

OPEN

Modeling somatic computation with non-neural bioelectric networks

Santosh Manicka & Michael Levin *

The field of basal cognition seeks to understand how adaptive, context-specific behavior occurs in non-neural biological systems. Embryogenesis and regeneration require plasticity in many tissue types to achieve structural and functional goals in diverse circumstances. Thus, advances in both evolutionary cell biology and regenerative medicine require an understanding of how non-neural tissues could process information. Neurons evolved from ancient cell types that used bioelectric signaling to perform computation. However, it has not been shown whether or how non-neural bioelectric cell networks can support computation. We generalize connectionist methods to non-neural tissue architectures, showing that a minimal non-neural Bio-Electric Network (BEN) model that utilizes the general principles of bioelectricity (electrodiffusion and gating) can compute. We characterize BEN behaviors ranging from elementary logic gates to pattern detectors, using both fixed and transient inputs to recapitulate various biological scenarios. We characterize the mechanisms of such networks using dynamical-systems and information-theory tools, demonstrating that logic can manifest in bidirectional, continuous, and relatively slow bioelectrical systems, complementing conventional neural-centric architectures. Our results reveal a variety of non-neural decision-making processes as manifestations of general cellular biophysical mechanisms and suggest novel bioengineering approaches to construct functional tissues for regenerative medicine and synthetic biology as well as new machine learning architectures.

Biological systems have long served as an inspiration and a design challenge for the engineering of artificial intelligence and machine learning^{1,2}, with a special focus on the brain³. However, many biological phenomena, ranging from maze solving by cells and slime molds to complex regulative morphogenesis and regeneration, can be viewed as processes involving information-processing and decision-making⁴, in the absence of a brain⁴⁻¹². Memory, anticipation, and problem solving have been demonstrated in sperm, amoebae, yeast, and plants⁸⁻¹², and these capabilities scaled with the emergence of the Metazoa. For example, when craniofacial anatomical features of tadpoles are scrambled into abnormal configurations, they normalize their aberrant positions over time to regain a correct frog face morphology and then cease remodeling¹³. Thus, cells and tissues functionally ascertain the difference between the current, incorrect craniofacial morphology and the frog's native craniofacial morphology and undertake corrective movements to reduce the error. Similarly, the regeneration of entire limbs in animals such as salamanders can be understood as being driven by a 'test-operate-text-exist' model⁷, in which cells act to implement an invariant anatomical outcome from diverse starting conditions, and stop once this target morphology has been achieved^{7,14}. This theme is reinforced by classical observations such as the fact that tails grafted onto the side of salamanders slowly remodel into limbs¹⁵, demonstrating the ability of tissue to ascertain its position within the whole, compare its organ-level anatomy with that dictated by the target morphology, and remodel toward that correct anatomical setpoint¹⁶.

Importantly, unicellular life forms and somatic cells of multicellular organisms were making flexible decisions based on inputs in their microenvironment long before neurons appeared^{11,17}. Nerves may have speed-optimized ancient bioelectric processes that, since the time of bacterial biofilms¹⁸, were already exploited by evolution to implement memory, long-range coordination, and decision-making utilized for maintenance and construction of anatomical structures¹⁹. In multicellular organisms, these same functions are used to control large-scale patterning²⁰⁻²³. For example, bioelectric signals mediate important aspects of the long-range coordination that keep cells harnessed towards maintenance of a body-plan and away from tumorigenesis⁵. Stable bioelectric circuits also maintain the information needed for fragments of planaria to regenerate the correct number and distribution

Allen Discovery Center, 200 College Ave., Tufts University, Medford, MA, 02155, USA. *email: michael.levin@tufts.edu

of heads and tails^{24,25}, and spatial distributions of resting potentials that are critical for the correct formation of hearts^{26,27}, eyes²⁸, and brains²⁹. Bioelectric signaling is involved at many scales, from decisions of stem cells as to when and how to differentiate³⁰, to the control of size³¹ and regeneration of entire organs^{32,33}. Transient inputs that shift the endogenous bioelectric network states into alternate stable modes can permanently change regenerative and developmental morphology³⁴; because of this, mutations in ion channel genes are an important cause of human birth defects^{35,36}, while modulation of bioelectric circuit state has been used to repair brain structure³⁷. Thus, a number of important fields converge on the need to understand how tissues process information via bioelectric processes: evolutionary developmental biology (plasticity in cell behavior contributes to evolvability of body plans), regenerative medicine (induction of complex repair by modulating the patterns to which cells build), and synthetic bioengineering (the pursuit of novel synthetic ‘living machines’).

Decision-making often exploits some form of logic. The need to understand tissue functions as computation mediated by ancient biophysical mechanisms led us to seek a quantitative model of how non-neural networks could possess logical abilities like those commonly studied in brains. The simplest realizations of logic are known as “logic gates”, which are tiny circuits that perform elementary logic operations like AND, OR, NOT, etc. that comprise any modern digital computer. A unique feature of logic gates is that, although they are typically realized as electronic circuits, the underlying mathematical formulations have been used as coarse-grained models of neurons^{38,39}, genes^{40,41}, and even physiology¹². Logic networks can be assembled to compute any computable function⁴², and thus can be valuable building blocks of intelligent information-processing systems.

Here, we show that non-neural bioelectric networks can compute logic functions, suggesting one way in which evolution can exploit biophysics for decision-making in cellular systems. This provides a new connection between non-neural physiology and a common kind of computational task, thus expanding the known capabilities of developmental bioelectricity. While defining necessary and sufficient conditions for “computation” is a profound question debated elsewhere^{43–46}, logic gates are widely accepted as a non-controversial example of computation. Thus, we extend established connectionist approaches to a more general physiological setting and analyze them to reveal how ionic dynamics in non-neuronal cells could implement both simple and complex logic gates.

A New Bioelectric Network Model

To facilitate the analysis of dynamic, context-dependent biological processes (e.g., morphogenesis and remodeling) as cognitive tasks⁴, where networks of non-neural cells collectively make decisions, we constructed a minimal BioElectric Network (BEN) that comprises the simplest components and processes of bioelectrical signaling. BEN is inspired by a sophisticated and realistic model known as the *Bioelectric Tissue Simulation Engine*⁴⁷ (BETSE), which has been used to show how bioelectric patterns can be created and sustained^{21,48–50} and how such a system might interact with genetic networks to give rise to morphological patterns⁵¹. Our goal was to define a generic, minimal biophysical system using realistic yet simple signaling dynamics, and study its computational properties in the absence of the specialized, highly-derived specifications that define neural networks. Thus, BEN is a simplified version of BETSE that is minimal enough to aid the investigation of the computational capabilities of a bioelectric system that have not yet been understood. The inputs and outputs of this network, representing the logic values, are represented by the bioelectric states of the cells (resting potential or V_{mem}).

BEN is defined by a set of bioelectric components and processes that dictate their production, transport, and decay. BEN consists of two types of regions: an environment and a network of cells. A single cell in a BEN network consists of different types of proteins: ion channels and ion pumps. There are two type of molecules: ions (charged) and signaling molecules (generic uncharged molecules that may represent a ligand, a neurotransmitter, a secondary messenger, etc.). BEN contains three types of ions: sodium (Na^+), potassium (K^+), and chloride (Cl^-). Furthermore, there are two types of ion channels (IC) that each selectively allow the passage of Na^+ and K^+ ions respectively between the cell and the environment; no such channel exists for Cl^- . There is a single potassium-potassium ion pump that actively strives to maintain a non-zero membrane potential (V_{mem}). Two cells in a BEN network may be connected by an electrical synapse known as a “gap junction”⁵² (GJ), represented as an undirected edge in the network, which allows the passage of molecules between the cells. The process that governs the passage of ions through either the GJ or IC is called “electrodifusion”, a combination of electrophoresis and regular diffusion, where the flux is driven by voltage gradients in the former and concentration gradients in the latter. Following the convention adopted in BETSE, we modeled electrodifusion through GJs using the Nernst-Planck equation, and the transmembrane flux of ions through ion channels using the Goldman-Hodgkin-Katz flux equation⁴⁷. However, since BEN is a networked system, where the processes occur at discrete points in space (the cells in Fig. 1a), we used simple discretized versions of the same equations. The flux of the signaling molecules in BEN is governed by a generic nonlinear reaction-diffusion process consisting of two layers of sigmoidal transformations. In the first transformation layer, the flux across each GJ is calculated using a sigmoidal transformation of the concentration difference of the signaling molecule. In the second layer, the transformed flux across each GJ is collectively transformed using a second sigmoid, constituting the net concentration change of the signaling molecule. These two layers minimally represent the multi-layered complexity prevalent in cell signaling networks^{53,54}. For example, calcium (Ca^{2+}) transport within a cell is mediated by clusters of inositol 1,4,5-triphosphate (IP_3) receptors distributed inside the cell in a multi-layered fashion^{55–57}. Moreover, such a two-layer transformation is also found in neuronal cells, where the first stage involves nonlinear transformations of local dendritic potentials, and a second stage involving a linear summation of the outputs from the first stage to determine the overall cellular response^{58–60}. Thus, the two-layer signal transformation architecture of BEN is representative of the multi-layered signaling complexity characteristic of multiple cell types. Moreover, the dynamics of the signal may be viewed as a reaction-diffusion system; see the ‘Methods’ section for more details. Finally, the extracellular environment simulated in BEN includes the same three types of ions as the cells (Na^+ , K^+ , and Cl^-),

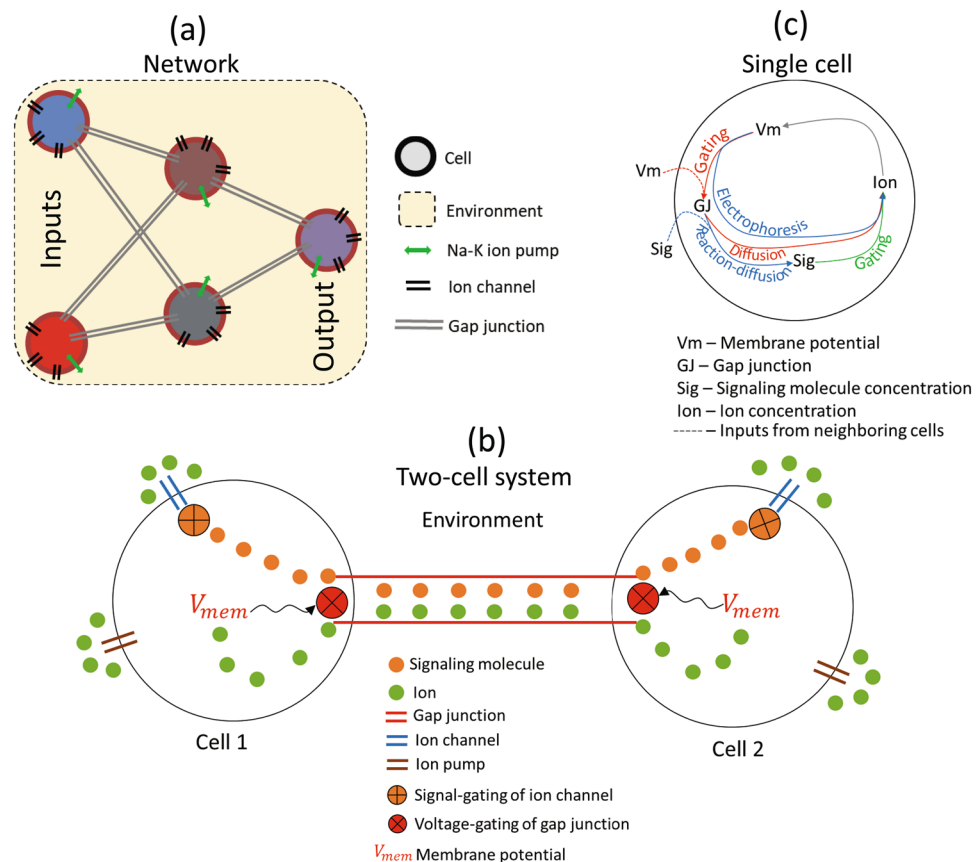


Figure 1. Schematics of the BEN architecture and its constituents. **(a)** The general architecture of a BEN network; **(b)** A BEN network consisting of two cells; **(c)** The network of components (black) related by processes (blue, red and green) within a single cell, some of which receive inputs from adjacent cells. For example, the V_{mem} of a cell together with that of the adjacent cell gates the gap junction which in turn affects the concentrations of the ion and the signaling molecule, and the ion concentration ultimately determines the V_{mem} , closing the loop. This highlights the recurrent nature of the dynamics in BEN.

but initialized with different concentrations. We assume that the environment is effectively infinite in size, hence the ion concentrations there remain constant.

There are two types of gating mechanisms in BEN: chemical-gating of ion channels and voltage-gating of gap junctions. Chemical-gating of IC is a mechanism by which the permeability of an IC is modulated by the binding of a gating molecule (typically a ‘ligand’) to the channel, as a function of the concentration of the molecule^{51,61(Ch.31)}. Rather than the more conventional Hill function to model it⁵¹, we used a simpler sigmoid function that maps the concentration of the signaling molecule to the proportion of the maximum permeability of the IC. In BEN, a higher concentration of the signal tends to depolarize the cell, and lower concentrations hyperpolarize it. Voltage-gating of GJ is a mechanism by which the permeability of a GJ is modulated by the V_{mem} of the connected cells and also the difference between them (known as the ‘transjunctional voltage’)^{51,62,63}. We adopted a sigmoidal transformation that maps the individual V_{mem} of the connected cells that are then averaged over to compute the net GJ permeability, in contrast to the more complex but similar Hill function based model adopted in BETSE⁵¹. In BEN, positive V_{mem} levels tend to make the GJ more permeable, and negative V_{mem} make them less permeable. The mathematical details of BEN are described in the ‘Methods’ section.

There are two types of *learnable* parameters in BEN: weight and bias, following the convention of artificial neural networks⁶⁴ (ANN). The weight is associated with a GJ, and it helps modulate the flux of the signal as well as the voltage-gating of GJ. Negative weights tend to lower the signal concentration in a cell, and positive weights tend to increase it. With regards to voltage-gating, smaller weights diminish the effect of V_{mem} , and larger weights tend to magnify it. The bias is associated with a single cell that helps modulate signal flux: it specifies the threshold at which the signal concentration switches direction of change. By linking these parameters to the core features of BEN, we have assumed that they are not special to ANNs.

BEN networks have a layered architecture (Fig. 1a), a scheme that is widely adopted for ANNs⁶⁴ but which also reflects the tissue layering of many different types of body structures. In this scheme, a network consists of multiple layers of cells, with different choices for the inter-layer and intra-layer connectivity. Some layers have clearly designated roles—for example, the first layer is the ‘input’ and the last layer is the ‘output’. This follows the scheme of various biological systems with specific roles assigned to the layers. For example, the retina is the ‘sensory’ layer that directly receives signals from the environment and sends it to the upstream inter-neuron

layers that integrates the input⁶⁵. The layered architecture is inspired by biological counterparts as diverse as the mammalian visual cortex^{1,2,65} and cellular signaling networks⁶⁶, whose benefits include better organization and efficiency of information processing⁶⁴ and prediction⁶⁷.

A simulation of BEN proceeds as follows. The input layer cells are set to specific V_{mem} levels at the beginning, while all other cells are set at $V_{\text{mem}} = 0$. From that point onward, the V_{mem} levels of the network will dynamically change according to the network parameters, until the output settles at some V_{mem} . The input to the network is either *fixed* (clamped throughout a simulation) or *transient* (set at only the beginning of the simulation and then allowed to change). These two conditions represent the nature of biological information processing where the outcome may depend on the duration of the stimulus. For example, the induction of certain proteins in cellular signaling networks require long-duration input signals^{68,69}; while a brief stimulus (progesterone) is sufficient to induce long-term regenerative response in *Xenopus* limb⁷⁰, a brief exposure to a gap-junction blocker is sufficient to cause stochastic phenotypic shifts in planaria²⁴, a transient invasion into a bacterial ecosystem can dramatically switch its character⁷¹ and transient alterations to V_{mem} cause permanent alterations to gene expression and phenotypic patterning in planaria⁷².

Results

The ability of somatic tissues outside the brain to compute is still controversial, in part because there is not a quantitative model available that demonstrates how basic operations of cognition could be implemented by generic cells. We demonstrate the logic-capabilities of BEN networks by constructing (1) small elementary logic gates; (2) larger “tissue-level” elementary logic gates; (3) compound logic gates composed from elementary gates; and (4) a pattern detector (showing how a complex regenerative response function can be implemented with these components) using standard machine learning methods. Below we show that BEN networks can indeed function as logic gates. We then analyze the behavior of a successfully trained BEN logic gate using the tools of information theory.

BEN networks can implement small elementary logic gates: AND, XOR. A logic gate is a circuit that computes a binary-valued output from binary-valued inputs, according to a set of rules. For example, the AND gate outputs a “HIGH” signal only if both inputs are “HIGH”, otherwise it outputs a “LOW”. The OR gate, on the other hand, outputs a “HIGH” signal if at least one of the inputs has a “HIGH” signal. Thus, a logic gate is defined by a “rule table” that maps an input configuration to a unique output. Many biological systems appear to implement logic functions to facilitate, for example, integration of the various inputs they receive from the environment with the required specificity^{54,73}.

Here we demonstrate that BEN networks can function as logic gates. We used standard machine learning methods to identify the parameters of BEN networks that can perform a desired logical function. This discovery phase in the biological world can be implemented either by evolution (which tweaks the parameters on a phylogenetic time scale) or within the lifetime of a single animal by the dynamic adjustment of ion channel and gap junctional open states as a function of experience (plasticity)⁷⁴.

To provide a proof-of-principle that somatic bioelectric networks can perform logical operations, we sought to identify BEN networks with specific behaviors. Since it is difficult to find the requisite parameters manually, we used machine learning to automatically discover suitable parameter values. We used “backpropagation” (BP) for that purpose. BP is a training method, often used in machine learning, that starts with a random network (defined with a random set of parameters) and gradually tweaks the parameters using gradient descent until the network performs the desired function. Since BEN networks are recurrent by definition (edges are bidirectional), we used “backpropagation through time” (BPTT) to train them; see ‘Methods’ section for more details. We do not claim that BP or BPTT is the method that organisms use to learn in the real world; they are just convenient tools that we used to discover parameters that illustrate the power of BEN networks (see Supplementary 3 for alternative training methods). Below, we show examples of BEN networks implementing the AND gate, and the more difficult XOR gate. First, we show the results of training for a set of 100 training runs for each gate. Then, we describe the behavior of the best gate in each set using time series and dynamical phase space illustrations.

We first sought to determine if bioelectric networks can implement an AND gate, by training a small five-cell BEN network (Fig. 2a) to follow the AND rules that specifies the expected output for a given pair of inputs (Fig. 2b). We trained 100 instances of this network, each starting from different initial conditions of the parameters, discovering that some were indeed able to attain good performance, illustrating that BEN networks can perform the AND function (Fig. 2c). Out of the hundred networks we trained, seven achieved an error of 0.3 mV or less, while a total of 33 networks achieved an error of 1 mV or less. The behavior of the best AND gate we identified is illustrated below by way of time-series plots and phase space diagrams (Figs. 2d, 2e) by considering various input-output conditions.

We next discovered a small five-cell BEN network that can function as an XOR gate by following the same training procedure as for the AND gate above (we report the parameter details in Supplementary 1). The rule table, training performances, behavior of the best XOR gate and the phase space diagram are shown below (Fig. 3). Of the fifty networks we trained, only one network achieved an error of 1 mV. Compared to the AND gate, clearly training the XOR gate is more difficult. This is consistent with the known difficulty of training any dynamical network to behave as an XOR function⁶⁴, since it is a nonlinear function compared to AND, which is linear.

BEN networks can implement larger “tissue-level” elementary logic gates. We next asked whether larger networks, which contain more cells than strictly necessary, could still perform the logic functions. These larger networks are like tissues and organs, which can contain a range of diverse cells that communicate. Thus, we next describe a “tissue-level” AND gate. First, we show the results of training for a set of 100 training

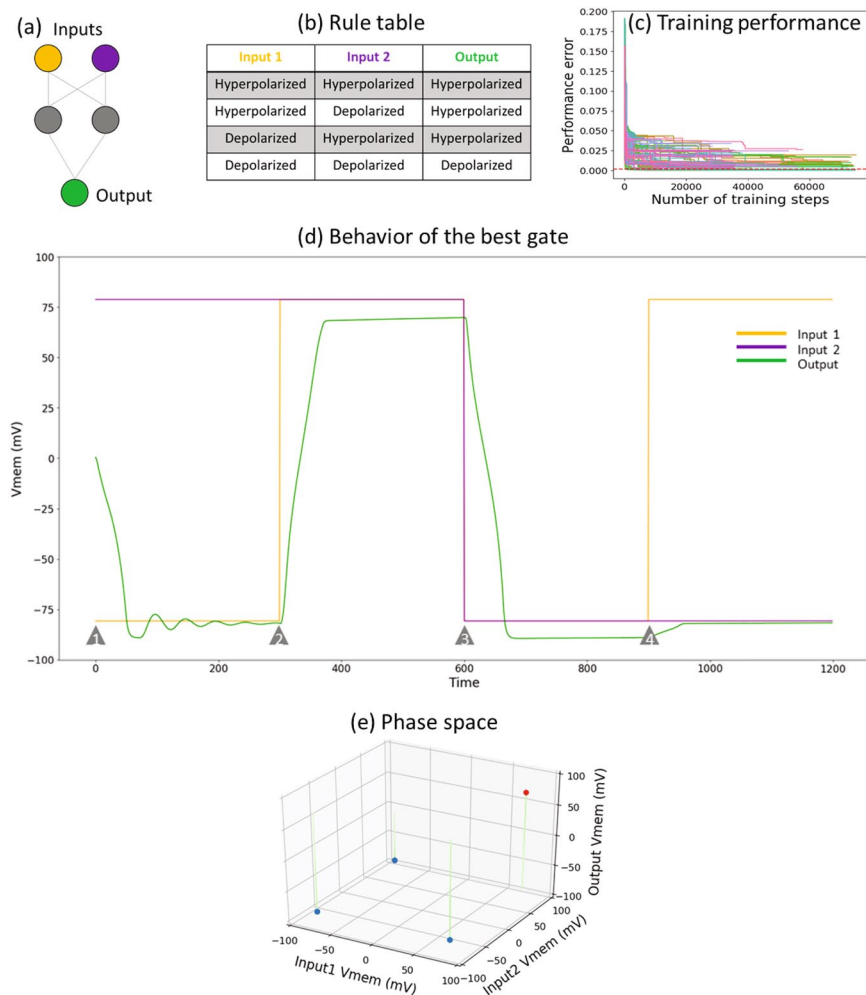


Figure 2. The AND gate. **(a)** A schematic of the AND logic gate and its rule table. The orange and purple cells are the inputs, and the green cell is the output; the grey cells are intermediate cells offering computational support. **(b)** Specification of the output V_{mem} for various combinations of input V_{mem} levels. This is known as the “truth table” in the Boolean logic literature, where the “hyperpolarized” state (around -80 mV) is indicated as “0”, “False” or “OFF” and the “depolarized” state (around $+80$ mV) as “1”, “True” or “ON”. This rule table essentially summarizes the mechanism of the BEN-based AND gate: the output is depolarized only when both of its inputs are depolarized; in Boolean logic terms the output is ON only when both inputs are ON. **(c)** Pareto front of training errors over time (one unit is equal to a single training epoch) for the AND gate. This plot depicts the “front” with the best errors achieved over time. This figure demonstrates that the training does indeed result in learning. **(d)** The behavior of the best AND gate shown in the form of time series of the input and output nodes of the gate, shown for all four input-output conditions generated in a random sequence. The red and blue lines represent the states of the two input nodes, and green represents the output. The grey triangles mark the time points at which the inputs are switched to a different state. For example, both inputs are hyperpolarized at time point (1), while at time point (2) the red input is depolarized. **(e)** The dynamical phase of the logic gate: a depiction of a set of trajectories in the input-output space, illustrated in a time-lapse style. This dynamical system has two attractors in the output space, highlighted in filled red (depolarized state) and blue (hyperpolarized state) circles. The trajectories look straight because the inputs are fixed, and only the output changes.

runs for each gate. Then, we describe the behavior of the best gate in each set, using time series and dynamical phase space illustrations.

We used a combination of genetic algorithm (GA) and BP for identifying this kind of circuit; the GA discovers the network structure (example structure in Fig. 4 inset) and BP finds the parameters of the network we sought. The best network was obtained at the end of 233 generations, which achieved an average error of about 0.6 mV. More details of the training are described in the ‘Methods’ section.

We conclude from the above results that it is indeed possible for large BEN circuits to compute simple logic functions, suggesting that this type of signaling can be carried out by developmental compartments with varying cell numbers.

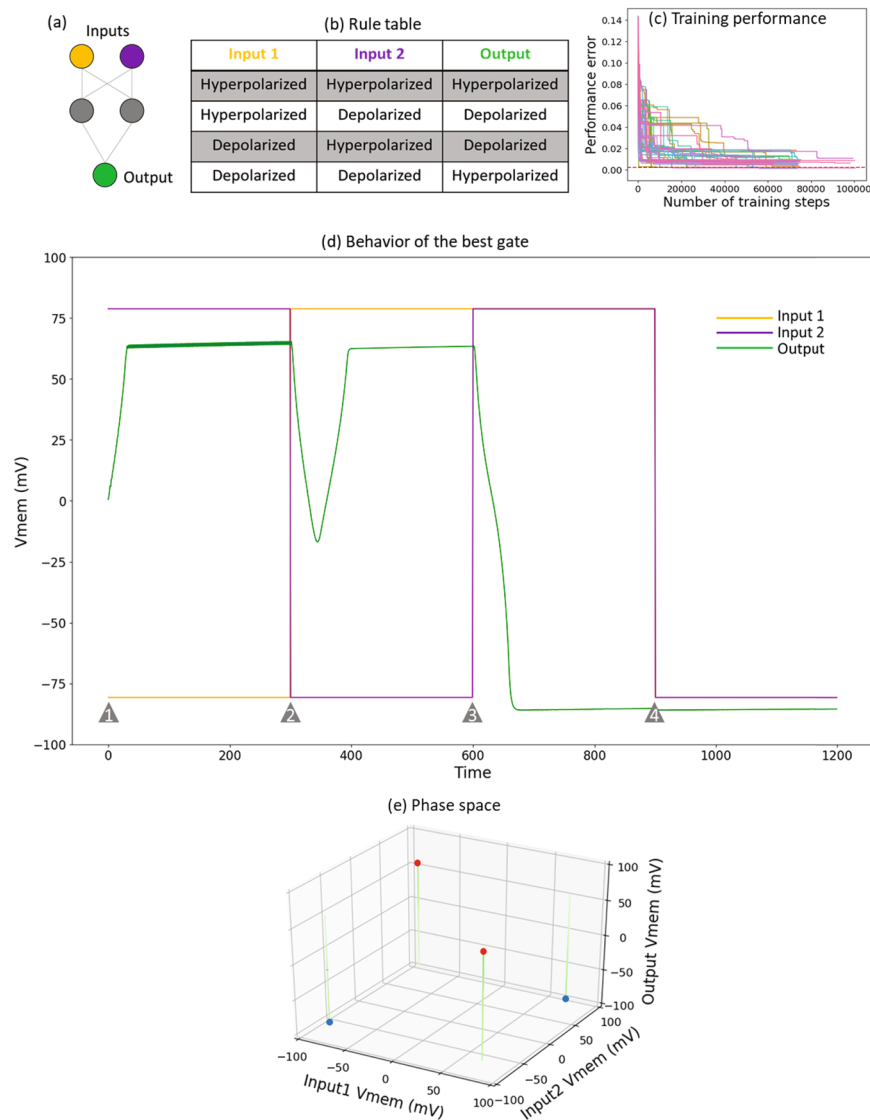


Figure 3. The XOR gate. **(a)** Schematic of the XOR logic gate; the orange and purple cells are the inputs, and the green cell is the output; the grey cells are intermediate cells offering computational support. **(b)** The XOR rule table: output is depolarized only when one of its inputs, but not both, is depolarized. Table specifies the output V_{mem} for various combinations of input V_{mem} levels; this is known as the “truth table” in the Boolean logic literature, where the “hyperpolarized” state (around -80 mV) is indicated as “0”, “False” or “OFF” and the “depolarized” state (around $+80$ mV) as “1”, “True” or “ON”. **(c)** Pareto front of training errors over time (one unit is equal to a single training epoch) for the XOR gate. This plot depicts the “front” with the best errors achieved over time. This figure demonstrates that the training does indeed result in learning, but fewer networks are successful compared to AND (Fig. 2). **(d)** Behavior of the best XOR gate. Shown here are the time series of the input and output nodes of the gate, shown for all four input-output conditions generated in a random sequence. The red and blue lines represent the states of the two input nodes, and green represents the output. The grey triangles mark the time points at which the inputs are switched to a different state. **(e)** The dynamical phase space of the gate: a depiction of a set of trajectories in the input-output space, illustrated in a time-lapse style. This dynamical system has two attractors in the output space, highlighted in filled red (depolarized state) and blue (hyperpolarized state) circles. The trajectories look straight because the inputs are fixed, and only the output changes.

BEN can implement a complex “tissue-level” logic gate: a pattern detector. One of the most impressive computational tasks that tissues undertake is the set of decisions that enable large-scale regeneration of organs and appendages in some species^{19,75}. Specifically, to repair after (unanticipated) injury and stop growth and remodeling when the anatomy has been corrected, cells need to make decisions about the physiological and geometric state of other tissues. Cells need to ascertain whether a large-scale morphology is correct or not, in order to regulate regenerative pathways or cease further change (errors in achieving morphostasis can manifest as cancer). Much work has gone into characterizing the physiological events that signal the binary event of “injury”, but it is now clear that even in the absence of trauma, some complex organ systems such as the craniofacial

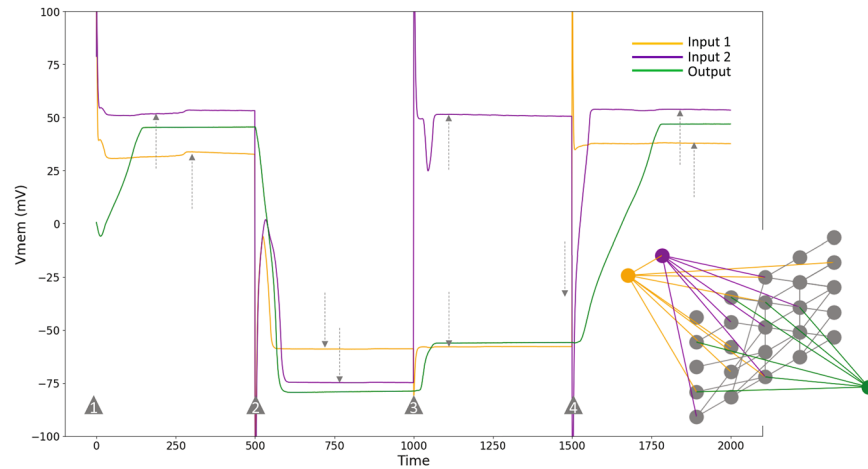


Figure 4. The structure and behavior of the best evolved tissue-level AND gate. Following the convention of the color-coding, orange and purple represent input nodes, while green represents the output node. Behavior correctly follows the AND rule (Fig. 2b). Inputs are transient: they are set (externally) to their respective states at the time steps marked by the grey triangles and then removed. As can be seen, they slightly oscillate for a small period of time after they are set initially, before dynamically fixing themselves (marked by grey dashed arrows) during every simulation. Thus, the network has evolved the capacity for memory: it remembers the inputs (at least their qualitative levels) even after they are removed. Inset shows the structure of the network.

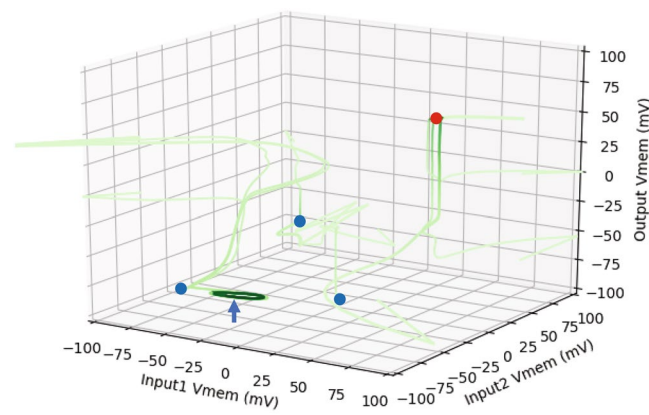


Figure 5. The dynamical phase space of the best tissue-level AND gate. A depiction of a set of trajectories in the input-output space, illustrated in a time-lapse style. This dynamical system has two attractors in the output space, highlighted in filled blue (hyperpolarized state) and red (depolarized state) circles. As can be seen, there are three standard hyperpolarized attractors (fixed-point-like), and a fourth cyclic hyperpolarized attractor (marked with a blue arrow). Even though the cyclic attractor is in the appropriate region of the phase space, it was not actually required for during the training—it accidentally emerged. Furthermore, as opposed to the straight trajectories of the previous phase space diagrams, the trajectories maneuver freely here, and occasionally spill over the usual V_{mem} range which is about $[-100, 100]$ mV. This is because the inputs are transient and not fixed, hence they change states as well over time.

structures^{13,76} can begin drastic remodeling when the configuration is incorrect. Despite the advances in molecular biology that have identified genes *necessary* for this to occur, it is entirely unclear how cells recognize correct vs. incorrect patterns on a spatial scale much larger than themselves. Thus, we next sought to demonstrate that realistic biophysical mechanisms can implement computations which are *sufficient* to enable this crucial capability of living systems.

A pattern detector can be thought of as a complex (more than two inputs, thus non-elementary) AND gate, since its output is ON only when each of the inputs is at a specific desired state—in other words, only when *all* input conditions are satisfied. This is a more realistic setting that the tissue-level AND described in the previous section, since it has many more than two inputs (a tissue typically consists of many more than two cells). We identified, as proof-of-principle, a relatively large (44 cells) non-elementary logic gate that recognizes French-flag-like patterns; we used the French-flag as the pattern of interest due to its prominence in developmental biology as a simple morphogen gradient⁷⁷. We define a French-flag pattern as a particular configuration of V_{mem} levels in the input layer (3×6): the leftmost two columns (band) are blue (hyperpolarized V_{mem}), the middle band is grey (intermediate V_{mem}), and the rightmost band is red (depolarized V_{mem}). This network recognizes the French-flag

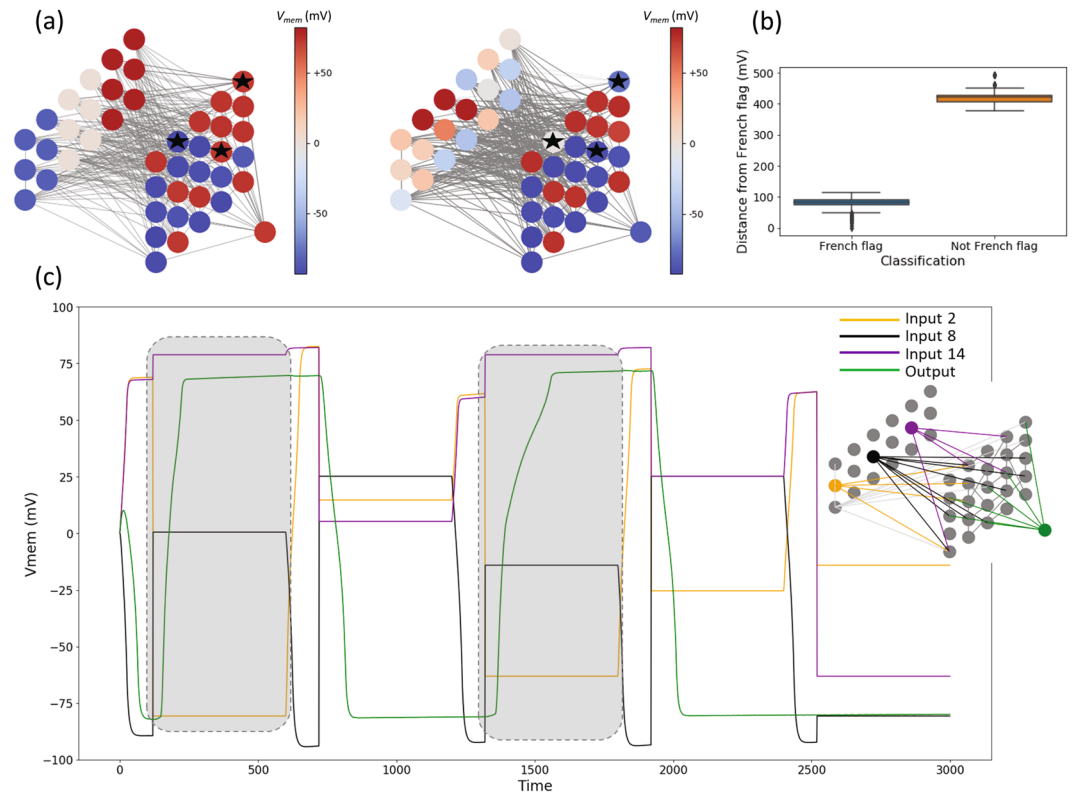


Figure 6. The French-flag detector. **(a)** Snapshots of the equilibrium (final) network states (V_{mem} levels) in two detection scenarios. Left: input pattern is the French flag, and the output is depolarized (representing “correct” pattern). Right: input pattern is a highly distorted version of the French flag, and the output is hyperpolarized (representing “incorrect” pattern). Colors represent polarity levels of the cells in units of mV. Notice that the only three intermediate cells that differ in their states between the two cases are those marked with a black star, suggesting that the detector makes minimal use of information to distinguish between patterns. Also notice that the edges are thicker in the right than in the left, suggesting that the voltage-gating dynamics of the edges (representing gap junctions) play a crucial role. **(b)** The overall performance of the best evolved pattern detector, with data collected from a set of 1000 simulations: 100 parallel sets each with a random sequence of 10 simulations. As expected, the pattern detector classifies patterns similar to the French flag (a total of 500 sample inputs) as “French flag”, and those that are dissimilar (500 inputs) as “not French flag”. This suggests that the pattern detector is robust to noise to a sensible extent. The width of the classification boundary was set implicitly due to the way sample input patterns from the two classes were generated (details in the ‘Methods’ section). **(c)** The behavior of the best French-flag detector shown for a set of four representative cells (three inputs out of a total of eighteen and one output) for a random sequence of five patterns. Inset highlights the four nodes whose colors correspond with those in the time series. Highlighted in grey squares are the cases where a French-flag-like pattern is input for which the output is depolarized, as expected; for all other cases where a random pattern is shown, the output is hyperpolarized.

pattern and slightly noisy versions of it by expressing a depolarized V_{mem} in the output cell, while expressing a hyperpolarized V_{mem} in the output as a response to all other patterns.

The pattern detector is set up similar to the tissue-level AND gate. The main differences between the two are: (1) inputs are fixed in the pattern detector, as was the case for the small logic gates described in Section 4.1; (2) inputs are set after an initial time period during which the network is allowed to settle at its intrinsic “baseline” state (this captures the essence of biological systems that have intrinsic activity even in the absence of external stimuli); and (3) input patterns are generated randomly where two sets are generated, one consisting of the French-flag and noisy variants of it, and the other consisting fully randomized versions of the French-flag (details in the ‘Methods’ section). We used the combined GA-BP search-train method, as before, to find this network; see more details in the ‘Methods’ section. The best evolved pattern detector is the individual with the highest fitness score in the last generation of the GA, which was discovered relatively quickly at the end of 11 generations (three other networks achieved similar errors). Even though the pattern detector is larger than the tissue-level AND described above, it was easier to train than the latter. This is due to the advantage of the inputs being fixed in the case of the pattern detector but transient in the case of the tissue-level AND gate: the fixed inputs provide a constant supply of information that the network does not have to remember unlike the tissue-level AND gate.

The above search successfully discovered a successful French-flag detector. The behavior of this detector when it sees a French-flag pattern and a random pattern in its input layer is depicted in Fig. 6a: in the first case the

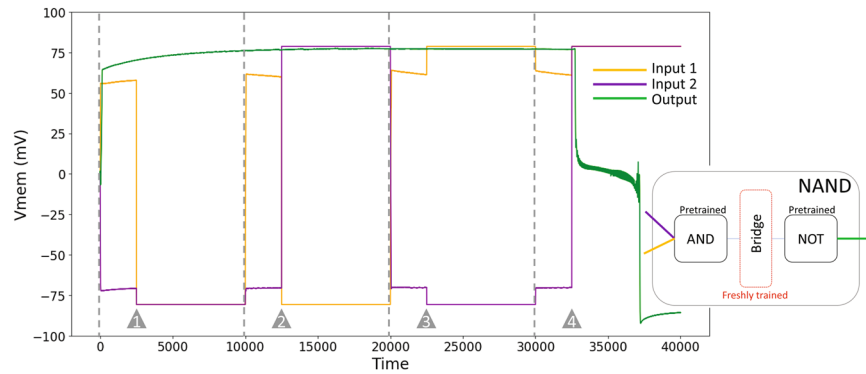


Figure 7. The behavior of a successfully trained NAND gate for a random sequence of four inputs. Orange and purple lines represent the activities of the inputs, while green represents the output. The vertical dashed lines mark the beginning of each trial in the sequence, however the inputs are applied, after an initial transient, at the time points marked by the grey triangles. Inset is a schematic of the architecture of the gate: pretrained AND and NOT gates are connected by a “bridge”, a single layer of cells, that is alone trained.

output is depolarized (red), while in the second case it is hyperpolarized (blue), both as expected. Furthermore, this detector recognizes slightly distorted variants of the French-flag where the distance of a pattern from the French-flag is the Euclidean distance (Fig. 6b). We observe that input patterns up to a distance of about 150 mV from the French-flag are recognized as French-flag, while those at a distance of about 350 mV and above are classed as not French-flag. Finally, the detector also responds correctly to a *sequence* of randomly chosen input patterns (Fig. 6c), showing that the behavior of the network only depends on the state of the inputs and not on the rest of the network.

We conclude that BEN has in principle the ability to distinguish between patterns. The biological implication is that somatic tissues can in principle have the same ability, and thus be a part of much larger pattern regulation mechanisms.

BEN can implement compound logic gates. Biological processes are complex by nature. The rampant complexity is partly managed by nature by way of modularity⁷⁸, in gene regulation⁷⁹ and the brain^{80,81}, for example, at multiple scales. Modularity is beneficial because the modules can be independently tinkered with for the purpose of large-scale outcomes⁷⁹. Thus, it is important to understand whether bioelectric circuits can implement logic functions that are too complex to be described by a single gate but are actually combinations of elementary logic primitives.

Compound logic gates are compositions of modules of logic gates, and their biological equivalents underlie several processes including sporulation in *B. subtilis* and the neuronal dynamics of *C. elegans*^{82,83}. They can also have pharmaceutical applications—for example, the control of bacterial invasion of tumor cells and diagnosis of cellular environments and automatic release of drugs⁸².

One way to design compound networks, in general, is to compose them from appropriate pre-designed modules. For example, a “NAND” gate can be constructed by combining the AND and NOT gates. Compound genetic circuits have been constructed in this fashion^{84,85}, as well as designed *de novo* from scratch⁸². Here, we show how to construct compound logic gates in physiological networks by composing pre-trained BEN logic gate modules. Unlike genetic networks and neural networks, which are directed, BEN networks are bidirectional, due to the symmetric nature of gap junctions. Thus, unlike genetic networks, compound logic gates in BEN cannot be constructed simply by connecting the modules. Put more precisely, if the modules were simply connected together, then the output of one module will also receive signals from the input of the connected module. This could potentially result in the upstream module outputting the wrong state, and thus the downstream module receiving and outputting the wrong states as well. To mitigate this, we included a “bridge” layer that interfaces the modules and trained it so that the compound gate as a whole behaves as desired: the downstream module outputs the correct state for every set of inputs received by the upstream module. For more details on the architecture and training of the bridge, see Supplementary 4. Figure 7 shows an example of a successfully trained NAND gate by composing pre-trained AND and NOT gates.

We conclude that it is possible to construct compound logic gates by composing pre-trained modules and training only an interface bridge connecting the modules. This method can in principle be extended to more complex circuits.

Discussion

Various forms of natural and artificial computing systems with unconventional mechanisms have been proposed⁸⁶. Even the modern von Neumann computer architecture was inspired by the mechanisms of DNA transcription and translation⁸⁷. Other examples include “liquid computers” that involve interactions of fluid streams and droplet flows to perform logic operations⁸⁸; “chemical computers” that utilize the dynamics and equilibrium-behavior of chemical reactions^{89–93} for logic operations and learning^{94,95}; DNA-based computers that utilize various forms of catalytic reactions such as ‘seesawing’ and ‘strand-displacement’ for pattern recognition⁹⁶, protein-DNA interactions to implement logic functions⁹⁷ and even for general-purpose computing⁹⁸;

gene regulatory networks that take advantage of the multistability of gene activity for learning⁹⁹, memory^{100–102} and logic^{84,85,101,102}; “bacterial computers” that use enzyme-based computing to implement logic gates¹⁰³; electric current flow making use of Kirchhoff’s law for solving mazes¹⁰⁴; and even “sandstone computers” that utilize natural erosion processes to implement logic gates¹⁰⁵. BEN is an unconventional computing system, since it is a non-neural system like the examples above.

Our work does not require a commitment to the view that every biological question must be dealt with via a computational perspective, despite the fact that there is a robust body of new experimental work driven by the hypothesis that some tissues are indeed implementing processes we would recognize as memory, comparison, or measurement^{53,106–108}. The key question we answer here is: are pre-neural physiological networks able, in principle, to perform simple computations such as logical operations? Prior to these results, it was not known, and many developmental biologists’ intuitions are that such networks cannot perform such functions - they are almost never considered for model-building. Our data provide the first proof-of-existence showing that the class of biological systems where a computational approach *could* be useful, such as evolved or synthetic regulative morphogenesis, needs to be expanded to include somatic bioelectric networks.

BEN is also a type of patterning system. Patterning is the ability of a system to generate, sustain, and recognize patterns in the states, whether physical, chemical, or biological, of the system. It is a widespread phenomenon reported in biological systems, including pattern recognition of pathogen molecules by cellular receptors¹⁰⁹, axial patterning in planaria¹¹⁰, spatial patterning during neural patterning¹¹¹, cell polarity¹¹², genetic patterning in bacterial populations¹¹³ and voltage patterning in neural tubes⁴⁹. Here we focus on bioelectric non-neural networks of patterning. Various non-neural network models of patterning have been proposed. Some examples include the “packet-routing” system that uses an internet-like intercellular messaging system¹¹⁴, auto-associative dynamical systems¹¹⁵, and cellular automata¹¹⁶ based systems that combine local and global communication to model pattern regeneration. Our laboratory has produced sophisticated non-neural bioelectric network models of tissue patterning, like the *Bioelectric Tissue Simulation Engine* (BETSE) that incorporates detailed biophysical mechanisms of ion channels and transport processes^{47,51,117}, as well as relatively simpler models that incorporate only the high-level functionality of the bioelectric components^{48,118,119}. These models have been used to show that networks of ordinary tissues can give rise to gradient-patterns^{47,51}, Turing patterns^{21,119}, as well as oscillatory patterns^{118,120} that are thought to be essential for long-distance communication. BEN differs from these models on two fronts: (1) it is biophysically simpler than BETSE, serving as a minimal model of dynamics sufficient for computation; and (2) it is more realistic than the model of Reference¹¹⁸, which uses “equivalent circuits” where the equilibrium V_{mem} is explicitly specified in the equations, whereas in BEN the equilibrium V_{mem} emerges from the ion channel parameters.

We have shown here that BEN networks can implement elementary logic gates, more complex tissue-level logic gates, pattern detectors and compound logic gates. Furthermore, we have demonstrated that logic can be implemented in circuits with bidirectional connections that is typical of non-neural tissues, in contrast to the conventional directed circuits like neural networks and digital electronic circuits. This implies that even though non-neural tissues may find it harder to implement formal logic (due to lower control over information flow; see Supplementary Fig. S2b, for example, where information flow is recurrent even in a small network, making it hard to control), they can achieve it. Not only can BEN networks compute, but they can also be robust to damage. One of the most important, and heretofore unexplained, properties of biological control circuits is that they continue to function computationally under significant perturbation (as seen by work on metamorphosis, stability of memory and behavioral programs during drastic tissue remodeling, and invariance of morphogenesis to changes in cell number^{76,121,122}). BEN networks can exhibit a similar behavior, retaining function even after the removal of cells. We found that BEN achieves this by distributing information in the network (Supplementary 2).

The particular methods by which BEN networks were trained is not of central importance in this work and do not impact the main result – that BENs can be readily found which implement logic functions. We used BPTT simply as a way to discover the parameters of (train) BEN networks that perform specific logic functions or recognize patterns. In the biological world, training (learning) may happen in other ways that may be broadly classified as offline and online. Offline learning happens outside of the lifetime of an individual and at a population level, while online learning happens during the lifetime of an individual. Evolution is an example of the former, while Hebbian learning is an example of the latter. BPTT may be classified as a form of online learning, but it has been claimed to be not biologically realistic^{123,124}. More biologically realistic forms of online learning have been proposed, for example, forward propagation of eligibility traces¹²³, extended Hebbian learning¹²⁵, dynamic self-organizing maps¹²⁶ and instantaneously trained neural networks¹²⁷. Such methods could potentially advance biologically realistic training of BEN networks, which we will develop in future research, but they are not necessary to find examples demonstrating that BEN networks can compute.

BEN models the bioelectric *principles* of a generic bioelectric network, and not *all* the physiological details, as these can vary widely across tissues. Individual cell types can be modeled in future effort simply by altering the ion channel parameters while retaining the basic form of the equations. Although BEN is a model of a non-neural bioelectric network, it has certain features that resemble those of a neural network, as described above (the weight and bias parameters, and the two-level nonlinear signal transformation). This may suggest that neural-like features are necessary for computation even in a non-neural network. On the other hand, it also suggests that non-neural features may support neural computation. Indeed, this is the subject matter of ongoing debates in neuroscience about how much glia contribute to neural computation^{128,129}.

Our work contributes to the field of basal cognition by describing how a simple network of aneural cells may perform both basic and complex logic operations. However, our goal is not to validate basal cognition as a field – it is just one context (besides others, such as developmental neurobiology, evolutionary cognitive science, and synthetic bioengineering) which our findings impact. Even if some computational views of specific biological systems turn out to be wrong, our results provide value in showing how physiological circuits can be made with

specific (and useful) behaviors, both for synthetic biology applications and to understand functional aspects of developmental physiology, the evolution of brain circuits, etc. In prior work, a computational perspective of cell behavior^{14,16,130} allowed our lab and other groups to formulate (1) ways of experimentally controlling large-scale anatomical outcomes¹³, and (2) formulate and solve the “inverse problem”¹⁴ of how the cells might actually achieve specific anatomical configurations, given cellular constraints and a range of starting conditions^{76,131}. Our current work addresses (2), by demonstrating a possible approach that cells could use to implement logic functions under realistic cellular constraints.

Overall, BEN has the potential to shed light on as-yet-unexplained non-neural bioelectric information-processing. For example, when certain planarian species (highly regenerative animals) are cut they sometimes develop two heads, instead of a head and a tail, as a response to a temporary exposure to 1-octanol, a gap junction blocker. Without the gap junction blocker, they always develop a wild type morphology consisting of a single head and a tail. It has been shown that this decision to switch morphologies is bioelectrically controlled and non-neural in nature^{24,132}. We hypothesize that the planarian bioelectric system encodes a pattern that either consists of a record of the gap junction blocker or not, and then maps it into different morphological decisions (wild-type or double-head) in a systematic way. We currently have a prototype BEN model that reproduces this phenomenon in a qualitative way, which we plan to report in the future. Finally, we have demonstrated in this work that the slower (reason described in ‘Methods’), continuous mode of non-neural bioelectric signaling, compared to the faster, pulsating mode of neural signaling, is sufficient to perform logical computations usually associated with the brain. This firmly supports the possibility of somatic computation in real biological systems. How it compares to the timescales of genetic signaling and how it could facilitate bioelectricity to be upstream of genetics^{53,103} is an open question that our future investigations will answer.

Our work paves the foundation for future research in training actual biological tissues for somatic computation. Ongoing research in our laboratory has identified ways by which gap junctions can be manipulated and controlled, further opening up the possibilities on this front. Overall, our research provides the conceptual and modeling foundations to understand and manipulate development and regeneration, and to construct computational synthetic living machines.

Methods

Mathematical details of BEN. The equations that define a BEN model are shown in Fig. 8 using an example 2-cell system. The equations are shown for cell-2 only; the same equations with appropriate change in subscripts apply to cell-1 and likewise to any other cell in larger networks. The processes that the equations represent, like electrodiffusion, reaction-diffusion and gating, are marked appropriately in the figure, thus self-explanatory. The following remarks serve to better illustrate the model. As described in the introductory section, electrodiffusion utilizes two types of gradients, namely voltage and concentration, to drive the flux. Those gradients are represented by the terms $v_1(t) - v_2(t)$ and $c_1(t) - c_2(t)$ respectively in the electrodiffusion equations. The ion-pump equations are approximate versions of the full mechanism modeled in equations (23–29) of BETSE⁵¹. BEN makes use of just the terms involving the ion concentrations in equation (27) of BETSE⁵¹, as it lacks the other biochemical components like ATP, and by choosing appropriate term coefficients to compensate for the approximation it effectively preserves the relationship between ion concentrations and the pump flux.

The dynamics of the signaling molecule is viewed as a nonlinear reaction-diffusion process as it can be abstractly represented as: $R(D(\nabla c))$, where ∇c is the concentration gradient, $D(\cdot)$ represents the 1st layer of sigmoid approximating a nonlinear diffusion, and $R(\cdot)$ represents the 2nd layer of sigmoid approximating a nonlinear reaction. Moreover, the nested form of $R(D(\cdot))$ departs from the conventional form of reaction-diffusion which is $\frac{dc}{dt} = D(c) + R(c)$, for example, $\frac{dc}{dt} = \nabla c + c^2$. Nevertheless, it qualifies as reaction-diffusion since it clearly involves diffusion (∇c) and a nonlinear component (the sigmoid) that alters the net mass of c , representing production or decay depending on the sign of $\frac{dc}{dt}$. Another important point to note is that even though sigmoid functions, which define the signal flux, also constitute the activation functions of neurons in neural network models^{38,64}, there is a crucial difference: while neuron models are input-activated since neuronal synaptic junctions are directional, a cell in BEN is gradient-activated since gap-junctions are bidirectional. This is also partly the reason why non-neuronal cells tend to be slower than neural cells—diffusion is a slower process than directed currents.

There’s no set range of V_{mem} levels that a given BEN network will not exceed (as there are no explicit bounds on the ion concentration levels), but there are typically set to lie within $[-80 \text{ mV}, +80 \text{ mV}]$; see for example Figs. 2d, 3d, 4, 6 and 7. The concentration of the signaling molecule, on the other hand, is forced to lie with $[0, 1]$, that is, within the generic minimum and maximum possible values of the actual concentration which is not explicitly considered in BEN.

The software to simulate, train and evolve BEN networks is available at: <https://gitlab.com/smanicka/BEN>.

Details of the simulation. We used the standard Euler integration method for integration the equations. We used a time step of 0.01 for the bioelectric dynamics and 0.02 for the signaling molecule dynamics. Each simulation was set up to run for a number of steps ranging from 300 to 600; smaller networks were run for fewer steps. Thus, the total simulation ranged between $300 * 0.01 = 3$ simulation seconds to $600 * 0.01 = 6$ simulation seconds.

The backpropagation method for training elementary logic gates. Regardless of the gate, a training run was set up as follows. A single BEN network was instantiated with randomized parameters. The initial weight parameters were chosen in the range $[-1, 1]$, and the biases in the range $[0, 1]$. It was then fed with a batch of four inputs and was simulated for about 300 time-steps, with the inputs being held *fixed* throughout the simulation. An “input” constitutes a specific V_{mem} pattern of the input cells. For example, input cell-1 may be set to a



Figure 8. Mathematical details of the BEN model. (Top) The equations that describe the dynamics in a 2-cell system illustrated in the figure on the left. (Bottom) A glossary of variables used in the equations, the constants and initial conditions.

hyperpolarized V_{mem} and cell-2 to a depolarized V_{mem} . The input cells were then randomly set to a different state, drawn from the training batch, without disturbing the state of the rest of the network (a method referred to as “continuous computation”¹⁰¹) and the simulation was continued for another 300 time-steps; this was repeated for ten times. At the end of each of 10 simulations, the observed states of the output cell during the last 10 time-steps were matched against the desired output corresponding to the input in the training batch (see Figs. 2b and 3b for examples). An error was then calculated based on the difference for each of the 10 simulations for each input sequence in the batch and averaged over. The derivatives of the final average error (gradient) with respect to all the parameters were then calculated. Due to the recurrent nature of the dynamics (Fig. 1), the simulation takes many more steps than the number of layers in the network before the output reaches an equilibrium. Thus, the gradients would in principle have to be computed over all those steps—a method known as “backpropagation through time” (BPTT). However, due to computational constraints and known problems like “vanishing gradient”, the gradients are typically computed over fewer time steps—this version of backpropagation is known as “truncated” BPTT. In this work, the gradients were computed over only the last 100 steps of a simulation. The quantum of changes for all the parameters were then calculated from the gradients after scaling it with a “learning rate” that set the pace of learning. The weights and bias were assigned separate “dynamic” learning rates, where the rates were fixed in such a way that the maximum change of a weight parameter was 0.1 and the maximum change of a bias parameter was 0.01. We adopted this strategy mainly to mitigate the vanishing gradient problem, and the assumption that small changes in the parameters should cause a smooth change in behavior. We also employed a “momentum” factor, a method often used in backpropagation, that determines the extent to which the previous parameter-change vector contributed to the following vector. We used a momentum of 0.5 for the first 100 training iterations, and 0.9 for the rest; the allowed range is [0, 1]. Finally, the parameter-updates were applied (thus error was backpropagated), and the whole process was repeated for about 600,000 “epochs”. The weights were allowed to change

infinitely in both directions, whereas the biases were limited within the range [0, 1]. We deemed a network as successfully trained if it achieved an average performance error of 0.0002 or below at some point during the training.

We used the Python software package called *Pytorch* for computing derivatives during backpropagation. This package uses a method called “automatic differentiation” to compute derivatives based on “computational graphs”—a graph that keeps track of every computation performed during a simulation, which is then swept backwards for computing the derivatives, essentially constituting an algorithm for the “chain rule” of differential calculus.

The combined backpropagation-genetic-algorithm method for training tissue-level logic gate and the pattern detector. A population of 50 BEN “genomes” was instantiated. A genome consists of a combination of the network’s parameters, namely the weight matrix and the bias vector, that define a network. Henceforth, by “genome” we refer to a BEN network, for clarity. Each network consists of the following structure: an input layer consisting of 2 nodes, an intermediate “tissue” layer consisting of 25 nodes, and the output layer consisting of a single node. Initially the tissue layer was designed to be a two-dimensional lattice. Next, the tissue layer of each individual in the population was randomly rewired with a rewiring probability chosen uniformly from the range [0, 1]. Thus, the population consisted of the full spectrum of tissue layer structures ranging from lattice to random. Next, the input layer and the output layers of each individual in the population were respectively connected to approximately 50% of the nodes in the tissue layer; this gave the GA a chance to explore both denser and sparser inter-layer connectivity. Every individual in the population was then assigned randomized weight and bias parameters chosen in the same range as backpropagation. They were each then backpropagated for 80 iterations in the same fashion as described above for the elementary gates, with an important difference: the inputs for these gates were *transient*, as they are set for the first time-step only. The final average error of each individual became its fitness score. The GA then goes through the conventional steps of selection, mutation, creating a new generation of individuals.

We used a simple flavor of GA known as the “microbial genetic algorithm”¹³³ that capitalizes on the notion of horizontal gene transfer observed in microbes. It is essentially a form of tournament selection where two individuals in a population are picked and pitted against each other (their fitness scores are compared). We used a “geographical selection” method where individuals are placed on a 1D ring, and only geographically close individuals are picked for the tournament. The size of selection-neighborhood is known as the “deme size”, which was set to 10% in this work. The genome of the winner is transmitted to the loser at a certain “recombination rate”, which was set to 0.1. The genome of the loser is then mutated at a certain “mutation rate” (set to 0.05) and reinserted into the population. This process is repeated for 1000 generations. The individual with the best fitness score in the final generation was deemed as the best individual—the most successfully trained tissue-level logic gate.

Generation of the input patterns during the training of the pattern detector. The set of input patterns used for training the pattern detector consists of two categories: (1) French-flag (FF) and (2) Not-French-flag (NFF). The subset FF consists of the French-flag pattern itself and its slightly noisy variants. The latter is generated by perturbing each pixel of the French-flag approximately within the range [−24 mV, +24 mV]. Any perturbation that takes a pixel below the bounds of [−80 mV, +80 mV] are reset to the closest boundary values. The subset NFF consists of the more distorted variants of the French-flag. It is generated in the same way as FF except the perturbation range is [−40 mV, +40 mV]. A total of 20 samples in each set were generated during each iteration of the GA-BP search.

Data availability

All parameters used for the networks in this manuscript, as well as the source code, can be downloaded at <https://gitlab.com/smanicka/BEN>.

Received: 21 June 2019; Accepted: 13 November 2019;

Published online: 09 December 2019

References

- Wiener, N. & Schadé, J. P. *Cybernetics of the nervous system*. (Elsevier Pub. Co., 1965).
- Wiener, N. & Schadé, J. P. *Nerve, brain, and memory models*. (Elsevier Pub. Co., 1963).
- Simon, H. A. Studying human intelligence by creating artificial intelligence. *Am Sci* **69**, 300–309 (1981).
- Manicka, S. & Levin, M. The Cognitive Lens: a primer on conceptual tools for analysing information processing in developmental and regenerative morphogenesis. *Philos Trans R Soc Lond B Biol Sci* **374**, 20180369, <https://doi.org/10.1098/rstb.2018.0369> (2019).
- Moore, D., Walker, S. I. & Levin, M. Cancer as a disorder of patterning information: computational and biophysical perspectives on the cancer problem. *Convergent Science Physical. Oncology* **3**, 043001 (2017).
- Levin, M. & Martyniuk, C. J. The bioelectric code: An ancient computational medium for dynamic control of growth and form. *Biosystems*, <https://doi.org/10.1016/j.biosystems.2017.08.009> (2017).
- Pezzulo, G. & Levin, M. Top-down models in biology: explanation and control of complex living systems above the molecular level. *J R Soc Interface* **13**, <https://doi.org/10.1098/rsif.2016.0555> (2016).
- Baluška, F. & Levin, M. On Having No Head: Cognition throughout Biological Systems. *Front Psychol* **7**, 902, <https://doi.org/10.3389/fpsyg.2016.00902> (2016).
- Tang, S. K. Y. & Marshall, W. F. Cell learning. *Curr Biol* **28**, R1180–R1184, <https://doi.org/10.1016/j.cub.2018.09.015> (2018).
- Mitchell, A. & Lim, W. Cellular perception and misperception: Internal models for decision-making shaped by evolutionary experience. *Bioessays* **38**, 845–849, <https://doi.org/10.1002/bies.201600090> (2016).
- Lyon, P. The cognitive cell: bacterial behavior reconsidered. *Front Microbiol* **6**, 264, <https://doi.org/10.3389/fmicb.2015.00264> (2015).
- Silva-Rocha, R., Tamames, J., dos Santos, V. M. & de Lorenzo, V. The logicome of environmental bacteria: merging catabolic and regulatory events with Boolean formalisms. *Environ Microbiol* **13**, 2389–2402, <https://doi.org/10.1111/j.1462-2920.2011.02455.x> (2011).

13. Vandenberg, L. N., Adams, D. S. & Levin, M. Normalized shape and location of perturbed craniofacial structures in the *Xenopus* tadpole reveal an innate ability to achieve correct morphology. *Dev Dyn* **241**, 863–878, <https://doi.org/10.1002/dvdy.23770> (2012).
14. Pezzulo, G. & Levin, M. Re-membering the body: applications of computational neuroscience to the top-down control of regeneration of limbs and other complex organs. *Integr Biol (Camb)* **7**, 1487–1517, <https://doi.org/10.1039/c5ib00221d> (2015).
15. Fariella-Ferruzza, N. The transformation of a tail into a limb after xenoplastic transformation. *Experientia* **15**, 304–305 (1956).
16. Friston, K., Levin, M., Sengupta, B. & Pezzulo, G. Knowing one's place: a free-energy approach to pattern regulation. *J R Soc Interface* **12**, <https://doi.org/10.1098/rsif.2014.1383> (2015).
17. Lyon, P. The biogenic approach to cognition. *Cogn Process* **7**, 11–29, <https://doi.org/10.1007/s10339-005-0016-8> (2006).
18. Prindle, A. *et al.* Ion channels enable electrical communication in bacterial communities. *Nature*, <https://doi.org/10.1038/nature15709> (2015).
19. Levin, M. Morphogenetic fields in embryogenesis, regeneration, and cancer: non-local control of complex patterning. *Bio Systems* **109**, 243–261, <https://doi.org/10.1016/j.biosystems.2012.04.005> (2012).
20. Bates, E. Ion Channels in Development and Cancer. *Annu Rev Cell Dev Biol* **31**, 231–247, <https://doi.org/10.1146/annurev-cellbio-100814-125338> (2015).
21. Pietak, A. & Levin, M. Bioelectrical control of positional information in development and regeneration: A review of conceptual and computational advances. *Prog Biophys Mol Biol* **137**, 52–68, <https://doi.org/10.1016/j.pbiomolbio.2018.03.008> (2018).
22. McLaughlin, K. A. & Levin, M. Bioelectric signaling in regeneration: Mechanisms of ionic controls of growth and form. *Dev Biol* **433**, 177–189, <https://doi.org/10.1016/j.ydbio.2017.08.032> (2018).
23. Mathews, J. & Levin, M. The body electric 2.0: recent advances in developmental bioelectricity for regenerative and synthetic bioengineering. *Curr Opin Biotechnol* **52**, 134–144, <https://doi.org/10.1016/j.copbio.2018.03.008> (2018).
24. Durant, F. *et al.* Long-Term, Stochastic Editing of Regenerative Anatomy via Targeting Endogenous Bioelectric Gradients. *Biophys J* **112**, 2231–2243, <https://doi.org/10.1016/j.bpj.2017.04.011> (2017).
25. Beane, W. S., Morokuma, J., Adams, D. S. & Levin, M. A chemical genetics approach reveals H,K-ATPase-mediated membrane voltage is required for planarian head regeneration. *Chemistry & Biology* **18**, 77–89 (2011).
26. Pitcairn, E. *et al.* Coordinating heart morphogenesis: A novel role for Hyperpolarization-activated cyclic nucleotide-gated (HCN) channels during cardiogenesis in *Xenopus laevis*. *Communicative & Integrative Biology* **10**, e1309488, <https://doi.org/10.1080/19420889.2017.1309488> (2017).
27. Pai, V. P. *et al.* HCN4 ion channel function is required for early events that regulate anatomical left-right patterning in a nodal and lefty asymmetric gene expression-independent manner. *Biology Open* **6**, 1445–1457, <https://doi.org/10.1242/bio.025957> (2017).
28. Pai, V. P., Aw, S., Shomrat, T., Lemire, J. M. & Levin, M. Transmembrane voltage potential controls embryonic eye patterning in *Xenopus laevis*. *Development* **139**, 313–323, <https://doi.org/10.1242/dev.073759> (2012).
29. Pai, V. P. *et al.* Endogenous Gradients of Resting Potential Instructively Pattern Embryonic Neural Tissue via Notch Signaling and Regulation of Proliferation. *The Journal of Neuroscience* **35**, 4366–4385, <https://doi.org/10.1523/JNEUROSCI.1877-14.2015> (2015).
30. Sundelacruz, S., Levin, M. & Kaplan, D. L. Membrane potential controls adipogenic and osteogenic differentiation of mesenchymal stem cells. *PLoS One* **3**, e3737 (2008).
31. Perathoner, S. *et al.* Bioelectric signaling regulates size in zebrafish fins. *PLoS genetics* **10**, e1004080, <https://doi.org/10.1371/journal.pgen.1004080> (2014).
32. Tseng, A. S., Beane, W. S., Lemire, J. M., Masi, A. & Levin, M. Induction of vertebrate regeneration by a transient sodium current. *J Neurosci* **30**, 13192–13200, 30/39/13192 [pii], <https://doi.org/10.1523/JNEUROSCI.3315-10.2010> (2010).
33. Adams, D. S., Masi, A. & Levin, M. H⁺ pump-dependent changes in membrane voltage are an early mechanism necessary and sufficient to induce *Xenopus* tail regeneration. *Development* **134**, 1323–1335 (2007).
34. Oviedo, N. J. *et al.* Long-range neural and gap junction protein-mediated cues control polarity during planarian regeneration. *Dev Biol* **339**, 188–199, S0012-1606(09)01402-X [pii], <https://doi.org/10.1016/j.ydbio.2009.12.012> (2010).
35. Adams, D. S. *et al.* Bioelectric signalling via potassium channels: a mechanism for craniofacial dysmorphogenesis in KCNJ2-associated Andersen-Tawil Syndrome. *J Physiol* **594**, 3245–3270, <https://doi.org/10.1113/JP271930> (2016).
36. Messori, A. *et al.* Keppen-Lubinsky Syndrome Is Caused by Mutations in the Inwardly Rectifying K(+) Channel Encoded by KCNJ6. *Am J Hum Genet* **96**, 295–300, <https://doi.org/10.1016/j.ajhg.2014.12.011> (2015).
37. Pai, V. P. *et al.* HCN2 Rescues brain defects by enforcing endogenous voltage pre-patterns. *Nature Communications* **9**, <https://doi.org/10.1038/s41467-018-03334-5> (2018).
38. McCulloch, W. S. & Pitts, W. A logical calculus of the ideas immanent in nervous activity. *Bulletin of Mathematical Biophysics* **5** (1943).
39. Goldental, A., Guberman, S., Vardi, R. & Kanter, I. A computational paradigm for dynamic logic-gates in neuronal activity. *Front Comput Neurosci* **8**, 52, <https://doi.org/10.3389/fncom.2014.00052> (2014).
40. Albert, R. E. In *Complex Networks. Lecture Notes in Physics* Vol. 650 (ed Frauenfelder H. Ben-Naim E., Toroczkai Z.) (Springer, 2004).
41. Albert, R. & Thakar, J. Boolean modeling: a logic-based dynamic approach for understanding signaling and regulatory networks and for making useful predictions. *Wiley Interdisciplinary Reviews: Systems Biology and Medicine* **6**, 353–369, <https://doi.org/10.1002/wsbm.1273> (2014).
42. Crama, Y. & Peter L. Hammer. *Boolean functions: Theory, algorithms, and applications*. (Cambridge University Press, 2011).
43. Copeland, B. J. What is computation. *Synthese* **108**, 335–359 (1996).
44. Horsman, C., Stepney, S., Wagner, R. C. & Kendon, V. When does a physical system compute? *Proc Math Phys Eng Sci* **470**, 20140182, <https://doi.org/10.1098/rspa.2014.0182> (2014).
45. Nicholson, D. J. Is the cell really a machine? *J Theor Biol* **477**, 108–126, <https://doi.org/10.1016/j.jtbi.2019.06.002> (2019).
46. Harvey, I. Cognition is Not Computation; Evolution is Not Optimisation (1996).
47. Pietak, A. & Levin, M. Exploring Instructive Physiological Signaling with the Bioelectric Tissue Simulation Engine. *Front Bioeng Biotechnol* **4**, 55, <https://doi.org/10.3389/fbioe.2016.00055> (2016).
48. Javier Cervera, A. P. & Levin, M. Salvador Mafe. Bioelectrical coupling in multicellular domains regulated by gap junctions - A conceptual approach. *Bioelectrochemistry*, 45–61, <https://doi.org/10.1016/j.bioelechem.2018.04.013> (2018).
49. Pai, V. P. *et al.* HCN2 Rescues brain defects by enforcing endogenous voltage pre-patterns. *Nat Commun* **9**, 998, <https://doi.org/10.1038/s41467-018-03334-5> (2018).
50. Levin, M., Pietak, A. M. & Bischof, J. Planarian regeneration as a model of anatomical homeostasis: Recent progress in biophysical and computational approaches. *Semin Cell Dev Biol*, <https://doi.org/10.1016/j.semcdb.2018.04.003> (2018).
51. Pietak, A. & Levin, M. Bioelectric gene and reaction networks: computational modelling of genetic, biochemical and bioelectrical dynamics in pattern regulation. *J R Soc Interface* **14**, <https://doi.org/10.1098/rsif.2017.0425> (2017).
52. Mathews, J. & Levin, M. Gap junctional signaling in pattern regulation: Physiological network connectivity instructs growth and form. *Developmental neurobiology* **77**, 643–673, <https://doi.org/10.1002/dneu.22405> (2017).
53. Koseska, A. & Bastiaens, P. I. Cell signaling as a cognitive process. *EMBO J* **36**, 568–582, <https://doi.org/10.15252/embj.201695383> (2017).
54. Barnard, A., Wolfe, A. & Busby, S. Regulation at complex bacterial promoters: how bacteria use different promoter organizations to produce different regulatory outcomes. *Curr Opin Microbiol* **7**, 102–108, <https://doi.org/10.1016/j.mib.2004.02.011> (2004).

55. Brodskiy, P. A. & Zartman, J. J. Calcium as a signal integrator in developing epithelial tissues. *Physical biology* **15**, 051001, <https://doi.org/10.1088/1478-3975/aabb18> (2018).
56. Guillermo Solovey, S. P. D. Intra-Cluster Percolation of Calcium Signals. *PLoS One* **5**, <https://doi.org/10.1371/journal.pone.0008997> (2010).
57. Lopez, L., Piegari, E., Sigaut, L. & Ponce Dawson, S. Intracellular calcium signals display an avalanche-like behavior over multiple lengthscales. *Front Physiol* **3**, 350, <https://doi.org/10.3389/fphys.2012.00350> (2012).
58. Polsky, A., Mel, B. W. & Schiller, J. Computational subunits in thin dendrites of pyramidal cells. *Nat Neurosci* **7**, 621–627, <https://doi.org/10.1038/nn1253> (2004).
59. Panayioti Poirazi, T. B. & Bartlett, W. Mel. Pyramidal neuron as two-layer neural network. *Neuron* **37**, 989–999 (2003).
60. Antic, S. D., Hines, M. & Lytton, W. W. Embedded ensemble encoding hypothesis: The role of the “Prepared” cell. *J Neurosci Res*, <https://doi.org/10.1002/jnr.24240> (2018).
61. Sperelakis, N. *Cell physiology source book: essentials of membrane biophysics*. (Elsevier, 2012).
62. Bukauskas, F. F. & Verselis, V. K. Gap junction channel gating. *Biochim Biophys Acta* **1662**, 42–60, <https://doi.org/10.1016/j.bbame.2004.01.008> (2004).
63. Brink, P. Gap Junction Voltage Dependence - a clear picture emerges. *J. Gen. Physiol* **116**, 11–12 (2000).
64. Nielsen, M. A. *Neural networks and deep learning*. Vol. 25 (Determination press, 2015).
65. Brodmann, K. *Brodmann's Localisation in the cerebral cortex*. (Springer Science & Business Media, 2007).
66. Hinczewski, M. & Thirumalai, D. Cellular signaling networks function as generalized Wiener-Kolmogorov filters to suppress noise. *Physical Review X* **4** (2014).
67. Becker, N. B., Mugler, A. & Ten Wolde, P. R. Optimal Prediction by Cellular Signaling Networks. *Phys Rev Lett* **115**, 258103, <https://doi.org/10.1103/PhysRevLett.115.258103> (2015).
68. Sagner, A. & Briscoe, J. Morphogen interpretation: concentration, time, competence, and signaling dynamics. *Wiley Interdiscip Rev Dev Biol* **6**, <https://doi.org/10.1002/wdev.271> (2017).
69. Ebisuya, M. & Briscoe, J. What does time mean in development? *Development* **145**, <https://doi.org/10.1242/dev.164368> (2018).
70. Herrera-Rincon, C. *et al.* Brief Local Application of Progesterone via a Wearable Bioreactor Induces Long-Term Regenerative Response in Adult Xenopus Hindlimb. *Cell Rep* **25**, 1593–1609 e1597, <https://doi.org/10.1016/j.celrep.2018.10.010> (2018).
71. Amor, D. R., Ratzke, C. & Gore, J. Transient invaders can induce shifts between alternative stable states of microbial communities. *BioRxiv*, <https://doi.org/10.1101/659052> (2019).
72. Durant, F. *et al.* The role of early bioelectric signals in the regeneration of planarian anterior-posterior polarity. *Biophys J*, <https://doi.org/10.1016/j.bpj.2019.01.029> (2019).
73. Davidson, S. I. A. E. H. Logic functions of the genomic cis-regulatory code. *PNAS* **102**, 4954–4959 (2004).
74. Palacios-Prado, N. & Bukauskas, F. F. Heterotypic gap junction channels as voltage-sensitive valves for intercellular signaling. *Proc Natl Acad Sci USA* **106**, 14855–14860, 0901923106 [pii], <https://doi.org/10.1073/pnas.0901923106> (2009).
75. Birnbaum, K. D. & Alvarado, A. S. Slicing across kingdoms: regeneration in plants and animals. *Cell* **132**, 697–710 (2008).
76. Pinet, K. & McLaughlin, K. A. Mechanisms of physiological tissue remodeling in animals: Manipulating tissue, organ, and organism morphology. *Dev Biol*, <https://doi.org/10.1016/j.ydbio.2019.04.001> (2019).
77. Wolpert, L. Positional information and the spatial pattern of cellular differentiation. *J. Theor. Biol* **25**, 1–47 (1969).
78. Dawkins, R. *The blind watchmaker: Why the evidence of evolution reveals a universe without design*. (WW Norton & Company, 1996).
79. Espinosa-Soto, C. On the role of sparseness in the evolution of modularity in gene regulatory networks. *PLoS computational biology* **14** (2018).
80. Hilger, K., Ekman, M., Fiebach, C. J. & Basten, U. Intelligence is associated with the modular structure of intrinsic brain networks. *Scientific reports* **7** (2017).
81. Meunier, D., Renaud, L. & Edward T. Bullmore. Modular and hierarchically modular organization of brain networks. *Frontiers in neuroscience* **4** (2010).
82. Marchisio, M. A. & Stelling, J. Automatic design of digital synthetic gene circuits. *PLoS Comput Biol* **7**, e1001083, <https://doi.org/10.1371/journal.pcbi.1001083> (2011).
83. Alon, U. *An Introduction to Systems Biology: Design Principles of Biological Circuits*. (Chapman and Hall/CRC Press, 2006).
84. Wang, B., Kitney, R. I., Joly, N. & Buck, M. Engineering modular and orthogonal genetic logic gates for robust digital-like synthetic biology. *Nat Commun* **2**, 508, <https://doi.org/10.1038/ncomms1516> (2011).
85. Amos, A. G. N.-M. A. M. A reconfigurable NAND-NOR genetic logic gate. *BMC Syst Biol* **6** (2012).
86. Sole, R., Moses, M. & Forrest, S. Liquid brains, solid brains. *Philos Trans R Soc Lond B Biol Sci* **374**, 20190040, <https://doi.org/10.1098/rstb.2019.0040> (2019).
87. Chaitin, G. J. *Proving Darwin: Making Biology Mathematical*. (Pantheon Books, 2012).
88. Adamatzky, A. A brief history of liquid computers. *Philos Trans R Soc Lond B Biol Sci* **374**, 20180372, <https://doi.org/10.1098/rstb.2018.0372> (2019).
89. Allen Hjelmfelt, E. D. W. & Ross, J. Chemical implementation of finite-state machines. *PNAS* **89**, 383–387 (1992).
90. Allen Hjelmfelt, E. D. W. & Ross, J. Chemical implementation of neural networks and Turing machines. *PNAS* **88**, 10983–10987 (1991).
91. Egbert, M., Gagnon, J. S. & Perez-Mercader, J. Dynamic modulation of external conditions can transform chemistry into logic gates. *J R Soc Interface* **15**, <https://doi.org/10.1098/rsif.2018.0169> (2018).
92. Holley, J., Jahan, I., Costello Bde, L., Bull, L. & Adamatzky, A. Logical and arithmetic circuits in Belousov-Zhabotinsky encapsulated disks. *Phys Rev E Stat Nonlin Soft Matter Phys* **84**, 056110 (2011).
93. Privman, V., Arugula, M. A., Halámek, J., Pita, M. & Katz, E. Network Analysis of Biochemical Logic for Noise Reduction and Stability - A System of Three Coupled Enzymatic AND Gates. *J. Phys. Chem. B*, 5301–5310 (2009).
94. McGregor, S., Vasas, V., Husbands, P. & Fernando, C. Evolution of associative learning in chemical networks. *PLoS computational biology* **8**, e1002739, <https://doi.org/10.1371/journal.pcbi.1002739> (2012).
95. Fernando, C. T. *et al.* Molecular circuits for associative learning in single-celled organisms. *J R Soc Interface* **6**, 463–469, <https://doi.org/10.1098/rsif.2008.0344> (2009).
96. Cherry, K. M. & Qian, L. Scaling up molecular pattern recognition with DNA-based winner-take-all neural networks. *Nature* **559**, 370–376, <https://doi.org/10.1038/s41586-018-0289-6> (2018).
97. Nicolas E. B. U. G. & Terence H. On schemes of combinatorial transcription logic. *PNAS* **100**, 5136–5141 (2003).
98. Woods, D. *et al.* Diverse and robust molecular algorithms using reprogrammable DNA self-assembly. *Nature* **567**, 366–372, <https://doi.org/10.1038/s41586-019-1014-9> (2019).
99. Sorek, M., Balaban, N. Q. & Loewenstein, Y. Stochasticity, bistability and the wisdom of crowds: a model for associative learning in genetic regulatory networks. *PLoS computational biology* **9**, e1003179, <https://doi.org/10.1371/journal.pcbi.1003179> (2013).
100. Watson, R. A., Wagner, G. P., Pavlicev, M., Weinreich, D. M. & Mills, R. The evolution of phenotypic correlations and “developmental memory”. *Evolution* **68**, 1124–1138, <https://doi.org/10.1111/evo.12337> (2014).
101. Goni-Moreno, A. & Amos, M. Continuous computation in engineered gene circuits. *Biosystems* **109**, 52–56, <https://doi.org/10.1016/j.biosystems.2012.02.001> (2012).

102. Bradley, R. W., Buck, M. & Wang, B. Recognizing and engineering digital-like logic gates and switches in gene regulatory networks. *Curr Opin Microbiol* **33**, 74–82, <https://doi.org/10.1016/j.mib.2016.07.004> (2016).
103. TerAvest, M. A., Li, Z. & Angenent, L. T. Bacteria-based biocomputing with Cellular Computing Circuits to sense, decide, signal, and act. *Energy & Environmental Science* **4**, <https://doi.org/10.1039/c1ee02455h> (2011).
104. Ayrinhac, S. Electric current solves mazes. *Phys. Educ* **49** (2014).
105. Safonov, A. A. Computing via natural erosion of sandstone. *International Journal of Parallel, Emergent and Distributed Systems* **33**, 742–751, <https://doi.org/10.1080/17445760.2018.1455836> (2018).
106. Ashikaga, H. *et al.* Modelling the heart as a communication system. *J R Soc Interface* **12**, <https://doi.org/10.1098/rsif.2014.1201> (2015).
107. Srinivasa V. & Chakravarthy, J. G. On Hebbian-like adaptation in heart muscle: a proposal for ‘cardiac memory’ (1997).
108. Gatenby, R. A. & Frieden, B. R. Cellular information dynamics through transmembrane flow of ions. *Sci Rep* **7**, 15075, <https://doi.org/10.1038/s41598-017-15182-2> (2017).
109. Couto, D. & Zipfel, C. Regulation of pattern recognition receptor signalling in plants. *Nat Rev Immunol* **16**, 537–552, <https://doi.org/10.1038/nri.2016.77> (2016).
110. Stuckemann, T. *et al.* Antagonistic Self-Organizing Patterning Systems Control Maintenance and Regeneration of the Anteroposterior Axis in Planarians. *Dev Cell* **40**, 248–263 e244, <https://doi.org/10.1016/j.devcel.2016.12.024> (2017).
111. Glen, C. M., McDevitt, T. C. & Kemp, M. L. Dynamic intercellular transport modulates the spatial patterning of differentiation during early neural commitment. *Nat Commun* **9**, 4111, <https://doi.org/10.1038/s41467-018-06693-1> (2018).
112. Gross, P. *et al.* Guiding self-organized pattern formation in cell polarity establishment. *Nature Physics*, <https://doi.org/10.1038/s41567-018-0358-7> (2018).
113. Karig, D. *et al.* Stochastic Turing patterns in a synthetic bacterial population. *Proc Natl Acad Sci USA*, <https://doi.org/10.1073/pnas.1720770115> (2018).
114. Ferreira, G. B., Matthias, S. & Levin, M. In *European Conference on Artificial Life*. 170–177 (MIT Press).
115. Minh-Thai, T. N., Aryal, J., Samarasinghe, S. & Levin, M. In *AI 2018: Advances in Artificial Intelligence Lecture Notes in Computer Science* Ch. Chapter 16, 153–159 (2018).
116. De, A., Chakravarthy, V. S. & Levin, M. A computational model of planarian regeneration. *International Journal of Parallel, Emergent and Distributed Systems* **32**, 331–347, <https://doi.org/10.1080/17445760.2016.1185521> (2016).
117. Levin, M. Z. B. A. M. In *Artificial Life* 351–358 (MIT Press, 2018).
118. Cervera, J., Manzanares, J. A., Mafe, S. & Levin, M. Synchronization of Bioelectric Oscillations in Networks of Nonexcitable Cells: From Single-Cell to Multicellular States. *J Phys Chem B* **123**, 3924–3934, <https://doi.org/10.1021/acs.jpcc.9b01717> (2019).
119. Brodsky, M. Turing-like patterns can arise from purely bioelectric mechanisms. *bioRxiv*, <https://doi.org/10.1101/336461> (2018).
120. Cervera, J., Manzanares, J. A. & Mafe, S. Cell-cell bioelectrical interactions and local heterogeneities in genetic networks: a model for the stabilization of single-cell states and multicellular oscillations. *Phys Chem Chem Phys* **20**, 9343–9354, <https://doi.org/10.1039/C8CP00648B> (2018).
121. Blackiston, D., Shomrat, T. & Levin, M. The Stability of Memories During Brain Remodeling: a Perspective. *Communicative & Integrative Biology* **8**, e1073424, <https://doi.org/10.1080/19420889.2015.1073424> (2015).
122. Cooke, J. Scale of body pattern adjusts to available cell number in amphibian embryos. *Nature* **290**, 775–778 (1981).
123. Bellec, G. *et al.* Biologically inspired alternatives to backpropagation through time for learning in recurrent neural nets. *ArXiv* (2019).
124. Stork, D. G. Is backpropagation biologically plausible? *International Joint Conference on Neural Networks* (1989).
125. Öfjäll, K. & Felsberg, M. In *British Machine Vision Conference* 137–156 (BMVA Press, Nottingham, UK, 2014).
126. Schillaci, G., Hafner, V. V. & Lara, B. In *4th International Conference on Development and Learning and on Epigenetic Robotics* 130–136 (IEEE, 2014).
127. Kak, S. C. On generalization by neural networks. *Information Sciences* **111**, 293–302 (1998).
128. Araque, A. & Navarrete, M. Glial cells in neuronal network function. *Philosophical Transactions of the Royal Society B: Biological Sciences* **365**, 2375–2381 (2010).
129. De Pittà, M. B. H. In *Computational Glioscience* (ed. Berry H. & De Pittà M.) (Springer, Cham, 2019).
130. Levin, M. & Martyniuk, C. J. The bioelectric code: An ancient computational medium for dynamic control of growth and form. *Biosystems* **164**, 76–93, <https://doi.org/10.1016/j.biosystems.2017.08.009> (2018).
131. Pinet, K., Deolankar, M., Leung, B. & McLaughlin, K. A. Adaptive correction of craniofacial defects in pre-metamorphic *Xenopus laevis* tadpoles involves thyroid hormone-independent tissue remodeling. *Development* **146**, <https://doi.org/10.1242/dev.175893> (2019).
132. Emmons-Bell, M. *et al.* Gap Junctional Blockade Stochastically Induces Different Species-Specific Head Anatomies in Genetically Wild-Type *Girardia dorotocephala* Flatworms. *Int J Mol Sci* **16**, 27865–27896, <https://doi.org/10.3390/ijms161126065> (2015).
133. Harvey, I. In *Advances in Artificial Life. Darwin Meets von Neumann. ECAL 2009*. (ed. Karsai, I., Kampis, G. & Szathmáry, E.) 126–133 (Springer, Berlin, Heidelberg).

Acknowledgements

We thank Alexis Pietak and many members of the Levin lab for many helpful discussions, as well as Nima Dehghani and Joshua Finkelstein for helpful comments on a draft of the manuscript. This research was supported by the Allen Discovery Center program through The Paul G. Allen Frontiers Group (12171), and M. L. gratefully acknowledges support from the Templeton World Charity Foundation (TWCF0089/AB55), the Barton Family Foundation, and the National Science Foundation (DBI-1152279 and Emergent Behaviors of Integrated Cellular Systems subaward CBET-0939511). This research was also sponsored by the Defense Advanced Research Projects Agency (DARPA) under Cooperative Agreement Number HR0011-18-2-0022, the Lifelong Learning Machines program from DARPA/MTO. The content of the information does not necessarily reflect the position or the policy of the Government, and no official endorsement should be inferred. Approved for public release; distribution is unlimited.

Author contributions

M.L. and S.M. conceptualized the project together. S.M. wrote all the code and performed the computational experiments and data collection. M.L. and S.M. analyzed the data and wrote the paper.

Competing interests

The authors declare no competing interests.

Additional information

Supplementary information is available for this paper at <https://doi.org/10.1038/s41598-019-54859-8>.

Correspondence and requests for materials should be addressed to M.L.

Reprints and permissions information is available at www.nature.com/reprints.

Publisher's note Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.



Open Access This article is licensed under a Creative Commons Attribution 4.0 International License, which permits use, sharing, adaptation, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons license, and indicate if changes were made. The images or other third party material in this article are included in the article's Creative Commons license, unless indicated otherwise in a credit line to the material. If material is not included in the article's Creative Commons license and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a copy of this license, visit <http://creativecommons.org/licenses/by/4.0/>.

© The Author(s) 2019