

Klotho: a possible role in the pathophysiology of nephrotic syndrome

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ABSTRACT

Klotho, encoded by the *klotho* gene, is associated with phosphate homeostasis. Klotho acts as a co-receptor for FGF23 for binding to its receptors. With FGF23, klotho regulates the systemic mineral homeostasis by regulation of vitamin D and parathyroid hormone. The anti-inflammatory, antifibrotic and antioxidant properties of klotho give it a cardinal role in the development of various renal diseases. The protective effect of klotho has been evident in different types of nephropathy, including diabetic nephropathy, cyclosporine A-induced nephropathy, Calcineurin inhibitors-induced nephropathy, and renal ischemic-reperfusion injury. Nephrotic syndrome is distinguished by hypoproteinemia, proteinuria, and hypercholesterolemia as a result of the aberration of the glomerular filtration barrier. The various factors and pathways associated with the pathophysiology of the nephrotic syndrome have similarities with other types of nephropathy. Despite these similarities,

the role of klotho in the pathology of nephrotic syndrome remains still unexplored. This mini-review builds the case for the possible role of klotho in nephrotic syndrome. The review explores the possible pathways where klotho can play a major role by identifying the similarities in the pathophysiology of nephrotic syndrome and other types of nephropathy.



INTRODUCTION

Klotho, a 135 kDa transmembrane protein, is associated with the aging process and is involved in phosphate metabolism, and regulates the activity of fibroblast growth factors (FGF) [1]. The word klotho has been derived from the name of the Greek goddess Clotho who is believed to spin the “thread” of human fate. The derivation of the word signifies a metaphor for the life span of an individual as the *klotho* gene (KL) is associated with aging. The role of klotho in aging was first reported in mouse studies where KL deficient mice had a decreased life span of fewer than eight weeks [2]. Similarly, an increased expression of the KL gene had positive effects on the life span of the organism [3]. Animal studies demonstrated an association between klotho protein deficiency and an increase in phosphate levels in the blood that is attributed to be one of the mechanisms involved in the decreasing life span in the animals.

The changes observed in klotho deficient animals are comparable to the age-related senile changes observed in humans [4, 5]. These major changes included plaque formation in the arteries, weakening of bones, fat and muscle loss, hasty shrinkage of the thymus resulting in changes in the architecture of the thymus, and decrease in tissue mass leading to gradual deterioration of the immune system; which are similar to senile changes observed in the elderly.

The klotho gene family comprises α -klotho, β -klotho, and γ -klotho [6]. α -KL gene, located on Chromosome 13, comprises three exons and two introns [7] and encodes a transmembrane protein with a smaller cytoplasmic domain and a larger extracellular domain. There are three distinct types of α -klotho protein: transmembrane klotho, secretory klotho, and soluble klotho [7]. Metalloproteinases enzymes ADAM 10 and 17 act on the extracellular domain of klotho to release it from its membrane sites [8-10]. The enzymatic cleavage by the sheddase enzyme at the extracellular domain leads to the generation of soluble klotho form. Secretory klotho, having a molecular weight of 70 kDa, is another form of klotho protein formed by alternate splicing of KL exons. Fibroblast Growth Factor-23 (FGF23) plays a crucial role in phosphate and vitamin D metabolism requires transmembrane α -klotho protein as its co-receptor [11]. β -klotho and γ -Klotho are also transmembrane proteins [6]. β -klotho acts as a co-receptor for FGF19 and FGF21 regulating bile acid synthesis and energy metabolism [12]. Both the transmembrane form of α -klotho protein (acting as a co-receptor for FGF23) and the soluble form of klotho has been demonstrated to be involved in pathways whose aberration can lead to nephrogenic effects [13,14].

Klotho has been directly implicated in the development of chronic kidney disease (CKD) [15]. A decrease in klotho levels succeeded by a rise in serum FGF23 indicated deterioration of kidney function in chronic kidney disease. Further, FGF23, produced from bones, regulates mineral metabolism [11].

Increased excretion of protein in urine and the resultant edema and hypoalbuminemia characterize the nephrotic syndrome. Of all types of nephrotic syndrome, some are steroid-resistant and some are steroid responsive. Minimal change disease, the commonest cause, is steroid responsive.

Steroid resistance includes focal segmental glomerulosclerosis that has a significant risk of kidney failure [16]. Systemic diseases such as Lupus can also cause nephrotic syndrome.

The primary and secondary causes of nephrotic syndrome should be distinguished and management strategies appropriately tailored. Immunosuppressive medications are the mainstay of treatment. The various causes of nephrotic syndrome have been categorized and listed in Table 1.

Klotho has significant beneficial effects on the kidney by alleviating oxidative stress and by its antiapoptotic properties. This protective effect of klotho in different types of nephropathy, including diabetic nephropathy, has been demonstrated in multiple studies.

The pathologic alterations observed in different types of nephropathy such as increased oxidative stress, and inflammation, have also been demonstrated in nephrotic syndrome. However few studies have explored the role of klotho in nephrotic syndrome.

KLOTHO IN DIABETIC NEPHROPATHY

The role of klotho in the development of diabetic nephropathy has been explored in various animal models. A decreased klotho level in animal models increased the purinergic receptor P2X₇, culminating in cell death by apoptosis or necrosis in diabetic nephropathy [18]. Klotho was also demonstrated to suppress the hyperglycemia-mediated glomerular endothelial injury and activation of the Wnt/ β -catenin pathway in mice models of diabetic nephropathy [19]. The aetiological role of klotho in DN was further expounded by the attenuation of apoptosis of renal tubular cells by the drug atrasentan that acts by decreasing the expression of miR-199b-5p and thus increasing its target, klotho [20]. A reduction in the odds of early nephropathy in T2DM patients was observed at a higher concentration of FGF-23, which correlated positively with sKL in diabetic patients [21]. Interestingly, nephrotic syndrome, when compared with controls is associated with decreased levels of FGF23. The reduction in Vitamin D levels and the loss of FGF23 in urine are thought to contribute to lowered levels observed [22].

Table 1 List of various causes of nephrotic syndrome categorized based on etiology

Genetic	Infectious causes	Idiopathic	Others
<ul style="list-style-type: none"> Diffuse mesangial sclerosis (DMS) Epidermolysis bullosa associated Steroid-resistant nephrotic syndrome Familial focal segmental glomerulosclerosis (FSGS) 	<ul style="list-style-type: none"> Congenital infections including syphilis, toxoplasmosis, and HIV Cytomegalovirus HIV-associated nephropathy 	<ul style="list-style-type: none"> Minimal change nephropathy Focal segmental glomerulosclerosis Diffuse mesangial hypercellularity Membranous glomerulonephritis Membranoproliferative GN 	<ul style="list-style-type: none"> Lupus nephropathy IgA nephropathy Drugs Malignancies Hemolytic uremic syndrome (HUS)

INFLAMMATION AND OXIDATIVE STRESS IN NEPHROPATHY

The protective effect of klotho has also been explored in other types of nephropathy. Increased klotho levels, in cyclosporine A-induced nephropathy, regulate cytokine expression and modulate the inflammation via PDLIM2/NF- κ B p65 pathway resulting in beneficial effects [23]. It also has a crucial role in protection against Calcineurin inhibitors-induced nephropathy [24]. The decreased klotho levels make the kidney vulnerable to oxidative stress-induced organ injury [25]. Klotho mitigates oxidative stress by increasing the manganese superoxide dismutase expression via suppression of the PI3K-AKT signaling pathway [26]. Further, klotho also prevents Calcineurin inhibitors-induced nephropathy by improving autophagy clearance and preventing autophagy cell death [27]. In renal ischemic-reperfusion injury, klotho was demonstrated to have an inhibitory effect on oxidative stress in tubular epithelial cells thus preventing necroptosis [28]. Besides, the inflammatory environment also adversely affects the klotho expressions via NF κ B-dependent mechanism as shown by the downregulation of klotho expression by inflammatory cytokines, such as TWEAK and TNF α [29].

Similar to the aforementioned types of nephropathy, the nephrotic syndrome also has been correlated with increased oxidative stress and inflammatory state. Patients with steroid-sensitive nephrotic syndrome had higher plasma levels of advanced oxidation protein products and malondialdehyde indicating oxidative stress in them [30]. The increased pro-oxidant status in nephrotic children leads to considerable change in antioxidant concentrations [31]. The presence of oxidative stress and abnormality in the antioxidative system has been verified in adult nephrotic syndrome patients also [32, 33]. The increased activity of GSH-Px and

selenium content in polymorphonuclear leukocytes (PMNLs) in nephrotic syndrome also indicates the presence of oxidative stress in these patients [34]. The decrease in NF- κ B p65 in addition to the up-regulation of IL-2 are mechanisms hypothesized to initiate glucocorticoid resistance in steroid-resistant nephrotic syndrome [35]. Similarly, elevated serum TNF α levels in nephrotic syndrome are associated with a lack of response to steroids [36]. Further, In vitro study has demonstrated the increased expression by TWEAK of PLA2R as well as NFKB1 and IRF4 which are linked to membranous nephropathy [37].

FACTORS AFFECTING KLOTHO EXPRESSION

Albuminuria, in cultured tubular cells, decreased the expression of klotho [38]. Concurrently, in CKD animal models with frank albuminuria, the klotho expressions were found to be suppressed indicating a possible role that klotho may have in the pathogenesis of CKD.

Proteinuric kidney disease is associated with endoplasmic reticulum (ER) stress. Animal models of albuminuria demonstrated features of ER stress in renal tubular cells which are instigated by the albumin and mediated via ATF3/ATF4 activation. The induction of ATF3 and ATF4 leads to enhanced binding to the promoter region effectuating altered transcription of the klotho gene culminating in the suppression of klotho expression [39]. ATF3 is also induced in renal ischemia-reperfusion injury, where klotho is observed to be downregulated [40]. Further, ER stress accentuates klotho degradations via proteasome and lysosome. Hence, proteinuria (especially albuminuria) leads to decreased klotho protein half-life as well as the genetic expression [41].

Further, the downregulation of klotho in tubular epithelial cells is also associated with the aggravation of renal fibrosis [42]. Although solu-

ble α -klotho positively correlated with eGFR in patients with CKD, the efficacy of klotho in improving renal function in CKD patients is still under investigation [43]. Further, the expression of klotho in kidneys is also affected by excessive renin–angiotensin–aldosterone system (RAAS) activation. In vitro experiments demonstrated Angiotensin II to have a suppressive effect on klotho through PPAR- γ downregulation [44] (Figure 1).

The aforementioned factors affecting klotho expression (albuminuria and Angiotensin II) have been implicated in the pathophysiology of nephrotic syndrome in various studies. The glomerular-derived angiotensinogen has been thought to have a significant effect on glomerular dysfunction in nephrotic syndrome. As a result, the ARB treatment is beneficial in slit diaphragm injury by inhibiting the positive feedback loop of the activated local Ang II action [45]. During podocyte injury, a vicious loop that stimulates the intrarenal generation of Angiotensin II is activated aggravating the development of glomerular dysfunction [46]. Further, PPAR γ

agonists decrease the proteinuria in acute nephrotic syndrome by regulating the expression of multiple genes like actinin-4 and nephrin and leading to the restoration of podocyte structure [47]. Hence, activation of PPAR γ and inhibition of ANGPTL4 is associated with a better prognosis in patients with nephrotic syndrome [48].

CONCLUSION

Figure 2 summarizes the possible mechanisms by which klotho may play a cardinal role in the pathophysiology of nephrotic syndrome.

The various factors associated with the pathophysiology of nephrotic syndrome, including oxidative stress and inflammation, are intertwined with klotho expression and its downstream effects. The multitude of studies demonstrating the association of klotho and these factors in other types of nephropathy warrants its investigation into the nephrotic syndrome. The insights thus obtained would help in designing therapeutic strategies involving klotho and its downstream effectors for nephrotic syndrome.

Figure 1 Depiction of factors and their modes of influencing the expression of klotho. A: Albuminuria; B: Angiotensin II

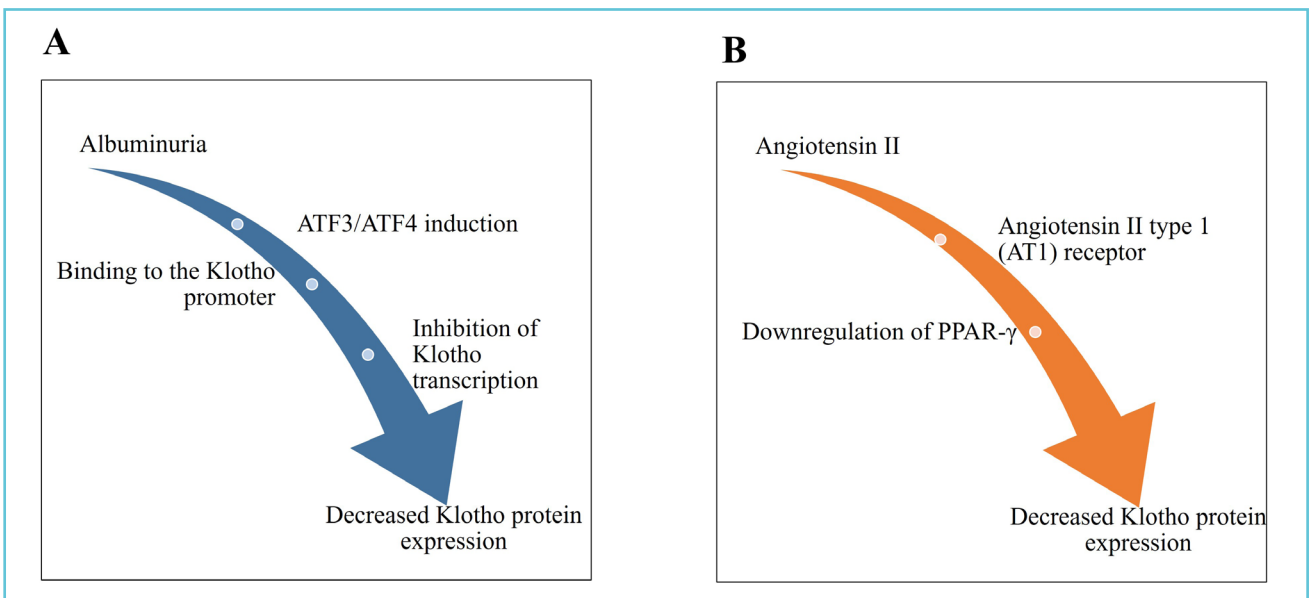
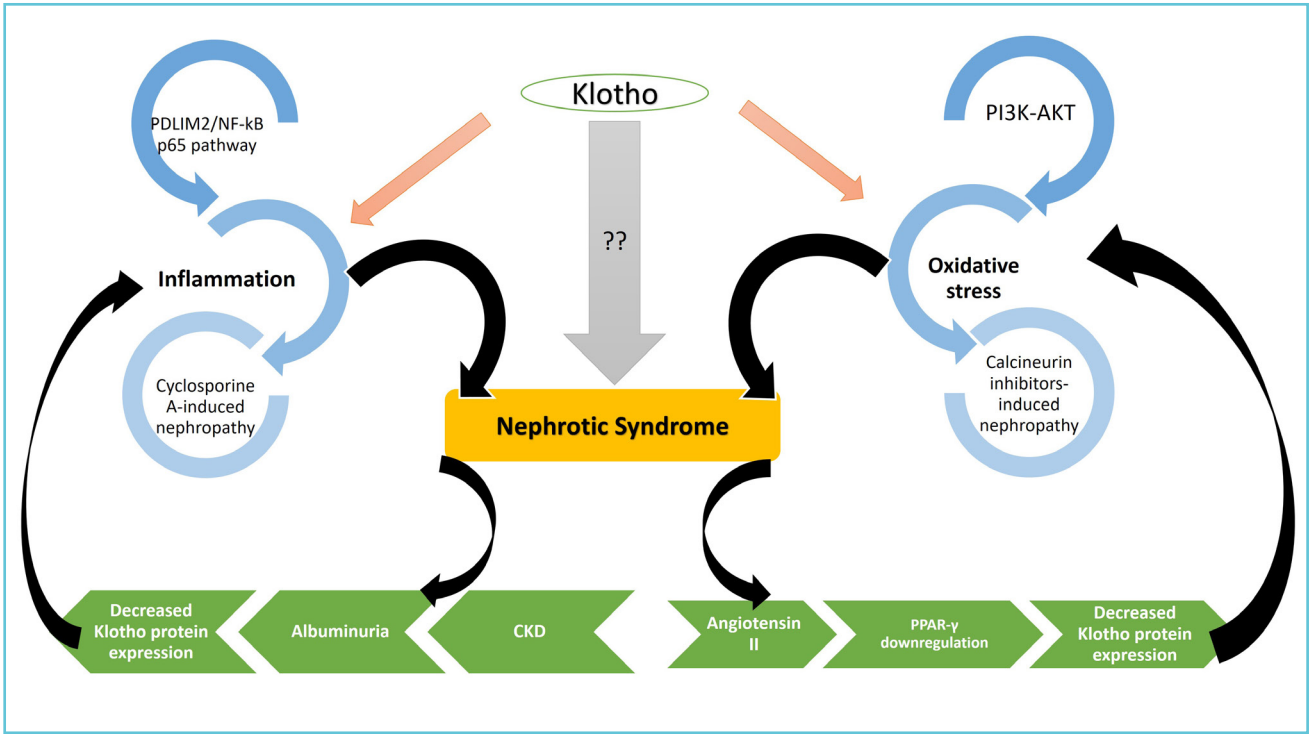


Figure 2 Illustration of the possible roles of klotho in nephrotic syndrome



REFERENCES

1. Vo HT, Laszczyk AM, King GD. Klotho, the Key to Healthy Brain Aging?. *Brain Plast* 2018;3(2):183-194.
2. Kuro-o M, Matsumura Y, Aizawa H, Kawaguchi H, Suga T, Utsugi T, et al. Mutation of the mouse KLOTHO gene leads to a syndrome resembling ageing. *Nature*. 1997; 390(6655):45–51.
3. Kurosu H, Yamamoto M, Clark JD, Pastor JV, Nandi A, Gurnani P, et al. Suppression of aging in mice by the hormone KLOTHO. *Science*.2005;309(5742):1829–33.
4. Urakawa I, Yamazaki Y, Shimada T, Iijima K, Hasegawa H, Okawa K, et al. KLOTHO converts canonical FGF receptor into a specific receptor for FGF23. *Nature*. 2006;444(7120): 770–74.
5. Kurosu H, Ogawa Y, Miyoshi M, Yamamoto M, Nandi A, Rosenblatt KP, et al. Regulation of fibroblast growth factor-23 signaling by KLOTHO. *J Biol Chem*. 2006;281(10): 6120–3.
6. Ito S, Fujimori T, Hayashizaki Y, Nabeshima Y. Identification of a novel mouse membrane-bound family 1 glycosidase-like protein, which carries an atypical active site structure. *Biochim Biophys Acta*. 2002;1576:341–5.
7. Shiraki-Iida T, Aizawa H, Matsumura Y, Sekine S, Iida A, Anazawa H, et al. Structure of the mouse KLOTHO gene and its two transcripts encoding membrane and secreted protein. *FEBS Lett*. 1998;424(1-2):6-10.
8. Imura A, Tsuji Y, Murata M, Maeda R, Kubota K, Iwano A, et al. alpha-KLOTHO as a regulator of calcium homeostasis. *Science*. 2007;316(5831):1615–8.
9. Chen C-D, Podvin S, Gillespie E, Leeman SE, Abraham CR. Insulin stimulates the cleavage and release of the extracellular domain of KLOTHO by ADAM10 and ADAM17. *Proc Natl Acad Sci U S A*. 2007;104(50):19796-801.
10. Chen C-D, Tung TY, Liang J, Zeldich E, Tucker Zhou TB, Turk BE, et al. Identification of cleavage sites leading to the shed form of the anti-aging protein KLOTHO. *Biochemistry*. 2014;53(34):5579-87.
11. Navarro-García JA, Fernández-Velasco M, Delgado C, Delgado JF, Kuro-O M, Ruilope LM, et al. PTH, vitamin D, and the FGF-23-klotho axis and heart: Going beyond the confines of nephrology. *Eur J Clin Invest* 2018; 48(4).
12. Kurosu H, Kuro OM. The klotho gene family as a regulator of endocrine fibroblast growth factors. *Mol Cell Endocrinol*. 2009; 299:72–8
13. Zou, D., Wu, W., He, Y. et al. The role of klotho in chronic kidney disease. *BMC Nephrol*. 2018;19:285.
14. Dalton GD, Xie J, An SW, Huang CL. New Insights into the Mechanism of Action of Soluble Klotho. *Front Endocrinol (Lausanne)*. 2017;8:323

15. Komaba H, Lanske B. Role of Klotho in bone and implication for CKD. *Curr Opin Nephrol Hypertens* 2018;27(4): 298-304.
16. Wang CS, Greenbaum LA. Nephrotic Syndrome. *Pediatr Clin North Am* 2019;66(1):73-85.
17. Ranganathan S. Pathology of Podocytopathies Causing Nephrotic Syndrome in Children. *Front Pediatr*. 2016 Mar 31;4:32.
18. Rodrigues AM, Serralha RS, Farias C, Punaro GR, Fernandes MJS, Higa EMS. P2X7 receptor and klotho expressions in diabetic nephropathy progression. *Purinergic Signal* 2018;14(2):167-176.
19. Wang Q, Ren D, Li Y, Xu G. Klotho attenuates diabetic nephropathy in db/db mice and ameliorates high glucose-induced injury of human renal glomerular endothelial cells. *Cell Cycle* 2019;18(6-7):696-707.
20. Kang WL, Xu GS. Atrasentan increased the expression of klotho by mediating miR-199b-5p and prevented renal tubular injury in diabetic nephropathy. *Sci Rep* 2016;6: 19979.
21. Fariás-Basulto A, Martínez-Ramírez HR, Gómez-García EF, Cueto-Manzano AM, Cortés-Sanabria L, Hernández-Ramos LE, et al. Circulating Levels of Soluble Klotho and Fibroblast Growth Factor 23 in Diabetic Patients and Its Association with Early Nephropathy. *Arch Med Res* 2018;49(7):451-455.
22. Yadav AK, Ramachandran R, Aggarwal A, Kumar V, Gupta KL, Jha V. Fibroblast growth factor 23 in untreated nephrotic syndrome. *Nephrology (Carlton)* 2018;23(4): 362-365.
23. Jin M, Lv P, Chen G, Wang P, Zuo Z, Ren L, et al. Klotho ameliorates cyclosporine A-induced nephropathy via PDLIM2/NF- κ B p65 signaling pathway. *Biochem Biophys Res Commun* 2017;486(2):451-457.
24. Jin J, Jin L, Lim SW, Yang CW. Klotho deficiency aggravates tacrolimus-induced renal injury via the phosphatidylinositol 3-kinase-Akt-forkhead box protein O pathway. *Am J Nephrol* 2016;43(5):357-365.
25. Luo K, Lim SW, Quan Y, Cui S, Shin YJ, Ko EJ, et al. Role of Klotho in Chronic Calcineurin Inhibitor Nephropathy. *Oxid Med Cell Longev*. 2019;2019:1825018.
26. Lim SW, Jin L, Luo K, Jin J, Shin YJ, Hong SY, et al. Klotho enhances FoxO3-mediated manganese superoxide dismutase expression by negatively regulating PI3K/AKT pathway during tacrolimus induced oxidative stress. *Cell Death Dis* 2017;8(8):e2972.
27. Lim SW, Shin YJ, Luo K, Quan Y, Ko EJ, Chung BH, Yang CW. Effect of Klotho on autophagy clearance in tacrolimus-induced renal injury. *FASEB J* 2019;33(2):2694-2706.
28. Qian Y, Guo X, Che L, Guan X, Wu B, Lu R, et al. Klotho Reduces Necroptosis by Targeting Oxidative Stress Involved in Renal Ischemic-Reperfusion Injury. *Cell Physiol Biochem* 2018;45(6):2268-2282.
29. Moreno JA, Izquierdo MC, Sanchez-Niño MD, Suárez-Alvarez B, Lopez-Larrea C, Jakubowski A, et al. The inflammatory cytokines TWEAK and TNF α reduce renal klotho expression through NF κ B. *J Am Soc Nephrol* 2011;22(7): 1315-25.
30. Fan A, Jiang X, Mo Y, Tan H, Jiang M, Li J. Plasma levels of oxidative stress in children with steroid-sensitive nephrotic syndrome and their predictive value for relapse frequency. *Pediatr Nephrol* 2016;31(1):83-8.
31. Kamireddy R, Kavuri S, Devi S, Vemula H, Chandana D, Harinarayanan S, et al. Oxidative stress in pediatric nephrotic syndrome. *Clin Chim Acta* 2002;325(1-2):147-50.
32. Soyoral YU, Aslan M, Emre H, Begenik H, Erdur FM, Turkel A, et al. Serum paraoxonase activity and oxidative stress in patients with adult nephrotic syndrome. *Atherosclerosis* 2011;218(1):243-6.
33. Bulucu F, Vural A, Aydin A, Sayal A. Oxidative stress status in adults with nephrotic syndrome. *Clin Nephrol* 2000; 53(3):169-73.
34. Akyol T, Bulucu F, Sener O, Yamanel L, Aydin A, Inal V, et al. Functions and oxidative stress status of leukocytes in patients with nephrotic syndrome. *Biol Trace Elem Res* 2007;116(3):237-48.
35. Aviles DH, Matti Vehaskari V, Manning J, Ochoa AC, Zea AH. Decreased expression of T-cell NF-kappaB p65 subunit in steroid-resistant nephrotic syndrome. *Kidney Int* 2004;66(1):60-7.
36. Weissbach A, Garty BZ, Lagovsky I, Krause I, Davidovits M. Serum Tumor Necrosis Factor-Alpha Levels in Children with Nephrotic Syndrome: A Pilot Study. *Isr Med Assoc J* 2017;19(1):30-33.
37. Cuarental L, Valiño-Rivas L, Mendonça L, Saleem M, Mezzano S, Sanz AB, et al. Tacrolimus Prevents TWEAK-Induced PLA2R Expression in Cultured Human Podocytes. *J Clin Med* 2020;9(7):2178.
38. Fernandez-Fernandez B, Izquierdo MC, Valiño-Rivas L, Nastou D, Sanz AB, Ortiz A, Sanchez-Niño MD. Albumin downregulates Klotho in tubular cells. *Nephrol Dial Transplant* 2018;33(10):1712-1722.
39. de Seigneux S, Wilhelm-Bals A, Courbebaisse M. On the relationship between proteinuria and plasma phosphate. *Swiss Med Wkly* 2017;147:w14509.
40. Hu MC, Shi M, Zhang J, Quiñones H, Kuro-o M, Moe OW. Klotho deficiency is an early biomarker of renal ischemia-reperfusion injury and its replacement is protective. *Kidney Int* 2010;78(12):1240-51.

41. Delitsikou V, Jarad G, Rajaram RD, Ino F, Rutkowski JM, Chen CD, et al. Klotho regulation by albuminuria is dependent on ATF3 and endoplasmic reticulum stress. *FASEB J* 2020;34(2):2087-2104.
42. Liu Y, Bi X, Xiong J, Han W, Xiao T, Xu X, et al. MicroRNA-34a Promotes Renal Fibrosis by Downregulation of Klotho in Tubular Epithelial Cells. *Mol Ther* 2019;27(5): 1051-1065.
43. Wang Q, Su W, Shen Z, Wang R. Correlation between Soluble α -Klotho and Renal Function in Patients with Chronic Kidney Disease: A Review and Meta-Analysis. *Biomed Res Int* 2018;2018:9481475.
44. Maquigussa E, Paterno JC, de Oliveira Pokorny GH, da Silva Perez M, Varela VA, da Silva Novaes A, et al. Klotho and PPAR Gamma Activation Mediate the Renoprotective Effect of Losartan in the 5/6 Nephrectomy Model. *Front Physiol* 2018;9:1033.
45. Yamazaki M, Fukusumi Y, Kayaba M, Kitazawa Y, Takamura S, Narita I, et al. Possible role for glomerular-derived angiotensinogen in nephrotic syndrome. *J Renin Angiotensin Aldosterone Syst* 2016;17(4):1470320316681223.
46. Koizumi M, Ueda K, Niimura F, Nishiyama A, Yanagita M, Saito A, et al. Podocyte Injury Augments Intrarenal Angiotensin II Generation and Sodium Retention in a Megalin-Dependent Manner. *Hypertension* 2019;74(3): 509-517.
47. Zuo Y, Yang HC, Potthoff SA, Najafian B, Kon V, Ma LJ, et al. Protective effects of PPAR γ agonist in acute nephrotic syndrome. *Nephrol Dial Transplant* 2012;27(1):174-81.
48. Lu R, Zhou J, Liu B, Liang N, He Y, et al. Paeoniflorin ameliorates Adriamycin-induced nephrotic syndrome through the PPAR γ /ANGPTL4 pathway in vivo and vitro. *Biomed Pharmacother* 2017;96:137-147.