



Perspective

Safely Targeting Cancer, the Wound That Never Heals, Utilizing CBP/Beta-Catenin Antagonists

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Simple Summary: Unresolved wound healing is a critical factor in cancer. More generally, inappropriate wound healing can promote chronic progressive fibroinflammatory diseases, including organ fibrosis and neurodegeneration. Transient loss of lineage fidelity is important for wound healing however, persistent lineage infidelity is associated with cancer and fibrosis. CBP/ β -catenin antagonists can safely target stem cells to induce differentiation and restore lineage fidelity to treat cancer, fibrosis and neurodegenerative diseases.

Abstract: Stem cells, both normal somatic (SSC) and cancer stem cells (CSC) exist in minimally two states, i.e., quiescent and activated. Regulation of these two states, including their reliance on different metabolic processes, i.e., FAO and glycolysis in quiescent versus activated stem cells respectively, involves the analysis of a complex array of factors (nutrient and oxygen levels, adhesion molecules, cytokines, etc.) to initiate the epigenetic changes to either depart or enter quiescence. Quiescence is a critical feature of SSC that is required to maintain the genomic integrity of the stem cell pool, particularly in long lived complex organisms. Quiescence in CSC, whether they are derived from mutations arising in SSC, aberrant microenvironmental regulation, or via dedifferentiation of more committed progenitors, is a critical component of therapy resistance and disease latency and relapse. At the beginning of vertebrate evolution, approximately 450 million years ago, a gene duplication generated the two members of the Kat3 family, CREBBP (CBP) and EP300 (p300). Despite their very high degree of homology, these two Kat3 coactivators play critical and non-redundant roles at enhancers and super-enhancers via acetylation of H3K27, thereby controlling stem cell quiescence versus activation and the cells metabolic requirements. In this review/perspective, we discuss the unique regulatory roles of CBP and p300 and how specifically targeting the CBP/β-catenin interaction utilizing small molecule antagonists, can correct lineage infidelity and safely eliminate quiescent CSC.

Keywords: CBP; p300; β -catenin; somatic stem cell (SSC); cancer stem cell (CSC); fatty acid oxidation (FAO); glycolysis; fibrosis; cancer; quiescence



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1. Introduction

Rudolf Virchow in 1858 first proposed the concept that "cancer is a wound that never heals" [1]. More recently the similarities between cancer and wounds have reemerged as a topic of investigation [2,3]. Patients with chronic wounds have increased cancer susceptibility [4,5] and in mice, mutations that retard the activation of stem cells, display reduced efficiency in wound closure, with increased cancer resistance [6].

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Tissue resident adult mammalian somatic stem cells (SSC) are responsible for both homeostasis and wound repair. The accumulation of mutations in normal SSC that causes them to lose their homeostatic balance, favoring self-renewal with subsequent tissue overgrowth at the expense of differentiation, is associated with tumorigenesis. A transient growth versus differentiation imbalance is created during wound repair. In SSC, Wnt signaling contributes to wound repair and tissue regeneration that is hijacked by cancer stem cells (CSC) [7-9]. Stem cell lineage infidelity occurs transiently in wounds, however it persists in cancer [3]. Cellular plasticity and lineage infidelity are associated with differential super-enhancer (SE) activation and critical in both wound repair and malignancies [10–12]. Multiple factors contribute to stem cell lineage infidelity, including cellular metabolism, hypoxia, inflammation and mechanical features of the tumor microenvironment (TME) [13]. Stem cell lineage infidelity and the epigenetic reprogramming associated with it, mediate immune escape in CSC [13,14]. Tumor-intrinsic Wnt/ β -catenin signaling is associated with cancer immune evasion, immunosuppressive cell subsets, defective dendritic cell presentation and T-effector cell recruitment [15,16]. Wnt signaling is critical in stem cell biology and the lineage infidelity associated with it in wound healing and cancer [17–19], fibrosis [20] and neurodegeneration [21].

Aging effects Wnt/ β -catenin signaling [22,23] and significantly negatively impacts healing by prolonging the inflammatory phase, which can lead to chronic wound healing [24]. Age-related lineage infidelity or precocious differentiation has been seen in various tissues, related to both intrinsic changes in SSC and to the stem cell niche microenvironment [25–28]. The role of cellular plasticity and lineage infidelity in an array of diseases, which is further exacerbated with aging, highlights the importance of developing therapies to safely manipulate endogenous "stemness" (both normal and cancerous) and correct lineage infidelity via modulation of Wnt/ β -catenin-regulated transcription [22,29,30].

2. Discussion

2.1. Wnt Signaling in Wound Healing

There are three important components to wound healing: epithelial movement, cell proliferation, and contraction (or remodeling) [31]. Wounding activated Wnt signaling is important in all stages of the healing process, including stem cell mobilization within the wound site, the control of inflammation, and remodeling [32]. At the wound edge, cells dedifferentiate to repair the wound [33–35]. The Wnt/ β -catenin, Hippo/YAP-TEAD, TGF- β /Smad, SOX and Notch pathways can all play a role in the dedifferentiation of more committed cells as part of a regenerative program required for wound healing [10,36–39]. Blocking Wnt signaling impairs normal fin regeneration after amputation in zebrafish [40] and inhibiting Wnt signaling results in abrupt cessation of regeneration in animals with continually regenerating retinas [41–44]. Wnt signaling disruption blocks the recruitment of stem/progenitor cells to the wound [41,45–47] adversely affecting the proliferative phase of the wound healing process [48,49]. Wnt signaling clearly has dichotomous roles in the inflammatory response to wounding, playing both pro- and anti-inflammatory roles [32]. Wnt signaling is critical in normal wound healing, however chronic injury can aberrantly prolong Wnt signaling, thereby increasing the risk of oncogenic transformation [50,51].

2.2. Stem Cells; Normal Somatic Stem Cells (SSC) and Cancer Stem Cells (CSC)

With aging, significant deterioration in stem cell functionality occurs, leading to reduced (e.g., thinning of the epidermis and dermis) and/or aberrant tissue regeneration (e.g., fibrosis), increased degenerative diseases of the brain [52] (e.g., Alzheimer's disease), skeletal muscle [53], skin [54,55] and cancer [56]. A limited pool exists of essentially immortal, generally quiescent SSC that serve as a reservoir for tissue regeneration. SSC

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exhibit inherent plasticity and exist in different states, minimally, quiescent versus activated. Infrequently entering the cell cycle during homeostasis, upon injury to repair damaged tissue, long-lived quiescent SSC do so more frequently. Through acquired mutations, SSC can become CSC, representing a "dark side" to their immortality. Similar to SSC, CSC can self-renew and differentiate, thereby maintaining, renewing and propagating tumors. Therapy resistance, disease relapse, and metastasis constitute the central challenges to more effective cancer therapy. The ability to eliminate CSC is the key to overcoming these challenges [57]. CSC may derive from long lived quiescent SSC. However, they can also originate from more differentiated cells via dedifferentiation to become CSC, particularly within the context of wounding and inflammation [58,59]. For example, the combination of oncogenic Kras and the activation of NF-kB, strongly activates Wnt signaling in intestinal epithelial cells (IEC), inducing dedifferentiation to a progenitor/stem cell-like state to generate stem-like IEC [60]. Niche inflammatory signals protect stem cells from cytotoxic stress and prevent differentiation via NF-kB induced Wnt signaling [61]. Cells with stem cell-like properties can also be generated via epithelial to mesenchymal transition (EMT) [62]. The tumor microenvironment via secretion of TGF- β inducing ZEB1 expression drives an EMT transition, critical for the conversion of non-CSC to CSC and the maintenance of the CSC-like state [63]. Secretion of hepatocyte growth factor by myofibroblasts inducing nuclear translocation of β -catenin activates Wnt signaling, thereby generating stem-like features in more differentiated colorectal tumor cells. This demonstrates that stromal cells by secreting extrinsic factors can activate Wnt signaling, creating a microenvironment that supports the dedifferentiation of colon cancer cells to a CSC phenotype [64].

CSC have far more similarities than differences to normal SSC, greatly complicating the safe elimination of CSC, regardless of how they arose. SSC and CSC express similar "stemness" markers and both reside in specialized niches making elimination of this cell population extremely difficult without deleterious effects to normal SSC [65–67].

2.3. Stem Cell Modes of Division and Implications

Rarely dividing [68,69] during normal tissue homeostasis, long-term SSC spend the majority of their lifetime in a quiescent state. Once activated, quiescent SSC enter the cell cycle and undergo mitosis, generating two daughter cells. Two basic modes of stem cell division exist, i.e., symmetric or asymmetric (Figure 1). Asymmetric division, whereby one daughter cell remains in its niche, while the other daughter proceeds forward initiating a differentiation process to maintain tissue homeostasis is ideal (Figure 1, upper panel). However, SSC can also undergo two modes of symmetric divisions. Symmetric non-differentiative divisions generate two daughter stem cells that remain in their niche, whereas in symmetric differentiative divisions both cells leave their niche and differentiate, and lose their "stemness", (Figure 1, lower panel). Both modes of symmetric division are considered deleterious to the pool of long-lived SSC. Symmetric differentiative division leads to premature exhaustion of the stem cell pool [70], whereas symmetric non-differentiative division increases the number of DNA mutations accumulated in the SSC pool, a feature associated for example with Clonal Hematopoiesis of Indeterminate Potential (CHIP) [71].

Normal tissue homeostasis requires timely activation and asymmetric division of the SSC, however, with aging, this process is corrupted due to the accumulation of mutations in the SSC pool, chronic or acute injury, reversion of differentiated daughters to SSC, serum factors, changes in the niche microenvironment and SSC senescence [72]. The activation of quiescent SSC, and subsequent asymmetric versus symmetric division, is the most critical cellular decision in adult organisms underlying diseases of aging, including impaired wound healing, sarcopenia, fibrosis and osteoporosis, cancer and neurodegeneration. Stem

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cells (either normal SSC or CSC) undergoing mitosis read and must integrate an enormously complex array of information from their niche microenvironment to arrive at what in principle is a simple binary decision [29]. A fundamental intrinsic difference between CSC (and even pre-CSC for example in CHIP or myelodysplastic syndrome (MDS)) and normal SSC, is that CSC preferentially divide symmetrically rather than asymmetrically. Mutations in the gene p53 in breast cancer stem cells induce preferential symmetric cell division [73]. Premature exhaustion of normal hematopoietic stem cells (HSC) (due to increased symmetric differentiative divisions), with expansion of the leukemic stem cell (LSC) population due to increased symmetric non-differentiative divisions, is associated with loss of function of the tumor suppressor PTEN [74]. Genetic activation of Hedgehog signaling via indirect perturbation of Notch signaling, causes increased neural stem cell (NSC) symmetric divisions [75]. Preferential symmetric non- differentiative versus symmetric differentiative division is another intrinsic difference between CSC and normal SSC due to critical mutations (i.e., p53, p73, PTEN, etc.) or aberrant Wnt mediated mTOR activation [74,76]. However, in some instances, symmetric differentiative division may provide a mechanism to eliminate defective SSC [77].

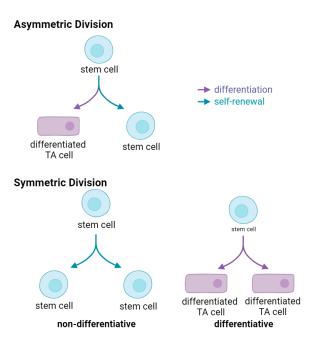


Figure 1. Modes of Stem Cell Division. Upper, an asymmetric division results in the production of two daughter cells with different fates—one remains a stem cell, the other becomes a differentiated transient amplifying (TA) cell. Lower left, symmetric non-differentiative division generates to two daughter stem cells. Lower right, symmetric differentiative division produces two differentiated transient amplifying daughter cells.

2.4. Stem Cell Heterogeneity

Somatic stem cells are heterogeneous and exist minimally in two distinct states. When activated deeply quiescent SSC in G_0 transition into a so-called G_{alert} phase before becoming fully activated and entering the cell cycle [78,79]. Although it is widely accepted that quiescent stem cells are arrested in G_0 , quiescent neural stem cells (NSC), at least in vitro, are found arrested in either G_0 or G_2 , where G_2 quiescent NSC reactivate prior to the activation of G_0 NSC [80]. After injury, organismal survival often requires rapid tissue repair. Under these conditions strict stem cell hierarchy is often compromised with the occurrence of cellular plasticity, fate conversion, lineage infidelity and "dedifferentiation" and reacquisition of "stem cell" characteristics [30,81]. For example, minimally two distinct intestinal stem cell (ISC) populations: Lgr5+ columnar basal cells (CBC) and a deeply

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quiescent, asymmetrically dividing, less radiosensitive, +4 Bmi1⁺ ISC, exist [82–85]. Due to their mode of chromosomal segregation [86], asymmetric cell division, deeply quiescent status and enhanced radio resistance, +4 Bmi1⁺ ISC appear to behave more like bona fide long-lived SSC than Lgr5⁺ CBC [84]. However, these populations are inter-convertible, with both possessing the capacity to generate all intestinal epithelial lineages [87,88]. Additionally, ISC that have a transcriptional profile similar to fetal intestinal stem cells that are important in intestinal regeneration after injury, termed "revival CSC", have been described. They express high levels of *Anxa1*, *Clu* and *Sca1* and are characterized by high YAP signaling and increased TNF- α , TGF- β , INF- γ and NF- κ B signaling [71,89–91]. Another recent report demonstrated that after chemical injury, reprogramming of Lgr5⁻ but Lgr4⁺ differentiated colonic epithelial cells, via Rspo3 induced Wnt signaling, is critical for epithelial regeneration [92]. In other stem cell populations including the lung epithelium and mammary gland [30,61] and in the hematopoietic system [93], similar situations exist. In hair follicles, both bulge cells (BC) and neighboring hair germ (HG) possess "stemness" features, BC and HG cells can both regenerate the seven distinct lineages of the hair follicle. Bulge cells normally generate HG cells under homeostatic conditions, however, after laserablation HG can replenish depleted bulge cells, [94]. Multiple pathways exist whereby cellular plasticity can induce a "stem-like" state from partially committed or differentiated cells, which has important implications for wound repair, chronic inflammation, tumorigenesis and aging [10,11,58,81]. Wound healing and inflammation play critical roles in cellular plasticity. Experiments in 1990 first demonstrated, that targeting oncogenic H-ras to murine differentiated epidermis generated papillomas preferentially at sites of irritation and wounding [95]. Introduction of a single oncogenic mutation, without inflammatory insult, only initiate intestinal tumor formation within one of the putative ISC populations (i.e., Lgr5/prominin/Bmi1 positive populations) [84,96,97], and targeting transient amplifying (TA) cells either had no effect or generated only microadenomas [96]. Cellular plasticity, associated with wounding and chronic inflammation, provides a mechanism whereby more differentiated progeny, with increased levels of DNA damage, can revert to a "stem-like" state. In the colon, the NF-kappa B (NF-kB) inflammatory pathway is dominant. Epidemiological studies have demonstrated that patients with chronic inflammation are predisposed to cancer. Non-steroidal anti-inflammatory drug administration decreases the incidence of colorectal cancer [98,99]. Immortalization of differentiated human cells by DNA viruses, including Epstein-Barr virus (EBV), Kaposi sarcoma-associated herpesvirus (KSHV), human papillomaviruses (HPV), hepatitis B virus (HBV), and more recently Merkel cell polyomavirus (MCPyV) and RNA viruses such as Hepatitis C Virus (HCV) and human T lymphotropic virus (HTLV-1) has been demonstrated [100]. Persistent chronic infection can lead to oncogenesis and induced dedifferentiation thereby generating cells with CSC characteristics. "Corruption" of the stem cell pool via chronic inflammation and viral infection significantly increases the risk of cancer.

Epigenetic landscape alterations in tissues occur after injury licensing normally restricted cell fate transitions. DNA methylation and histone modifications play critical roles in regulating this plasticity [11]. Rewiring of the epigenetic landscape and environmental perturbation after injury, allows for pathways that normally do not exist in embryonic or steady state adult tissues. Super-enhancers (SE) control the identity, lineage commitment and plasticity of adult SSC. SE represent a small fraction of total enhancers, which contain dense clusters ('epicenters') of transcription factor (TF) binding sites [12]. Understanding the biology of therapy resistance is critical to attain complete cancer cures. CSC, regardless of their cell of origin, represent the major cause of therapy resistance, metastatic disease and relapse. As discussed above, the many shared features of CSC and SSC, their heterogeneity

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(i.e., quiescent versus activated), and plasticity (i.e., EMT, lineage infidelity), complicates the safe elimination of CSC [101].

A variety of nutrient sources, akin to dining at a Smorgasbord table, can be sampled by stem cells (germ line and somatic) and CSC to suit their "behaviors" and "life-styles" and match their metabolic requirements [102]. The tumor stromal microenvironment, including endothelial cells, fibroblasts and adipocytes, serves up an assortment of "dishes" including glucose, lactate, ketone bodies, fatty acids and glutamine, to heterogeneous CSC [103–105]. Quiescence, low metabolic activity, and a protective niche microenvironment better safeguard stem cells, both SSC and CSC, against the accumulation of DNA damage compared to more differentiated activated cells [106]. Regulation of metabolic balance and the synthesis of antioxidants maintains intracellular redox homeostasis in conjunction with CSC-specific metabolism. In long-lived stem cells, reactive oxygen species (ROS)-induced damage to nuclear and/or mitochondrial DNA leads to mutations and therefore must be avoided. Interestingly, low levels of ROS are maintained in quiescent stem cells, despite primarily utilizing mitochondrial fatty acid oxidation (FAO) and oxidative phosphorylation (OXPHOS) [102]. The primary energetic contributor to the maintenance of deeply quiescent SSC and CSC is FAO [107-110]. How do quiescent stem cells (normal or cancerous) avoid the generation of ROS, which is deleterious to genomic integrity and also maintenance of quiescence while utilizing FAO? Primordial oocytes, a long-lived, in humans more than 40 years, quiescent stem cells that primarily utilize FAO, maintain functionality and genomic integrity to produce healthy offspring. They maintain low mitochondrial activity with a minimum of ROS generation. They accomplish this by silencing of the expression of mitochondrial complex I (MC1). MC1 is responsible for the oxidation of NADH to NAD+ with the release and leakage of electrons, which when received by oxygen generate ROS. They bypass the need for MC1 by utilizing mitochondrial complex II (MC2). MC2 catalyzes the oxidation of FADH2 to FAD2⁺ thereby serving as an entry point for electrons [111]. This process, albeit less efficiently, supports ATP production without generating large amounts of ROS [112]. Interestingly, we recently reported that the most deeply quiescent subset of a therapy resistant population of CD34⁻CD38⁻ chronic myeloid leukemic stem cells (LSC) [113], utilize a similar approach to primordial oocytes to maintain quiescence [108]. These LSC suppress the expression of MC1 complex genes, providing a mechanism to rely primarily on FAO while minimizing ROS. Pharmacologic differentiation of these quiescent LSC with ICG-001, a small molecule CBP/β-catenin antagonist, initiates the expression of MC1 genes and exit from quiescence. More generally, we believe that long lived quiescent stem cells rely on mitochondrial FAO, without complex I expression, thereby mitigating vulnerability to ROS. This allows deeply quiescent CSC, which are highly resistant to cancer chemo- and immunotherapy and a reservoir for disease recurrence and relapse, to persist for many years [114,115].

2.5. Wnt Signaling in Stem Cells; Low and High

The Wnt pathway is critical for the specification and maintenance of SSC in multiple tissues and organs, including the intestines, heart, blood, brain, and mammary gland [116]. Therefore, not surprisingly, a recurrent theme in cancer biology is the involvement of aberrant Wnt signaling [117,118]. In partnership with other key development pathways, Wnt regulation of SSC is important throughout development and organogenesis [119,120]. However, whether Wnt signaling is critical for self-renewal and maintenance of potency or stem cell differentiation and lineage commitment is debated [57]. The maintenance of ES cell pluripotency and the expansion of neural stem/progenitors is Wnt/ β -catenin regulated [121,122]. However, ES cell differentiation [123] and lineage determination in neural crest stem cells is also controlled by Wnt/ β -catenin signaling [124,125]. However,

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the dichotomous roles of Wnt/ β -catenin signaling in stem cell biology is not adequately explained by the 'Goldilocks' concept of a "just right" level of nuclear Wnt/ β -catenin transcription [126–129]. Similarly in cancer, a poorer prognosis is associated with increased Wnt/ β -catenin signaling in colorectal tumors [130–132], however, increased nuclear β -catenin in tumors is associated with a more favorable melanoma prognosis [133–136]. However, a recent report outlined how a TCF4-dependent gene regulatory network conferred immunotherapy resistance in melanoma [137], consistent with previous reports of enhanced Wnt signaling in immunotherapy resistance [16,138].

2.6. Wnt Signaling and Metabolism

In stem cells, cellular and tissue differentiation and in immune cell biology, integration of cellular metabolism and cellular function is critical and highly evolutionarily conserved. In addition to playing a significant role in stem/progenitor cell biology [9,29,139], the Wnt signaling cascade regulates glucose metabolism, de novo lipogenesis and mitochondrial physiology [140,141], thereby providing a mechanism to couple metabolism with cellular status, i.e., quiescence, activation and differentiation [142]. The nuclear receptor (NR) family also plays essential regulatory roles in stem cell maintenance, differentiation and lineage commitment and metabolism [143]. Fatty acid oxidation (FAO), gluconeogenesis, lipogenesis and glycolysis are regulated by NR family members [144]. The nuclear receptors PPAR α and ESRRA, in proximal tubule kidney cells, coordinate metabolism with differentiation. Their dysfunction is associated with kidney fibrosis [145]. Wnt signaling is also important in kidney cell differentiation [146], and aberrant Wnt activation and blocked differentiation with renal fibrosis [147]. Interferon activation of the signal transducer and activator of transcription 1 (STAT1)/Janus kinase (JAK) pathway, regulates immunometabolism and immune cell polarization [148]. Interferon also plays dichotomous roles in that it possesses antifibrotic effects via differentiation [149], yet also contributes to renal fibrosis and chronic kidney disease [150]. Wnt/β-catenin crosstalk and multiple signaling cascades converge at the amino-termini of the Kat3 coactivators to regulate metabolism and differentiation to maintain homeostasis or to resolve inflammatory processes after injury. Aberrant coordination is associated with cancer, fibrosis and neurodegeneration [22].

2.7. Differential Kat3 Coactivator Usage

β-catenin must recruit either cAMP response element-binding protein (CREBbinding protein (CREBPP or CBP) or its closely related homolog, p300 (E1A-binding protein, 300 KDa), in addition to the basal transcriptional apparatus to activate gene transcription [151–153]. Just prior to the vertebrate radiation, gene duplication gave rise to CBP and p300, very large proteins, both over 300 kD, encoded over 33 exons and 31 exons respectively [154]. Despite diverging over 450 million years ago, they retain an extremely high degree of identity, up to 93%, particularly over a large central core that includes the CH1, KIX, Bromodomain, and CH2 and CH3 regions (Figure 2) [155,156]. CBP and p300 are master orchestrators of transcription interacting with hundreds of proteins in this role. Perhaps not surprisingly, they are often considered redundant and interchangeable due to their high degree of protein sequence identity and even higher similarity. However, the retention of more than 150 Kb of redundant DNA for over 450 million years, is not something 'Mother Nature' would do without a very good reason! and numerous studies have clearly shown that CBP and p300 have definitive and unique roles both in vitro and in vivo [29,157–160]. Furthermore, the divergent roles of CBP and p300 are critical for the maintenance of fidelity in long-lived SSC (CBP) and the timely activation and proliferation of differentiated TA cells (p300) [29].

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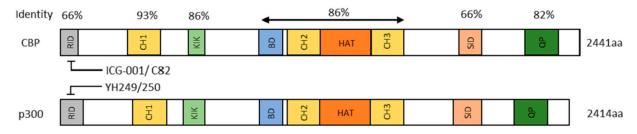


Figure 2. CBP and p300. An extremely high percentage of identity and even higher homology. between various regions of these large Kat3 coactivators at the amino acid level, despite their divergence over 450 million years ago. The CBP and p300 amino termini, the regions to which CBP/ β -catenin (ICG-001 and C82 the active agent derived from PRI-724 dephosphorylation) and the direct p300/ β -catenin antagonists (YH249, 250) bind respectively, are the most divergent regions.

Over 25 years ago, utilizing a cell-based TopFlash Wnt reporter assay in SW480 cells, with a library of 5000 secondary structure mimetics, my lab originally identified ICG-001 (IC $_{50}$ ~3 μ M) (Figure 3) [161]. We identified and validated that the molecular target of ICG-001 was the Kat3 coactivator CBP. With high affinity (~1 nM in vitro), ICG-001 binds the extreme N-terminus of CBP specifically, thereby directly inhibiting the CBP/ β -catenin interaction. Critically important, ICG-001 does not bind to p300 [161,162]. We later identified the structurally related small molecule p300 direct binders, YH249 and YH250, which block p300/ β -catenin driven transcription (Figure 3) [163].

Figure 3. Structures of ICG-001, PRI-724 and YH-249/250.

The extreme N-termini of CBP and p300 are specifically and directly bound by ICG-001 and YH249/250 respectively. These regions of CBP and p300, with only 66% identity between them, are the least conserved regions (Figure 2). It appears that after initial evolutionary divergence of these N-terminal regions, within the individual N-terminal regions of both CBP and p300, >98% identity has been retained at the amino acid level for more than 100 million years! This confirms the crucial roles that these evolutionarily conserved regions have in vertebrate biology.

Selectively blocking the CBP/ β -catenin interaction leads to enhanced p300/ β -catenin transcription, resulting in the initiation of differentiation in a wide variety of stem/progenitor cells, including CSC [113,164–167]. Directly or indirectly blocking the p300/ β -catenin interaction enhances CBP/ β -catenin transcription in both mouse and human ES, iPS, and SSC, both in vitro and in vivo, which is critical for stem cell self-renewal [163,164,168–170]. These studies led to the development and validation of our model, in which CBP/ β -catenin maintains stemness/enhances and self-renewal and

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p300/ β -catenin transcription initiates a differentiative program, providing distinct roles for CBP/ and p300/ β -catenin-mediated transcription in stem/progenitor cell biology (Figure 4) [29,167,168].

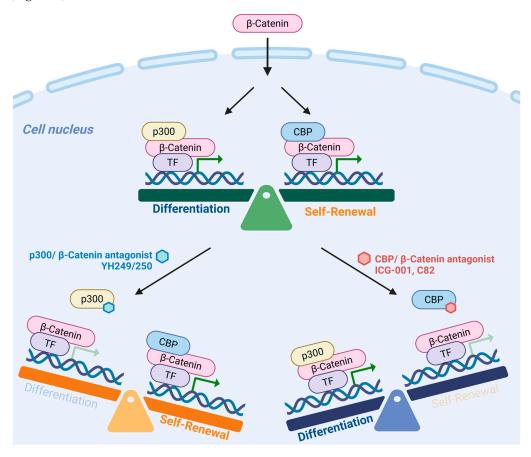


Figure 4. CBP/β-catenin antagonists ICG-001/C82 selectively block the CBP/β-catenin interaction leading to enhanced p300 usage thereby initiating differentiation. YH-249/250 selectively block the p300/β-catenin interaction, thereby enhancing CBP usage, resulting in maintenance of a program supporting self-renewal.

2.8. CBP/β-Catenin Antagonists Are Safe and Efficacious

CBP/ β -catenin antagonists have been extensively investigated in a variety of preclinical tumor models. CBP/ β -catenin antagonists safely eliminate quiescent drug-resistant CSC, via forced stochastic symmetric differentiation, without deleterious effects to normal endogenous SSC [77,113,171–174]. CBP/ β -catenin antagonists have proven efficacious in a wide array of disease and injury models; including pulmonary, renal, hepatic and systemic fibrosis [175–178], myocardial infarction [179], neuro-development and neuro-degeneration [77,167]. The beneficial effects of CBP/ β -catenin antagonists in these preclinical models, are associated with enhanced activation and subsequent asymmetric differentiation of SSC, thereby correcting lineage infidelity and initiating proper repair and healing [29,129,180].

Based on these studies, PRI-724 (IC50 ~150 nM) (Figure 3), a second-generation CBP/ β -catenin specific antagonist was developed and proved extremely safe both pre-clinically and clinically. The non-adverse event level in IND enabling toxicology studies in dogs, given 28-day continuous infusion of PRI-724, was 120 mg/kg/day. Plasma concentrations roughly 300 times the IC50 were maintained for 28 days. In a first-in-human clinical trial in cancer patients, PRI-724 was administered by 7 days of continuous i.v. infusion, with dose escalation from 40 to 1280 mg/m²/day. No dose limiting toxicities were observed. Reduction of the biomarker *survivin/BIRC5* with upregulation of the differentiation antigen

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CK20 in circulating tumor cells (CTC), strongly correlated with plasma concentrations of drug (R = 0.97) [181], demonstrating on target efficacy. Likely due to the critical role of the highly conserved CBP N-terminus in stem cell maintenance, resistance to the PRI-724 was not observed [172]. PRI-724, in HCV-induced cirrhosis patients, for which no currently approved treatment exists, demonstrated encouraging anti-fibrotic activity, [182]. However, the lack of oral bioavailability of PRI-724 hampered its development. Eisai developed a next generation orally available analog E7386, that is purportedly a CBP/beta-catenin antagonist, however, comparative analysis of E7386 with the highly specific bona fide CBP/β-catenin antagonists, ICG-001 and C82 (the active agent derived from PRI-724) cast significant doubt that the mechanism of action of E7386 is via specific CBP/β-catenin antagonism [183].

Taking advantage of an intrinsic and critical difference between CSC and SSC, CBP/ β -catenin antagonists enforce either symmetric (in CSC) or asymmetric (in SSC) differentiative cell division, providing efficacy and safety. Due to various mutations (e.g., p53, PTEN, etc.), CSC preferentially divide symmetrically whereas SSC divide asymmetrically [29,73,74]. CBP/ β -catenin antagonists can thereby stochastically eliminate CSC via forced symmetric differentiation, whereas SSC in their niche divide asymmetrically and are maintained (Figure 5).

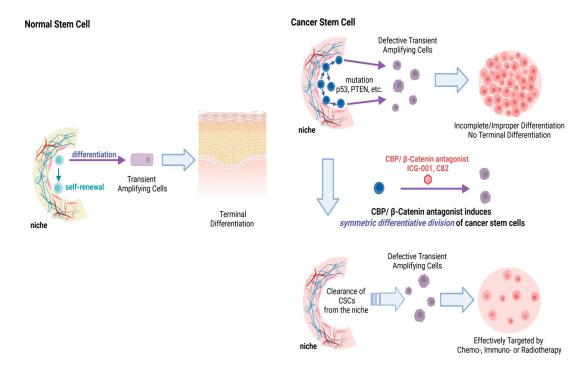


Figure 5. Upper panel Normal somatic stem cells (SSC) undergo asymmetric divisions—with the original stem cell remaining in the niche and the differentiated daughter TA cell going on to eventually become part of a fully differentiated tissue or organ system. Normal SSC are encouraged to undergo asymmetric divisions upon CBP/ β -catenin antagonism and thus are not depleted from their niche. **Middle panel** Cancer stem cells (CSC) have a preference for symmetric divisions, thus leading to an increase in CSC in the niche over time with increased mutational burden. **Lower panel** CBP/ β -catenin antagonists (ICG-001 or C82) induce symmetric differentiative divisions of the CSC population thereby stochastically clearing CSC from their niche.

2.9. Pleiotropic Effects of CBP/β-Catenin Antagonists

ICG-001 was initially identified as a TCF/ β -catenin antagonist that modulated Wnt signaling by selectively blocking the recruitment of the Kat3 coactivator CBP. However, after 25 years of research, it is now clear that effects of CBP/ β -catenin antagonists involve

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far more than classical Wnt target genes and are highly pleiotropic. β-catenin, independent of TCFs, recruits CBP and p300 to enhancers (E) and super-enhancers (SE). Super-enhancers constitute a subset of enhancers that regulate genes controlling cellular identity and lineage fidelity [184–188]. Acetylated H3K27 (H3K27Ac), which is uniquely acetylated by Kat3 coactivators, demarcate E and SE [184,189–193]. The N and C-terminal intrinsically disordered regions (IDR) of β-catenin, independent of TCF/LEF interactions, can recruit either CBP or p300 into E/SE-driven complexes [194]. Already more than 20 years ago, differential Kat3 recruitment, with a predominant bias toward p300-mediated regulation to licensed enhancers, was noted [195]. CBP/p300 acetylation at enhancers simultaneously promote transcription initiation and elongation via pre-initiation complex (PIC) formation and RNAPII recruitment, independent of their roles in BRD4-dependent pause release [196]. The formation of the mouse cardiovascular system, lung and the small intestine are strongly impaired by p300 HAT mutation but significantly less by similar mutation of CBP, despite their highly conserved HAT domains [197]. This confirms the differential roles of CBP and p300 at critical E/SE during development and the critical role of p300 HAT activity during organogenesis. Experiments on human myoblast differentiation [157], cellular senescence [198], immune cell function [199] and in mouse embryonic fibroblasts at the single-cell genome-wide level [200], further attest to differential roles for Kat3 coactivators at E and SE.

SE in stem cells, provide for rapid fate switching when the niche local environment is modified [201], playing critical roles in development and disease [186,202]. Terminal transcription factors of the Wnt (i.e., TCF/LEF), TGF- β (Smad3), and LIF (Stat3) pathways often occupy SE and transcriptionally control stem cell states and can promote tumorigenesis [203,204]. Small molecule CBP/ β -catenin antagonists by targeting 'Wnt/ β -catenin'-driven oncogenic 'stemness' and correcting lineage infidelity via SE modulation, provide an opportunity to target drug resistant CSC [205].

The CBP/ β -catenin antagonist ICG-001, in diffuse intrinsic pontine gliomas, down-regulates genes involved in stemness maintenance (i.e., ID1 and ID3) by targeting SE, yet increase the expression of invasion promoting genes, via p300 recruitment, which can be effectively targeted by JQ-1, a BET inhibitor [206].

CBP and p300 are seen as 'molecular interpreters that can parse and/or conjugate the regulatory words, phrases, and sentences of the genome' [207]. The amino terminal intrinsically disordered regions (IDR) of both Kat3 coactivators allows for promiscuous interaction with hundreds of cellular transcription factors, via their own IDRs to 'interpret' the gene encoded regulatory language. Within distinct tissue environments, differential activation of a common enhancer repertoire and the expression of divergent secondary transcription factors (TFs) that collaborate with core TFs, can establish tissue-specific enhancers [208]. β -catenin IDRs, via SE occupancy and interactions with both TFs and the N-termini of either Kat3 coactivator, assists in interpreting signal transduction cascades and extracellular information required for cell-specific responses [209–213]. In addition to canonical TCF/LEF family members [214–219], β -catenin interacts with a large number of TFs with assemblage of tissue-specific SEs fostered by the recruitment of either CBP or p300 [205] (Figure 6).

Treatment of pancreatic cancer cells with ICG-001 demonstrated wide ranging effects on super-enhancers and chromatin architecture over particularly broad epigenomic domains based on Hi-C analyses [220,221]. Insulin signaling genes were enriched in the altered chromatin structure and insulin signaling chromatin loops were significantly weakened by ICG-001, with strongly diminished IRS1 looping, in these cells [222].

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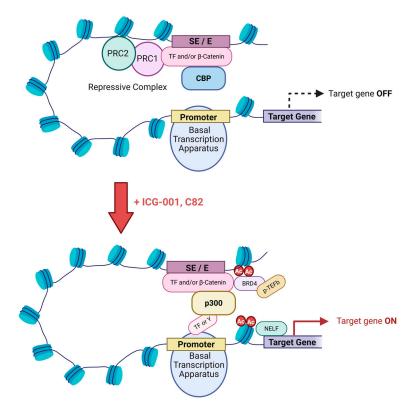


Figure 6. Intrinsically disordered regions (IDRs) of β-catenin connect transcription factor (TF)-interacting domains with the N-termini of either CBP or p300 in enhancer (E) and super-enhancer (SE) loci to interpret extracellular information and complex signaling cascades to orchestrate cell specific responses. The model depicts a small molecule specific CBP/β-catenin antagonist (ICG-001 or C82) dismissing CBP occupancy from a repressive complex with recruitment of p300 enhancing the assembly of tissue specific E/SE with pleiotropic effects on differentiation, lineage identity and fidelity, coupled to cellular metabolism.

3. Conclusions

Blocking recruitment of the N-terminal of CBP to various E and SE, with or without a corresponding increase in p300 recruitment, and the de novo formation of many new E/SE, broadly affects stem cell differentiation, lineage commitment, hypoxia, immune cell function, metabolism, cellular senescence, etc., leading to the wide array of pleiotropic effects observed [22,29,223,224]. Additionally, selective disruption of CBP recruitment via its very N-terminal domain with small molecule CBP/β-catenin antagonists, frees up the limited amount per cell of CBP for recruitment to new E/SE, via alternative domains (e.g., KIX domain), to drive differentiative programs [225,226] (Figure 6). Wnt signaling cascade intrinsic complexity and further crosstalk with multiple pathways, clearly represents a major obstacle to safe therapeutic targeting [22,29]. Specific small molecule CBP/β-catenin antagonists can safely eliminate CSC via forced symmetric differentiation, thereby sensitizing resistant tumors to conventional or immunotherapy. Preclinically CBP/β-catenin antagonists by targeting CSC, dramatically sensitized tumors to chemotherapy and prevented secondary tumor engraftment, however, they did not affect tumor growth [171]. Therefore, unsurprisingly, in the PRI-724 Phase I clinical study, no objective RECIST criteria responses were seen, despite on-target effects in circulating tumor cells, a surrogate "stem cell" population [181]. Furthermore, PRI-724's excellent safety profile and promising on target clinical effects both in oncology and liver fibrosis bode well for specific CBP/β-catenin antagonist development, to maintain or regain lineage fidelity, commonly defective in diseases of aging including cancer, fibrosis and neurodegeneration. Intriguingly, *survivin/Birc5* expression has been shown to be critical for both hES cell generation of teratomas [227] and oncogene

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targeted stem cell initiation of basal cell carcinoma [228]. The excellent safety profile of CBP/beta-catenin antagonists coupled with their capacity to down-regulate *survivin/Birc5* expression auger well for the prospects of utilizing CBP/beta-catenin antagonists to prevent malignancies.

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Abbreviations

ROS

CREBBP/CBP	CREB Binding Protein
SSC	Somatic stem cell
CSC	Cancer stem cell
LSC	Leukemic stem cell
CTC	Circulating Tumor Cell
E/SE	Enhancer/Super-Enhancer
TME	Tumor Microenvironment
CHIP	Clonal Hematopoiesis of Indeterminate Potential
FAO	Fatty Acid Oxidation
OXPHOS	Oxidative Phosphorylation
MC1	Mitochondrial Complex 1

References

1. Virchow, R. Die Cellularpathologie in Ihrer Begrundung Auf Physiologische und Pathologische Gewebelehre; Verlag von August Hirschwald: Berlin, Deutschland, 1861.

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- 2. Arwert, E.N.; Hoste, E.; Watt, F.M. Epithelial stem cells, wound healing and cancer. *Nat. Rev. Cancer* **2012**, *12*, 170–180. [CrossRef] [PubMed]
- 3. Dvorak, H.F. Tumors: Wounds that do not heal. Similarities between tumor stroma generation and wound healing. *N. Engl. J. Med.* **1986**, 315, 1650–1659. [PubMed]
- 4. Lee, S.; Haddow, A. Extraosseous extension of vertebral hemangioma, a rare cause of spinal cord compression. *Spine* **1999**, 24, 2111–2114. [CrossRef] [PubMed]
- 5. Dunham, L.J. Cancer in man at site of prior benign lesion of skin or mucous membrane: A review. Cancer Res. 1972, 32, 1359–1374.
- 6. Schober, M.; Fuchs, E. Tumor-initiating stem cells of squamous cell carcinomas and their control by TGF-β and integrin/focal adhesion kinase (FAK) signaling. *Proc. Natl. Acad. Sci. USA* **2011**, *108*, 10544–10549. [CrossRef]
- 7. Kahn, M. Chapter Seven—Wnt Signaling in Stem Cells and Cancer Stem Cells: A Tale of Two Coactivators. In *Progress in Molecular Biology and Translational Science*; Larraín, J., Olivares, G., Eds.; Elsevier: Amsterdam, The Netherlands, 2018; Volume 153, pp. 209–244.
- 8. Holland, J.D.; Klaus, A.; Garratt, A.N.; Birchmeier, W. Wnt signaling in stem and cancer stem cells. *Curr. Opin. Cell Biol.* **2013**, 25, 254–264. [CrossRef]
- 9. Reya, T.; Clevers, H. Wnt signalling in stem cells and cancer. Nature 2005, 434, 843–850. [CrossRef]
- 10. Ge, Y.; Gomez, N.C.; Adam, R.C.; Nikolova, M.; Yang, H.; Verma, A.; Lu, C.P.-J.; Polak, L.; Yuan, S.; Elemento, O.; et al. Stem Cell Lineage Infidelity Drives Wound Repair and Cancer. *Cell* 2017, 169, 636–650.e14. [CrossRef]
- 11. Paksa, A.; Rajagopal, J. The epigenetic basis of cellular plasticity. Curr. Opin. Cell Biol. 2017, 49, 116–122. [CrossRef]

Cancers 2025, 17, 1503 14 of 22

12. Adam, R.C.; Yang, H.; Rockowitz, S.; Larsen, S.B.; Nikolova, M.; Oristian, D.S.; Polak, L.; Kadaja, M.; Asare, A.; Zheng, D.; et al. Pioneer factors govern super-enhancer dynamics in stem cell plasticity and lineage choice. *Nature* **2015**, *521*, 366–370. [CrossRef]

- 13. Miranda, A.; Hamilton, P.T.; Zhang, A.W.; Pattnaik, S.; Becht, E.; Mezheyeuski, A.; Bruun, J.; Micke, P.; de Reynies, A.; Nelson, B.H. Cancer stemness, intratumoral heterogeneity, and immune response across cancers. *Proc. Natl. Acad. Sci. USA* **2019**, *116*, 9020–9029. [CrossRef] [PubMed]
- 14. Malta, T.M.; Sokolov, A.; Gentles, A.J.; Burzykowski, T.; Poisson, L.; Weinstein, J.N.; Kamińska, B.; Huelsken, J.; Omberg, L.; Gevaert, O.; et al. Machine Learning Identifies Stemness Features Associated with Oncogenic Dedifferentiation. *Cell* **2018**, 173, 338–354.e15. [CrossRef] [PubMed]
- 15. Luke, J.J.; Bao, R.; Sweis, R.F.; Spranger, S.; Gajewski, T.F. WNT/β-catenin Pathway Activation Correlates with Immune Exclusion across Human Cancers. *Clin. Cancer Res.* **2019**, 25, 3074–3083. [CrossRef] [PubMed]
- 16. Spranger, S.; Bao, R.; Gajewski, T.F. Melanoma-intrinsic β-catenin signalling prevents anti-tumour immunity. *Nature* **2015**, *523*, 231–235. [CrossRef]
- 17. Langille, E.R.; Al-Zahrani, K.N.; Ma, Z.; Malik, A.; Loganathan, S.K.; Trcka, D.; Liu, J.; Kozma, K.; Tsai, R.; Teng, K.; et al. Loss of epigenetic regulation disrupts lineage integrity, reactivates multipotency and promotes breast cancer. *bioRxiv* 2021. [CrossRef]
- 18. Perekatt, A.O.; Shah, P.P.; Cheung, S.; Jariwala, N.; Wu, A.; Gandhi, V.; Kumar, N.; Feng, Q.; Patel, N.; Chen, L.; et al. SMAD4 Suppresses WNT-Driven Dedifferentiation and Oncogenesis in the Differentiated Gut Epithe-lium. *Cancer Res.* **2018**, *78*, 4878–4890. [CrossRef]
- 19. Ordóñez-Morán, P.; Dafflon, C.; Imajo, M.; Nishida, E.; Huelsken, J. HOXA5 Counteracts Stem Cell Traits by Inhibiting Wnt Signaling in Colorectal Cancer. *Cancer Cell* **2015**, *28*, 815–829. [CrossRef]
- 20. Jeon, H.Y.; Choi, J.; Kraaier, L.; Kim, Y.H.; Eisenbarth, D.; Yi, K.; Kang, J.; Kim, J.W.; Shim, H.S.; Lee, J.; et al. Airway secretory cell fate conversion via YAP-mTORC1-dependent essential amino acid metabolism. *EMBO J.* **2022**, *41*, e109365. [CrossRef]
- 21. Xiong, X.; James, B.T.; Boix, C.A.; Park, Y.P.; Galani, K.; Victor, M.B.; Sun, N.; Hou, L.; Ho, L.-L.; Mantero, J.; et al. Epigenomic dissection of Alzheimer's disease pinpoints causal variants and reveals epigenome erosion. *Cell* **2023**, *186*, 4422–4437.e21. [CrossRef]
- 22. Kahn, M. Taking the road less traveled—The therapeutic potential of CBP/β-catenin antagonists. *Expert Opin. Ther. Targets* **2021**, 25, 701–719. [CrossRef]
- 23. Ogamino, S.; Yamamichi, M.; Sato, K.; Ishitani, T. Dynamics of Wnt/β-catenin reporter activity throughout whole life in a naturally short-lived vertebrate. *NPJ Aging* **2024**, *10*, 23. [CrossRef] [PubMed]
- 24. Khalid, K.A.; Nawi, A.F.M.; Zulkifli, N.; Barkat, A.; Hadi, H. Aging and Wound Healing of the Skin: A Review of Clinical and Pathophysiological Hallmarks. *Life* 2022, 12, 2142. [CrossRef] [PubMed]
- 25. Nalapareddy, K.; Nattamai, K.J.; Kumar, R.S.; Karns, R.; Wikenheiser-Brokamp, K.A.; Sampson, L.L.; Mahe, M.M.; Sundaram, N.; Yacyshyn, M.-B.; Yacyshyn, B.; et al. Canonical Wnt Signaling Ameliorates Aging of Intestinal Stem Cells. *Cell Rep.* **2017**, *18*, 2608–2621. [CrossRef] [PubMed]
- 26. Beerman, I.; Bhattacharya, D.; Zandi, S.; Sigvardsson, M.; Weissman, I.L.; Bryder, D.; Rossi, D.J. Functionally distinct hematopoietic stem cells modulate hematopoietic lineage potential during aging by a mechanism of clonal expansion. *Proc. Natl. Acad. Sci. USA* **2010**, *107*, 5465–5470. [CrossRef]
- 27. Cho, R.H.; Sieburg, H.B.; Muller-Sieburg, C.E. A new mechanism for the aging of hematopoietic stem cells: Aging changes the clonal composition of the stem cell compartment but not individual stem cells. *Blood* **2008**, *111*, 5553–5561. [CrossRef]
- 28. Sudo, K.; Ema, H.; Morita, Y.; Nakauchi, H. Age-Associated Characteristics of Murine Hematopoietic Stem Cells. *J. Exp. Med.* **2000**, 192, 1273–1280. [CrossRef]
- 29. Thomas, P.D.; Kahn, M. Kat3 coactivators in somatic stem cells and cancer stem cells: Biological roles, evolution, and pharmacologic manipulation. *Cell Biol. Toxicol.* **2016**, *32*, 61–81. [CrossRef]
- 30. Blanpain, C.; Fuchs, E. Plasticity of epithelial stem cells in tissue regeneration. Science 2014, 344, 1242281. [CrossRef]
- 31. Akaiwa, H. A Quantitative Study of Wound Healing in the Rat: I. Cell Movements and Cell Layers during Wound Healing. *J. Med. Res.* **1919**, *40*, 311–351.
- 32. Whyte, J.L.; Smith, A.A.; Helms, J.A. Wnt Signaling and Injury Repair. Cold Spring Harb. Perspect. Biol. 2012, 4, a008078. [CrossRef]
- 33. Azevedo, A.S.; Grotek, B.; Jacinto, A.; Weidinger, G.; Saúde, L. The Regenerative Capacity of the Zebrafish Caudal Fin Is Not Affected by Repeated Amputations. *PLoS ONE* **2011**, *6*, e22820. [CrossRef] [PubMed]
- 34. Kragl, M.; Knapp, D.; Nacu, E.; Khattak, S.; Maden, M.; Epperlein, H.H.; Tanaka, E.M. Cells keep a memory of their tissue origin during axolotl limb regeneration. *Nature* **2009**, *460*, 60–65. [CrossRef] [PubMed]
- 35. Straube, W.L.; Brockes, J.P.; Drechsel, D.N.; Tanaka, E.M. Plasticity and Reprogramming of Differentiated Cells in Amphibian Regeneration: Partial Purification of a Serum Factor that Triggers Cell Cycle Re-Entry in Differentiated Muscle Cells. *Cloning Stem Cells* 2004, 6, 333–344. [CrossRef] [PubMed]
- 36. ter Steege, E.J.; Bakker, E.R.M. The role of R-spondin proteins in cancer biology. Oncogene 2021, 40, 6469–6478. [CrossRef]

Cancers 2025, 17, 1503 15 of 22

37. Chen, L.; Qiu, X.; Dupre, A.; Pellon-Cardenas, O.; Fan, X.; Xu, X.; Rout, P.; Walton, K.D.; Burclaff, J.; Zhang, R.; et al. TGFB1 induces fetal reprogramming and enhances intestinal regeneration. *Cell Stem Cell* **2023**, *30*, 1520–1537.e8. [CrossRef]

- 38. Shi, Y.; Shu, B.; Yang, R.; Xu, Y.; Xing, B.; Liu, J.; Chen, L.; Qi, S.; Liu, X.; Wang, P.; et al. Wnt and Notch signaling pathway involved in wound healing by targeting c-Myc and Hes1 separately. *Stem Cell Res. Ther.* **2015**, *6*, 120. [CrossRef]
- 39. Gregorieff, A.; Liu, Y.; Inanlou, M.R.; Khomchuk, Y.; Wrana, J.L. Yap-dependent reprogramming of Lgr5+ stem cells drives intestinal regeneration and cancer. *Nature* **2015**, *526*, 715–718. [CrossRef]
- 40. Kawakami, Y.; Esteban, C.R.; Raya, M.; Kawakami, H.; Martí, M.; Dubova, I.; Belmonte, J.C.I. Wnt/β-catenin signaling regulates vertebrate limb regeneration. *Genes Dev.* **2006**, *20*, 3232–3237. [CrossRef]
- 41. Ramachandran, R.; Zhao, X.-F.; Goldman, D. Ascl1a/Dkk/β-catenin signaling pathway is necessary and glycogen synthase kinase-3β inhibition is sufficient for zebrafish retina regeneration. *Proc. Natl. Acad. Sci. USA* **2011**, *108*, 15858–15863. [CrossRef]
- 42. Stephens, W.Z.; Senecal, M.; Nguyen, M.; Piotrowski, T. Loss of *adenomatous polyposis coli (apc)* results in an expanded ciliary marginal zone in the zebrafish eye. *Dev. Dyn.* **2010**, 239, 2066–2077. [CrossRef]
- 43. Cho, S.-H.; Cepko, C.L. Wnt2b/β-catenin-mediated canonical Wnt signaling determines the peripheral fates of the chick eye. *Development* **2006**, 133, 3167–3177. [CrossRef] [PubMed]
- 44. Kubo, F.; Takeichi, M.; Nakagawa, S. Wnt2b controls retinal cell differentiation at the ciliary marginal zone. *Development* **2003**, *130*, 587–598. [CrossRef] [PubMed]
- 45. Denayer, T.; Locker, M.; Borday, C.; Deroo, T.; Janssens, S.; Hecht, A.; van Roy, F.; Perron, M.; Vleminckx, K. Canonical Wnt Signaling Controls Proliferation of Retinal Stem/Progenitor Cells in Postembryonic *Xenopus* Eyes. *Stem Cells* **2008**, *26*, 2063–2074. [CrossRef] [PubMed]
- 46. Das, A.V.; Bhattacharya, S.; Zhao, X.; Hegde, G.; Mallya, K.; Eudy, J.D.; Ahmad, I. The Canonical Wnt Pathway Regulates Retinal Stem Cells/Progenitors in Concert with Notch Signaling. *Dev. Neurosci.* **2008**, *30*, 389–409. [CrossRef]
- 47. Liu, H.; Xu, S.; Wang, Y.; Mazerolle, C.; Thurig, S.; Coles, B.L.; Ren, J.-C.; Taketo, M.M.; van der Kooy, D.; Wallace, V.A. Ciliary margin transdifferentiation from neural retina is controlled by canonical Wnt signaling. *Dev. Biol.* **2007**, *308*, 54–67. [CrossRef]
- 48. Stoick-Cooper, C.L.; Weidinger, G.; Riehle, K.J.; Hubbert, C.; Major, M.B.; Fausto, N.; Moon, R.T. Distinct Wnt signaling pathways have opposing roles in appendage regeneration. *Development* **2007**, *134*, 479–489. [CrossRef]
- Qyang, Y.; Martin-Puig, S.; Chiravuri, M.; Chen, S.; Xu, H.; Bu, L.; Jiang, X.; Lin, L.; Granger, A.; Moretti, A.; et al. The Renewal and Differentiation of Isl1+ Cardiovascular Progenitors Are Controlled by a Wnt/β-Catenin Pathway. *Cell Stem Cell* 2007, 1, 165–179. [CrossRef]
- 50. Moparthi, L.; Koch, S. Wnt signaling in intestinal inflammation. Differentiation 2019, 108, 24–32. [CrossRef]
- 51. Nabhan, A.N.; Brownfield, D.G.; Harbury, P.B.; Krasnow, M.A.; Desai, T.J. Single-cell Wnt signaling niches maintain stemness of alveolar type 2 cells. *Science* **2018**, *359*, 1118–1123. [CrossRef]
- 52. Molofsky, A.V.; Slutsky, S.G.; Joseph, N.M.; He, S.; Pardal, R.; Krishnamurthy, J.; Sharpless, N.E.; Morrison, S.J. Increasing p16INK4a expression decreases forebrain progenitors and neurogenesis during ageing. *Nature* **2006**, *443*, 448–452. [CrossRef]
- 53. Conboy, I.M.; Conboy, M.J.; Wagers, A.J.; Girma, E.R.; Weissman, I.L.; Rando, T.A. Rejuvenation of aged progenitor cells by exposure to a young systemic environment. *Nature* **2005**, *433*, 760–764. [CrossRef] [PubMed]
- 54. Sharpless, N.E.; DePinho, R.A. Cancer biology: Gone but not forgotten. Nature 2007, 445, 606–607. [CrossRef] [PubMed]
- 55. Nishimura, E.K.; Granter, S.R.; Fisher, D.E. Mechanisms of Hair Graying: Incomplete Melanocyte Stem Cell Maintenance in the Niche. *Science* 2005, 307, 720–724. [CrossRef] [PubMed]
- 56. Chen, J.; Astle, C.M.; Harrison, D.E. Genetic regulation of primitive hematopoietic stem cell senescence. *Exp. Hematol.* **2000**, 28, 442–450. [CrossRef]
- 57. Reya, T.; Morrison, S.J.; Clarke, M.F.; Weissman, I.L. Stem cells, cancer, and cancer stem cells. Nature 2001, 414, 105–111. [CrossRef]
- 58. Gupta, P.B.; Pastushenko, I.; Skibinski, A.; Blanpain, C.; Kuperwasser, C. Phenotypic Plasticity: Driver of Cancer Initiation, Progression, and Therapy Resistance. *Cell Stem Cell* **2019**, 24, 65–78. [CrossRef]
- 59. Friedmann-Morvinski, D.; Verma, I.M. Dedifferentiation and reprogramming: Origins of cancer stem cells. *EMBO Rep.* **2014**, *15*, 244–253. [CrossRef]
- 60. Schwitalla, S.; Fingerle, A.A.; Cammareri, P.; Nebelsiek, T.; Göktuna, S.I.; Ziegler, P.K.; Canli, O.; Heijmans, J.; Huels, D.J.; Moreaux, G.; et al. Intestinal Tumorigenesis Initiated by Dedifferentiation and Acquisition of Stem-Cell-like Properties. *Cell* 2013, 152, 25–38. [CrossRef]
- 61. Liu, C.; Xu, Y.; Yang, G.; Tao, Y.; Chang, J.; Wang, S.; Cheung, T.H.; Chen, J.; Zeng, Y.A. Niche inflammatory signals control oscillating mammary regeneration and protect stem cells from cytotoxic stress. *Cell Stem Cell* **2023**, *31*, 89–105.e6. [CrossRef]
- 62. Mani, S.A.; Guo, W.; Liao, M.-J.; Eaton, E.N.; Ayyanan, A.; Zhou, A.Y.; Brooks, M.; Reinhard, F.; Zhang, C.C.; Shipitsin, M.; et al. The Epithelial-Mesenchymal Transition Generates Cells with Properties of Stem Cells. *Cell* 2008, 133, 704–715. [CrossRef]
- 63. Chaffer, C.L.; Marjanovic, N.D.; Lee, T.; Bell, G.; Kleer, C.G.; Reinhardt, F.; D'Alessio, A.C.; Young, R.A.; Weinberg, R.A. Poised Chromatin at the ZEB1 Promoter Enables Breast Cancer Cell Plasticity and Enhances Tumorigenicity. *Cell* **2013**, *154*, 61–74. [CrossRef] [PubMed]

Cancers 2025, 17, 1503 16 of 22

64. Vermeulen, L.; Felipe De Sousa, E.M.; Van Der Heijden, M.; Cameron, K.; De Jong, J.H.; Borovski, T.; Tuynman, J.B.; Todaro, M.; Merz, C.; Rodermond, H.; et al. Wnt activity defines colon cancer stem cells and is regulated by the microenvironment. *Nat. Cell Biol.* **2010**, *12*, 468–476. [CrossRef] [PubMed]

- 65. Cullen, S.M.; Mayle, A.; Rossi, L.; Goodell, M.A. Hematopoietic stem cell development: An epigenetic journey. *Curr. Top. Dev. Biol.* **2014**, 107, 39–75.
- 66. Clevers, H. The Intestinal Crypt, A Prototype Stem Cell Compartment. Cell 2013, 154, 274–284. [CrossRef]
- 67. Gage, F.H.; Temple, S. Neural Stem Cells: Generating and Regenerating the Brain. Neuron 2013, 80, 588-601. [CrossRef]
- 68. Foudi, A.; Hochedlinger, K.; Van Buren, D.; Schindler, J.W.; Jaenisch, R.; Carey, V.; Hock, H. Analysis of histone 2B-GFP retention reveals slowly cycling hematopoietic stem cells. *Nat. Biotechnol.* **2008**, 27, 84–90. [CrossRef]
- 69. Baker, A.M.; Cereser, B.; Melton, S.; Fletcher, A.G.; Rodriguez-Justo, M.; Tadrous, P.J.; Humphries, A.; Elia, G.; McDonald, S.A.; Wright, N.A.; et al. Quantification of crypt and stem cell evolution in the normal and neoplastic human colon. *Cell Rep.* **2014**, *8*, 940–947. [CrossRef]
- 70. Castilho, R.M.; Squarize, C.H.; Chodosh, L.A.; Williams, B.O.; Gutkind, J.S. mTOR Mediates Wnt-Induced Epidermal Stem Cell Exhaustion and Aging. *Cell Stem Cell* **2009**, *5*, 279–289. [CrossRef]
- 71. Park, D.S.; Akuffo, A.A.; Muench, D.E.; Grimes, H.L.; Epling-Burnette, P.K.; Maini, P.K.; Anderson, A.R.A.; Bonsall, M.B. Clonal hematopoiesis of indeterminate potential and its impact on patient trajectories after stem cell transplantation. *PLoS Comput. Biol.* **2019**, *15*, e1006913. [CrossRef]
- 72. O'reilly, S.; Tsou, P.-S.; Varga, J. Senescence and tissue fibrosis: Opportunities for therapeutic targeting. *Trends Mol. Med.* **2024**, *30*, 1113–1125. [CrossRef]
- 73. Cicalese, A.; Bonizzi, G.; Pasi, C.E.; Faretta, M.; Ronzoni, S.; Giulini, B.; Brisken, C.; Minucci, S.; Di Fiore, P.P.; Pelicci, P.G. The Tumor Suppressor p53 Regulates Polarity of Self-Renewing Divisions in Mammary Stem Cells. *Cell* **2009**, *138*, 1083–1095. [CrossRef] [PubMed]
- 74. Lee, J.Y.; Nakada, D.; Yilmaz, O.H.; Tothova, Z.; Joseph, N.M.; Lim, M.S.; Gilliland, D.G.; Morrison, S.J. mTOR Activation Induces Tumor Suppressors that Inhibit Leukemogenesis and Deplete Hematopoietic Stem Cells after Pten Deletion. *Cell Stem Cell* 2010, 7, 593–605. [CrossRef] [PubMed]
- 75. Ferent, J.; Cochard, L.; Faure, H.; Taddei, M.; Hahn, H.; Ruat, M.; Traiffort, E. Genetic Activation of Hedgehog Signaling Unbalances the Rate of Neural Stem Cell Renewal by Increasing Symmetric Divisions. *Stem Cell Rep.* **2014**, *3*, 312–323. [CrossRef]
- 76. Gonzalez-Cano, L.; Herreros-Villanueva, M.; Fernandez-Alonso, R.; Ayuso-Sacido, A.; Meyer, G.; Garcia-Verdugo, J.M.; Silva, A.; Marques, M.M.; Marin, M.C. p73 deficiency results in impaired self renewal and premature neuronal differentiation of mouse neural progenitors independently of p53. *Cell Death Dis.* **2010**, *1*, e109. [CrossRef]
- 77. Lukaszewicz, A.I.; Nguyen, C.; Melendez, E.; Lin, D.P.; Teo, J.-L.; Lai, K.K.Y.; Huttner, W.B.; Shi, S.-H.; Kahn, M. The Mode of Stem Cell Division Is Dependent on the Differential Interaction of β-Catenin with the Kat3 Coactivators CBP or p300. *Cancers* **2019**, *11*, 962. [CrossRef]
- 78. Llorens-Bobadilla, E.; Zhao, S.; Baser, A.; Saiz-Castro, G.; Zwadlo, K.; Martin-Villalba, A. Single-Cell Transcriptomics Reveals a Population of Dormant Neural Stem Cells that Become Activated upon Brain Injury. *Cell Stem Cell* 2015, 17, 329–340. [CrossRef]
- 79. Rodgers, J.T.; King, K.Y.; Brett, J.O.; Cromie, M.J.; Charville, G.W.; Maguire, K.K.; Brunson, C.; Mastey, N.; Liu, L.; Tsai, C.-R.; et al. mTORC1 controls the adaptive transition of quiescent stem cells from G0 to GAlert. *Nature* **2014**, *510*, 393–396. [CrossRef]
- 80. Otsuki, L.; Brand, A.H. Cell cycle heterogeneity directs the timing of neural stem cell activation from quiescence. *Science* **2018**, 360, 99–102. [CrossRef]
- 81. Davies, A.; Zoubeidi, A.; Beltran, H.; Selth, L.A. The Transcriptional and Epigenetic Landscape of Cancer Cell Lineage Plasticity. *Cancer Discov.* **2023**, *13*, 1771–1788. [CrossRef]
- 82. Montgomery, R.K.; Carlone, D.L.; Richmond, C.A.; Farilla, L.; Kranendonk, M.E.G.; Henderson, D.E.; Baffour-Awuah, N.Y.; Ambruzs, D.M.; Fogli, L.K.; Algra, S.; et al. Mouse telomerase reverse transcriptase (mTert) expression marks slowly cycling intestinal stem cells. *Proc. Natl. Acad. Sci. USA* **2010**, *108*, 179–184. [CrossRef]
- 83. Takeda, N.; Jain, R.; LeBoeuf, M.R.; Wang, Q.; Lu, M.M.; Epstein, J.A. Interconversion between intestinal stem cell popu-lations in distinct niches. *Science* **2011**, *334*, 1420–1424. [CrossRef] [PubMed]
- 84. Sangiorgi, E.; Capecchi, M.R. Bmi1 is expressed in vivo in intestinal stem cells. *Nat. Genet.* 2008, 40, 915–920. [CrossRef] [PubMed]
- 85. Barker, N.; Van Es, J.H.; Kuipers, J.; Kujala, P.; Van Den Born, M.; Cozijnsen, M.; Haegebarth, A.; Korving, J.; Begthel, H.; Peters, P.J.; et al. Identification of stem cells in small intestine and colon by marker gene Lgr5. *Nature* **2007**, 449, 1003–1007. [CrossRef]
- 86. Potten, C.S.; Hume, W.J.; Reid, P.; Cairns, J. The segregation of DNA in epithelial stem cells. Cell 1978, 15, 899–906. [CrossRef]
- 87. Ritsma, L.; Ellenbroek, S.I.J.; Zomer, A.; Snippert, H.J.; de Sauvage, F.J.; Simons, B.D.; Clevers, H.; van Rheenen, J. Intestinal crypt homeostasis revealed at single-stem-cell level by in vivo live imaging. *Nature* **2014**, *507*, 362–365. [CrossRef]
- 88. De Mey, J.R.; Freund, J.N. Understanding epithelial homeostasis in the intestine: An old battlefield of ideas, recent breakthroughs and remaining controversies. *Tissue Barriers* **2013**, *1*, e24965. [CrossRef]

Cancers 2025, 17, 1503 17 of 22

89. Vazquez, E.G.; Nasreddin, N.; Valbuena, G.N.; Mulholland, E.J.; Belnoue-Davis, H.L.; Eggington, H.R.; Schenck, R.O.; Wouters, V.M.; Wirapati, P.; Gilroy, K.; et al. Dynamic and adaptive cancer stem cell population admixture in colorectal neoplasia. *Cell Stem Cell* 2022, 29, 1612. [CrossRef]

- 90. Ayyaz, A.; Kumar, S.; Sangiorgi, B.; Ghoshal, B.; Gosio, J.; Ouladan, S.; Fink, M.; Barutcu, S.; Trcka, D.; Shen, J.; et al. Single-cell transcriptomes of the regenerating intestine reveal a revival stem cell. *Nature* **2019**, *569*, 121–125. [CrossRef]
- 91. Yui, S.; Azzolin, L.; Maimets, M.; Pedersen, M.T.; Fordham, R.P.; Hansen, S.L.; Larsen, H.L.; Guiu, J.; Alves, M.R.; Rundsten, C.F.; et al. YAP/TAZ-Dependent Reprogramming of Colonic Epithelium Links ECM Remodeling to Tissue Regeneration. *Cell Stem Cell* 2018, 22, 35–49.e7. [CrossRef]
- 92. Harnack, C.; Berger, H.; Antanaviciute, A.; Vidal, R.; Sauer, S.; Simmons, A.; Meyer, T.F.; Sigal, M. R-spondin 3 promotes stem cell recovery and epithelial regeneration in the colon. *Nat. Commun.* **2019**, *10*, 4368. [CrossRef]
- 93. Oh, I.; Humphries, R. Concise Review: Multidimensional Regulation of the Hematopoietic Stem Cell State. *Stem Cells* **2011**, *30*, 82–88. [CrossRef] [PubMed]
- 94. Rompolas, P.; Mesa, K.R.; Greco, V. Spatial organization within a niche as a determinant of stem-cell fate. *Nature* **2013**, *502*, 513–518. [CrossRef] [PubMed]
- 95. Bailleul, B.; Surani, M.; White, S.; Barton, S.C.; Brown, K.; Blessing, M.; Jorcano, J.; Balmain, A. Skin hyperkeratosis and papilloma formation in transgenic mice expressing a ras oncogene from a suprabasal keratin promoter. *Cell* **1990**, *62*, *697*–708. [CrossRef]
- 96. Barker, N.; Ridgway, R.A.; Van Es, J.H.; Van De Wetering, M.; Begthel, H.; van den Born, M.; Danenberg, E.; Clarke, A.R.; Sansom, O.J.; Clevers, H. Crypt stem cells as the cells-of-origin of intestinal cancer. *Nature* **2009**, 457, 608–611. [CrossRef]
- 97. Zhu, L.; Gibson, P.; Currle, D.S.; Tong, Y.; Richardson, R.J.; Bayazitov, I.T.; Poppleton, H.; Zakharenko, S.; Ellison, D.W.; Gilbertson, R.J. Prominin 1 marks intestinal stem cells that are susceptible to neoplastic transformation. *Nature* **2008**, 457, 603–607. [CrossRef]
- 98. Mantovani, A.; Allavena, P.; Sica, A.; Balkwill, F. Cancer-related inflammation. Nature 2008, 454, 436–444. [CrossRef]
- Chan, A.T.; Ogino, S.; Fuchs, C.S. Aspirin and the Risk of Colorectal Cancer in Relation to the Expression of COX-2. N. Engl. J. Med. 2007, 356, 2131–2142. [CrossRef]
- 100. Saha, A.; Kaul, R.; Murakami, M.; Robertson, E.S. Tumor viruses and cancer biology: Modulating signaling pathways for therapeutic intervention. *Cancer Biol. Ther.* **2010**, *10*, 961–978. [CrossRef]
- 101. Liu, S.; Cong, Y.; Wang, D.; Sun, Y.; Deng, L.; Liu, Y.; Martin-Trevino, R.; Shang, L.; McDermott, S.P.; Landis, M.D.; et al. Breast cancer stem cells transition between epithelial and mesenchymal states reflective of their normal counterparts. *Stem Cell Rep.* **2014**, *2*, 78–91. [CrossRef]
- 102. Kahn, M. A Metabolic Smorgasbord Drives and Sustains CSC Heterogeneity. Cancers 2023, 15, 343. [CrossRef]
- 103. Tanabe, A.; Sahara, H. The Metabolic Heterogeneity and Flexibility of Cancer Stem Cells. *Cancers* **2020**, *12*, 2780. [CrossRef] [PubMed]
- 104. Sancho, P.; Barneda, D.; Heeschen, C. Hallmarks of cancer stem cell metabolism. *Br. J. Cancer* **2016**, *114*, 1305–1312. [CrossRef] [PubMed]
- 105. Dando, I.; Pozza, E.D.; Biondani, G.; Cordani, M.; Palmieri, M.; Donadelli, M. The metabolic landscape of cancer stem cells. *IUBMB Life* **2015**, *67*, *687*–*693*. [CrossRef] [PubMed]
- 106. Derks, L.L.M.; van Boxtel, R. Stem cell mutations, associated cancer risk, and consequences for regenerative medicine. *Cell Stem Cell* 2023, 30, 1421–1433. [CrossRef]
- 107. van der Weijden, V.A.; Bulut-Karslioğlu, A. Embryos burn fat in standby. Trends Cell Biol. 2024, 34, 700-702. [CrossRef]
- 108. Chimge, N.-O.; Chen, M.-H.; Nguyen, C.; Zhao, Y.; Wu, X.; Gonzalez, P.G.; Ogana, H.; Hurwitz, S.; Teo, J.-L.; Chen, X.; et al. A Deeply Quiescent Subset of CML LSC depend on FAO yet Avoid Deleterious ROS by Suppressing Mitochondrial Complex I. *Curr. Mol. Pharmacol.* **2023**, 17, e060923220758. [CrossRef]
- 109. Kohli, L.; Passegué, E. Surviving change: The metabolic journey of hematopoietic stem cells. *Trends Cell Biol.* **2014**, 24, 479–487. [CrossRef]
- 110. Ito, K.; Carracedo, A.; Weiss, D.; Arai, F.; Ala, U.; Avigan, D.E.; Schafer, Z.T.; Evans, R.M.; Suda, T.; Lee, C.H.; et al. A PML-PPAR-delta pathway for fatty acid oxidation regulates hematopoietic stem cell maintenance. *Nat. Med.* **2012**, *18*, 1350–1358. [CrossRef]
- 111. Rodríguez-Nuevo, A.; Torres-Sanchez, A.; Duran, J.M.; De Guirior, C.; Martínez-Zamora, M.A.; Böke, E. Oocytes maintain ROS-free mitochondrial metabolism by suppressing complex I. *Nature* **2022**, *607*, 756–761. [CrossRef]
- 112. Brand, M.D. The sites and topology of mitochondrial superoxide production. Exp. Gerontol. 2010, 45, 466–472. [CrossRef]
- 113. Zhao, Y.; Masiello, D.; McMillian, M.; Nguyen, C.; Wu, Y.; Melendez, E.; Smbatyan, G.; Kida, A.; He, Y.; Teo, J.-L.; et al. CBP/catenin antagonist safely eliminates drug-resistant leukemia-initiating cells. *Oncogene* **2015**, *35*, 3705–3717. [CrossRef] [PubMed]
- 114. Lytle, N.K.; Barber, A.G.; Reya, T. Stem cell fate in cancer growth, progression and therapy resistance. *Nat. Rev. Cancer* **2018**, *18*, 669–680. [CrossRef] [PubMed]

Cancers 2025, 17, 1503 18 of 22

115. Plaks, V.; Kong, N.; Werb, Z. The Cancer Stem Cell Niche: How Essential Is the Niche in Regulating Stemness of Tumor Cells? *Cell Stem Cell* **2015**, *16*, 225–238. [CrossRef]

- 116. Kühl, S.J.; Kühl, M. On the role of Wnt/β-catenin signaling in stem cells. *Biochim. Biophys. Acta* (*BBA*)-*Gen. Subj.* **2013**, 1830, 2297–2306. [CrossRef]
- 117. Zhan, T.; Rindtorff, N.; Boutros, M. Wnt signaling in cancer. Oncogene 2017, 36, 1461–1473. [CrossRef]
- 118. Polakis, P. Drugging Wnt signalling in cancer. EMBO J. 2012, 31, 2737–2746. [CrossRef]
- 119. Veltri, A.; Lang, C.; Lien, W.-H. Concise Review: Wnt Signaling Pathways in Skin Development and Epidermal Stem Cells. *Stem Cells* **2017**, *36*, 22–35. [CrossRef]
- 120. Komiya, Y.; Habas, R. Wnt signal transduction pathways. Organogenesis 2008, 4, 68–75. [CrossRef]
- 121. Sato, N.; Meijer, L.; Skaltsounis, L.; Greengard, P.; Brivanlou, A.H. Maintenance of pluripotency in human and mouse embryonic stem cells through activation of Wnt signaling by a pharmacological GSK-3-specific inhibitor. *Nat. Med.* **2003**, *10*, 55–63. [CrossRef]
- 122. Chenn, A.; Walsh, C.A. Regulation of Cerebral Cortical Size by Control of Cell Cycle Exit in Neural Precursors. *Science* **2002**, 297, 365–369. [CrossRef]
- 123. Otero, J.J.; Fu, W.; Kan, L.; Cuadra, A.E.; Kessler, J.A. Beta-catenin signaling is required for neural differentiation of em-bryonic stem cells. *Development* 2004, 131, 3545–3557. [CrossRef] [PubMed]
- 124. Van Camp, J.K.; Beckers, S.; Zegers, D.; Van Hul, W. Wnt Signaling and the Control of Human Stem Cell Fate. *Stem Cell Rev. Rep.* **2013**, *10*, 207–229. [CrossRef] [PubMed]
- 125. Hari, L.; Brault, V.; Kléber, M.; Lee, H.-Y.; Ille, F.; Leimeroth, R.; Paratore, C.; Suter, U.; Kemler, R.; Sommer, L. Lineage-specific requirements of *β-catenin* in neural crest development. *J. Cell Biol.* **2002**, *159*, 867–880. [CrossRef] [PubMed]
- 126. Famili, F.; Brugman, M.H.; Taskesen, E.; Naber, B.E.; Fodde, R.; Staal, F.J. High Levels of Canonical Wnt Signaling Lead to Loss of Stemness and Increased Differentiation in Hematopoietic Stem Cells. *Stem Cell Rep.* **2016**, *6*, 652–659. [CrossRef]
- 127. Essex, A.; Pineda, J.; Acharya, G.; Xin, H.; Evans, J.; Reproducibility Project: Cancer Biology. Replication Study: Wnt activity defines colon cancer stem cells and is regulated by the microenvironment. *eLife* **2019**, *8*, e45426. [CrossRef]
- 128. De Sousa e Melo, F.; Vermeulen, L. Wnt Signaling in Cancer Stem Cell Biology. Cancers 2016, 8, 60. [CrossRef]
- 129. Ring, A.; Kim, Y.-M.; Kahn, M. Wnt/Catenin Signaling in Adult Stem Cell Physiology and Disease. *Stem Cell Rev. Rep.* **2014**, *10*, 512–525. [CrossRef]
- 130. Lugli, A.; Zlobec, I.; Minoo, P.; Baker, K.; Tornillo, L.; Terracciano, L.; Jass, J.R. Prognostic significance of the wnt signalling pathway molecules APC, beta-catenin and E-cadherin in colorectal cancer: A tissue microarray-based analysis. *Histopathology* **2007**, *50*, 453–464. [CrossRef]
- 131. Takada, T.; Yagi, Y.; Maekita, T.; Imura, M.; Nakagawa, S.; Tsao, S.; Miyamoto, K.; Yoshino, O.; Yasugi, T.; Taketani, Y.; et al. Methylation-associated silencing of the Wnt antagonist *SFRP1* gene in human ovarian cancers. *Cancer Sci.* **2004**, *95*, 741–744. [CrossRef]
- 132. Cao, X.; Eu, K.W.; Seow-Choen, F.; Zao, Y.; Cheah, P.Y. APC mutation and phenotypic spectrum of Singapore familial adenomatous polyposis patients. *Eur. J. Hum. Genet.* **2000**, *8*, 42–48. [CrossRef]
- 133. Chien, A.J.; Moore, E.C.; Lonsdorf, A.S.; Kulikauskas, R.M.; Rothberg, B.G.; Berger, A.J.; Major, M.B.; Hwang, S.T.; Rimm, D.L.; Moon, R.T. Activated Wnt/β-catenin signaling in melanoma is associated with decreased proliferation in patient tumors and a murine melanoma model. *Proc. Natl. Acad. Sci. USA* 2009, *106*, 1193–1198. [CrossRef] [PubMed]
- 134. Bachmann, I.M.; Straume, O.; Puntervoll, H.E.; Kalvenes, M.B.; Akslen, L.A. Importance of P-Cadherin, β-Catenin, and Wnt5a/Frizzled for Progression of Melanocytic Tumors and Prognosis in Cutaneous Melanoma. *Clin. Cancer Res.* **2005**, *11*, 8606–8614. [CrossRef]
- 135. Maelandsmo, G.M.; Holm, R.; Nesland, J.M.; Fodstad, Ø.; Flørenes, V.A. Reduced beta-catenin expression in the cytoplasm of advanced-stage superficial spreading malignant melanoma. *Clin. Cancer Res.* **2003**, *9*, 3383–3388.
- 136. Kageshita, T.; Hamby, C.; Ishihara, T.; Matsumoto, K.; Saida, T.; Ono, T. Loss of beta-catenin expression associated with disease progression in malignant melanoma. *Br. J. Dermatol.* **2001**, *145*, 210–216. [CrossRef] [PubMed]
- 137. Pozniak, J.; Pedri, D.; Landeloos, E.; Van Herck, Y.; Antoranz, A.; Vanwynsberghe, L.; Nowosad, A.; Roda, N.; Makhzami, S.; Bervoets, G.; et al. A TCF4-dependent gene regulatory network confers resistance to immunotherapy in melanoma. *Cell* 2024, 187, 166–183.e25. [CrossRef]
- 138. Luke, J.J.; Bao, R.; Spranger, S.; Sweis, R.F.; Gajewski, T. Correlation of WNT/β-catenin pathway activation with immune exclusion across most human cancers. *J. Clin. Oncol.* **2016**, *34*, 3004. [CrossRef]
- 139. MacDougall, M.S.; Clarke, R.; Merrill, B.J. Intracellular Ca²⁺ Homeostasis and Nuclear Export Mediate Exit from Naive Pluripotency. *Cell Stem Cell* **2019**, 25, 210–224.e6. [CrossRef]
- 140. Abou Ziki, M.; Mani, A. The interplay of canonical and noncanonical Wnt signaling in metabolic syndrome. *Nutr. Res.* **2019**, *70*, 18–25. [CrossRef]
- 141. Yoon, J.C.; Ng, A.; Kim, B.H.; Bianco, A.; Xavier, R.J.; Elledge, S.J. Wnt signaling regulates mitochondrial physiology and insulin sensitivity. *Genes Dev.* **2010**, 24, 1507–1518. [CrossRef]

Cancers 2025, 17, 1503

142. Hu, X.; Ono, M.; Chimge, N.-O.; Chosa, K.; Nguyen, C.; Melendez, E.; Lou, C.-H.; Lim, P.; Termini, J.; Lai, K.K.Y.; et al. Differential Kat3 Usage Orchestrates the Integration of Cellular Metabolism with Differentiation. *Cancers* **2021**, *13*, 5884. [CrossRef]

- 143. Jeong, Y.; Mangelsdorf, D.J. Nuclear receptor regulation of stemness and stem cell differentiation. *Exp. Mol. Med.* **2009**, 41, 525–537. [CrossRef] [PubMed]
- 144. Preidis, G.A.; Kim, K.H.; Moore, D.D. Nutrient-sensing nuclear receptors PPARα and FXR control liver energy balance. *J. Clin. Investig.* **2017**, 127, 1193–1201. [CrossRef] [PubMed]
- 145. Dhillon, P.; Park, J.; del Pozo, C.H.; Li, L.; Doke, T.; Huang, S.; Zhao, J.; Kang, H.M.; Shrestra, R.; Balzer, M.S.; et al. The Nuclear Receptor ESRRA Protects from Kidney Disease by Coupling Metabolism and Differentiation. *Cell Metab.* 2020, 33, 379–394.e8. [CrossRef]
- 146. Kato, H.; Gruenwald, A.; Suh, J.H.; Miner, J.H.; Barisoni-Thomas, L.; Taketo, M.M.; Faul, C.; Millar, S.E.; Holzman, L.B.; Susztak, K. Wnt/β-catenin pathway in podocytes integrates cell adhesion, differentiation, and survival. *J. Biol. Chem.* **2011**, 286, 26003–26015. [CrossRef]
- 147. He, W.; Dai, C.; Li, Y.; Zeng, G.; Monga, S.P.; Liu, Y. Wnt/β-Catenin Signaling Promotes Renal Interstitial Fibrosis. *J. Am. Soc. Nephrol.* **2009**, 20, 765–776. [CrossRef]
- 148. Ivashkiv, L.B. IFNγ: Signalling, epigenetics and roles in immunity, metabolism, disease and cancer immunotherapy. *Nat. Rev. Immunol.* **2018**, *18*, 545–558. [CrossRef]
- 149. Vu, T.N.; Chen, X.; Foda, H.D.; Smaldone, G.C.; Hasaneen, N.A. Interferon-γ enhances the antifibrotic effects of pirfenidone by attenuating IPF lung fibroblast activation and differentiation. *Respir. Res.* **2019**, *20*, 206. [CrossRef]
- 150. Law, B.M.; Wilkinson, R.; Wang, X.; Kildey, K.; Lindner, M.; Rist, M.J.; Beagley, K.; Healy, H.; Kassianos, A.J. Interferon-γ production by tubulointerstitial human CD56bright natural killer cells contributes to renal fibrosis and chronic kidney disease progression. *Kidney Int.* **2017**, *92*, 79–88. [CrossRef]
- 151. Teo, J.-L.; Kahn, M. The Wnt signaling pathway in cellular proliferation and differentiation: A tale of two coactivators. *Adv. Drug Deliv. Rev.* **2010**, *62*, 1149–1155. [CrossRef]
- 152. Hecht, A.; Vleminckx, K.; Stemmler, M.P.; van Roy, F.; Kemler, R. The p300/CBP acetyltransferases function as transcrip-tional coactivators of beta-catenin in vertebrates. *EMBO J.* **2000**, *19*, 1839–1850. [CrossRef]
- 153. Takemaru, K.-I.; Moon, R.T. The Transcriptional Coactivator Cbp Interacts with β-Catenin to Activate Gene Expression. *J. Cell Biol.* **2000**, 149, 249–254. [CrossRef] [PubMed]
- 154. Mi, H.; Muruganujan, A.; Thomas, P.D. PANTHER in 2013: Modeling the evolution of gene function, and other gene attributes, in the context of phylogenetic trees. *Nucleic Acids Res.* **2013**, *41*, D377–D386. [CrossRef] [PubMed]
- 155. Arany, Z.; Sellers, W.R.; Livingston, D.M.; Eckner, R. E1A-associated p300 and CREB-associated CBP belong to a conserved family of coactivators. *Cell* 1994, 77, 799–800. [CrossRef] [PubMed]
- 156. Eckner, R.; Arany, Z.; Ewen, M.; Sellers, W.; Livingston, D. The Adenovirus E1A-associated 300-kD Protein Exhibits Properties of a Transcriptional Coactivator and Belongs to an Evolutionarily Conserved Family. *Cold Spring Harb. Symp. Quant. Biol.* **1994**, *59*, 85–95. [CrossRef]
- 157. Fauquier, L.; Azzag, K.; Parra, M.A.M.; Quillien, A.; Boulet, M.; Diouf, S.; Carnac, G.; Waltzer, L.; Gronemeyer, H.; Vandel, L. CBP and P300 regulate distinct gene networks required for human primary myoblast differentiation and muscle integrity. *Sci. Rep.* **2018**, *8*, 12629. [CrossRef]
- 158. Roth, J.; Shikama, N.; Henzen, C.; Desbaillets, I.; Lutz, W.; Marino, S.; Wittwer, J.; Schorle, H.; Gassmann, M.; Eckner, R. Differential role of p300 and CBP acetyltransferase during myogenesis: p300 acts upstream of MyoD and Myf5. *EMBO J.* 2003, 22, 5186–5196. [CrossRef]
- 159. Yamauchi, T.; Oike, Y.; Kamon, J.; Waki, H.; Komeda, K.; Tsuchida, A.; Date, Y.; Li, M.-X.; Miki, H.; Akanuma, Y.; et al. Increased insulin sensitivity despite lipodystrophy in Crebbp heterozygous mice. *Nat. Genet.* **2002**, *30*, 221–226. [CrossRef]
- 160. Kung, A.L.; Rebel, V.I.; Bronson, R.T.; Ch'Ng, L.-E.; Sieff, C.A.; Livingston, D.M.; Yao, T.-P. Gene dose-dependent control of hematopoiesis and hematologic tumor suppression by CBP. *Genes Dev.* **2000**, *14*, 272–277. [CrossRef]
- 161. Emami, K.H.; Nguyen, C.; Ma, H.; Kim, D.H.; Jeong, K.W.; Eguchi, M.; Moon, R.T.; Teo, J.; Oh, S.W.; Kim, H.Y.; et al. A small molecule inhibitor of beta-catenin/CREB-binding protein transcription. *Proc. Natl. Acad. Sci. USA* 2004, 101, 12682–12687, Erratum in *Proc. Natl. Acad. Sci. USA* 2004, 101, 16707. [CrossRef]
- 162. McMillan, M.; Kahn, M. Investigating Wnt signaling: A chemogenomic safari. Drug Discov. Today 2005, 10, 1467–1474. [CrossRef]
- 163. Higuchi, Y.; Nguyen, C.; Yasuda, S.-Y.; McMillan, M.; Hasegawa, K.; Kahn, M. Specific Direct Small Molecule p300/β-Catenin Antagonists Maintain Stem Cell Potency. *Curr. Mol. Pharmacol.* **2016**, *9*, 272–279. [CrossRef] [PubMed]
- 164. Hasegawa, K.; Yasuda, S.-Y.; Teo, J.-L.; Nguyen, C.; McMillan, M.; Hsieh, C.-L.; Suemori, H.; Nakatsuji, N.; Yamamoto, M.; Miyabayashi, T.; et al. Wnt Signaling Orchestration with a Small Molecule DYRK Inhibitor Provides Long-Term Xeno-Free Human Pluripotent Cell Expansion. *Stem Cells Transl. Med.* 2011, 1, 18–28. [CrossRef] [PubMed]

Cancers 2025, 17, 1503 20 of 22

165. Rieger, M.E.; Zhou, B.; Solomon, N.; Sunohara, M.; Li, C.; Nguyen, C.; Liu, Y.; Pan, J.-H.; Minoo, P.; Crandall, E.D.; et al. p300/β-Catenin Interactions Regulate Adult Progenitor Cell Differentiation Downstream of WNT5a/Protein Kinase C (PKC). *J. Biol. Chem.* **2016**, 291, 6569–6582. [CrossRef] [PubMed]

- 166. Banerjee, E.R.; Laflamme, M.A.; Papayannopoulou, T.; Kahn, M.; Murry, C.E.; Henderson, W.R. Human Embryonic Stem Cells Differentiated to Lung Lineage-Specific Cells Ameliorate Pulmonary Fibrosis in a Xenograft Transplant Mouse Model. *PLoS ONE* **2012**, 7, e33165. [CrossRef]
- 167. Teo, J.-L.; Ma, H.; Nguyen, C.; Lam, C.; Kahn, M. Specific inhibition of CBP/β-catenin interaction rescues defects in neuronal differentiation caused by a presentiin-1 mutation. *Proc. Natl. Acad. Sci. USA* **2005**, *102*, 12171–12176. [CrossRef]
- 168. Miyabayashi, T.; Teo, J.-L.; Yamamoto, M.; McMillan, M.; Nguyen, C.; Kahn, M. Wnt/β-catenin/CBP signaling maintains long-term murine embryonic stem cell pluripotency. *Proc. Natl. Acad. Sci. USA* **2007**, 104, 5668–5673. [CrossRef]
- 169. Zhao, Y.; Wu, K.; Nguyen, C.; Smbatyan, G.; Melendez, E.; Higuchi, Y.; Chen, Y.; Kahn, M. Small molecule p300/catenin antagonist enhances hematopoietic recovery after radiation. *PLoS ONE* **2017**, *12*, e0177245. [CrossRef]
- 170. Marson, A.; Foreman, R.; Chevalier, B.; Bilodeau, S.; Kahn, M.; Young, R.A.; Jaenisch, R. Wnt Signaling Promotes Reprogramming of Somatic Cells to Pluripotency. *Cell Stem Cell* **2008**, *3*, 132–135. [CrossRef]
- 171. Manegold, P.; Lai, K.K.Y.; Wu, Y.; Teo, J.-L.; Lenz, H.-J.; Genyk, Y.S.; Pandol, S.J.; Wu, K.; Lin, D.P.; Chen, Y.; et al. Differentiation Therapy Targeting the β-Catenin/CBP Interaction in Pancreatic Cancer. *Cancers* **2018**, *10*, 95. [CrossRef]
- 172. Kim, Y.-M.; Gang, E.-J.; Kahn, M. CBP/Catenin antagonists: Targeting LSCs' Achilles heel. Exp. Hematol. 2017, 52, 1–11. [CrossRef]
- 173. Gang, E.J.; Hsieh, Y.-T.; Pham, J.; Zhao, Y.; Nguyen, C.; Huantes, S.; Park, E.; Naing, K.; Klemm, L.; Swaminathan, S.; et al. Small-molecule inhibition of CBP/catenin interactions eliminates drug-resistant clones in acute lymphoblastic leukemia. *Oncogene* **2013**, 33, 2169–2178. [CrossRef] [PubMed]
- 174. Wend, P.; Fang, L.; Zhu, Q.; Schipper, J.H.; Loddenkemper, C.; Kosel, F.; Brinkmann, V.; Eckert, K.; Hindersin, S.; Holland, J.D.; et al. Wnt/β-catenin signalling induces MLL to create epigenetic changes in salivary gland tumours. *EMBO J.* **2013**, 32, 1977–1989. [CrossRef] [PubMed]
- 175. Osawa, Y.; Oboki, K.; Imamura, J.; Kojika, E.; Hayashi, Y.; Hishima, T.; Saibara, T.; Shibasaki, F.; Kohara, M.; Kimura, K. Inhibition of Cyclic Adenosine Monophosphate (cAMP)-response Element-binding Protein (CREB)-binding Protein (CBP)/β-Catenin Reduces Liver Fibrosis in Mice. *eBioMedicine* **2015**, *2*, 1751–1758. [CrossRef] [PubMed]
- 176. Beyer, C.; Reichert, H.; Akan, H.; Mallano, T.; Schramm, A.; Dees, C.; Palumbo-Zerr, K.; Lin, N.Y.; Distler, A.; Gelse, K.; et al. Blockade of canonical Wnt signalling ameliorates experimental dermal fibrosis. *Ann. Rheum. Dis.* **2013**, 72, 1255–1258. [CrossRef]
- 177. Hao, S.; He, W.; Li, Y.; Ding, H.; Hou, Y.; Nie, J.; Hou, F.F.; Kahn, M.; Liu, Y. Targeted Inhibition of β-Catenin/CBP Signaling Ameliorates Renal Interstitial Fibrosis. *J. Am. Soc. Nephrol.* **2011**, 22, 1642–1653. [CrossRef]
- 178. Henderson, W.R.; Chi, E.Y.; Ye, X.; Nguyen, C.; Tien, Y.-T.; Zhou, B.; Borok, Z.; Knight, D.A.; Kahn, M. Inhibition of Wnt/β-catenin/CREB binding protein (CBP) signaling reverses pulmonary fibrosis. *Proc. Natl. Acad. Sci. USA* **2010**, *107*, 14309–14314. [CrossRef]
- 179. Sasaki, T.; Hwang, H.; Nguyen, C.; Kloner, R.A.; Kahn, M. The Small Molecule Wnt Signaling Modulator ICG-001 Improves Contractile Function in Chronically Infarcted Rat Myocardium. *PLoS ONE* **2013**, *8*, e75010. [CrossRef]
- 180. Lai, K.K.; Hu, X.; Chosa, K.; Nguyen, C.; Lin, D.P.; Lai, K.K.; Kato, N.; Higuchi, Y.; Highlander, S.K.; Melendez, E.; et al. P300 Serine 89: A Critical Signaling Integrator and Its Effects on Intestinal Homeostasis and Repair. *Cancers* 2021, 13, 1288. [CrossRef]
- 181. El-Khoueiry, A.B.; Ning, Y.; Yang, D.; Cole, S.; Kahn, M.; Zoghbi, M.; Berg, J.; Fujimori, M.; Inada, T.; Kouji, H.; et al. A phase I first-in-human study of PRI-724 in patients (pts) with advanced solid tumors. *J. Clin. Oncol.* **2013**, *31*, 2501. [CrossRef]
- 182. Kimura, K.; Ikoma, A.; Shibakawa, M.; Shimoda, S.; Harada, K.; Saio, M.; Imamura, J.; Osawa, Y.; Kimura, M.; Nishikawa, K.; et al. Safety, Tolerability, and Preliminary Efficacy of the Anti-Fibrotic Small Molecule PRI-724, a CBP/β-Catenin Inhibitor, in Patients with Hepatitis C Virus-related Cirrhosis: A Single-Center, Open-Label, Dose Escalation Phase 1 Trial. *eBioMedicine* **2017**, 23, 79–87. [CrossRef]
- 183. Higuchi, Y.; Nguyen, C.; Chimge, N.-O.; Ouyang, C.; Teo, J.-L.; Kahn, M. E7386 is not a Specific CBP/β-Catenin Antagonist. *Curr. Mol. Pharmacol.* **2024**, *17*, e290523217409. [CrossRef] [PubMed]
- 184. Hnisz, D.; Abraham, B.J.; Lee, T.I.; Lau, A.; Saint-André, V.; Sigova, A.A.; Hoke, H.A.; Young, R.A. Super-Enhancers in the Control of Cell Identity and Disease. *Cell* 2013, 155, 934–947. [CrossRef] [PubMed]
- 185. Dowen, J.M.; Fan, Z.P.; Hnisz, D.; Ren, G.; Abraham, B.J.; Zhang, L.N.; Weintraub, A.S.; Schuijers, J.; Lee, T.I.; Zhao, K.; et al. Control of Cell Identity Genes Occurs in Insulated Neighborhoods in Mammalian Chromosomes. *Cell* 2014, 159, 374–387. [CrossRef]
- 186. Lovén, J.; Hoke, H.A.; Lin, C.Y.; Lau, A.; Orlando, D.A.; Vakoc, C.R.; Bradner, J.E.; Lee, T.I.; Young, R.A. Selective Inhibition of Tumor Oncogenes by Disruption of Super-Enhancers. *Cell* 2013, 153, 320–334. [CrossRef] [PubMed]
- 187. Whyte, W.A.; Orlando, D.A.; Hnisz, D.; Abraham, B.J.; Lin, C.Y.; Kagey, M.H.; Rahl, P.B.; Lee, T.I.; Young, R.A. Master Transcription Factors and Mediator Establish Super-Enhancers at Key Cell Identity Genes. *Cell* 2013, 153, 307–319. [CrossRef]

Cancers 2025, 17, 1503 21 of 22

188. Parker, S.C.J.; Stitzel, M.L.; Taylor, D.L.; Orozco, J.M.; Erdos, M.R.; Akiyama, J.A.; van Bueren, K.L.; Chines, P.S.; Narisu, N.; Black, B.L.; et al. Chromatin stretch enhancer states drive cell-specific gene regulation and harbor human disease risk variants. *Proc. Natl. Acad. Sci. USA* **2013**, *110*, 17921–17926. [CrossRef]

- 189. Creyghton, M.P.; Cheng, A.W.; Welstead, G.G.; Kooistra, T.; Carey, B.W.; Steine, E.J.; Hanna, J.; Lodato, M.A.; Frampton, G.M.; Sharp, P.A.; et al. Histone H3K27ac separates active from poised enhancers and predicts developmental state. *Proc. Natl. Acad. Sci. USA* 2010, 107, 21931–21936. [CrossRef]
- 190. Azad, F.M.; Atlasi, Y. WNT-Regulated Transcriptional Enhancers and Stem Cell Plasticity. *Trends Cell Biol.* **2021**, *31*, 525–528. [CrossRef]
- 191. Cho, S.; Lee, G.; Pickering, B.F.; Jang, C.; Park, J.H.; He, L.; Mathur, L.; Kim, S.-S.; Jung, S.; Tang, H.-W.; et al. mTORC1 promotes cell growth via m6A-dependent mRNA degradation. *Mol. Cell* **2021**, *81*, 2064–2075.e8. [CrossRef]
- 192. Martire, S.; Nguyen, J.; Sundaresan, A.; Banaszynski, L.A. Differential contribution of p300 and CBP to regulatory element acetylation in mESCs. *BMC Mol. Cell Biol.* **2020**, *21*, 55. [CrossRef]
- 193. Spitz, F.; Furlong, E.E.M. Transcription factors: From enhancer binding to developmental control. *Nat. Rev. Genet.* **2012**, *13*, 613–626. [CrossRef] [PubMed]
- 194. Rada-Iglesias, A.; Bajpai, R.; Swigut, T.; Brugmann, S.A.; Flynn, R.A.; Wysocka, J. A unique chromatin signature uncovers early developmental enhancers in humans. *Nature* **2010**, 470, 279–283. [CrossRef] [PubMed]
- 195. Zamudio, A.V.; Dall'agnese, A.; Henninger, J.E.; Manteiga, J.C.; Afeyan, L.K.; Hannett, N.M.; Coffey, E.L.; Li, C.H.; Oksuz, O.; Sabari, B.R.; et al. Mediator Condensates Localize Signaling Factors to Key Cell Identity Genes. *Mol. Cell* **2019**, *76*, 753–766.e6. [CrossRef]
- 196. Vo, N.; Goodman, R.H. CREB-binding Protein and p300 in Transcriptional Regulation. *J. Biol. Chem.* **2001**, 276, 13505–13508. [CrossRef] [PubMed]
- 197. Narita, T.; Ito, S.; Higashijima, Y.; Chu, W.K.; Neumann, K.; Walter, J.; Satpathy, S.; Liebner, T.; Hamilton, W.B.; Maskey, E.; et al. Enhancers are activated by p300/CBP activity-dependent PIC assembly, RNAPII recruitment, and pause release. *Mol. Cell* 2021, 81, 2166–2182.e6. [CrossRef]
- 198. Shikama, N.; Lutz, W.; Kretzschmar, R.; Sauter, N.; Roth, J.F.; Marino, S.; Wittwer, J.; Scheidweiler, A.; Eckner, R. Essential function of p300 acetyltransferase activity in heart, lung and small intestine formation. *EMBO J.* **2003**, 22, 5175–5185. [CrossRef]
- 199. Sen, P.; Lan, Y.; Li, C.Y.; Sidoli, S.; Donahue, G.; Dou, Z.; Frederick, B.; Chen, Q.; Luense, L.J.; Garcia, B.A.; et al. Histone Acetyltransferase p300 Induces De Novo Super-Enhancers to Drive Cellular Senescence. *Mol. Cell* 2019, 73, 684–698.e8. [CrossRef]
- 200. Vahedi, G.; Kanno, Y.; Furumoto, Y.; Jiang, K.; Parker, S.C.J.; Erdos, M.R.; Davis, S.R.; Roychoudhuri, R.; Restifo, N.P.; Gadina, M.; et al. Super-enhancers delineate disease-associated regulatory nodes in T cells. *Nature* **2015**, *520*, *558*–*562*. [CrossRef]
- 201. Etchegaray, J.-P.; Lee, C.; Wade, P.A.; Reppert, S.M. Rhythmic histone acetylation underlies transcription in the mammalian circadian clock. *Nature* **2002**, 421, 177–182. [CrossRef]
- 202. Ge, Y.; Fuchs, E. Stretching the limits: From homeostasis to stem cell plasticity in wound healing and cancer. *Nat. Rev. Genet.* **2018**, 19, 311–325. [CrossRef]
- 203. Chapuy, B.; McKeown, M.R.; Lin, C.Y.; Monti, S.; Roemer, M.G.; Qi, J.; Rahl, P.B.; Sun, H.H.; Yeda, K.T.; Doench, J.G.; et al. Discovery and Characterization of Super-Enhancer-Associated Dependencies in Diffuse Large B Cell Lymphoma. *Cancer Cell* 2013, 24, 777–790. [CrossRef] [PubMed]
- 204. Young, R.A. Control of the Embryonic Stem Cell State. Cell 2011, 144, 940–954. [CrossRef] [PubMed]
- 205. Ng, H.-H.; Surani, M.A. The transcriptional and signalling networks of pluripotency. *Nat. Cell Biol.* **2011**, *13*, 490–496. [CrossRef] [PubMed]
- 206. Biehs, B.; Dijkgraaf, G.J.P.; Piskol, R.; Alicke, B.; Boumahdi, S.; Peale, F.; Gould, S.E.; de Sauvage, F.J. A cell identity switch allows residual BCC to survive Hedgehog pathway inhibition. *Nature* **2018**, *562*, 429–433. [CrossRef]
- 207. Wiese, M.; Hamdan, F.H.; Kubiak, K.; Diederichs, C.; Gielen, G.H.; Nussbaumer, G.; Carcaboso, A.M.; Hulleman, E.; Johnsen, S.A.; Kramm, C.M. Combined treatment with CBP and BET inhibitors reverses inadvertent activation of detrimental super enhancer programs in DIPG cells. *Cell Death Dis.* 2020, 11, 673. [CrossRef]
- 208. Smith, J.L.; Freebern, W.J.; Collins, I.; De Siervi, A.; Montano, I.; Haggerty, C.M.; McNutt, M.C.; Butscher, W.G.; Dzekunova, I.; Petersen, D.W.; et al. Kinetic profiles of p300 occupancy in vivo predict common features of promoter structure and coactivator recruitment. *Proc. Natl. Acad. Sci. USA* **2004**, *101*, 11554–11559. [CrossRef]
- 209. Hnisz, D.; Shrinivas, K.; Young, R.A.; Chakraborty, A.K.; Sharp, P.A. A Phase Separation Model for Transcriptional Control. *Cell* **2017**, *169*, 13–23. [CrossRef]
- 210. David, C.J.; Massagué, J. Contextual determinants of TGFβ action in development, immunity and cancer. *Nat. Rev. Mol. Cell Biol.* **2018**, *19*, 419–435. [CrossRef]
- 211. Mullen, A.C.; Wrana, J.L. TGF-β Family Signaling in Embryonic and Somatic Stem-Cell Renewal and Differentiation. *Cold Spring Harb. Perspect. Biol.* **2017**, *9*, a022186. [CrossRef]

Cancers 2025, 17, 1503 22 of 22

212. Nusse, R.; Clevers, H. Wnt/β-Catenin Signaling, Disease, and Emerging Therapeutic Modalities. *Cell* **2017**, *169*, 985–999. [CrossRef]

- 213. Lee, T.I.; Young, R.A. Transcriptional Regulation and Its Misregulation in Disease. Cell 2013, 152, 1237–1251. [CrossRef] [PubMed]
- 214. Tian, X.; Zhang, J.; Tan, T.K.; Lyons, J.G.; Zhao, H.; Niu, B.; Lee, S.R.; Tsatralis, T.; Zhao, Y.; Wang, Y.; et al. Association of beta-catenin with P-Smad3 but not LEF-1 dissociates in vitro profibrotic from anti-inflammatory effects of TGF-beta1. *J. Cell Sci.* 2013, 126, 67–76. [CrossRef] [PubMed]
- 215. Kelly, K.F.; Ng, D.Y.; Jayakumaran, G.; Wood, G.A.; Koide, H.; Doble, B.W. β-Catenin Enhances Oct-4 Activity and Reinforces Pluripotency through a TCF-Independent Mechanism. *Cell Stem Cell* **2011**, *8*, 214–227. [CrossRef] [PubMed]
- 216. Kaidi, A.; Williams, A.C.; Paraskeva, C. Interaction between beta-catenin and HIF-1 promotes cellular adaptation to hypoxia. *Nat. Cell Biol.* **2007**, *9*, 210–217. [CrossRef]
- 217. Essers, M.A.G.; de Vries-Smits, L.M.M.; Barker, N.; Polderman, P.E.; Burgering, B.M.T.; Korswagen, H.C. Functional Interaction Between \(\mathbb{B} \)-Catenin and FOXO in Oxidative Stress Signaling. *Science* **2005**, *308*, 1181–1184. [CrossRef]
- 218. Nateri, A.S.; Spencer-Dene, B.; Behrens, A. Interaction of phosphorylated c-Jun with TCF4 regulates intestinal cancer development. *Nature* **2005**, 437, 281–285. [CrossRef]
- 219. Kouzmenko, A.P.; Takeyama, K.-I.; Ito, S.; Furutani, T.; Sawatsubashi, S.; Maki, A.; Suzuki, E.; Kawasaki, Y.; Akiyama, T.; Tabata, T.; et al. Wnt/β-Catenin and Estrogen Signaling Converge In Vivo. *J. Biol. Chem.* **2004**, 279, 40255–40258. [CrossRef]
- 220. Gerrard, D.L.; Wang, Y.; Gaddis, M.; Zhou, Y.; Wang, J.; Witt, H.; Lin, S.; Farnham, P.J.; Jin, V.X.; Frietze, S.E. Three-dimensional analysis reveals altered chromatin interaction by enhancer inhibitors harbors TCF7L2-regulated cancer gene signature. *J. Cell. Biochem.* 2018, 120, 3056–3070. [CrossRef]
- 221. Gerrard, D.L.; Boyd, J.R.; Stein, G.S.; Jin, V.X.; Frietze, S. Disruption of Broad Epigenetic Domains in PDAC Cells by HAT Inhibitors. *Epigenomes* **2019**, *3*, 11. [CrossRef]
- 222. Zhou, Y.; He, Z.; Li, T.; Choppavarapu, L.; Hu, X.; Cao, R.; Leone, G.W.; Kahn, M.; Jin, V.X. 3D Chromatin Alteration by Disrupting β-Catenin/CBP Interaction Is Enriched with Insulin Signaling in Pancreatic Cancer. *Cancers* **2024**, *16*, 2202. [CrossRef]
- 223. Ring, A.; Nguyen, C.; Smbatyan, G.; Tripathy, D.; Yu, M.; Press, M.; Kahn, M.; Lang, J.E. CBP/β-Catenin/FOXM1 Is a Novel Therapeutic Target in Triple Negative Breast Cancer. *Cancers* **2018**, *10*, 525. [CrossRef] [PubMed]
- 224. Holmqvist, P.-H.; Mannervik, M. Genomic occupancy of the transcriptional co-activators p300 and CBP. *Transcription* **2012**, *4*, 18–23. [CrossRef] [PubMed]
- 225. Didier, S.; Sauvé, F.; Domise, M.; Buée, L.; Marinangeli, C.; Vingtdeux, V. AMP-activated Protein Kinase Controls Immediate Early Genes Expression Following Synaptic Activation Through the PKA/CREB Pathway. *Int. J. Mol. Sci.* 2018, 19, 3716. [CrossRef] [PubMed]
- 226. Kandel, E.R. The molecular biology of memory: cAMP, PKA, CRE, CREB-1, CREB-2, and CPEB. Mol. Brain 2012, 5, 14. [CrossRef]
- 227. Blum, B.; Bar-Nur, O.; Golan-Lev, T.; Benvenisty, N. The anti-apoptotic gene survivin contributes to teratoma formation by human embryonic stem cells. *Nat. Biotechnol.* **2009**, *27*, 281–287. [CrossRef]
- 228. Canato, S.; Sarate, R.; Carvalho-Marques, S.; Soares, R.M.; Song, Y.; Monteiro-Ferreira, S.; Vieugué, P.; Liagre, M.; Grossi, G.; Cardoso, E.; et al. Survivin promotes stem cell competence for skin cancer initiation. *Cancer Discov.* 2025, 15, 427–443. [CrossRef]

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