected the binding of mircoRNA, thus affecting the activity of the GI tract.

 $5\text{-HT}_{3\mathrm{E}}$ is abundantly expressed in colonic mucosal glands, but with less expression level in the stroma. The $5\text{-HT}_{3\mathrm{E}}$ protein expression level was significantly higher in IBS-D patients with the GA genotype than in patients with the GG genotype. However,



Figure 3. Relative expression of microRNA-510 (miR-510) in colonic mucosal tissues of diarrhea predominant irritable bowel syndrome (IBS-D) patients and controls. (A) The relative expression of miR-510 in colonic mucosal tissues of IBS-D patients (n = 53) or normal controls (n = 53) was determined using quantitative reverse-transcription polymerase chain reaction (qRT-PCR). (B) The relative expression of miR-510 in 45 IBS-D tissue with the GG genotype and 8 IBS-D tissue samples with the GA genotype. (C) Relative expression of 5-HT_{3E} mRNA in 45 IBS-D tissue with GG genotype and 8 IBS-D tissue samples with the GA genotype. The expression level of miR-510 was normalized against U6 small nuclear RNA. The expression level of 5-HT_{3E} mRNA was normalized against glyceraldehyde-3-phosphate dehydrogenase (GAPDH) mRNA. *******P* < 0.01.

there was no significant difference of miR-510 and 5-HT_{3E} mRNA expression in colonic mucosal tissues in neither the GA nor GG genotype of IBS-D patients. This demonstrates that less binding of miR-510 to the 5-HT_{3E} mRNA contributes to high expression of 5-HT_{3E} protein in the GA genotype. In addition, it is still not clear why miR-510 was down-regulated in colonic mucosal tissues of IBS-D patients compared with controls, which may be associated with local chronic inflammation.

In conclusion, our studies confirm that the SNP rs56109847 is a susceptibility genetic polymorphism for the development of IBS-D in the South Chinese Han female population, and provide insights into the regulation of miR-510 on 5-HT_{3E} expression

of colonic tissues in patients with gastrointestinal disorders. The molecular mechanism that underlies the association of *HTR3E* SNP rs56109847 with IBS-D is also elucidated. Down-regulated expression of miR-510 in colonic mucosa would be a marker of the increased risk for developing IBS-D.

Supplementary Materials

Note: To access the supplementary figure mentioned in this article, visit the online version of *Journal of Neurogastroenterology* and Motility at http://www.jnmjournal.org/, and at http://dx.doi. org/10.5056/jnm15138.



Figure 4. Serotonin receptor 3E (*HTR3E*) variant rs56109847 significantly reduces binding and inhibitory effects of miR-510 in HEK293 and HT29 cells. (A) The *HTR3E* variant rs56109847 significantly reduces the binding and inhibitory effects of microRNA 510 (miR-510) in HEK293 cells (*HTR3E* 3'-UTR-c.*76G versus c.*76A, *HTR3E* 3'-UTR-c.*76G transfected with miR-510 versus with negative control). (B) The *HTR3E* variant rs56109847 significantly reduces the binding and inhibitory effects of miR-510 versus with negative control). (B) The *HTR3E* 3'-UTR-c.*76G transfected with miR-510 in HT29 cells (*HTR3E* 3'-UTR-c.*76G versus c.*76A, *HTR3E* 3'-UTR-c.*76G transfected with miR-510 in HT29 cells (*HTR3E* 3'-UTR-c.*76G versus c.*76A, *HTR3E* 3'-UTR-c.*76G, pRL-TK-*HTR3E* 3'-UTR-c.*76A and blank EK293) (P > 0.05). (D) No significant difference in the relative luciferase activity among pRL-TK-*HTR3E* 3'-UTR-G, pRL-TK-*HTR3E* 3'-UTR-A and blank (HT29) (P > 0.05). Renilla luciferase (pRL-TK) activity was normalized against firefly luciferase (pGL3 control). Values are expressed as the mean \pm SEM for 3 transfections, each measuring 3-fold. ***P < 0.001.

Acknowlegements: Editorial support was provided by School of Medicine, Yangzhou University. The first author thanks all Yangzhou women who participated in this study and the research staff Yaoyao Li, Zhenfeng Hao. The first author thanks Weijuan Gong especially for her support in the modification process.

Financial support: This study was supported by funding obtained from Jiangsu province, P.R. China (Grant No. 14KJD330004).

Conflicts of interest: None.

Author contributions: Ping Bo, Weijuan Gong, and Xiang-

ming Li designed the research study; Yu Zhang, Yaoyao Li, and Zhenfeng Hao performed the experiment; Yu Zhang and Xiangming Li analyzed the data; and Yu Zhang drafted the manuscript.

References

- Futagami S, Yamawaki H, Shimpuku M, et al. Impact of coexisting irritable bowel syndrome and non-erosive reflux disease on postprandial abdominal fullness and sleep disorders in functional dyspepsia. J Nippon Med Sch 2013;80:362-370.
- Longstreth GF, Thompson WG, Chey WD, Houghton LA, Mearin F, Spiller RC. Functional bowel disorders. Gastroenterology 2006;130:1480-1491.

- Tack J, Piessevaux H, Coulie B, Caenepeel P, Janssens J. Role of impaired gastric accommodation to a meal in functional dyspepsia. Gastroenterology 1998;115: 1346-1352.
- Holtmann G, Goebell H, Jockenhoevel F, Talley NJ. Altered vagal and intestinal mechanosensory function in chronic unexplained dyspepsia. Gut 1998;42:501-506.
- Nourrisson C, Scanzi J, Pereira B, et al. Blastocystis is associated with decrease of fecal microbiota protective bacteria: comparative analysis between patients with irritable bowel syndrome and control subjects. PLoS One 2014;9:e111868.
- Zheng Z, Tang H. Decreased neuroplasticity may play a role in irritable bowel syndrome: implication from the comorbidity of depression and irritable bowel syndrome. J Neurogastroenterol Motil 2015; 21:298-299.
- Arisawa T, Tahara T, Shiroeda H, et al. Genetic polymorphisms of SCN10A are associated with functional dyspepsia in Japanese subjects. J Gastroenterol 2013;48:73-80.
- Bengtson MB, Rønning T, Vatn MH, Harris JR. Irritable bowel syndrome in twins: genes and environment. Gut 2006;55:1754-1759.
- Jiang Y, Nie Y, Li Y, Zhang L. Association of cannabinoid type 1 receptor and fatty acid amide hydrolase genetic polymorphisms in Chinese patients with irritable bowel syndrome. J Gastroenterol Hepatol 2014;29:1186-1191.
- Bartel DP. MicroRNAs: target recognition and regulatory functions. Cell 2009;136:215-233.
- 11. Fourie NH, Peace RM, Abey SK, et al. Elevated circulating miR-150 and miR-342-3p in patients with irritable bowel syndrome. Exp Mol Pathol 2014;96:422-425.
- Kapeller J, Houghton LA, Mönnikes H, et al. First evidence for an association of a functional variant in the microRNA-510 target site of the serotonin receptor-type 3E gene with diarrhea predominant irritable bowel syndrome. Hum Mol Genet 2008;17:2967-2977.
- Arisawa T, Tahara T, Fukuyama T, et al. Genetic polymorphism of primicroRNA 325, targeting SLC6A4 3'-UTR, is closely associated with the risk of functional dyspepsia in Japan. J Gastroenterol 2012;47:1091-1098.
- Gershon MD. Nerves, reflexes, and the enteric nervous system: pathogenesis of the irritable bowel syndrome. J Clin Gastroenterol 2005; 39(suppl 3):S184-S193.
- 15. Niesler B, Frank B, Kapeller J, Rappold GA. Cloning, physical mapping and expression analysis of the human 5-HT₃ serotonin receptor-like

genes HTR3C, HTR3D and HTR3E. Gene 2003;310:101-111.

- Karnovsky AM, Gotow LF, McKinley DD, et al. A cluster of novel serotonin receptor 3-like genes on human chromosome 3. Gene 2003;319: 137-148.
- Si JM, Wang LJ, Chen SJ, Sun LM, Dai N. Irritable bowel syndrome consulters in Zhejiang province: the symptoms pattern, predominant bowel habit subgroups and quality of life. World J Gastroenterol 2004; 10:1059-1064.
- Gwee KA, Chua AS. Functional dyspepsia and irritable bowel syndrome, are they different entities and does it matter? World J Gastroenterol 2006; 12:2708-2712.
- Fujiwara Y, Kubo M, Kohata Y, et al. Cigarette smoking and its association with overlapping gastroesophageal reflux disease, functional dyspepsia, or irritable bowel syndrome. Intern Med 2011;50:2443-2447.
- Miller SA, Dykes DD, Polesky HF. A simple salting out procedure for extracting DNA from human nucleated cells. Nucleic Acids Res 1988;16:1215.
- Davies PA, Pistis M, Hanna MC, et al. The 5-HT_{3B} subunit is a major determinant of serotonin-receptor function. Nature 1999;397:359-363.
- Miyake A, Mochizuki S, Takemoto Y, Akuzawa S. Molecular cloning of human 5-hydroxytryptamine3 receptor: heterogeneity in distribution and function among species. Mol Pharmacol 1995;48:407-416.
- Feng N, Xu B, Tao J, et al. A miR-125b binding site polymorphism in bone morphogenetic protein membrane receptor type IB gene and prostate cancer risk in China. Mol Biol Rep 2012;39:369-373.
- Yang P, Tang R, Zhu J, et al. A functional variant at miR-24 binding site in B7-H2 alters susceptibility to gastric cancer in a Chinese Han population. Mol Immunol 2013;56:98-103.
- 25. Papagregoriou G, Erguler K, Dweep H, et al. A miR-1207-5p binding site polymorphism abolishes regulation of HBEGF and is associated with disease severity in CFHR5 nephropathy. PLoS One 2012;7:e31021.
- Yang L, Zhang R, Li M, et al. A functional MiR-124 binding-site polymorphism in IQGAP1 affects human cognitive performance. PLoS One 2014;9:e107065.
- Zhou Q, Souba WW, Croce CM, Verne GN. MicroRNA-29a regulates intestinal membrane permeability in patients with irritable bowel syndrome. Gut 2010;59:775-784.
- Zhou Q, Costinean S, Croce CM et al. MicroRNA 29 targets nuclear factor-KB–repressing factor and Claudin 1 to increase intestinal permeability. Gastroenterology 2015;148:158-169.