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Telmisartan Activates PPAR δ to Improve Symptoms of Unpredictable Chronic Mild Stress-Induced Depression in Mice

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Major depression is a common mental disorder that has been established to be associated with a decrease in serotonin and/or serotonin transporters in the brain. Peroxisome proliferator-activated receptor δ (PPAR δ) has been introduced as a potential target for depression treatment. Telmisartan was recently shown to activate PPAR δ expression; therefore, the effectiveness of telmisartan in treating depression was investigated. In unpredictable chronic mild stress (UCMS) model, treatment with telmisartan for five weeks notably decrease in the time spent in the central and the reduced frequency of grooming and rearing in open field test (OFT) and the decreased sucrose consumption in sucrose preference test (SPT) compared with the paradigms. Telmisartan also reversed the decrease in PPAR δ and 5-HTT levels in the hippocampus of depression-like mice. Administration of PPAR δ antagonist GSK0660 and direct infusion of sh-PPAR δ into the brain blocked the effects of telmisartan on the improvement of depression-like behavior in these mice. Moreover, telmisartan enhanced the expression of PPAR δ and 5HTT in H19-7 cells. In conclusion, the obtained results suggest that telmisartan improves symptoms of stress-induced depression in animals under chronic stress through activation of PPAR δ . Therefore, telmisartan may be developed as a potential anti-depressant in the future.

The chronic and stressful life events are associated with the onset of major depression, which is the most prevalent psychiatric disorder with high morbidity and mortality rates¹. Efforts to reduce the prevalence of depression continue due to its public health significance. Therefore, the model of unpredictable chronic mild stress (UCMS) was developed to investigate depressive phenomena and drug treatment in animals. Clinical and experimental data have shown that the disturbances in the serotonergic system and stress play a key role in depressive disorders². Serotonin (5-HT) released from serotonergic terminals is selectively taken up from the synaptic cleft into these terminals via the serotonin transporter (5-HTT)³. In depression, the extensive degeneration of serotonergic neurons corresponds to the loss of 5-HTT⁴. Additionally, 5-HTT knockout mice show several behavioral changes, including increased anxiety-like behavior, increased sensitivity to stress, and inhibited exploratory locomotion⁵.

Peroxisome proliferator-activated receptors δ (PPAR δ), as one of the receptors in the PPAR nuclear receptor family, is a ligand-activated transcription factor. PPAR δ regulates energy metabolism and mitochondrial biogenesis in skeletal muscle⁶. PPAR δ shows a widespread brain distribution, it is least two-fold more highly expressed in brain than in muscle⁷. Recently, PPAR δ was shown to play an important role in repress stress-induced depressive behaviors⁸ in addition to the regulation of serotonin transporter expression in hippocampus⁹. Moreover, PPAR δ activation also produces neuroprotection and reverses neurodegeneration in Alzheimer's disease^{10,11}, Parkinson's disease¹² and Huntington's disease¹³. Generally, the hippocampus has been widely selected to investigate 5-HTT

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and PPAR δ expression levels, as this brain region has been strongly implicated in the cause and consequences of both depression and chronic stress¹⁴.

Telmisartan, an angiotensin II type 1 receptor blocker (ARB), is widely used to treat hypertension with the expectation of a decrease in the onset of cardiovascular and cerebrovascular disease. As the most lipophilic agent with the longest half-life among ARBs¹⁵, telmisartan is known to cross the brain-blood barrier (BBB) for blockade of central AT1 receptors¹⁶. Telmisartan was identified to play a role in neurological system. Since BBB permeability is increased due to stress¹⁷, the effect of peripherally administered telmisartan on cerebral function seems sufficient to attenuate the stress-induced cognitive decline. Telmisartan exhibited anti-apoptosis, anti-inflammatory, and antioxidant benefits in the intracerebral hemorrhage rat model¹⁸. In Parkinson's disease, telmisartan was reported to protect mouse dopaminergic neurons and inhibit the microglial response¹⁹. Telmisartan has been recently discovered to activate PPAR δ for the promotion of glucose uptake to improve insulin sensitivity and hyperglycemia-induced cardiac fibrosis^{20,21}.

In the present study, we investigated the effect of telmisartan on stress-induced depression in animals. In the UCMS mice model, the behavior performances including open field test (OFT) and the sucrose preference test (SPT) were evaluated. The effect of telmisartan and losartan, a selective AT receptor antagonist, also compared. Moreover, the expression levels of PPAR δ and 5-HTT in the hippocampus between vehicle-treated group and telmisartan-treatment group were determined in UCMS mice. To further understanding the relationship between telmisartan and PPAR δ , the expression of PPAR δ were knockdown or knockout using PPAR δ specific antagonist GSK0660 or ShRNA. Additionally, the effects of telmisartan on expressions of PPAR δ and 5-HTT were further studied in the H19-7 cell line.

Results

Telmisartan ameliorated anxiety- and depression-like behavior in UCMS mice. The open field test was performed to evaluate locomotion and anxiety-like behaviors; stress affected the time spent in the center and the periphery. Reduced time spent in the OFT center is used to measure the anxiety-like behavior. Mice that received daily oral administration of telmisartan showed a significant increase in the total distance travelled [$F_{(4,35)} = 62.311$, $P < 0.05$; Fig. 1A] and time spent in the central and was more likely to explore the environment than the vehicle-treated group [$F_{(4,35)} = 84.202$, $P < 0.05$; Fig. 1B]. The number of grooming [$F_{(4,35)} = 24.598$, $P < 0.05$] and rearing behavior [$F_{(4,35)} = 69.947$, $P < 0.05$] were also markedly suppressed in the UCMS group [Fig. 1C,D]. Additionally, mice with UCMS showed less preference in sucrose intake, but the sucrose consumption was reversed in both telmisartan- and fluoxetine-treated groups [$F_{(4,35)} = 16.071$, $P < 0.05$, Fig. 1E]. Moreover, in stress condition, the telmisartan produced significantly different results than that of the losartan, a selective AT receptor antagonist ($p < 0.05$). Overall, telmisartan showed a slightly smaller anti-depressant ability than fluoxetine, which is a selective serotonin reuptake inhibitor (SSRI) used to treat depression in clinics.

Furthermore, the PPAR δ and 5-HTT expression levels in the hippocampus were significantly reduced in mice receiving UCMS. Chronic administration of telmisartan reversed the UCMS-induced changes in hippocampal PPAR δ [$F_{(4,25)} = 8.180$, $P < 0.05$, Fig. 1E] and 5-HTT expressions [$F_{(4,25)} = 6.662$, $P < 0.05$, Fig. 1F] seen with Western blots. Losartan treatment did not affect PPAR δ and 5-HTT expressions in UCMS mice. These results indicated that telmisartan effectively attenuates the adverse effect of stress on depression-like behavior, and this action seems different with other AT1 blockade, associated with increased PPAR δ activation in the hippocampus.

PPAR δ antagonist inhibited the anti-depression-like effect of telmisartan in UCMS mice. For further investigation of the interaction between telmisartan and PPAR δ in depression, PPAR δ antagonist GSK0660 was administered into UCMS mice. Compared to the improvement in behavior observed in the telmisartan group, the total distance travelled [$F_{(5,42)} = 49.285$, $P < 0.05$], the time spent in the central [$F_{(5,42)} = 46.881$, $P < 0.05$], novelty-seeking behavior including rearings [$F_{(5,42)} = 18.898$, $P < 0.05$] and grooming [$F_{(4,35)} = 26.573$, $P < 0.05$] and percentile of sucrose preference [$F_{(5,42)} = 54.628$, $P < 0.05$] in the UCMS group co-treated with telmisartan and GSK0660 was markedly decreased. However, the results also showed that the administration of GSK0660 only in the UCMS groups did not further deteriorate the depression-like behavior (Fig. 2A,B,C,D,E).

Compared to the control group, the expression of hippocampal PPAR δ was significantly reduced in mice receiving UCMS, and this was reversed by telmisartan. Once PPAR δ was blocked by GSK0660, the effect was extinguished [$F_{(5,42)} = 6.349$, $P < 0.05$]. Likewise, a similar change in hippocampal 5-HTT expression [$F_{(5,42)} = 5.864$, $P < 0.05$] was observed [Fig. 2F,G]. Additionally, the treatment of GSK0660 decreased 5-HTT expression but did not affect behavior performances in control group. Taken together, the anti-depressive effect of telmisartan is likely to be PPAR δ -dependent. Additionally, administration of GSK0660 alone in mice receiving UCMS did not cause any changes in behavioral performance or expression levels compared to that observed in the vehicle-treated group.

The anti-depressive effect of telmisartan disappeared in mice with hippocampal PPAR δ knock-down. Then, we knocked down the expression of PPAR δ using shRNA constructs in control and UCMS groups respectively. On the 7th day after the transfection of shRNA into the brain, decreased PPAR δ expression in the hippocampus was identified. One week later, mice were treated following the procedure noted above. Changes in behavioral performance were then compared with that in animals injected with a scramble control. Mice with PPAR δ knockdown in hippocampus displayed a significantly decreased the total distance travelled and the time spent in the central. When PPAR δ is silenced in control mice, telmisartan could not reverse the inactivated behavior (Fig. 3A,B,C,D,E). Two-way ANOVA indicated telmisartan effects were dependent of the stress condition [effect of group, $F_{(4,70)} = 81.419$, $P < 0.05$; effect of stress, $F_{(1,70)} = 338.827$, $P < 0.05$; effect of group-by-stress interaction, $F_{(4,70)} = 53.853$, $P < 0.05$]. Moreover, telmisartan increased the grooming and rearing in the OFT and the sucrose consumption in the SPT in the scramble group but not in that of the PPAR δ knockdown animals

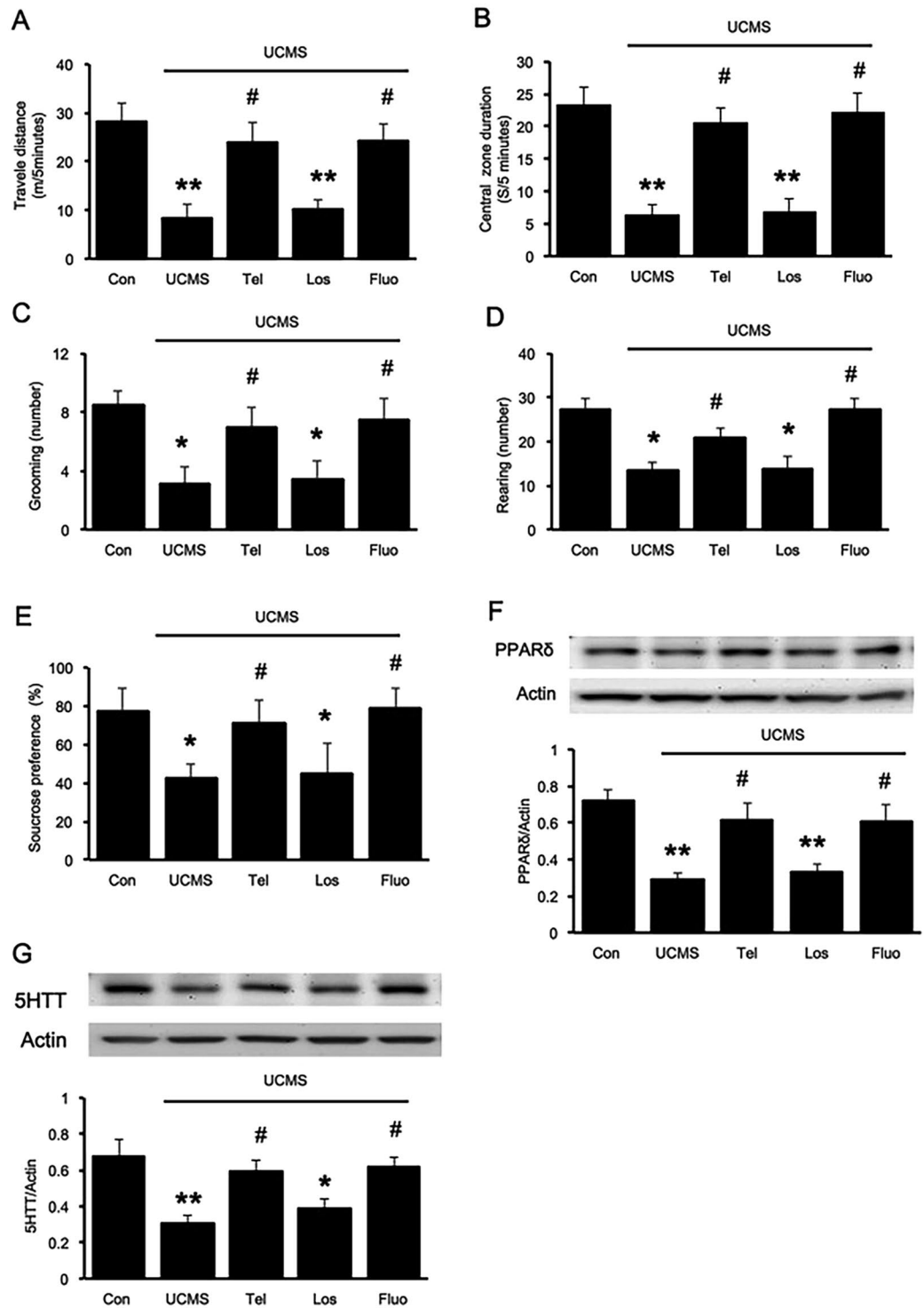


Figure 1. Telmisartan ameliorated the depression-like behavior through the PPAR δ pathway in mice receiving UCMS. The animals were allowed to explore the open field arena for 5 min. **(A)** The total distance travelled; **(B)** the time spent in the central; **(C)** frequency of grooming; **(D)** frequency of rearing; **(E)** the percentage of sucrose solution consumed (%) in the sucrose preference test (SPT); **(F)** PPAR δ expression levels in the hippocampus; **(G)** 5-HTT expression levels in the hippocampus. Data are presented as the mean \pm SEM ($n = 8$ per group). * $p < 0.05$ and ** $p < 0.01$ compared to the corresponding control; # $p < 0.05$ and ## $p < 0.01$ compared to the UCMS group. Full-length blots are presented in Supplementary Fig. S1.

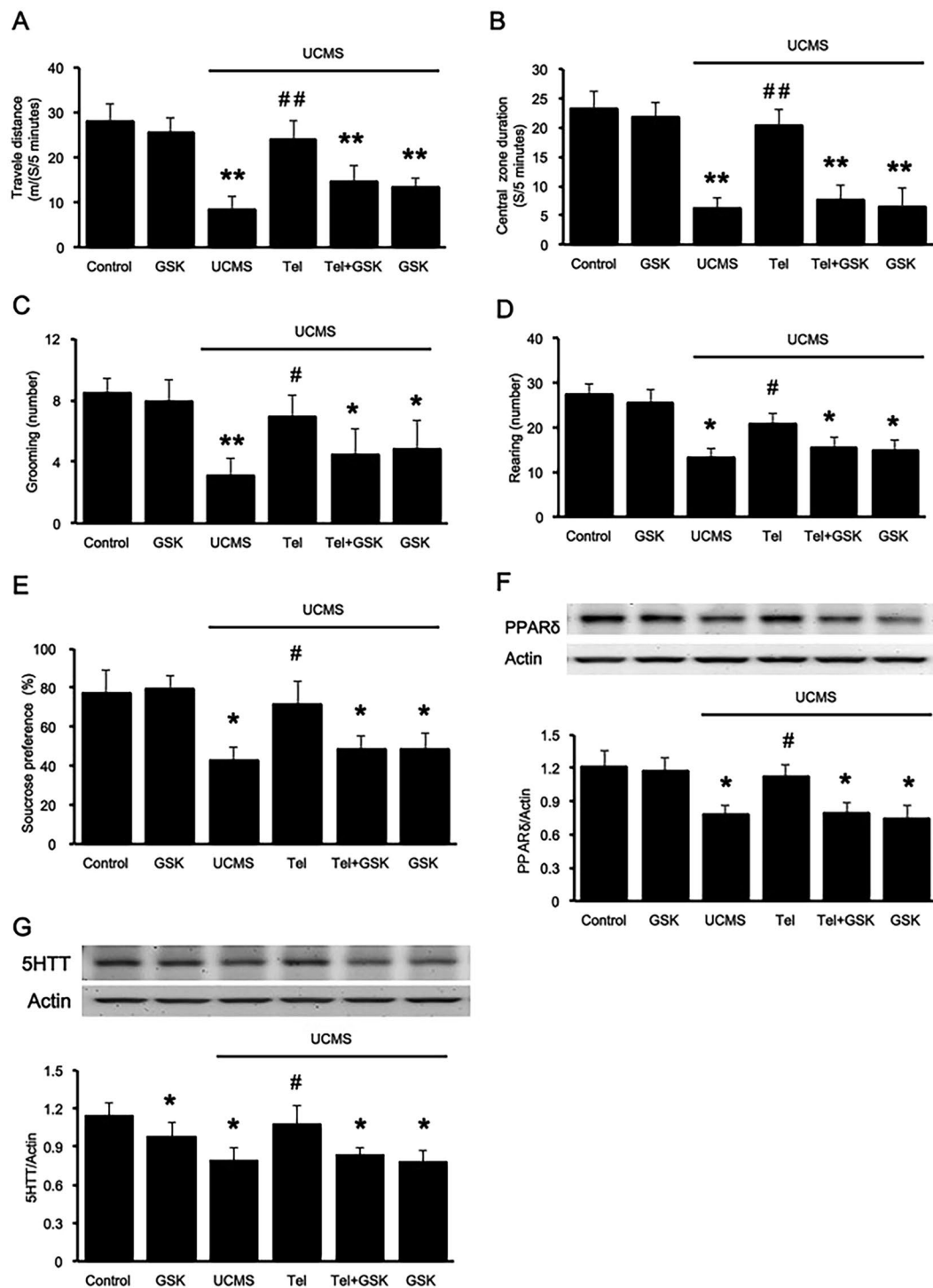


Figure 2. The PPAR δ antagonist inhibited the anti-depression-like action of telmisartan in mice receiving UCMS. The PPAR δ antagonist, GSK0660 (5 nmol.L⁻¹, i.c.v), was administered 30 min before telmisartan treatment. In the open field test, the animals were allowed to explore the open field arena for 5 min. (A) the total distance travelled; (B) the time spent in the central; (C) frequency of grooming; (D) frequency of rearing; (E) the percentage of sucrose solution consumed (%) in the sucrose preference test (SPT); (F) PPAR δ expression levels in the hippocampus of the various treatment groups; (G) 5-HTT expression levels in the hippocampus of the various treatment groups. Data are presented as the mean \pm SEM (n = 8 per group). *p < 0.05 and **p < 0.01 compared to the corresponding control; #p < 0.05 and ##p < 0.01 compared to the UCMS group. Full-length blots are presented in Supplementary Fig. S2.

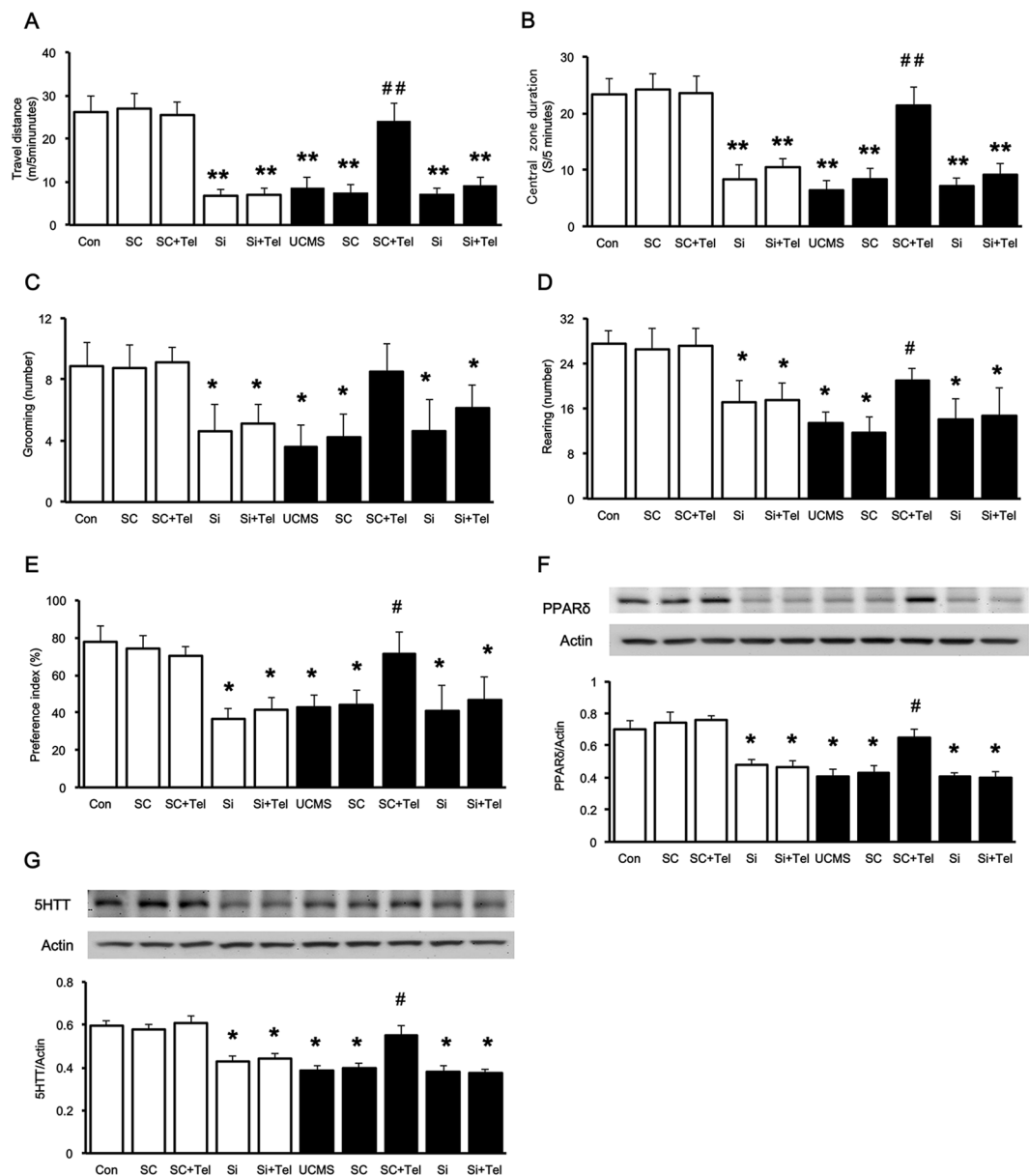


Figure 3. Changes of telmisartan-induced effects in the mice received hippocampal PPAR δ knockdown. Normal mice and UCMS mice were transfected with scrambled shRNA or PPAR δ shRNA, respectively. In the open field test, the animals were allowed to explore the open field arena for 5 min. (A) The total distance travelled; (B) the time spent in the central; (C) frequency of grooming; (D) frequency of rearing; (E) the percentage of sucrose solution consumed (%); (F) PPAR δ expression levels in the hippocampus; (G) 5-HTT expression levels in the hippocampus. White columns indicated normal control groups, and black columns indicated UCMS groups. Data are presented as the mean \pm SEM ($n = 8$ per group). * $p < 0.05$ and ** $p < 0.01$ compared to the corresponding control; # $p < 0.05$ and ## $p < 0.01$ compared to the UCMS group. Full-length blots are presented in Supplementary Fig. S3.

only in UCMS mice [grooming: effect of group, $F_{(4,70)} = 15.791$, $P < 0.05$; effect of stress, $F_{(1,70)} = 23.421$, $P < 0.05$; effect of group-by-stress interaction, $F_{(4,70)} = 10.695$, $P < 0.05$; rearing: effect of group, $F_{(4,70)} = 22.557$, $P < 0.05$; effect of stress, $F_{(1,70)} = 65.24$, $P < 0.05$; effect of group-by-stress interaction, $F_{(4,70)} = 10.155$, $P < 0.05$; and glucose consumption: effect of group, $F_{(4,70)} = 15.345$, $P < 0.05$; effect of stress, $F_{(1,70)} = 60.92$, $P < 0.05$; effect of group-by-stress interaction, $F_{(4,70)} = 8.528$, $P < 0.05$].

Furthermore, the reduction in the PPAR δ levels in the hippocampus of scramble mice was reversed by telmisartan only in UCMS group [effect of group, $F_{(4,50)} = 13.325$, $P < 0.05$; effect of stress, $F_{(1,50)} = 79.649$, $P < 0.05$; effect of group-by-stress interaction, $F_{(4,50)} = 3.232$, $P < 0.05$; Figs 3E, 4E]. Similar to the changes in PPAR δ , the lower 5-HTT levels in the scramble group were also reversed after telmisartan treatment [effect of group, $F_{(4,50)} = 15.277$, $P < 0.05$; effect of stress, $F_{(1,50)} = 35.710$, $P < 0.05$; effect of group-by-stress interaction, $F_{(4,50)} = 3.589$, $P < 0.05$; Figs 3F, 4G]. However, telmisartan failed to restore the PPAR δ and 5-HTT expression levels in the PPAR δ

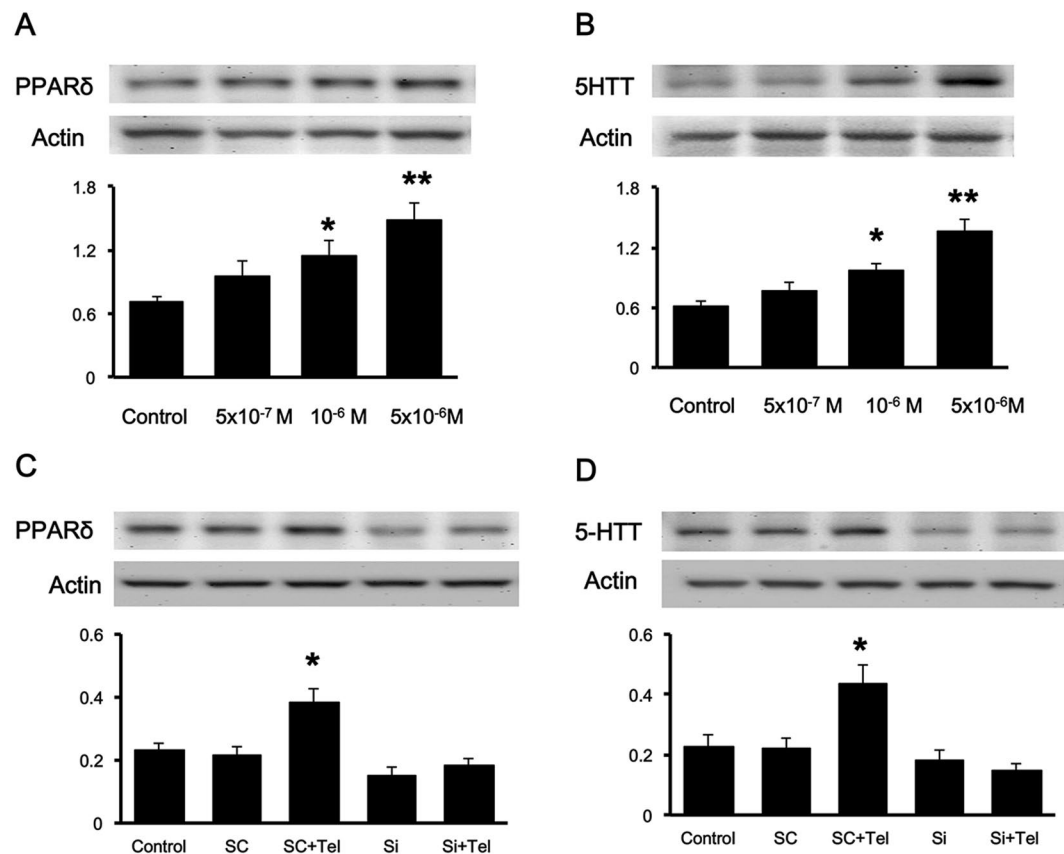


Figure 4. Effect of telmisartan on PPAR δ and 5-HTT expression levels in cultured H19-7 cells. Cultured hippocampal H19-7 cells were incubated with different doses of telmisartan at 5×10^{-7} M, 10^{-6} M, and 5×10^{-6} M for 24 h. (A) PPAR δ expression level and (B) 5-HTT expression level. Cells were transfected with scramble shRNA (Sc) or PPAR δ shRNA (Si) for 24 h and then treated with 5×10^{-6} M telmisartan for 24 h. (C) PPAR δ expression level and (D) 5-HTT-expression level. Each column represents the mean \pm SEM ($n = 6$). * $p < 0.05$ and ** $p < 0.01$ compared to the corresponding control. Full-length blots are presented in Supplementary Fig. S4.

Groups	Stage for Treatment	
	Before	After
Control	84.92 \pm 2.94	84.04 \pm 2.32
UCMS + Vehicle	82.54 \pm 2.03	85.88 \pm 2.95
UCMS + Telmisartan	81.42 \pm 1.67	81.67 \pm 2.88

Table 1. Changes in systolic blood pressure of mice receiving telmisartan treatment. Data (mean \pm SEM, $n = 8$) show the value of systolic blood pressure (mmHg). Telmisartan was treated at $1 \text{ mg} \cdot \text{kg}^{-1}$ and the solvent used to dissolve telmisartan was treated at same volume in vehicle-treated group.

knockdown mice with or without UCMS. It indicated that PPAR δ deficiency correlated with the development of depression. The effects of telmisartan in the improvement of depression induced by UCMS seem to be associated with the selective enhancement of PPAR δ and 5-HTT expression levels.

Effect of telmisartan on blood pressure in UCMS mice. The mean systolic blood pressures in the vehicle-treated normal control group, telmisartan-treated normal group, UCMS + vehicle control group, and UCMS + telmisartan group were calculated at before, and the end of telmisartan treatment. As shown in Table 1, no statistical difference ($P > 0.05$) can be obtained in telmisartan-treated group at before and after stage, as that in vehicle-treated control (Table 1). Therefore, the behavioral effect of telmisartan seems to not be related to its antihypertensive action.

Telmisartan increased PPAR δ and 5-HTT expressions in the H19-7 cell line. To understand the direct effect of telmisartan on PPAR δ and 5-HTT expression levels, H19-7 cells were treated with telmisartan ($3 \times 10^{-7} \text{ mol} \cdot \text{L}^{-1}$, $10^{-6} \text{ mol} \cdot \text{L}^{-1}$ and $3 \times 10^{-6} \text{ mol} \cdot \text{L}^{-1}$) or vehicle (control) for 24 h. The results showed that telmisartan increased PPAR δ expression in a concentration-dependent fashion (Fig. 4A). Increased expression of

5-HTT was also observed in cells treated with telmisartan in the same manner (Fig. 4B). However, 5-HTT levels were not increased in PPAR δ -silenced cells, even when they were treated with telmisartan (Fig. 4C,D). These results show that PPAR δ is able to regulate 5-HTT expressions. Therefore, telmisartan failed to affect 5-HTT expressions when PPAR δ was absent in hippocampal cells.

Discussion

In the present study, we found that telmisartan is useful for alleviating the symptoms of depression. BALB/c mice are known to exhibit depressive-related behaviors when subjected to selected stress paradigms, offer much promise for the study of the stress response, and are good models for depression and the antidepressant treatment response in humans²². UCMS has been widely used in mice to mimic a depression-like disorder and is recognized as a reliable model of depression in humans²³. Mice were exposed to UCMS and exhibited significant depressive behaviors, as shown by the decreased locomotor activity and suppressed grooming and rearing behavior in the OFT and reduced sucrose intake in the SPT. Additionally, food and water deprivation were applied in SPT, as described previously^{24,25}. Chronic treatment with telmisartan significantly ameliorated the depressive-like behaviors in the chronic stress mice. Telmisartan effectively reversed the changes in the locomotor activity and frequency of grooming and rearing in the OFT and increased sucrose intake in the SPT. We have previously conducted some preliminary experiments and found no effect of telmisartan itself in normal mice. However, the anti-depressive action of telmisartan was still less than that of fluoxetine. In addition, telmisartan at the treated dosage did not affect blood pressure in mice that received UCMS. This is consistent with a previous report that reported that telmisartan at a non-hypotensive dose had beneficial effects on cognitive impairment²⁶. A non-hypotensive dose of telmisartan preferentially promoted the expression of 5-HTT by activating PPAR δ . It possibly because the wide distribution of PPAR δ receptors in the brain, particularly the hippocampus.

Moreover, inhibition of brain AT1 receptor activity was approved to reduce stress responses and anxiety²⁷. ARBs might have neuroprotective effect in addition to slow the progression of Alzheimer's disease²⁸. The antidepressant-like effect of ARBs has also been previously reported in depressed patients²⁹. Additionally, our data showed that telmisartan was more effective on ameliorating depression-like behaviors in UCMS mice as compared with losartan. It means that telmisartan might act different with the other ARBs in depression treatment. Telmisartan can block cerebral AT1 receptors, which have a higher expression in the brain³⁰. In contrast to classical ARBs, telmisartan is able to cross BBB³¹. Peripheral administration of losartan was less effective on cognitive function compared with telmisartan in diabetic mice³². However, directly injection of losartan into the amygdala showed anxiolytic-like effect in acute stressed rats³³. Therefore, our results indicated that the absence of efficacy of losartan to counteract the UCMS effect is also probably due to the lack of BBB permeability.

Furthermore, ligands of PPAR δ are known to interfere with 5-HTT signaling³⁴. After a UCMS challenge, PPAR δ and 5-HTT expression levels were both markedly reduced in the hippocampus as shown in the present study, which is in agreement with previous reports^{34,35}. The levels of PPAR δ and 5-HTT were effectively up regulated by telmisartan in the model group, indicating that the antidepressant-like action of telmisartan seems to be related to increased expression of PPAR δ or 5-HTT. Notably, the amelioration of the behavioral performance in the OFT and SPT, as well as hippocampal PPAR δ and 5-HTT expression levels, induced by telmisartan was inhibited by a PPAR δ antagonist, GSK0660. Furthermore, down regulation of PPAR δ alone reproduced the phenotypes of the UCMS as indicated in our previous study⁹. In PPAR δ knockdown mice with UCMS, the depression-like behavior was significantly higher than those mice infected with the scrambled shRNA. The anti-depressive effect of telmisartan also disappeared once hippocampal PPAR δ s were silenced. Therefore, PPAR δ is involved in the behavioral performance of mice, and telmisartan can promote PPAR δ expression to improve depression-like behaviors.

PPAR δ shows a relatively high neuronal expression compared with that of the other PPAR subtypes, both PPAR α and PPAR γ ³⁶. It has been mentioned that activation of PPAR δ induces oligodendrocyte differentiation and enhances neuronal differentiation in the peripheral nervous system^{37,38}. Additionally, a decrease in 5-HTT activity in the presynaptic membrane has been identified in depression patients³⁸. Furthermore, depressive behaviors have also been observed in 5-HTT-knockout mice³⁹. 5-HTT is one of the major modulators of 5-HT neurotransmission as it determines the magnitude and duration of 5-HT signaling. In the present study, changes in 5-HTT expressions were associated with PPAR δ expression in both stress-induced depression mice and PPAR δ -silenced mice. Therefore, PPAR δ seems to interact with 5-HTT in the hippocampus. In cultured H19-7 hippocampus cells, we also demonstrated that PPAR δ activation by telmisartan could increase 5-HTT expressions.

Chronic stress is associated with oxidative stress and promotes the production of reactive oxygen species, resulting in impaired function in the central nervous system⁴⁰. Telmisartan as a unique ARB with a partial PPAR δ agonistic property; has been shown to be neuroprotective and improve cognitive decline by reducing the levels of interleukin and TNF- α ⁴¹⁻⁴³. A recent study indicated that treatment with telmisartan reduced the levels of proinflammatory mediators and ameliorated the depression-like behaviors in diabetes-induced depression rats⁴⁴. In addition, PPAR δ activation in hypertensive rats is considered to contribute to the protection against cognitive decline, although it also up-regulates the expression of brain-derived neurotrophic factor in the hippocampus⁴⁵. These findings probably explain why oral administration of the PPAR δ antagonist GSK0660 or administration of PPAR δ shRNA into the brain completely blocked the actions of telmisartan both *in vivo* and *in vitro*.

Conclusions

Overall, we provided promising and novel evidence that up-regulating hippocampal PPAR δ by telmisartan results in an anti-depressive effect through the elevation of 5-HTT expressions. Our study suggests that hippocampal PPAR δ is an important therapeutic target for depression. Telmisartan could be used for the development of treatments for depressive disorders in clinics.

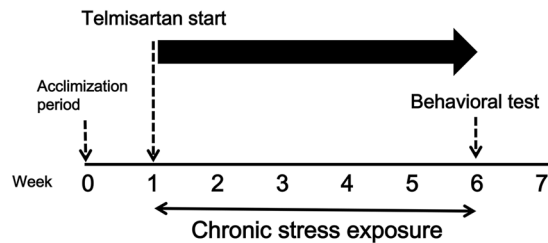


Figure 5. Timeline of experimental procedures.

Materials and Methods

Animals. A total of 142 male BALB/c mice weighing 22–30 g were obtained from the National Animal Center (Taipei, Taiwan) and maintained in the animal center of Chi Mei Medical Center (Tainan, Taiwan). The animals were housed 3–4 mice per cage on a 12/12-hr light/dark cycle with ad libitum access to food and water except during behavioral tests. Mice were introduced to the experiment room at least 1 h before the behavioral tests. This project was approved by the Institutional Animal Care and Use Committee of Chi Mei Medical Center (No. 105111531). All of the animal procedures were performed according to the Guide for the Care and Use of Laboratory Animals published by the US National Institutes of Health (NIH Publication No. 85-23, revised 1996).

Treatment schedule. This study was conducted according to the experimental protocols described in Fig. 5.

To investigate the effect of telmisartan on depression-like behavior in UCMS mice, mice were randomly divided into five groups ($n = 8$): control group, UCMS model group, UCMS + telmisartan (1 mg.kg^{-1}) group²⁶, UCMS + losartan (1 mg.kg^{-1}) group, UCMS + fluoxetine (20 mg.kg^{-1}) group⁴⁶.

To investigate the role of telmisartan in PPAR δ antagonist treated UCMS mice, mice were randomly divided into six groups ($n = 8$): control group, control group + GSK0660 (10 mg.kg^{-1}), UCMS model group, UCMS + telmisartan (1 mg.kg^{-1}) group, UCMS + GSK0660 (10 mg.kg^{-1}) + telmisartan (1 mg.kg^{-1}) group, and UCMS + GSK0660 (10 mg.kg^{-1}).

To investigate the role of the role of telmisartan in PPAR δ knockdown mice mice were randomly divided into 10 groups ($n = 8$): control, control + scramble, control + scramble + telmisartan, control + PPAR δ shRNA, control + PPAR δ shRNA + telmisartan, UCMS, UCMS + scramble, UCMS + scramble + telmisartan, UCMS + PPAR δ shRNA, UCMS + PPAR δ shRNA + telmisartan.

After an adaptation period (week 0 to week 1), mice were exposed to UCMS for 5 weeks (week 1 to week 6). Telmisartan, losartan and fluoxetine was administered 30 min by intragastric gavage before behavioral test and/or chronically 30 min before UCMS procedure to stressed as well as to unstressed control mice; while GSK0660 was administered by intraperitoneal injection, 30 min before drug treatment. All the animals were treated with respective drugs from week 1 to week 6. Behavioral testing was done in independent groups of mice in the week 7; all mice were subjected to one test daily, always in the same sequence. Blood pressure was measured every week during the experiment. Finally, all mice were sacrificed by cervical dislocation, and each hippocampus was removed, immediately frozen in liquid nitrogen, and kept at -80°C for protein assays.

The establishment of the depression-like mouse model. The UCMS model was used to explore depressive-like behaviors in mice as described previously^{47,48}. Experimental mice ($n = 8$ per group) were exposed to unpredictable mild stressors randomly every day in one week. The stressors applied included the following: water deprivation (24 h), food deprivation (24 h), reversed light/dark cycle (24 h), overnight illumination (12 h), soiled cage (12 h), and cage tilt (18 h, 45°). Each stressor was randomly assigned two or three times over a 5-week period. Stressors continued to be applied during the testing phase, except on testing days to avoid effects of acute stress. The non-stressed control mice were housed in groups (3–4 per cage), and the stressed mice were singly housed⁴⁹. At least 12 h of rest was provided between a stressor and a test⁵⁰. All of the procedures were organized in a random in order to ensure the unpredictable characteristic of the experiment.

Behavioral testing. *OPT.* Mice were placed in an open field area made of a $70 \times 70 \times 40$ cm wooden box and equipped with an infrared floor to measure locomotor activity. The arena was subdivided into a central and a peripheral zone. Mice were placed in the open field boxes for 5 min under normal light conditions, and the locomotor activity of mice was automatically scored with a camera connected to a computerized system (Viewpoint, Lyon, France). Individual animals were gently placed in the same corner of the apparatus in all trials. Time stay in central, rearing (number of times the mice stood on their hind legs), grooming (total seconds of the mice spent licking or scratching itself) and excretion were observed⁵¹.

SPT. SPT is widely used to measure the anhedonic response, which was defined as a reduction in sucrose preference relative to baseline levels^{52,53}. The mice were exposed to bottles, the one containing 1% sucrose and the other containing tap water for 24 h. After the deprivation of food and water overnight²⁴, mice were used to receive the bottle of 1% (w/v) sucrose or the bottle of tap water for 1 hour. Then, the sucrose preference was evaluated according to the formula: sucrose preference = [sucrose intake/(sucrose intake + water intake)] \times 100, as described previously⁵³.

Blood pressure measurement. To investigate the possible effect of telmisartan on blood pressure in depression-like mice, the systolic blood pressures were measured in mice received telmisartan treatment and others using the tail-cuff method by a sphygmomanometer without animal heating (Muromachi Kikai Co., Ltd., Tokyo, Japan). The blood pressure of mice under anesthesia was measured at 15-min intervals. Each value was calculated as the average of 3 measurements.

Intracerebroventricular (ICV) injection. Mice were held in a towel with the dummy cannula to inject the testing agent as described previously⁵⁴. Mice were anesthetized with a mixture of isoflurane in oxygen (2%) and placed in a Kopf stereotaxic instrument equipped with blunt ear bars. A dummy cannula was placed into the guide⁵⁵. Mice were allowed to recover for 7 days.

The infusion cannula (22 gauge), attached to PE-10 tubing, was inserted into the guide cannula and extended 0.5 mm beyond the guide. A 10.0- μ l Hamilton syringe was used to manually deliver saline or drugs over a two-minute period⁵⁶. The infusion cannula was kept in place for an additional 1 min following infusion.

Moreover, the solution containing shRNA specific to PPAR δ (Gene ID 25682) with an expression vector (pCMV6-Entry) was administered via ICV injection into mice using 25 μ l of the prepared solution (0.12 μ g μ l⁻¹), while mice receiving a similar injection of an empty vector at the same volume were used as a control.

Cell Cultures. Rat-derived hippocampus H19-7 cell line cells (CRL-2526; American Type Culture Collection, Manassas, VA) were maintained at 37 °C and 5% CO₂ in Dulbecco's modified Eagle's medium (DMEM; HyClone, South Logan, UT, USA) with 4 mM l-glutamine that was adjusted with sodium bicarbonate (1.5 g/L), glucose (4.5 g/L), G418 (200 μ g/mL), and puromycin (1 μ g/mL) and supplemented with 10% fetal bovine serum⁵⁷. Cells (1 \times 10⁶) were plated on 60-mm culture dishes, and at 80% confluence, they were differentiated by culturing for 6–7 days in DMEM containing 2% fetal bovine serum. Medium was changed every other day.

Western Blotting Analysis. Western blotting analysis was performed as previous⁵⁸. Total protein lysates from mouse hippocampus or cells were extracted in lysis buffer (1% Triton X-100, 150 mM NaCl, 10 mM Tris [pH 7.5] and 5 mM ethylenediaminetetraacetic acid), containing a protease and phosphatase inhibitor cocktail (Sigma-Aldrich, MO, USA). The protein concentration was determined with the BCA assay kit (Pierce Biotechnology, Rockford, IL, USA). The following primary antibodies were used at 4 °C overnight: anti-PPAR δ (1:1000) (Abcam, Cambridge, UK); anti-5-HTT (1:1000) (Merck Millipore, Darmstadt, Germany); anti- β actin (1:5000) (Merck Millipore) was used as an internal control. The next day, the blots were incubated with a 1/5000 dilution of horseradish peroxidase-conjugated secondary antibodies at 25 °C for 1 h. Protein bands were visualized using the enhanced chemiluminescence kit (PerkinElmer, Boston, MA, USA). The optical densities of the bands were determined using software (Gel-Pro Analyzer version 4.0 software; Media Cybernetics Inc., Silver Spring, MD, USA).

Statistical analysis. Data were expressed as the mean-standard error of the mean. Statistical analyses one-way ANOVA was used to investigate the differences between groups with pharmacological treatments. Among multiple groups were analyzed by two-way ANOVA with “stress” and “drugs” are the factors to evaluate data in the knockdown experiments. If an interaction and/or main effect were observed, pairwise comparisons following ANOVA were made using Bonferroni post-hoc test. Data sets of two sample groups were analyzed using independent Student's t-tests. All analyses were carried out by SPSS, version 21. Statistical differences were accepted at $p < 0.05$.

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Author Contributions

Y.L., K.C.C. and K.F.L. designed the study and performed experiments. Y.L., K.C.C. and K.F.L. managed the literature searches and analyses. Y.L., K.C.C., W.H.P. undertook the statistical analysis. All authors contributed to and have approved the final manuscript.

Additional Information

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