



Supporting Information

for *Adv. Sci.*, DOI: 10.1002/adv.202101031

IRF8 impacts self-renewal of hematopoietic stem cells by
regulating TLR9 signaling pathway of innate immune cells

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Supporting Information

Title

IRF8 impacts self-renewal of hematopoietic stem cells by regulating TLR9 signaling pathway of innate immune cells

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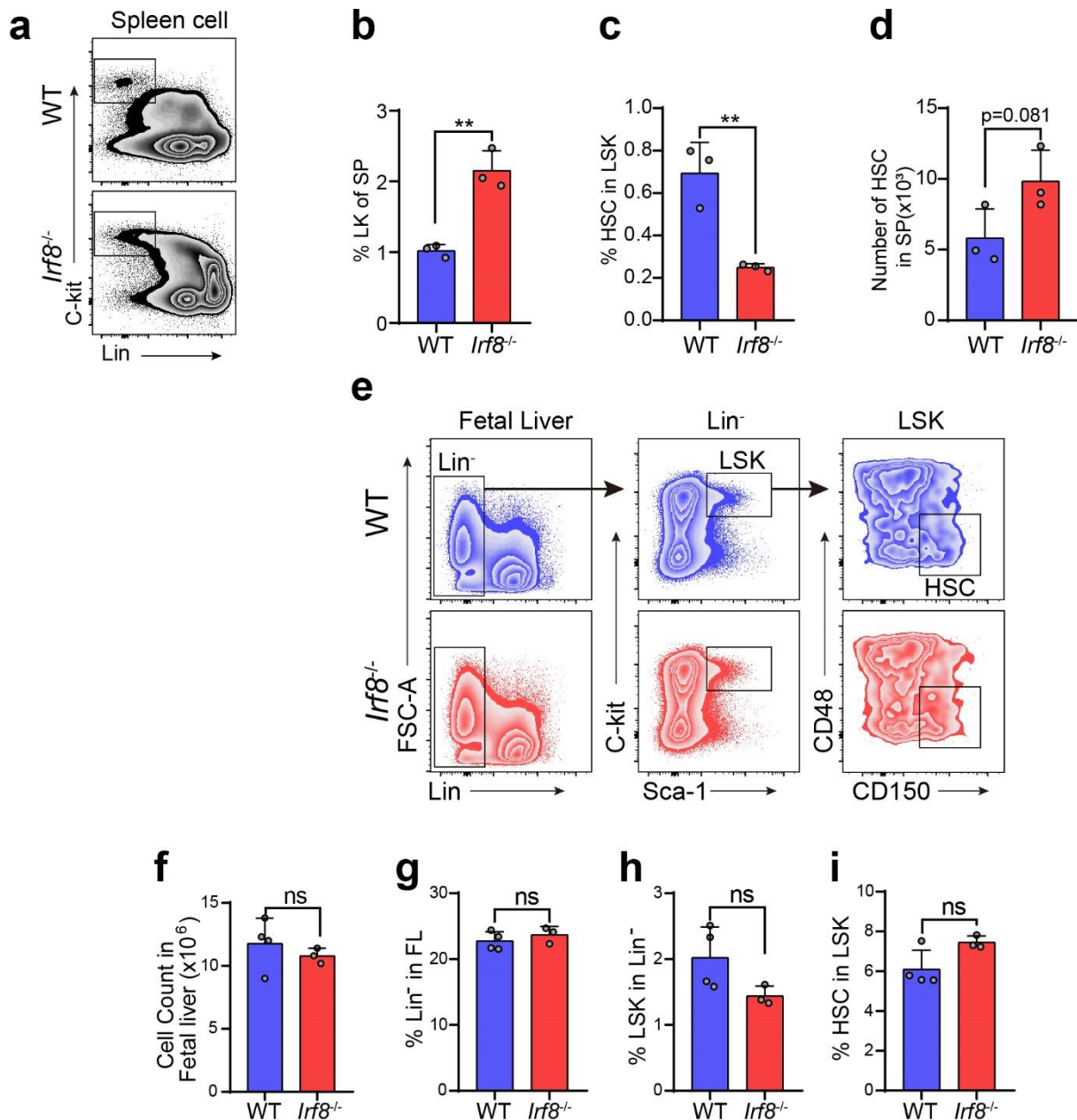


Figure S1. Loss of IRF8 increased the numbers of spleen-derived LT-HSCs from 4-week-old mice and did not change the numbers of LT-HSCs derived from E14.5 mice fetal livers. a) Representative FACS plots of LKs in spleen cells from 4-week-old WT or *Irf8*^{-/-} mice. b) Proportions of LKs in spleen cells and c) the HSC proportions in LSKs. d) Absolute numbers of HSCs in spleen of 4-week-old WT or *Irf8*^{-/-} mice. e) Gating strategy for the analysis of HSCs in the fetal liver cells from E14.5 embryos of WT or *Irf8*^{-/-} mice. f) Total cell numbers in E14.5 fetal liver, and g) the proportions of lineage-negative cells in it. h) Proportions of LSKs in Lin⁻ cells and (i) the HSC proportions in LSKs of WT or *Irf8*^{-/-} fetal livers. Error bars, mean ± s.e.m. ns, not significant, ** *P* < 0.01, data were analyzed with unpaired Student's *t*-test.

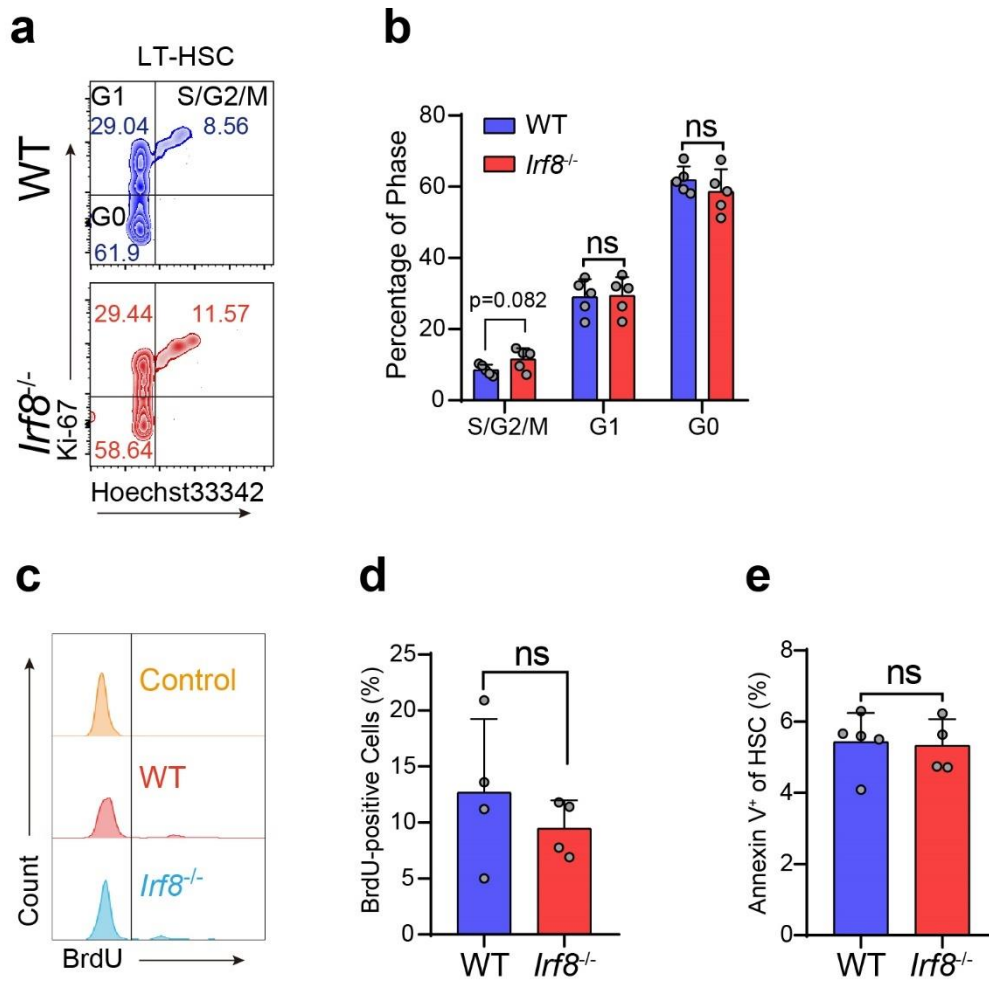


Figure S2. Loss of IRF8 did not affect the cell cycle and apoptosis of LT-HSCs. a) Representative flow-cytometric cell-cycle analysis of HSCs (LSK CD48⁻ CD150⁺) from 4-week-old WT or *Lrf8*^{-/-} mice at steady-state. G0: Hoechst33342⁻, Ki-67⁻; G1: Hoechst33342⁻, Ki-67⁺; S/G2/M: Hoechst33342⁺, Ki-67⁺. b) Percentage of cells in different cell-cycle phases (G0, G1, and S/G2/M) of HSCs. c) Representative FACS histograms of HSCs in BrdU incorporation assay. d) Percentage of BrdU-positive cells in HSCs from 4-week-old WT or *Lrf8*^{-/-} mice at steady-state, after 16 hours of BrdU incorporation. e) Apoptosis analysis of HSCs from 4-week-old WT or *Lrf8*^{-/-} mice at steady-state by Annexin V staining. Error bars, mean±s.e.m. ns, not significant, data were analyzed with unpaired Student's t-test.

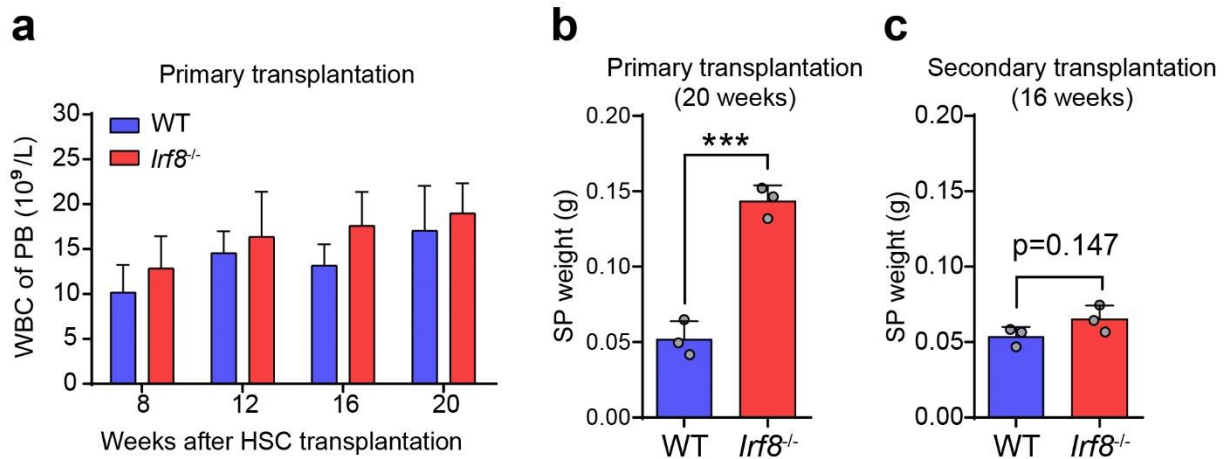


Figure S3. IRF8-deficient HSCs showed increased repopulation capacity. a) Monthly analysis of white blood cell counts in PB of primary HSC transplanted mice (WT: n=10; KO: n=10). b) Spleen weight of primary HSC transplanted mice. c) Spleen weight of secondary transplanted recipient mice.

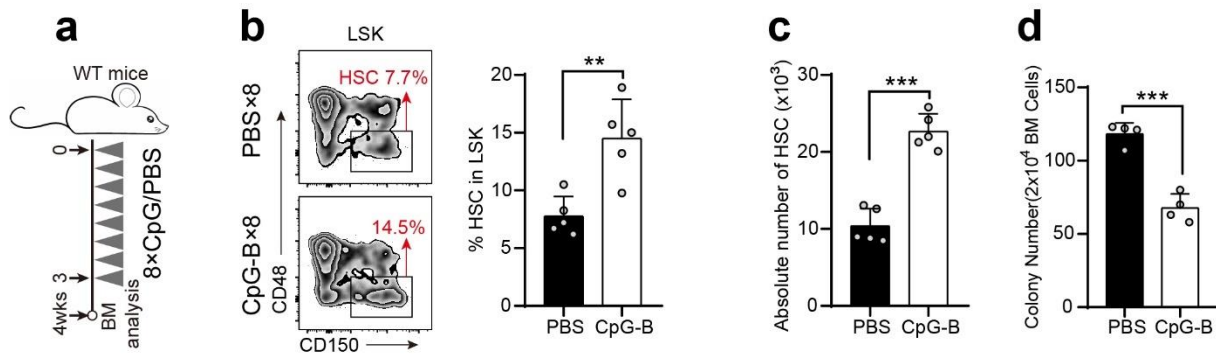
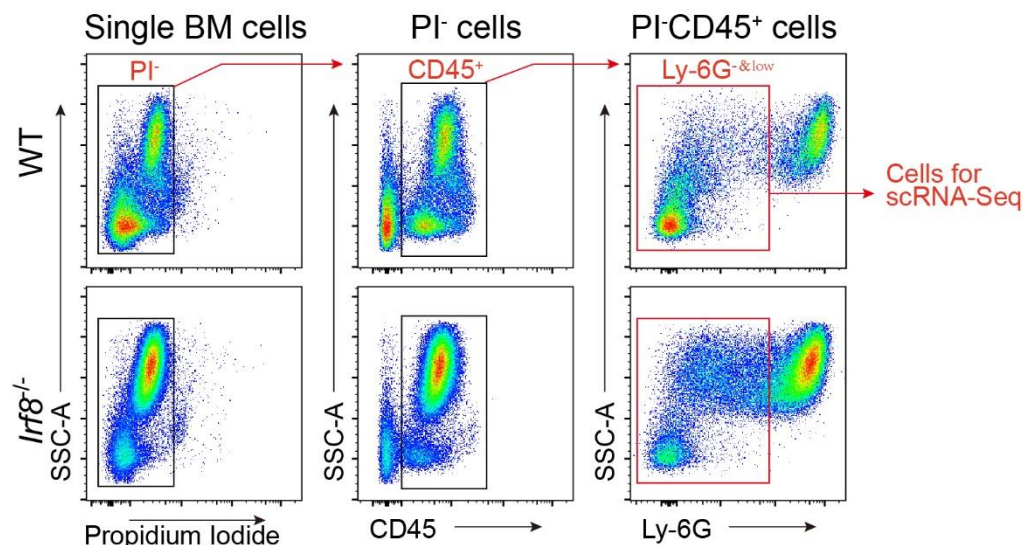


Figure S4. Long-term treatment with CpG-B increased proportions of HSCs in LSKs and absolute numbers of HSCs, while impaired the colony-forming capacity. a) Experimental scheme for long-term treatment with PBS or CpG-B. b) Representative flow plots gated on HSCs in LSKs after 8 successive doses of PBS or CpG were injected (left) and the percentages of HSCs within the LSKs (right). c) Absolute numbers of HSCs after 8 successive doses of PBS or CpG were injected. d) Colony numbers counted in the colony-forming unit (CFU) assay of total BM cells isolated from mice treated 8 times with PBS or CpG. Error bars, mean \pm s.e.m. ** $P < 0.01$, *** $P < 0.001$, data were analyzed with unpaired Student's t-test.

a

Sorting strategy of Cells for Single-Cell RNA sequencing



b

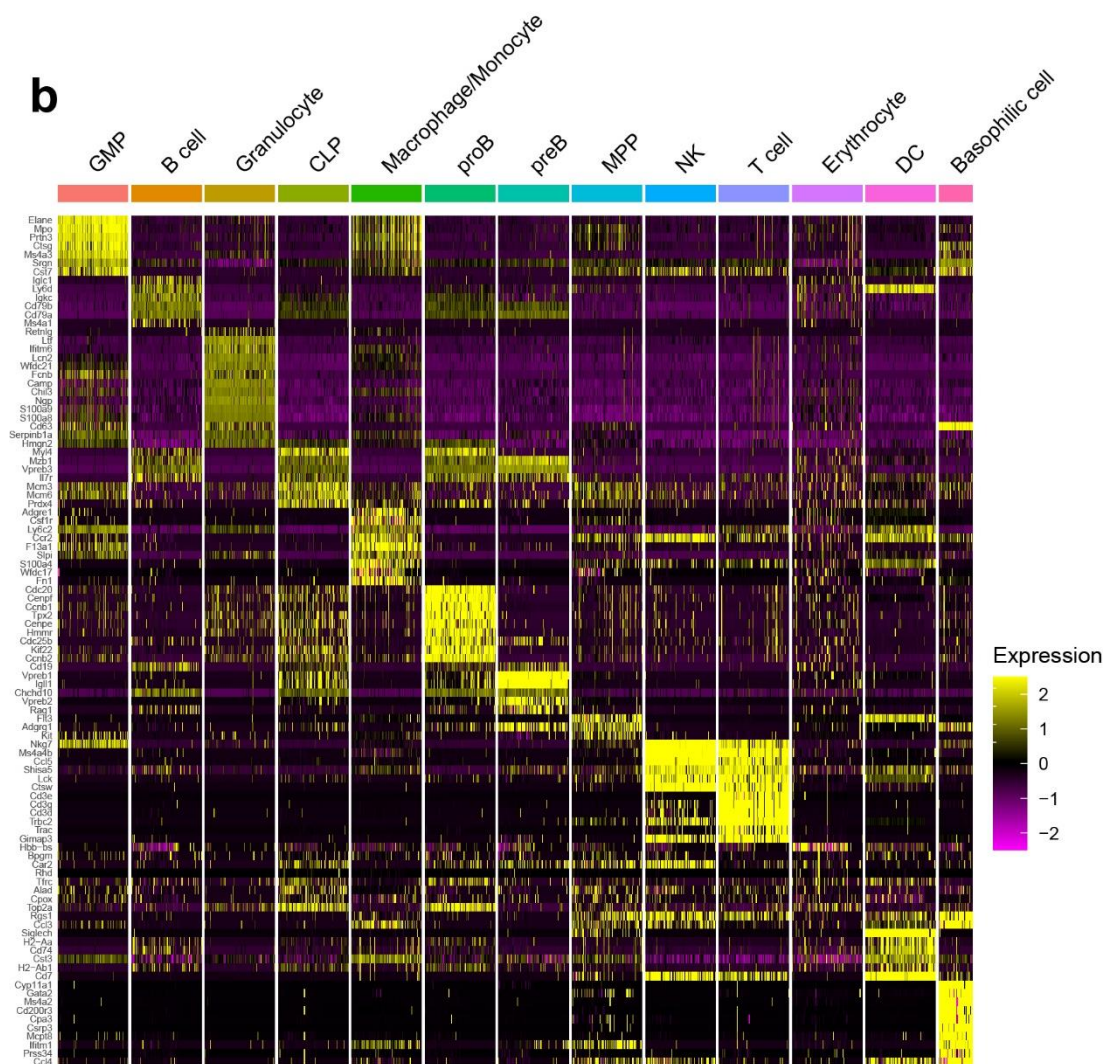


Figure S5. Single-cell RNA sequencing of *Irf8*^{-/-} Ly-6G^{hi} depleted BM cells. a) Sorting strategy for Ly-6G^{hi} & low BM cells from WT or *Irf8*^{-/-} mice by FACS for single-cell RNA sequencing. b) Unsupervised hierarchical clustering of cells based on the gene expression profiles from single-cell RNA sequencing.

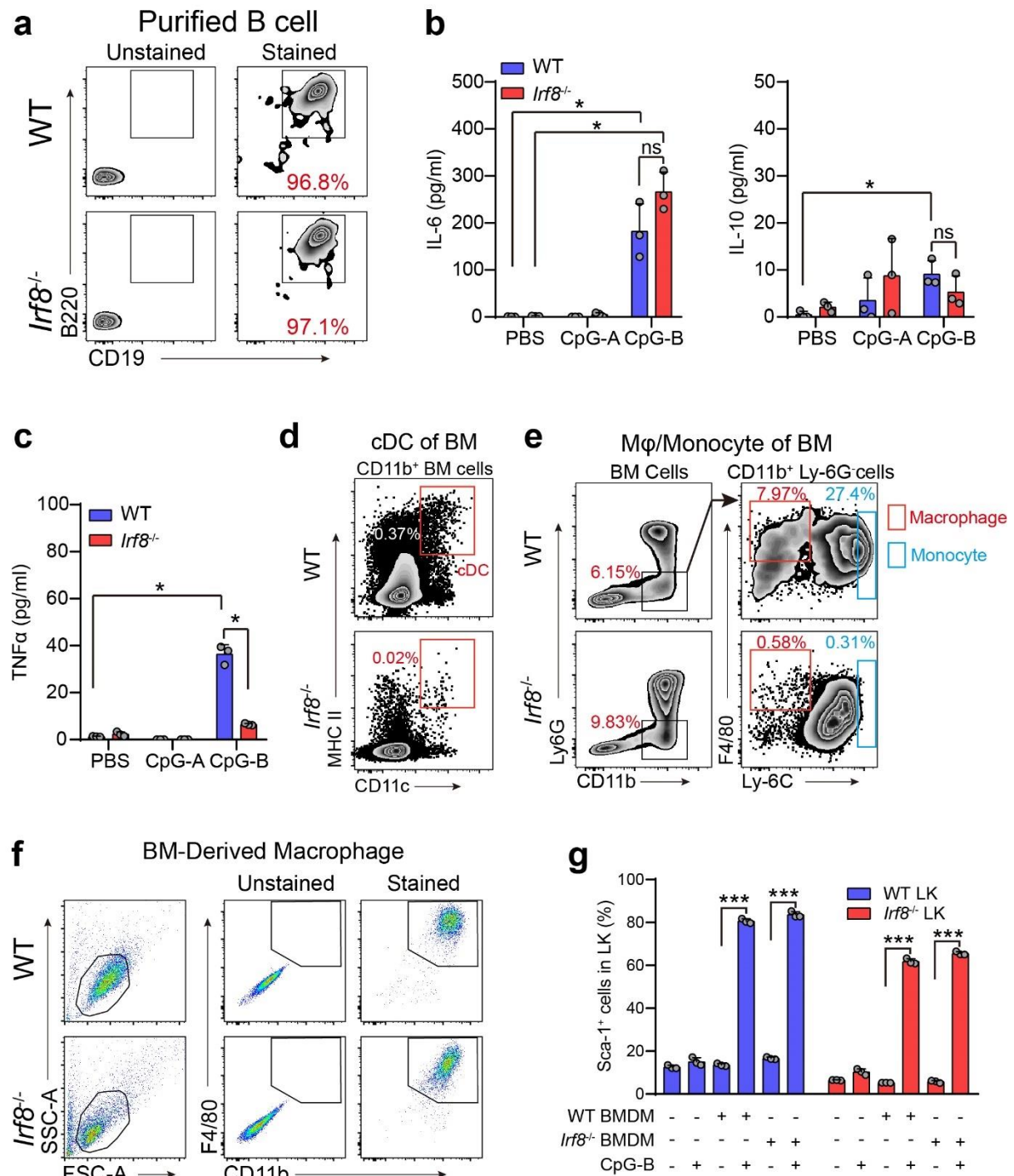


Figure S6. IRF8 is essential for TLR9 signaling in IRF8-dependent immune cells. a) The purity of B cells was verified by flow cytometry after isolation. b) The levels of IL-6, IL-10, and c) TNFα detected in purified B cell culture supernatant at 24 hours after stimulation with PBS, CpG-A or CpG-B. d) Representative flow plots gated on BM cDCs (CD11b⁺, MHCII⁺, CD11c⁺). e) Representative flow plots

gated on BM monocytes (Ly-6G⁻ CD11b⁺ Ly-6C^{hi}) and BM macrophages (Ly-6G⁻ CD11b⁺ Ly-6C⁻ F4/80⁺). f) Flow cytometric analysis of expression of F4/80 and CD11b of BMDMs generated from WT and *Irf8*^{-/-} BM. g) Proportions of Sca-1-positive cells in purified WT and *Irf8*^{-/-} LKs at 16 hours after PBS or CpG-B stimulation, co-culturing with or without WT or *Irf8*^{-/-} BMDMs *in vitro*. Error bars, mean±s.e.m. ns, no significant, *P<0.05, ***P<0.001, data were analyzed with unpaired Student's t-test.

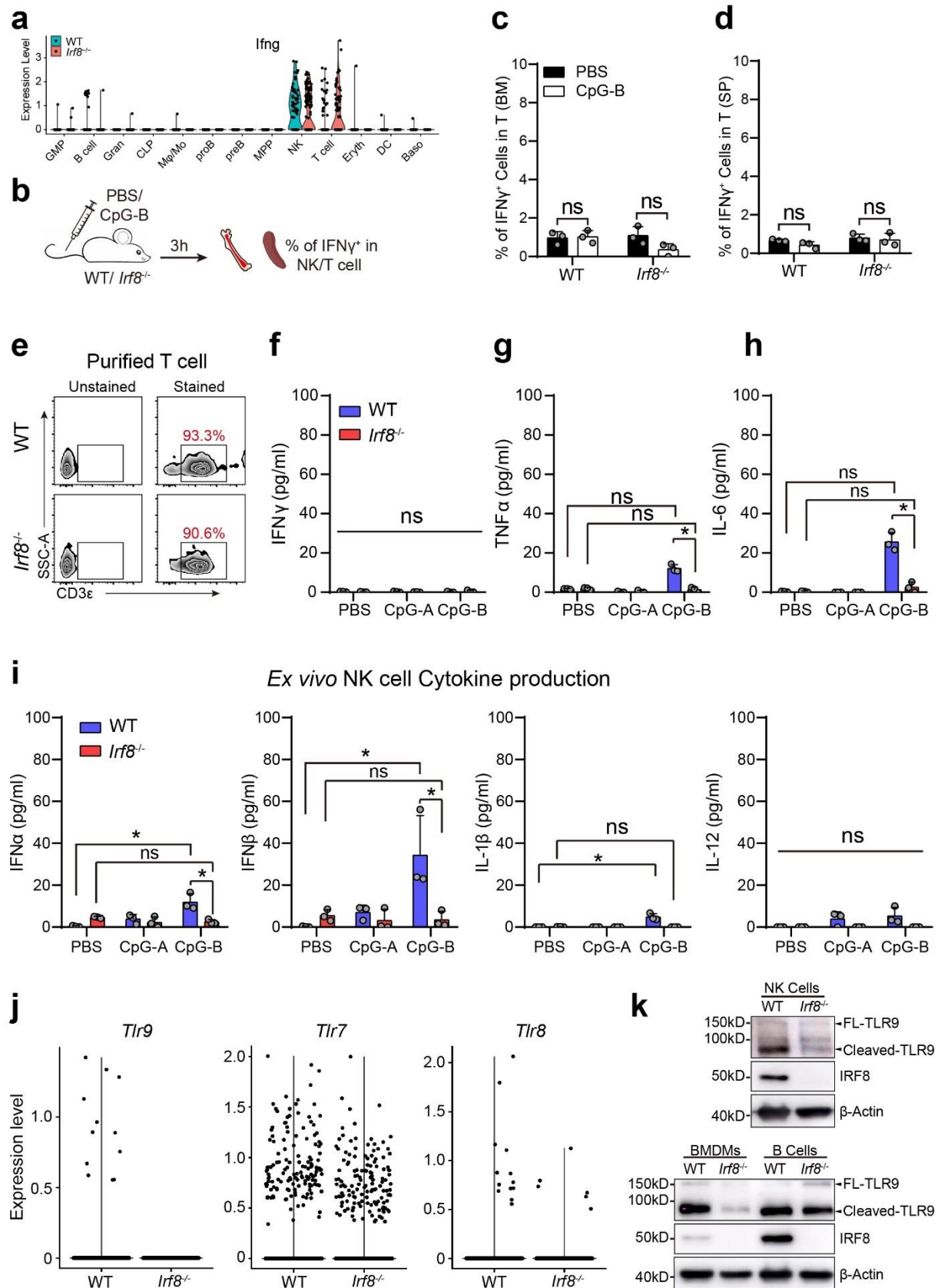


Figure S7. The TLR9 signaling pathway of BM NK cells was abolished in *Lrf8*^{-/-} mice. a) The *Ifng* expression level from single-cell RNA-seq data. b) Experimental scheme for measuring the

percentages of IFN γ ⁺ cells in T cells and NK cells by flow cytometry at 3 hours after treating WT and *Irf8*^{-/-} mice with PBS or CpG-B (i.v.). c) Percentages of IFN γ ⁺ cells in BM T cells and d) spleen T cells at 3 hours after PBS and CpG-B stimulation. e) Purified T cells were verified by flow cytometry. f-h) The production of IFN γ , TNF α , and IL-6 by WT or *Irf8*^{-/-} splenic T cells at 24 hours after PBS, CpG-A or CpG-B stimulation. i) IFN α , IFN β , IL-1 β and IL-12 production of WT or *Irf8*^{-/-} BM NK cells at 24 hours after PBS, CpG-A or CpG-B stimulation, respectively. j) The expression level of *Tlr9*, *Tlr7* and *Tlr8* of Ly-6G^{low} BM cells from single-cell RNA-seq data. k) The protein levels of TLR9 were down-regulated in NK cells, BMDMs and B cells. FL-TLR9, full-length TLR9; Cleaved-TLR9, truncated and functional TLR9. Error bars, mean \pm s.e.m. ns, no significant; *P<0.05; data were analyzed with unpaired Student's t-test.