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Research Article

Ameliorative Effects of *Pueraria lobata* Extract on Postmenopausal Symptoms through Promoting Estrogenic Activity and Bone Markers in Ovariectomized Rats

Mi-Ra Lee , Bohye Kim , Yongjun Lee , Soung-Yong Park , Jae-Hoon Shim , Bong-Hwan Chung , and Seong-Il Heo

Correspondence should be addressed to Seong-Il Heo; h.seongil@himh.re.kr

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Pueraria lobata (Willd.) Ohwi, known as kudzu, is one of the most popular traditional medicines in Asian countries. It has been widely used as a natural alternative to hormone replacement therapy for treating postmenopausal symptoms. This study aimed to investigate the estrogenic effect of P. lobata extract (PE) against postmenopausal osteoporosis in ovariectomized (OVX) rats. OVX rats were treated with PE (25–1600 mg/kg) for 8 weeks. Biochemical parameters, estradiol, and bone turnover markers (e.g., osteocalcin, C-terminal telopeptide fragment of type I collagen, deoxypyridinoline, and pyridinoline) were measured in plasma samples. In addition, estrogen receptor-alpha (ER-α) protein expression and morphology of uterine were evaluated. Long-term treatment with PE did not cause liver damage in OVX rats. PE supplementation reduced body weight gain in obese rats with high lipid accumulation induced by ovariectomy. Furthermore, PE exhibited a protective effect against insulin resistance, hyper-lipidemia, and hepatic lipid peroxidation. PE treatment increased uterine weight and thickness of the uterine layers in cases of uterus atrophy due to removal of ovaries. The levels of bone turnover markers, which were significantly increased in OVX rats, were decreased by PE treatment. Western blotting analysis showed that ER-α protein expression was upregulated in PE-treated rats compared with OVX rats. These results suggest that PE could be a promising alternative functional food for improving menopausal symptoms.

1. Introduction

Menopause, also known as climacteric, is a biological stage that represents the end of the reproductive lifespan in women [1]. Menopause-induced estrogen deficiency may increase the risk of metabolic diseases, such as obesity, osteoporosis, cardiovascular disease, and diabetes [2, 3]. Hormone replacement therapy (HRT) is commonly used to treat menopausal symptoms, but long-term HRT can lead to an increased risk of carcinogenesis and cardiovascular diseases [4, 5]. Thus, there is a need for natural alternative therapies, including dietary supplements and new herbal medicines, to reduce adverse side effects. Phytoestrogens are

estrogen-like compounds derived from plants with a structure similar to that of 17β -estradiol [6]. The antipostmenopausal effects of phytoestrogens have been demonstrated to have various biological activities, such as bone-protective effects and improvement in menopausal symptoms, as a potential alternative to HRT [7–9].

Pueraria lobata (Willd.) Ohwi, known as kudzu, has been traditionally used in combination with other herbs to produce synergistic and additive effects through a multitarget approach for disease [10]. It contains high levels of several isoflavones, such as puerarin, daidzin, daidzein, genistin, and genistein [11]. Pueraria lobata is known to have multiple pharmacological activities that help alleviate

¹Hongcheon Institute of Medicinal Herb, Hongcheon-gun 25142, Gangwon, Republic of Korea

²Sejun F & B Co. Ltd., Hongcheon-gun 25107, Gangwon, Republic of Korea

³Department of Food Science and Nutrition, Hallym University, Chuncheon 24252, Gangwon, Republic of Korea

various diseases such as liver diseases, cardiovascular diseases, diabetes, and osteoporosis [12, 13]. In particular, puerarin, the primary active component of P. lobata, has been demonstrated to have preventive activity against osteoporosis in several studies using ovariectomized (OVX) animal models [14, 15]. An OVX rat model is considered appropriate for investigating problems related to postmenopausal bone loss, a primary risk factor for osteoporosis [16]. However, there is little information on the effect of P. lobata on dyslipidemia and bone metabolism in OVX rats. Based on the estrogenic activity of puerarin, we predicted that P. lobata extract (PE) could have beneficial effects on menopausal metabolic dysfunction, the endocrine system, and osteoporosis in OVX rats. Therefore, we investigated the protective effects of long-term supplementation with PE against metabolic parameters in OVX rats. In addition, white adipose tissue weight, atrophic uterus morphology, bone turnover markers, and estrogen levels were evaluated to clarify the underlying mechanisms of action of PE extract.

2. Materials and Methods

- 2.1. Preparation of P. lobata Extract. PE was provided by Sejun F & B Co., Ltd. (Gangwon, Korea). It was in the form of a spray-dried powder, which was prepared as follows. To separate the starch, dried P. lobate was soaked in 50% ethanol for 1 h and then centrifuged. The residue of P. lobate was extracted in five volumes of 50% ethanol for 1 h at 60°C. The extract was filtered and concentrated under reduced pressure. The PE was powdered in the spray drying system by mixing 30% dextrin. The content of puerarin, which is a major isoflavone in PE, was 42.38 mg/g.
- 2.2. Animals and Treatment. Nine-week-old female Sprague Dawley rats, which were ovariectomized or sham-operated at 8 weeks of age, were purchased from Chemon Inc. (Gyeonggido, Korea) and maintained under controlled laboratory conditions (23°C \pm 1°C, humidity 55% \pm 5%, 12 h light/dark cycle) for 2 weeks before the experiments. The rats were randomly divided into the following treatment groups (n = 8per group): (1) sham group, (2) OVX group, (3) OVX+PE 25 mg/kg group, (4) OVX + PE 100 mg/kg group, (5) OVX + PE 400 mg/kg group, and (6) OVX + PE 1600 mg/kg group. All animal experiment protocols in this study were reviewed and approved by the Institutional Animal Care and Use Committee of the Hongcheon Institute of Medicinal Herb (No. 19-07). PE was suspended in phosphate buffered saline (PBS) for oral administration at the desired dose in a volume of 5 mL/kg once daily for 8 weeks. The food intake and body weight of rats were measured twice a week. After 16 h of fasting on the last day of the experiment, blood was collected from the portal vein. The uterus and white adipose tissue were collected and weighed.
- 2.3. Biochemical Parameters. The plasma samples were prepared by centrifuging the collected blood in heparinized tubes (4,000 rpm for 15 min at 4°C) and then stored at -80°C for assay. Aspartate aminotransferase (AST), alanine

aminotransferase (ALT), Ca, P, total cholesterol (TC), high-density lipoprotein cholesterol (HDL-C), low-density lipoprotein cholesterol (LDL-C), and triglyceride (TG) levels were measured using an automated clinical chemistry analyser (Konelab 20XT; Thermo Fisher Scientific, Waltham, MA, USA). Free fatty acid (FFA; BioAssay Systems, Hayward, CA, USA), insulin, estradiol (E2), and serotonin levels were determined using enzyme-linked immunosorbent assay (ELISA) kits (Cusabio, Wuhan, China) according to the manufacturer's instructions.

- 2.4. Liver Lipid Peroxidation Assay. The liver sample (120 mg) was homogenized in 1 mL of PBS and then centrifuged at 10,000 rpm for 15 min at 4°C. The supernatant obtained was used a source for the assay. Malondialdehyde (MDA) level was determined using the thiobarbituric acid reactive substance (TBARS) assay according to the manufacturer's (Dogenbio, Seoul, Korea) instructions.
- 2.5. Bone Turnover Markers in Plasma Samples. The bone formation marker Osteocalcin (OC) was determined using a rat osteocalcin ELISA kit (Cusabio). The bone resorption markers, such as C-terminal telopeptide fragment of type I collagen (CTX-1; Cusabio), deoxypyridinoline (DPD; Cusabio), and pyridinoline (PYD; MyBioSource, San Diego, USA), were determined using commercial ELISA kits.
- 2.6. Immunoblot Analysis. The total protein in the uterus was extracted using RIPA lysis buffer containing phosphatase and protease inhibitor cocktails. Equal amounts of protein ($15\,\mu g$) were separated on 10% sodium dodecyl sulfate polyacrylamide gels and then transferred on to polyvinylidene fluoride membranes. The membranes were blocked with 5% skim milk for 1 h and then probed with specific primary antibodies at a final dilution of 1:1000. The immunoreactive protein bands were detected using an enhanced chemiluminescence western blot detection kit (Clarity MaxTM Western ECL Substrate; Bio-Rad Laboratories, Hercules, CA, USA) and visualized using the Chemidoc Touch image system (ChemiDocTM XRS+system; Bio-Rad Laboratories).
- 2.7. Histological Analysis. The uterus samples were fixed in 10% formalin and embedded in paraffin. The tissue sections were mounted on glass slides and stained with haematoxylin and eosin. The static parameters of the uterus, endometrial, myometrial, and epithelial lengths were calculated using ImageJ software (NIH, USA).
- 2.8. Statistical Analysis. All data are expressed as mean±standard error of the mean (SEM). They were analysed using the analysis of variance followed by Dunnett's multiple comparison test. Differences were considered statistically significant at p < 0.05. All statistical analyses were performed using GraphPad Prism 7.05.

3. Results

3.1. Effects of PE on Body Weight, Food Efficiency Ratio, and White Adipose Tissue in OVX Rats. As shown in Figure 1(a), the OVX group showed a significant increase in body weight compared with the sham group throughout the experimental period. However, PE treatment (400 and 1600 mg/kg) significantly reduced body weight gain from week 6 by 15% and 35%, respectively, compared with the OVX group (Figure 1(b)). The food efficiency ratio in the PE-1600 group was $6.28\% \pm 0.88\%$, which was significantly reduced by 28% compared to that in the OVX group (Figure 1(c)). The weight of mesenteric white adipose tissue (mWAT) increased significantly in the OVX group compared with the sham group. PE treatment significantly suppressed the increase in mWAT weight in the OVX rats (Figure 1(d)).

3.2. Effects of PE on Uterus Weight and Histological Analysis. The OVX rats presented with prominent atrophy of the uterus as shown in Figure 2(e). The uterus weight of rats in the OVX group $(0.27\pm0.22 \, \text{mg/g})$ was 86% lower than that in the sham group $(2.03\pm0.49 \, \text{mg/g})$ (Figure 2(a)). Although not statistically significant, PE treatment recovered the decrease in uterus weight by 9%–46% in a dose-dependent manner compared with the OVX group. Figures 2(b)–2(d) show the results of histological analysis of the stained uterine sections. The thickness of the endometrium, myometrium, and epithelium in the OVX group was significantly decreased compared with that in the sham group. However, PE treatment showed a marked reversal of the OVX-induced uterine atrophy.

3.3. Effects of PE on Plasma Biochemical Parameters. The results of plasma biochemical parameters are summarised in Table 1. The plasma levels of ALT and AST were not significantly different among the groups. The blood Ca level in the OVX group decreased by 10% compared with that in the sham group, but it was not statistically significant. Treatment with either PE 400 or 1600 mg/kg significantly increased the blood Ca level compared with the OVX group. The blood P level did not differ significantly among the groups. The OVX group showed a considerable increase in fasting plasma FFA and insulin levels compared with the sham group. The PE-1600 group showed a significant decrease in FFA level by 50% compared with the OVX group. The elevated insulin level in OVX rats was significantly reduced in all PE-treated groups except for the PE-100 group.

3.4. Effects of PE on Plasma Lipids and Liver MDA Levels in OVX Rats. To further investigate whether PE treatment improves the high lipid accumulation induced by ovariectomy, we measured the changes in plasma lipid profiles and MDA levels in the liver homogenates (Table 2). The plasma TC and LDL-C levels did not change in the OVX- and PEtreated groups. However, chronic PE treatment increased the HDL-C levels, with the PE-400 group, especially showing a statistically significant increase compared with the OVX

group. The OVX group showed significantly elevated TG levels compared with the sham group. PE-treated groups inhibited increase of TG levels by 16% to 43%. Furthermore, the PE-25 group significantly reduced TG level to the similar results of the sham group (p<0.001). Liver MDA levels significantly increased by 25% in the OVX group compared with the sham group. Treatment with PE significantly reduced these increases in MDA levels to values comparable to those in the sham group.

3.5. Effects of PE on Bone Mineral Density, OC, E2, and ER- α Protein Levels. The bone mineral density (BMD) of the femur is shown in Figure 3(a). The BMD in the OVX group was decreased by 5% compared with that in the sham group. PE treatment did not affect ovariectomy-induced decrease in BMD. OC levels were markedly increased in the OVX group compared to the sham group (Figure 3(b)). However, PE treatment (100, 400, or 1600 mg/kg) significantly inhibited the elevated OC levels to values similar to those in the sham group. Plasma E2 levels were significantly lower in the OVX group than in the sham group. However, PE treatment, except for 1600 mg/kg, significantly increased E2 levels in OVX rats (Figure 3(c)). The expression of ER- α was decreased in OVX rats, and PE treatment upregulated ER- α expression in a dose-dependent manner (Figure 3(d)).

3.6. Effects of PE on Serotonin and Bone Turnover Markers. Table 3 summarises the changes in the levels of serotonin and bone turnover markers, such as CTX-1, DPD, and PYD. The OVX group showed a considerable increase in serotonin, DYP, and PYD levels compared with the sham group. There was no significant difference in CTX-1 levels between the sham and OVX groups. PE treatment of OVX rats significantly decreased these bone turnover markers and serotonin levels.

4. Discussion

Estrogen deficiency in postmenopausal women and OVX rats promotes metabolic syndromes [17]. The OVX rat model has been widely used to evaluate the estrogenic activity of natural compounds [18]. In this study, we investigated the estrogenic activity of PE against postmenopausal symptoms such as obesity, osteoporosis, and dyslipidemia in OVX rats.

The OVX rats present with weight gain, uterine atrophy, bone loss, and estrogen reduction, all of which commonly occur in postmenopausal women. As described previously [19], OVX rats showed a significant increase in body weight gain, distinct uterine atrophy, and uterine weight loss compared with sham-operated rats, indicating that ovariectomy successfully induced menopause. However, PE treatment inhibited uterine weight loss and body weight gain and increased endometrial thickness in OVX rats. These results suggest that PE could improve postmenopausal symptoms in OVX rats.

We also confirmed that PE supplementation elevated the expression of ER- α protein in the uterus and plasma E2

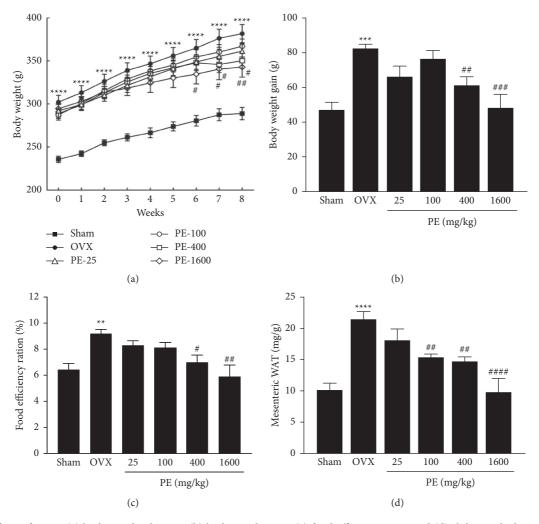
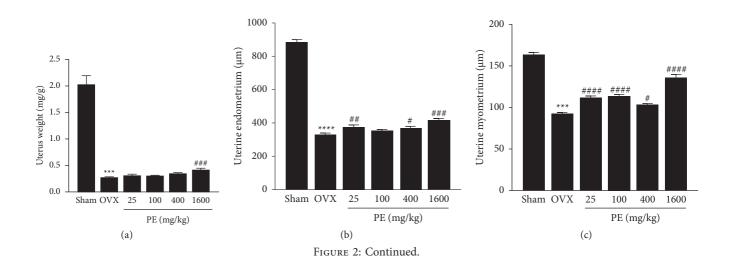
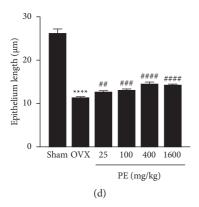


FIGURE 1: Effects of PE on (a) body weight changes, (b) body weight gain, (c) food efficiency ratio, and (d) abdominal white adipose tissue weight in OVX rats. Data represent the mean \pm standard error of mean (SEM). ** p < 0.01, *** p < 0.001, and **** p < 0.0001 vs. sham group. # p < 0.05, ## p < 0.01, *** p < 0.001, and **** p < 0.001, and **** p < 0.001, and **** p < 0.001, *** p < 0.001, ** p < 0.001, *** p < 0.





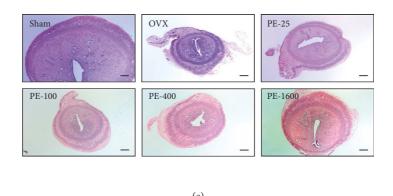


FIGURE 2: Effects of PE on uterus weight and histological analysis in OVX rats. (a) Uterus weight, (b) uterine endometrial, (c) uterine myometrial, (d) uterine epithelial length, and (e) haematoxylin and eosin stain (50×, bar = 200 μ m). Each column represents the mean \pm standard error of mean (SEM). *** p < 0.001 and **** p < 0.0001 vs. sham group; *# p < 0.01, *## p < 0.001, and *### p < 0.0001 vs. OVX group.

TABLE 1: Effects of PE on plasma biochemical parameters in OVX rats.

	Sham	OVX	PE-25	PE-100	PE-400	PE-1600
ALT (U/L)	17.46 ± 1.27	24.70 ± 2.51	24.87 ± 1.99	23.76 ± 1.31	20.77 ± 0.98	22.01 ± 1.60
AST (U/L)	76.48 ± 5.11	109.00 ± 16.84	99.76 ± 6.44	93.73 ± 6.51	80.73 ± 6.37	74.90 ± 5.29
Ca (mg/dL)	4.95 ± 0.31	4.47 ± 0.40	5.28 ± 0.12	5.28 ± 0.14	$5.73 \pm 0.13^{##}$	$5.72 \pm 0.12^{\#\#}$
P (mg/dL)	4.06 ± 0.22	4.49 ± 0.56	4.37 ± 0.30	4.28 ± 0.32	4.91 ± 0.20	4.45 ± 0.25
FFA (μM)	197.3 ± 14.96	$346.7 \pm 35.42^*$	254.2 ± 33.97	247.7 ± 37.05	321.40 ± 37.74	$173.7 \pm 42.8^{\#\#}$
Insulin (ng/mL)	0.92 ± 0.04	1.23 ± 0.03 ***	$1.00 \pm 0.04^{\#\#}$	1.11 ± 0.05	$1.03 \pm 0.05^{\#\#}$	$0.94 \pm 0.04^{\#\#}$

Data are presented as the mean \pm standard error of mean (SEM). Sham, sham operation; OVX, ovariectomy; PE-25, OVX + PE 25 mg/kg; PE-100, OVX + PE 100 mg/kg; PE-400, OVX + PE 400 mg/kg; PE-1600, OVX + PE 1600 mg/kg; FFA, free fatty acid; ALT, alanine aminotransferase; AST, aspartate aminotransferase. *p < 0.005 and *** p < 0.001 vs. OVX group; *#p < 0.001 vs. OVX group.

TABLE 2: Effects of PE on plasma lipid profiles and liver MDA levels in OVX rats.

	Sham	OVX	PE-25	PE-100	PE-400	PE-1600
TC (mg/dL)	48.25 ± 3.09	60.10 ± 4.34	55.45 ± 5.62	65.64 ± 1.88	61.81 ± 2.92	55.52 ± 3.78
HDL-C (mg/dL)	55.53 ± 2.68	59.30 ± 4.85	72.52 ± 5.01	71.20 ± 3.15	$74.46 \pm 4.18^{\#}$	65.60 ± 3.16
LDL-C (mg/dL)	7.76 ± 0.38	10.96 ± 0.82	10.83 ± 1.14	11.23 ± 0.51	9.80 ± 0.79	9.27 ± 0.91
TG (mg/dL)	38.71 ± 3.71	59.55 ± 3.96**	$33.74 \pm 5.49^{###}$	$44.13 \pm 2.27^{\#}$	$42.84 \pm 2.01^{\#}$	49.78 ± 4.03
$MDA(\mu M)$	47.95 ± 2.46	60.22 ± 3.39 *	$42.05 \pm 2.64^{###}$	$39.9 \pm 2.23^{####}$	$40.21 \pm 2.11^{####}$	$42.02 \pm 4.02^{\#\#}$

Data are presented as the mean \pm standard error of mean (SEM). Sham, sham operation; OVX, ovariectomy; PE-25, OVX + PE 25 mg/kg; PE-100, OVX + PE 100 mg/kg; PE-400, OVX + PE 1600 mg/kg; TC, total cholesterol; HDL-C, high-density lipoprotein cholesterol; LDL-C, low-density lipoprotein cholesterol; triglyceride, TG; MDA, malondialdehyde. *p < 0.05 and **p < 0.01 vs. sham group; *p < 0.05, *p < 0.001, and **p < 0.0001 vs. OVX group.

levels in OVX rats. ER- α is essential for the induction of uterine muscular disorganization and vaginal epithelial cell proliferation by estrogen during critical developmental stages [20]. These results imply that PE might exert estrogenic effects by increasing E2 levels and improving the uterus.

The supplementation with PE up to 1600 mg/kg did not change ALT and AST levels, which are biomarkers of hepatic injury. This result is similar to that of a previous study, which reported that *kudzu* roots are not highly toxic based on toxicity assessment [21]. These data suggest that PE intake at this dose did not cause any adverse effects on the livers of OVX rats.

Estrogen deprivation by ovariectomy leads to increased fat accumulation and hepatic steatosis [22]. The ER- α knockout female mice are reported to exhibit an obese phenotype with increased visceral adiposity, decreased energy expenditure, and insulin resistance [23, 24]. Similar to previous studies, the OVX group showed a significant increase in visceral fat mass, plasma FFA, insulin levels, and hepatic lipid peroxidation. The ER- α is also necessary for the attenuation of body weight gain after OVX [25]. In this study, PE treatment significantly suppressed body weight gain, abdominal fat accumulation, and blood lipid levels. These results suggest that PE acts as a phytoestrogen in lipid metabolism in OVX rats.

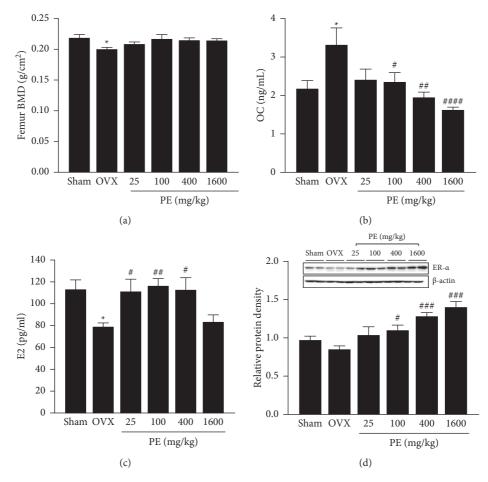


FIGURE 3: Effects of PE on (a) femur bone mineral density (BMD), (b) estradiol (E2), (c) osteocalcin (OC), and (d) estrogen receptor-alpha (ER- α) protein expression. Each column represents the mean ± standard error of mean (SEM). *p < 0.05 vs. sham group; #p < 0.05, ##p < 0.01, ### p < 0.001, and #### p < 0.0001 vs. OVX group.

TABLE 3: Effects of PE on serotonin and bone turnover markers in OVX rats.

	Serotonin (ng/mL)	CTX-1 (pg/mL)	DYP (nmol/L)	PYD (ng/mL)
Sham	200.10 ± 28.19	167.10 ± 14.10	2.10 ± 0.10	27.07 ± 0.97
OVX	$390.20 \pm 67.26^*$	155.5 ± 8.14	$3.07 \pm 0.18^{***}$	$34.92 \pm 3.20^*$
PE-25	264.30 ± 38.04	$106.80 \pm 7.95^{\#\#}$	$2.37 \pm 0.07^{###}$	$27.47 \pm 1.51^{\#}$
PE-100	$205.70 \pm 22.92^{\#}$	110.60 ± 7.14 [#]	$2.23 \pm 0.12^{####}$	$25.86 \pm 1.42^{##}$
PE-400	$222.60 \pm 40.00^{\#}$	$104.10 \pm 11.25^{##}$	$2.34 \pm 0.08^{\#\#}$	$25.18 \pm 1.06^{###}$
PE-1600	$159.40 \pm 45.05^{\#\#}$	$96.24 \pm 9.87^{###}$	$2.09 \pm 0.10^{\#\#\#}$	$25.31 \pm 0.88^{\#\#}$

Data are presented as the mean \pm standard error of mean (SEM). Sham, sham operation; OVX, ovariectomy; PE-25, OVX + PE 25 mg/kg; PE-100, OVX + PE 100 mg/kg; PE-400, OVX + PE 400 mg/kg; PE-1600, OVX + PE 1600 mg/kg; CTX-1, C-terminal telopeptide fragment of type I collagen; DYP, deoxypyridinoline; PYD, pyridinoline. *p < 0.05 and **** p < 0.001 vs. sham group; *p < 0.05, *p < 0.01, *p < 0.001, and **** p < 0.001 vs. OVX group.

Blood Ca and P levels reflect the degree of bone loss [26]. This study showed that Ca levels in the OVX group were slightly lower than those in the sham group. However, PE treatment (400 and 1600 mg/kg) significantly increased the blood Ca levels in OVX rats. Biochemical markers of bone turnover have been widely used to assess the metabolic activity of bone remodeling [27, 28]. The results indicated that femur BMD was lower in OVX rats than in sham rats. Bone turnover was increased by OVX-induced estrogen deficiency, as evidenced by higher bone resorption (CTX, DPD, and PYD) and higher bone formation (OC) in OVX

rats than in sham rats. These findings are consistent with those of previous studies [29, 30]. In contrast, PE supplementation significantly suppressed the increase in bone turnover marker levels, suggesting that PE may have sufficient estrogenic activity to affect the bone turnover rate in OVX rats.

It has been reported that blood serotonin levels increase with an increase in bone turnover in humans and in an OVX rat model of postmenopausal osteoporosis [31]. In the present study, we found that circulating serotonin significantly increased in the OVX group. However, PE treatment

significantly inhibited the increase in serotonin levels. Therefore, serotonin levels may be used as a marker of postmenopausal osteoporosis.

5. Conclusions

In the present study, it was demonstrated that PE exhibits phytoestrogen-like activities in OVX rats. PE supplementation increased the plasma estradiol level, uterine layer thickness, and ER- α protein expression, without affecting serum levels of hepatic injury biomarkers. Moreover, PE treatment inhibited the increase in bone turnover markers such as CTX-1, OC, DYP, and PYD in OVX rats. Overall, PE might be a potential functional food for controlling menopausal symptoms.

Data Availability

The data used to support the findings of this study are included within the article.

Conflicts of Interest

The authors declare that they have no conflicts of interest.

Acknowledgments

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References

- [1] J. V. Pinkerton, C. J. Guico-Pabia, and H. S. Taylor, "Menstrual cycle-related exacerbation of disease," *American Journal of Obstetrics and Gynecology*, vol. 202, no. 3, pp. 221–231, 2010.
- [2] H. J. Cho, H.-j. Jun, J. H. Lee et al., "Acute effect of high-dose isoflavones from Pueraria lobata (Willd.) Ohwi on lipid and bone metabolism in ovariectomized mice," *Phytotherapy Research*, vol. 26, no. 12, pp. 1864–1871, 2012.
- [3] A. C. Thorup, M. N. Lambert, H. S. Kahr, M. Bjerre, and P. B. Jeppesen, "Intake of novel red clover supplementation for 12 weeks improves bone status in healthy menopausal women," *Evidence-Based Complementary and Alternative Medicine*, vol. 2015, Article ID 689138, 11 pages, 2015.
- [4] L. J. Martin, S. Minkin, and N. F. Boyd, "Hormone therapy, mammographic density, and breast cancer risk," *Maturitas*, vol. 64, no. 1, pp. 20–26, 2009.
- [5] J. C. Stevenson, H. N. Hodis, J. H. Pickar, and R. A. Lobo, "Coronary heart disease and menopause management: the swinging pendulum of HRT," *Atherosclerosis*, vol. 207, no. 2, pp. 336–340, 2009.
- [6] I. M. C. M. Rietjens, J. Louisse, and K. Beekmann, "The potential health effects of dietary phytoestrogens," *British Journal of Pharmacology*, vol. 174, no. 11, pp. 1263–1280, 2017.
- [7] A. L. Russell, J. M. Grimes, D. F. Cruthirds et al., "Dietary isoflavone-dependent and estradiol replacement effects on body weight in the ovariectomized (OVX) rat," Hormone and metabolic research = Hormon-und Stoffwechselforschung = Hormones et metabolisme, vol. 49, no. 6, pp. 457–465, 2017.

- [8] M.-n. Chen, C.-c. Lin, and C.-f. Liu, "Efficacy of phytoestrogens for menopausal symptoms: a meta-analysis and systematic review," *Climacteric*, vol. 18, no. 2, pp. 260–269, 2015
- [9] H. M. A. Abdelrazek, M. M. A. Mahmoud, H. M. Tag et al., "Soy isoflavones ameliorate metabolic and immunological alterations of ovariectomy in female wistar rats: antioxidant and estrogen sparing potential," Oxidatvie Medicine and Celularl Longevity, vol. 2019, Article ID 5713606, 13 pages, 2019
- [10] D. Wing-Shing Cheung, C.-M. Koon, C.-F. Ng et al., "The roots of Salvia miltiorrhiza (Danshen) and Pueraria lobata (Gegen) inhibit atherogenic events: a study of the combination effects of the 2-herb formula," *Journal of Ethnopharmacology*, vol. 143, no. 3, pp. 859–866, 2012.
- [11] J. K. Prasain, K. Jones, M. Kirk et al., "Profiling and quantification of isoflavonoids in kudzu dietary supplements by high-performance liquid chromatography and electrospray ionization tandem mass spectrometry," *Journal of Agricultural and Food Chemistry*, vol. 51, no. 15, pp. 4213–4218, 2003.
- [12] S. Wang, S. Zhang, S. Wang et al., "A comprehensive review on Pueraria: insights on its chemistry and medicinal value," *Biomedicine & Pharmacotherapy*, vol. 131, Article ID 110734, 2020.
- [13] P. Loutchanwoot and T. Vortherms, "Effects of puerarin on estrogen-regulated gene expression in gonadotropin-releasing hormone pulse generator of ovariectomized rats," *Steroids*, vol. 135, pp. 54–62, 2018.
- [14] B. Li, M. Liu, Y. Wang et al., "Puerarin improves the bone micro-environment to inhibit OVX-induced osteoporosis via modulating SCFAs released by the gut microbiota and repairing intestinal mucosal integrity," *Biomedicine & Pharmacotherapy*, vol. 132, Article ID 110923, 2020.
- [15] H. Liu, W. Li, X. Ge, S. Jia, and B. Li, "Coadministration of puerarin (low dose) and zinc attenuates bone loss and suppresses bone marrow adiposity in ovariectomized rats," *Life Sciences*, vol. 166, pp. 20–26, 2016.
- [16] D. D. Thompson, H. A. Simmons, C. M. Pirie et al., "FDA Guidelines and animal models for osteoporosis," *Bone*, vol. 17, no. 4, pp. 125S–133S, 1995.
- [17] R. A. Lobo, S. R. Davis, T. J. De Villiers et al., "Prevention of diseases after menopause," *Climacteric*, vol. 17, no. 5, pp. 540–556, 2014.
- [18] P. P. Lelovas, T. T. Xanthos, S. E. Thoma, G. P. Lyritis, and I. A Dontas, "The laboratory rat as an animal model for osteoporosis research," *Comparative Medicine*, vol. 58, no. 5, pp. 424–430, 2008.
- [19] T. Tanaka, Y. Yokota, H. Tang, N. Zaima, T. Moriyama, and Y. Kawamura, "Anti-hyperglycemic effect of a kudzu (pueraria lobata) vine extract in ovariectomized mice," *Journal of Nutritional Science & Vitaminology*, vol. 62, no. 5, pp. 341–349, 2016.
- [20] T. Nakamura, Y. Katsu, H. Watanabe, and T. Iguchi, "Estrogen receptor subtypes selectively mediate female mouse reproductive abnormalities induced by neonatal exposure to estrogenic chemicals," *Toxicology*, vol. 253, no. 1-3, pp. 117–124, 2008.
- [21] J. Liu, Y.-C. Shi, and D. Y.-W. Lee, "Applications of Pueraria lobata in treating diabetics and reducing alcohol drinking," *Chinese Herbal Medicines*, vol. 11, no. 2, pp. 141–149, 2019.
- [22] S. Panneerselvam, R. M. Packirisamy, Z. Bobby, S. Elizabeth Jacob, and M. G. Sridhar, "Soy isoflavones (Glycine max) ameliorate hypertriglyceridemia and hepatic steatosis in high fat-fed ovariectomized Wistar rats (an experimental model of

- postmenopausal obesity)," The Journal of Nutritional Biochemistry, vol. 38, pp. 57-69, 2016.
- [23] V. Ribas, M. T. A. Nguyen, D. C. Henstridge et al., "Impaired oxidative metabolism and inflammation are associated with insulin resistance in ERα-deficient mice," American Journal of Physiology Endocrinology And Metabolism, vol. 298, no. 2, pp. E304–E319, 2010.
- [24] A. Yasrebi, J. A. Rivera, E. A. Krumm, J. A. Yang, and T. A. Roepke, "Activation of estrogen response element-independent ERα signaling protects female mice from dietinduced obesity," *Endocrinology*, vol. 158, no. 2, pp. 319–334, 2017
- [25] S. Thammacharoen, N. Geary, T. A. Lutz, S. Ogawa, and L. Asarian, "Divergent effects of estradiol and the estrogen receptor-α agonist PPT on eating and activation of PVN CRH neurons in ovariectomized rats and mice," *Brain Research*, vol. 1268, pp. 88–96, 2009.
- [26] D.-W. Guo, Y.-X. Han, L. Cong, D. Liang, and G.-J. Tu, "Resveratrol prevents osteoporosis in ovariectomized rats by regulating microRNA-338-3p," *Molecular Medicine Reports*, vol. 12, no. 2, pp. 2098–2106, 2015.
- [27] F. R. Singer and D. R. Eyre, "Using biochemical markers of bone turnover in clinical practice," *Cleveland Clinic Journal of Medicine*, vol. 75, no. 10, pp. 739–750, 2008.
- [28] J. Han and W. Wang, "Effects of tanshinol on markers of bone turnover in ovariectomized rats and osteoblast cultures," *PLoS One*, vol. 12, no. 7, Article ID e0181175, 2017.
- [29] D.-F. Ma, L.-Q. Qin, P.-Y. Wang, and R. Katoh, "Soy iso-flavone intake inhibits bone resorption and stimulates bone formation in menopausal women: meta-analysis of randomized controlled trials," *European Journal of Clinical Nutrition*, vol. 62, no. 2, pp. 155–161, 2008.
- [30] S. B. Mada, S. Reddi, N. Kumar et al., "Antioxidative peptide from milk exhibits antiosteopenic effects through inhibition of oxidative damage and bone-resorbing cytokines in ovariectomized rats," *Nutrition*, vol. 43-44, no. 44, pp. 21–31, 2017.
- [31] U. I. Mödder, S. J. Achenbach, S. Amin, B. L. Riggs, L. J. Melton, and S. Khosla, "Relation of serum serotonin levels to bone density and structural parameters in women," *Journal* of Bone and Mineral Research, vol. 25, no. 2, pp. 415–422, 2010.