

Evolutionary divergence and functions of the human acyl-CoA thioesterase gene (*ACOT*) family

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Abstract

The acyl-CoA thioesterase gene (*ACOT*) family encodes enzymes that catalyse the hydrolysis of acyl-CoA thioester compounds, also known as activated fatty acids, to their corresponding non-esterified (free) fatty acid and coenzyme A (CoASH). These enzymes play a very important role in lipid metabolism by maintaining cellular levels and proper ratios of free and activated fatty acids, as well as CoASH. Within the acyl-CoA family there are two distinct subgroups, type I and type II. Despite catalysing the same reaction, the two groups are not structurally similar and do not share sequence homology, strongly suggesting convergent evolution. This suggestion is further supported if one compares the human with the mouse and rat *ACOT* gene families. To date, four human type I *ACOT*s have been identified which belong to the α/β -hydrolase fold enzyme superfamily. Type II *ACOT*s fall into the 'hot dog' fold superfamily. There are currently six human type II genes; however, two homologous proteins, thioesterase superfamily members 4 (THEM4) and 5 (THEM5) share common type II structural features and, in the case of THEM4, acyl-CoA thioesterase activity — suggesting that the family may be larger than previously realised. Although recent studies have greatly expanded the current understanding of these proteins and their physiological importance, there are a number of members whose functions are relatively unexplored and which warrant further investigation.

Keywords: *ACOT*, acyl-CoA, thioesterase, hydrolase, gene family, evolution, human genome

Introduction

Acyl-CoA thioesterases (*ACOT*s) represent a group of enzymes that metabolise acyl-CoA esters to their corresponding non-esterified fatty acid and coenzyme A (CoASH). In addition to 'acyl-CoA thioesterases', these enzymes also have been described as 'acyl-CoA hydrolases', 'acyl-CoA thioester hydrolases' and 'deacylases'. The reaction catalysed by *ACOT* enzymes is very important during fatty acid metabolism. As such, *ACOT* genes are found throughout prokaryotic and eukaryotic organisms. In mammals, acyl-CoA

thioesterase expression is widespread and activity is detectable within many tissues and cell types. *ACOT* activity helps to regulate cellular pools, as well as proper ratios, of activated fatty acyl-CoAs (acyl-CoA esters), free fatty acids (non-esterified fatty acids) and CoASH.

These enzymes are found within a number of subcellular locations, including peroxisomes, mitochondria and the cytosol. Peroxisomal and mitochondrial *ACOT*s are thought to play a major role in fatty acid degradation via β -oxidation, which provides energy through the citric acid cycle, as

well as in ketone body formation. In order for a fatty acid to be transported into the peroxisome for β -oxidation, it must first be activated to its coenzyme A ester by acyl-CoA synthetases. Some acyl-CoA esters undergo β -oxidation much more slowly than others — which can effectively sequester large amounts of CoASH and limit the activity of acyl-CoA synthetases and the rate of subsequent fatty acid β -oxidation. Under these conditions, ACOTs may act to increase the levels of free CoASH, thereby restoring fatty acid β -oxidation.¹ Mammalian ACOTs are capable of metabolising a wide variety of compounds, including short-, medium- and long-chain acyl-CoAs. They can also metabolise polyunsaturated fatty acyl-CoAs, branched-chain acyl-CoAs and aromatic-acyl-CoAs.² These compounds are not only utilised during β -oxidation and fatty acid degradation, but also have roles in inflammation, ion channel opening, signal transduction, lipid biosynthesis and gene regulation.^{2,3}

Ten human *ACOT* genes have been identified to date, compared with 13 in mice and 12 in rats. These genes can be divided into two groups, based upon differences in sequence and structure. The first group includes type I ACOT proteins. These enzymes are members of the α/β -hydrolase fold enzyme superfamily, which includes other esterase-activity-exhibiting enzymes such as carboxylesterases and lipases.

Type II ACOTs make up the second group; these enzymes are members of the 'hot dog' fold enzyme superfamily, which includes a wide variety of functionally diverse proteins. Despite catalysing the same reaction, type I and type II enzymes share no similarity in terms of structure or sequence — indicating that the two groups are analogous and not homologous. This observation is reminiscent of the cytochrome P450 (*CYP*) and nitric oxide synthase (*NOS*) gene families. Studies with purified *NOS* proteins revealed that they possess P450-like activity; however, detailed alignment analyses showed <15 per cent amino acid identity, thereby confirming that these two gene families were not evolutionarily related.⁴ It was thus concluded that this was an example of convergent evolution. In

other words, during the evolution of enzyme activities needed at a particular subcellular location (probably more than 1 billion years ago) Mother Nature needed a 'P450-like activity' and assigned that task to the nearest protein, which happened to be an *NOS*. The same thing most likely happened to the type I versus type II ACOTs.

The revised nomenclature for human, mouse and rat acyl-CoA thioesterase genes was established by Hunt *et al.* and has been accepted by the HUGO Gene Nomenclature Committee.⁵ In recent years, a number of human ACOTs have been recombinantly expressed and enzymatically characterised, although the exact physiological function and importance of many of these enzymes remain unknown.

Human ACOT genes and evolution

To date, a total of ten genes have been identified within the human genome, all of which exhibit acyl-CoA thioesterase activity (Table 1). These ten genes can be sub-divided into two distinct groups, type I and type II, based on sequence and structural homology. Type I ACOTs comprise the smaller group, which contains only four genes: *ACOT1*, *ACOT2*, *ACOT4* and *ACOT6*. The larger group includes six genes, and these members are referred to as type II. There is no sequence homology between the two groups, and the translated proteins have very different domain architecture, indicating two distinct, evolutionarily unrelated, origins (Figure 1) and, most likely, an example of convergent evolution.

The type I proteins, *ACOT1*, *ACOT2*, *ACOT4* and *ACOT6*, share a high degree of sequence homology (Table 2). *ACOT1* and *ACOT2* are the most closely related, having 98 per cent amino acid sequence identity. *ACOT4* is 70 per cent identical to both *ACOT1* and *ACOT2*. *ACOT6* is the least homologous, but still shares 54–57 per cent sequence identity with the other members. None of these proteins shares any significant sequence homology with the type II proteins. The four type I genes form an 80 kilobase gene cluster on chromosome 14q24.3, indicating that they most likely have arisen as the result of gene duplication.

Table 1. List of all human (*ACOT*) with full gene name, aliases, chromosomal location, isoforms, NCBI RefSeq mRNA accession number, NCBI RefSeq protein accession number and total amino acid number (AA#)

Gene	Full gene name	Aliases	Chromosome	Isoforms	Ref mRNA	Ref protein	AA#
<i>ACOT1</i>	Acyl-CoA thioesterase 1	ACH2, CTE-I, LACH2	14q24.3		NM001037161.1	NP001032238	421
<i>ACOT2</i>	Acyl-CoA thioesterase 2	MTE1, PTE2, CTE1A, PTE2A, ZAP128	14q24.3		NM006821.4	NP006812.3	483
<i>ACOT4</i>	Acyl-CoA thioesterase 4	FLJ31235, PTE-Ib, PTE1B, PTE2B	14q24.3		NM152331.3	NP689544.3	421
<i>ACOT6</i>	Acyl-CoA thioesterase 6	C14orf42, c14_5530, MGCI61656, MGCI61658	14q24.3		NM001037162.1	NP001032239.1	207
<i>ACOT7</i>	Acyl-CoA thioesterase 7	BACH, ACH1, ACT, CTE-II, LACH, LACH1, MGCI126	1p36.31-p36.11	hBACHa	NM007274.3	NP009205.3	370
				hBACHb	NM181864.2	NP863654.1	380
				hBACHc	NM181865.2	NP863655.1	350
				hBACHd	NM181866.2	NP863656.1	329
<i>ACOT8</i>	Acyl-CoA thioesterase 8	hTE, PTE1, PTE2, PTE-2, HNAACTE, hACTE-III	20q13.12		NM005469.2	NP005460.2	319
<i>ACOT9</i>	Acyl-CoA thioesterase 9	CGI-16, MT-ACT48, ACATE2	Xp22.11	Isoform a	NM001037171.1	NP001032248.1	448
				Isoform b	NM001033583.2	NP001028755.2	439
<i>ACOT11</i>	Acyl-CoA thioesterase 11	BFIT, THEA, BFIT1, BFIT2, THEM1, STARD14, KIAA0707	1p32.3	BFIT1	NM015547.2	NP056362.1	607
				BFIT2	NM147161.2	NP671517.1	594

Continued

Table 1. Continued

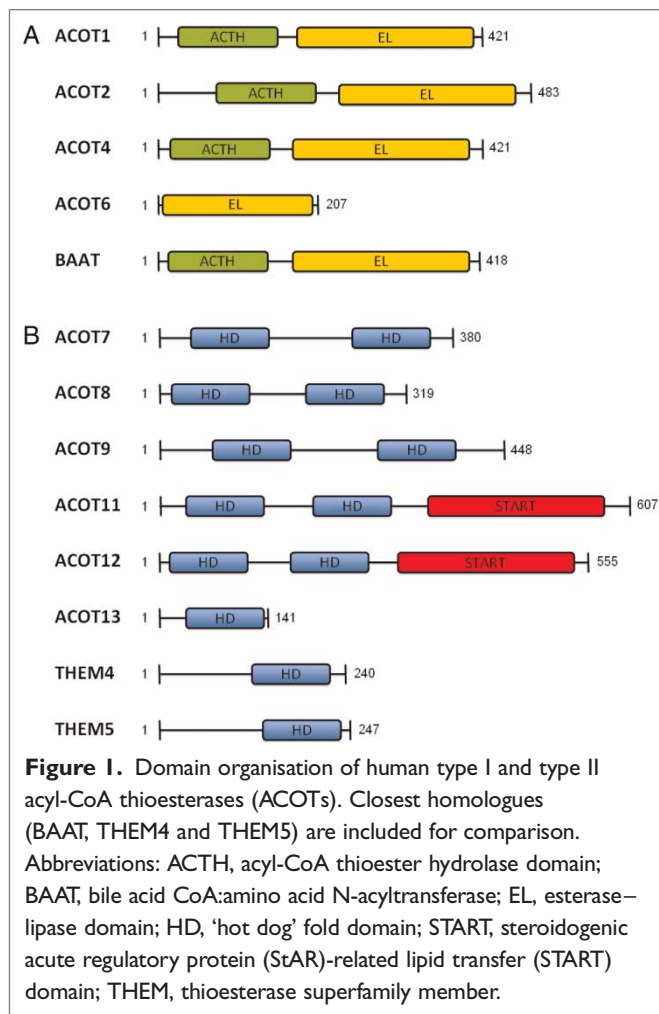
Gene	Full gene name	Aliases	Chromosome	Isoforms	Ref mRNA	Ref protein	AA#
ACOT12	Acyl-CoA thioesterase 12	Cach, THEAL, CACH-1, STARD15, MGCI05114	5q14.1		NM130767.2	NP570123.1	555
ACOT13	Acyl-CoA thioesterase 13	THEM2, HT012, MGC4961, PNAS-27	6p22.3	Isoform 1	NM018473.3	NP060943.1	140
				Isoform 2	NM018473.3	NP060943.1	117
THEM4	Thioesterase superfamily member 4	CTMP, MGC29636	1q21		NM053055.3	NP444283.2	240
THEM5	Thioesterase superfamily member 5	FLJ37964; MGC126659; MGC138444	1q21.3		NM182578.3	NP872384.1	247

A similar situation is observed within the mouse and rat orthologues *Acot1*, *Acot2*, *Acot3*, *Acot4*, *Acot5* and *Acot6*, which are clustered on chromosomes 12 D3 and 6q31, respectively. A cladogram depicting the evolutionary relationships observed between human, mouse and rat type I proteins is shown in Figure 2A. This cladogram clearly shows that, prior to the mammalian radiation approximately 75 million years ago (MYA), there was a single ancestral *ACOT1* (or -2) gene; after the mammalian radiation but before the mouse–rat split at approximately 15 MYA, this single ancestor divided twice to become four rodent *Acot* functional genes (now named *Acot1*, *Acot3*, *Acot4* and *Acot5*). By contrast, after the mammalian radiation, the human single ancestor duplicated into a second functioning gene, leading to *ACOT1* and *ACOT2*. On the other hand, *ACOT4* and *ACOT6* existed before the mammalian radiation and remain conserved to this day in human, mouse and rat.

In the mouse, the *ACOT3*, *ACOT4* and *ACOT5* enzymes each exhibit specific expression patterns and enzyme substrate specificity. Human *ACOT4* carries out the reactions catalysed by all three of these mouse enzymes. The lack of a human orthologue to mouse

Acot3 and *Acot5* was suggested to be the result of convergent evolution; it was hypothesised that convergent, functional evolution led to a reduced need for multiple enzymes in humans.⁶ The Protein Basic Local Alignment Search Tool (BLASTP) analysis of all non-redundant protein sequences, however, failed to identify orthologues for *ACOT3* or *ACOT5* in non-rodent species. This finding suggests that *ACOT4* was an ancestral gene encoding an *ACOT4* enzyme with broad substrate recognition, and that gene duplication events led to *ACOT1* (or -2), and that further duplication events led to the mouse *Acot3* and *Acot5* genes, encoding the *ACOT3* and *ACOT5* enzymes, which have similar substrate specificity to that of mouse *ACOT4* and human *ACOT4*.

Differences exist among the human, mouse and rat *ACOT6* orthologues. Analysis of the genomic *ACOT6* sequence indicates transcription of a complete mRNA containing three exons; however, a full-length mRNA is not detectable.⁶ A shorter transcript, in which the first exon was omitted, was identified. Subsequent translation begins at a methionine located within the second exon, producing a truncated protein that lacks the N-terminal acyl-CoA thioester hydrolase domain found within



other type I proteins. The truncated protein contains all residues required for catalysis and could therefore still produce an active acyl-CoA thioesterase. This is not true for the mouse or rat ACOT.

The BLAST results indicate that bile acid CoA:amino acid N-acyltransferase (BAAT) is the only enzyme to share significant sequence homology with type I ACOTs. BAAT shares 45 per cent sequence identity with ACOT1, ACOT2 and ACOT4. ACOT6 shares 37 per cent sequence identity with BAAT. BAAT is known to participate in the transfer of bile acids from the acyl-CoA thioester to either glycine or taurine, which are used for bile salt secretion. BAAT contains the same domain architecture and ACOT1, ACOT2 and ACOT4; however, the catalytic triad found within the esterase–lipase domain contains a cysteine, instead of the highly conserved serine

residue. This substitution may explain the difference in activity (ie transferase versus hydrolase) observed between the two enzymes. Despite sharing catalytic domains, other α/β -hydrolase fold enzyme superfamily esterases, such as carboxylesterase 1 (CES1) and acetylcholinesterase (ACHE), share very little sequence homology with their acyl-CoA thioesterase relatives. CES1 has thus been added as an evolutionarily unrelated gene to Figure 2A simply to ‘ground’ the dendrogram.

Type II ACOTs are strikingly less related than type I proteins. None of the type II proteins shares more than 25 per cent identity and many do not share any significant sequence similarity (Table 2). Unlike the type I genes, type II ACOTs are widely distributed throughout the genome. With the exception of ACOT13, all type II proteins contain double ‘hot dog’ domains, which may have evolved as a gene duplication event that allowed for the accommodation of bulky substrates.⁷ ACOT7 shares 19 per cent identity with ACOT9, 22 per cent with ACOT11 and 24 per cent identity with ACOT12. ACOT9 shares 22 per cent identity with ACOT12 but displays no homology with ACOT11. ACOT8 has no significant sequence homology with any of the other type II proteins. ACOT11 and ACOT12 share 51 per cent sequence identity and both enzymes contain a C-terminal steroidogenic acute regulatory protein (StAR)-related lipid transfer (START) domain, which seems to be specific to mammalian evolution.⁷

The human genome contains 15 START-containing proteins, including ACOT11 and ACOT12.⁸ The N-terminal region of both ACOTs is unrelated to any of the other START-containing enzymes, suggesting that these proteins may be the result of a gene fusion event between an acyl-thioesterase and START domain-containing sequences. Like ACOT8, ACOT13 shares no significant sequence similarity to the other type II proteins. BLAST results did identify 22 per cent sequence identity with thioesterase superfamily member 4 (THEM4 is the official name) — also known as Akt C-terminal modulator protein (CTMP). This small degree of homology suggests that the two proteins may share common evolutionary ties. Another

Table 2. Amino acid sequence identity between type I and type II human acyl-CoA thioesterases (ACOTs). Identity was determined using BLASTP and non-redundant human protein sequences. The predominant isoform sequences were used for ACOTs having multiple protein variants. Homology was considered not significant and left blank if the observed identity was less than 15 per cent

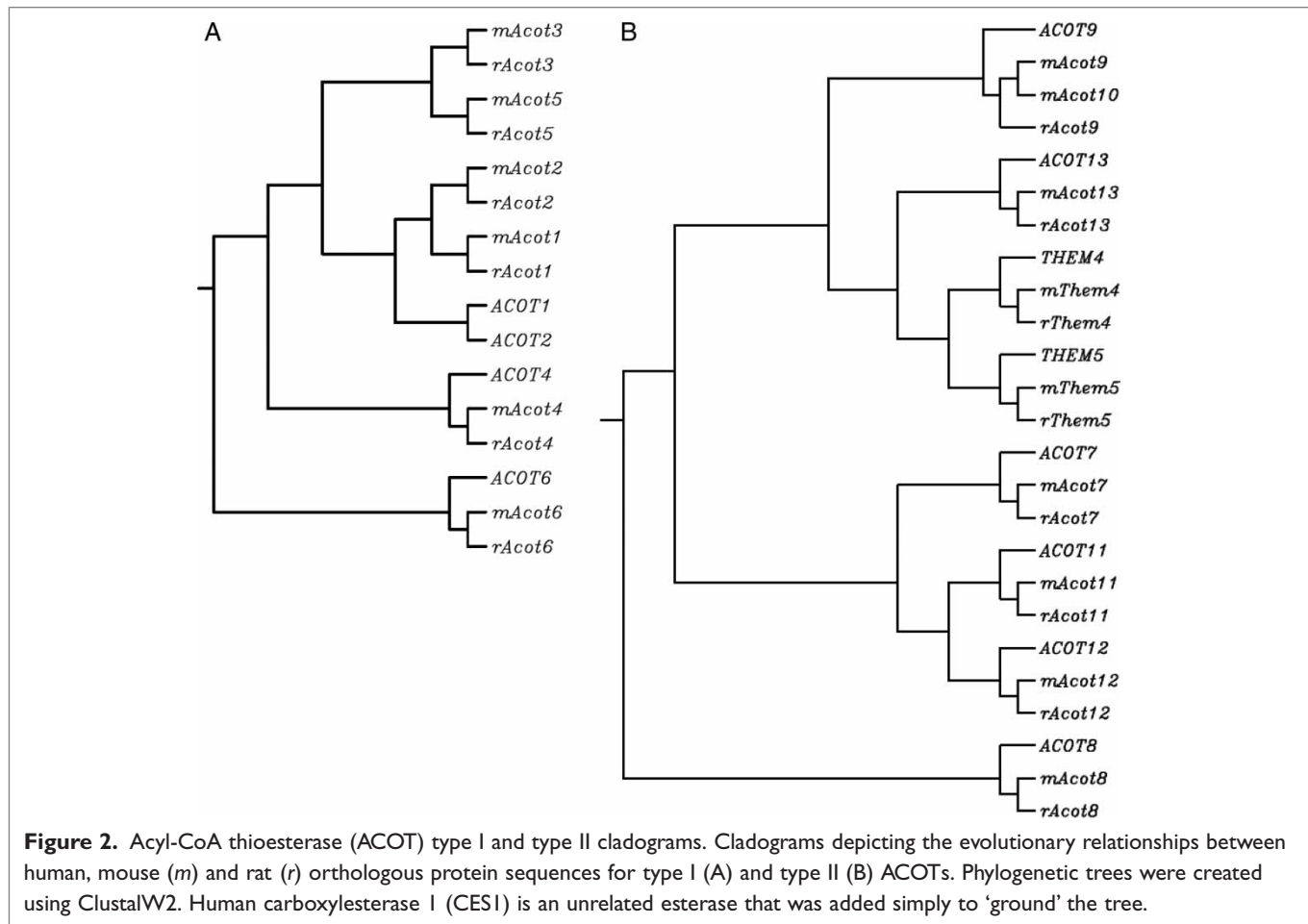
Type I ACOTs								
	ACOT1	ACOT2	ACOT4	ACOT6	BAAT			
ACOT1								
ACOT2	98%							
ACOT4	70%	70%						
ACOT6	57%	57%	54%					
BAAT	45%	45%	45%	37%				
Type II ACOTs								
	ACOT7	ACOT8	ACOT9	ACOT11	ACOT12	ACOT13	THEM4	THEM5
ACOT7								
ACOT8	—							
ACOT9	19%	—						
ACOT11	22%	—	—					
ACOT12	24%	—	22%	51%				
ACOT13	—	—	—	—	—			
THEM4	—	—	—	—	—	22%		
THEM5	—	—	—	—	—	—	37%	

Abbreviations: BAAT, bile acid CoA:amino acid N-acyltransferase; THEM, thioesterase superfamily member

protein, thioesterase superfamily member 5 (THEM5) shares more than 37 per cent identity with THEM4 but lacks significant sequence homology with ACOT13. Similar to ACOT13, both THEM4 and THEM5 contain a single ‘hot dog’ domain. Furthermore, type II proteins, THEM4 and THEM5 do not share sequence homology with other ‘hot dog’ domain superfamily members. The ‘hot dog’ fold-containing proteins possess highly conserved structural elements that are created by highly divergent sequences.⁹ This high degree of divergence makes evolutionary comparisons difficult without three-dimensional structures, because conservation of structural interactions is not directly associated with residue conservation.

BLAST results indicate that mammalian type II ACOTs are evolutionarily conserved. A cladogram

depicting the evolutionary relationships observed between human, mouse and rat type II protein is shown in Figure 2B. The mouse genome contains an additional type II ACOT, *Acot10*, which shares approximately 95 per cent mRNA nucleotide identity with ACOT9.¹⁰ Unlike *ACOT3* and *ACOT5*, however, which are found in both mouse and rat, *ACOT10* has only been identified in mice, indicating that the gene duplication event occurred less than 17 MYA — that is, after the mouse–rat split. In mouse, *Acot9* is located at chromosomal location X F3 and *Acot10* is found on 15 A2. The amplification and movement of genes to a new chromosomal location is often facilitated by transposable elements, as opposed to duplications occurring within close proximity on a chromosome, which are attributed to errors during meiosis.¹¹ The remaining seven



type II *ACOT* genes are highly conserved among human, mouse and rat, confirming that these genes were all present in the ancestor prior to the mammalian radiation.

Type I and type II acyl-CoA thioesterases represent two structurally distinct enzyme groups. These groups catalyse the same reaction but have distinct evolutionary origins and reaction mechanisms. An unrooted dendrogram — created using protein sequences from both type I and type II enzymes, closest-related homologues (BAAT, THEM4 and THEM5), distant α/β -hydrolase fold homologues (CES1, ACHE) and β -actin (ACTB) as an unrelated sequence — exemplifies how dissimilar the two groups are from one another (Figure 3). It also illustrates how the type II proteins must have originated from a common, evolutionarily distant, predecessor (as reflected in individual branch proximity to the origin of the tree).

Type I ACOT structures

Type I ACOTs share a high degree of sequence and structural homology. ACOT1, ACOT2 and ACOT4 contain an N-terminal acyl-CoA thioester hydrolase domain (Pfam 04775). This domain does not participate directly in catalysis, but studies indicate that the N-terminus plays an important role in regulating enzyme activity.³ All type I proteins also contain a C-terminus esterase–lipase superfamily domain. All residues required for catalysis are located within this domain, which is also known as the α/β -hydrolase fold.

Proteins that share this common structural element make up the functionally diverse α/β -hydrolase fold superfamily of enzymes. Within this family are a number of other distantly related proteins that exhibit esterase activity, including carboxylesterases, ACHE and lipases. The superfamily has diverged greatly and

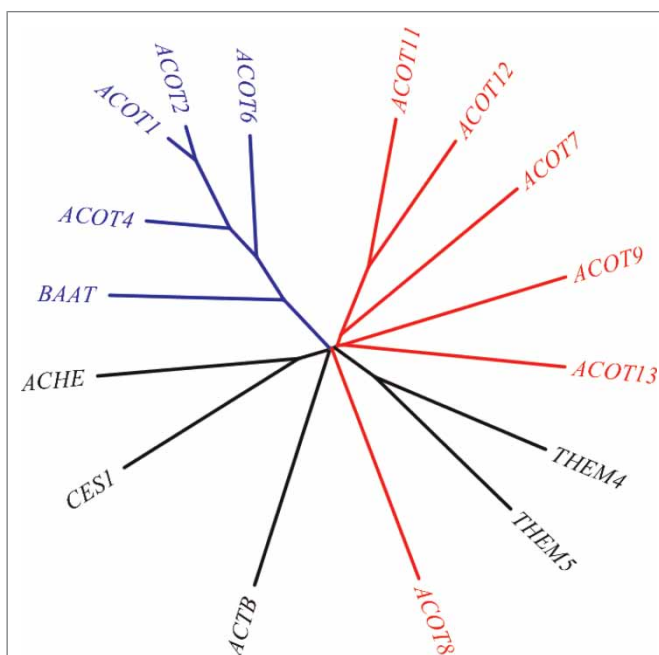


Figure 3. Unrooted dendrogram of human acyl-CoA thioesterases (ACOTs). Phylogenetic tree created using ClustalW2 and edited using TreeIllustrator. ACHE and CES1 were included as distant homologues belonging to the α / β -hydrolase fold superfamily. Bile acid acetyl Co-A:amino acid N-acyltransferase (BAAT), thioesterase superfamily members 4 and 5 (THEM4 and THEM5) are the closest-related homologues to type I (BAAT) and type II sequences (THEM4 and THEM5). ACTB was included as an unrelated sequence used to ground the tree. Type I ACOT branches are highlighted in blue, whereas type II are in red.
Abbreviations: ACHE, acetylcholinesterase; ACTB; β -actin; CES1, carboxylesterase I

many members share very little sequence homology but are considered evolutionarily linked, based on structural homology. The enzyme activity is dependent on a catalytic triad composed of highly conserved serine, histidine and aspartate residues found within the active site.¹² These three residues are conserved throughout type I ACOTs.

The crystalline structure for ACOT2 indicates that the enzyme is most likely biologically active as a monomer.¹³ The structure also reveals that the N-terminal acyl-CoA thioester hydrolase domain is connected to the active site by a long loop; it is postulated that this contributes substrate specificity, although this has yet to be verified experimentally.³ The four human type I ACOT genes are located on chromosome 14q24.3. ACOT1, ACOT2 and

ACOT3 each encode a transcript containing three exons. Genomic analysis suggests that ACOT6 should encode a similar 3 exon transcript; however, the only detectable transcript is truncated and utilises a start codon located within the 3' end of exon two, resulting in the synthesis of a truncated protein.⁶ In addition to the four protein-coding genes, there is one intronless pseudogene, referred to as ACOT4P, located on chromosome 19q13.12. ACOT4P is most likely the result of retroviral action that has inserted this DNA segment elsewhere in the genome. Attempts to translate the pseudogene results in numerous in-frame stop codons.

ACOT1, ACOT2 and ACOT4 have distinct sub-cellular localisation patterns and substrate specificity. ACOT1 is cytosolic and primarily responsible for the metabolism of long-chain acyl-CoAs.^{1,14} ACOT2 and ACOT1 can metabolise a similar range of substrates; however, ACOT2 contains an N-terminal mitochondrial targeting signal which directs its transport to mitochondria.⁶ ACOT4 contains a C-terminal peroxisomal targeting signal, directing transport to peroxisomes, where it exhibits high activity against succinyl-CoA, as well as a wide variety of short-chain dicarboxylic-CoAs. ACOT4 can also metabolise medium- and long-chain saturated- and unsaturated-CoAs.^{6,15} As previously mentioned, ACOT6 lacks the N-terminal acyl-CoA thioester hydrolase domain.

Although studies have yet to confirm the enzymatic activity of ACOT6, the fact that it contains a complete C-terminal esterase-lipase domain supports its role as a functional esterase.¹³ ACOT6 is predicted to be cytosolic, due to a missense mutation which alters the peroxisomal targeting signal that is found in the *Acot6* mouse ortholog.⁶ The subcellular localisation and substrate specificity for each enzyme suggest that each plays an important, non-redundant function within the cell.

Type II ACOT structures

The type II enzymes are more distantly related than type I ACOTs, as indicated by the relatively low degree of sequence conservation observed between proteins. Despite the low degree of sequence

homology, the enzymes are structurally related and capable of metabolising similar substrates. ACOT7, formerly called brain acyl-CoA hydrolase (BACH), has been extensively characterised, owing to recent studies that indicate arachidonoyl-CoA as a preferred substrate. The metabolism of arachidonoyl-CoA by ACOT7 gives rise to arachidonic acid and coenzyme A. Arachidonic acid serves as a precursor for a number of compounds associated with inflammation, including prostaglandins, leukotrienes and thromboxanes.^{16,17} ACOT7 also efficiently metabolises a number of long-chain-acyl-CoAs.¹⁸ ACOT8 is found in peroxisomes, where it metabolises a diverse range of acyl-CoA compounds.¹ Substrates include short-, medium- and long-chain acyl-CoAs. ACOT8 also has high activity towards branched- and methyl-branched chain acyl-CoAs.

ACOT9 contains a mitochondrial-targeting signal and metabolises medium- to long-chain acyl-CoAs. ACOT9 exhibits the highest activity towards myristoyl-CoA, which can then be used by N-myristoyltransferase to modify proteins posttranslationally.¹⁰ Expression of mouse ACOT11, formerly referred to as brown fat-inducible thioesterase (BFIT), increases after cold shock. This protein was found to play a role in regulating thermogenesis, which helps to maintain body temperature.¹⁹ The human homologue, ACOT11, may be a risk factor for obesity.¹⁹ As previously mentioned, ACOT11 and ACOT12 contain a C-terminal START domain. The START domain in animals is commonly found within homeodomain transcription factors, where lipid-binding regulates transcription.²⁰ The binding of lipids to the START domain in ACOT11 or ACOT12 acts as a mechanism for regulating activity and for allowing for rapid enzyme activation. The kinetic properties of ACOT11 have yet to be determined. ACOT12 has highest activity towards acetyl-CoA.²¹

ACOT13, previously called thioesterase superfamily member 2 (THEM2), differs from the other type II ACOTs, in that it contains a single 'hot dog' domain. Immunocytochemistry identified ACOT13 within the cytosol and nucleus.²² Another study indicated that the enzyme is mitochondrial.²³ The crystal structure and substrate specificity were recently

determined using recombinant protein; results indicate that ACOT13 prefers medium- to long-chain-acyl-CoA thioesters.²⁴ Interestingly, ACOT13 interacts with a START domain-containing protein named StarD2. The addition of StarD2 decreases the K_{cat} of ACOT13 for a number of substrates.²⁵ This finding suggests that the START domains, identified in ACOT11 and ACOT12, could have a similar effect on enzyme activity. BLAST results indicate >22 per cent sequence identity between ACOT13 and THEM4.

Two separate studies have revealed that THEM4 is a mitochondrial protein.^{26,27} Recent *in vitro* kinetic activity studies revealed that THEM4 is a broad-range, high-activity acyl-CoA thioesterase.²⁸ Similar to ACOT13, THEM4 preferentially metabolises medium- to long-chain acyl-CoA thioesters. As such, we recommend that the enzyme be renamed ACOT14 according to the accepted nomenclature established by Hunt *et al.*⁵ Acyl-CoA thioesterase activity has not been verified for THEM5; however, activity is probably based on the high degree of sequence identity with THEM4, but the exact role that this protein might play in fatty acid metabolism has yet to be determined.

Conclusions

The human acyl-CoA gene family represents a diverse group of enzymes that catalyse the hydrolysis of acyl-CoA thioesters to their corresponding free fatty acids and coenzyme A. These enzymes are involved in a number of cellular processes but are primarily thought to play an important role in lipid metabolism. Despite catalysing the same enzymatic reaction, the acyl-CoA thioesterase family members can be divided into two distinct groups, type I and type II. The two groups do not share common structural elements or sequence homology and are therefore considered analogous, rather than homologous, enzymes; this is probably another example of convergent evolution.

Type I proteins belong to the α/β -hydrolase fold superfamily, whereas type II proteins fall into the 'hot dog' fold superfamily. To date, four type I and six type II genes have been identified within the human

genome. There are two additional genes, *THEM4* and *THEM5*, which gene products share both structural and sequence homology with other type II proteins. *THEM4* was identified as having acyl-CoA thioesterase activity and should be considered as a new family member. In the future, perhaps *THEM5* should also be considered as a type II ACOT family member. Recent studies have greatly expanded the current understanding of acyl-CoA thioesterases; however, many precise physiological functions are not completely understood and warrant further investigation.

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